CALIFORNIA REGIONAL WATER QUALITY CONTROL BOARD
LOS ANGELES REGION

MONITORING AND REPORTING PROGRAM NO. 5720
for
SAINT-GOBAIN CONTAINERS
(Formerly Ball-Foster Glass Container Corporation)
(CA0000884)

I. Reporting Requirements

A. Saint-Gobain Containers (hereinafter SGC or Discharger) shall implement this monitoring program on the effective date of this Order. All monitoring reports shall be submitted semi-annually and must be received by the Regional Board by the dates in the following schedule. All monitoring reports should be addressed to the Regional Board, Attention: Information Technology Unit. The first monitoring report under this Program is due by February 1, 2006.

Reporting Period	Report Due
January – June	August 1
July-December	February 1
Annual Summary Report	March 1

If there is no discharge during any reporting period, the report shall so state.

B. B. The Discharger shall submit an annual report (for both dry and wet weather discharges from Discharge Serial No. 001), containing a discussion of the previous year's effluent and receiving water monitoring data, as well as graphical and tabular summaries of the data. The data shall be submitted to the Regional Board on hard copy and on a 3 ½ " computer diskette. Submitted data must be IBM compatible, preferably using EXCEL software. This annual report is to be received by the Regional Board by March 1 of each year following the calendar year of data collection. The Regional Board and the State Water Resources Control Board (State Board) are developing a database compliance monitoring management system that may require the Discharger to submit the monitoring and annual summary reports electronically when it becomes fully operational.

C. Each monitoring report shall contain a separate section titled "Summary of Non-Compliance" which discusses the compliance record and corrective actions taken or planned that may be needed to bring the discharge into full compliance with waste discharge requirements. This section shall clearly list all non-compliance with waste discharge requirements, as well as all excursions of effluent limitations.

D. The discharger shall inform the Regional Board well in advance of any proposed construction activity that could potentially affect compliance with applicable requirements.

II. Effluent Monitoring Requirements

A A sampling station shall be established for each point of discharge and shall be located where representative samples of effluent can be obtained. This monitoring shall occur at Discharge Serial No. 001 (Latitude 34° 04' 46" North and Longitude 118° 02' 34" West), prior to entering the storm drain located north of Valley Boulevard.

B. This Regional Board shall be notified in writing of any change in the sampling stations once established or in the methods for determining the quantities of pollutants in the individual waste streams.

C. Pollutants shall be analyzed using the analytical methods described in 40 CFR sections 136.3, 136.4, and 136.5 (revised May 14, 1999); or, where no methods are specified for a given pollutant, by this Regional Board or the State Board. Laboratories analyzing effluent samples and receiving water samples shall be certified by the California Department of Health Services Environmental Laboratory Accreditation Program (ELAP) or approved by the Executive Officer and must include quality assurance/quality control (QA/QC) data in their reports. A copy of the laboratory certification shall be provided each time a new certification and/or renewal of the certification is obtained from ELAP.

The monitoring reports shall specify the analytical method used, the Method Detection Limit (MDL), and the Minimum Level (ML) for each pollutant. For the purpose of reporting compliance with numerical limitations, performance goals, and receiving water limitations, analytical data shall be reported by one of the following methods, as appropriate:

1. An actual numerical value for sample results greater than or equal to the ML; or,
2. "Detected, but Not Quantified (DNQ)" if results are greater than or equal to the laboratory's MDL but less than the ML; or,
3. "Not-Detected (ND)" for sample results less than the laboratory's MDL with the MDL indicated for the analytical method used.

Current MLs (Attachment B) are those published by the State Water Resources Control Board in the *Policy for the Implementation of Toxics Standards for Inland Surface Waters, Enclosed Bays, and Estuaries of California, March 2, 2000*.

D. Where possible, the MLs employed for effluent analyses shall be lower than the Order limitations established for a given parameter. If the ML value is not below the effluent limitation, then the lowest ML value and its associated analytical method shall be selected for compliance purposes. At least once a year, the Discharger shall submit a list of the analytical methods employed for each test

and associated laboratory QA/QC procedures.

