Changes in Forced Expiratory Flow Due to Air Pollution From Fireworks

Preliminary Report

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Spirometry in humans and air sampling have been performed during a brief but intense bout of air pollution due to exploding fireworks on New Year's Eve in Honolulu, Hawaii. An integrating nephelometer recorded a peak concentration of respirable particles in excess of 3.8 mg/m^3. X-ray dispersive analysis, combined with scanning electron microscopy of particles collected with an Anderson cascade impactor, indicated that most of the small particles were probably crystals of KCl. Conditions were ideal for a possible SO_2–KCl aerosol synergism which may have led to the measured changes in maximal midexpiratory flow ($FEV_{25-75\%}$). Two male subjects with a history of chronic respiratory disease experienced an average decrease of 26% in $FEV_{25-75\%}$ when compared to that measured the previous night. Three normal males experienced a 4.7% decrease approaching statistical significance ($0.1 > P > 0.05$), but the overall difference in $FEV_{25-75\%}$ in healthy male and female subjects combined was not statistically significant. Thus, while susceptible people may be measurably affected, the general population of Honolulu probably experienced little, if any, change in $FEV_{25-75\%}$ with the air pollution levels reported here.

INTRODUCTION

Each New Year's Eve the people of Honolulu are exposed to a brief but intense bout of air pollution characterized by increased SO_2 and particulate matter of unknown composition as a result of the legal burning of fireworks on private property. Especially under conditions of low wind velocity, the smoke from the possibly more than $1 million worth of fireworks markedly decreases the visibility in Honolulu (Bach *et al.*, 1972). In a recent survey of 175 patients suffering from chronic respiratory disease, 68% reported that they took special precautions at home to avoid the smoke and 73% found it more difficult to breathe during the fireworks.[1] In the only published study of possible adverse health effects due to one of these severe air pollution episodes, Bach *et al.* (1972)

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[1] Houghtby, G. Director, Health Education, Hawaii Thoracic Society. Personal Communication, 1974.

reported that while mortality data were unchanged, five Honolulu hospitals experienced a 113% increase in emergency room visits due to respiratory distress in people with a history of chronic respiratory disease.

Other than hospital admissions, no data exist concerning possible adverse effects of fireworks air pollution on people with or without a history of chronic respiratory disease. The present study was performed on the New Year 1972–1973 to determine if acute changes in forced expiratory flow occurred in healthy residents of Honolulu exposed to the fireworks smoke. In addition, the instantaneous respirable particle concentration and 18 hour average concentration, size distribution, and composition of the particulate matter produced by burning fireworks are presented, together with selected scanning electron microscope (SEM) photomicrographs of the particles.

MATERIALS AND METHODS

Pulmonary Function

Some physical characteristics of the subjects used in this study are presented in Table 1. Of the nine acceptable subjects, all except one smoker (subject 5) were near normal limits on control tests of forced vital capacity (FEV), one sec-

TABLE 1
PHYSICAL CHARACTERISTICS AND RESPIRATORY DISEASE HISTORY OF SUBJECTS

Subject	Sex	Age	Height (meters)	Weight (kg)	Control measurements (Percent of predicted)[a]			History of adult respiratory disease and comments
					FEV	$FEV_{25-75\%}$	FEV_1	
1	M	38	1.58	61.4	104.9	82.8	94.0	Allergic asthma (Drixoral medication). Gas trapping in control spirogram.
2	M	27	1.82	79.5	108.8	128.2	94.2	Hospitalized for recurrent bronchitis at age 21. Gas trapping in control spirogram.
3	M	23	1.90	79.5	131.2	124.0	99.3	None
4	M	24	1.81	75.0	121.5	146.3	98.1	None, competitive swimmer.
5	M	44	1.72	70.0	107.2	62.5	82.9	None, smokes pipe and 1–10 cigarettes per day for 17 years.
6	F	26	1.90	68.2	114.0	118.6	99.1	None, childhood asthma.
7	F	18	1.52	42.3	95.7	—	100.0	None, childhood asthma.
8	F	26	1.70	56.8	99.4	111.2	94.3	None
9	F	27	1.73	56.8	118.5	165.0	113.0	None, smokes infrequently.

[a] Calculated from the following equations obtained from normal standing subjects (Altman and Dittmer, 1971): FEV: 0.052 Height—0.022 Age—3.60 (Males); 0.052 Height—0.018 Age—4.36 (Females); $FEV_{25-75\%}$: 5.85—0.0523 Age (Males); 5.63—0.0549 Age (Females); and FEV_1: 0.201 Height—0.400 Age + 129 (Males); 0.253 Age + 93 (Females).

ond forced expiratory volume (FEV_1), and maximal midexpiratory flow rate ($FEV_{25-75\%}$), measured on January 20, 1973. However, a detailed personal history filled out by each subject indicated that male subjects 1 and 2 had a history of chronic respiratory disease persisting to adulthood. Both of these subjects had control spirograms indicative of gas trapping at the end of forced expiratory maneuvers. Since none of the other subjects (3–9) exhibited marked gas trapping or a history of chronic respiratory disease, they were treated separately from subjects one and two.

Measurements of pulmonary function in the standing position were performed between 11:35 PM and 12:30 AM on three occasions. The first two took place in the open garage of a home in residential Manoa Valley (about two miles from the H-1 highway). This site is about 200 feet above sea level and is 80 feet above the valley floor. Two Collins spirometers (9 and 13 l) and the air sampling instruments were set up about 15 and 20 feet from the curb, respectively. Unfortunately, between the first control measurements on December 30 (called Control I) and the experimental measurements on December 31, a cold front passed over Hawaii and the air temperature (T_a) measured at the experimental site fell from 20 to 15°C. Therefore, we performed a second control experiment (Control II) at a T_a of 15°C well after the air pollution episode in order to separate possible temperature effects from air pollution effects. We used a large climate-controlled chamber (Forma Scientific, Inc.) to produce the 15°C air.

On all three testing occasions the subjects arrived at the measurement site at least 30 min before testing began. The subjects' activity before arrival was not restricted except to refrain from heavy drinking or exercise. No smoking was allowed for 30 min before testing began. Each subject performed three to six forced expiratory maneuvers into the same Collins spirometer on each occasion under the direct supervision of one of us. Encouragement was given to ensure that each expiratory maneuver began at total lung capacity and ended at residual volume, and the suggestions of Hyatt, Schilder, and Fry (1958) were followed. All of the curves obtained on each occasion were analysed for FEV, FEV at 1, 2 and 3 seconds, and $FEV_{25-75\%}$. The FEV measure used for each subject is the largest of the several recorded on each occasion and the $FEV_{1,2,3}$ are taken from the same curve and are expressed as % FEV. The $FEV_{25-75\%}$ is the maximal value obtained from several efforts so long as this FEV was at least 90% as large as the largest FEV recorded on that occasion. In exactly one-half of the FEV curves used in this paper, the maximal FEV curve also yielded the maximal $FEV_{25-75\%}$.

All gas volumes were measured at ambient temperature (T_a) and converted to BTPS assuming a body temperature of 37°C. Since T_a changed significantly between testing dates, we tested that the gas in the spirometer bells after several forced expirations at 20 and 15°C was at T_a, using small YSI thermistors in the bells. In both 15 and 20°C air, repeated rapid FEV maneuvers did not cause the bell air temperature to deviate significantly from T_a.

Air Sampling

The air-borne mass concentration of particulate matter was determined with an Andersen (nonviable) cascade impactor (Anderson 2000, Inc.). Air was sucked through the eight stage sampler at 1.5 ft³/minute (cfm) and the particles were

collected on preweighed thin aluminum foil disks 3.125 in. in diameter and 0.25 mil
in thickness. The foil disks were handled only with forceps and were weighed on a
Cahn electrobalance with a stated accuracy of ± 2 μg within the required weighing
range of 20 mg. After the gravimetric determination, the aluminum disks were cut
into small sections, mounted on blocks, and shadowed with gold or palladium for
observation with a Cambridge S_{24}-10 SEM. Of the some 400 impaction deposits per
stage, 4 to 5 deposits were scanned and representative areas were photographed.
The SEM is equipped with an EDAX X-ray dispersive analysis attachment which
allowed us to determine the partial elemental composition of the smoke particles.

Instantaneous air particulate concentrations were estimated with an in-
tegrating nephelometer (Meteorology Research, Inc.) which measures the con-
centration of "respirable" particles from 0.2 to 1.2 μm in diameter (roughly
comparable to stages 5, 6, and 7 of the Andersen sampler). The amplified
nephelometer output was continuously recorded on a strip chart recorder
(Hewlett-Packard, Model 680) with a chart speed of 0.07 in./minute. The air was
sampled at 10 cfm. The nephelometer was zeroed immediately before each run,
using filtered dry air, and the manufacturer's conversion was used to convert
scattering coefficient to average mass concentration. The relative accuracy of the
scattering coefficient is about 4% (Porch, Charlson and Radke, 1970), but we did
not attempt an error estimate of the absolute concentrations reported here.

RESULTS

Air Sampling

The time course of the air pollution due to fireworks on New Year's Eve as
measured by the nephelometer in Manoa Valley, Honolulu is shown in Fig. 1. At
8 PM the average mass concentration was already about 150 μg/m^3 due to sporadic
explosions of fireworks (Fig. 1,a). At about 11:35 PM, when we began the
pulmonary function tests, the concentration increased sharply, exceeding the 380
μg/m^3 level, and forcing the change of scale on the nephelometer (Fig. 1,b). During
the fifteen minutes before midnight the pollution level increased from less than 1
mg/m^3 to 3.8 mg/m^3 and continued beyond the nephelometer measurement
capability. Fifteen minutes after midnight the concentration fell low enough to be
measurable and continued to fall rapidly so that the lower scale could be used at
12:30 AM, by which time we had completed the pulmonary function tests. One
hour later the concentration of respirable particles in the air had returned to near
normal values for Honolulu. During the half hour around midnight when most of
our pulmonary function testing was done, the average concentration was at least
2.15 mg/m^3.

The 18 hour average concentration and size distribution of suspended par-
ticulate matter in the air on Control I and New Year's Eve is shown in Table 2.
The Control I run of the Andersen Sampler which started at 6 PM on December
30 and ended at 12 AM on December 31 yielded an average concentration of
20 μg/m^3, with a fairly even distribution among the stages except for stage 2 which
contributed relatively more than the others. The experimental run covering the
New Year's celebration which started at 8 PM on December 31 and ended at 1 PM
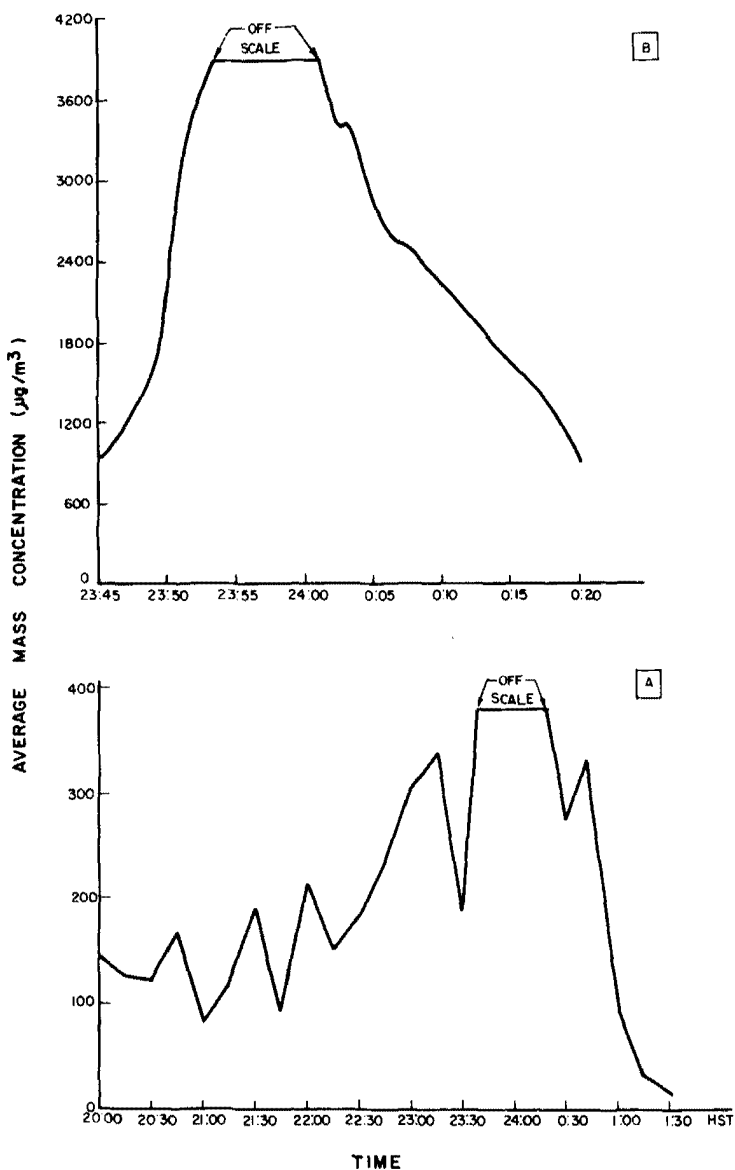on January 1, yielded a value of 113 μg/m^3. This increase of more than five times

FIG. 1. Fifteen (a) and five (b) minute average mass concentration of respirable particles measured on December 31, 1972–January 1, 1973 with an integrating nephelometer.

over the previous day's value was mainly due to a sharp increase in the smaller particles collected on stages 5, 6, and 7. This result, which tends to corroborate the nephelometer result, indicates that most of the air pollution due to fireworks is composed of particles between 0.3 and 2.0 μm (or smaller); these particles are known to penetrate and be retained by the lung (Dennis, 1971).

