**CALIFORNIA REGIONAL WATER QUALITY CONTROL BOARD**
**LOS ANGELES REGION**

**MONITORING AND REPORTING PROGRAM NO. 6895**
**for**
**PLASKOLITE WEST, INC.**
**(CA0060798)**

## I. Reporting Requirements

A. Plaskolite West, Inc. (hereinafter Plaskolite or Discharger) shall implement this monitoring program on the effective date of this Order. All monitoring reports shall be submitted quarterly and must be received by the Regional Board by the dates in the following schedule. All monitoring reports should be addressed to the Regional Board, Attention: Information Technology Unit. The first monitoring report under this Program is due by January 15, 2003

| Reporting Period | Report Due |
|---|---|
| January-March | April 15 |
| April-June | July 15 |
| July-September | October 15 |
| October-December | January 15 |

If there is no discharge during any reporting period, the report shall so state.

B. The Discharger shall submit an annual summary report (for both dry and wet weather discharges), containing a discussion of the previous year's effluent and receiving water monitoring data, as well as graphical and tabular summaries of the data. The data shall be submitted to the Regional Board on hard copy and on a 3 ½ " computer diskette. Submitted data must be IBM compatible, preferably using EXCEL software. In addition, the Discharger shall discuss the compliance record and the corrective actions taken or planned that may be needed to bring the discharge into full compliance with waste discharge requirements. This annual report is to be received by the Regional Board by March 1 of each year following the calendar year of data collection.

C. The Discharger shall inform the Regional Board well in advance of any proposed construction activity that could potentially affect compliance with applicable requirements.

## II. Effluent Monitoring Requirements

A. A sampling station shall be established for each point of discharge and shall be located where representative samples of that effluent can be obtained.

B. This Regional Board shall be notified in writing of any change in the sampling stations once established or in the methods for determining the quantities of pollutants in the individual waste streams.

C. Pollutants shall be analyzed using the analytical methods described in 40 CFR 136.3, 136.4, and 136.5 (revised May 14, 1999); or, where no methods are specified for a given pollutant, by methods approved by this Regional Board or the State Board. Laboratories analyzing effluent samples and receiving water samples shall be certified by the California Department of Health Services and must include quality assurance/quality control (QA/QC) data in their reports.

The monitoring reports shall specify the analytical method used, the Method Detection Limit (MDL), and the Minimum Level (ML) for each pollutant. For the purpose of reporting compliance with numerical limitations, performance goals, and receiving water limitations, analytical data shall be reported by one of the following methods, as appropriate:

1. An actual numerical value for sample results greater than or equal to the ML; or,
2. "Detected, but Not Quantified (DNQ)" if results are greater than or equal to the laboratory's MDL but less than the ML; or,
3. "Not-Detected (ND)" for sample results less than the laboratory's MDL with the MDL indicated for the analytical method used.

Current MLs (Attachment T-1) are those published by the State Water Resources Control Board in the *Policy for the Implementation of Toxics Standards for Inland Surface Waters, Enclosed Bays, and Estuaries of California, March 2, 2000*.

D. Where possible, the MLs employed for effluent analyses shall be lower than the permit limitations established for a given parameter. If the ML value is not below the effluent limitation, then the lowest ML value and its associated analytical method shall be selected for compliance purposes. At least once a year, the Discharger shall submit a list of the analytical methods employed for each test and associated laboratory QA/QC procedures.

The Regional Board, in consultation with the State Board Quality Assurance Program, shall establish an ML that is not contained in Attachment T-1 to be included in the Discharger's permit in any of the following situations:

1. When the pollutant under consideration is not included in Attachment T-1;

2. When the Discharger and Regional Board agree to include in the permit a test method that is more sensitive than that specified in 40 CFR Part 136 (revised May 14, 1999);

3. When the Discharger agrees to use an ML that is lower than that listed in Attachment T-1;

4. When the Discharger demonstrates that the calibration standard matrix is sufficiently different from that used to establish the ML in Attachment T-1, and proposes an appropriate ML for their matrix; or,

5. When the Discharger uses a method whose quantification practices are not consistent with the definition of an ML. Examples of such methods are the USEPA-approved method 1613 for dioxins and furans, method 1624 for volatile organic substances, and method 1625 for semi-volatile organic substances. In such cases, the Discharger, the Regional Board, and the State Board shall agree on a lowest quantifiable limit and that limit will substitute for the ML for reporting and compliance determination purposes.