The Regional Board, in consultation with the State Board Quality Assurance Program, shall establish an ML that is not contained in Attachment B to be included in the Discharger's Order in any of the following situations:

1. When the pollutant under consideration is not included in Attachment B;

2. When the Discharger and Regional Board agree to include in the Order a test method that is more sensitive than that specified in 40 CFR Part 136 (revised May 14, 1999);

3. When the Discharger agrees to use an ML that is lower than that listed in Attachment B;

4. When the Discharger demonstrates that the calibration standard matrix is sufficiently different from that used to establish the ML in Attachment B, and proposes an appropriate ML for their matrix; or,

5. When the Discharger uses a method whose quantification practices are not consistent with the definition of an ML. Examples of such methods are the U.S. EPA-approved method 1613 for dioxins and furans, method 1624 for volatile organic substances, and method 1625 for semi-volatile organic substances. In such cases, the Discharger, the Regional Board, and the State Board shall agree on a lowest quantifiable limit and that limit will substitute for the ML for reporting and compliance determination purposes.

E. Water/wastewater samples must be analyzed within allowable holding time limits as specified in 40 CFR section 136.3. All QA/QC items must be run on the same dates the samples were actually analyzed, and the results shall be reported in the Regional Board format, when it becomes available, and submitted with the laboratory reports. Proper chain of custody procedures must be followed, and a copy of the chain of custody shall be submitted with the report.

F. All analyses shall be accompanied by the chain of custody, including but not limited to data and time of sampling, sample identification, and name of person who performed sampling, date of analysis, name of person who performed analysis, QA/QC data, method detection limits, analytical methods, copy of laboratory certification, and a perjury statement executed by the person responsible for the laboratory.

G. Temperature is a pollutant of concern for the furnace water discharge. Temperature shall be monitored continuously by a probe, placed at the discharge point and the results recorded on a chart for the entire duration of each discharge. The recorded chart shall be submitted with the monitoring report.

III. Effluent Monitoring Program

A. The effluent monitoring program for the discharge of glass quenching water through Discharge Serial No. 001 (Latitude 34° 04' 46" North and Longitude 118° 02' 34" West) is provided in the Table below.

B. In addition to monitoring to determine compliance with effluent limitations, the Discharger must monitor the effluent for priority pollutants at Discharge Serial No. 001 to determine reasonable potential. Pursuant to the California Water Code, section 13267, the Discharger is required to submit data sufficient for: (1) determining if WQBELs for priority pollutants are required, and (2) to calculate effluent limitations, if required. The *Policy for the Implementation of Toxics Standards for Inland Surface Waters, Enclosed Bays, and Estuaries of California* (March 2, 2000) requires that the Regional Boards require periodic monitoring for pollutants for which criteria or objectives apply and for which no effluent limitations have been established. Accordingly, the Regional Board is requiring that the Discharger monitor the effluent annually for the priority pollutants listed in Section VI. The results of monitoring for reasonable potential determination shall be submitted in accordance with Section I.A.

Pollutant	Units	Type of Sample	Sampling Frequency
Total waste flow	Gallons/day	Meter or Best estimation	All discharges
Temperature	°F	Continous recording [3]	All discharges
pH	Standard units	Grab	Once per discharge event
Boron	Mg/L	Grab	Once per discharge event
Total Suspended Solids	Mg/L, Lbs/day [1]	Grab	Once per discharge event
Settleable Solids	Ml/L	Grab	Once per discharge event
Oil and Grease	Mg/L, Lbs/day [1]	Grab	Once per discharge event
Ammonia	Mg/L	Grab	Once per discharge event
Biochemical Oxygen Demand (BOD) [2]	Mg/L, Lbs/day [1]	Grab	Once per discharge event
Turbidity	TU	Grab	Once per discharge event
Total Dissolved Solids	Mg/L, Lbs/day [1]	Grab	Once per discharge event
Chemical Oxygen Demand	Mg/L	Grab	Once per discharge event
Priority Pollutants (as listed in Section VI)	ì g/l	Grab	Once per discharge event
Acute Toxicity	% Survival	Grab	Once per discharge event

