

**ERRATA SHEET
TENTATIVE ORDER NO. R9-2009-0001 NPDES NO. CA0107409**

**WASTE DISCHARGE REQUIREMENTS AND
NATIONAL POLLUTANT DISCHARGE ELIMINATION SYSTEM PERMIT
FOR THE CITY OF SAN DIEGO E.W. BLOM
POINT LOMA METROPOLITAN WASTEWATER TREATMENT PLANT
DISCHARGE TO THE PACIFIC OCEAN THROUGH THE
POINT LOMA OCEAN OUTFALL, SAN DIEGO COUNTY**

The following changes have been made to Tentative Order No. R9-2009-0001. Changes below are shown in **bold and underline**/~~strikeout~~ format to indicate added and removed language, respectively.

ERRATA #	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)
1	12	III.B	<p><i>Based on Comment 14 (a):</i></p> <p>Discharge through the PLOO from the Facility in excess of an <u>monthly</u> average daily-flow rate of 240 MGD is prohibited.</p>
2	47	VI.C.6.b	<p><i>Based on Comment 14 (d):</i></p> <p><u>The Discharger shall prepare a feasibility study that assesses behavior of the PLOO wastewater plume and means of tracking the plume. The feasibility study shall present a recommended plan for plume tracking which includes identifying recommended modifications in receiving water sampling parameters, locations, and/or sampling protocols. The feasibility study shall be submitted to the Executive Director within two years of the effective date of this Order.</u></p>

**ERRATA SHEET FOR TENTATIVE ORDER NO. R9-2009-0001
CITY OF SAN DIEGO E.W. BLOM POINT LOMA METROPOLITAN WASTEWATER TREATMENT PLANT**

ERRATA #	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)																																																																		
3	E-21 and E-22	Table E-5 (MRP)	<p><i>Based on Comment 14 (d):</i></p> <p>Table E-5. General Water Quality Monitoring Requirements</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th rowspan="2" style="text-align: center;">Parameter</th> <th rowspan="2" style="text-align: center;">Units</th> <th rowspan="2" style="text-align: center;">Sample Type</th> <th colspan="3" style="text-align: center;">Minimum Sampling Frequency</th> <th rowspan="2" style="text-align: center;">Required Analytical Test Method</th> </tr> <tr> <th style="text-align: center;">Offshore Stations</th> <th style="text-align: center;">Kelp Stations</th> <th style="text-align: center;">Shoreline Stations</th> </tr> </thead> <tbody> <tr> <td>Temperature</td> <td style="text-align: center;">°C</td> <td>Profile</td> <td>1/Quarter</td> <td>5/Month</td> <td style="text-align: center;">--</td> <td style="text-align: center;">1</td> </tr> <tr> <td>Salinity</td> <td style="text-align: center;">ppt</td> <td>Profile</td> <td>1/Quarter</td> <td>5/Month</td> <td style="text-align: center;">--</td> <td style="text-align: center;">1</td> </tr> <tr> <td>Dissolved Oxygen</td> <td style="text-align: center;">mg/L</td> <td>Profile</td> <td>1/Quarter</td> <td>5/Month</td> <td style="text-align: center;">--</td> <td style="text-align: center;">1</td> </tr> <tr> <td>Light Transmittance</td> <td style="text-align: center;">%</td> <td>Profile</td> <td>1/Quarter</td> <td>5/Month</td> <td style="text-align: center;">--</td> <td style="text-align: center;">1</td> </tr> <tr> <td>Chlorophyll a</td> <td style="text-align: center;">m</td> <td>Profile</td> <td>1/Quarter</td> <td>5/Month</td> <td style="text-align: center;">--</td> <td style="text-align: center;">1</td> </tr> <tr> <td>pH</td> <td style="text-align: center;">units</td> <td>Profile</td> <td>1/Quarter</td> <td>5/Month</td> <td style="text-align: center;">--</td> <td style="text-align: center;">1</td> </tr> <tr> <td>Ammonium (NH₄⁺)</td> <td style="text-align: center;">mg/L</td> <td>Profile</td> <td>1/Quarter</td> <td>5/Month</td> <td style="text-align: center;">--</td> <td style="text-align: center;">1,3</td> </tr> <tr> <td>Visual Observations²</td> <td style="text-align: center;">--</td> <td>Visual</td> <td>1/Quarter</td> <td>5/Month</td> <td style="text-align: center;">5/Month</td> <td style="text-align: center;">--</td> </tr> </tbody> </table> <p>As specified in 40 CFR 136.3.</p> <p>² Visual observations shall note the presence or absence of floatable materials of sewage origin. Observations of wind (direction and speed), weather (e.g., cloudy, sunny, or rainy), and tidal conditions (e.g., high or low tide) shall be recorded. Observations of water color, discoloration, oil and grease, turbidity, odor, materials of sewage origin in the water or on the beach shall be recorded. These observations shall be recorded whenever a sample is collected. Further, the nature and extent of primary contact recreation use in federal waters must be noted and reported.</p> <p>³ <u>Shall be monitored at the same discrete depths specified for bacterial monitoring in Table E-1.</u></p>	Parameter	Units	Sample Type	Minimum Sampling Frequency			Required Analytical Test Method	Offshore Stations	Kelp Stations	Shoreline Stations	Temperature	°C	Profile	1/Quarter	5/Month	--	1	Salinity	ppt	Profile	1/Quarter	5/Month	--	1	Dissolved Oxygen	mg/L	Profile	1/Quarter	5/Month	--	1	Light Transmittance	%	Profile	1/Quarter	5/Month	--	1	Chlorophyll a	m	Profile	1/Quarter	5/Month	--	1	pH	units	Profile	1/Quarter	5/Month	--	1	Ammonium (NH ₄ ⁺)	mg/L	Profile	1/Quarter	5/Month	--	1,3	Visual Observations ²	--	Visual	1/Quarter	5/Month	5/Month	--
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4	F-47	VI.D.X.1.a (Fact Sheet)	<p><i>Based on Comment 14 (d):</i></p> <p>A grid of 36 offshore stations is monitored quarterly and 8 kelp bed stations are monitored five times per month for the following parameters: salinity, temperature, density, pH, transmissivity, dissolved oxygen, ammonium (NH₄⁺), and chlorophyll a. These parameters are measured throughout the entire water column, <u>with the exception of ammonium, which shall be monitored at the same discrete depths specified for bacterial monitoring.</u></p>																																																																		