E. Laboratory analyses – all chemical, bacteriological, and toxicity analyses shall be conducted at a laboratory certified for such analyses by the California Department of Health Services Environmental Laboratory Accreditation Program (ELAP). A copy of the laboratory certification shall be submitted with the Annual Report.

F. Water/wastewater samples must be analyzed within allowable holding time limits as specified in 40 CFR 136.3. All QA/QC items must be run on the same dates the samples were actually analyzed, and the results shall be reported in the Regional Board format, when it becomes available, and submitted with the laboratory reports. Proper chain of custody procedures must be followed, and a copy of the chain of custody shall be submitted with the report.

G. If applicable, annual effluent analyses shall be performed during the month of February. Results of annual analyses shall be reported in the appropriate quarterly monitoring report.

H. All analyses shall be accompanied by the chain of custody, including but not limited to data and time of sampling, sample identification, and name of person who performed sampling, date of analysis, name of person who performed analysis, QA/QC data, method detection limits, analytical methods, copy of laboratory certification, and a perjury statement executed by the person responsible for the laboratory.

I. Quarterly effluent analyses shall be performed during the months of February, May, August and November. Annual effluent analyses shall be performed during the month of February.

J. For parameters that both monthly average and daily maximum limits are specified and the monitoring frequency is less than four times a month, the following shall apply. If an analytical result is greater than the monthly average limit, the sampling frequency shall be increased (within one week of receiving the test results) to a minimum of once

weekly at equal intervals, until at least four consecutive weekly samples have been obtained, and compliance with the monthly average limit has been demonstrated. The Discharger shall provide for the approval of the Executive Officer a program to ensure future compliance with the monthly average limit.

## III. Effluent Monitoring Program

A. The following shall constitute the effluent monitoring program for the final effluent from Discharge No. 001:

| Constituent | Units | Type of Sample | Sampling Frequency |
|---|---|---|---|
| Flow | gal/day | ---- | quarterly |
| pH | standard units | grab | quarterly |
| Temperature | °F | grab | quarterly |
| Oil and Grease | mg/L | grab | quarterly |
| $BOD_5$ @ 20°C | mg/L | grab | quarterly |
| Total suspended solids | mg/L | grab | quarterly |
| Settleable Solids | ml/L | grab | semiannually |
| Methyl methacrylate | µg/L | grab | semiannually |
| Phosphorous | mg/L | grab | annually |
| Ethyl acrylate | µg/L | grab | annually |
| Polymethyl methacrylate | µg/L | grab | annually |
| Acrylonitrile | µg/L | grab | semiannually |
| Benzene | µg/L | grab | semiannually |
| Carbon Tetrachloride | µg/L | grab | semiannually |
| Chlorobenzene | µg/L | grab | semiannually |
| Chloroethane | µg/L | grab | semiannually |
| Chloroform | µg/L | grab | semiannually |
| 1,1-Dichloroethane | µg/L | grab | semiannually |
| 1,2-Dichloroethane | µg/L | grab | semiannually |
| 1,1-Dichloroethylene | µg/L | grab | semiannually |
| 1,2-Dichloropropane | µg/L | grab | semiannually |
| 1,3-Dichloropropylene | µg/L | grab | semiannually |
| Ethylbenzene | µg/L | grab | semiannually |
| Methyl Chloride | µg/L | grab | semiannually |
| Methylene Chloride | µg/L | grab | semiannually |
| Tetrachloroethylene | µg/L | grab | semiannually |
| Toluene | µg/L | grab | semiannually |
| 1,2-trans-Dichloroethylene | µg/L | grab | semiannually |
| 1,1,1-Trichloroethane | µg/L | grab | semiannually |
| 1,1,2-Trichloroethane | µg/L | grab | semiannually |
| Trichloroethylene | µg/L | grab | semiannually |
| Vinyl Chloride | µg/L | grab | semiannually |
| 2,4-Dimethylphenol | µg/L | grab | semiannually |
| 2,4-Dinitrophenol | µg/L | grab | semiannually |
| 2-Nitrophenol | µg/L | grab | semiannually |
| 4-Nitrophenol | µg/L | grab | semiannually |
| Phenol | µg/L | grab | semiannually |
| Acenaphthene | µg/L | grab | semiannually |