1. The mass emission (in lbs/day) for the discharge shall be calculated and

reported using the reported concentration and the actual flow rate measured at the time of the discharge, using the formula:

$m = 8.34C_iQ$

where:

m = mass for a pollutant in lbs/day

C_i = actual measured concentration for a pollutant, mg/L

Q = actual discharge flow rate in MGD

2. 5-day biochemical oxygen demand at 20 °C

3. Recorded chart of continuous temperature monitoring for the entire discharge shall be submitted with the monitoring report.

IV. Toxicity Monitoring Requirements

A. Acute Toxicity Effluent Monitoring Program

1. The Discharger shall conduct acute toxicity tests on effluent grab samples by methods specified in 40 CFR Part 136 which cites U.S. EPA's *Methods for Measuring the Acute Toxicity of Effluents and Receiving Waters to Freshwater and Marine Organisms*, Fifth Edition, October 2002, U.S. EPA, Office of Water, Washington D.C. (EPA/821-R-02-012) or a more recent edition to ensure compliance in 100 % effluent.

2. The fathead minnow, *Pimephales promelas*, shall be used as the test species for fresh water discharges and the topsmelt, *Atherinops affinis*, shall be used as the test species for brackish effluent. The method for topsmelt is found in U.S. EPA's *Short-term Method for Estimating the Chronic Toxicity of Effluents and Receiving Waters to West Coast Marine and Estuarine Organisms*, Third Edition, October 2002 (EPA/821-R-02-014).

3. In lieu of conducting the standard acute toxicity testing with the fathead minnow, the Discharger may elect to report the results or endpoint from the first 48 hours of the chronic toxicity test as the results of the acute toxicity test.

4. Effluent samples shall be collected after all treatment processes and before discharge to the receiving water.

B. Quality Assurance

1. Concurrent testing with a reference toxicant shall be conducted. Reference toxicant tests shall be conducted using the same test conditions as the effluent toxicity tests (e.g., same test duration, etc).

2. If either the reference toxicant test or effluent test does not meet all test acceptability criteria (TAC) as specified in the test methods manuals (EPA/821-R-02-013 and EPA/821-R-02-014), then the Discharger must re-sample and re-test at the earliest time possible.

3. Control and dilution water should be receiving water or laboratory water, as appropriate, as described in the manual. If the dilution water used is different from the culture water, a second control using culture water shall be used.

C. Accelerated Monitoring

1. If toxicity exceeds the limitations (as defined in Order No. R4-2005-0049, Sections I.B.3.a.i.), then the Discharger shall immediately implement accelerated testing as specified in Sections I.B.3.a.ii. The Discharger shall ensure that they receive results of a failing acute toxicity test within 24 hours of the close of the test and the additional tests shall begin within three business days of the receipt of the result. If the accelerated testing shows consistent toxicity, the Discharger shall immediately implement the Initial Investigation of the Toxicity Reduction Evaluation (TRE) Workplan.

2. If implementation of the initial investigation TRE Workplan indicates the source of toxicity (e.g., a temporary plant upset, etc.), then the Discharger may discontinue the Toxicity Identification Evaluation (TIE).

3. The first step in the initial Investigation TRE Workplan for downstream receiving water toxicity can be a toxicity test protocol designed to determine if the effluent from Discharge Serial No. 001 causes or contributes to the measured downstream acute toxicity. If this first step TRE testing shows that the Discharge Serial No. 001 effluent does not cause or contribute to downstream acute toxicity, using U.S. EPA's *Methods for Measuring the Acute Toxicity of Effluents and Receiving Waters to Freshwater and Marine Organisms*, Fifth Edition, October 2002, U.S. EPA, Office of Water, Washington D.C. (EPA/821-R-02-012), then a report on this testing shall be submitted to the Board and the TRE will be considered to be completed. Routine testing in accordance with MRP No.1558 shall be continued thereafter.