All stages of the Andersen samples collected on December 31–January 1 were viewed with the scanning electron microscope, but only selected results from stages 0, 2, 4, 6 and 7 are presented in Fig. 2. Stages 0, 2, and 4 (Fig. 2: a, b, c)

TABLE 2
Size Distribution of Suspended Particulate Matter in
Honolulu on the 1972–1973 New Year

Andersen impactor stage	Aerodynamic particle diameter by stage[a] (μm)	Weight (μg/m^3)	
		Sample 1 December 30–December 31	Sample 2 December 31–January 1
0	>8,8	2	5
1	5.6 –8.8	1	4
2	3.8 –5.6	7	7
3	2.7 –3.8	3	10
4	1.7 –2.7	2	7
5	0.84–1.7	1	14
6	0.52–0.84	2	40
7	0.34–0.52	2	28
	Total	20 μg/m^3	113 μg/m^3

[a] Normal sampler flow rate is 1 cfm. We used 1.5 cfm and estimated size distribution for an impaction efficiency of 50% by stage at this flow rate from a calibration chart provided by Carl H. Erickson, Director of Engineering, Anderson 2000, Inc.

show that most of the larger particles collected (about 28% by weight of the total collected) are unidentified fungal spores, and explains why stages 0–4 did not show a marked weight change from December 30 to December 31. In addition to the large biological particles, large crystals were seen in stages 2–4 (Fig. 2, b).

Most of the material collected on stages 5–7 was composed of crystals and unidentified amorphous material (Fig. 2: d, e). X-ray microanalysis of single crystals from stage 7 (Fig. 2, f) indicated the presence of four major energy peaks representing from left to right the following elements: aluminum, gold, chlorine, and potassium. The flash powder used in fireworks is composed of about 25% aluminum powder.[2] However, the particles are deposited on aluminum disks, so it is impossible to know how much of the aluminum peak is due to particulate aluminum from the air sample and how much is background due to the aluminum disk. The gold peak is due to coating the particles with gold. The chlorine and potassium peaks indicate that much of these crystals is KCl. However, energy emission counts from two different crystals from stage 7 (shown as solid bars and dots respectively in Fig. 2, f) indicate that the potassium and chlorine do not always appear in constant proportions, so that other salts of potassium may be present. The crystals were hygroscopic since they deliquesced in an atmosphere saturated with water vapor at 24°C.

Pulmonary Function

The results of the pulmonary function tests are shown in Table 3. Two subjects with a history of adult respiratory disease showed a significant decrease in

[2] Miller, D. C. Bureau of Compliance, Consumer Product Safety Commission. Washington, DC 20016. Personal Communication, 1973.
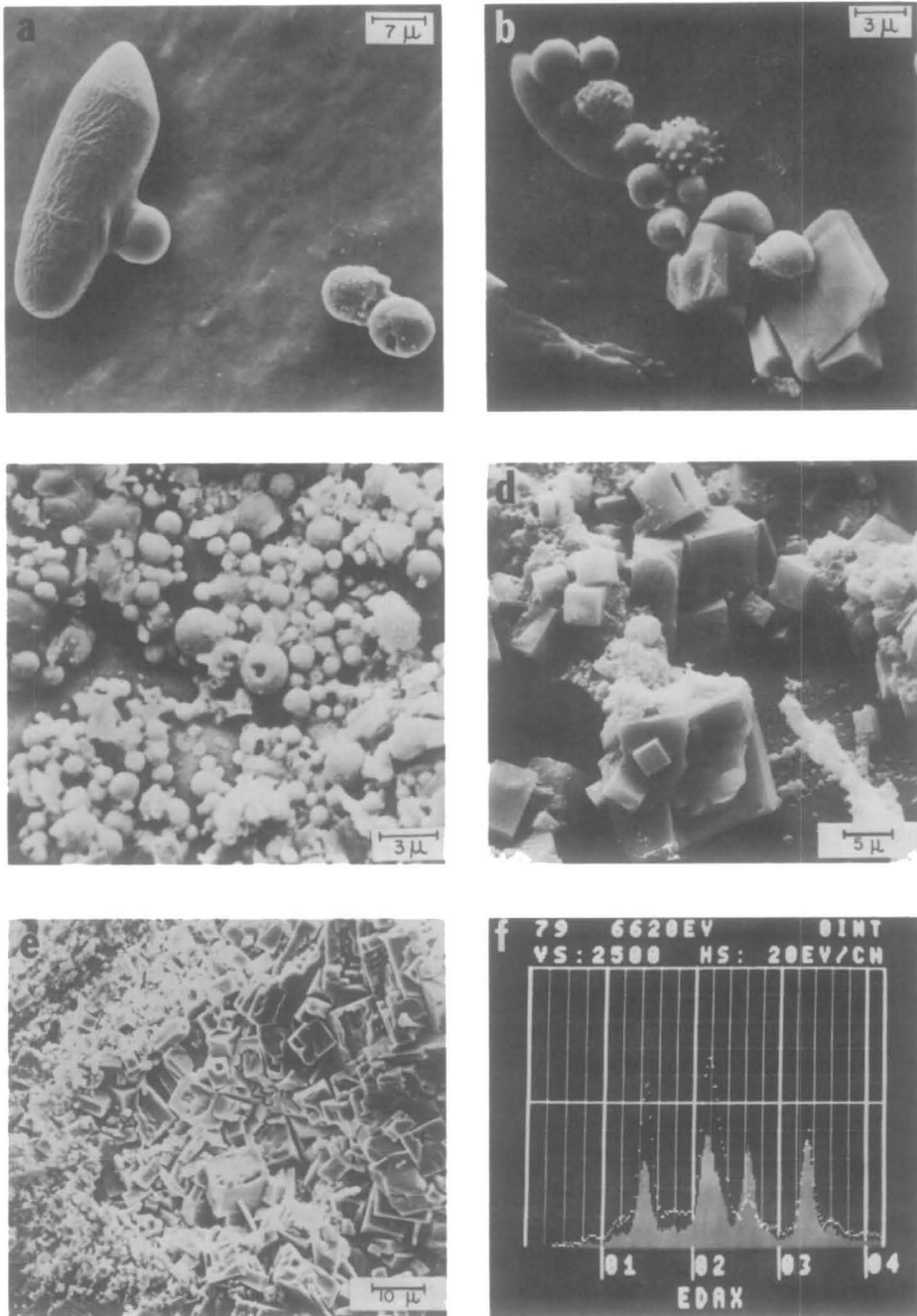
FIG. 2. Photographs of particles collected with an Andersen sampler in Honolulu on December 31, 1972—January 1, 1973, and viewed with a scanning electron microscope; a, × 1440; b, × 3240; c, × 3000; d, × 2000; e, × 1125. Fig. 2, f is an energy count from crystals shown in 2, e. See text for description.

TABLE 3

COMPARISON OF FEV MEASUREMENTS BEFORE AND DURING FIREWORKS

Subjects and analysis[a]	December 30–31, 1972			December 31–January 1, 1972–73		
	FEV (1)	FEV_1 (% FEV)	$FEV_{25-75\%}$ (1/sec)	FEV (1)	FEV_1 (% FEV)	$FEV_{25-75\%}$ (1/sec)
History of disease						
1	3.770	76.4	2.818	3.699	74.1	2.366
	3.754	77.0	2.423	3.504	74.8	2.297
2	4.560	77.0	5.108	4.318	75.6	3.290
	4.438	80.5	4.193	4.264	59.1	2.807
$\bar{x}$	4.131	77.7	3.636	3.946	70.9	2.690
$\bar{d}$	0.184	6.83	0.945			
P	$0.025 > P > 0.01$	$0.15 > P > 0.1$	$0.05 > P > 0.025$			
Normal males						
3	6.271	80.9	5.326	5.906	79.1	5.208
4	6.469	81.4	6.678	6.203	85.1	6.277
5	4.096	63.7	2.197	3.947	68.0	2.058
$\bar{x}$	5.612	75.33	4.734	5.352	77.40	4.514
$\bar{d}$	0.260	−2.07	0.219			
P	$0.025 > P > 0.01$	NS	$0.10 > P > 0.05$			
Normal females						
6	3.858	85.6	4.237	3.783	86.2	4.552
7	2.889	88.3	3.542	2.728	84.7	3.140
8	4.217	81.5	4.335	4.044	85.5	4.448
9	4.471	96.8	5.724	4.560	88.8	5.580
$\bar{x}$	3.859	88.05	4.460	3.779	86.3	4.430
$\bar{d}$	0.080	1.75	0.030			
P	$0.15 > P > 0.1$	NS	NS			
Combined normal males and females						
$\bar{x}$	4.610	82.6	4.577	4.453	82.5	4.466
$\bar{d}$	0.157	0.114	0.111			
P	$0.025 > P > 0.01$	NS	$0.15 > P > 0.1$			

[a] One-tailed paired design t test ($df = n - 1$); NS, $P < 0.15$; P values > 0.15 are indicated.

the two highest FEV and $FEV_{25-75\%}$ measurements between control and New Year's Eve. The FEV_1 did not show a significant change. $FEV_{2,3}$ were nearly identical on the two nights for all subjects and have not been included in the table.

Seven normal subjects had a significant reduction in the FEV, but no change in the FEV_1. The $FEV_{25-75\%}$ decreased but not to generally accepted levels of statistical significance. While each of the three normal males showed a reduction in $FEV_{25-75\%}$ which approached statistical significance ($0.10 > P > 0.05$), two of the four females showed an increase. Among the normal males, it is interesting to note that the fit young swimmer (subject 4) and the older sedentary smoker (subject 5) both experienced a 6% decrease in $FEV_{25-75\%}$.

DISCUSSION

Forced Expiratory Volume

We suspected that our most consistent finding, a decrease in FEV, was due to the 5°C drop in environmental temperature between December 30 and December 31, since very high concentrations of particulate matter are known to have very little effect on pulmonary compliance (Dubois and Dautrebaude, 1958). We tested this hypothesis on January 20, 1973 in an environmental chamber preset to 15°C. A 48-hour run with the Andersen sampler indicated no measurable particulate matter in the chamber. The two maximal FEV efforts for the two subjects with a history of adult respiratory disease were the same on New Year's Eve and on Jan. 20 ($\bar{d} = 0.0026$, $t = 0.05$). Similarly, the seven normal subjects showed no significant difference between these two test dates ($T_a = 15°C$): $\bar{d} = 0.0442$, $t = 1.120$, $.4 > P > .3$. However, for all 9 subjects combined, there was a nearly statistically significant decrease from Control I (20°C): $\bar{d} = 0.111$, $t = 1.78$, $.1 > P > .05$). Thus, we conclude that the significant change in FEV between December 30 and December 31 (Table 3) was probably due to temperature and not fireworks smoke. The effect of cold to reduce the FEV is explicable if peripheral vasoconstriction induced a blood shift of about 150 ml into the pulmonary circulation, displacing an equal volume of gas. While no subjects felt cold in 20°C air, most complained of the cold in 15°C air.

Forced Expiratory Flow

This study presents the first direct evidence that fireworks air pollution in Honolulu can alter pulmonary function in susceptible people (Table 3, subjects 1 and 2). This result was not due to the 5°C decrease in T_a between Control I and New Year's Eve since the $FEV_{25-75\%}$ was also significantly ($P < 0.05$) larger on Control II than on New Year's Eve. Moreover, we know of no data which would indicate that the slight change in RH between December 30 and December 31 (75% and 82% RH, respectively) could have caused the 26% reduction in $FEV_{25-75\%}$ seen in subjects 1 and 2. Thus, the most likely explanation for this observation is the smoke from burning fireworks. Such a result was anticipated in light of clinical observations and a published report on hospital admissions for chronic pulmonary patients on New Year's Eve (Bach et al. 1972).

Although five of the seven healthy subjects tended to show a decrease in $FEV_{25-75\%}$ from December 30 to December 31, the average change was not significant (Table 3). Thus, the population of Honolulu unburdened by chronic respiratory disease probably experiences little, if any, change in $FEV_{25-75\%}$ with the pollution levels reported here. However, Frank et al. (1962) exposed humans to 4–15 ppm SO_2 and also found a negligible change in FEV_1 and only a 7% decrease in peak flow rate while the resting tidal flow resistance increased 89%. They suggest that the deep inspiration required for all FEV tests may reverse the small airway narrowing detectable during quiet breathing. Thus, the possibility exists that a more sensitive test of airway resistance would have disclosed a significant change in the healthy subjects in the present study.

At 80 feet above the valley floor, our subjects may not have been exposed to the highest concentration of smoke, which tends to settle in the valleys in the absence of trade winds. The wind velocity at 11 PM on December 31 was about 4 mph, (anonymous, 1972) which is well below the average of 8–11 mph for Honolulu. Bach *et al.* (1972) used the same nephelometer the previous year on the floor of Palolo Valley, adjacent to Manoa Valley. They found the lower scale limit (380 μg/m^3) was exceeded before 9 PM and for most of five hours remained above this limit. That year the wind velocity was 2 mph or less and the RH about 90%. Thus, the smoke concentrations reported here were probably less than those of the previous year and may have been less than that in other areas during the 1972–73 New Year. However, even assuming that higher smoke concentrations may have existed and that many susceptible people were affected, the significance of an acute reduction in FEV$_{25-75\%}$ with respect to longer-term health remains unknown.

