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Auspices Statement

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EXECUTIVE SUMMARY

A critical component of the California State Water Board’s Groundwater Ambient Monitoring and Assessment (GAMA) Program is to assess the major threats to groundwater resources that supply drinking water to Californians (BELITZ et al., 2003). Nitrate is the most pervasive and intractable contaminant in California groundwater and is a focus of special studies under the GAMA program.

This report assesses the impact of Central Valley dairy operations on underlying groundwater quality and on groundwater processes using new tools developed during the course of the study. During the investigation, samples were collected and analyzed from a total of five dairies in the San Joaquin-Tulare Basins of California: three in Kings County, one in Stanislaus County, and one in Merced County (Figure 1). The study investigated water samples from production wells, monitor wells, and manure lagoons.

The three primary findings of this research are that dairy operations do impact underlying groundwater quality in California’s San Joaquin Valley, that dairy operations also appear to drive denitrification of dairy-derived nitrate in these groundwaters, and that new methods are available for characterization of nitrate source, transport and fate in the saturated zone underlying dairy operations.

This study demonstrated groundwater quality impact at three sites using a multi-disciplinary approach, and developed a new tool for source attribution in dairy groundwater. Negative groundwater quality impacts from dairy-derived nitrate were demonstrated using groundwater chemistry, nitrate isotopic composition, groundwater age, and transport modeling. A significant advance in characterization of groundwaters for nitrate source determination was the use of groundwater dissolved gas content to distinguish dairy wastewater irrigation from dairy wastewater lagoon seepage, both of which contributed to dairy groundwater contamination.

The demonstration of saturated-zone denitrification in dairy groundwaters is important in assessing the net impact of dairy operations on groundwater quality. The extent of denitrification can be characterized by measuring “excess” nitrogen and nitrate isotopic composition while the location of denitrification can be determined using a bioassay for denitrifying bacteria that developed in this research. In both northern and southern San Joaquin Valley sites, saturated-zone denitrification occurs and mitigates the impact of nitrogen loading on groundwater quality.

Other new methods developed during the course of this study include the field determination of denitrification in groundwater (allowing siting of monitor wells and mapping of denitrifying zones) and characterization of aquifer heterogeneity using direct-push drilling and geostatistics (allowing development of more accurate groundwater transport models). Application of these new methods in conjunction with traditional hydrogeologic and agronomic methods will allow a more complete and accurate understanding of the source, transport and fate of dairy-derived nitrogen in the subsurface.
STUDY SITES: HYDROGEOLOGIC SETTING

Two concentrations of dairies exist in the Central Valley of California, which is a low relief structural basin that is from 60 to 100 km wide and 700 km long. Both centers are in the southern two-thirds of the basin - the northern concentration is in Merced and Stanislaus Counties, and the southern concentration is in Kings and Tulare Counties. Both concentrations of dairies occur in the San Joaquin Valley Groundwater Basin, as designated by the California Department of Water Resources (2003). The San Joaquin Valley groundwater basin comprises two of the Central Valley’s three large structural sub-basins: the San Joaquin Basin and the Tulare Basin. In this document, we will use “San Joaquin Valley Basin” and “San Joaquin-Tulare Basin” interchangeably.

During the investigation, samples were collected and analyzed from a total of five dairies in the San Joaquin-Tulare Basins of California: three in Kings County, one in Stanislaus County, and one in Merced County (Figure 1). Groundwater samples were collected from production wells on each of the dairies. On three of the dairies, samples were also collected from monitoring wells: one of sites in Kings County was instrumented by LLNL, and the two sites in Stanislaus and Merced Counties were instrumented by UC-Davis. Samples were collected from manure lagoons at four of the sites.

Northern Sites

The two northern sites (SCD and MCD) are part of an extensive shallow groundwater monitoring network on five representative dairies set up by Thomas Harter of UC-Davis and the UC Cooperative Extension. The following description of the study area and the dairies is adapted from Harter et al. (2002).

The northern sites study area is in the central-eastern portion of the northern San Joaquin Valley, an area of low alluvial plains and fans bordered by the San Joaquin River to the west, tertiary upland terraces to the east, the Stanislaus River to the north, and the Merced River to the south. The region has a long history of nitrate and salt problems in groundwater (LOWRY, 1987; PAGE and BALDING, 1973).

The main regional aquifer is in the upper 100-200 m of basin deposits, which consist of Quaternary alluvial and fluvial deposits with some interbedded hardpan and lacustrine deposits. Groundwater generally flows from the ENE to the WSW following the slope of the landscape. The average regional hydraulic gradient ranges from approximately 0.05% to 0.15%. The water table at the selected facilities is between 2 and 5 m below ground surface. Measured K values range from 0.1 to 2 x 10^{-3} m/s, as consistent with the predominant texture of the shallow sediments.

The dominant surface soil texture is sandy loam to sand underlain by silty lenses, some of which are cemented with lime. Water holding capacity is low and water tables are locally high (and maintained by community drainage systems and shallow groundwater pumping). Border flood irrigation of forage crops has historically been the dominant cropping system among dairies in
the study area. Low-salinity (0.1–0.2 μS/cm) surface water from the Sierra Nevada is the main source of irrigation water.

Figure 1. Dairy Field Sites in the Central Valley.
Dairy Field Sites in the Central Valley Dairy study sites in Kings County (KCD1, KCD2, and KCD3), Merced County (MCD) and Stanislaus County (SCD) are shown with red triangles. Other sites where LLNL has conducted groundwater nitrate studies are shown with blue triangles.

A number of hydrogeologic criteria make the area suitable as a field laboratory for investigating recharge water quality from dairies: 1) Groundwater in the area is highly vulnerable because of the sandy soils with high infiltration rates and shallow water tables. 2) The shallow groundwater table and small long-term fluctuations in water level (1-2 m) allow sampling from vertically narrow groundwater zones with well-defined recharge source areas. 3) These same two factors also allow installation of a relatively inexpensive fixed-depth monitoring well network that is also inexpensive to sample.

The five dairy facilities in the UC-Davis network are progressive with respect to herd health, product quality, and overall operations. Improvements in manure and pond management have continually occurred since the inception of the project. The dairies are located in a geographic and hydrogeologic environment that is representative of many other dairies on the lowlands of the northern San Joaquin Valley. The manure management practices employed at these dairies over the past 35 years, particularly with respect to corral design, runoff capture, and lagoon
management, have been recognized by industry, regulators, and university extension personnel as typical or even progressive relative to other California dairies (see references in Harter et al., 2002). Over the past 30–40 years, the herd size on these dairies has continually grown from less than 100 at their inception to over 1000 animal units in the 1990s.

In 1993, UC-Davis installed 6 to 12 monitoring wells on each dairy for a total of 44 wells. Monitoring wells are strategically placed upgradient and downgradient from fields receiving manure water, near wastewater lagoons (ponds), and in corrals, feedlots, and storage areas (henceforth referred to as “corrals”). Wells are constructed with PVC pipe (3 or 5 cm diameter) and installed to depths of 7–10 m. The wells are screened from a depth of 2–3 m below ground surface to a depth of 10 m. Water samples collected from monitoring wells are representative of only the shallowest “first-encounter” groundwater.

**Southern Sites**

To augment the UC-Davis dairy monitoring network, LLNL chose to establish sites in the southern San Joaquin Valley groundwater basin. LLNL developed a list of five potential cooperators, sampled three sites, and chose to instrument one site. The cooperators were chosen with the expertise and assistance of the University of California Cooperative Extension (Thomas Harter, Carol Collar and Carol Frate). Sampling sites were chosen from the list of cooperative dairies using regional water quality data, including NAWQA data from the USGS and water quality dairy data from the Central Regional Water Quality Control Board (Fresno office). The site chosen for more extensive instrumentation was chosen with the following criteria: 1) a cooperative operator, 2) a shallow depth to groundwater to allow cost-effective installation of multi-level wells and synoptic soil-groundwater surveys, 3) a dairying operation typical for the region, and 4) regional evidence for nitrate contamination and denitrification.

The three dairies sampled are within the Tulare Lake Groundwater Subbasin of the San Joaquin Valley Groundwater Basin (California DWR, 2003) (Figure 1). The sites are located south of the Kings River and north-northeast of the Tulare Lake basin, the natural internal drainage for this hydrologically closed system. Groundwater hydraulic gradients are regionally from the Kings River toward Tulare Lake, but are generally low and are locally influenced by recharge from unlined irrigation canals and by agricultural and municipal groundwater extraction. Surface soils at these sites are predominantly Nord series (USDA National Resource Conservation Service, 2006), and are developed on distal Kings River alluvial fan deposits (Weissmann et al., 2003; Weissmann et al., 1999; Weissmann and Fogg, 1999; Weissmann et al., 2002a), which in general are less sandy and have more fine-grained interbeds than the sediments in the northern UC-Davis monitoring network. Groundwater levels in the area are in general deeper (50-200' below ground surface) and more variable (50' over 2-5 years) than in the north. A deeper depth to groundwater and heavier textured soils indicate that southern groundwaters should be less vulnerable to contamination than northern groundwaters. The regional groundwater is highly impacted by agricultural activities and contains elevated concentrations of nitrate and pesticides (Burrow et al., 1998b; Burrow et al., 1998).

Two of the three dairies sampled (KCD2 and KCD3) have deep water tables typical of the region. The one dairy that LLNL instrumented is located in an area to the west of Hanford.
characterized by a shallow perched aquifer, with depth to groundwater on the order of 15 feet. California Department of Water Resources (DWR) water level data for wells in the area indicate that this perched aquifer developed in the mid-1960’s in response to local groundwater overdrafting (CARLE et al., 2005), and is separated by an unsaturated zone from the deeper regional aquifer (that is sampled by wells on KCD2 and KCD3 to the east and south of Hanford).

The three dairy sites sampled by LLNL in Kings County each have close to the average of 1000 dairy cows, fed in free stalls with flush lanes. The manure management practices employed at these dairies, with respect to corral design, runoff capture, and lagoon management, are typical or progressive relative to other California dairies (see references in HARTER et al., 2002). The most intensively studied dairy, KCD1, operates three clay-lined wastewater lagoons that receive wastewater after solids separation. Wastewater is used for irrigation of 500 acres of forage crops (corn and alfalfa) on the dairy and on neighboring farms; dry manure is exported to neighboring farms. This dairy is also immediately adjacent to another dairy operation, and many of the conclusions regarding nitrate impact apply to dairy practices shared by both operations.

**STUDY SITES: SAMPLING AND INSTRUMENTATION**

**Kings County Dairy Site 1 (KCD1)**

Kings County Dairy #1 (KCD1; see Figure 1, Appendix A-Figure 1, and Appendix B-Figure 1), was the primary site in Kings County, and was sampled on multiple occasions, from existing production wells, from LLNL-installed monitor wells, from manure lagoons and irrigation canals, and with direct push soil and water sampling methods. A total of 31 days were devoted to collecting 139 water samples at the site, including 29 direct push samples, 17 surface water samples from 3 manure lagoons and a nearby irrigation canal, 16 groundwater samples from 9 production wells, and 60 groundwater samples from 17 monitor wells. A large number of subsurface soil samples were also collected, both as continuous drill core and as depth-discrete grab samples. Production and monitor wells were sampled on semi-regular intervals between August 2003 and August 2005.

KCD1 was instrumented with five sets of multi-level monitoring wells and one “up-gradient” well near an irrigation canal (Figure 2). The multi-level well “clusters” consisted of wells installed in separate boreholes approximately 5’ apart. A first set of three nested 2” wells in one cluster was installed in September 2003. In August 2004, three new well clusters were installed, each with four 2” wells. Also at that time, an upgradient 2” well was installed, and a small cluster of three 1.25” wells were installed. Two aquifers underlie the KCD1 dairy site, a shallow perched aquifer and a more regionally extensive deep aquifer. The deep aquifer is instrumented with one 2” well screened at 178-180’ below ground surface (bgs) that was installed in September 2003. The remaining monitor wells are all in the shallow perched aquifer and are screened between 18” and 65” bgs.

In August 2004, shortly before the second sets of well clusters were installed, a CPT/DP survey (see methods section) was conducted across the site (Figure 3). Depth discrete water and soils
samples were collected at this time, after which the holes were grouted and abandoned. With the exception of the upgradient monitor well near the canal, CPT/DP sites included locations near all of the multi-level monitor well clusters.

Figure 2. KCD1 Dairy Field Site.
KCD1 site, showing monitor wells and direct-push locations. Sites 1, 2, 3, and 4 (S1 through S4) are all multi-level two-inch monitor well clusters; site 5 (S5) is a single two-inch first-encounter well. The Site 1 cluster (S1) also includes a well in the deep aquifer. Direct-push (DP) and cone penetrometer (CPT) holes are also shown. CPT/DP was done at all multi-level well sites; it was not done at the single-level 5S site. Inset shows application of manure lagoon wastewater for furrow irrigation of silage corn crops at the site.
The production wells are screened in both the shallow and deep aquifer, and have 20-30’ long screens. Domestic supply wells, one of which was sampled, are screened in the deep aquifer, and typically have 20’ long screens. Agricultural supply wells, eight of which were sampled, typically have 30’ long screens, with the top of the screen at 30’ bgs. Information on screen length and depth is from conversations with the water well company which installed the more recent wells and has extensive experience in the region.

Figure 3. KCD1 field site with CPT/DP locations. Soil Behavior Type (SBT) profiles from Direct-Push Cone Penetrometer Testing on the KCD1 dairy field site. Large inset shows direct-push rig.
Kings County Dairy Sites 2 and 3 (KCD2 and KCD3)

The second and third Kings County dairy sites (Figure 1) were sampled during initial screening of Kings County sites in August 2003. At each site, groundwater pumped from a domestic supply well was analyzed for inorganic cations and anions (including nitrate, nitrite and ammonia), dissolved gases by membrane-inlet mass spectrometry, and tritium/helium-3 mean groundwater age by noble gas mass spectrometry. Groundwater in the area is 120-150 feet below ground surface, and the Corcoran Clay is generally 400-450' below ground surface and 90-100' thick. At each site, groundwater was sampled from wells screened between 200 and 300 feet below ground surface.

The second dairy was sampled again in April 2005. On this occasion, groundwater from the same domestic supply well sampled in 2003 was re-sampled, and manure lagoon and field water from six sampling locations was sampled. The groundwater was analyzed as before; while the lagoon water samples were analyzed for inorganic cations and anions (including nitrate, nitrite and ammonia), and dissolved gases by membrane-inlet mass spectrometry.