D. Steps in TRE and TIE procedures:

1. Following a TRE trigger, the Discharger shall initiate a TRE in accordance with the facility's initial investigation TRE workplan. The Discharger shall use EPA manuals EPA/600/2-88/070 (industrial) or EPA/833B-99/002 (municipal) as guidance or current versions. At a minimum, the TRE workplan must contain the provision in Attachment C. The Discharger shall expeditiously develop a more detailed TRE workplan for submittal to the

Executive Officer within 30 days of the trigger, which will include, but not be limited to:

a. Further actions to investigate and identify the cause of toxicity;

b. Actions the Discharger will take to mitigate the impact of the discharge and prevent the recurrence of toxicity;

c. Standards the Discharger will apply to consider the TRE complete and to return to normal sampling frequency; and,

d. A schedule for these actions.

2. The following is a stepwise approach in conducting the TRE:

a. Step 1 - Basic data collection. Data collected for the accelerated monitoring requirements may be used to conduct the TRE:

b. Step 2 - Evaluates optimization of the treatment system operation, facility housekeeping, and the selection and use of in-plant process chemicals;

c. If Steps 1 and 2 are unsuccessful, Step 3 implements a TIE and employment of all reasonable efforts and using currently available TIE methodologies. The objective of the TIE is to identify the substance or combination of substances causing the observed toxicity;

d. Assuming successful identification or characterization of the toxicant(s), Step 4 evaluates final effluent treatment options;

e. Step 5 evaluates in-plant treatment options; and,

f. Step 6 consists of confirmation once a toxicity control method has been implemented.

Many recommended TRE elements parallel source control, pollution prevention, and storm water control program best management practices (BMPs). To prevent duplication of efforts, evidence of implementation of these control measures may be sufficient to comply with TRE requirements. By requiring the first steps of a TRE to be accelerated testing and review of the facility's TRE workplan, a TRE may be ended in its early stages. All reasonable steps shall be taken to reduce toxicity to the required level. The TRE may be ended at any stage if monitoring indicates there is no longer toxicity (or six consecutive chronic toxicity results are less than or equal to 1.0 TU_c).

3. The Discharger may initiate a TIE as part of the TRE process to identify the cause(s) of toxicity. The Discharger shall use the EPA acute and chronic manuals, EPA/600/6-91/005F (Phase I)/EPA/600/R-96-054 (for marine), EPA/600/R-92/080 (Phase II), and EPA-600/R-92/081 (Phase III) as guidance.

4. If a TRE/TIE is initiated prior to completion of the accelerated testing schedule required by Part I.B.3.a.ii of this permit, then the accelerated testing schedule may be terminated, or used as necessary in performing the TRE/TIE, as determined by the Executive Officer.

5. Toxicity tests conducted as part of a TRE/TIE may also be used for compliance, if appropriate.

6. The Board recognizes that toxicity may be episodic and identification of causes of and reduction of sources of toxicity may not be successful in all cases. Consideration of enforcement action by the Board will be based in part on the Discharger's actions and efforts to identify and control or reduce sources of consistent toxicity.

E. Reporting

1. The Discharger shall submit a full report of the toxicity test results, including any accelerated testing conducted during the month as required by this permit. Test results shall be reported as % survival with the discharge monitoring reports (DMR) for the month in which the test is conducted.

2. If an initial investigation indicates the source of toxicity and accelerated testing is unnecessary, then those results also shall be submitted with the DMR for the period in which the investigation occurred.

   a. The full report shall be submitted on or before the end of the month in which the DMR is submitted.

   b. The full report shall consist of (1) the results; (2) the dates of sample collection and initiation of each toxicity test; (3) the acute toxicity average limit or chronic toxicity limit or trigger.