The combustion products of fireworks taken alone are probably not highly toxic. The typical compostion of the flash powder formulated in Hong Kong and used in fireworks exploded in Hawaii is: aluminum powder, 25%; potassium chlorate, 45% (potassium perchlorate may also be used); and sulfur, 30%.[2] Thus, we expected SO$_2$ gas, KC1 crystals, and oxides of aluminum to be produced. The average SO$_2$ level from 10 PM to 2 AM December 31–January 1 was 108 μg/m^3 at the fifth story level in downtown Honolulu.[3] These levels of SO$_2$ (0.04 ppm) and particulate matter (up to 4.0 mg/m^3) are below those found to cause acute effects in humans (Frank *et al.*, 1962; McDermott, 1962). However, the SO$_2$ level in Manoa Valley around midnight was probably much higher than 0.04 ppm.

Amdur and Underhill (1968) reported that 10 mg/m^3 of KC1 aerosol, while having no effect on airway resistance in guinea pigs, caused a 3.3-fold potentiation of the effect of 2.3 ppm SO$_2$. They related this effect to the solubility of SO$_2$ in various aerosols. Potassium chloride was found to be 1.4 times as effective as NaCl in potentiating the response to SO$_2$. Recently, McJilton, Frank, and Charlson, (1973) confirmed Amdur and Underhill's results in guinea pigs using low concentrations of SO$_2$ and NaCl. They found that 1 ppm SO$_2$ in combination with 1 mg/m^3 NaCl aerosol increased pulmonary flow resistance in guinea pigs by 45%, but only when the RH was high (80%). They argued that if the RH is high enough the hydration of the aerosol particles and hence their uptake of SO$_2$ proceed too rapidly for the SO$_2$ to be scrubbed from the air by the nose. The particles then carry the irritating SO$_2$ into the lung. The RH at the time of the New Year fireworks was 82% at the Honolulu airport. Thus, it seems probably that the 26% decrease in FEV$_{25-75\%}$ in susceptible people was the result of an SO$_2$–KCl aerosol interaction. However, the pollution was apparently subthreshold for spirometric effects to be seen in subjects without a respiratory disease history. Nevertheless, the potential exists for nearly one million people to be affected in future years should the use of fireworks continue to grow or should Honolulu experience very calm wind conditions on New Year's Eve.

[3] Data provided by the Hawaii State Health Department, Division of Air Pollution. Normal 24 hour average concentration of SO$_2$ at this site is 15–20 μg/m^3.

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Perchlorate Environmental Contamination: Toxicological Review and Risk Characterization

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Preface

The purpose of this review is to provide scientific support and rationale for hazard identification and dose-response assessments based on the emerging data for both human health and ecological effects caused by exposures to perchlorate. It is not intended to be a comprehensive treatise on the chemical or the toxicological nature of perchlorate.

In Chapter 10, the U.S. Environmental Protection Agency (EPA) has characterized its overall confidence in the quantitative and qualitative aspects of hazard and dose-response (U.S. Environmental Protection Agency, 1995) for both the human health and ecotoxicological effects of perchlorate. Matters considered in this characterization include knowledge gaps, uncertainties, quality of data, and scientific controversies. This characterization is presented in an effort to make apparent the limitations of the individual assessments and to aid and guide the risk assessor in the ensuing steps of the risk assessment process.

Development of these hazard identifications and dose-response assessments for perchlorate have followed the general guidelines for risk assessments set forth by the National Research Council (1983). Other EPA guidelines that were used in the development of this health risk assessment include the Assessment of Thyroid Follicular Cell Tumors (U.S. Environmental Protection Agency, 1998a), Guidelines for Neurotoxicity Risk Assessment (U.S. Environmental Protection Agency, 1998b), 1996 Proposed Guidelines for Carcinogen Risk Assessment (Federal Register, 1996), Guidelines for Reproductive Toxicity Assessment (U.S. Environmental Protection Agency, 1996a), Use of the Benchmark Dose Approach in Health Risk Assessment (Crump et al., 1995), Methods for Derivation of Inhalation Reference Concentrations and Application of Inhalation Dosimetry (U.S. Environmental Protection Agency, 1994), Proposed Interim Policy for Particle Size and Limit Concentration Issues in Inhalation Toxicology Studies (Whalan and Redden, 1994), Guidelines for Developmental Toxicity Risk Assessment (Federal Register, 1991), Recommendations for and Documentation of Biological Values for Use in Risk Assessment (U.S. Environmental Protection Agency, 1988), The Risk Assessment Guidelines of 1986 (U.S. Environmental Protection Agency, 1987), and the Guidelines for Ecological Risk Assessment (U.S. Environmental Protection Agency, 1998c).

The document presents the hazard identification or dose-response assessment for noncancer toxicity for each route of exposure, either the oral reference dose (RfD) or the inhalation

reference concentration (RfC). The RfD and RfC are meant to provide information on long-term effects other than carcinogenicity, although more recently, the value of mode-of-action information to inform the potential for a continuum from noncancer toxicity as precursor lesions to carcinogenicity presented as tumors has been recognized (Federal Register, 1996; Wiltse and Dellarco, 1996). Consideration of this continuum is especially pertinent to the evaluation of the potential toxicity of perchlorate. When such a continuum can be characterized, the dichotomous approaches to "noncancer" versus "cancer" toxicity can be harmonized into one route-specific estimate. The objective is to select a prominent toxic effect or key event that is pertinent to the chemical's key mode of action, defined as a chemical's influence on molecular, cellular, and physiological functions (Wiltse and Dellarco, 1996). A harmonized approach to the neurodevelopmental and neoplastic effects of perchlorate is proposed herein.

In a default characterization without mode-of-action information, the RfD typically is based, in part, on the assumption that a threshold exists for certain toxic effects, both for the individual and the population; whereas, a threshold may not exist for other carcinogenic effects. Thus, if the critical toxic effect is prevented, then all toxic effects are prevented. In general, the RfD or RfC is an estimate (with uncertainty spanning perhaps an order of magnitude) of a daily oral exposure or continuous inhalation exposure to the human population (including sensitive subpopulations) that is likely to be without deleterious noncancer effects during a lifetime. The oral RfD is expressed in units of milligrams per kilogram per day. The inhalation RfC considers toxic effects for both the respiratory tract as the portal of entry, as well as for effects remote to the respiratory tract (extra-respiratory or systemic effects). The RfC is expressed in units of milligrams per cubic meter.

The carcinogenicity assessment is meant to provide information on three aspects of the carcinogenic risk assessment for perchlorate: the EPA classification and quantitative estimates of risk from both oral and inhalation exposure. The classification reflects a weight-of-evidence judgment of the likelihood that the agent is a human carcinogen and the conditions under which the carcinogenic effects may be expressed.

The screening-level ecological risk assessment of environmental contamination by perchlorate follows the Guidelines for Ecological Risk Assessment (U.S. Environmental Protection Agency, 1998c).

Acknowledgments

EXECUTIVE SUMMARY

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2

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4   The purposes of this document is to present an assessment that updates previous

5   provisional values issued by the U.S. Environmental Protection Agency (EPA) for an oral

6   reference dose (RfD) for perchlorate and revises the assessment previously released as a draft

7   external review document (U.S. Environmental Protection Agency, 1998d). The objective of this

8   assessment is to derive a human health risk estimate, based on an evaluation of its potential to

9   cause toxicity or cancer, and to provide a screening-level ecological risk assessment for

10   perchlorate based on all toxicity data that recently have become available to the Agency as of fall

11   2001. Another important objective was to evaluate the evidence for indirect exposures, i.e.,

12   those exposures not by direct ingestion of contaminated water. This revised assessment

13   incorporates data from new studies and analyses in response-level to recommendations made at a

14   previous peer review of the 1998 draft (Research Triangle Institute, 1999). Most of these data

15   were obtained as results of a testing strategy that was designed with knowledge of the mode of

16   action for perchlorate toxicity that identified major data gaps in the data available prior to 1997.

17   This executive summary concisely presents key findings from the present assessment.

18

19   **SUMMARY FINDINGS**

20   **Sources of Perchlorate Contamination and Occurrence**

21   • Perchlorate is an oxidizing anion that originates as a contaminant in ground and surface waters

22   from the dissolution of ammonium, potassium, magnesium, or sodium salts. Perchlorate is

23   exceedingly mobile in aqueous systems and can persist for many decades under typical ground

24   and surface water conditions.

25   • Ammonium perchlorate is manufactured for use as the oxidizer component and primary

26   ingredient in solid propellant for rockets, missiles, and fireworks. Because it is a reducing

27   agent, it can undergo a variety of intramolecular redox reactions that lead to the release of

28   gaseous products. Through such reactions, it acts as a thrust booster.

29   • Perchlorate salts are also used on a large scale as a component of air bag inflators. Perchlorate

30   salts are also used in nuclear reactors and electronic tubes, as additives in lubricating oils, in

1   tanning and finishing leather, as a mordant for fabrics and dyes, in electroplating, in aluminum

2   refining, and in rubber manufacture, as a mordant for fabrics and dyes, and in the production of

3   paints and enamels. Chemical fertilizer had been reported to be a potential source of

4   perchlorate contamination, but new investigations by the Agency have determined that this is

5   not an issue for agricultural applications.

6 • Large-scale production of perchlorate-containing chemicals in the United States began in the

7   mid-1940s. Because of its shelf life, perchlorate must be washed out of the United States'

8   missile and rocket inventory to be replaced with a fresh supply. Thus, large volumes have been

9   disposed of in various states since the 1950s.

10 • Perchlorate began to be discovered at various manufacturing sites and in well water and

11   drinking water supplies within the months following the April 1997 development of an ion

12   chromatography analytical method that achieved a method detection limit (MDL) of

13   approximately 1 ppb and a minimum reporting limit (MRL) of 4 ppb. There are 20 states with

14   confirmed releases in ground or surface water. There are 40 states that have confirmed

15   perchlorate manufacturers or users based on EPA Information Request responses.

16   In California, most of the locations where perchlorate has been detected are associated with

17   facilities that have manufactured or tested solid rocket fuels for the Department of Defense or

18   the National Aeronautics and Space Administration.

19 • To date, there has not been a systematic national survey of perchlorate occurrence and a

20   National Primary Drinking Water Regulation for perchlorate does not currently exist.

21   Perchlorate was placed on the Contaminant Candidate List (CCL) in March 1998. The CCL

22   lists priority contaminants (defined as either known or anticipated to occur in public water

23   systems) in need of research, guidance development, regulatory determinations, or monitoring

24   by the states. Perchlorate was listed as a contaminant that required additional research and

25   occurrence information before regulatory determinations could be considered.

26 • Perchlorate was placed on the Unregulated Contaminants Monitoring Rule (UCMR) in March

27   1999 (Federal Register, 1999) to gather needed exposure information. Under the UCMR, all

28   large public water systems and a representative sample of small public water systems were

29   required to monitor for perchlorate beginning in January 2001. This effort does not extend to

30   investigating potential sources in ground and surface water that have not migrated into public

31   water supplies. Identification of the magnitude and extent of perchlorate occurrence in the

1   environment is important in assessing the routes of exposure to humans and for determining the

2   different types of organisms and ecosystems that may be affected.

3   • In early 2000, an analytical method to detect perchlorate in drinking water (EPA Method 314.0)

4   using ion chromatography was published as a direct final rule (Federal Register, 2000). The

5   EPA Method 314.0 was also approved as a monitoring method for the UCMR (Federal

6   Resister, 2000). The MDL for the method is 0.53 ppb and the MRL is 4 ppb. Improvements

7   developed commercially in the analytical capabilities may lower the MRL to the sub-part per

8   billion level in the near future.

9   • Adequate exposure characteristics of transport and transformation in the environment are also

10   absent. Preliminary biotransport studies at six contaminated sites indicate a potential for

11   uptake into plant and animal tissues in ecosystems. Extension of analytical methods to detect

12   perchlorate in plant and animal tissues awaits validation before a conclusive determination can

13   be made.

14

An Integrated Approach to Comprehensive Risk Characterization

16   • Perchlorate is of concern for several reasons. First, there were uncertainties in the toxicological

17   database available that could be used to evaluate the potential for perchlorate to produce human

18   health effects when present at low levels in drinking water. The purpose of the targeted

19   toxicity testing strategy was to develop a database to address key data gaps. Secondly, the

20   actual extent of the occurrence of perchlorate in ground and surface waters is not known at this

21   time. Additionally, the efficacy of different treatment technologies for various water uses (such

22   as drinking water or agricultural applications) and different scales (i.e., large or small volumes)

23   is still being determined. Finally, the extent and nature of ecological impact or transport and

24   transformation phenomena in various environmental media have only, as yet, been studied

25   superficially.

26   • To adequately and comprehensively characterize the risks posed by perchlorate contamination

27   and to develop scientifically-based management strategies that effectively mitigate the potential

28   risks posed by perchlorate contamination, several advances are essential. The analytical

29   methods used to characterize various exposures must be accurate and precise. The exposure

30   estimates cannot be gauged with respect to their risk unless robust health and ecological risk

31   estimates are available. Treatment technologies should be targeted to levels of concern and

1   tailored to the intended water use. Technology transfer is necessary so that all affected parties

2   and concerned citizens are apprised of accurate and reliable information that is up to date with

3   the evolving state of the science.

4 • The toxicity testing strategy was expedited through a unique partnership between the

5   Department of Defense and EPA, together with members of an Interagency Perchlorate

6   Steering Committee (IPSC), which includes other governmental representatives from the

7   National Institute for Environmental Health Sciences (NIEHS) and affected state, tribal, and

8   local governments.