Merced and Stanislaus Dairy Sites (MCD and SCD)

MCD and SCD (Figure 1, Appendix A-Figure 1: The Merced County and Stanislaus County Dairies (MCD and SCD) were sampled on three occasions: August 2003, April 2005 and June 2005. Almost 40 samples were taken broken down as follows: 30 MCD samples and 9 SCD samples; 28 groundwater samples from 22 wells, 1 lagoon water sample, and 1 tile drain sample. Groundwater samples were analyzed for field parameters (temperature, conductivity, dissolved oxygen and ORP); inorganic cations and anions (including nitrate, nitrite and ammonia), dissolved gases by membrane-inlet mass spectrometry, tritium/helium-3 mean groundwater age by noble gas mass spectrometry, stable isotopic composition of nitrate and water, and organic co-contaminants. Tritium/helium-3 samples were not taken from the surface water sampling sites. These sites and data from these sites are described in Harter et al. (2002)

METHODS

Cone Penetrometer (CPT) and Direct Push (DP) Methods

Standard cone penetrometer/direct push methods were used to characterize the shallow hydrostratigraphy at the site. The survey was accomplished using a 20-25 ton CPT rig and accompanying support rig. The dead weight of the CPT rig was used to push the cone penetrometer to depths up to 90 feet using a hydraulic ram located at the center of the truck. Soil parameters such as cone bearing, sleeve friction, friction ratio and pore water pressure were measured as the cone penetrometer was advanced. These measurements were sent through the cone rods to the CPT rig's on-board data acquisition system. All data was processed in real time in the field, and CPT plots of tip resistance, sleeve friction, friction ratio and pore pressure were provided in the field along with a table of interpreted soil parameters. For development of
geostatistical models of subsurface hydraulic properties, soil behavior types determined by CPT (ROBERTSON et al., 1983) were calibrated and validated against a 200-foot continuous core log recovered from the first site (Figure 4.)

After CPT logging, a second hole was developed for collecting depth-discrete groundwater and soil samples using direct push methods. For water, a Hydropunch groundwater sample was taken at specified depth intervals. The Hydropunch operates by pushing 1.75-inch diameter hollow rods with a steel tip. A filter screen is attached to the tip. At the desired sampling depth, the rods are retracted, exposing the filter screen and allowing for groundwater infiltration. A small diameter bailer is then used to collect groundwater samples through the hollow rod. Typically, 4 or more 40 ml VOA vials were collected. For soil, a piston-type soil sampler was used to collect undisturbed soil samples (12" long x 1" diameter) that were stored on ice or dry ice immediately upon retrieval. After completion of logging and sampling, CPT/DP sampling holes were grouted under pressure with bentonite using the support rig.

![Figure 4. KCD Field Site CPT Logs.](image)

Comparison of soil behavior type (SBT) profile derived from CPT data to sediment texture profile as logged by a State of California certified drilling geologist at the KCD Site 1. Depth is shown in feet below ground surface. The thick sequence of sand between 25 and 55 feet shows up in both profiles, as does the confining unit at about 80 feet.

**Standard Drilling Methods**

Monitor wells were emplaced using standard methods. The first and deepest 200-foot bore-hole was drilled with a mud-rotary rig; subsequent wells were drilled using hollow-stem auger. In the
deep 200-foot hole, continuous log core was recovered and logged by a State-certified geologist (Figure 4) and down-hole geophysical data were obtained, including caliper, gamma ray, electromagnetic induction, and spontaneous potential and resistivity logs. Wells were cased with either 2" or 1.25" PVC pipe with short (generally 2') slotted screens and sand packs, and completed with a sanitary seal. Early wells (installed in 2003) were completed with stovepipe installation, which were subsequently converted to ground-level flush-mount installations in 2004 to accommodate farm activities. All wells installed in 2004 were completed with a flush-mount installation. The 2"-diameter wells were developed using standard bail, surge and pump methods.

**Sample Collection and Field Parameters**

Groundwater samples were collected after purging the well by either pumping or bailing, after determining water level against a marked datum. Groundwater from production wells was sampled, whenever possible, from upstream of any storage or pressure tank. A variety of methods were used to draw samples from monitor wells, depending on their diameter. Two-inch diameter monitor wells were sampled with a Grundfos MP-1 submersible pump and Teflon-lined sample line. Smaller 1.25"-diameter monitor wells were sampled with small-diameter Teflon bailers or with a bladder pump and Teflon sample line.

When practical, field measurements of temperature (°C), conductivity (µS/cm), pH, dissolved oxygen (mg/L) and oxidation reduction potential (mV using Ag/AgCl with 3.33 mol/L KCl as the reference electrode) were carried out using a Horiba U-22 ® water quality analyzer.

Sampling protocols were specific for different sets of analytes (see sampling sheet in Appendix C), and differed with regard to filtration, sample volume and container, the presence of headspace, and the use of gloves.

**Chemical Composition Analysis**

Samples for anions and cations were filtered in the field to 0.45 µm, and stored cold and dark until analysis. Anion (NO₃⁻, SO₄²⁻, Cl⁻, F⁻, Br⁻, PO₄³⁻, NO₂⁻) and cation (Ca²⁺, Mg²⁺, Na⁺, K⁺, Li⁺, NH₄⁺) concentrations were determined by ion chromatography using a Dionex DX-600. Total inorganic and organic carbon (TIC/TOC) was determined on unfiltered samples poisoned with mercuric chloride using a carbon analyzer (OI Analytical TOC Analyzer 1010). Dissolved inorganic carbon (DIC) concentrations were estimated in the water samples by employing the PHREEQC geochemical model (PARKHURST and APPELO, 2002) to achieve charge balance in the samples by adjusting and speciating DIC at the measured pH values. Dissolved organic carbon was also measured in a subset of samples as CO₂ gas pressure after acidification with orthophosphoric acid.

Sediment sulfur and carbon content was determined by elemental analysis by Actlabs (Ancaster, Ontario, Canada). Total C and S were determined on an ELTRA CS 2000 carbon sulfur analyzer. A weighed sample is mixed with iron chips and a tungsten accelerator and is then combusted in an oxygen atmosphere at 1370°C. The moisture and dust are removed and the CO₂ gas and SO₂...
gas are measured by a solid-state infrared detector. Sulphate S was determined by elemental analysis of the residue from roasting at 850° C. Reduced S was determined by difference. Carbonate C was determined by digestion of the sample in 2 N perchloric acid followed by coulometric titration. Graphitic C was determined by elemental analysis of the residue from roasting at 600° C. Organic C was determined by difference.

**Stable Isotope Mass Spectrometry**

Samples for nitrate N and O isotopic compositions are filtered in the field to 0.45 μm, and stored cold and dark until analysis. Anion and cation concentrations are determined by ion chromatography using a Dionex DX-600. The nitrogen and oxygen isotopic compositions (δ¹⁵N and δ¹⁸O) of nitrate in 26 groundwater samples from KCDI and MCD were measured at Lawrence Berkeley National Laboratory’s Center for Isotope Geochemistry using a version of the denitrifying bacteria procedure (CascioTTi et al., 2002) as described in Singleton et al. (Singleton et al., 2005). In addition, the nitrate from 34 samples were extracted by ion exchange procedure of (Silva et al., 2000) and analyzed for δ¹⁵N at the University of Waterloo. Analytical uncertainty is 0.3‰ for δ¹⁵N of nitrate and 0.5‰ for δ¹⁸O of nitrate.

Isotopic compositions of hydrogen and oxygen in water (δ²H and δ¹⁸O) were determined at LLNL using a VG Prism II® isotope ratio mass spectrometer, and are reported in per mil values relative to the Vienna Standard Mean Ocean Water (VSMOW). Isotopic composition of oxygen in water using the CO₂ equilibration method (Epstein and Mayeda, 1953), and have an analytical uncertainty of 0.1‰. Hydrogen isotope compositions were determined using the Zn reduction method (Coleman et al., 1982).

**Membrane Inlet Mass Spectrometry (Excess N₂)**

Previous studies have used gas chromatography and/or mass spectrometry to measure dissolved N₂ gas (BohIke and Denver, 1995; McMahon and BohlKE, 1996; Vogel et al., 1981; Wilson et al., 1990; Wilson et al., 1994). Both methods require extraction of a gas sample, which adds time and can limit precision. Membrane inlet mass spectrometry (MIMS) allows precise and fast determination of the concentrations of nitrogen, oxygen and argon dissolved in groundwater samples without a separate extraction step. This method has been used to document denitrification in estuarine and ocean settings (An et al., 2001; Kana et al., 1994), as well as for detection of volatile organic compounds in water (Ketola et al., 2002). The MIMS technique has also proven useful for determining excess N₂ from denitrification in groundwater systems (Beller et al., 2004).

Samples for N₂, O₂, Ar, CO₂ and CH₄ concentration were analyzed by MIMS. A water sample at atmospheric pressure is drawn into the MIMS through a thin silicone rubber tube inside a vacuum manifold. Dissolved gases readily permeate through the tubing into the analysis manifold, and are analyzed using a quadrupole mass spectrometer. Water vapor that permeates through the membrane is frozen in a dry ice cold trap before reaching the quadrupole. The gas abundances are calibrated using water equilibrated with air under known conditions of
temperature, altitude and humidity (typically 18 °C, 183 m, and 100% relative humidity). A small isobaric interference from CO₂ at mass 28 (N₂) is corrected based on calibration with CO₂-rich waters with known dissolved N₂, but is negligible for most samples. Typical sample size is 5 mL, and each analysis takes approximately 3 minutes. Dissolved oxygen, methane, carbon dioxide and argon content are measured at the same time as nitrogen. Samples are collected for MIMS analysis in 40 mL amber glass VOA vials, with no headspace, and kept cold during transport. Samples are analyzed within 24 hours to minimize the risk of gas loss or biological fractionation of gas in the sample container. The MIMS is field portable, and can be used on site when fieldwork requires extended time away from the laboratory, or when samples cannot be readily transported to the laboratory.

**Noble Gas Mass Spectrometry (³H/³He dating)**

Dissolved noble gas samples are collected in copper tubes, which are filled without bubbles and sealed with a cold weld in the field. Dissolved noble gas concentrations were measured at LLNL after gas extraction on a vacuum manifold and cryogenic separation of the noble gases. Concentrations of He, Ne, Ar and Xe were measured on a quadrupole mass spectrometer. Calculations of excess air and recharge temperature from Ne and Xe measurements are described in detail in Ekwurzel (2004), using an approach similar to that of Aeschbach-Hertig et al. (2000). The ratio of ³He to ⁴He was measured on a VG5400 mass spectrometer.

Tritium samples are collected in 1 L glass bottles. Tritium was determined by measuring ³He accumulation after vacuum degassing each sample and allowing three to four weeks accumulation time. After correcting for sources of ³He not related to ³H decay (Aeschbach-Hertig et al., 1999; Ekwurzel et al., 1994), the measurement of both tritium and its daughter product ³He allows calculation of the initial tritium present at the time of recharge, and apparent ages can be determined from the following relationship based on the production of tritogenic helium (³He₄₃):

\[
\text{Groundwater Apparent Age (years) = -17.8 x ln (1 + ³He₄₃/³H)}
\]

The reported groundwater age is the mean age of the mixed sample, and furthermore, is only the age of the portion of the water that contains measurable tritium. Average analytical error for the age determinations is ±1 year, and samples with ³H that is too low for accurate age determination (<1 pCi/L) are reported as >50 years. Loss of ³He from groundwater is not likely in this setting given the relatively short residence times, lack of water table fluctuations, and high infiltration rates from irrigation. Groundwater age dating has been applied in several studies of basin-wide flow and transport (Ekwurzel et al., 1994; Poreda et al., 1988; Schlosser et al., 1988; Solomon et al., 1992). Mean ³H-³He apparent ages are determined for water produced from 20 KCD monitor wells at depths of 6 m to 54 m, and from 14 sites at MCD. The apparent ages give a measure of the time elapsed since water entered the saturated zone, but only of tritium-containing portion of the groundwater sample. Apparent ages therefore give the mean residence time of the fraction of recently recharged water in a sample, and are especially useful for comparing relative ages of water from different locations at each site. The absolute mean age of
groundwater may be obscured by mixing along flow paths due to heterogeneity in the sediments (WEISSMANN et al., 2002b).

Quantitative Real-Time Polymerase Chain Reaction (rt-qPCR)

We have developed a simple bioassay to quantify populations of denitrifying bacteria in moderate amounts of aquifer material (on the order for a few grams of sediment or filtrate). The method detects the presence of bacterial genes that encode nitrite reductase, a central enzyme involved in denitrification. The assay is not species-specific, but rather a functional test for the presence of bacterial populations capable of nitrite reduction. Nitrite reduction is considered to be the "committed" step in denitrification, and bacteria capable of nitrite reduction are generally also capable of nitric and nitrous oxide reduction to nitrogen gas (TIEDJE, 1988). Currently, the assay provides valuable information on the distribution of denitrifying bacteria populations in aquifers. Ultimately, data on denitrifier populations (i.e., biomass) can be used in combination with specific (i.e., biomass-normalized) denitrification rate constants to determine subsurface denitrification rates.

Real-time, quantitative Polymerase Chain Reaction (rt-qPCR) analysis (Gibson et al., 1996; Heid et al., 1996; Holland et al., 1991), specifically the 5'-nuclease or TaqMan® assay, was chosen for this assay because it offers many advantages over traditional methods used to detect specific bacterial populations in environmental samples, such as DNA: DNA hybridization (Beller et al. 2002). Although most real-time PCR applications to date have involved the detection and quantification of pathogenic bacteria in food or animal tissue, the technique has recently been used to quantify specific bacteria in environmental samples (Hristova et al., 2001; Suzuki et al., 2000; Takai and Horikoshi, 2000).

Real-time qPCR is a rapid, sensitive, and highly specific method. The rt-qPCR assay developed targets two variants of the nitrite reductase gene: nirS (Fe-containing nitrite reductase) and nirK (Cu-containing nitrite reductase). Homologous gene sequences were used to develop a primer/probe set that encompasses functional nir genes of known denitrifying soil bacteria (including heterotrophic and autotrophic species) and that does not result in false positive detection of genes that are not associated with denitrification. The rt-qPCR primers and probes were designed based on multiple alignments of 14 nirS and 20 nirK gene sequences available in GenBank. During development of the assay, the first nitrite reductase gene (nirS) reported in an autotrophic denitrifying bacterium (T. denitrificans) was sequenced and amplified, and demonstrated to have high homology to nirS in a phylogenetically diverse set of heterotrophic denitrifying bacteria.

Real-time PCR was also be used to quantify total eubacterial population, based on detection of the sequence encoding the eubacterial 16S rRNA subunit, which is specific for bacteria.

Wastewater Co-Contaminants
A number of co-contaminants expected to occur on a dairy farm from the dairy operation proper or from associated field crop production were determined using GC-MS or LC-MS. Co-contaminants targeted included herbicides, pesticides, VOCs, fecal sterols, caffeine and nonylphenol. The analysis of these compounds and a discussion of their distribution at the dairy sites is in Moran et al. (2006).

DATA

Chemical, isotopic, dissolved gas, and groundwater age data for the KCD1 and MCD sites are discussed in Appendix A and Appendix B, and are tabulated in Table 1 of Appendix A and Table 1 of Appendix B. Chemical composition, stable isotope, and groundwater age data for KCD2, KCD3 and SCD2 are tabulated in Table 1 of the main report. In addition, membrane inlet mass spectrometry data for KCD2 is presented graphically in Figures 8 and 9. Neither Appendix A nor Appendix B contains sediment C and S data or bacterial population data, which are discussed below.