3. Test results for toxicity tests also shall be reported according to the appropriate manual chapter on Report Preparation and shall be attached to the DMR. Routine reporting shall include, at a minimum, as applicable, for each test:

   a. Sample date(s);

   b. Test initiation date;

c. Test species;

d. End point values for each dilution (e.g., number of young, growth rate, percent survival);

e. NOEC value(s) in percent effluent;

f. IC_{15}, IC_{25}, IC_{40} and IC_{50} values in percent effluent;

g. TU_c values $\left(TU_c = \frac{100}{NOEC} \right)$;

h. Mean percent mortality ($\pm$standard deviation) after 96 hours in 100% effluent (if applicable);

i. NOEC and LOEC values for reference toxicant test(s);

j. C_{25} value for reference toxicant test(s);

k. Any applicable charts; and

l. Available water quality measurements for each test (e.g., pH, D.O., temperature, conductivity, hardness, salinity, ammonia).

4. The Discharger shall provide a compliance summary, which includes a summary table of toxicity data from all samples collected during that year.

The Discharger shall notify by telephone or electronically, this Regional Board of any toxicity exceedance of the limit or trigger within 24 hours of receipt of the results followed by a written report within 14 calendar days of receipt of the results. The verbal or electronic notification shall include the exceedance and the plan the Discharger has taken or will take to investigate and correct the cause(s) of toxicity. It may also include a status report on any actions required by the permit, with a schedule for actions not yet completed. If no actions have been taken, the reasons shall be given.

V. Receiving Water (Rio Hondo Channel) Monitoring

A. Temperature Monitoring

Receiving water shall be monitored (during each effluent discharge event) at two points one hour after the start of discharge to storm drain. The two points of monitoring are one approximately 50 feet upstream and the second one 50 feet downstream of the discharge point (i.e., storm drain) into the receiving water.

B. Priority Pollutant Monitoring

Pursuant to the California Water Code, section 13267, the Discharger is required to submit data sufficient for: (1) determining if WQBELs for priority pollutants are required, and (2) to calculate effluent limitations, if required. The *Policy for the Implementation of Toxics Standards for Inland Surface Waters, Enclosed Bays, and Estuaries of California* (March 2, 2000) requires that the Regional Boards require periodic monitoring of the receiving water. Accordingly the Regional Board is requiring that the Discharger conduct annual receiving water monitoring for the priority pollutants listed in Section VI at the same time as effluent monitoring. Not more than two receiving water samples shall be collected during the life of the permit. The results of monitoring for reasonable potential determination shall be submitted in accordance with Section I.A of this Monitoring and Reporting Program. Receiving water sampling shall be conducted at the same time as the effluent monitoring. The receiving water monitoring location shall be within 50 feet upstream of the discharge point (i.e., storm drain) into the receiving water (i.e., Rio Hondo Channel).

The required monitoring frequency and type of sample for pH, hardness, salinity, and toxic pollutants are listed in Section VI of this Monitoring and Reporting Program.

VI. Effluent and Receiving Water Monitoring for Reasonable Potential

A. As described in Sections III.B and V of this Monitoring and Reporting Program, in accordance with the SIP, the Discharger is required to monitor both the effluent and receiving water for priority pollutants listed in the Table below to determine reasonable potential.

Receiving water sampling shall be conducted at the same time as the effluent sampling. Further, the Discharger must analyze pH, salinity, and hardness of the receiving water at the same time as priority pollutants in the receiving water.

This monitoring shall occur at the following locations:

- Discharge Serial No. 001 (Latitude 34° 04' 46" North and Longitude 118° 02' 34" West), prior to entering the storm drain north of Valley Boulevard.

- Receiving water. The monitoring stations shall be within 50 feet upstream of the discharge point (i.e., storm drain) into the receiving water (Rio Hondo Channel).