9 • The charge of the IPSC is to facilitate and coordinate accurate accounts of related technological

10   issues (occurrence surveys, health assessment, ecotoxicology assessment, treatability, waste

11   stream handling, and analytical detection). This assessment is intended to address the need for

12   evaluation of perchlorate's potential to cause human health effects or impact on ecological

13   systems, based on currently available data.

14

15 **Physicochemical Characteristics**

16 • As an oxidant, perchlorate is kinetically nonlabile. This means the reduction of the central

17   chlorine atom from an oxidation state of $+7$ (perchlorate) to -1 (chloride ion) occurs extremely

18   slowly. Sorption is not expected to attenuate perchlorate because it absorbs weakly to most soil

19   minerals. Natural chemical reduction in the environment is not expected to be significant.

20   These two factors account for perchlorate being both very mobile in aqueous systems and

21   persistent for many decades under typical ground and surface water conditions.

22 • The activation energy to perchlorate reduction is so high that it cannot be expected to act as an

23   oxidant under human physiological conditions (i.e., dilute solution, unelevated temperatures,

24   neutral pH). This is supported by absorption, distribution, metabolism, and elimination studies

25   that show perchlorate is excreted virtually unchanged in the urine after absorption.

26

27 **Hazard Identification and Mode of Action Testing Strategy**

28 • The health effects and toxicity database available in the spring of 1997 was determined to be

29   inadequate for quantitative risk assessment by an independent (non-EPA) peer review. A

30   testing strategy was developed based on a hazard identification using the available data and the

31   suspected mode of action for perchlorate to target testing on potential effects of perchlorate.

1   Data from this effort was used to support the previous EPA draft assessment and this revised

2   assessment in 2002.

3 • To design a testing strategy based on the mode of action for a chemical, it is necessary to

4   understand its toxicokinetics and toxicodynamics. Perchlorate is readily absorbed from the

5   intestinal tract, and oral uptake is considered to be the major route of exposure. Because of its

6   high charge, perchlorate does not pass readily through the skin. Exposure via inhalation is

7   expected to be negligible because the vapor pressure of perchlorate salts and acids is expected

8   to be low at room temperatures. Droplet size during showering likely would preclude

9   inhalation of perchlorate-contaminated water as an aerosol. Perchlorate is known to inhibit the

10   uptake of iodide in the thyroid at the sodium (Na+)–iodide (I-) symporter, or NIS, thereby

11   causing a reduction in the hormones thyroxine (T4) and triiodothyronine (T3). When these

12   hormones enter the blood circulation, they are bound to plasma proteins. There may be other

13   locations of inhibition of iodide transport in the gland. Perchlorate itself is not metabolized in

14   the thyroid or peripheral tissues.

15 • Control of circulating concentrations of these hormones is regulated primarily by a negative

16   feedback known as the hypothalamic-pituitary-thyroid axis or feedback system involving three

17   organs: (1) the thyroid, which produces T4 and T3; (2) the pituitary gland which produces

18   TSH; and (3) the hypothalamus, which also responds to and helps to maintain optimal T4 and

19   T3 levels. The hypothalamus stimulates the pituitary gland through thyrotrophic-releasing

20   hormone (TRH) to produce thyroid stimulating hormone (TSH), which then prompts the

21   thyroid to produce T4 and T3. Cells in the hypothalamus and pituitary gland respond to the

22   levels of circulating T4 and T3, such that when thyroid production levels are low, there is a

23   signal to increase the output of TRH and TSH. Circulating hormone levels (T4, T3, and TSH)

24   can be monitored readily to serve as biomarkers of exposure and effect of agents that disrupt

25   the status of this negative feedback system.

26 • The hypothalamic-pituitary-thyroid feedback system for regulation of thyroid hormones is

27   conserved across species. Differences in plasma protein binding between rats and humans

28   account for differences in the circulating half-life of the hormones and in thyroid response to

29   TSH between the species. New studies since 1999 have confirmed that the inhibition of iodide

30   uptake by perchlorate at the NIS is essentially the same sensitivity across species. This is

1   important when considering decrements in T4 as important to neurodevelopmental effects

2   versus neoplasia that results in the gland due to stimulation by TSH.

3 • Given its mode of action as an inhibitor of iodide uptake that results in disturbances of the

4   hypothalamic-pituitary-thyroid axis, concerns arose about the potential for perchlorate to cause

5   carcinogenic, neurodevelopmental, developmental, reproductive, and immunotoxic effects.

6   Further, there is concern for ecotoxicology effects on various aquatic and terrestrial plants and

7   animals.

8 • The human health testing strategy for perchlorate developed in 1997 originally included eight

9   different recommended studies to address data gaps and enhance the mechanistic information

10   on the mode of action. The goal of these studies was to provide a comprehensive database on

11   which to arrive at a revised human health risk assessment with greater confidence than previous

12   recommended provisional values. These studies are described briefly below.

13   (1) A 90-day oral bioassay to identify other target tissues in young adult rats; to provide data

14   on the effects of repeated exposures to perchlorate on T3, T4, and TSH levels; to

15   evaluate recovery of effects after 30 days; and to screen for some reproductive

16   parameters. A genotoxicity assay also was performed on rats from the terminal sacrifice.

17   (2) A neurodevelopmental study in rats to evaluate the potential for functional and

18   morphological effects in offspring from the mother exposed during pregnancy and

19   lactation.

20   (3) A Segment II developmental study in rabbits to evaluate the potential for perchlorate to

21   cause birth defects and to provide data on thyroid hormone effects in a second species

22   other than the rat.

23   (4) A two-generation reproductive toxicity study to evaluate the potential for perchlorate to

24   cause deficits in reproductive performance in adult rats and for toxicity in the young

25   offspring.

26   (5) Absorption, distribution, metabolism, and elimination (ADME) studies to characterize

27   the pharmacokinetics of perchlorate in laboratory animals and humans and to provide

28   data necessary to allow construction of models for quantitative description of different

29   internal dose metrics and interspecies extrapolation.

(6)	Mechanistic studies that characterize the effects of perchlorate on the iodide uptake mechanism across species as a link with the ADME studies to aid in the quantitative extrapolation of dose across species.

(7)	A battery of genotoxicity assays to evaluate the potential for carcinogenicity by evaluating the potential for direct effects on deoxyribonucleic acid (DNA).

(8)	Immunotoxicity studies to evaluate the potential for perchlorate to disrupt immune function, including cell-mediated and humoral toxicity.

• After the External Peer Review in 1999, additional studies were performed to replicate the neurodevelopmental study (i.e., changes in brain morphometry and motor activity); determine the developmental toxicity potential in rats versus rabbits; investigate additional aspects of immunotoxicity; and develop a consistent nomenclature and scoring system for the histopathological lesions in the thyroid gland. Additional pharmacokinetic data was also developed into physiologically-based pharmacokinetic (PBPK) models of perchlorate and iodide distribution.

• A battery of ecological screening tests as part of the 1997 testing strategy was conducted as part of the 1997 testing strategy in laboratory organisms representative of ecological receptors across soil, sediment, and water to evaluate dose-response relationships. These were considered to be a tier of tests to give an idea of gross toxicity that would determine the need and types of tests to be performed in the next tier. The tests did not measure the amount of perchlorate in the tissues of the species being tested. Based on stakeholder input and the need for a more focused battery of tests, lettuce was substituted for duckweed because of Tribal concerns regarding the sizable lettuce crop along the Colorado river. The following species were selected for the first round of testing:

(1)	*Daphnia magna* (water flea) to represent an aquatic invertebrate

(2)	*Ceriodaphnia magna* (water flea) to represent an aquatic invertebrate

(3)	*Lactuca sativa* (lettuce) to represent a vascular plant

(4)	*Pimephales promelas* (fathead minnow) to represent an aquatic invertebrate

(5)	*Eisenia foetida* (earthworm) to represent a soil invertebrate

(6)	*Microtus pennsylvanicus* (meadow vole) to represent an herbivore

• Other studies in the set of tests included the Frog Embryo Teratogenesis Assay: *Xenopus* (FETAX) and a phytoremediation study to examine uptake, distribution, and degradation in

1   experimental systems with rooted cuttings of woody plants, including willow, Eastern

2   Cottonwood, and eucalyptus.

3   • Additional studies, some of chronic duration, on effect levels in aquatic animals, an aquatic

4   plant, a terrestrial plant, and a soil invertebrate have been performed since 1999. A study of

5   perchlorate occurrence in six selected sites with known or suspected contamination also

6   examined perchlorate concentrations in site media and in various ecological receptors.

7

8   **Human Health Assessment**

9   • The testing strategy confirmed that the target tissue for perchlorate toxicity was the thyroid

10   gland. Anti-thyroid effects included iodide uptake inhibition, perturbations of T3, T4, and TSH

11   hormones, and thyroid histopathology in adult, fetal, and postnatal rats across studies with a

12   range of experimental design. Thyroid weight in these studies was also effected. Other than

13   effects in the thyroid, no effects were observed in rabbits of the developmental study, but the

14   developmental study in rats identified 30 mg/kg-day as the lowest observed adverse effect level

15   (LOAEL).

16   • Competitive inhibition of iodide uptake at the NIS by perchlorate is the key event leading to

17   both potential neurodevelopmental and neoplastic sequelae. The decrement in iodide uptake

18   leads to subsequent drops in T4 and T3 that can lead to permanent neurodevelopmental

19   deficits. Because of strong correlations between changes in iodide uptake inhibition with

20   decrements in T3 and T4; between T3 and T4 with changes in TSH; and between changes in

21   T3, T4, or TSH with thyroid histopathology, an assessment model was proposed that used the

22   changes in T3, T4, and TSH as the precursor lesions to subsequent effects that potentially could

23   lead to thyroid tumors or to altered neurodevelopment. This assessment approach essentially

24   harmonizes noncancer and cancer approaches because it is presumed that the no-observed-

25   adverse-effect-level (NOAEL) for the precursor lesions will preclude any subsequent sequelae

26   at higher doses.

27   • Thyroid tumors were observed in previous studies in rats exposed in long-term bioassays at

28   high doses. Thyroid tumors were more recently also diagnosed in the first-generation (F1)

29   adults (second parental generation [P2]) at 19 weeks in a two-generation reproductive study.

30   Both the latency and incidence of these tumors were significant relative to the entirety of the

31   National Toxicology Program data base for this type of tumor and in this strain of rat. These

1   effects and the demostration of a progression with duration of effects on hormones and thyroid

2   histopathology in the 90-day study raised the concern that extended exposures to perchlorate

3   may change the hypothalamic-pituitary-feedback system or the cellular sensitivity and demand

4   for thyroid hormones.

5 • The rat model is considered relevant yet conservative for human health risk assessment of

6   potential thyroid neoplasia because of the differences in thyroid structure and hormone

7   half-lives. Perchlorate was demonstrated to be nongenotoxic in the testing battery employed,

8   suggesting the antithyroid effects are an indirect mode of action for thyroid tumor formation.

9 • Due to the age- and time-dependent nature of the key event of perchlorate toxicity and its

10   anti-thyroid effects, the revised RfD was based on weight-of-the-evidence approach to the

11   entire data base. The RfD is proposed to be protective of both neurodevelopmental and

12   neoplastic sequelae. An administered dose of 0.01 mg/kg-day was supported as a lowest-

13   observed-adverse-effect level (LOAEL) based on effects on brain morphometry in pups from a

14   PND21 sacrifice in a neurodevelopmental study that repeated similar observations made in a

15   similar 1998 study, hormonal effects indicative of hypothyroidism (decreased T4 and increased

16   TSH) in the dams of those same pups on GD21, thyroid histopathology and hormone changes

17   in these same pups at various developmental stages (GD21, PND4, PND9, and PND21),

18   thyroid histopathology and hormone changes at the 14- and 90-day sacrifice dates in a

19   subchronic study, and indications of immunotoxicity (dermal contact hypersensitivity).

20 • A human equivalent exposure (HEE) was calculated using physiologically-based

21   pharmacokinetic (PBPK) models for interspecies adjustment based on the area under the curve

22   (AUC) of perchlorate in the serum as the dose metric. The HEE for the maternal dams was

23   chosen for operational derivation because brain morphometry effects may have been

24   programmed *in utero* and because the dams of effected pups were hypothyroid.

25 • A composite uncertainty factor of 300 was used to address uncertainties in the extrapolations

26   required for the RfD derivation. A three-fold factor for intraspecies variability was retained

27   due to the variability observed in the data and PBPK modeling for the adult humans and

28   because the subjects used to develop the models did not provide kinetic data for the potentially

29   susceptible population. There was also uncertainty in the parallelogram approach to extending

30   the adult structures to predict doses for different life stages in the human. A full factor of ten

31   was applied to extrapolate the LOAEL for the adverse effects (brain morphometry, colloid

1   depletion and hormone changes) observed in various studies at the 0.01 mg/kg-day dosage

2   level. A three-fold factor for duration was applied due to the concern for the biological

3   importance of the statistically significant increase in tumors observed in the F1-generation pups

4   (second parental, P2 generation) at 19 weeks and the evidence for progression of effects with

5   extended exposure in the 90-day study. The finding of tumors at 19 weeks raised concern for

6   *in utero* programming, i.e., that disruption of thyroid hormones in the developing fetus may

7   predispose the developing neonate and adult to future insults to the thyroid gland. This factor

8   can also be viewed as part of a data base deficiency since there are no adequate long-term

9   bioassays of perchlorate. Finally, a three-fold factor was applied for inaccurate characterization

10   of immunotoxicity since recent studies reinforced concern for this potential endpoint. Because

11   the test article was ammonium perchlorate, an adjustment factor of 0.85 was made for the

12   percent of molecular weight of the salt from ammonium (15.35%), so that the RfD is expressed

13   for perchlorate as the anion alone. This was done to be compatible with the analytical methods

14   that measure the anion in environmental samples and because most perchlorate salts readily

15   dissolve in water. The resultant revised RfD value for perchlorate is 0.00003 mg/kg-day.