Sediment Data

In zones sampled for groundwater at the KCD1 site, sediment texture as determined from well logging, CPT and laser diffraction particle size analysis ranges from sand to clayey silt (with trace to >95% fines). Sedimentary carbonate C is extremely low (generally < 0.003 wt %); organic C is low but generally detectable (0.05-0.10 wt %), although occasional beds have 0.1-1.3% organic C; sulfate S ranges from nondetectable (<0.017) to 0.08 wt%; and reduced S is only detectable in a few wells (<0.01 to 0.15 wt %). For organic C and total S, no strong vertical gradients exist, and no significant difference exists between sediment in the oxic groundwater column, sediment in the anoxic water column, and sediment at the interface. Sediment data are summarized in Table 2, and represented graphically in Figures 5 and 6.

Bacterial Population Data

In this study we use the abundance of the nir gene, as determined by rt-qPCR, to map the vertical distribution of denitrifying bacterial populations in the saturated zone. We use the abundance of the eubacterial 16S rRNA gene, as determined by rt-PCR, to map the vertical distribution of total eubacteria in the subsurface. The analyses were performed on soil returned from four locations at the KCD1 dairy during the course of the DP sampling survey in August 2003. Soil samples were placed on ice upon recovery, and subsequently stored frozen until analysis. Total nir data are reported as gene copies per 5 g of sediment, and comprise both nirS and nirK assay results. Total eubacteria data are reported as cells per 5 g sediment. The data are tabulated in Table 3 and in Figure 7.

Relative abundances of nirS, nirK and eubacteria are consistent with previous studies in non-groundwater systems: nirS and nirK gene copies typically constitute ~5% and ~0.1% of total bacteria, respectively. Total nir abundance varies by almost four orders of magnitude and is not
well-correlated with total eubacteria ($R^2 \sim 0.19$ for 5 locations with multiple depths). Peak populations occur either at or below the redoxicline where strong vertical gradients exist in ORP, nitrate and excess nitrogen. Where nir abundance is high, total nir gene copies tend to constitute a larger fraction of total bacteria (up to 18%).

The presence of high and localized nir populations near the interface between oxic high-nitrate groundwater and suboxic low-nitrate groundwater indicates active denitrification is occurring near that interface.

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**Figure 5. KCD1 Well Cluster 1 sediment composition, texture & groundwater oxidation state**

Sediment composition and texture and groundwater oxidation state at KCD1 Site 1. From left to right are shown profiles of sediment organic carbon and total sulfur, sediment iron oxidation state as indicated by sediment color, a continuous core log of sediment texture (yellow sands, brown silty sands, and red silts), the location of the perched and deep aquifer along with groundwater oxidation state (as determined by dissolved oxygen and oxidation-reduction potential probes and the presence of hydrogen sulfide gas).
Figure 6. KCD1 depth profiles of sediment and water properties.

KCD1 soil behavior type, sediment organic carbon and total sulfur, $^3$H-$^3$He groundwater age and fraction pre-modern water, field oxidation-reduction potential (ORP) and dissolved chloride content. The dashed line indicates the transition from nitrate to dissolved nitrogen from denitrification.
Figure 7. KCD1 depth profiles of nitrogen speciation and bacterial populations.
KCD1 depth profiles of soil behavior type, nitrate, excess nitrogen, total nir gene copies, and total eubacteria. The colored fields indicated water oxidation state based on field ORP.
RESULTS AND DISCUSSION

*Saturated-Zone Denitrification at KCD1 and MCD*

Appendix A is a manuscript prepared for submittal to a peer-review journal. The manuscript addresses evidence for saturated-zone denitrification in groundwaters impacted by dairy operations. The manuscript abstract follows.

Results from field studies at two central California dairies (KCD1 and MCD) demonstrate the prevalence of saturated-zone denitrification in shallow groundwater with $^3$H/$^3$He apparent ages of 30 years or younger. Confined animal feeding operations are suspected to be major contributors of nitrate to groundwater but saturated zone denitrification could effectively mitigate their impact to groundwater quality. Denitrification is identified and quantified using stable isotope compositions of nitrate coupled with measurements of excess N$_2$ and residual NO$_3^-$.

Nitrate in dairy groundwater from this study has $\delta^{15}$N values (4.3–61‰), and $\delta^{18}$O values (-4.5–24.5‰) that plot with a $\delta^{18}$O/$\delta^{15}$N slope of 0.5, consistent with denitrification. Dissolved gas compositions, determined by noble gas mass spectrometry and membrane inlet mass spectrometry, are combined to document denitrification and to determine recharge temperature and excess air content. Dissolved N$_2$ is found at concentrations well above those expected for equilibrium with air or incorporation of excess air, consistent with reduction of nitrate to N$_2$.

Fractionation factors for oxygen and nitrogen isotopes appear to be smaller ($\varepsilon_N \approx -10\%$; $\varepsilon_O \approx -5\%$) at a location where denitrification is found in a laterally extensive anoxic zone 5 m below the water table, compared with a site where denitrification occurs near the water table and is strongly influenced by localized lagoon seepage ($\varepsilon_N \approx -50\%$; $\varepsilon_O \approx -25\%$).

*Spatial Distribution of Saturated-Zone Denitrification at KCD1*

At the KCD1 site, multiple lines of evidence indicate saturated-zone denitrification. These include the presence of excess nitrogen from denitrification at depth, the correlation between nitrate-$\delta^{15}$N and $\delta^{18}$O (which has a slope characteristic of denitrification), and the presence of denitrifying bacteria (which occur at above background levels only where excess nitrogen is present). The lateral extent of denitrification at the site and the excess nitrogen and isotopic evidence for denitrification at the site are discussed in Appendix B. Bacterial distributions give valuable evidence for the localization of denitrification.

Denitrifying bacteria populations at the KCD1 site have a high dynamic range, with peak populations occurring at the oxic-anoxic interface in the perched aquifer where strong gradients in oxidation-reduction potential, nitrate and excess nitrogen exist. Denitrifying bacteria populations are not well correlated with total bacteria ($R^2 \approx 0.19$ for 5 locations with multiple depths). The relative population abundances of *Nir* gene copies, however, are consistent with previous studies in non-groundwater systems: *nirS* and *nirK* gene copies typically constitute ~5% and ~0.1% of total bacteria.
The depth of oxic-anoxic interface is remarkably constant at 37-41 feet below ground surface (Figure 7). This transition is not strongly correlated with lithology or sediment composition (organic-C or total-S content), although it generally occurs in sand. At the irrigated field monitoring sites, the redox interface corresponds to the interface between shallower “young” groundwater (having young apparent $^{3}H$-$^{3}$He ages and low mixing ratios of pre-1955 water) and deeper “old” groundwater (with higher fractions of pre-modern water) (Figure 8). The depth of the zone corresponds to the top of several agricultural production pump screens in the area, suggesting that pumping may be a factor.
Saturated-Zone Denitrification at the Northern Dairy Sites

Both of the northern San Joaquin Valley dairy sites (MCD and SCD) are a part of the northern San Joaquin Valley monitoring network described in Harter et al. (2002). Chemical data from these sites have been used to calibrate and validate regional models for nitrogen loading to the shallow groundwater system (Van der Schans, 2001). The wells sampled are all shallow piezometers that draw first-encounter water, with the exception of one deeper domestic supply well (W-98, Table 1 of Appendix A). A significant finding of the current study is that evidence for saturated-zone denitrification at MCD and SCD only exists in first-encounter wells that are predicted by other criteria (groundwater gradient, the presence of ammonia, total dissolved solids, etc) to be impacted by recharge from lagoons or corrals, i.e. from the dairy operation proper. Wells so impacted include W02, W03, W16, W17, V01, and V21 on the MCD site (Table 1 of Appendix A), and Y03 and Y10 on the SCD site (Table 1). No evidence for denitrification exists in first-encounter wells that are impacted only by wastewater irrigation of either field crops (MCD) or of orchards (SCD). This finding is significant in two respects:

- The UC-Davis nitrate loading model for the region is in agreement with available spatial and time-series groundwater nitrate concentration data. The model does not explicitly consider denitrification of nitrogen fluxes from lagoons and corrals. The absence of evidence for denitrification in first encounter groundwater impacted by wastewater irrigation validates the model assumption that denitrification is not occurring and strengthens confidence in the model as a predictive tool.

- The deep domestic well W-98 is predicted by the UC-Davis model to have approximately 50 mg/L nitrate (T. Harter, personal communication). Groundwater from this well actually has very low nitrate (0.4 mg/L), but does have 45 mg/L nitrate-equivalent of excess N2 indicating that the mass fluxes and transport in the model are accurate. The mean 3He/3H groundwater age also matches well with model travel time predictions. The good agreement between predicted nitrate and excess nitrogen in W-98 is consistent with a groundwater impacted by wastewater irrigation in which denitrification is occurring at some depth below the water table, as is the case at KCD1 in Kings County.

- The association of denitrification with groundwater impacted by manure lagoon seepage is consistent with the findings from the KCD1 study (see Appendix B)

To the extent that saturated-zone denitrification is significant and is associated with nitrogen loading from wastewater irrigation from dairy operations (as has been shown on one site, and indicated on another), the process needs to considered when assessing total impact of dairy operations on the groundwater resource. The most effective way to characterize saturated-zone denitrification is the installation of multi-level monitor wells in conjunction with the determination of nitrate stable isotope composition and excess nitrogen content.
The Impact of Dairy Manure Lagoons on Groundwater Quality

Appendix B is a manuscript prepared for submission to a peer-review journal. The manuscript addresses the impact of dairy manure lagoon seepage on groundwater quality, and discusses a new tracer for manure lagoon seepage. The manuscript abstract follows.

Dairy facilities and similar confined animal operation settings pose a significant nitrate contamination threat to groundwater via oxidation of animal wastes and subsequent transport through the subsurface. While nitrate contamination resulting from application of animal manure as fertilizer to fields is well recognized, the impact of manure lagoon leakage on groundwater quality is less well characterized. For this study, a dairy facility located in the southern San Joaquin Valley of California (KCD1) has been instrumented with monitoring wells as part of a two-year multidisciplinary study to evaluate nitrate loading and denitrification associated with facility operations. Among the multiple types of data collected from the site, groundwater and surface water samples have been analyzed for major cations, anions, pH, oxidation-reduction potential, dissolved organic carbon, and selected dissolved gases (CO2, CH4, N2, Ar, Ne). Modeling of geochemical processes occurring within the dairy site manure lagoons suggests substantial off-gassing of CO2 and CH4 in response to mineralization of organic matter. Evidence for gas ebullition is evident in low Ar and Ne concentrations in lagoon waters and in groundwaters downgradient of the lagoon, presumably as a result of gas "stripping". Shallow groundwaters with Ar and Ne contents less than saturation with respect to atmosphere are extremely rare, making the fractionated dissolved gas signature an effective tracer for lagoon water in underlying shallow groundwater. Preliminary evidence suggests that lagoon water rapidly re-equilibrates with the atmosphere during furrow irrigation, allowing this tracer to also distinguish between seepage and irrigation as the source of lagoon water in underlying groundwater. Together with ion exchange and mineral equilibration reactions, identification of lagoon seepage helps to constrain key attributes of the local groundwater chemistry, including input and cycling of nitrogen, across the site.

A New Tracer for Manure Lagoon Seepage

The manuscript in Appendix B uses only data collected from the KCD1 site. We also see evidence for gas stripping in lagoon waters from the KCD2 site (Figure 9). To further test the hypothesis that gas stripping in biologically active manure lagoons, we sampled manure lagoon water from several locations at KCD2 site. At this site, manure-laden water flows from free stall flush lanes to a settling lagoon (Lagoon 1) through an intake near the bottom of the lagoon to a larger holding lagoon (Lagoon 2) to a distribution standpipe to furrows in nearby fields. Samples were collected from the surface of Lagoon 1 near the outtake from the flush lanes, from the outlet of Lagoon 1 into Lagoon 2, from the surface of Lagoon 2 near the intake to the field distribution system, from a distribution standpipe, and from a field furrow about halfway down the length of the furrow. At the time of sample collection in April 2005, water in the distribution standpipe and in the field furrows was entirely from the manure lagoon, and was not mixed with well water or canal water. The results are shown in Figure 10.
Atmospheric Gas in Dairy Lagoon Water

As discussed in Appendix B, biological activity in the lagoon consumes oxygen and strips atmospheric gases from the lagoon water through ebullition of carbon dioxide and methane. This effect of this activity is evident in the absence of detectable oxygen in any of the lagoon samples, and in lagoon water argon partial pressures that are close to or far below saturation argon partial pressures. For non-reactive gases such as argon, the “gas-stripping” effect is most evident in the sample drawn from the outlet of Lagoon 1 into Lagoon 2, which presumably represents water from near the bottom of Lagoon 1. This sample has extremely low argon, and may be representative of lagoon seepage through the bottom or sides of the lagoon. Atmospheric re-equilibration does not take place until the water is delivered to the field – the water sample drawn from the distribution standpipe has no detectable oxygen, while surface water from halfway down a furrow is at about 40% saturation. We suspect that percolation through the soil zone and through an oxic vadose zone, which is characterized by incorporation of excess air, will result in complete re-equilibration or over-equilibration with soil gases.
Dissolved gas samples from a number of manure lagoons on five dairy sites (KCD1, KCD2, MCD, and SCD) are characterized in general by deficiency in reactive and non-reactive atmospheric gases, and in detail by a wide range in non-reactive gas pressures from near equilibrium to far below equilibrium. The only other mechanism known to produce such signals is methane production either in marine sediments or in the deep subsurface in association with natural gas formation (see references in Appendix B). Currently the presence of an air “deficit” (i.e. atmospheric noble gases below saturation values) in shallow groundwater samples associated with dairy operations can be considered as indicative of the presence of a manure lagoon seepage component. To determine the mixing ratio of lagoon seepage with other water sources, however, will require a more quantitative understanding on the dissolved gas content in manure lagoons and manure lagoon seepage.

Source, Fate and Transport of Dairy Nitrate at KCD1

Harter et al. (2002) have demonstrated that dairy operations in the northern San Joaquin Valley strongly impact groundwater quality, resulting in first-encounter water that is high in salinity and inorganic nitrogen. On the KCD1 site in the southern San Joaquin Valley, a number of observations indicate that the dairy operation and associated wastewater irrigation are the source of high nitrate in first encounter groundwaters at the site:
• The isotopic composition of nitrate-N and -O is consistent with a manure or septic nitrogen source (see Appendix A).

• The young age of the first encounter waters (Figure 6 and 8), which we have accurately simulated using an irrigation recharge model (see groundwater transport discussion below) are inconsistent with transport from offsite locations.

• Nitrate co-contaminants can be traced to a specific application event on the site (see Moran, 2006). In a subset of wells on the site, norflurazon and its degradation product, desmethylnorflurazon, were detected. Norflurazon was applied to a corn field in excess of the intended amount approximately two years prior to sampling. The well closest to the field contains norflurazon; a more distal well contains the degradation product, desmethylnorflurazon.

The unconfined aquifer at KCDI is strongly stratified with respect to electron donor concentration (oxygen and nitrate), redox state (ORP), and excess nitrogen (Figures 5 and 6). The transition zone is sharp: nitrate levels can drop from significantly above maximum contaminant levels to nondetectable over a depth range of five feet. Our data indicate that the water immediately below the transition zone also has a significant wastewater component:

• Low-nitrate groundwaters nitrate isotopic compositions that are consistent with denitrification of manure or septic source nitrate.

• Some low-nitrate waters have below-saturation dissolved gas pressures that indicate a component of manure lagoon seepage (see Appendix B and discussion below.)