| | | | |
|---|---|---|---|
| Acenaphthylene | µg/L | grab | semiannually |
| Anthracene | µg/L | grab | semiannually |
| Benzo (a) Anthracene | µg/L | grab | semiannually |
| Benzo (a) Pyrene | µg/L | grab | semiannually |
| Benzo (k) Fluoranthene | µg/L | grab | semiannually |
| Bis (2-ethylhexyl) Phthalate | µg/L | grab | semiannually |
| Chrysene | µg/L | grab | semiannually |
| 1,2-Dichlorobenzene | µg/L | grab | semiannually |
| 1,3-Dichlorobenzene | µg/L | grab | semiannually |
| 1,4-Dichlorobenzene | µg/L | grab | semiannually |
| Diethyl Phthalate | µg/L | grab | semiannually |
| Dimethyl Phthalate | µg/L | grab | semiannually |
| Di-n-butyl Phthalate | µg/L | grab | semiannually |
| Fluoranthene | µg/L | grab | semiannually |
| Fluorene | µg/L | grab | semiannually |
| Hexachlorobenzene | µg/L | grab | semiannually |
| Hexachlorobutadiene | µg/L | grab | semiannually |
| Hexachloroethane | µg/L | grab | semiannually |
| Naphthalene | µg/L | grab | semiannually |
| Nitrobenzene | µg/L | grab | semiannually |
| Phenanthrene | µg/L | grab | semiannually |
| Pyrene | µg/L | grab | semiannually |
| 1,2,4-Trichlorobenzene | µg/L | grab | semiannually |
| 3,4-Benzofluoranthene | µg/L | grab | semiannually |
| 4,6-Dinitro-o-cresol | µg/L | grab | semiannually |
| Toxicity-acute | % survival | grab | semiannually |
| Toxicity-chronic | $TU_c$ | grab | semiannually |

## IV. Toxicity Monitoring Requirements

### A. Acute Toxicity Effluent Monitoring Program

1. The Discharger shall conduct acute toxicity tests on effluent grab samples by methods specified in 40 CFR Part 136 which cites USEPA's *Methods for Measuring the Acute Toxicity of Effluents to Freshwater and Marine Organisms*, Fourth Edition, August 1993 (EPA/600/4-90/027F) or a more recent edition to ensure compliance in 100 % effluent.

2. The fathead minnow, *Pimephales promelas*, shall be used as the test species for fresh water discharges and the topsmelt, *Atherinops affinis*, shall be used as the test species for brackish effluent. The method for topsmelt is found in USEPA's *Short-term Method for Estimating the Chronic Toxicity of Effluents and Receiving Waters to West Coast Marine and Estuarine Organisms*, First Edition, August 1995 (EPA/600/R-95/136).

3. In lieu of conducting the standard acute toxicity testing with the fathead minnow,

the Discharger may elect to report the results or endpoint from the first 48 hours of the chronic toxicity test as the results of the acute toxicity test.

4. Effluent samples shall be collected after all treatment processes and before discharge to the storm drain.

B. **Chronic Toxicity Effluent Monitoring Program**

1. The Discharger shall conduct critical life stage chronic toxicity tests on 24-hour composite 100 percent effluent samples in accordance with EPA's *Short Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms*, Third Edition, July 1994 (EPA/600/4-91/002) or EPA's *Short Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Marine and Estuarine Organisms*, August 1995, (EPA/600/R-95/136).

2. Effluent samples shall be collected after all treatment processes and before discharge to the storm drain.

   Test Species and Methods:

   a. The Discharger shall conduct tests as follows: with a vertebrate, an invertebrate, and a plant for the first three suites of tests. After the screening period, monitoring shall be conducted using the most sensitive species.

   b. Re-screening is required every 15 months. The Discharger shall re-screen with the three species listed above and continue to monitor with the most sensitive species. If the first suite of re-screening tests demonstrates that the same species is the most sensitive than re-screening does not need to include more than one suite of tests. If a different species is the most sensitive or if there is ambiguity then the Discharger shall proceed with suites of screening tests for a minimum of three, but not to exceed five suites.

   c. The presence of chronic toxicity shall be estimated as specified using West Coast marine organisms according to EPA's Short-Term Methods for Estimating Chronic Toxicity of Effluent and Receiving Waters to West Coast Marine and Estuarine Organisms, August 1995 (EPA/600/R-95/136).