16   Confidence in the principal study, the data base and the RfD were all designated as medium.

17

18   **Screening Ecological Risk Assessment**

19   • A secondary acute value of 5 mg/L (as perchlorate) was derived to be protective of 95% of

20   aquatic organisms during short-term exposures with 80% confidence. The secondary chronic

21   value of 0.6 (as perchlorate) likewise was derived to be protective of 95% of aquatic organisms

22   during short-term exposures with 80% confidence. These values were derived based on

23   sodium perchlorate and are probably protective even if ammonium perchlorate is the

24   contaminant released. Calculated ammonia-nitrogen concentrations corresponding to those

25   values are below the acute and chronic ambient water quality criteria for ammonia, regardless

26   of pH.

27   • For terrestrial plants, the quartile inhibitory concentrations for growth in soil and sand were

28   78 mg/kg (293 mg/L) and 41 mg/kg (160 mg/L), respectively. A factor of 10 was applied to

29   account for interspecies variance to obtain a screening benchmark of 4 mg/kg.

1 • Because of limited data on effects for soil invertebrates, a conservative estimate of a threshold

2   for soil community effects was derived at 1 mg/kg. The equivalent aqueous phase benchmark

3   is 2.8 mg/L.

4 • A factor of 10 for interspecies variance and LOAEL to NOAEL extrapolation was applied to

5   the human health risk LOAEL estimate based on rat data (0.01 mg/kg-day) to obtain a

6   screening benchmark of 0.001 mg/kg-day for the representative herbivore (meadow vole)

7   because it also is a rodent. The population-level implications of this effect are unknown, but it

8   seems likely that such effects on the thyroid could diminish survivorship and fecundity, which

9   would diminish population production.

10 • Data are available showing that perchlorate accumulates in the tissues of exposed fish,

11   amphibians, and invertebrates. However, data are insufficient to determine whether perchlorate

12   is concentrated in those tissues to levels exceeding the levels of exposure. By contrast, several

13   studies have shown that perchlorate is taken up and concentrated in aerial plant parts, especially

14   leaves, although studies designed for the purpose of quantifying plant concentration factors

15   have not yet been conducted.

16

17 **Uncertainties and Assessment Research Needs**

18 • Accurate exposure information is a requisite for risk characterization for both human and

19   ecological assessments. These data should include transport and transformation processes,

20   notably the fate of perchlorate in irrigated soils because of the potential for evaporative

21   concentration.

22 • Research concerning the human health risks of perchlorate needs to better characterize the

23   dose-response for perchlorate inhibition of iodide uptake in adults, fetuses, and neonates. More

24   definitive studies linking iodide uptake inhibition and the degree of perturbation of the

25   hypothalamic-pituitary-thyroid axis (i.e., changes in T3, T4, and TSH levels) and association

26   with neurobehavioral problems, thyroid changes, and neoplastic sequelae may continue to

27   improve the confidence in the assessment. Understanding the relative sensitivity of laboratory

28   animal assays of neurodevelopmental effects versus epidemiological studies of

29   neuropsychological development also needs to be advanced. Research on potential factors

30   influencing sensitivity is critically requisite. Animal models of thyroid impairment such as

31   iodide deficiency and "womb to tomb" exposure designs should be explored.

1 • Because only a screening tier of tests has been performed, the major uncertainty derives from

2   data gaps. Data on bioaccumulation in aquatic biota would allow evaluation of exposure of

3   organisms that feed on fish and other aquatic organisms. Effects of perchlorate on algae and

4   aquatic macrophytes are required to estimate risks to aquatic primary producers. Data on

5   bioaccummulation in aquatic plants are necessary to assess direct impact to primary consumers

6   (i.e., planktonic and benthic invertebrate communities). Data on accumulation in terrestrial

7   vascular plants also should be investigated further. The factor applied for the use of subchronic

8   data in fish could be addressed by chronic effect testing. Effects also should be determined in

9   nondaphnid invertebrates and of dietary exposure in birds and herbivorous or litter-feeding

10   invertebrates.

11

12 **Risk Characterization**

13 • As noted above, the lack of exposure information precludes comparison with the human health

14   and ecological toxicity assessment for accurate characterization of risk. Indirect human

15   exposure pathways can be addressed best by a new EPA document, Methodology for Assessing

16   Health Risks Associated with Multiple Pathway of Exposure to Combustor Emissions, which is

17   scheduled for final release in January 2002.

18 • Noncancer neurobehavioral effects have been shown at lower doses. The estimate for

19   perchlorate has been based on precursor effects considered protective for both the thyroid

20   neoplasia and neurodevelopmental effects. It is appropriate for comparison against direct oral

21   exposures. The frequency and magnitude of exposure are key attributes for characterization

22   compared with those assumptions of continuous lifetime exposure assumed in the derivation.

23   The degree to which the particular suspected population at risk fits with the assumptions used

24   in the RfD derivation should be kept in mind when performing any risk characterization.

25   Further, RfD estimates are not intended to serve as a "bright line" because, by definition, there

26   is an order-of-magnitude uncertainty around the estimate. This typically translates into a range

27   of threefold below to threefold above the RfD.

28 • Ecological risk could not be precluded nor accurately characterized because of the significant

29   data gaps described above.

30

1. INTRODUCTION

The purpose of this document is to revise the previous human health and ecological risk assessment external review draft (ERD) document (U.S. Environmental Protection Agency, 1998d). This revision is based on recommendations made at the 1999 external peer review (Research Triangle Institute, 1999). The peer review panel recommended some alternative analyses and several additional studies. This revised assessment addresses these recommendations and is based on all data made available to the Agency as of Fall 2001; including new studies from the perchlorate testing strategy. The purpose of this chapter is to provide background information on the current status of perchlorate (ClO_4^-) contamination in the United States and an historical perspective on how certain issues of concern have evolved to prominence. The role of this risk assessment will be placed in context with respect to the overall integrated approach to addressing the perchlorate contamination and regulatory readiness.

1.1 PRODUCTION USES AND SOURCES OF PERCHLORATE CONTAMINATION

Perchlorate is an oxidizing anion that originates as a contaminant in ground and surface waters from the dissolution of perchloric acid and of the salts including ammonium, potassium, magnesium, or sodium. With the exception of potassium perchlorate, each of these compounds is extremely soluble. Potassium perchlorate is regarded as sparingly soluble; however, it may dissolve completely under the appropriate environmental conditions.

Ammonium perchlorate is the oxidizer and primary ingredient (by mass) in solid propellant for rocket motors. For example, ammonium perchlorate (NH_4ClO_4) makes up 69.7% of the propellant for the space shuttle rocket motors and 65 to75% of the Stage I motors of the Minuteman III and 68% of the Titan missile motors (Rogers, 1998). Because the ammonium ion is a reducing agent, ammonium perchlorate can undergo a variety of intramolecular redox reactions that lead to the release of gaseous products. The explosive decomposition shown in Equation 1-1 is induced thermally and occurs at temperatures below 300 °C (Schilt, 1979a).

1	$4\ NH_4ClO_4(s) \rightarrow 2\ Cl_2(g) + 3\ O_2(g) + 2\ N_2O(g) + 8\ H_2O(g)$	(1-1)
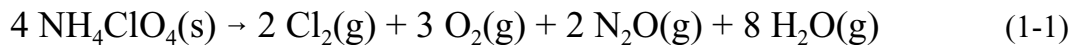
2

3   Through such reactions, ammonium perchlorate also acts as a thrust booster. Even after such

4   decomposition, the dichlorine and dioxygen thus produced remain capable of engaging in

5   subsequent redox reactions with fuels.

6   Specific uses of various perchlorate salts include: solid rocket fuel oxidizer, flares, and

7   pyrotechnics (potassium); solid rocket fuel oxidizer, explosives, chemical processes and

8   pyrotechnics (ammonium); precursor to potassium and ammonium perchlorate and in explosives

9   (sodium); and military batteries (magnesium) (Rogers, 1998). Perchlorate salts also are used on a

10   large scale as a component of air bag inflators. Other industrial or commercial applications of

11   perchlorate salts include their use in nuclear reactors and electronic tubes; as additives in

12   lubricating oils; in tanning and finishing leathers; as a mordant for fabrics and dyes;

13   in electroplating, aluminum refining, and rubber manufacture; and in the production of paints and

14   enamels (Siddiqui et al., 1998). A 1998 report raised the concern that chemical fertilizer is

15   a potential source of perchlorate contamination (TRC Environmental Corporation, 1998). More

16   recent studies limit concern to certain types of fertilizer containing Chilean caliche (Urbansky,

17   2000; U.S. Environmental Protection Agency, 2001a,b; Urbansky and Collette, 2001); however,

18   production practices have been changed to address that issue. Besides their large-scale

19   commercial uses, perchlorate salts often are employed on a small scale in laboratory chemical

20   studies as ionic strength adjustors or as noncomplexing counterions. Some still are used in

21   medical diagnostics in thyroid function tests. Perchloric acid is used for various laboratory

22   applications requiring a strong acid. Wet ashing organic matter with perchloric acid still is

23   performed today as a means of preparation for certain samples. Anhydrous magnesium

24   perchlorate ($Mg(ClO_4)_2$) is a strong desiccant; however, historically, Anhydrone[®], a slightly

25   hydrated form of $Mg(ClO_4)_2$, has been used to collect the water formed in combustion analyses.

26   The large-scale production of perchlorate-containing chemicals in the United States began

27   in the mid-1940s. The approximate percentages sold for specific end uses are 92% as an

28   oxidizer, 7% as an explosive, and 1% other uses (American Pacific Corporation, 1998). The

29   typical volume of production ranged from 1 to 15 million lb per year (Rogers, 1998) although

30   production in the 1980's was generally 20 to 30 million pounds per year (Kerr-McGee Chemical

31   LLC, 1998; American Pacific Corporation, 1998). Solid rocket fuel inventories are growing at a

1   significant rate as systems reach the end of their service life and as treaties mandate motor

2   disposal. The current disposal method for these motors is open burning or open detonation, both

3   of which are becoming increasingly difficult to perform under intense public and regulatory

4   pressure based, in part, on concern over incomplete destruction via these methods. Currently, the

5   large solid rocket motor disposal inventory shows 55 million lb of propellant awaits disposal, and

6   this number is expected to be over 164 million lb by the year 2005 (Siddiqui et al., 1998).

7   A significant portion of this inventory contains ammonium perchlorate, which now can be

8   reclaimed and recycled into new motor propellants. The accepted method for removal and

9   recovery of solid rocket propellant from rocket motors is high-pressure water washout. This

10   method generates large amounts of aqueous solution containing low concentrations of

11   ammonium perchlorate. Although ammonium perchlorate can be recovered from these aqueous

12   solutions, it is cost-prohibitive to remove it entirely. Most of the locations where perchlorate has

13   been detected in ground or surface waters are in areas associated with development, testing, or

14   manufacture of aerospace materials. Explosives and fireworks manufacturing and disposal

15   operations have also been implicated in a number of environmental releases. Laboratory

16   activities and fertilizer operations are potential sources of contamination in relatively few known

17   instances. Perchlorate contamination also may occur where mining activities use explosives

18   extensively (Siddiqui et al., 1998).

19   When ammonium perchlorate is released to water, the salt is highly soluble and dissociates

20   completely releasing ammonium (NH_4) and perchlorate (ClO_4^-):

21

22 $$NH_4ClO_4(s) \ \overset{H_2O}{\rightarrow} \ NH_4^+(aq) \ + \ ClO_4^-(aq). \qquad\qquad (1\text{-}2)$$

23
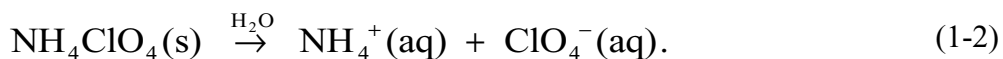
24   Its high solubility is not affected by pH or temperature. It is likely that most of the ammonium

25   has been biodegraded, and the cation in the environment is best viewed as mostly sodium (Na^+)

26   or possibly hydrogen (H^+), especially where contamination levels are below 100 ppb;

27   nevertheless, those regions with high concentrations of perchlorate ion probably retain a small

28   fraction of ammonium ion (Urbansky, 1998a). At those sites where contamination has occurred

29   for decades, very little (if any) ammonium ion has been found. To date, there has been no

30   quantitative determination of the cations responsible for the charge balance.

1   As an oxidant, perchlorate is kinetically nonlabile. This means that reduction of the central

2   chlorine atom from an oxidation state of +7 (perchlorate) to −1 (chloride ion) occurs extremely

3   slowly. This will be elaborated on in Chapter 2 in the discussion of physicochemical

4   characteristics. Sorption is not expected to attenuate perchlorate concentrations because it

5   absorbs weakly to most soil minerals. Natural chemical reduction in the environment is also not

6   expected to be significant. Together, these two factors account for perchlorate's high mobility

7   and persistence for many decades under typical groundwater and surface water conditions.

8   Figure 1-1 summarizes the various pathways through which perchlorate can reach ground and

9   surface water sources.