• Groundwater transport modeling (see discussion below) that assumes recharge dominated by wastewater irrigation accurately simulates the mean age and pre-modern mixing ratios for low-nitrate groundwaters below the transition zone.

The strong spatial association of high denitrifier bacterial populations (Figure 6) with the transition zone is consistent with active denitrification occurring in this zone and being at least one source of denitrified groundwater seen below the zone. We cannot currently convert nir gene copy populations into denitrification rates, and so cannot estimate what fraction of denitrification occurs in the transition zone and what fraction occurs upgradient (proximal to a manure lagoon seepage plume, for example). What is clear, however, is that active denitrification is currently occurring on the dairy site in localized subsurface zones.

The relationship of the dairy operation (including wastewater irrigation and manure lagoon seepage) to nitrate mitigation through the establishment of redox stratification and the enhancement of saturated-zone denitrification is more complex. Any model of the evolution of redox stratification and denitrification must first provide an electron donor and then produce a sharp transition zone (~5 feet in vertical extent) at a remarkably uniform depth across the site (~35-40 feet bgs). A number of hypotheses can be put forward:

• Lateral transport of manure lagoon seepage.
- Field irrigation with dairy wastewater (assuming vertical percolation through a homogeneous soil column that contains a solid-phase electron donor).
- Agricultural pumping and nitrogen loading from dairy operations (assuming strong lateral transport of nitrate through a heterogeneous aquifer).

**The Impact of Lagoon Seepage on Groundwater Quality**

The first hypothesis is discussed in McNab et al. (Appendix B and Figure 11).

![Diagram](image)

**Figure 11. Simulation of transport of lagoon seepage through groundwater.** Simulation of the influence of seepage from a dairy wastewater lagoon on groundwater chemistry. See Appendix B for details on modeling.
McNab et al. assume that oxidation of organic carbon derived from manure creates the reducing conditions and provides the electron donor necessary for denitrification. While manure lagoon seepage is associated with excess nitrogen and does appear to drive denitrification locally, reactive transport modeling of lagoon seepage shows that the modeled zone of denitrification does not extend far from the lagoon, and that the modeled zone of low redox potential (where pE < 0) is localized (Figure 11). These model results are driven by the relative magnitudes of lagoon seepage and wastewater irrigation percolation rates, and are consistent with dissolved gas evidence indicating that lagoon seepage is not a major component in most site groundwaters. We conclude that manure lagoon seepage is not the cause of the laterally extensive reduced zone observed at the KCD1 site.

**The Impact of Dairy Wastewater Irrigation on Groundwater Quality**

Reactive transport modeling of vertical flow under an irrigated field indicates that vertical redox stratification can be created without a lagoon influence when dairy wastewater percolates through a soil column containing organic carbon in low permeability micro-environments. Attempts to simulate the development of redox stratification in the absence of a sedimentary electron donor were not successful.

We employed a reactive modeling approach using PHREEQC that addresses multispecies solute transport, soil-water reactions (mineral phase equilibria and ion exchange), and reaction kinetics for redox reactions involving nitrogen species as means for identifying the potential roles of different electron donors in the denitrification process at the site. The model parameters are shown below:

**Parameters**
- 10-m column
  - 10 volume elements (mobile pore water)
  - 10 volume elements (immobile pore water)
- Initial sediment composition:
  - 25% Quartz
  - 15% Na-montmorillonite (ion exchanger)
  - 15% K-mica (“C” model; no K-mica = “X” model)
  - 1% Goethite (HFO surface)
  - 0.02 mol/kg organic carbon

**Step 1: Set up initial conditions**
- Flush column with 300 pore volumes:
  - 1 mM NaCl
  - mM KCl
- After flushing
  - Equilibrium with CO₂(g) and O₂(g), calcite, and dolomite
  - Undersaturated with gypsum

**Step 2: Simulate irrigation**
- Flush column with 2 pore volumes with a mixture of agricultural well water and lagoon water (~0.02 M NH4⁺; ~0.01 M K⁺) – agricultural well water.
- Allow equilibration with calcite, ion exchanger, and HFO surface.

Figure 12. Simulation of dairy wastewater percolation through sediment. Model results from simulation of vertical percolation of dairy wastewater through a sediment column containing organic carbon in low-permeability environments. See text for explanation.
Results from the reactive transport simulations results generally match most major cation and anion distributions with depth (Figure 12 and Figure 13). Moreover, the quantities of organic carbon required to produce a redox front (via diffusion-limited transport through low-permeability lenses) are consistent with measurements from soil samples (which are low). These results do not depend on any lagoon influence. Reactive transport modeling of vertical flow under the irrigated field demonstrates that general geochemistry in wells distal from the manure lagoons can be explained without postulating a lagoon influence, if the aquifer has reducing capacity.

\begin{figure}[h]
\centering
\includegraphics[width=\textwidth]{figure13.png}
\caption{Simulation of denitrification associated with dairy wastewater percolation. Saturated-zone denitrification in a simulation of vertical percolation of dairy wastewater through a sediment column containing organic carbon in low-permeability environments. See text for explanation.}
\end{figure}

A number of lines of evidence exist that indicate that reducing groundwater conditions are common in the region surrounding the KCD1 site. At a number of NAWQA sites in the region that are not believed to be impacted by dairy wastewater, nitrate in deeper waters is nondetectable and iron and manganese concentrations are high, an association consistent with suboxic or anoxic conditions (Burow et al., 1998a; Burow et al., 1998b). The most convincing evidence comes from the deep well at the KCD1 site (KCD1-1D, Table 1 in Appendix A). Groundwater in the lower aquifer sampled by this well is tritium dead with a mean groundwater age in excess of 50 years. Radiogenic \textsuperscript{3}He content indicates an age on the order of 100 years or more. Neither nitrate nor excess nitrogen is present, indicating that source waters were low in inorganic nitrogen species. This groundwater has extremely low chloride and has isotopically lighter water than water sampled in the perched aquifer. Finally, this groundwater is reduced as indicated by both field ORP and DO measurements, and measurements of volatile sulfide compounds in the water. These observations are consistent with recharge by source waters un-
impacted by agriculture and the occurrence of naturally reducing conditions along the flow path. The electron donor driving the evolution of the natural reducing system is unclear. The water is low in TOC (0.8 mg/L). Sediment organic C and reduced S contents are generally low (< 0.1 wt %), but are sufficient to produce reducing conditions, particularly since sediments with organic carbon contents of over 1 wt% have been characterized (Figures 5 and 6). Reducing conditions may have also been created during recharge (in the hyporheic zone during riverbank infiltration).

The existence of regionally reducing conditions is also evident in the redox state of sedimentary iron in site sediments. Above approximately 60’ bgs, sediment core is stained with orange, red and brown ferric iron oxides; below 60’, this stain is not present (Figures 5 and 8). The existence of a denitrification zone approximately 20-25’ above the iron reduction zone is consistent with the energetics of these reactions.

Given the presence of reducing conditions within the aquifer, one-dimensional transport through homogeneous media can drive the development of redox stratification and saturated-zone denitrification within the shallow aquifer. This process, however, can only reproduce the sharpness and uniform depth of the observed groundwater redox stratification 1) if a layer of laterally extensive reducing sediment exists at the groundwater redox boundary or 2) if a sharp transition in sediment reducing capacity exists at or near the depth of the water redox transition. Neither of these conditions is observed at the KCD1 site. The redox boundary is not correlated with sediment texture, nor do any gradients exist in sedimentary organic C, total S, or reduced S that correlate with the depth of the redox boundary.

The Impact of Pumping and Wastewater Irrigation on Groundwater Quality

A number of processes that may contribute to strong vertical stratification of groundwater flow and chemistry are not adequately simulated in a one-dimensional homogeneous model. To explore the effect of aquifer heterogeneity and lateral transport on groundwater flow and transport at the KCD1 site, we used the numerical flow and transport model NUFT to simultaneously simulate three-dimensional variably-saturated groundwater flow processes including canal recharge, agricultural pumping, and irrigation (CARLE et al., 2005). Heterogeneity of sandy, silty, and clayey zones in the system was characterized stochastically by applying transition probability geostatistics to data from 12 CPT logs that vertically transect the perched aquifer. In the first iteration of this model, nitrate in surface irrigation was simulated as a tracer rather than as a reactive species.

Groundwater Hydrology. In the distal reaches of the Kings River within the Tulare Lake Basin, groundwater is extracted from both a perched zone (less than ~ 25 m deep) and a deep zone. Before the 1950’s, water levels were nearly equal in both zones (DWR data). Overdraft in the deep zone has caused water level declines of over 100 feet (30 m). Perched zone water level elevations, where they exist, persist well above the deep zone, as evident from DWR water level elevation maps for 2001-2002. The Kings River, unlined ditches and canals, and irrigation appear to provide recharge to sustain the perched aquifer. Crop irrigation uses canal diversions and both shallow and deep groundwater.
At and near the KCD1 site, groundwater level elevations in different wells screened in the perched aquifer are remarkably similar over time and correlate to canal diversions. This suggests canal leakage and irrigation from canal diversions provides substantial recharge to the perched aquifer. Leakage from the canal is estimated at 10% by the irrigation district.

Several dairies are located within the area of the perched aquifer. KCD1 is located about one mile east of the canal. The dairy grows much of its own feed—corn and alfalfa. The crops are irrigated primarily with water pumped from the shallow aquifer. Crops are fertilized largely by mixing in effluent from the dairy operation that is collected in a lagoon. The lagoon water and other fertilizers provide sources of nitrate that appear to impact upper portions of the perched aquifer, but not lower portions of the perched aquifer or the deep aquifer. Other nearby farms also irrigate with canal diversions or groundwater pumped from the deep aquifer. Thus, overdraft from the deep aquifer helps, in part, to sustain the perched aquifer.

The modeling approach was designed to include consideration of the major factors and processes affecting groundwater flow, nitrate transport, and groundwater age dating:

- **Heterogeneity**: Use hydrofacies-based geostatistics.
- **Varibly Saturated Flow**: Couple vadose zone and saturated zone using LLNL’s NUFT code.
- **Boundary Head Conditions**: Use time-series DWR water levels in perched and deep zone.
- **Perched and Deep Zone**: Use modeling to determine leakage that maintains perched condition.
- **Canal Leakage and Irrigation**: Distinguish different sources with different tracer simulations.
- **Tritium/Helium-3 Age Dating**: Add decay to tracer simulations, simulate apparent age estimate.
- **Groundwater Mixing**: Keep track of proportions of groundwater from different sources.

**Heterogeneity.** Based on our interpretation of lithologic and CPT logs, we defined three hydrofacies: “sand”, “silt”, and “clayey” categories. We quantified vertical and horizontal spatial variability with a transition probability matrix using the CPT data categorized as hydrofacies. The solid lines in the probability matrices (Figure 14) represent 1-D Markov chain models used to develop stochastic simulations of hydrofacies architecture at the site.

The hydraulic properties of the hydrofacies categories were estimated from a combination of pump test analysis, soil core measurements, and model calibration.

<table>
<thead>
<tr>
<th>HYDROFACIES</th>
<th>K (m/d)</th>
<th>POROSITY</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sand</td>
<td>30</td>
<td>0.40</td>
</tr>
<tr>
<td>Silt</td>
<td>0.24</td>
<td>0.43</td>
</tr>
<tr>
<td>Clayey</td>
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<td>0.45</td>
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<tr>
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<td>Canal (sandy)</td>
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A Van Genuchten model was used to predict unsaturated hydraulic conductivity and capillary pressure. A continuous 1-m thick aquitard layer at 46-47 m elevation sustains the perched aquifer conditions. This aquitard layer correlates to a distinctive clay layer identified in our initial characterization lithologic log.

Figure 14. Geostatistical representation of the subsurface at KCD1. Transition probability matrices and geostatistical representation of hydrofacies architecture for the KCD1 site. See text for explanation.
Flow and transport simulation (Figure 15 and 16). We used LLNL’s NUFT code to simulate variably saturated flow according to the Richards equation (Figure 15). The simulation runs from late 1949 through 2001. Initial conditions are equilibrated to local head measurements and rainfall recharge of 1 cm/year. For boundary conditions, x-direction and bottom boundaries were conditioned to observed piezometric heads. A fully saturated initial condition is applied to the canal when canal diversions occur (between early April and early October). In the simulation, the six site production wells were pumped during irrigation season a rate greater and proportionate to crop evapotranspiration (ET). Recharge from irrigation was distributed proportionately to crop (ET), with about 25 cm/yr within the dairy crop fields and 10 cm/yr in surrounding areas.

In the simulation, piezometric head in the perched aquifer remains relatively steady, although in fall 1992 (during a drought) head is noticeably lower. However, head in the deep aquifer drops considerably since the 1950s, to the extent that the top of the deep zone begins to desaturate in the 1960s. In effect, the aquifer system near the dairy field site now functions like two unconfined aquifers stacked on top of each other. This is consistent with the observed separation of the DWR water levels between shallow and deep wells in the 1960s.

We used LLNL’s NUFT code to simulate tracer transport from different recharge sources (Figure 16). The three primary recharge sources near the dairy site are canal, dairy crop irrigation, and irrigation from surrounding areas. The transport simulation results indicate that nitrate entering the saturated zone from dairy crop irrigation is contained in the upper parts of the aquifer. Nitrate containment occurs within the high permeability sand-dominated perched aquifer because the dairy irrigation wells screened in the perched aquifer effectively capture nearly all recharge from dairy crop irrigation. The dairy irrigation wells pump groundwater at rates far higher than the recharge from dairy crop irrigation. The dairy irrigation wells also extract groundwater originating from irrigation of surrounding areas, canal leakage, and older groundwater.
Figure 15. Simulation of groundwater flow at KCD1.
Model validation. To validate the groundwater flow and transport model, we used the model to simulate \(^3\text{H}-\text{He}\) groundwater ages in the aquifer and compared the results of the simulation to measured values. Groundwater ages determined using the \(^3\text{H}-\text{He}\) method are apparent age estimates of the average age of a mixed groundwater. Such ages are affected by mixing of groundwater through diffusion and dispersion, transient flow, and sampling, and by the decay of atmospheric tritium activities since 1963 bomb pulse.

To simulate apparent age of groundwater, we used NUFT to tag all surface recharge sources. We then simulated apparent groundwater age for two scenarios: (1) for an “ideal source” that assumes constant tritium concentration over time and (2) for a “bomb source” where tritium concentration varies as measured. The simulated tritium/helium-3 ratios are backed out of the differences in simulated concentration.
These simulations of apparent age indicate variation in concentration of bomb source tritium will lead to some underestimation of groundwater age, particularly for older modern groundwater.

Figure 17. Simulation of apparent groundwater age at KCD1.
The simulation of apparent age show excellent agreement for the southern Site 1 and Site 4 wells south of the dairy operation (Figure 18). At these well cluster locations, simulated ages are less than measured tritium/helium-3 ages in shallow groundwater at these sites because the simulations assumed that $^{3}$He begins accumulating at the ground surface and not the water table. Current modeling efforts address this effort and produce better agreement for shallow groundwater. At Site 2 to the southeast of the dairy operation, measured groundwater ages are younger than simulated ages. This difference may indicate the absence of a shallow clayey zone at this location. These simulations of apparent age indicate variation in concentration of bomb source tritium will lead to some underestimation of groundwater age, particularly for older modern groundwater.