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5	E-30 and E-31	Table E-9 (MRP)	<p><i>Based on Comment 14 (e): Table E-9 replaced with the table below.</i></p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: center;"><u>Reports</u></th> <th style="text-align: center;"><u>Report Period</u></th> <th style="text-align: center;"><u>Report Due</u></th> </tr> </thead> <tbody> <tr> <td style="vertical-align: top;"> <u>MONTHLY REPORTS</u> <u>Influent and effluent</u> <u>Solids removal/disposal</u> <u>Receiving water quality</u> <u>Tijuana cross-border emergency connection (when flowing)</u> </td> <td style="text-align: center; vertical-align: top;"><u>Monthly</u></td> <td style="text-align: center; vertical-align: top;"><u>By the 1st day of 2nd following month (e.g., March 1 for January)</u></td> </tr> <tr> <td style="vertical-align: top;"> <u>QUARTERLY REPORTS</u> <u>Sludge analysis</u> </td> <td style="text-align: center; vertical-align: top;"> <u>January-March</u> <u>April-June</u> <u>July-September</u> <u>October-December</u> </td> <td style="text-align: center; vertical-align: top;"> <u>June 1</u> <u>September 1</u> <u>December 1</u> <u>March 1</u> </td> </tr> <tr> <td style="vertical-align: top;"> <u>SEMI-ANNUAL REPORTS</u> <u>Pretreatment report</u> </td> <td style="text-align: center; vertical-align: top;"> <u>January-June</u> <u>July-December</u> </td> <td style="text-align: center; vertical-align: top;"> <u>September 1</u> <u>March 1</u> </td> </tr> <tr> <td style="vertical-align: top;"> <u>ANNUAL REPORTS</u> <u>Pretreatment report</u> <u>Sludge analysis</u> <u>QA report</u> <u>Flow measurement</u> <u>Outfall inspection</u> <u>Receiving waters monitoring</u> <u>Kelp report</u> </td> <td style="text-align: center; vertical-align: top;"><u>January-December</u></td> <td style="text-align: center; vertical-align: top;"> <u>April 1</u> <u>April 1</u> <u>April 1</u> <u>July 1</u> <u>July 1</u> <u>July 1</u> <u>October 1</u> </td> </tr> </tbody> </table>	<u>Reports</u>	<u>Report Period</u>	<u>Report Due</u>	<u>MONTHLY REPORTS</u> <u>Influent and effluent</u> <u>Solids removal/disposal</u> <u>Receiving water quality</u> <u>Tijuana cross-border emergency connection (when flowing)</u>	<u>Monthly</u>	<u>By the 1st day of 2nd following month (e.g., March 1 for January)</u>	<u>QUARTERLY REPORTS</u> <u>Sludge analysis</u>	<u>January-March</u> <u>April-June</u> <u>July-September</u> <u>October-December</u>	<u>June 1</u> <u>September 1</u> <u>December 1</u> <u>March 1</u>	<u>SEMI-ANNUAL REPORTS</u> <u>Pretreatment report</u>	<u>January-June</u> <u>July-December</u>	<u>September 1</u> <u>March 1</u>	<u>ANNUAL REPORTS</u> <u>Pretreatment report</u> <u>Sludge analysis</u> <u>QA report</u> <u>Flow measurement</u> <u>Outfall inspection</u> <u>Receiving waters monitoring</u> <u>Kelp report</u>	<u>January-December</u>	<u>April 1</u> <u>April 1</u> <u>April 1</u> <u>July 1</u> <u>July 1</u> <u>July 1</u> <u>October 1</u>
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6	51	VII.I.2.e.ii	<p><i>Based on Comment 14 (i):</i></p> <p>For all bacterial analyses, sample dilutions should be performed so the range of values extends <u>as follows:</u></p> <ul style="list-style-type: none"> • <u>2 to 16,000/100ml colony-forming units (CFU) for total coliforms</u> • <u>2 to 12,000/100ml CFU for fecal coliforms</u> • <u>2 to 12,000/100ml CFU for enterococci</u> <p>from 2 to 16,000 CFU (colony-forming units).—The detection methods used for each analysis shall be reported with the results of the analysis. Detection methods used for coliform (total and fecal) shall be those presented in Table 1A of 40 CFR 136, unless alternate methods have been approved in advance by USEPA, pursuant to 40 CFR 136. Detection methods used for enterococcus shall be those presented in USEPA publication EPA 600/4-85/076, <i>Test Methods for Escherichia coli and Enterococci in Water by Membrane Filter Procedure</i>, listed under 40 CFR 136, or any improved method determined by the Regional Water Board or USEPA to be appropriate.</p>																				
7	B-1	Attachment B	<p><i>Based on Comment 14 (l): Attachment B replaced with the bathymetric map attached to the Errata Sheet.</i></p>																				
8	E-4 to E-6	Table E-1	<p><i>Based on Comment 14 (m): The F station locations have been updated in Table E-1 with the information below.</i></p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: center;">Discharge Point Name</th> <th style="text-align: center;">Monitoring Location Name</th> <th style="text-align: center;">Latitude</th> <th style="text-align: center;">Longitude</th> <th style="text-align: center;">Depth (m)</th> </tr> </thead> <tbody> <tr> <td colspan="5" style="text-align: center;">OFFSHORE MONITORING STATIONS</td> </tr> <tr> <td style="text-align: center;">--</td> <td style="text-align: center;">F-001</td> <td style="text-align: center;">32.637683</td> <td style="text-align: center;">-117.240316</td> <td style="text-align: center;">18¹</td> </tr> <tr> <td style="text-align: center;">--</td> <td style="text-align: center;">F-002</td> <td style="text-align: center;">32.756966</td> <td style="text-align: center;">-117.272733</td> <td style="text-align: center;">18¹</td> </tr> </tbody> </table>	Discharge Point Name	Monitoring Location Name	Latitude	Longitude	Depth (m)	OFFSHORE MONITORING STATIONS					--	F-001	32.637683	-117.240316	18 ¹	--	F-002	32.756966	-117.272733	18 ¹
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ERRATA #	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)				
			--	F-003	32.781833	-117.272416	18 ¹
			--	F-004	32.594533	-117.26875	60 ²
			--	F-005	32.611683	-117.26965	60 ²
			--	F-006	32.630833	-117.2736	60 ²
			--	F-007	32.651134	-117.279994	60 ²
			--	F-008	32.67215	-117.283	60 ²
			--	F-009	32.68555	-117.286316	60 ²
			--	F-010	32.705419	-117.290658	60 ²
			--	F-011	32.725544	-117.294632	60 ²
			--	F-012	32.746583	-117.302066	60 ²
			--	F-013	32.765383	-117.3072	60 ²
			--	F-014	32.781559	-117.311423	60 ²
			--	F-015	32.5941	-117.28645	80 ³
			--	F-016	32.611833	-117.290066	80 ³
			--	F-017	32.630016	-117.294166	80 ³
			--	F-018	32.649766	-117.298333	80 ³
			--	F-019	32.66785	-117.306833	80 ³
			--	F-020	32.685416	-117.310966	80 ³
			--	F-021	32.7038	-117.318687	80 ³
			--	F-022	32.72273	-117.320902	80 ³
			--	F-023	32.741883	-117.330416	80 ³
			--	F-024	32.761216	-117.33645	80 ³
			--	F-025	32.77895	-117.343583	80 ³
			--	F-026	32.593766	-117.3122	98 ⁴
			--	F-027	32.611783	-117.321383	98 ⁴
			--	F-028	32.629287	-117.323721	98 ⁴
			--	F-029	32.647815	-117.32493	98 ⁴
			--	F-030	32.66567	-117.32483	98 ⁴
			--	F-031	32.684668	-117.328353	98 ⁴
			--	F-032	32.701416	-117.334166	98 ⁴
			--	F-033	32.720466	-117.339916	98 ⁴