10

11

1.2 EVOLUTION OF ANALYTICAL DETECTION METHODS AND EMERGING OCCURRENCE DATA

14   The Region 9 Office of the U.S. Environmental Protection Agency (EPA) first became

15   aware of the potential contamination issues with perchlorate in 1985 when samples measured

16   with a colorimetric method reported contamination in 14 wells ranging from 0.11 to 2.6 ppm

17   (Takata, 1985). The Region 9 office requested assistance from the Centers for Disease Control

18   and Prevention (CDC) to evaluate the potential health effects of these levels of perchlorate

19   (Takata, 1985). In response the CDC recommended validation of the colorimetric measures but

20   could not address the potential for toxicity of the chemical because of toxicity data

21   insufficiencies (Margolis, 1986). The CDC also recommended additional testing to determine

22   potential target tissues and the effects from long-term, low-level exposures. The absence of a

23   valid analytical method to measure low concentrations of perchlorate and the lack of data with

24   which to characterize the risk of toxicity led Region 9 of EPA to focus on chemicals other than

25   perchlorate at these sites. By the early 1990s, however, perchlorate at detectable levels

26   (>1 mg/L) was found in monitoring wells at a California Superfund site, and EPA Region 9

27   increased its effort to establish a human-health-based reference dose (RfD) in order to help gauge

28   the risk of the contamination that was beginning to be characterized. In 1997, after perchlorate

29   was discovered in a number of California water supplies, the California Department of Health

30   Services (CA DHS) adopted 18 ppb as its provisional action level.

31
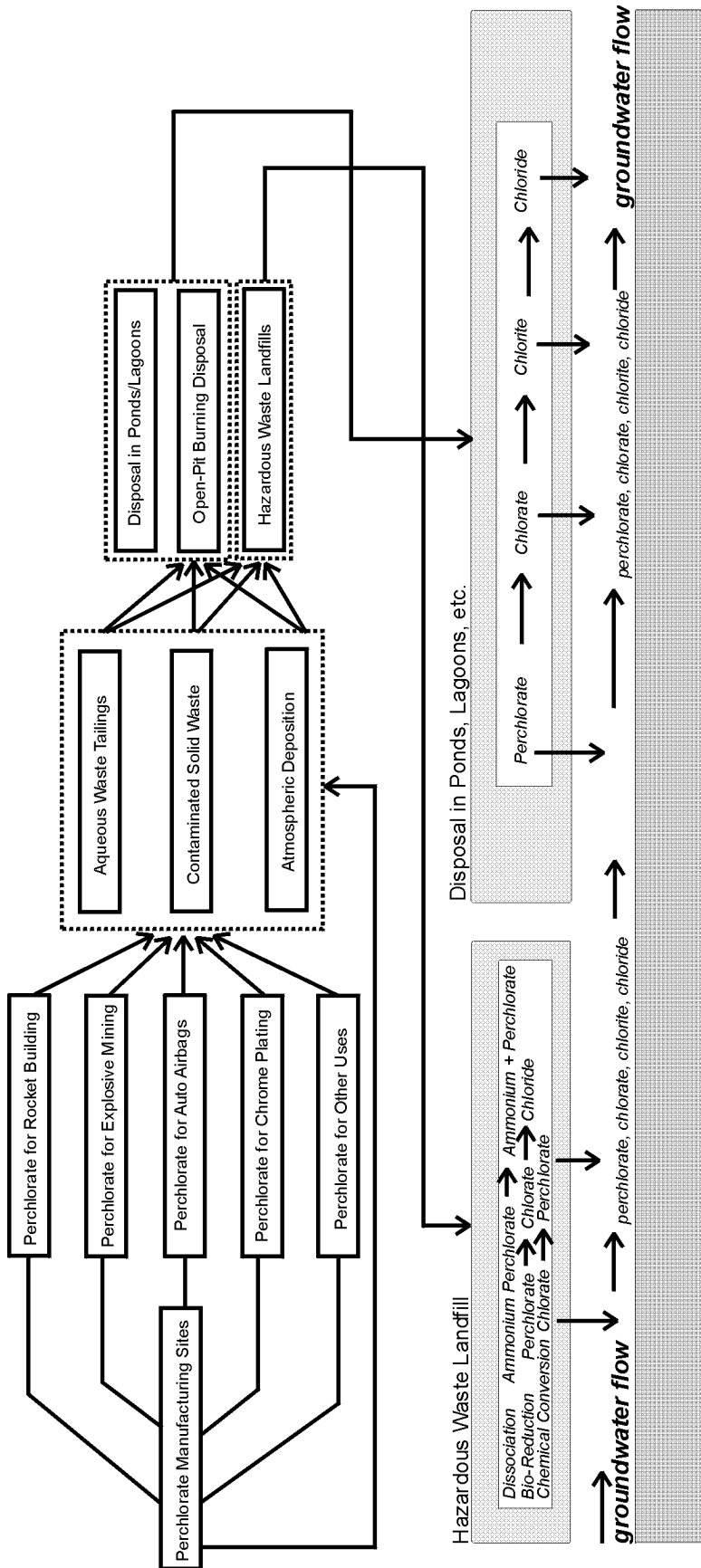
Figure 1-1. Sources and pathways of groundwater contamination for perchlorate. (Modified from Siddiqui et al., 1998.)

1   In January 1997, the California Department of Health Services' Division of Drinking Water

2   and Environmental Management requested the Sanitation and Radiation Laboratory Branch

3   (SRLB) test for perchlorate in drinking water wells potentially affected by groundwater migrating

4   from the Aerojet facility near Sacramento. Based on its provisional action level, Region 9 of

5   EPA indicated that a reporting limit of at least 4 ppb would be necessary. However, procedures

6   to measure perchlorate at such low levels were not available. An ion chromatographic (IC)

7   method was capable of detecting 400 ppb; and, during the previous year, Aerojet had improved

8   the method to detect 100 ppb. By March 1997, SRLB and an analytical equipment manufacturer

9   had developed an IC method that achieved a method detection limit of approximately 1 ppb and a

10   reporting limit of 4 ppb. Within several months following the March 1997 development of the

11   low-level (4 ppb) IC detection method, perchlorate was discovered at various manufacturing sites

12   and in well water and drinking water supplies in California, Nevada, and Utah.

13   Efforts in several additional laboratories helped improve the IC method (Eldridge et al.,

14   1999; Urbansky, 2000). Although IC is the dominant analytical method used at this time, a

15   variety of additional techniques are being refined for perchlorate analysis, including: mass

16   spectrometry, Raman spectrometry, capillary electrophoresis, and others (Urbansky, 2000).

17   Recent publications have reported detection of perchlorate in tap water at levels as low as 0.1 ppb

18   (Handy et al., 2000; Koester et al., 2000).

19   In March 1999, EPA included perchlorate in the Unregulated Contaminant Monitoring

20   Rule (UCMR) (Federal Register, 1999). Under the UCMR, all large public water systems and a

21   representative sample of small public water systems were required to monitor for perchlorate

22   beginning in January 2001. The EPA Method 314.0 for the analysis of perchlorate in drinking

23   water using IC methods was published in early 2000 as a direct final rule (Federal Register,

24   2000). The EPA Method 314.0 was also approved as a monitoring method for the UCMR

25   (Federal Register, 2000). However, this effort does not extend to investigating potential sources

26   in groundwater and surface water that have not migrated into public water supplies. Additional

27   information about the UCMR is available at the web site http://www.epa/gov/safewater/

28   ucmr.html.

29   The CA DHS adopted 18 ppb as its provisional action level in 1997 after perchlorate was

30   discovered in a number of California water supplies. The CA DHS also added perchlorate to the

31   list of unregulated chemicals for which monitoring is required in 1999 (Title 22, California Code

1	of Regulations §64450). By September 2001, over 2,800 sources of public water supply had
2	been sampled in California by water supply agencies responding to CA DHS requirements. Most
3	of these sources represent water supply wells. Of the sources sampled, 206 (over 7 percent) had
4	perchlorate concentrations greater than 5 ppb in at least two samples (Figure 1-2). Most of these
5	wells have as their source groundwater plumes that have spread as far as nine miles from the site
6	of original release.
7	At this time, there has not been a systematic national survey of perchlorate occurrence.
8	Several states and EPA regions are taking significant steps to test water supplies for perchlorate,
9	notably the states of Arizona, Utah, and Texas, EPA Regions 6 and 7, and Suffolk County,
10	New York. Figure 1-3 indicates states with confirmed perchlorate manufacturers or users, and
11	Figure 1-4 indicates those states in which facilities have, in response to reported releases, directly
12	measured perchlorate in groundwater or surface water. Table 1-1 describes these locations. The
13	data published in Siddiqui et al., 1998 (drinking water systems in New Mexico, Indiana,
14	Pennsylvania, and Iowa) are displayed in Figure 1-3 and in Table 1-1, but they have not been
15	independently confirmed.
16	Information on other potential sites across the country is being gathered from the
17	Department of Defense (DoD) and National Aeronautics and Space Administration (NASA)
18	searches and from EPA information requests made to perchlorate manufacturers. The EPA has
19	notified state, tribal, and local governments when it has evidence of perchlorate manufacture and
20	use in these governmental jurisdictions. The American Water Works Association Research
21	Foundation is coordinating a survey to characterize possible perchlorate contamination of
22	drinking water sources in areas of high risk. The EPA will build on these survey data and other
23	information to discover potential sources and evaluate threats to water resources.
24	Region 9 officials have collected information concerning detected perchlorate releases in
25	20 different states (Table 1-1). For two of these states, Pennsylvania and Indiana, the only
26	reported releases have not been confirmed by a state or federal agency and should be considered
27	questionable until the detections can be independently validated. In Washington State, propellant
28	was observed scattered around open burn/open detonation sites although results of solid rocket
29	chemical analyses of groundwater samples are not yet available. In California, most areas where
30	perchlorate has been detected are associated with facilities that have manufactured, tested, or
31	disposed of solid rocket fuels and propellants for DoD or NASA. Two facilities that
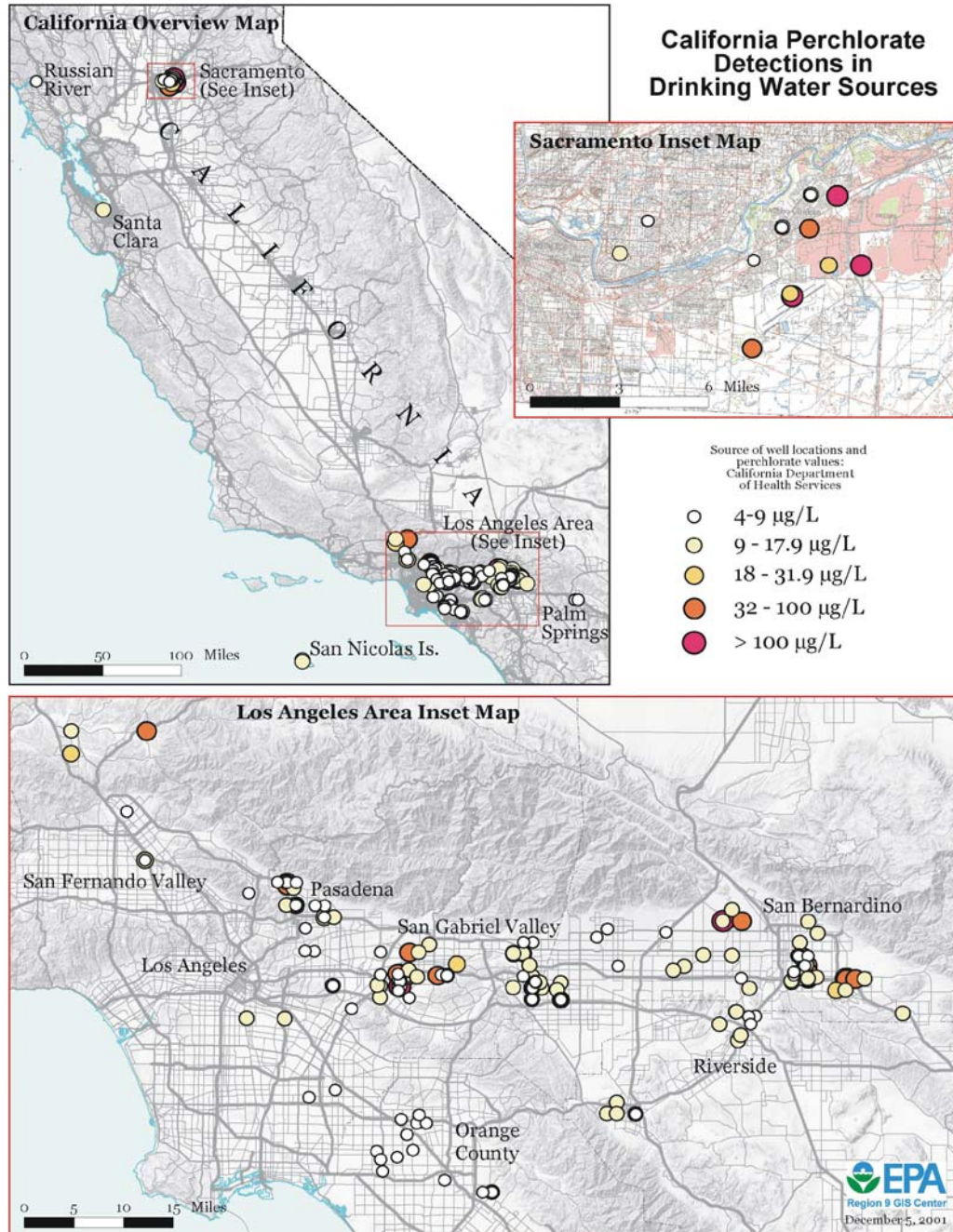
Figure 1-2. Distribution of perchlorate detected in public water supply sources in California. Also noted are several large sites of groundwater contamination that include perchlorate releases (Mayer, 2001).
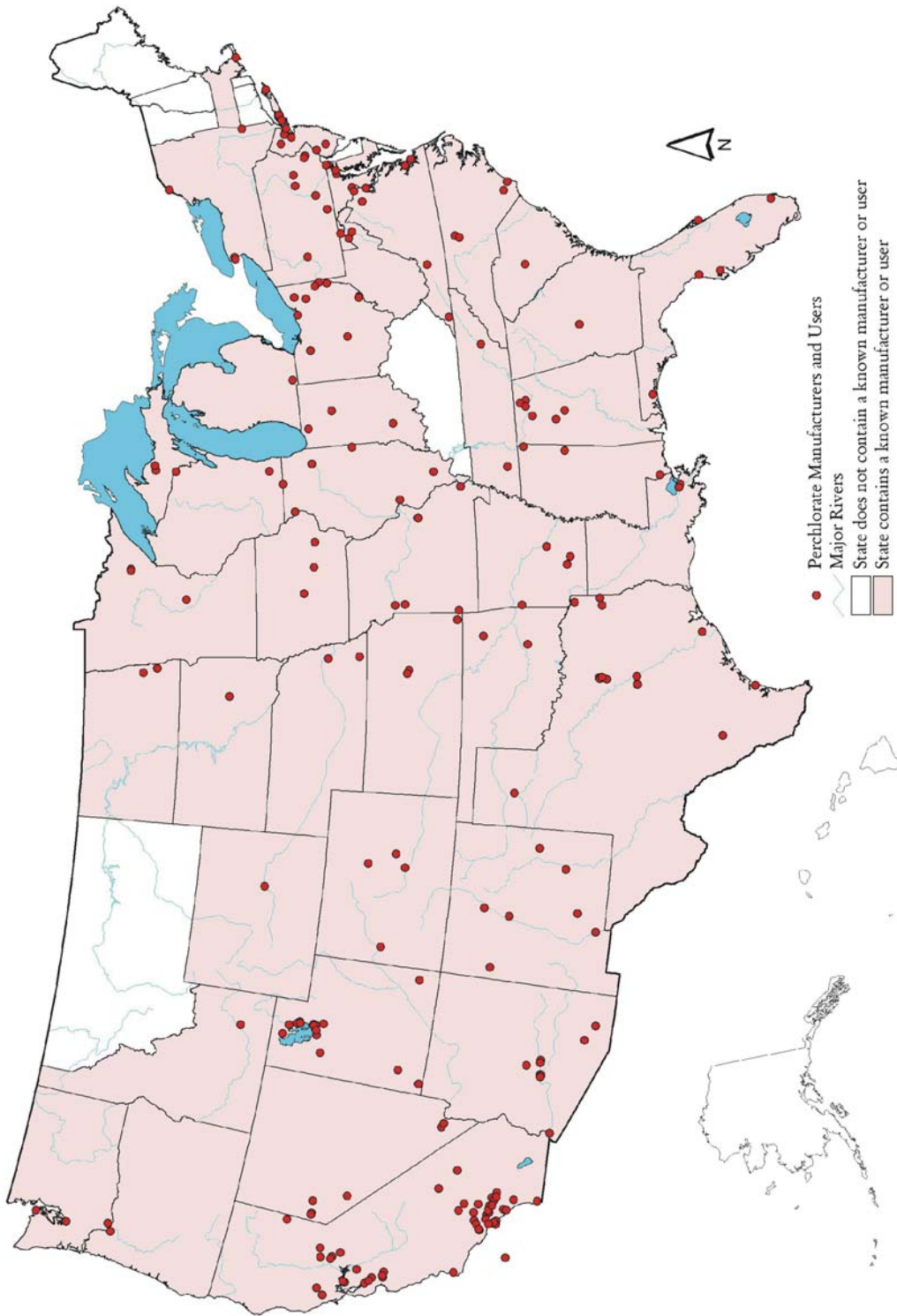
Figure 1-3. Locations of specific perchlorate manufacturers or users identified through responses to EPA Information Requests from current manufacturers (identifying shipments of at least 500 pounds in any year) and through investigations by state and local authorities (Mayer, 2001).