**Conclusions.** Coupling flow and transport simulations with groundwater age data and geostatistical simulations of hydraulic properties provides invaluable insights. Heterogeneity plays a large role in creating the perched aquifer and in causing vertical compartmentalization of flow patterns. The hydrofacies architecture consists of laterally continuous sand with interbeds of silt and clayey zones. Maintaining head and saturation in perched zone requires a continuous ~3 foot-thick clay layer at ~ 85 feet bgs. Flow simulation desaturates upper portions of the deep zone below the confining layer, and is consistent with observation of de-saturated zone below ~ 80 feet bgs.
The perched zone draws older water and recharge mostly from irrigation and less so from canal leakage. The dairy site pumps more groundwater from the perched aquifer than is recharged by crop irrigation, and thus physically contains lateral and vertical migration of nitrate contamination. High nitrate irrigation water penetrates to depths below the sharp redox gradient. Without denitrification, nitrate concentrations would be greater below the redox gradient, as is consistent with the presence of excess nitrogen in this zone.

The NUFT model presented here does not simulate transport of reactive constituents such as oxygen, nitrate, sulfate and organic carbon, and does not directly address the sharpness and uniform depth of the redox gradient in the shallow groundwater system. The strong vertical compartmentalization of the groundwater flow created by agricultural pumping and the location of the redox gradient close to the top of the production well screens, however, suggest that agricultural pumping and lateral groundwater flow may be important controls on the development of redox stratification in the shallow aquifer.

The Development of Reducing Conditions in Dairy Site Groundwaters

At three sites in this study (KCD, SCD, and MCD), dairy operations have been demonstrated to impact groundwater quality. At all three sites, nitrogen mitigation (either through denitrification or denitrification) has been demonstrated in groundwater impacted by manure lagoon seepage, a finding consistent with geochemical reactive transport modeling. At two of the sites (KCD and MCD), denitrification has also been demonstrated to occur in deeper waters impacted by irrigation with dairy wastewater. For denitrification to occur in the saturated zone, dissolved oxygen must be absent or present in very low concentrations. A key question, then, in assessing the ability of a groundwater to assimilate nitrate loading is what mechanism drives the development of reducing conditions necessary for denitrification to occur.

At the best studied site, KCD1, evidence exists for both natural and anthropogenic influence on the development of suboxic and anoxic groundwater. The deep aquifer at the KCD1 site consists of old water un-impacted by agricultural inputs. The water is tritium-dead and has a radiogenic $^4$He age of approximately 100 years. In addition to having a mean age that pre-dates the intensification of agricultural activities, especially with regards to fertilizer usage and manure production, the deep aquifer groundwater has a chemical composition that indicates the absence of significant agricultural input. Salinity, dissolved organic C, nitrate and excess nitrogen are all low. This water is also anoxic, with nondetectable dissolved oxygen, detectable hydrogen sulfide, and low ORP. The electron donor responsible for reducing conditions is not known. Groundwater DOC is low, as is sediment solid-phase total S and organic C. Reduced sediment phases, however, are sufficient to create reducing conditions, even for slow redox processes such as solid-phase autotrophy given the age of the water. These observations all indicate that regionally reducing conditions un-related to agricultural activities do exist at the KCD1 site. Rates of denitrification in this deep system are unconstrained but may be slow and controlled by the abundance or reactivity of solid-phase electron donors.

The perched shallow aquifer is impacted by agricultural operations. Total inorganic nitrogen ($NO_3 + NO_2 +$ excess $N_2$) shows a secular trend with apparent groundwater age, with the highest
concentrations in the youngest water. The isotopic composition of high-nitrate waters indicates a wastewater source. Groundwater transport modeling indicates that irrigation dominates recharge in the perched aquifer. Irrigation with dairy wastewater results in the percolation of high-nitrate water to the water table and the penetration of this water to a depth controlled by agricultural pumping (Figure 16). Both the vertical and later transport of irrigation water is controlled by agricultural pumping. The perched aquifer is also strongly stratified with respect to oxidation state, nitrate distribution, and denitrification activity. Denitrification under irrigated fields occurs where oxic high-nitrate irrigation water mixes with older anoxic water. The mixing or “reaction” zone is sharp and at constant depth, and may be controlled by agricultural pumping.

What is the electron donor for the denitrification observed at the oxic-anoxic interface? Sediment organic-C and total-S concentrations in the deep and perched aquifer are comparable and are sufficient (assuming most of the S to be present in reduced phases) to create reducing conditions and support denitrification. At one shallow site (Site 3) upgradient of the main dairy operation, PCR data do indicate the presence of autotrophic bacteria capable of using reduced S as an electron donor, and geochemical modeling is consistent with pyrite oxidation. This evidence is not seen at the other sites, however, and the vertical variability in sediment C and S, does not explain the sharpness or location of the oxic-anoxic interface. Total organic carbon in site groundwaters varies from < 1 to 20 mg/L. (Neither other potential dissolved-phase electron donors such as thiosulfate nor the reactivity or bioavailability of the dissolved organic carbon was characterized.) Geochemical modeling is consistent with organic C oxidation, although simple models that assume shallow and deep waters have similar initial chemical compositions do not match observed compositions tightly. These observations, coupled with the lack of evidence for widespread distribution of autotrophic denitrifying bacteria in active denitrification zones, indicate that heterotrophy dominates the observed denitrification in the agriculturally-impacted perched aquifer. Simulations of irrigation and pumping at the KCD1 site indicate that groundwater flow at this site is strongly vertically compartmentalized. The location of the redox gradient close to the top of the production well screens suggests that agricultural pumping and lateral groundwater flow in conjunction may be important controls on the development of chemical and redox stratification in the shallow aquifer.

The conceptual model, then, is of a regionally extensive deep aquifer that is naturally reducing and is unimpacted by agricultural operations overlain by a shallow aquifer that in its upper strata is strongly stratified, is reducing, and is the site of active denitrification of dairy-derived nitrate, and that these conditions in the shallow aquifer are driven by irrigation with dairy wastewater and groundwater pumping for dairy operations. This proposition, that denitrification in shallow nitrate-impacted aquifers is driven by dairy operations, is consistent with observations at not only the KCD1 site but also with evidence for denitrification at the MCD and SCD sites. The implication is that to assess net impact of dairy operations on groundwater quality, one must consider denitrification in the saturated zone.
CONCLUSIONS

The three primary findings of this research are that dairy operations do impact underlying groundwater quality in California’s San Joaquin Valley, that dairy operations also appear to drive denitrification of dairy-derived nitrate in these groundwaters, and that new methods are available for characterization of nitrate source, transport and fate in the saturated zone underlying dairy operations.

Groundwater quality impact has been demonstrated at three sites, with a site in the southern San Joaquin Valley, KCD1, being the best characterized. High nitrate in groundwaters underlying these dairy sites can be attributed to dairy operations using a number of methods, including

- Chemical composition and nitrogen speciation.
- Nitrate isotopic composition.
- Groundwater dissolved gas content and composition.
- Groundwater age
- Reactive transport and flow modeling

The use of chemical composition, nitrogen speciation, and nitrate isotopic composition are well described in the literature. The use of dissolved gas content to identify manure lagoon seepage is new, and is introduced in this research. Groundwater age and transport simulations can be used to trace contaminants back to their source.

In both northern and southern San Joaquin Valley sites, saturated-zone denitrification occurs and mitigates the impact of nitrogen loading on groundwater quality. At the southern KCD1 site, the location and extent of denitrification in the upper aquifer is driven by irrigation with dairy wastewater and groundwater pumping. The extent of denitrification can be characterized by measuring “excess” nitrogen and nitrate isotopic composition while the location of denitrification can be determined using a PCR bioassay for denitrifying bacteria that developed in this research. The demonstration of saturated-zone denitrification in dairy groundwaters is important in assessing the net impact of dairy operations on groundwater quality.

New tools available for research on dairy groundwater include the determination of groundwater dissolved gas content to distinguish dairy wastewater irrigation from dairy wastewater lagoon seepage, field determination of excess nitrogen to identify denitrification in synoptic surveys and to characterize the extent of denitrification in monitor and production well samples, bioassay of aquifer sediment and water samples for the presence of denitrifying bacteria, characterization of aquifer heterogeneity using direct-push drilling and geostatistical simulation methods. Application of these new methods in conjunction with traditional hydrogeologic and agronomic methods will allow a more complete and accurate understanding of the source, transport and fate of dairy-derived nitrogen in the subsurface, and allow more quantitative estimates of net impact of dairy operations on underlying groundwater.
PUBLICATIONS AND PRESENTATIONS

Peer-Reviewed Presentations


Conference presentations


Moran J. E., Moore K., McNab W., Esser B. K., Hudson B., and Ekwurzel B. (2005) Sources and transport of nitrate in the Livermore Valley Groundwater Basin (abstr.). *Joint GSA-AAPG Cordilleran Section Meeting (San Jose, April 29 - May 1, 2005)*.


REFERENCES


Morgan, J. E., 2006. California GAMA Program: Fate and transport of wastewater indicators: Results from ambient groundwater and from groundwater directly influenced by wastewater. Lawrence Livermore National Laboratory, UCRL-TR-222531-DRAFT.


Table 1: KCD2, KCD3, & SCD Site Data

Field Parameters, chemical composition, groundwater age, recharge temperature, excess air, stable isotopic composition, excess nitrogen

(Unless otherwise indicated, all analytes are reported as mg/L; nitrate is reported as nitrate)

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Saturated Zone Denitrification: Potential for Natural Attenuation of Nitrate Contamination in Shallow Groundwater Under Dairy Operations


Chemical Sciences Division, Lawrence Livermore National Laboratory, Environmental Restoration Division, Lawrence Livermore National Laboratory, and Department of Land, Air, and Water Resources, University of California at Davis

We present results from field studies at two central California dairies that demonstrate the prevalence of saturated-zone denitrification in shallow groundwater with 41/3He apparent ages of <35 years. Concentrated animal feeding operations are suspected to be major contributors of nitrate to groundwater, but saturated zone denitrification could mitigate their impact to groundwater quality. Denitrification is identified and quantified using N and O stable isotope compositions of nitrate coupled with measurements of excess N2 and residual NO3- concentrations. Nitrate in dairy groundwater from this study has δ15N values (4.3–61%), and δ18O values (−4.5–24.5%) that plot with δ18O/δ15N slopes of 0.47–0.66, consistent with denitrification. Noble gas mass spectrometry is used to quantify recharge temperature and excess air content. Dissolved N2 is found at concentrations well above those expected for equilibrium with air or incorporation of excess air, consistent with reduction of nitrate to N2. Fractionation factors for nitrogen and oxygen isotopes in nitrate appear to be highly variable at a dairy site where denitrification occurs in a laterally extensive anoxic zone 5 m below the water table, and at a second dairy site where denitrification occurs near the water table and is strongly influenced by localized lagoon seepage.

Introduction

High concentrations of nitrate, a cause of methemoglobinemia in infants (1), are a national problem in the United States (2), and nearly 10% of public drinking water wells in the state of California are polluted with nitrate at concentrations above the maximum contaminant level (MCL) for drinking water set by the U.S. Environmental Protection Agency (3). The federal MCL is 10 mg/L as N, equivalent to the California EPA limit of 45 mg/L as NO3- (all nitrate concentrations are hereafter given as NO3-). In the agricultural areas of California’s Central Valley, it is not uncommon to have nearly half the active drinking water wells produce groundwater with nitrate concentrations in the range considered to indicate anthropogenic impact (>13–18 mg/L (2, 4). The major sources of this nitrate are septic discharge, fertilizer application, sewage treatment plant discharges, and manure and synthetic nitrogen sources, and concentrated animal feeding operations. Dairies are the largest concentrated animal operations in California, with a total heard size of 1.7 million milking cows (5).

Denitrification is the microbially mediated reduction of nitrate to gaseous N2, and can occur in both unsaturated and saturated soils and below the water table where the presence of NO3-, denitrifying bacteria, low O2 concentrations, and electron donor availability exist. In the unsaturated zone, denitrification is recognized as an important process in manure and fertilizer management (6). Although a number of field studies have shown the potential of denitrification in the saturated zone (e.g., 7, 8–11), prior to this study it was not known whether saturated zone denitrification could mitigate the impact of nitrate loading at dairy operations. The combined use of tracers of denitrification and groundwater dating allows us to distinguish between nitrate dilution and denitrification, and to detect the presence of pre-modern water at two dairy operations in the Central Valley of California, referred to here as the Kings County Dairy (KCD) and the Merced County Dairy (MCD) (Figure 1). Detailed descriptions of the hydrogeologic settings and dairy operations at each site are included as Supporting Information.

Materials and Methods

Concentrations and Nitrate Isotopic Compositions. Samples for nitrate and O isotopic compositions were filtered in the field to 0.45 μm and stored cold and dark until analysis. Anion and cation concentrations were determined by ion chromatography using a Dionex DX-600. Field measurements of dissolved oxygen and oxidation reduction potential (using Ag/AgCl with 3.33 mol/L KCl as the reference electrode) were carried out using a Horiba U-22 water quality analyzer. The nitrogen and oxygen isotopic compositions (δ15N and δ18O) of nitrate in 23 groundwater samples from KCD and MCD were measured at Lawrence Berkeley National Laboratory’s Center for Isotope Geochemistry using a version of the denitrifying bacteria procedure (12) as described in Singleton et al. (13). In addition, the nitrate from 17 samples was extracted by ion exchange procedure of (14) and analyzed for δ15N at the University of Waterloo. Analytical uncertainty (1σ) is 0.3% for δ15N of nitrate and 0.5% for δ18O of nitrate. Isotopic compositions of oxygen in water were determined on a VG Prism isotope ratio mass spectrometer at Lawrence Livermore National Laboratory (LLNL) using the CO2 equilibration method (15), and have an analytical uncertainty of 0.1%.

Membrane Inlet Mass Spectrometry. Previous studies have used gas chromatography and/or mass spectrometry to measure dissolved N2 gas in groundwater samples (16–19). Dissolved concentrations of N2 and Ar for this study were analyzed by membrane inlet mass spectrometry (MIMS), which allows for precise and fast determination of dissolved gas concentrations in water samples without a separate extraction step, as described in Kana et al. (20, 21). The gas abundances are calibrated using water equilibrated with air under known conditions of temperature, humidity, and nitrogen content. A small isotopic interference from CO2 at mass 28 (N2) is corrected based on calibration with CO2-rich waters with known dissolved N2, but is negligible for most samples. Samples are collected for MIMS analysis in 40 mL amber
glass VOA vials with no headspace that are kept cold during transport, and then analyzed within 24 h.

Noble Gases and $^3$H/$^4$He Dating. Dissolved noble gas samples are collected in copper tubes, which are filled without bubbles and sealed with a cold weld in the field. Dissolved noble gas concentrations were measured at LLNL after gas extraction on a vacuum manifold and cryogenic separation of the noble gases. Concentrations of He, Ne, Ar, and Xe were measured on a quadrupole mass spectrometer. The ratio of $^3$He to $^4$He was measured on a VG5400 mass spectrometer. Calculations of excess air and recharge temperature from Ne and Xe measurements are described in detail in Ekwurzel (22), using an approach similar to that of Aeschbach-Hertig et al. (23).