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			--	F-034	32.7389	-117.349366	98 ⁴																																													
			--	F-035	32.7577	-117.363383	98 ⁴																																													
			--	F-036	32.776783	-117.374566	98 ⁴																																													
9	E-7	III.A (MRP)	<p><i>Based on Comment 14 (n): Text in Section III.A and the portion of Table E-2 shown below has been updated as follows.</i></p> <p>Monitoring Location INF-001 and EMG-001 Influent monitoring is required to determine the effectiveness of pretreatment and non-industrial source control programs, to assess the performance of treatment facilities, and to evaluate compliance with effluent limitations. As such, influent monitoring results must accurately characterize raw wastewater from the entire service area of the treatment facilities, unaffected by in-plant return or recycle flows or the addition of treatment chemicals. Influent monitoring shall be conducted at INF-001 and EMG-001 (when flow is present) as shown in the table below.</p> <p>Table E-2. Influent and Emergency Connection Monitoring at INF-001 and EMG-001</p> <table border="1"> <thead> <tr> <th>Parameter</th> <th>Units</th> <th>Sample Type</th> <th>Minimum Sampling Frequency</th> <th>Required Analytical Test Method</th> </tr> </thead> <tbody> <tr> <td>Flow rate</td> <td>MGD</td> <td>recorder/totalizer</td> <td>Continuous</td> <td></td> </tr> <tr> <td>Biochemical Oxygen Demand (5-day @20 °C) (BOD₅)</td> <td>mg/L</td> <td>24-hr composite</td> <td><u>1/Day at INF-001</u> <u>1/Week at EMG-001</u></td> <td>1</td> </tr> <tr> <td>Volatile Suspended Solids</td> <td>mg/L</td> <td>24-hr composite</td> <td><u>1/Day at INF-001</u> <u>1/Week at EMG-001</u></td> <td></td> </tr> <tr> <td>Total Dissolved Solids (TSS)</td> <td>mg/L</td> <td>24-hr composite</td> <td><u>1/Day at INF-001</u> <u>1/Week at EMG-001</u></td> <td></td> </tr> <tr> <td>Temperature</td> <td>°C</td> <td>grab</td> <td><u>1/Day at INF-001</u> <u>1/Week at EMG-001</u></td> <td></td> </tr> <tr> <td>Floating Particulates</td> <td>mg/L</td> <td>24-hr composite</td> <td><u>1/Day at INF-001</u> <u>1/Week at EMG-001</u></td> <td></td> </tr> <tr> <td colspan="5" style="text-align: center;">TABLE A PARAMETERS</td> </tr> <tr> <td>Oil and Grease</td> <td>mg/L</td> <td>grab</td> <td><u>1/Day at INF-001</u> <u>1/Week at EMG-001</u></td> <td></td> </tr> </tbody> </table>					Parameter	Units	Sample Type	Minimum Sampling Frequency	Required Analytical Test Method	Flow rate	MGD	recorder/totalizer	Continuous		Biochemical Oxygen Demand (5-day @20 °C) (BOD ₅)	mg/L	24-hr composite	<u>1/Day at INF-001</u> <u>1/Week at EMG-001</u>	1	Volatile Suspended Solids	mg/L	24-hr composite	<u>1/Day at INF-001</u> <u>1/Week at EMG-001</u>		Total Dissolved Solids (TSS)	mg/L	24-hr composite	<u>1/Day at INF-001</u> <u>1/Week at EMG-001</u>		Temperature	°C	grab	<u>1/Day at INF-001</u> <u>1/Week at EMG-001</u>		Floating Particulates	mg/L	24-hr composite	<u>1/Day at INF-001</u> <u>1/Week at EMG-001</u>		TABLE A PARAMETERS					Oil and Grease	mg/L	grab	<u>1/Day at INF-001</u> <u>1/Week at EMG-001</u>	
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			Total Suspended Solids	mg/L	24-hr composite	1/Day <u>at INF-001</u> 1/Week <u>at EMG-001</u>		
			Settleable Solids	ml/L	grab	1/Day <u>at INF-001</u> 1/Week <u>at EMG-001</u>		
			Turbidity	NTU	grab	1/Day <u>at INF-001</u> 1/Week <u>at EMG-001</u>		
			pH	units	grab	1/Day <u>at INF-001</u> 1/Week <u>at EMG-001</u>		
10	16	Table 10	<i>Based on Comment 14 (o): The portion of Table 10 shown below has been updated as follows.</i>					
			Chromium, Total Recoverable (III) ²	µg/L	--	--	--	3.9E+07
				lbs/day	--	--	--	6.7E+07
11	E-8	Table E-2 (MRP)	<i>Based on Comment 14 (o): The portion of Table E-2 shown below has been updated as follows.</i>					
			Chromium (III), Total Recoverable ²	µg/L	24-hr composite	1/Month	1	
12	E-11	Table E-3 (MRP)	<i>Based on Comment 14 (o):</i>					
			Chromium (III) ²	µg/L	24-hr composite	1/Month	1	
13	F-36	Table F-16 (Fact Sheet)	<i>Based on Comment 14 (o):</i>					