C. **Quality Assurance**

1. Concurrent testing with a reference toxicant shall be conducted. Reference toxicant tests shall be conducted using the same test conditions as the effluent toxicity tests (e.g., same test duration, etc).

2. If either the reference toxicant test or effluent test does not meet all test acceptability criteria (TAC) as specified in the test methods manuals (EPA/600/4-91/002 and EPA/600/R-95/136), then the Discharger must re-sample and re-test at the earliest time possible.

3. Control and dilution water should be receiving water or laboratory water, as appropriate, as described in the manual. If the dilution water used is different from the culture water, a second control using culture water shall be used.

D. **Accelerated Monitoring**

1. If toxicity is detected as defined in this Order, then the Discharger shall conduct six additional tests, approximately every 7 days, over a six-week period. The samples shall be collected and the tests initiated no less than 7 days apart. The Discharger shall ensure that they receive results of a failing acute toxicity test within 24 hours of the close of the test and the additional tests shall begin within 3 business days of the receipt of the result. If two of the six tests (including the initial test) exceed 1.0 $TU_c$, the Discharger shall immediately implement the Toxicity Identification Evaluation (TIE).

2. If toxicity is not detected in any of the six additional tests required above, then the Discharger may return to the normal sampling frequency required in Part III.B. of this monitoring and reporting program.

3. The Discharger shall obtain 6 consecutive chronic toxicity results less than or equal to 1 TUc in order to return to the normal sampling frequency required in Part III.B. of this monitoring and reporting program.

4. The Board recognizes that toxicity may be episodic and identification of causes of and reduction of sources of toxicity may not be successful in all cases. Consideration of enforcement action by the Board will be based in part on the Discharger's actions and efforts to identify and control or reduce sources of consistent toxicity.

E. **Steps in Toxicity Reduction Evaluation (TRE) and Toxicity Identification Evaluation (TIE)**

1. Following a TRE trigger, the Discharger shall initiate a TRE in accordance with the facility's initial investigation TRE workplan. At a minimum, the Discharger shall consult EPA manuals EPA/600/2-88/070 (industrial) or EPA/833B-99/002 (municipal) as guidance. The Discharger will expeditiously develop and implement a more detailed TRE workplan for submittal to the Executive Officer within 15 days of the trigger, which includes:

   a. Further actions to investigate and identify the cause of toxicity;

   b. Actions the Discharger will take to mitigate the impact of the discharge and prevent the recurrence of toxicity;

   c. Standards the Discharger will apply to consider the TRE complete and for the return to normal sampling frequency; and,

   d. A schedule for these actions.

2. The following is a stepwise approach in conducting the TRE:

   a. Step 1 includes basic data collection;

   b. Step 2 evaluates optimization of the treatment system operation, facility housekeeping, and the selection and use of in-plant process chemicals;

   c. If Steps 1 and 2 are unsuccessful, Step 3 implements a Toxicity Identification Evaluation (TIE) and employment of all reasonable efforts using currently available TIE methodologies. The objective of the TIE shall be to identify the substance or combination of substances causing the observed toxicity;

   d. Assuming successful identification or characterization of the toxicant(s), Step 4 evaluates final effluent treatment options;

   e. Step 5 evaluates within plant treatment options; and,

   f. Step 6 consists of confirmation once a toxicity control method has been implemented.

      Many recommended TRE elements parallel source control, pollution prevention, and storm water control program best management practices (BMPs). To prevent duplication of efforts, evidence of complying with those requirements may be sufficient to comply with TRE requirements. By requiring the first steps of a TRE to be accelerated testing and review of the facility's TRE workplan, a TRE may be ended in its early stages. All reasonable steps shall be taken to reduce toxicity to the required level. The TRE may be ended at any stage if monitoring finds there is no longer toxicity

(or six consecutive chronic toxicity results less than or equal to 1 $TU_c$).

3. The Discharger may initiate a TIE as part of the TRE process to identify the cause(s) of toxicity. The Discharger shall consult the EPA acute and chronic manuals, EPA/600/6-91/005F (Phase I)/EPA/600/R-96-054 (for marine), EPA/600/R-92/080 (Phase II), and EPA-600/R-92/081 (Phase III) or other methods as approved by the Executive Officer of the Regional Board as guidance.