Tritium samples were collected in 1 L glass bottles. Tritium was determined by measuring $^3$He accumulation after vacuum degassing each sample and allowing 3–4 weeks accumulation time. After correcting for sources of $^3$He not related to $^3$H decay (24, 25), the measurement of both tritium and its daughter product $^3$He allows calculation of the initial tritium present at the time of recharge, and apparent ages can be determined from the following relationship based on the production of tritiogenic helium ($^{3}{H}_{\text{ex}}$):

\[
\text{Groundwater Apparent Age (years)} = -17.8 \times \ln (1 + ^{3}\text{He}_{\text{ex}}/^{3}\text{H})
\]

Groundwater age dating has been applied in several studies of basin-wide flow and transport (25–27). The reported groundwater age is the mean age of the mixed sample, and furthermore, is only the age of the portion of the water that contains measurable tritium. Average analytical error for the age determinations is ±1 year, and samples with $^{3}$H that is too low for accurate age determination (<1 pCi/L) are reported as >50 years. Significant loss of $^3$He from groundwater is not likely in this setting given the relatively short residence times and high infiltration rates from irrigation. Apparent ages give the mean residence time of the fraction of recently recharged water in a sample, and are especially useful for comparing relative ages of water from different locations at each site. The absolute mean age of groundwater may be obscured by mixing along flow paths due to heterogeneity in the sediments (28).

Results and Discussion

Nitrate in Dairy Groundwater. Nitrate concentrations at KCD range from below detection limit (BDL, <0.07 mg/L) to 274 mg/L. Within the upper aquifer, there is a sharp boundary between high nitrate waters near the surface and deeper, low nitrate waters. Nitrate concentrations are highest between 6 and 13 m below ground surface (BGS) at all multilevel wells (0.5 m screened intervals), with an average concentration of 98 mg/L. Groundwater below 15 m has low nitrate concentrations ranging from BDL to 2.8 mg/L, and also has low or nondetectable ammonium concentrations. The transition from high to low nitrate concentration corresponds to decreases in field-measured oxidation-reduction potential (ORP) and dissolved oxygen (DO) concentration. ORP values are generally above 0 mV and DO concentrations are >1 mg/L in the upper 12 m of the aquifer, defining a more oxidizing zone (Figure 2). A reducing zone is indicated below.
12 m by ORP values as low as -196 mV and DO concentrations <1.2 mg/L. Vertical head varies by less than 10 cm in the upper aquifer multilevel wells.

Nitrate concentrations at MCD monitoring wells sampled for this study range from 2 to 426 mg/L with an average of 230 mg/L. Several wells (W-02, W-16, and W-17) located next to a lagoon and corral have lower nitrate but high ammonium concentrations (Table 1 in Supporting Information). The MCD wells are all screened at the top of the unconfined aquifer except W98, a supply well that is pumped from approximately 57 m BGS. Nitrate concentrations observed for this deeper well are <1 mg/L.

**Dissolved Gases.** Nitrogen gas, the comparatively conservative product of denitrification, has been used as a natural tracer to detect denitrification in the subsurface (16–18). Groundwater often also contains N₂ beyond equilibrium concentrations due to incorporation of excess air from physical processes at the water table interface (23, 29, 30). In the saturated zone, total dissolved N₂ is a sum of these three sources:

\[
\text{[N₂]_{dissolved}} = \text{[N₂]_{equilibrium}} + \text{[N₂]_{excess air}} + \text{[N₂]_{denitrification}}
\]

By normalizing the measured dissolved concentrations as N₂/Ar ratios, the amount of excess N₂ from denitrification can be calculated as

\[
\text{[N₂]_{denitrification}} = \frac{\text{[N₂]_{measured}} - \left(\frac{\text{[N₂]_{equilibrium}} + \text{[N₂]_{excess air}}}{\text{Ar}_{equilibrium} + \text{Ar}_{excess air}}\right)\text{Ar}_{measured}}{\text{Ar}_{measured}}
\]

where the N₂ and Ar terms for equilibrium are calculated from equilibrium concentrations determined by gas solubility. The N₂/Ar ratio is relatively insensitive to recharge temperature, but the incorporation of excess air must be constrained in order to determine whether denitrification has shifted the ratio to higher values (19). Calculations of excess N₂ based on the N₂/Ar ratio assume that any excess air entrapped during recharge has the ratio of N₂/Ar in the atmosphere (83, 5). Any partial dissolution of air bubbles would lower the N₂/Ar ratio (30, 31), thus decreasing the apparent amount of excess N₂.

For this study, Xe and Ne derived recharge temperature and excess air content were determined for 12 of the monitoring wells at KCD and 9 wells at MCD. For these sites, excess N₂ can be calculated directly, accounting for the contribution of excess air and recharge temperature. Site representative mean values of recharge temperature and excess air concentration are used for samples without noble gas measurements. Mean annual air temperatures at the KCD and MCD sites are 17 and 16 °C, respectively (32), and the Xe-derived average recharge temperatures for the KCD and MCD sites are 19 and 18 °C. Recharge temperatures are most likely higher than mean annual air temperature because most recharge is from excess irrigation during the summer months. The average amount of excess air indicated by Ne concentrations is 2.2 × 10⁻³ cm³(STP)/g H₂O for KCD and 1.7 × 10⁻³ cm³(STP)/g H₂O for MCD. From these parameters, we estimate the site representative initial N₂/Ar ratios including excess air to be 41.2 for KCD and 40.6 for MCD. Measured N₂/Ar ratios greater than these values are attributed to production of N₂ by denitrification.

The excess N₂ concentration can be expressed in terms of the equivalent reduced nitrate that it represents in mg/L NO₃⁻ based on the stoichiometry of denitrification. Considering excess N₂ in terms of equivalent NO₃⁻ provides a simple test to determine whether there is a mass balance between nitrate concentrations and excess N₂. From Figure 2, there does not appear to be a balance between nitrate concentrations and excess N₂ in KCD groundwater, since nitrate concentrations in the shallow wells are more than twice that of equivalent excess N₂ concentrations in the anoxic zone. There are multiple possible causes of the discrepancy between NO₃⁻ concentrations and excess N₂ concentrations including (1) the NO₃⁻ loading at the surface has increased over time, and denitrification is limited by slow vertical transport into the anoxic zone, (2) mixing with deeper, low initial NO₃⁻ waters has diluted both the NO₂⁻ and excess N₂ concentrations, or (3) some dissolved N₂ has been lost from the saturated zone. All three processes may play a role in N cycling at the dairies, but we can shed some light on their relative importance by considering the extent of denitrification and then constraining the time scale of denitrification as discussed in the following sections.

**Isotopic Compositions of Nitrate.** Large ranges in δ¹⁵N and δ¹⁸O values of nitrate are observed at both dairies (Figure 3). Nitrate from KCD has δ¹⁵N values of 4.3–61.1‰, and δ¹⁸O values of -0.7 to 24.5‰. At MCD, nitrate δ¹⁵N values range from 5.3 to 30.2‰, and δ¹⁸O values range from -0.7 to 13.1‰. The extensive monitoring well networks at these sites increase the probability that water containing residual nitrate from denitrification can be sampled.

Nitrate δ¹⁵N and δ¹⁸O values at both dairies are consistent with nitrification of ammonium and mineralized organic N
Compounds from manure-rich wastewater, which is stored and used as a fertilizer at both dairy sites. At some locations, nitrification has been followed by denitrification. Prior to nitrification, cow manure likely starts out with a bulk $\delta^{15}N$ value close to 5%, but is enriched in $^{15}N$ to varying degrees due to volatile loss of ammonia, resulting in $\delta^{15}N$ values of 10 – 22% in nitrate derived from manure. Culture experiments have shown that nitrification reactions typically combine 2 oxygen atoms from the local pore water and one oxygen atom from atmospheric $O_2$ (35, 36), which has a $\delta^{18}O$ of 23.5% (37). Different ratios of oxygen from water and atmospheric $O_2$ are possible for very slow nitrification rates and low ammonia concentrations (38), however for dairy wastewater we assume that the 2:1 relation gives a reasonable prediction of the starting $\delta^{18}O$ values for nitrate at the two dairies based on the average values for $\delta^{18}O$ of groundwater at each site (−12.6% at KCD and −9.9% at MCD). Based on this approach, the predicted initial values for $\delta^{15}N$ in nitrate are −0.7% at KCD and 1.1% at MCD. Samples with the lowest nitrate $\delta^{15}N$ values have $\delta^{18}O$ values in this range, and are consistent with nitrate derived from manure. There is no strong evidence for mixing with nitrate from synthetic nitrogen fertilizers, which are used occasionally at both sites, but typically have low $\delta^{15}N$ values (0 – 5%) and $\delta^{18}O$ values around 23% (39).

Denitrification drives the isotopic composition of the residual nitrate to higher $\delta^{15}N$ and $\delta^{18}O$ values. The stable isotopes of nitrogen are more strongly fractionated during denitrification than those of oxygen, leading to a slope of approximately 0.5 on a $\delta^{18}O$ vs $\delta^{15}N$ diagram (34). Nitrate $\delta^{15}N$ and $\delta^{18}O$ values at individual KCD multilevel well sites are positively correlated with calculated slopes ranging from 0.47 to 0.60; the slope of first encounter well data at MCD is 0.66 (Figure 3). These nitrate $\delta^{15}N$ and $\delta^{18}O$ values indicate that denitrification is occurring at both sites. Because a wide range of fractionation factors are known to exist for this process (40), it is not possible to determine the extent of denitrification using only the isotopic compositions of nitrate along a denitrification trend, even when the initial value for manure-derived nitrate can be measured or calculated.

**Extent of Denitrification.** The concentrations of excess N$_2$ and residual nitrate can be combined with the isotopic composition of nitrate in order to characterize the extent of denitrification. In an ideal system, denitrification leads to a regular decrease in nitrate concentrations, an increase in excess N$_2$, and a Rayleigh-type fractionation of N and O isotopes in the residual nitrate (Figure 4). In the Rayleigh fractionation model (41) the isotopic composition of residual nitrate depends on the fraction of initial nitrate remaining in the system ($f = C_{\text{initial}}/C_{\text{final}}$, the initial $\delta^{15}N$, and the fractionation factor ($\alpha$) for denitrification:

$$\delta^{15}N = \left(1000 + \delta^{15}N_{\text{initial}}/f^{(0-1)}\right) - 1000$$

The fractionation factor $\alpha$ is defined from the isotopic ratios of interest ($R = \delta^{15}N/\delta^{14}N$ and $\delta^{18}O/\delta^{16}O$):

$$\alpha = \frac{\left(R_{\text{Product}}\right)}{\left(R_{\text{Reactant}}\right)}$$

This fractionation can also be considered as an enrichment factor ($e$) in % units using the approximation $e = 1000 \ln \alpha$. The extent of denitrification can be calculated as 1 – $f$. Rather than relying on an estimate of initial nitrate concentration, the parameter $f$ is determined directly using field measurements of excess N$_2$ in units of equivalent reduced N$_2$:

$$f = C_{\text{NO}_2}/(C_{\text{NO}_2} + C_{\text{excess} \text{ N}_2})$$

Heterogeneity in groundwater systems can often complicate the interpretation of contaminant degradation using a Rayleigh model (42). Denitrified water retains a proportion of its excess N$_2$ concentration (and low values of $f$) during mixing, but the isotopic composition of nitrate may be disturbed by mixing since denitrified waters contain extremely low concentrations of nitrate (< 1 mg/L). The sample from 1S with a $f$ value close to zero and a $\delta^{15}N$ value of 7.6% was likely denitrified and is one example of this type of disturbance. However, in general, groundwater samples from the same multilevel well sites at KCD fall along similar Rayleigh fractionation curves, indicating that the starting isotopic composition of nitrate and the fractionation factor of denitrification vary across the site (Figure 4).

Values of $\delta^{15}N$ and $f$ calculated from nitrate and excess N$_2$ fall along Rayleigh fractionation curves with enrichment factors ($e$) ranging from −57% to −7% for three multilevel well sites at KCD and first encounter wells at MCD. As expected for denitrification, the enrichment factors indicated for oxygen are roughly half of those for nitrogen. The magnitude of these enrichment factors for N in residual nitrate are among the highest reported for denitrification, which typically range from −40% to −5% (34, 40). Partial gas loss near the water table interface at MCD could potentially increase the value of $f$, resulting in larger values of $e$. Gas loss is unlikely to affect fractionation factors at KCD since most excess N$_2$ is produced well below the water table. Considering the large differences observed for denitrification fractionation factors within and between the two dairy sites, it is not sufficient to estimate fractionation factors for denitrification at dairies based on laboratory-derived values or field-derived values from other sites. The appropriate fractionation factors must be determined for each area, and even then the processes of mixing and gas loss must be considered in the relation between isotopic values and the extent of denitrification. Nevertheless, direct determination of the original amount of nitrate using dissolved N$_2$ values significantly improves our ability to determine the extent of denitrification in settings where the initial nitrate concentrations are highly variable.
Time Scale of Denitrification. Modern water (i.e., groundwater containing measurable tritium) is found at all multi-level wells completed in the upper aquifer at KCD, the deepest of which is 20 m BGS. The upper aquifer below KCD has 3H/3He apparent ages of <35 years. At well 1D1 (54 m BGS), the lower aquifer has no measurable NO₃⁻ and tritium below 1 pCi/L, indicating a groundwater age of more than 50 years. The sum of nitrate and excess N₂ is highest in the young, shallow dairy waters at KCD. Samples with 3H/3He ages >29 years were below the MCL for nitrate prior to denitrification. These results are consistent with an increase in nitrate loading at the surface, which followed the startup of KCD operations in the early 1970s.

The extent of denitrification at KCD is related to both depth and groundwater residence times based on 3H/3He apparent ages (Figure 5). There is a sharp transition from high nitrate waters to denitrified waters between 11 and 13 m depth across the KCD site. This transition is also related to the apparent age of the groundwater, as the high nitrate waters typically have apparent ages of between 0 and 5 years, and most samples with ages greater than 8 years are significantly or completely denitrified. There are five samples...
that do not follow this pattern. These outliers are from sites 35 and 45 where the shallow groundwater has much higher $^{3}H/^{3}H$ apparent ages due to slow movement around clay zones at the screened intervals for these samples. The existence of older water that is not significantly impacted by denitrification indicates that it is the physical transport of water below the transition from oxic to anoxic conditions rather than the residence time that governs denitrification in this system.

At the MCD site, groundwater $^{3}H/^{3}H$ apparent ages indicate fast transit rates from the water table to the shallow monitoring wells. Most of the first encounter wells have apparent ages of $<2$ years, consistent with the hydraulic analysis presented by Harter et al. (5). The very fast transit times to the shallow monitoring wells at MCD allow for some constraints on minimum denitrification rates at this site. Based on the comparison of the calculated ages with the initial tritium curve, these shallow wells contain a negligible amount of old, $^{3}H$-decayed water. In shallow wells near lagoons (e.g., W-16 and V-21), the observed excess N$_2$ (equivalent to 71 and 40 mg/L of reduced N$_2$) accumulated over a duration of less than 1 year, indicating that denitrification rates may be very high at these sites. Complete denitrification of groundwater collected from well W-98 (excess N$_2$ equivalent to 51 mg/L N$_2$O) was attained within approximately 31 years, but may have occurred over a short period of time relative to the mean age of the water.