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			lbs/day	--	--	--
						6.7E+07
14	E-14	V.A.1 (MRP)	<p><i>Based on Comment 14 (p):</i></p> <p>The Discharger shall conduct monthly chronic toxicity tests on 24-hour composite effluent samples. Once each calendar year, at a different time of year from the previous years, the Discharger shall split a 24-hour composite effluent sample and concurrently conduct three toxicity tests using a fish, an invertebrate, and an alga species; the Discharger shall then continue to conduct routine monthly toxicity testing using the single, most sensitive species. <u>For the initial three suites of chronic toxicity tests, the Discharger shall split a 24-hour composite effluent sample and concurrently conduct toxicity tests using a fish, an invertebrate, and an alga species. After the initial screening period, the Discharger shall conduct routine monthly toxicity testing using the most sensitive species. Every other year, the Discharger shall re-screen at a different time from the prior years. Re-screening can be limited to one month, if results are the same as the previous three-month screening. However, if results of the re-screening are different, then the Discharger shall conduct two additional months of re-screening to determine the most sensitive species and then conduct routine monthly toxicity testing using the most sensitive species.</u></p>			
15	E-17	V.A.3.j (MRP)	<p><i>Based on Comment 14 (r):</i></p> <p>pH drift during the toxicity test may contribute to artifactual toxicity when pH-dependent toxicants (e.g., ammonia, metals) are present in an effluent. To determine whether or not pH drift during the toxicity test is contributing to artifactual toxicity, the Discharger shall conduct three sets of parallel toxicity tests, in which the pH of one treatment is controlled at the pH of the effluent and the pH of the other treatment is not controlled, as described in Section 11.3.6.1 of the test methods manual, <i>Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms</i> (EPA/821/R-02/013, 2002). Toxicity is confirmed to be artifactual and due to pH drift when no toxicity above the chronic toxicity effluent limit is observed in the treatments controlled at the pH of the effluent. If toxicity is</p>			