4. If a TRE/TIE is initiated prior to completion of the accelerated testing schedule required by Parts I.B.4.a.2. or I.B.4.b.2 of Order No. R4-2002-0126, then the accelerated testing schedule may be terminated, or used as necessary in performing the TRE/TIE, as determined by the Executive Officer.

5. Toxicity tests conducted as part of a TRE/TIE may also be used for compliance, if appropriate.

6. The Board recognizes that toxicity may be episodic and identification of causes of and reduction of sources of toxicity may not be successful in all cases. Consideration of enforcement action by the Board will be based in part on the Discharger's actions and efforts to identify and control or reduce sources of consistent toxicity.

F. **Reporting**

1. The Discharger shall submit a full report of the toxicity test results, including any accelerated testing conducted during the month as required by this permit. Test results shall be reported as % survival with the discharge monitoring reports (DMR) for the month in which the test is conducted.

2. If an initial investigation indicates the source of toxicity and accelerated testing is unnecessary, then those results also shall be submitted with the DMR for the period in which the investigation occurred.

3. The full report shall be submitted by the end of the month in which the DMR is submitted.

4. The full report shall consist of (1) the results; (2) the dates of sample collection and initiation of each toxicity test; (3) the acute toxicity average limit or chronic toxicity limit or trigger.

5. Test results for toxicity tests also shall be reported according to the appropriate manual chapter on Report Preparation and shall be attached to the DMR. Routine reporting shall include, at a minimum, as applicable, for each test:

a) sample date(s);
b) test initiation date;
c) test species;
d) end point values for each dilution (e.g., number of young, growth rate, percent survival);
e) NOEC value(s) in percent effluent;
f) $IC_{15}$, $IC_{25}$, $IC_{40}$ and $IC_{50}$ values in percent effluent;
g) $TU_c$ values $\left( TU_c = \frac{100}{NOEC} \right)$ ;
h) Mean percent mortality (±standard deviation) after 96 hours in 100% effluent (if applicable);
i) NOEC and LOEC values for reference toxicant test(s);
j) $C_{25}$ value for reference toxicant test(s);
k) Any applicable charts; and
l) Available water quality measurements for each test (e.g., pH, D.O., temperature, conductivity, hardness, salinity, ammonia).

6. The Discharger shall provide a compliance summary, which includes a summary table of toxicity data from at least eleven of the most recent samples.

7. The Discharger shall notify this Regional Board immediately of any toxicity exceedance and in writing 14 days after the receipt of the results of a monitoring limit or trigger. The notification will describe actions the Discharger has taken or will take to investigate and correct the cause(s) of toxicity. It may also include a status report on any actions required by the permit, with a schedule for actions not yet completed. If no actions have been taken, the reasons shall be given.

## V. Receiving Water Monitoring Requirements

The receiving water monitoring program shall consist of periodic surveys of Dominguez Channel and shall include studies of those physical-chemical characteristics of the receiving water that may be impacted by the discharge.

### A. Receiving Water Monitoring

1. Receiving Water Observations. General observations of the receiving water shall be made at each discharge point on a monthly basis and shall be reported in the quarterly monitoring report. If no discharge occurred during the observation period, this shall be reported.

   Observations shall be descriptive where applicable, such that colors, approximate amounts, or types of materials are apparent. The following observations shall be made:

a) Tidal stage, time, and date of monitoring
b) Weather conditions
c) Color of water
d) Appearance of oil films or grease, or floatable materials
e) Extent of visible turbidity or color patches
f) Direction of tidal flow
g) Description of odor, if any, of the receiving water
h) Presence and activity of California Least Tern and California Brown Pelican.

## VI. Interim Monitoring

Pursuant to the California Water Code, Section 13267 and in accordance with the SIP, the Discharger is hereby directed to conduct four quarters (from March 2002 to March 2003) of effluent and receiving water sampling/monitoring for all the constituents listed in Attachment A. To the extent there is any conflict between the requirements contained in this provision VI and a previous directive issued by the Regional Board on February 21, 2002, the requirements of this provision VI control. The Discharger may use any data previously collected and analyses previously performed in response to the aforementioned directive to comply with the requirements of this provision VI.