Occurrence of Denitrification at Dairy Sites. The depth at which denitrified waters are encountered is remarkably similar across the KCD site. This transition is not strongly correlated with a change in sediment texture. The denitrified waters at all KCD wells coincide with negative ORP values and generally low dissolved O$_2$ concentrations. Total organic carbon (TOC) concentrations in the shallow groundwaters range from 1.1 to 15.7 mg/L at KCD, with the highest concentrations of TOC found in wells adjacent to lagoons. The highest concentrations of excess N$_2$ are found in nested well-set 25, which is located in a field downgradient from the lagoons. However, sites distal to the lagoons (35 and 45) that are apparently not impacted by lagoon seepage (45) also show evidence of denitrification, suggesting that direct lagoon seepage is not the sole driver for this process.

The chemical stratification observed in multilevel wells at the KCD site demonstrates the importance of characterizing vertical variations within aquifers for nitrate monitoring studies. Groundwater nitrate concentrations are integrated over the high and low nitrate concentration zones by dairy water supply wells, which have long screened intervals from 9 to 18 m BOS. Water quality samples from these supply wells underestimate the actual nitrate concentrations present in the uppermostoxic aquifer. Similarly, first encounter monitoring wells give an overestimate of nitrate concentrations found deep in the aquifer, and thus would miss entirely the impact of saturated zone denitrification in mitigating nitrate transport to the deep aquifer.

Monitoring wells at MCD sample only the top of the aquifer, so the extent of denitrification at depth is unknown, except for the one deep supply well (W98), which has less than 1 mg/L nitrate and an excess N$_2$ content consistent with reduction of 51 mg/L N$_2$ to N$_2$. This supply well would be above the MCL for nitrate without the attenuation of nitrate by denitrification. The presence of ammonium at several of the wells with excess N$_2$ indicates a component of wastewater seepage in wells located near lagoons, where mixing of oxic waters with anoxic lagoon seepage may induce both nitrification and denitrification. Wells that are located in the surrounding fields have high NO$_3$ concentrations, and do not have any detectable excess N$_2$, a result consistent with mass-balance models of nitrate loading and groundwater nitrate concentration (5).

While dairy operations seem likely to establish conditions conducive to saturated zone denitrification, the prevalence of the phenomenon is not known. Major uncertainties include the spatial extent of anaerobic conditions, and transport of organic carbon under differing hydrogeologic conditions and differing nutrient management practices. Lagoon seepage may also increase the likelihood of denitrification in dairy aquifers. The extent to which dairy animal and field operations affect saturated zone denitrification is an important consideration in determining the assimilative capacity of underlying groundwater to nitrogen loading associated with dairy operations.

Acknowledgments

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Supporting Information Available

A table of chemical, isotopic, and dissolved gas results from this study, a plot of apparent age with depth, and detailed descriptions of the study sites. This material is available free of charge via the Internet at http://pubs.acs.org.

Literature Cited


(28) Heaton, T. H. E.; Vogel, J. C. Excess air in groundwater.

Supporting Information for "Saturated Zone Denitrification: Potential for Natural Attenuation of Nitrate Contamination in Shallow Groundwater Under Dairy Operations" by M. J. Singleton¹, B. K. Esser¹, J. E. Moran¹, G. B. Hudson¹, W. W. McNab², and T. Harter³

Contents: 7 Pages, 1 Figure, and 1 Table
Description of Dairy Sites

*Study Site 1:*

Study Site #1 is located at a dairy operation in Kings County, CA (KCD). Manure management practices employed at KCD, with respect to corral design, runoff capture and lagoon management are typical of practices employed at other dairies in the region. KCD has close to the 1000-cow average for dairies in the area, and operates three clay-lined wastewater lagoons that receive wastewater after solids separation. Wastewater is used for irrigation of 500 acres of forage crops (corn and alfalfa) on the dairy and on neighboring farms; dry manure is exported to neighboring farms.

KCD is located in the Kings River alluvial fan, a sequence of layered sediments transported by the Kings River from the Sierra Nevada to the low lying southern San Joaquin Valley of California (1, 2). The site overlies an unconfined aquifer, which has been split into an upper aquifer from 3m to 24m below ground surface (BGS) and a lower aquifer (>40 m BGS) that are separated by a gap of unsaturated sediments. Both aquifers are predominantly composed of unconsolidated sands with minor clayey sand layers. The lower unsaturated gap was likely caused by intense regional groundwater pumping, and a well completed in this unsaturated zone has very low gas pressures. There are no persistent gradients in water table levels across the KCD site, but in general, regional groundwater flow is from the NW to SE due to topographic flow on the Kings River fan. The water table is located about 5 m BGS. Local recharge is dominated by vertical fluxes from irrigation, and to a lesser extent, leakage from adjacent unlined canals. Transient cones of depression are induced during groundwater pumping from dairy operation wells.
The regional groundwater is highly impacted by agricultural activities and contains elevated concentrations of nitrate and pesticides (3, 4).

KCD was instrumented with five sets of multi-level monitoring wells and one “up-gradient” well near an irrigation canal. These wells were installed in 2002, and sampled between Feb. 2002 and Aug. 2005. The multi-level wells have short (0.5 m) screened intervals in order to detect heterogeneity and stratification in aquifer chemistry. One monitoring well was screened in the lower aquifer, 54m BGS. The remaining monitoring wells are screened in the upper aquifer from 5m to 20m BGS. In addition, there are eight dairy operation wells that were sampled over the course of this study. These production wells have long screens, generally between 9 to 18 meters below ground surface (BGS).

Study Site 2:

The second dairy field site is located in Merced County, CA. The Merced County dairy (MCD) lies within the northern San Joaquin Valley, approximately 160 km NNW from the KCD site. The site is located on the low alluvial fans of the Merced and Tuolumne Rivers, which drain the north-central Sierra Nevada. Soils at the site are sand to loamy sand with rapid infiltration rates. The upper portion of the unconfined alluvial aquifer is comprised of arkosic sand and silty sand, containing mostly quartz and feldspar, with interbedded silt and hardpan layers. Hydraulic conductivities were measured with slug tests and ranged from $1 \times 10^{-4}$ m/s to $2 \times 10^{-3}$ m/s with a geometric mean of $5 \times 10^{-4}$ m/s (5). Regional groundwater flow is towards the valley trough with a
gradient of approximately 0.05% to 0.15%. Depth to groundwater is 2.5 m to 5 m BGS. The climate is Mediterranean with annual precipitation of 0.5 m, but groundwater recharge is on the order of 0.5–0.8 m per year with most of the recharge originating from excess irrigation water (3). Transit times in the unsaturated zone are relatively short due to the shallow depth to groundwater and due to low water holding capacity in the sandy soils. Shallow water tables are managed through tile drainage and groundwater pumping specifically for drainage. The MCD site is instrumented with monitoring wells that are screened from 2-3 m BGS to a depth of 7-9 m BGS. The wells access the upper-most part of the unconfined aquifer, hence, the most recently recharged groundwater (6). Recent investigations showed strongly elevated nitrate levels in this shallow groundwater originating largely from applications of liquid dairy manure to field crops, from corrals, and from manure storage lagoons (6). For this study, a subset of 18 wells was sampled. A deep domestic well was also sampled at MCD. This domestic well is completed to 57 m BGS, and thus samples a deeper part of the aquifer than the monitoring well network.
Figure S1. Groundwater $^3$H/$^3$He apparent ages from multilevel monitoring wells at KCD. Error bars show analytical error.
Table S1. Chemical, dissolved gas, and isotopic compositions for multilevel groundwater monitoring wells and lagoons. Average values are given for wells sampled more than once.

<table>
<thead>
<tr>
<th>Site</th>
<th>Depth of multilevel well (m)</th>
<th>C (mg/L)</th>
<th>NO₃ (mg/L)</th>
<th>NH₄ (mg/L)</th>
<th>ORP (mg/L)</th>
<th>DOC (mg/L)</th>
<th>TOC (mg/L)</th>
<th>δ₁₈O H₂O (‰)</th>
<th>δ¹⁵N NO₃ (%)</th>
<th>δ¹⁸O NO₃ (%)</th>
<th>Excess air determined from Ne (ce STP/cm³)</th>
<th>Recharge Temp. (°C)</th>
<th>¹⁸O/H pCi/L</th>
<th>N₂ Ar</th>
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<tbody>
<tr>
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<td>1.5</td>
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<td>0.2</td>
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<td>3.0</td>
<td>1.1</td>
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<td>21.85 ± 0.64</td>
<td>13 ± 1.2</td>
<td>0.1 ± 0.2</td>
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<tr>
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<td>2.0</td>
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<td>3.0</td>
<td>1.1</td>
<td>1.1</td>
<td>1.1</td>
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<td>13 ± 1.2</td>
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<td>28</td>
</tr>
<tr>
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<td>10.0</td>
<td>2.0</td>
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<td>3.0</td>
<td>1.1</td>
<td>1.1</td>
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<td>21.85 ± 0.64</td>
<td>13 ± 1.2</td>
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<td>1.1</td>
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<td>21.85 ± 0.64</td>
<td>13 ± 1.2</td>
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References


Assessing the Impact of Animal Waste Lagoon Seepage on the Geochemistry of an Underlying Shallow Aquifer

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Evidence of seepage from animal waste holding lagoons at a dairy facility in the San Joaquin Valley of California is assessed in the context of a process geochemical model that addresses reactions associated with the formation of the lagoon water as well as reactions occurring upon the mixture of lagoon water with underlying aquifer material. Comparison of model results with observed concentrations of NH4+, K+, PO43-, dissolved inorganic carbon, pH, Ca2+, Mg2+, SO42-, Cl-, and dissolved Ar in lagoon water samples and groundwater samples suggests three key geochemical processes: (i) off-gassing of significant quantities of CO2 and CH4 during mineralization of manure in the lagoon water, (ii) ion exchange reactions that remove K+ and NH4+ from seepage water as it migrates into the underlying anaerobic aquifer material, and (iii) mineral precipitation reactions involving phosphate and carbonate minerals in the lagoon water in response to an increase in pH as well as in the underlying aquifer from elevated Ca2+ and Mg2+ levels generated by ion exchange. Substantial off-gassing from the lagoons is further indicated by dissolved argon concentrations in lagoon water samples that are below atmospheric equilibrium. As such, Ar may serve as a unique tracer for lagoon water seepage since under-saturated Ar concentrations in groundwater are unlikely to be influenced by any processes other than mechanical mixing.

Introduction

Animal waste management at dairy facilities often entails storing dairy wastewater in manure lagoons. Irrigation with such lagoon water is a common practice that utilizes readily available fertilizer for forage crops while reducing the stored wastewater volume. The transfer of anoxic lagoon water to aerated unsaturated zone soils leads to the nitrification of ammonia to nitrate, as well as the mineralization of organic nitrogen, and can impact underlying groundwater when nitrogen is added to the fields in excess of the assimilation capacity of the crops (1–3).

The impact of manure lagoon seepage on groundwater quality is a separate problem from that of fertilizer application but is nonetheless also a groundwater protection concern. Previous studies have indicated that manure lagoons can leak at rates on the order of a few millimeters per day or more based on soil type, construction, and operation (4–10). Geochanical interactions between the seepage water and groundwater may differ from those involving fertilizer application (6, 11–13). For example, nitrate loading from the lagoon will depend on the rate of oxidation of NH4+ and organic nitrogen released from the lagoon that, in turn, are affected by subsurface oxidation–reduction conditions and ion exchange characteristics. Distinguishing lagoon seepage from applied manure fertilizer in monitoring wells is difficult because the multitude of possible geochemical reactions create ambiguities with respect to potential tracers.

This study has sought to understand the effects of lagoon seepage on underlying groundwater quality in the context of a putative set of geochemical reactions characterizing the formation of lagoon water as well as the interaction of lagoon water with the groundwater environment. Our study entailed evaluating water quality data collected at an anonymous dairy facility located in Kings County, CA, in the southern San Joaquin Valley (Figure 1). The dairy holds approximately 1000 cows. Three manure lagoons have been active at the dairy since the 1970s, two of which have liners with a 10% clay content while the third is unlined. The largest lagoon measures approximately 100 m x 20 m. The lagoons receive runoff water from the flushing of animal stalls with water pumped from onsite agricultural wells. In turn, lagoon water is mixed with additional pumped groundwater and applied to onsite corn and alfalfa fields. Water depth within the lagoons varies temporally, depending on site operations, but is constrained to a maximum of approximately 3 m to prevent overflow. The site climatic setting is semi-arid, with a mean annual rainfall of approximately 220 mm/year, most of it falling from November through April. The daily summer average temperature is approximately 26 °C, although maximum daytime temperatures of 35 °C are common, while daily average winter temperatures are on the order of 7 °C (14).

Groundwater is first encountered in a perched aquifer extending from depths of approximately 3–24 m, separated by an unsaturated zone from a regional aquifer below a 40 m depth. Both aquifers consist of alluvial fan deposits. Measured oxidation–reduction potentials and dissolved gas data delineate the perched aquifer into an upper, aerobic zone above a depth of approximately 11 m below the ground surface (Shallow zone) and a lower, anaerobic zone (Deep zone) subject to denitrification (13). Recharge to the perched aquifer stems from nearby unlined irrigation canals, with a mean groundwater flow direction from northwest to southeast. However, agricultural pumping dominates the shallow hydrologic system, so groundwater flow directions are spatially and temporally variable.

Experimental Procedures

Lagoon water and groundwater samples were collected during six sampling events, from the locations indicated in Figure 1, between August 2004 and May 2005. Samples were analyzed for cations (Ca2+, Mg2+, Na+, K+, Li+, and NH4+) and anions (NO3-, SO42-, Cl−, F−, Br−, PO43−, and NO2−) by ion chromatography using a Dionex DX-600. pH, DO, and oxidation–reduction potential were measured in the field using a Horiba U-22 water quality parameter field meter. Dissolved inorganic carbon (DIC) concentrations were estimated in the water samples from charge imbalances and pH using the PHREEQC geochemical model. DIC was also...
quantified in a subset of samples as CO₂ gas pressure after acidification with orthophosphoric acid. δH and δ¹⁸O were determined using a VG Prism II isotope ratio mass spectrometer and are reported in permil values relative to the Vienna Standard Mean Ocean Water (VSMOW). Oxygen isotope compositions were determined using the CO₂ equilibration method (15), and hydrogen isotope compositions were determined using the Zn reduction method (16). Dissolved gases (O₂, N₂, CO₂, CH₄, and Ar) were measured by membrane inlet mass spectroscopy (MIMS (17)) or noble gas mass spectrometry.

Geochemical trends in water quality data were interpreted using the PHREEQC geochemical model (18). PHREEQC calculates equilibrium water chemistry compositions given an initial water composition, a set of postulated mineral and/or gas phases, and a thermodynamic database of equilibrium reaction constants. For this study, PHREEQC and its associated PHREEQC.DAT database were used to formulate two geochemical processes models: (i) a lagoon water formation model based upon dairy operating practices and a set of assumptions concerning evolution of a multi-component gas phase, oxidation–reduction reaction equilibria, and mineral precipitation and (ii) a seepage model that considers possible ion exchange interactions and mineral precipitation that could occur when seepage water contacts aquifer sediments.