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			<p>confirmed to be artifactual and due to pH drift, then, following written approval by the Executive Officer and USEPA, the Discharger may use the procedures outlined in Section 11.3.6.2 of the test methods manual to control sample pH during the toxicity test.</p>
16	E-20	V.B.3 (MRP)	<p><i>Based on Comment 14 (r): The text below has been deleted from Section V.A.3.j in the MRP and added as Section V.B.3.k in the MRP.</i></p> <p><u>k. pH drift during the toxicity test may contribute to artifactual toxicity when pH-dependent toxicants (e.g., ammonia, metals) are present in an effluent. To determine whether or not pH drift during the toxicity test is contributing to artifactual toxicity, the Discharger shall conduct three sets of parallel toxicity tests, in which the pH of one treatment is controlled at the pH of the effluent and the pH of the other treatment is not controlled, as described in Section 11.3.6.1 of the test methods manual, <i>Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms</i> (EPA/821/R-02/013, 2002). Toxicity is confirmed to be artifactual and due to pH drift when no toxicity above the toxicity effluent limit is observed in the treatments controlled at the pH of the effluent. If toxicity is confirmed to be artifactual and due to pH drift, then, following written approval by the Executive Officer and USEPA, the Discharger may use the procedures outlined in Section 11.3.6.2 of the test methods manual to control sample pH during the toxicity test.</u></p>
17	E-17	V.A.4.a (MRP)	<p><i>Based on Comment 14 (s):</i></p> <p>A full laboratory report for all toxicity testing shall be submitted as an attachment to the DMR for the month in which the toxicity test was conducted and shall also include: the toxicity test results as in NOEC; $TU_c = 100/NOEC$; and as EC_{25} (or IC_{25}); and $TU_c = 100/EC_{25}$ (or IC_{25})—reported according to the test methods manual chapter on report preparation and test review; the dates of sample collection and initiation of each toxicity test; all results for effluent parameters monitored concurrently with the toxicity test(s); and progress reports on accelerated testing and TRE/TIE investigations.</p>

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18	E-18	V.B.1 (MRP)	<p><i>Based on Comment 14 (t):</i></p> <p>The Discharger shall conduct semi-annual acute toxicity tests on 24-hour composite effluent samples. Once each calendar year, at a different time of year from the previous years, the Discharger shall split a 24-hour composite effluent sample and concurrently conduct two toxicity tests using a fish and an invertebrate species; the Discharger shall then continue to conduct routine semi-annual toxicity testing using the single, most sensitive species. <u>For the initial three suites of acute toxicity tests, the Discharger shall split a 24-hour composite effluent sample and concurrently conduct toxicity tests using a fish and an invertebrate. After the initial screening period, the Discharger shall conduct routine semi-annual toxicity testing using the most sensitive species. Every other year, the Discharger shall re-screen at a different time from the prior years. Re-screening can be limited to one month, if results are the same as the previous three-month screening. However, if results of the re-screening are different, then the Discharger shall conduct two additional months of re-screening to determine the most sensitive species and then conduct routine semi-annual toxicity testing using the most sensitive species.</u></p>
19	E-25	VIII.A.3 (MRP)	<p><i>Based on Comment 14 (u):</i></p> <p>Twice per year (January and July), sediment samples for benthic infauna community structure shall be collected from the offshore sediment monitoring locations specified in Table E-1, which consists of 12 primary stations and an additional 10 secondary stations. Two replicate samples shall be taken using a 0.1 square meter modified Van Veen grab sampler. These samples shall be separate from those collected for grain size and chemistry. The samples shall be sieved using a 1.0-mm mesh screen. The benthic organisms retained on the sieve shall be fixed in 105 percent buffered formalin and transferred to <u>at least</u> 70 percent ethanol within two to seven days for storage. All retained benthic infauna organisms shall be counted and identified to as low a taxon as possible. This enumeration and identification of organisms continues to use the historical database developed by the Discharger.</p>

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			<p>Analysis of benthic community structure shall include determination of the number of species, number of individuals per species, and total numerical abundance present. The following parameters shall be calculated for each grab sample and summarized by station as appropriate summarized for each station:</p> <p>a. Average number of species (species richness) per 0.1 m²; b. Total number of species per station; c. Total numerical abundance; d. Infaunal trophic index (ITI); e. Benthic response index (BRI); f. Swartz' 75% dominance index; g. Shannon-Weiner's diversity index (H'); and h. Pielou evenness (J')</p> <p><u>a. Number of species per 0.1m² (species richness);</u> <u>b. Total (cumulative) number of species per station;</u> <u>c. Total numerical abundance;</u> <u>d. Benthic response index (BRI);</u> <u>e. Swartz's 75% dominance index;</u> <u>f. Shannon's diversity index (H'); and</u> <u>g. Pileou's evenness index (J').</u></p>
20	E-26	VIII.A.4 (MRP)	<p><i>Based on Comment 14 (v):</i></p> <p>Chemical analyses of fish tissues shall be performed annually on target species collected at or near the trawl and rig fishing stations. The various stations are classified into zones for the purpose of collecting sufficient numbers of fish for tissue analyses. Trawl Zone 1 represents the nearfield zone, defined as the area within a 1-km radius of stations SD-010 and/or SD-012; Trawl Zone 2 is considered the northern farfield zone, defined as the area within a 1-km radius of stations SD-013 and/or SD-014; Trawl Zone 3 represents the LA-5 disposal site zone, and is defined as the area centered within a 1-km radius of station SD-008; Trawl Zone 4 is</p>