A. Effluent monitoring shall be conducted for all pollutants at the NPDES discharge point during periods when a discharge occurs.

B. Receiving water monitoring stations shall be at 50 feet upstream from the discharge point.

The required monitoring frequency and type of sample of the effluent and the receiving water for toxic pollutants are listed below:

| Constituent | Units | Type of Sample | Monitoring Frequency |
|---|---|---|---|
| pH | Standard Units | grab | monthly |
| Hardness (as $CaCO_3$) | mg/L | grab | monthly |
| PAHs | µg/L | grab | semiannually |
| Antimony | µg/L | grab | semiannually |
| Arsenic | µg/L | grab | semiannually |
| Beryllium | µg/L | grab | semiannually |
| Cadmium | µg/L | grab | semiannually |
| Chromium (III) | µg/L | grab | semiannually |
| Chromium (VI) | µg/L | grab | semiannually |
| Copper | µg/L | grab | semiannually |
| Lead | µg/L | grab | semiannually |
| Mercury | µg/L | grab | semiannually |
| Nickel | µg/L | grab | semiannually |
| Selenium | µg/L | grab | semiannually |
| Silver | µg/L | grab | semiannually |
| Thallium | µg/L | grab | semiannually |

| Constituent | Units | Type of Sample | Monitoring Frequency |
|---|---|---|---|
| Zinc | µg/L | grab | semiannually |
| Cyanide | µg/L | grab | semiannually |
| Asbestos | µg/L | grab | semiannually |
| Acrolein | µg/L | grab | semiannually |
| Acrylonitrile | µg/L | grab | semiannually |
| Benzene | µg/L | grab | semiannually |
| Bromoform | µg/L | grab | semiannually |
| Carbon tetrachloride | µg/L | grab | semiannually |
| Chlorobenzene | µg/L | grab | semiannually |
| Chlorodibromomethane | µg/L | grab | semiannually |
| Chloroethane | µg/L | grab | semiannually |
| 2-Chloroethylvinyl ether | µg/L | grab | semiannually |
| Chloroform | µg/L | grab | semiannually |
| Dichlorobromomethane | µg/L | grab | semiannually |
| 1,1-Dichloroethane | µg/L | grab | semiannually |
| 1,2-Dichloroethane | µg/L | grab | semiannually |
| 1,1-Dichloroethylene | µg/L | grab | semiannually |
| 1,2-Dichloropropane | µg/L | grab | semiannually |
| 1,3-Dichloropropylene | µg/L | grab | semiannually |
| Ethylbenzene | µg/L | grab | semiannually |
| Methyl bromide | µg/L | grab | semiannually |
| Methyl chloride | µg/L | grab | semiannually |
| Methylene chloride | µg/L | grab | semiannually |
| 1,1,2,2-Tetrachloroethane | µg/L | grab | semiannually |
| Tetrachloroethylene | µg/L | grab | semiannually |
| Toluene | µg/L | grab | semiannually |
| 1,2-Trans-dichloroethylene | µg/L | grab | semiannually |
| 1,1,1-Trichloroethane | µg/L | grab | semiannually |
| 1,1,2-Trichloroethane | µg/L | grab | semiannually |
| Trichloroethylene | µg/L | grab | semiannually |
| Vinyl chloride | µg/L | grab | semiannually |
| 2-Chlorophenol | µg/L | grab | semiannually |
| 2,4-Dichlorophenol | µg/L | grab | semiannually |
| 2,4-Dimethylphenol | µg/L | grab | semiannually |
| 2-Methyl-4,6-Dinitrophenol | µg/L | grab | semiannually |
| 2,4-Dinitrophenol | µg/L | grab | semiannually |
| 2-Nitrophenol | µg/L | grab | semiannually |
| 4-Nitrophenol | µg/L | grab | semiannually |
| 3-Methyl-4-Chlorophenol | µg/L | grab | semiannually |
| Pentachlorophenol | µg/L | grab | semiannually |
| Phenol | µg/L | grab | semiannually |
| 2,4,6-Trichlorophenol | µg/L | grab | semiannually |
| Acenaphthene | µg/L | grab | semiannually |
| Acenaphthylene | µg/L | grab | semiannually |
| Anthracene | µg/L | grab | semiannually |
| Benzidine | µg/L | grab | semiannually |
| Benzo (a) Anthracene | µg/L | grab | semiannually |
| Benzo (a) Pyrene | µg/L | grab | semiannually |
| Benzo (b) Fluoranthene | µg/L | grab | semiannually |