Constituent	Units	Type of Sample	Sampling frequency [1]
pH	Standard units	Grab	Once per discharge event [2]
Hardness (as $CaCO_3$)	mg/L	Grab	Once per discharge event [2]
Salinity	g/L	Grab	Once per discharge event [2]
Antimony	ì g/L	Grab	Once per discharge event
Arsenic [3]	ì g/L	Grab	Once per discharge event
Beryllium	ì g/L	Grab	Once per discharge event
Cadmium [3]	ì g/L	Grab	Once per discharge event
Chromium (III) [3]	ì g/L	Grab	Once per discharge event
Chromium (VI) [3]	ì g/L	Grab	Once per discharge event
Copper [3]	ì g/L	Grab	Once per discharge event
Lead [3]	ì g/L	Grab	Once per discharge event
Mercury	ì g/L	Grab	Once per discharge event
Nickel [3]	ì g/L	Grab	Once per discharge event
Selenium	ì g/L	Grab	Once per discharge event
Silver [3]	ì g/L	Grab	Once per discharge event
Thallium	ì g/L	Grab	Once per discharge event
Zinc [3]	ì g/L	Grab	Once per discharge event
Cyanide	ì g/L	Grab	Once per discharge event
Asbestos	ì g/L	Grab	Once per discharge event
Acrolein	ì g/L	Grab	Once per discharge event
Acrylonitrile	ì g/L	Grab	Once per discharge event
Benzene	ì g/L	Grab	Once per discharge event
Bromoform	ì g/L	Grab	Once per discharge event
Carbon tetrachloride	ì g/L	Grab	Once per discharge event
Chlorobenzene	ì g/L	Grab	Once per discharge event
Chlorodibromomethane	ì g/L	Grab	Once per discharge event
Chloroethane	ì g/L	Grab	Once per discharge event
2-Chloroethylvinyl ether	ì g/L	Grab	Once per discharge event
Chloroform	ì g/L	Grab	Once per discharge event
Dichlorobromomethane	ì g/L	Grab	Once per discharge event
1,1-Dichloroethane	ì g/L	Grab	Once per discharge event
1,2-Dichloroethane	ì g/L	Grab	Once per discharge event
1,1-Dichloroethylene	ì g/L	Grab	Once per discharge event
1,2-Dichloropropane	ì g/L	Grab	Once per discharge event
1,3-Dichloropropylene	ì g/L	Grab	Once per discharge event
Ethylbenzene	ì g/L	Grab	Once per discharge event
Methyl bromide	ì g/L	Grab	Once per discharge event
Methyl chloride	ì g/L	Grab	Once per discharge event
Methylene chloride	ì g/L	Grab	Once per discharge event