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			<p>considered the southern farfield zone, and is defined as the area centered within a 1-km radius of station SD-007. The two rig fishing stations also represent two distinct zones. Rig #Fishing zZone 1 is the nearfield area centered within a 1-km radius of Station RF-001; Rig #Fishing zZone 2 is considered the farfield area centered within a 1-km radius of station RF-002. <u>There are no depth requirements for these six zones with regards to the collection of fishes for tissue analysis.</u></p> <p>Liver tissues shall be analyzed semiannually (January and July) <u>annually (i.e., during October)</u> from fishes collected in each of the above four trawl zones. Each trawl station may be trawled up to a maximum of five times in order to acquire sufficient numbers of fish for composite samples within a zone; trawls subsequent to the initial community trawl discussed above (i.e., trawls 2-5/site) may occur anywhere within a defined zone. <u>No more than a maximum of five 10-minute (bottom time) trawls shall be required per zone in order to acquire sufficient numbers of fish for composite samples; these trawls may occur anywhere within a defined zone.</u> Three replicate composite samples shall be prepared from each trawl zone, with each composite consisting of tissues from as least three <u>individual</u> fish of the same species collected within a zone. These liver tissues shall be analyzed for the presence <u>and concentrations of</u> lipids, PCB (congeners), chlorinated pesticides, and the <u>following three</u> metals: mercury, arsenic and selenium. The species <u>of fish</u> targeted for <u>tissue</u> analysis at <u>from</u> the trawl sites shall be primarily flatfish, and include <u>including, but not limited to,</u> the longfin sanddab (<i>Citharichthys xanhostigma</i>) and the Pacific sanddab (<i>Citharichthys sordidus</i>). If sufficient numbers of these primary target species are not present in a zone, secondary candidate species such as other flatfish or rockfish may be collected as necessary.</p> <p>Rig fishing shall be performed <u>Muscle tissues shall be analyzed</u> annually (<u>i.e., during</u> October) to monitor the uptake of pollutants in fish species which are <u>from fishes collected in each of the above two rig fishing zones in order to monitor the uptake of pollutants in species and tissues that are</u> consumed by humans. These fish species shall be representative of those caught by recreational and/or commercial fishery activities in the region. All fish shall be collected by hook and line or by setting baited lines or traps within the two <u>rig fishing</u> zones described above. The species targeted for analysis at the rig fishing sites shall be primarily rockfish, <u>which may include, but are not limited to,</u> and include the vermilion</p>

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			<p>rockfish (<i>Sebastes miniatus</i>) and the copper rockfish (<i>Sebastes caurinum</i>). If sufficient numbers of these primary fish-species are not present <u>or cannot be caught in a particular zone</u>, other <u>secondary target</u> species (e.g., rockfish, scorpionfish) may be collected <u>and analyzed</u> as necessary. Three replicate composite samples of the target species shall be obtained from each zone, with each composite consisting of a minimum of three individual fish. Muscle tissues shall be removed from the composites and chemically analyzed for the presence <u>and concentrations</u> of lipids, PCB (congeners), chlorinated pesticides, and the <u>following nine metals</u>: arsenic, cadmium, chromium, copper, lead, mercury, selenium, tin and zinc.</p>
21	F-48	VI.D.1.d (Fact Sheet)	<p><i>Based on Comment 14 (v):</i></p> <p>Twice each year, in January and July, eEpibenthic trawls at four trawl zone stations are used to assess the structure of demersal fish and megabenthic invertebrate communities and to evaluate compliance with narrative water quality standards in the Ocean Plan. Semiannually, in January and July, cChemical analyses of fish tissues are performed on target species collected at the four trawl zone stations and two rig fishing stations. Species targeted for analysis are selected based on their ecological and/or commercial importance. Liver tissue is monitored at trawl stations to assess general fish health. Muscle tissue is monitored at rig fishing stations annually, in October, to assess the uptake of pollutants in fish species commonly consumed by humans in the region. Fish tissues are monitored for lipids, metals, PCBs, and chlorinated pesticides.</p>
22	E-30	IX.B.1 (MRP)	<p><i>Based on Comment 14 (x):</i></p> <p>At any time during the term of this permit, the State or Regional Water Board may notify the Discharger to electronically submit Self-Monitoring Reports (SMRs) using the State Water</p>

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			<p>Board's California Integrated Water Quality System (CIWQS) Program Web site (http://www.waterboards.ca.gov/ciwqs/index.html). Until such notification is given, the Discharger shall submit hard copy SMRs. <u>For this purpose, a hard copy signed penalty of perjury statement accompanying a CD with a single file in PDF format (including the certification specified in Section V.B. 5 of Attachment D) shall qualify as a hard copy SMR.</u> The CIWQS Web site will provide additional directions for SMR submittal in the event there will be service interruption for electronic submittal.</p>
23	-	-	<p><i>Based on Comment 14 (aa): Typographical errors and other minor corrections to the wording in the tentative order have been or will be made prior to sending out the final version.</i></p>
24	43	VI.C.5.c.v	<p><i>Based on a recommendation by the United States Environmental Protection Agency (USEPA) Region 9 Clean Water Act Compliance Office:</i></p> <p>By April March 1 of each year, the Discharger shall submit an annual report to the Regional Water Board; USEPA Region 9; the State Water Board, Division of Water Quality, Regulations Unit; and the San Diego County Department of Health Services, Hazardous Materials Division, describing its pretreatment activities over the previous calendar year.</p>
25	C-1	Attachment C	<p><i>Attachment C figures replaced with schematic diagrams (including disinfection) attached to the Errata Sheet.</i></p>