Results

Ideally, a tracer for lagoon seepage should (i) be transported conservatively in groundwater and (ii) be unique to the lagoon environment. While partial pressures of CH₄ and CO₂ measured in site water samples may reflect mineralization of organic matter under anaerobic conditions in the lagoon water (Figure 2), neither indicator is likely to be conservative in groundwater (e.g., CH₄ could be subject to oxidation, while CO₂ is affected by pH). Alternatively, δ¹⁸O and Cl⁻ are elevated in lagoon water (Figure 2) as a result of evaporation and, for Cl⁻, the composition of manure, but both indicators will exist in lagoon seepage as well as applied fertilizer and thus would not provide an unequivocal means of distinguishing the two.

Given these limitations, an alternative approach for identifying lagoon seepage is to evaluate multiple geochemical parameters—major cations, anions, pH, and dissolved gases—together in the context of a geochemical process.
and introduced by addition of manure as well as the complexation. To model lagoon water formation, we assumed gas bubbles will form when the total gas pressure exceeds the gaseous phases in the vicinity of the lagoon are supersaturated with argon, because it can partition into an evolved gas phase, as explained next.

The geochemical modeling scheme is illustrated in Figure 3. Modeling lagoon water formation entailed simulating the mineralization of manure in a starting water composition given by the mean agricultural well water composition (i.e., the water used to flush the animal stalls). Dairy manure is compositionally variable and depends on feed composition, degree of mixing with urine, and storage issues affecting decomposition and preferential loss of volatiles. Reported manure compositions describe nutrient content (nitrogen, phosphorus, and potassium) per unit weight, which is typically less than 5% for dry manure and contains roughly equivalent amounts of nitrogen and potassium with a much smaller phosphorus component (19, 20). We assumed a manure stoichiometry of Ca(HCO₃)₂(PO₄)₀.₁₀₂(KOH)₀.₉ₐ₆, which has a carbon/nitrogen ratio of approximately 34:1 on a per weight basis, similar to the value of 28:1 reported by Cameron et al. (1).

In this formulation, both organic nitrogen and NH₄⁺ are represented by NH₄⁺.

PHREEQC models aqueous species concentrations under an assumption of thermodynamic equilibrium in the presence of user-selected heterogeneous reactions involving gas phases, mineral equilibria, and ion exchange or surface complexation. To model lagoon water formation, we assumed (i) precipitation of calcium- and magnesium-carbonates (idealized as calcite, CaCO₃, and magnesite, MgCO₃) as well as hydroxyapatite, Ca₅(PO₄)₃OH, upon supersaturation and (ii) evolution of a mixed gas phase consisting of CO₂, CH₄, NH₃, H₂S, and Ar when the sum of the partial pressures of the gas components exceeds a threshold pressure. Ideally, gas bubbles will form when the total gas pressure exceeds local hydrostatic pressure in the lagoon; active gas bubble formation is indeed readily observed in the dairy site lagoons. However, mechanical mixing of the lagoon water during water transfer and the natural movement of air across the surface of the lagoon both facilitate diffusive transport, so a loss of gas phase components at a total pressure less than 1 atm is reasonable given the very low ambient partial pressures of all of the listed gas species in air. Separately, evaporative loss during lagoon water formation was simulated by removing half of the fluid volume as pure H₂O concurrent with the mineralization of the manure.

Lagoon seepage simulation entailed mixing the lagoon water with the mean composition of anaerobic groundwater (i.e., from depths greater than 11 m) in the presence of an ion exchanger initially in equilibrium with the same anaerobic groundwater. In the absence of site-specific ion exchange data, an exchange capacity of 0.15 mol of charge/kg of soil (21) and the default cation exchange selectivity coefficient (as utilized by the PHREEQC database for Na⁺, K⁺, NH₄⁺, Ca₂⁺, and Mg²⁺) were assumed. In addition, calcite and magnesite were modeled to precipitate upon supersaturation.

By setting the gas evolution threshold to 0.1 atm, manure loading to 0.45 mol/L, evaporative loss from the lagoon to 50%, and the mixing ratio of lagoon water/groundwater to 1:1, the proposed geochemical model provides a reasonable semiquantitative match to the water quality data set, at an ambient temperature of 25°C, as indicated in Figure 4. The agricultural water (i.e., starting composition for the model, and background groundwater compositions) is also shown in Figure 4 for comparison. Several key processes are suggested by the modeling results and the observed data.

(i) Gas evolution and mineral precipitation can account for the observed concentrations of mineralized manure components (PO₄³⁻ and DIC), pH, and Ca²⁺ and Mg²⁺ concentrations measured in the lagoon water. The model shows that hydroxyapatite precipitation is a plausible sink for PO₄³⁻ introduced by addition of manure as well as the Ca²⁺ present in the agricultural water. Ca²⁺, along with Mg²⁺, can also be removed as carbonates, explaining the low Mg²⁺ content of the lagoon water. Modeling suggests that DIC may be removed from solution by off-gassing (as CO₂ and CH₄) and by precipitation of carbonate minerals in such a manner as to reproduce the observed lagoon water pH.

(ii) Seepage modeling suggests that the high concentrations of NH₄⁺ and K⁺ found in the lagoon water diminish via ion exchange and dilution after a 1:1 mixing event, with the exchange reactions releasing Ca²⁺ and Mg²⁺, which results in calcite and magnesite precipitation and, as a consequence, a pH decline. Calculated calcite saturation indices among site water samples suggest that calcite precipitation is more likely in the lagoon water and in the Near-Lagoon Well than in groundwater at other locations (Figure 5).

Dissolved Ar warrants special mention. In a well-mixed model system, Ar initially dissolved in the agricultural water in equilibrium with the atmosphere partitions into the gas phase generated during lagoon water formation (consisting mainly of a CO₂–CH₄ mixture with a volumetric equivalent of approximately 10.7 L of gas per liter of lagoon water at standard temperature and pressure). Such gas stripping phenomena have been reported for coal bed methane environments (23) and ocean sediment pore waters (24). MIMS data indicate Ar concentrations in the lagoon water, and while not reduced to negligible levels as predicted by the model, they nonetheless appear to be depleted with respect to the atmosphere even at elevated temperature (Figure 5). In comparison, groundwater samples from both shallow and deep portions of the permeable aquifer beyond the vicinity of the lagoon are supersaturated with argon, indicating excess air entrapped during recharge (25). The Near-Lagoon water composition is intermediate between two, supporting the 1:1 mixing assumption used in the seepage model.

Groundwater encountered below a depth of 11 m in Well 25, some 100 m to the east–southeast of the manure lagoons, exhibits indications of lagoon impact such as comparatively low pH and Ar (Figure 5). δ¹⁸O–DIC, quantified in a subset
Discussion

The geochemical model for manure lagoon water formation and seepage proposed in this study is based on idealized assumptions that may lead to error. In our judgment, the most problematic assumptions include the following.

of the data, appears to be elevated in association with the pH and Ar signatures. While δ13C was not addressed in the geochemical model, isotopically heavy DIC residue in the lagoon water is qualitatively consistent with extensive off-gassing of CO2 and/or CH4. As such, data from Well 2S below 11 m were not included in the previous comparisons.
FIGURE 5. Thermodynamic saturation indices for calcite in site water samples, calculated with PHREEQC (left) and Ar concentrations and solubility (22) (right). The box-whisker marks correspond to the minimum, maximum, median, lower quartile, and upper quartile values for each group. Deep samples exclude groundwater samples from Well 2S.

FIGURE 6. Distributions of pH (top), Ar (middle), and δ13C (bottom) in site groundwater, each consistent with lagoon seepage that may have impacted Well 2S at depths greater than 11 m. Iso-surface values for pH correspond to 6.75, 6.8, and 7.3. The isosurface value for Ar corresponds to 3.6 x 10⁻⁴ mol/L. The isosurface values for δ13C correspond to -6.4 and 2.3 per mil.

Perfectly Well-Mixed Lagoon. Some stratification of the lagoons with regard to oxidation-reduction reactions and temperature seems likely, so gas evolution at the surface may reflect a superposition of biogeochemical regimes. Moreover, bubble formation and diffusive gas component losses are separate mechanisms that may operate differently on individual gas phase components depending on the respective diffusion coefficients and other factors. Seasonal and diurnal differences in temperature, microbiological activity in the lagoons, and even the lagoon operation itself will all exert various effects on the rate of off-gassing. This departure from ideality may explain, in part, the inability of the model, with a gas evolution threshold of 0.1 atm, to reproduce the measured CH₄ partial pressures approaching 1 atm (Figure 2).

Thermodynamic Equilibrium within the Lagoon. It is well-recognized that oxidation-reduction processes and some mineral precipitation reactions are slow kinetically. This constraint pertains to all oxidation-reduction reactions occurring in the lagoon—including the assumption of complete mineralization of manure—as well as the precipitation of Mg-rich carbonates that can be kinetically slow (26).

Complexation of Ions with Organic Matter. High concentrations of partially degraded manure constituents in the form of organic acids could complex cations such as Ca²⁺ and Mg²⁺ in the lagoon water, affecting their speciation but not considered by the model (27, 28).

Cation Exchange Model Used for the Aquifer Material. Hypothetical cation exchange characteristics were assumed.

Solute Transport beneath Lagoons. The compartmentalized geochemical model assumes that lagoon water mixes directly with underlying groundwater without passing through an aerobic vadose zone. While the geochemical data appear consistent with this assumption, there is an absence of soil boring data directly beneath the lagoons to support this assertion.

Despite these caveats, we believe that the proposed model has likely identified evidence of three major processes that affect lagoon water formation and seepage: (i) off-gassing of significant quantities of CO₂ and/or CH₄ during mineralization of manure in the lagoon water, (ii) ion exchange reactions that remove K⁺ and NH₄⁺ from seepage water in the underlying aquifer, and (iii) phosphate and carbonate mineral precipitation reactions occurring in the lagoon water resulting from an increase in pH and in the underlying aquifer— including the assumption of some mineral precipitation reactions are slow kinetically.

These results are consistent with findings reported in previous studies. For example, significant fluxes of CH₄ (up to 19 mol m² day⁻¹) were measured from an anaerobic waste lagoon at a swine operation in southwestern Kansas (29), while ion exchange reactions were found to retard the movement of NH₄⁺ in lagoon seepage through soils in both field and laboratory studies (12, 30), with NH₄⁺ occupying more than 20% of the exchange sites in some cases (hence displacing cations such as Ca²⁺). Moreover, the off-gassing process has suggested a new diagnostic tool—dissolved Ar—to detect gas stripped lagoon water that has migrated into ground-
water. At and other noble gases could be particularly useful in distinguishing lagoon seepage from applied fertilizer since lagoon water applied to fields will equilibrate with atmospheric argon prior to infiltration.

**Acknowledgments**

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**Supporting Information Available**

Additional details of our analysis. This material is available free of charge via the Internet at http://pubs.acs.org.

**Literature Cited**

(20) Christensen, P.; Peacock, B. Manure as a Fertilizer; NG7-97; University of California Cooperative Extension: Tulare, CA, 1998.

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TITLE Titration and mixing KCD water quality data set

SOLUTION_MASTER_SPECIES
   Ar   Ar   0   1   1

SOLUTION_SPECIES
   Ar = Ar
   log_k  0

PHASES
Manure
   $\text{CH}_2\text{O} (\text{NH}_3) 0.025 (\text{P}_{2}\text{O}_5) 0.002 (\text{K}_2\text{O}) 0.006 + \text{O}_2 = \text{HCO}_3^- + 0.025\text{NH}_4^+ + 0.004\text{PO}_4^{3-} + 0.012\text{K}^+ + 0.975\text{H}^+$
   log_k  100
Magnesite
   $\text{MgCO}_3 + \text{H}^+ = \text{HCO}_3^- + \text{Mg}^{2+}$
   log_k  2.2936
Ar(g)
   Ar = Ar
   log_k  -2.854

SOLUTION_SPECIES

2 $\text{NO}_3^- + 12 \text{H}^+ + 10 \text{e}^- = \text{N}_2 + 6 \text{H}_2\text{O}$
   #log_k  207.080
   log_k  203.
   delta_h  312.130 kcal

$\text{CO}_3^{2-} + 10 \text{H}^+ + 8 \text{e}^- = \text{CH}_4 + 3 \text{H}_2\text{O}$
   log_k  41.071
   #log_k  45.
   delta_h  61.039 kcal

SOLUTION 1
#Mean agricultural well water
   temp  22
   pH  6.83
   pe  4
   redox $\text{O}(-2)/\text{O}(0)$
   units mg/l
   density  1
   F  0.23
   Cl  156.03
   Br  0.13
   N  72.42 as $\text{NO}_3^-$
   S(6)  440.52 as $\text{SO}_4^{2-}$
   S(-2)  1e-010 as $\text{SO}_4^{2-}$
   P  0.02 as $\text{PO}_4^{3-}$
   Li  0.0067
Na 216.6
K 6.39
Mg 75.99
Ca 209.61
C(-4) 1e-010
C(4) 100 charge
O(0) 1
Ar 1e-010 Ar(g) -2.027
-water 1 # kg

EQUILIBRIUM PHASES
Calcite 0 0
Magnesite 0 0
Hydroxyapatite 0 0

GAS PHASE
-fixed_pressure
-pressure 0.1
-volume 100
-temperature 25
CH4(g) 0
CO2(g) 0
H2S(g) 0
NH3(g) 0
Ar(g) 0

REACTION 1
Manure 0.45
H2O -22
1 moles in 200 steps

SELECTED_OUTPUT
-file titrate.txt
-reset false
-solution true
-distance true
-time true
-step true
-ph true
-pec true
-totals C(4) S(6) C(-4) Fe(2) S(-2) Ca Mg
-Na K F P Ar Cl
-molalities O2 NH4+ NH3 NO3-
-N2
-equilibrium_phases Calcite Magnesite Hydroxyapatite
-saturation_indices CH4(g) CO2(g) H2S(g) NH3(g) N2(g) Ar(g)
SAVE Solution 1

END

SOLUTION 2 #Deep field groundwater

- gases CH4(g) CO2(g) H2S(g) NH3(g) Ar(g)

 temp 22
 pH 7.07
 pe 4
 redox N(0)/N(5)
 units mg/l
 density 1
 F 0.28
 Cl 42.32
 Br 0.08
 N(0) 34.87 as NO3-
 N(5) 1.75 as NO3-
 S(6) 169.39 as SO4-2
 P 0.02 as PO4-3
 Li 0.0033
 Na 65.18
 K 4.83
 Mg 29.62
 Ca 68.91
 Fe 0.001 Goethite
 C(4) 100 charge
 Ar 1e-010 Ar(g) -2.027
 - water 1 # kg

EXCHANGE 1

- equilibrate with solution 2

SAVE Solution 2
SAVE Exchange 1

END

USE Solution 1
USE Solution 2
USE Exchange 1

MIX 1
1 1
2 1
EQUILIBRIUM_PHASES 2
Calcite 0 0
Magnesite 0 0
Hydroxyapatite 0 0
END