Constituent	Units	Type of Sample	Sampling frequency [1]
1,1,2,2-Tetrachloroethane	ì g/L	Grab	Once per discharge event
Tetrachloroethylene	ì g/L	Grab	Once per discharge event
Toluene	ì g/L	Grab	Once per discharge event
1,2-Trans-dichloroethylene	ì g/L	Grab	Once per discharge event
1,1,1-Trichloroethane	ì g/L	Grab	Once per discharge event
1,1,2-Trichloroethane	ì g/L	Grab	Once per discharge event
Trichloroethylene	ì g/L	Grab	Once per discharge event
Vinyl chloride	ì g/L	Grab	Once per discharge event
2-Chlorophenol	ì g/L	Grab	Once per discharge event
2,4-Dichlorophenol	ì g/L	Grab	Once per discharge event
2,4-Dimethylphenol	ì g/L	Grab	Once per discharge event
2-Methyl-4,6-Dinitrophenol	ì g/L	Grab	Once per discharge event
2,4-Dinitrophenol	ì g/L	Grab	Once per discharge event
2-Nitrophenol	ì g/L	Grab	Once per discharge event
4-Nitrophenol	ì g/L	Grab	Once per discharge event
3-Methyl-4-Chlorophenol	ì g/L	Grab	Once per discharge event
Pentachlorophenol	ì g/L	Grab	Once per discharge event
Phenol	ì g/L	Grab	Once per discharge event
2,4,6-Trichlorophenol	ì g/L	Grab	Once per discharge event
Acenaphthene	ì g/L	Grab	Once per discharge event
Acenaphthylene	ì g/L	Grab	Once per discharge event
Anthracene	ì g/L	Grab	Once per discharge event
Benzidine	ì g/L	Grab	Once per discharge event
Benzo (a) Anthracene	ì g/L	Grab	Once per discharge event
Benzo (a) Pyrene	ì g/L	Grab	Once per discharge event
Benzo (b) Fluoranthene	ì g/L	Grab	Once per discharge event
Benzo (g,h,i) Perylene	ì g/L	Grab	Once per discharge event
Benzo (k) Fluoranthene	ì g/L	Grab	Once per discharge event
Bis (2-Chloroethoxy) Methane	ì g/L	Grab	Once per discharge event
Bis (2-Chloroethyl) Ether	ì g/L	Grab	Once per discharge event
Bis (2-Chloroisopropyl) Ether	ì g/L	Grab	Once per discharge event
Bis (2-Ethylhexyl) Phthalate	ì g/L	Grab	Once per discharge event
4-Bromophenyl Phenyl Ether	ì g/L	Grab	Once per discharge event
Butylbenzyl Phthalate	ì g/L	Grab	Once per discharge event
2-Chloronapthalene	ì g/L	Grab	Once per discharge event
4-Chlorophenyl Phenyl Ether	ì g/L	Grab	Once per discharge event
Chrysene	ì g/L	Grab	Once per discharge event
Dibenzo (a,h) Anthracene	ì g/L	Grab	Once per discharge event
1,2-Dichlorobenzene	ì g/L	Grab	Once per discharge event
1,3-Dichlorobenzene	ì g/L	Grab	Once per discharge event
1,4-Dichlorobenzene	ì g/L	Grab	Once per discharge event

Constituent	Units	Type of Sample	Sampling frequency [1]
3,3'-Dichlorobenzidine	ì g/L	Grab	Once per discharge event
Diethyl Phthalate	ì g/L	Grab	Once per discharge event
Dimethyl Phthalate	ì g/L	Grab	Once per discharge event
Di-n-Butyl Phthalate	ì g/L	Grab	Once per discharge event
2,4-Dinitrotoluene	ì g/L	Grab	Once per discharge event
2,6-Dinitrotoluene	ì g/L	Grab	Once per discharge event
Di-n-Octyl Phthalate	ì g/L	Grab	Once per discharge event
1,2-Diphenylhydrazine	ì g/L	Grab	Once per discharge event
Fluoranthene	ì g/L	Grab	Once per discharge event
Fluorene	ì g/L	Grab	Once per discharge event
Hexachlorobenzene	ì g/L	Grab	Once per discharge event
Hexachlorobutadiene	ì g/L	Grab	Once per discharge event
Hexachlorocyclopentadiene	ì g/L	Grab	Once per discharge event
Hexachloroethane	ì g/L	Grab	Once per discharge event
Indeno (1,2,3-cd) Pyrene	ì g/L	Grab	Once per discharge event
Isophorone	ì g/L	Grab	Once per discharge event
Naphthalene	ì g/L	Grab	Once per discharge event
Nitrobenzene	ì g/L	Grab	Once per discharge event
N-Nitrosodimethylamine	ì g/L	Grab	Once per discharge event
N-Nitrosodi-n-Propylamine	ì g/L	Grab	Once per discharge event
N-Nitrosodiphenylamine	ì g/L	Grab	Once per discharge event
Phenanthrene	ì g/L	Grab	Once per discharge event
Pyrene	ì g/L	Grab	Once per discharge event
1,2,4-Trichlorobenzene	ì g/L	Grab	Once per discharge event
Aldrin	ì g/L	Grab	Once per discharge event
alpha-BHC	ì g/L	Grab	Once per discharge event
beta-BHC	ì g/L	Grab	Once per discharge event
gamma-BHC	ì g/L	Grab	Once per discharge event
delta-BHC	ì g/L	Grab	Once per discharge event
Chlordane	ì g/L	Grab	Once per discharge event
4,4'-DDT	ì g/L	Grab	Once per discharge event
4,4'-DDE	ì g/L	Grab	Once per discharge event
4,4'-DDD	ì g/L	Grab	Once per discharge event
Dieldrin	ì g/L	Grab	Once per discharge event
alpha-Endosulfan	ì g/L	Grab	Once per discharge event
beta-Endosulfan	ì g/L	Grab	Once per discharge event
Endosulfan Sulfate	ì g/L	Grab	Once per discharge event
Endrin	ì g/L	Grab	Once per discharge event
Endrin Aldehyde	ì g/L	Grab	Once per discharge event
Heptachlor	ì g/L	Grab	Once per discharge event
Heptachlor Epoxide	ì g/L	Grab	Once per discharge event