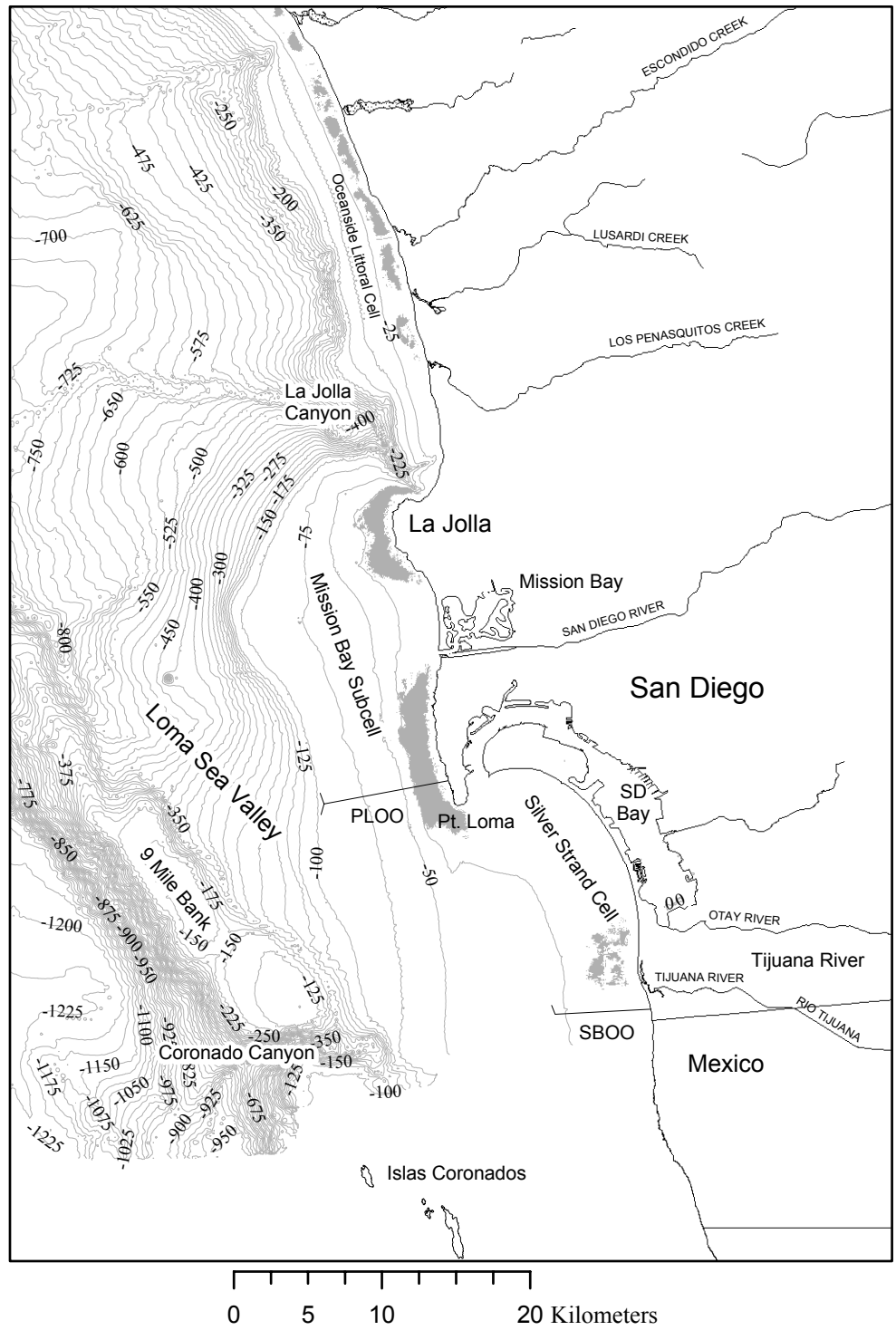
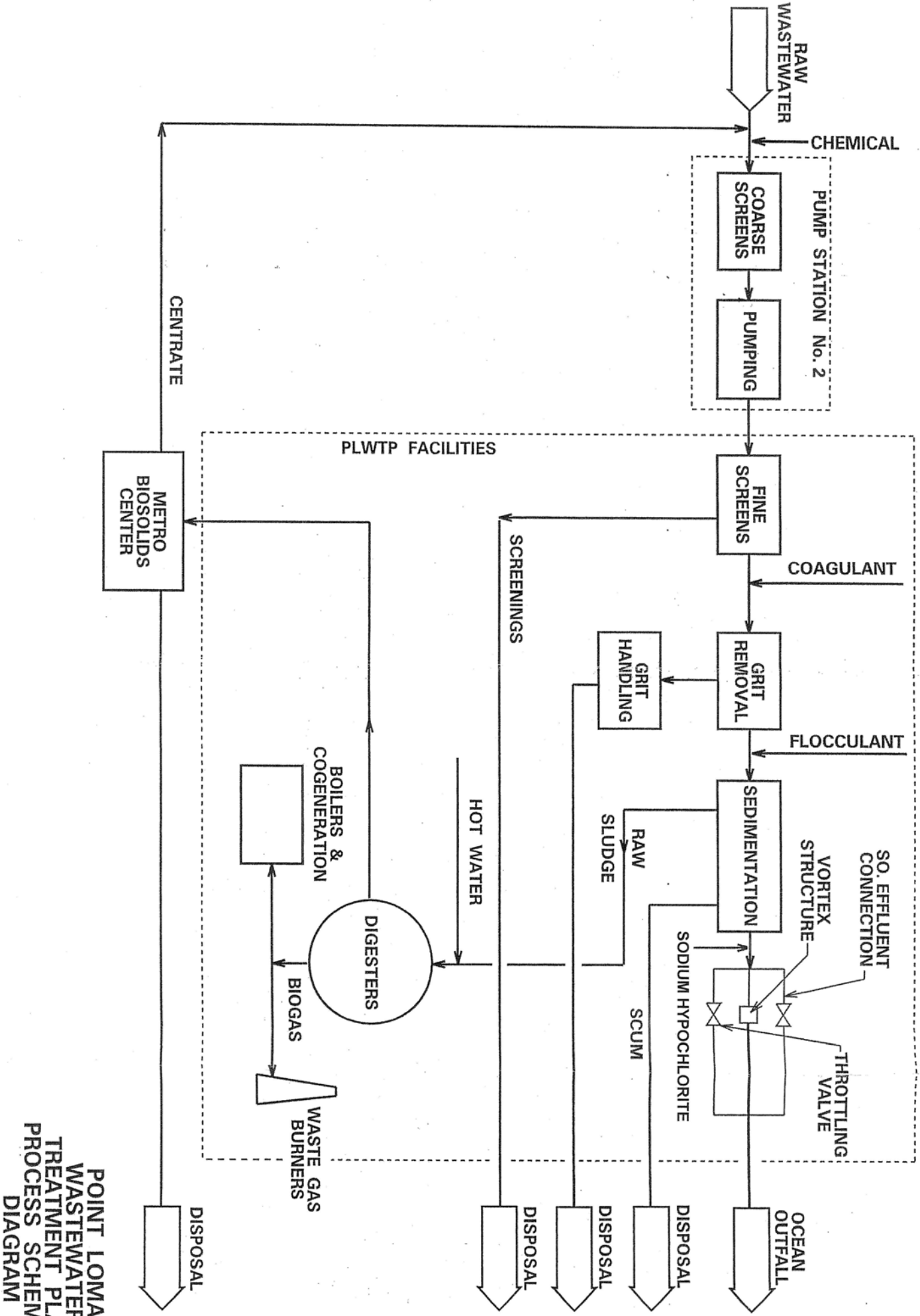
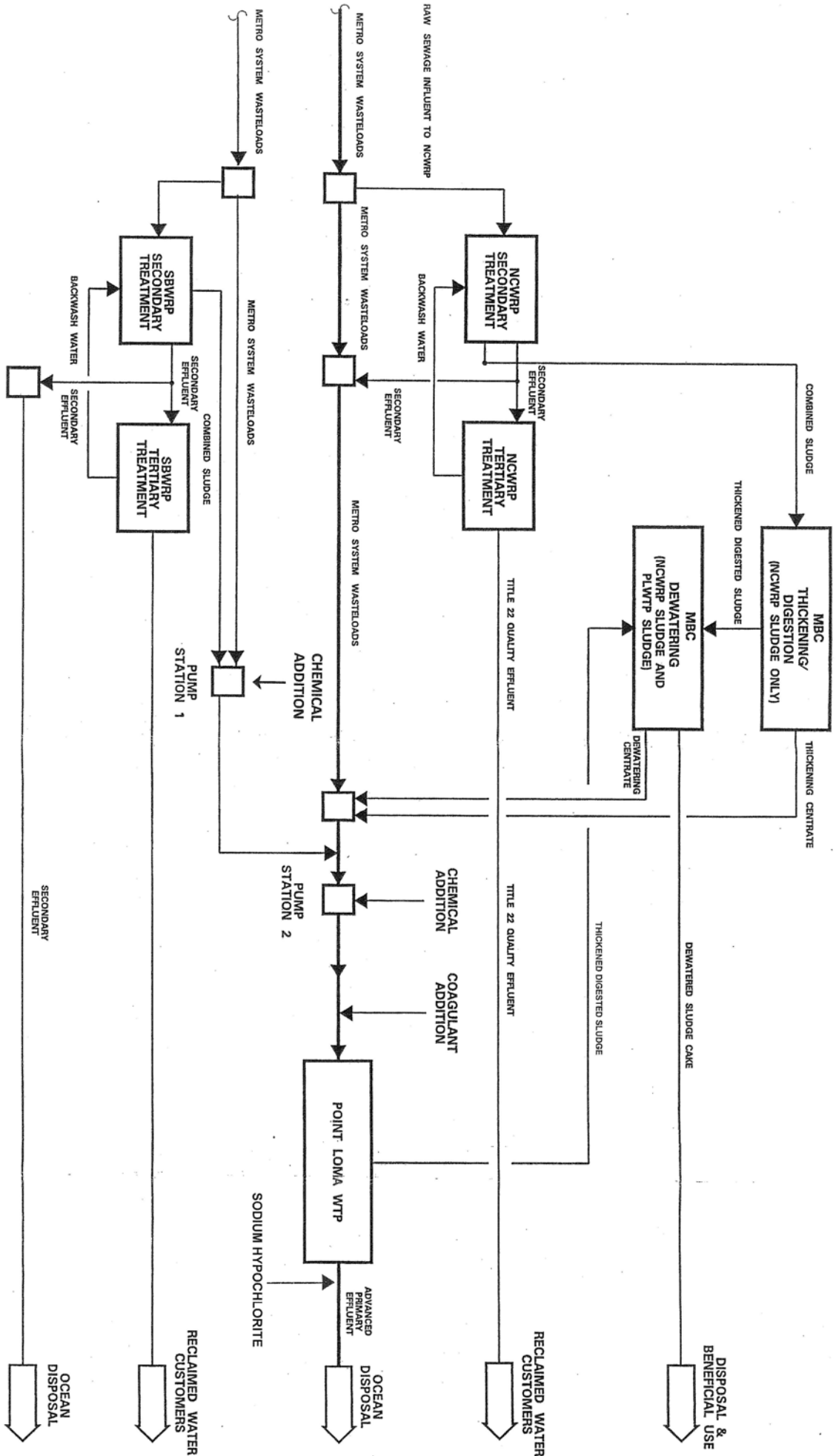


Figure 1. Map of marine shelf of San Diego County. Bathymetric units are meters. Locations of littoral cells, submarine canyons, outfalls (PLOO and SBOO), rivers, and Kelp Forests (shaded areas close to shore) are indicated.



POINT LOMA
WASTEWATER
TREATMENT PLANT
PROCESS SCHEMATIC
DIAGRAM



METRO SYSTEM PROCESS SCHEMATIC