Legend:
- Perchlorate Manufacturers and Users
- Major Rivers
- State does not contain a known manufacturer or user
- State contains a known manufacturer or user
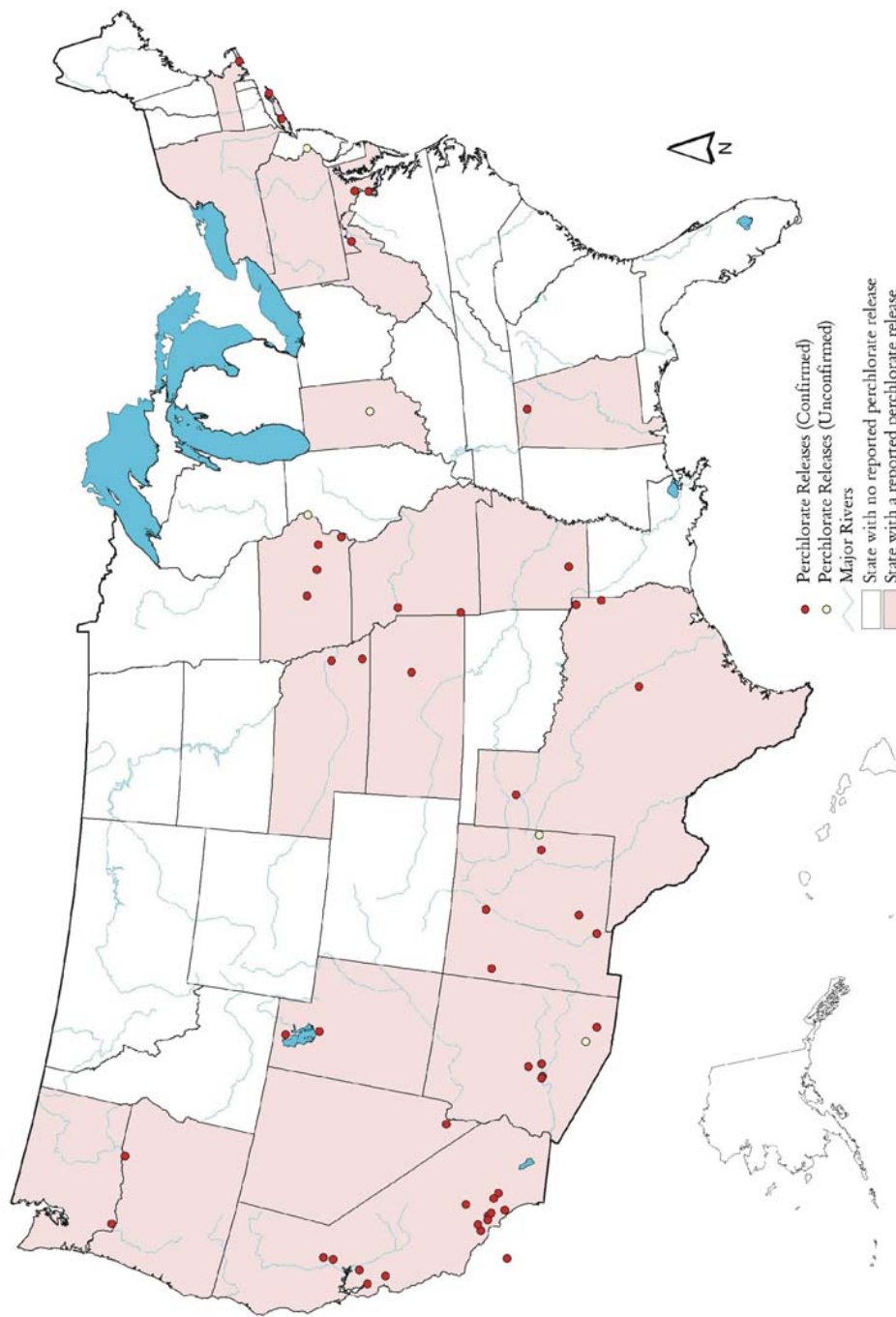
Figure 1-4. Locations of reported environmental releases of perchlorate to groundwater, surface water, or soil. Perchlorate measured in four water supplies in New Mexico, Iowa, Indiana, and Pennsylvania has been published in Siddiqui et al., 1998, but has not been confirmed independently by EPA or state authorities. Monitoring for perchlorate releases in most states is very limited or nonexistent (Mayer, 2001).

TABLE 1-1. OCCURRENCE AND POTENTIAL SOURCES OF PERCHLORATE RELEASES TO THE ENVIRONMENT AS OF NOVEMBER, 2OO1ª (Mayer, 2001)

State	Location	Suspected Source	Type of Contamination	Max. Conc. ppb
AL	Redstone Army Arsenal - NASA Marshall Space Flight Center Huntsville, AL	Propellant Manufacturing, Testing, Research, Disposal	Monitoring Well Springs/Seeps	19,000 37
AZ	Apache Nitrogen Products Benson, AZ	Explosives Manufacturing	Monitoring Well	670
AZ	Aerodyne Gila River Ind. Res., Chandler, AZ	Propellant Testing	Monitoring Well	18
AZ	Davis Monthan AFB Tucson, AZ	Explosives/Propellant Disposal	Soil	Not confirmed
AZ	Unidynamics Phoenix Inc. Phoenix Goodyear Airport, Goodyear, AZ	Explosives/Ordnance Manufacturing	Monitoring Well	80
AZ	Universal Propulsion Phoenix, AZ	Rocket Manufacturing	Soil	—
AZ	Unidynamics Phoenix Inc. Whiter Tanks Disposal Area Maricopa County, AZ	Explosives/Ordnance Disposal	Public Water Supply Well (Unconfirmed Report) Soil	(4) —
AR	Atlantic Research East Camden, AR	Rocket Manufacturing Disposal - Open Burn/Open Detonation	Monitoring Well Surface Water Soil	1,500 480,000 —
CA	Aerojet General also affects Mather AFB Rancho Cordova, CA	Rocket Manufacturing	Public Water Supply Well Monitoring Well	260 640,000
CA	Alpha Explosives Lincoln, CA	Explosives Manufacturing	Monitoring Well Reported in Surface Water	67,000
CA	Boeing/Rocketdyne, NASA at Santa Susana Field Lab U.S. DOE Santa Susana, CA	Rocket Research, Testing and Production	Monitoring Well	750
CA	Edwards AFB Jet Propulsion Lab, North Base Edwards, CA	Rocket Research	Monitoring Well	300
CA	El Toro Marine Corps Air Station Orange Co., CA	Explosives Disposal	Monitoring Well	380
CA	Lawrence Livermore National Laboratory Site 300 Tracy, CA	U.S. DOE Explosives Research	Monitoring Well	84

TABLE 1-1 (cont'd). OCCURRENCE AND POTENTIAL SOURCES OF PERCHLORATE RELEASES TO THE ENVIRONMENT AS OF NOVEMBER, 2OO1[a] (Mayer, 2001)

State	Location	Suspected Source	Type of Contamination	Max. Conc. ppb
CA	Lockheed Propulsion Upper Santa Ana Valley Redlands, CA	Rocket Manufacturing	Public Water Supply Well	87
CA	NASA - Jet Propulsion Lab Raymond Basin Pasadena, CA	Rocket Research	Public Water Supply Well	54
CA	Rialto, CA	Fireworks Facility (?) B.F. Goodrich (?) Rocket Research and Manufacturing	Public Water Supply Well (inactive)	811
CA	San Fernando Valley Glendale, CA	Grand Central Rocket (?) Rocket Manufacturing	Monitoring Well	84
CA	San Gabriel Valley Baldwin Park, CA	Aerojet Rocket Manufacturing	Public Water Supply Well Monitoring Well	159 2,180
CA	San Nicholas Island Ventura Co., CA	U.S. Navy Firing Range	Public Water Supply (Springs)	12
CA	Stringfellow Superfund Site Glen Avon, CA	Hazardous Waste Disposal Facility	Monitoring Well Private Well	682,000 37
CA	UTC (United Technologies) San Jose, CA	Rocket Testing	Monitoring Well	180,000
CA	Whittaker-Bermite Ordnance Santa Clarita, CA	Ordnance Manufacturing	Public Water Supply Well	47
CA	Whittaker Ordnance Hollister, CA	Ordnance Manufacturing	Private Well Monitoring Well	810 88
IN	American Water Works Service Greenwood, IN	Unknown Source	Public Water Supply Well (Unconfirmed Report)	(4)
IA	American Water Works Service Clinton, IA	Unknown Source	Public Water Supply Well (Unconfirmed Report)	(6)
IA	Ewart, IA	Unknown Source	Livestock Well	29
IA	Hills, IA	Unknown Source	Private Well	30
IA	Napier, IA	Agriculture (?)	Private Well	10
KS	Herington, KS	Ammunition Facility	Monitoring Well	9
MA	Massachusetts Military Res. Barnstable Co., MA	Disposal - Open Burn/ Open Detonation	Monitoring Well	300
MD	Naval Surface Warfare Center Indian Head, MD	Propellant Handling	Waste Discharge to Surface Water	>1,000

TABLE 1-1 (cont'd). OCCURRENCE AND POTENTIAL SOURCES OF PERCHLORATE RELEASES TO THE ENVIRONMENT AS OF NOVEMBER, 2OO1[a] (Mayer, 2001)