Constituent	Units	Type of Sample	Sampling frequency [1]
Polychlorinated Biphenyls [5]	ì g/L	Grab	Once per discharge event
Toxaphene	ì g/L	Grab	Once per discharge event

1. During periods of furnace drainage of glass quenching water, samples shall be collected during the first hour of the discharge.

2. Sampling for pH, hardness, and salinity of receiving water shall be concurrent with sampling for priority pollutants in receiving water.

3. Measured as total recoverable.

4. Refers to the sum of PCB Aroclors 1016, 1221, 1232, 1242, 1248, 1254, and 1260.

B. The Discharger must monitor the effluent and receiving water for the presence of the 16 congeners of 2,3,7,8-TCDD listed below, once during the first discharge. The Discharger is required to report for each congener the analytical results of the effluent monitoring, including the quantifiable limit and the Method Detection Limit (MDL), and the measured or estimated concentration. The Discharger must multiply each measured or estimated congener concentration by its respective Toxicity Equivalent Factors (TEFs) and report the sum of these values.

Congeners	TEF
2,3,7,8-Tetra CDD	1.0
1,2,3,7,8-penta CDD	1.0
1,2,3,4,7,8-hexa CDD	0.1
1,2,3,6,7,8-hexa CDD	0.1
1,2,3,7,8,9-hexa CDD	0.1
1,2,3,4,6,7,8-hepta CDD	0.01
Octa CDD	0.0001
2,3,7,8-tetra CDF	0.1
1,2,3,7,8 penta CDF	0.05
2,3,4,7,8-penta CDF	0.5
1,2,3,4,7,8-hexa CDF	0.1
1,2,3,6,7,8-hexa CDF	0.1
1,2,3,7,8,9-hexa CDF	0.1
2,3,4,6,7,8-hexa CDF	0.1
1,2,3,4,6,7,8-hepta CDF	0.01
1,2,3,4,7,8,9-hepta CDF	0.01
Octa CDF	0.0001

C. SWRCB-approved laboratory methods and the corresponding MLs for the examination of each priority pollutant are listed in Attachments B-1. Reporting

requirements for the data to be submitted are listed in Attachment D. We recommend that you select the analytical method from Attachment B capable of achieving the lowest ML for each pollutant as listed on Attachment B-1. ML is necessary for determining compliance for a priority pollutant when an effluent limit is below the MDL.

D. The laboratory analytical data shall include applicable MLs, MDL, quality assurance/quality control data, and shall comply with the reporting requirements contained in the Attachments B & C.

E. Please forward all interim monitoring data/reports to The Regional Board, Attn: Industrial Permitting Unit, and please include a reference to "Compliance File No. CI-5720 and NPDES No. CA0000884".

Ordered by: _____ Date: July 7, 2005
Jonathan S. Bishop
Executive Officer