| Constituent | Units | Type of Sample | Monitoring Frequency |
|---|---|---|---|
| Benzo (g,h,i) Perylene | µg/L | grab | semiannually |
| Benzo (k) Flouranthene | µg/L | grab | semiannually |
| Bis (2-Chloroethoxy) Methane | µg/L | grab | semiannually |
| Bis (2-Chloroethyl) Ether | µg/L | grab | semiannually |
| Bis (2-Chloroisopropyl) Ether | µg/L | grab | semiannually |
| Bis (2-Ethylhexyl) Phthalate | µg/L | grab | semiannually |
| 4-Bromophenyl Phenyl Ether | µg/L | grab | semiannually |
| Butylbenzyl Phthalate | µg/L | grab | semiannually |
| 2-Chloronapthalene | µg/L | grab | semiannually |
| 4-Chlorophenyl Phenyl Ether | µg/L | grab | semiannually |
| Chrysene | µg/L | grab | semiannually |
| Dibenzo (a,h) Anthracene | µg/L | grab | semiannually |
| 1,2-Dichlorobenzene | µg/L | grab | semiannually |
| 1,3-Dichlorobenzene | µg/L | grab | semiannually |
| 1,4-Dichlorobenzene | µg/L | grab | semiannually |
| 3,3'-Dichlorobenzidine | µg/L | grab | semiannually |
| Diethyl Phthalate | µg/L | grab | semiannually |
| Dimethyl Phthalate | µg/L | grab | semiannually |
| Di-n-Butyl Phthalate | µg/L | grab | semiannually |
| 2,4-Dinitrotoluene | µg/L | grab | semiannually |
| 2,6-Dinitrotoluene | µg/L | grab | semiannually |
| Di-n-Octyl Phthalate | µg/L | grab | semiannually |
| 1,2-Diphenylhydrazine | µg/L | grab | semiannually |
| Fluoranthene | µg/L | grab | semiannually |
| Fluorene | µg/L | grab | semiannually |
| Hexachlorobenzene | µg/L | grab | semiannually |
| Hexachlorobutadiene | µg/L | grab | semiannually |
| Hexachlorocyclopentadiene | µg/L | grab | semiannually |
| Hexachloroethane | µg/L | grab | semiannually |
| Indeno (1,2,3-cd) Pyrene | µg/L | grab | semiannually |
| Isophorone | µg/L | grab | semiannually |
| Napthalene | µg/L | grab | semiannually |
| Nitrobenzene | µg/L | grab | semiannually |
| N-Nitrosodimethylamine | µg/L | grab | semiannually |
| N-Nitrosodi-n-Propylamine | µg/L | grab | semiannually |
| N-Nitrosodiphenylamine | µg/L | grab | semiannually |
| Phenanthrene | µg/L | grab | semiannually |
| Pyrene | µg/L | grab | semiannually |
| 1,2,4-Trichlorobenzene | µg/L | grab | semiannually |
| Aldrin | µg/L | grab | semiannually |
| alpha-BHC | µg/L | grab | semiannually |
| beta-BHC | µg/L | grab | semiannually |
| gamma-BHC | µg/L | grab | semiannually |
| delta-BHC | µg/L | grab | semiannually |
| Chlordane | µg/L | grab | semiannually |
| 4,4'-DDT | µg/L | grab | semiannually |
| 4,4'-DDE | µg/L | grab | semiannually |
| 4,4'-DDD | µg/L | grab | semiannually |
| Dieldrin | µg/L | grab | semiannually |

| Constituent | Units | Type of Sample | Monitoring Frequency |
|---|---|---|---|
| alpha-Endosulfan | µg/L | grab | semiannually |
| beta-Endosulfan | µg/L | grab | semiannually |
| Endosulfan Sulfate | µg/L | grab | semiannually |
| Endrin | µg/L | grab | semiannually |
| Endrin Aldehyde | µg/L | grab | semiannually |
| Heptachlor | µg/L | grab | semiannually |
| Heptachlor Epoxide | µg/L | grab | semiannually |
| Polychlorinated Biphenyls[1] | µg/L | grab | semiannually |
| Toxaphene | µg/L | grab | semiannually |

[1] The sum of Arochlors 1242, 1254, 1221, 1232, 1248, 1260, and 1016.

C. You may conduct the quarterly/semi-annually sampling during the periods prescribed in the monitoring and reporting section of your current permit, but the data must be submitted according to the Monitoring and Reporting Schedule which follows. However, if quarterly/semi-annually sampling is not required in your current permit, you must sample your effluent and the receiving water, and submit a report according to the Monitoring and Reporting Schedule below. Please note that the report for this required monitoring must be submitted separately from the self-monitoring reports.

| Monitoring and Reporting Schedule | |
|---|---|
| **Monitoring Period** | **Report Due Date** |
| January – March | April 15 |
| April – June | July 15 |
| July – September | October 15 |
| October – December | January 15 |
| Semi-annual sampling (to be conducted during October to March, and during April to September) | April 15 & October 15, respectively |

Semi-annual sampling results shall be reported in the quarterly reports submitted on the 15th day of April and October respectively

D. SWRCB-approved laboratory methods and the corresponding minimum levels (MLs) for the examination of each priority pollutant are listed in Attachment B. Reporting requirements for the data to be submitted are listed in Attachment C. We recommend that you select the analytical method from Attachment A capable of achieving the lowest ML for each pollutant as listed on Attachment B. ML is necessary for determining compliance for a priority pollutant when an effluent limit is below the MDL.

E. The laboratory analytical data shall include applicable MLs, MDL, quality assurance/quality control data, and shall comply with the reporting requirements

contained in the Attachments B & C.

F. The first monitoring report shall be due January 15, 2003 and the last report is due pursuant to this MRP on January 15, 2005, to this Regional Board. The last monitoring data shall include all the analytical data from the previous sampling events under this program. You must provide these analytical results in both electronic format (available as a Microsoft Excel Spreadsheet on our Web site **http://www.swrcb.ca.gov/rwqcb4/html/programs/watershed_reg.html**) and in paper format.

G. Please forward all interim monitoring data/report to The Regional Board, Attn: Industrial Permitting Unit, and please include a reference to "Compliance File No. CI-6895 and NPDES No. CA0060798".

H. Monitoring for TCDD Equivalents –The Discharger shall conduct effluent/receiving water monitoring for the presence of the 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD or Dioxin) congeners. The monitoring shall be a grab sample with a minimum frequency of once during dry weather and once during wet weather for 1 year. The Discharger shall calculate Toxic Equivalence (TEQ) for each congener by multiplying its analytical concentration by the appropriate Toxicity Equivalence Factors (TEF). Compliance with the dioxin limitation shall be determined by the summation of the 17 individual TEQs.

| Congeners | TEF |
|---|---|
| 2,3,7,8-tetra CDD | 1.0 |
| 1,2,3,7,8-penta CDD | 1.0 |
| 1,2,3,4,7,8-hexa CDD | 0.1 |
| 1,2,3,6,7,8-hexa CDD | 0.1 |
| 1,2,3,7,8,9-hexa CDD | 0.1 |
| 1,2,3,4,6,7,8-hepta CDD | 0.01 |
| Octa CDD | 0.0001 |
| 2,3,7,8-tetra CDF | 0.1 |
| 1,2,3,7,8-penta CDF | 0.05 |
| 2,3,4,7,8-penta CDF | 0.5 |
| 1,2,3,4,7,8-hexa CDF | 0.1 |
| 1,2,3,6,7,8-hexa CDF | 0.1 |
| 1,2,3,7,8,9-hexa CDF | 0.1 |
| 2,3,4,6,7,8-hexa CDF | 0.1 |
| 1,2,3,4,6,7,8-hepta CDF | 0.01 |
| 1,2,3,4,7,8,9-hepta CDF | 0.01 |
| Octa CDF | 0.0001 |

## VII. Storm Water Monitoring and Reporting

The Discharger shall implement the attached Storm Water Pollution Prevention Plan Requirements (Attachment D). The monitoring reports shall be received at the

Regional Board by July 1 of each year. Please reference Compliance File CI-6895 in the storm water monitoring reports submitted.

Ordered by: _____ Date: July 11, 2002

Dennis A. Dickerson
Executive Officer