State	Location	Suspected Source	Type of Contamination	Max. Conc. ppb
MD	White Oak Fed. Research Center (Naval Surface Warfare Center) White Oak, MD	Propellant Handling	Monitoring Well	72
MO	ICI Explosives Joplin, MO	Explosives Facility	Monitoring Well	107,000
MO	Lake City Army Amm. Plant Independence, MO	Propellant Handling	Monitoring Well	70
NE	Lewiston, NE	Agricultural Chemical Facility	Shallow Private Well	5
NE	Mead, NE	Fireworks Facility	Monitoring Well	24
NV	Kerr-McGee/BMI Henderson, NV	Chemical Manufacturing	Public Water Supply Monitoring Well Surface Water	24 3,700,000 120,000
NV	PEPCON Henderson, NV	Chemical Manufacturing	Monitoring Well	600,000
NM	American Water Works Service Clovis, NM	Unknown	Public Water Supply Well (Unconfirmed Report)	(4)
NM	Ft. Wingate Depot Activity Gallup, NM	Explosives Disposal	Monitoring Well	2,860
NM	Holloman AFB Alamogordo, NM	Rocket Testing	Monitoring Well Seasonal Surface Water Soil	40 16,000 —
NM	Los Alamos National Lab Los Alamos, NM	U.S. Dept. of Energy Lab Chemicals	Public Water Supply Well Monitoring Well Deep Borehold Water	3 220 1,662
NM	Melrose Air Force Range Melrose, NM	Explosives	Public Water Supply Well	25
NM	White Sands Missile Range White Sands, NM	Rocket Testing	Monitoring Well Soil	21,000 —
NY	West Hampton Suffolk County, NY	Unknown Source(s)	Public Water Supply Well Monitoring Well	16 3,370
NY	Yaphank Suffolk County, NY	Fireworks	Private Well Monitoring Well	26 122
PA	American Water Works Service Yardley, PA	Unknown	Public Water Supply Well (Unconfirmed Report)	(5)

TABLE 1-1 (cont'd). OCCURRENCE AND POTENTIAL SOURCES OF PERCHLORATE RELEASES TO THE ENVIRONMENT AS OF NOVEMBER, 2OO1[a]
(Mayer, 2001)

State	Location	Suspected Source	Type of Contamination	Max. Conc. ppb
TX	Longhorn Army Ammunition Depot Karnak, TX	Propellant Handling	Monitoring Well Reported in Surface Water Soil	169,000 — —
TX	McGregor Naval Weapons Plant McGregor, TX	Propellant Handling	Monitoring Well Reported in Surface Water Soil	91,000 — —
TX	PANTEX Plant (USDOE) Amarillo, TX	Explosives	Monitoring Well	5
TX	Red River Army Depot Texarkana, TX	Propellant Handling	Monitoring Well	80
UT	Alliant Tech Systems Magna, UT	Rocket Manufacturing	Public Water Supply Well	16
UT	Thiokol Promontory, UT	Rocket Manufacturing	Well Supply Well (Inactive)	42
WA	Camp Bonneville near Vancouver, WA	Explosives/Propellant Disposal	Soil	—
WV	Allegheny Ballistics Lab Rocket Center, WV	Rocket Research, Production, Open Burn/Open Detonation	Surface Discharge of Groundwater Extraction	400

[a]Data reported to EPA Region 9 as of November 2001. All reports have been confirmed by federal, state, or county agencies except where noted. Soil concentrations are not listed.

1 manufactured ammonium perchlorate in Nevada were found to have released perchlorate to
2 groundwater resulting in low levels (4 to 24 ppb) in Lake Mead and the Colorado River. This
3 water is used for drinking, irrigation, and recreation for millions of people in Nevada, California,
4 Arizona, and by Native American tribes.
5   The concentrations reported in wells and surface water vary widely. At one facility near
6 Henderson, NV, perchlorate in groundwater monitoring wells was measured as high as 0.37%
7 (3.7 million ppb). The highest level of perchlorate reported in any public water supply well was
8 800 ppb in an inactive well in California. Few active public water supply wells have perchlorate
9 greater than 100 ppb, and none are reported at this level outside of California.

1   Perchlorate was found in a number of water supply wells on Long Island, NY, including

2   several downgradient from a fireworks facility. It has been speculated that the wide distribution

3   pattern of the New York contamination could be a result of low levels of perchlorate contained in

4   fertilizer imported from Chile (TRC Environmental Corporation, 1998; Urbansky, 2000; Suffolk

5   County Department of Health Services, 2001a,b). Agricultural chemicals also have been

6   implicated as a potential source of perchlorate contamination in Nebraska at a shallow well near

7   a speciality fertilizer facility (Williams, 2000). After state and federal officials in Region 7

8   added perchlorate analyses in their program testing hundreds of rural wells for fertilizers and

9   agricultural chemicals. Their results showed that fertilizer application to farmlands is an unlikely

10   source of perchlorate in Midwestern states.

11   In addition to discoveries at facilities involved with rocket propellants, explosives, and

12   fireworks, a number of perchlorate detections have been made at current or former military

13   facilities where propellants and explosives were disposed of by detonation and burning.

14   Cooperation from Department of Defense (DoD) and Department of Energy (DoE) officials will

15   continue to be important for examining these types of potential sources.

16   In the past three years, the increasing interest in investigating the environment has resulted

17   in increasing detections. It is likely that regional positive efforts at detection may largely explain

18   the distribution of known areas of release to the environment (Figure 1-4) when compared to the

19   potential distribution suggested in Figure 1-3. As the efforts for detection become more uniform

20   nationwide, the occurrence of perchlorate in the environment may more closely resemble the

21   pattern of perchlorate usage.

22   It is important to distinguish between minimum detection limit (MDL) and the minimum

23   reporting limit (MRL), which is also called the practical quantitation limit (PQL). MDLs are

24   calculated from the precision of replicate low level measurements and are assumed to reflect

25   99% confidence that a trace concentration above zero can be detected. MRLs are higher values

26   that reflect actual quantifiable concentrations. The EPA calculated and published an MDL for

27   Method 314 (Ion Chromatography) at 0.53 μg/L (Federal Register, 2000). This was derived

28   through the analysis of 7 replicate samples fortified at 2.0 μg/L. Based upon this result, an MRL

29   for perchlorate was established at 4.0 μ/L. Dionex, the manufacturer of the ion chromatography

30   column, published an MDL of 0.2 μg/L and MRL of 2.0 μg/L (Dionex, 2000).

1   Method 314 does not represent the lowest possible MRL or MDL. Unpublished

2   improvements in the ion chromatography method may lower the MRL to the sub-part per billion

3   level (Yates, 2001). Several research and commercial laboratories have been developing mass

4   spectrometry methods to detect sub-ppb levels of perchlorate (Urbansky et al., 1999; Magnuson

5   et al., 2000 a,b; Urbansky, 2000; Handy et al., 2000; Koester et al., 2000; Winkler, 2001). It is

6   reasonable to expect that a reliable sub-ppb MRL for perchlorate will be commercially available

7   in the very near future. The Agency encourages development of these emerging methods (e.g.,

8   LC/MS/MS) to eliminate interferences that can be encountered by extending IC methods for

9   low-level analysis in a variety of matrices (e.g., soil or plants and animal tissues). The market

10   demand for this capability may determine the commercial availability and expense of this

11   method. Regulatory pressure to ensure protection or water supplies and to maintain treatment

12   process control is also a factor driving the development of lower reporting limits for perchlorate.

13   Thorough method validation and quality assurance information must be complied to establish a

14   standard analytical method in the sub-ppb range for various media.

15

16

1.3   HEALTH AND ECOTOXICOLOGY RISK ASSESSMENTS—
HISTORICAL OVERVIEW

19   This section briefly summarizes how the assessments for the health and ecotoxicology risks

20   of perchlorate contamination have evolved. This document represents the revised assessment

21   that incorporates additional data and analyses recommended at the external peer review convened

22   by the Agency in February, 1999 (Research Triangle Institute, 1999).

23

1.3.1   Overview of Perchlorate Health Risk Assessment

25   The EPA Region 9 office requested evaluation of the toxicology data from the EPA

26   Superfund Technical Support Center (Stralka, 1992). The EPA Superfund Technical Support

27   Center issued a provisional RfD in 1992 (Dollarhide, 1992) and a revised provisional RfD in

28   1995 (Dollarhide, 1995) based on a literature review (Environmental Resources Management,

29   Inc., 1995) submitted by the Perchlorate Study Group (PSG). Ideally, an RfD is based on a

30   database that evaluates an array of endpoints that address potential toxicity during various critical

31   life stages, from developing fetus through adult and reproductive stages. The provisional RfD

1 values (1992 and 1995) were based on an acute study in which single doses of potassium

2 perchlorate caused the release of iodide (I⁻) from the thyroids of patients with Graves' disease, an

3 autoimmune condition that results in hyperthyroidism. It was difficult to establish a

4 dose-response for the effects on thyroid function from daily or repeated exposures in normal

5 humans from the data on patients with Graves' disease because of a variety of confounding

6 factors, including that the disease itself has effects; that often only a single exposure, rather than

7 repeated exposures was tested; that only one or two doses were employed; and that often the only

8 effect monitored was iodide release from the thyroid or control of the hyperthyroid state.

9 Nevertheless, a no-observed-adverse-effect-level (NOAEL) was determined to be

10 0.14 mg/kg-day based on release of iodide in the thyroid, followed by incomplete inhibition of

11 iodide uptake. Uncertainty factors that ranged from 300 to 1,000 were applied to account for

12 data missing on additional endpoints and extrapolations required to calculate a lifetime human

13 exposure level. The provisional RfD values issued are listed as such by EPA because they did

14 not undergo the internal EPA and external peer review required of estimates available on the

15 EPA's Integrated Risk Information System (IRIS). Standard assumptions for ingestion rate and

16 body weight were applied to the RfD to calculate the reported range in the groundwater cleanup

17 guidance levels of 4 to 18 ppb.

18   In recognition of the potential influence of the reduced analytical detection limit, a

19 reevaluation of the provisional 1992 and 1995 RfDs that serve as the basis of the provisional

20 action level was warranted. An external non-EPA peer review convened in March 1997 to assess

21 an analogous RfD derivation by an independent organization (Toxicology Excellence for Risk

22 Assessment, 1997) determined that the health effects and toxicity data were insufficient for a

23 credible quantitative risk analysis (Toxicology Excellence for Risk Assessment, 1998a). The

24 external peer review panel concluded that the limited database was insufficient to rule out effects

25 of perchlorate on other organs, so it could not be determined unequivocally that the effect on the

26 thyroid was the critical effect. In particular, the reviewers were concerned that developmental

27 toxicity, notably neurological development affected by hypothyroidism during pregnancy, could

28 be another critical effect of perchlorate that had not been examined adequately in studies to date.

29 In response to the March 1997 external peer review of the provisional RfD value, a subsequent

30 external peer review of experts was convened in May 1997 to recommend and prioritize a set of

31 studies to address the key data gaps and to reduce uncertainties in various extrapolations

1   (Toxicology Excellence for Risk Assessment, 1998b). The objective of the new studies is to

2   provide a comprehensive database that will support development of a robust RfD estimate that

3   reduces the uncertainties inherent in the provisional values. The strategical basis of the new

4   battery of toxicity studies is discussed in Chapter 3. These data featured prominently in the

5   external peer review draft of the assessment issued by the EPA in December 1998. At the

6   subsequent external peer review convened by the Agency in February 1999, recommendations

7   were made for additional studies and analyses, including completion of those on studies that were

8   only available as preliminary data at that time (Research Triangle Institute, 1999). The EPA

9   committed to a second external peer review and a revised risk assessment in order to benefit from

10   the additional insights that these data might bring to bear. The purpose of this current revised

11   document is to incorporate all of the data from new studies and to respond to recommendations

12   made at the previous external peer review.

13   Because the Agency is committed to utilizing the latest available science to support its

14   human and ecotoxicological risk estimates, the Office of Research and Development (ORD)

15   issued interim guidance in 1999 to its risk assessors and risk managers to be followed until this

16   revised assessment became publicly available (Noonan, 1999). The recommendation was to

17   continue using the standing provisional RfD range of 0.0001 to 0.0005 mg/kg-day for

18   perchlorate-related risk assessment activities because of the significant concerns and

19   uncertainties that remained to be addressed. This recommendation was based on the

20   determination that important new analyses on emerging data would likely have an impact on the

21   previously proposed health risk benchmark in the 1998 external review draft (U.S.

22   Environmental Protection Agency, 1998d) and that, while the new estimates would reflect greater

23   accuracy, the resultant revised risk estimate could be either higher or lower.

24   This recommendation helped to ensure that the Agency bases its risk management

25   decisions on the best available peer reviewed science and was in keeping with the full and open

26   participatory process embodied by the proposed series of external peer review workshops.

27   It should be noted that, due to the uncertainty of whether the final proposed revised oral human

28   health risk benchmark would increase or decrease based on the new data and analyses, the

29   standing provisional RfD range was the more conservative of the estimates available at the time

30   of the interim guidance and, therefore, more likely to be protective of public health. The

31   recommendation was also consistent with Agency practice that existing toxicity estimates remain

1   in effect until the review process to revise them is completed. The steps necessary to complete

2   this assessment are outlined in Section 1.4. Once completed, this assessment will be included on

3   the Agency's Integrated Risk Information System (IRIS).

4

1.3.2 Overview of Ecotoxicology Screening Level Assessment

6   The mobility and persistence of perchlorate discussed in the beginning of this chapter also

7   may pose a threat to ecological receptors and whole ecosystems either by direct harm to

8   organisms or by indirectly affecting their ability to survive and reproduce. There were very

9   limited data in 1997 with which to evaluate the effects of perchlorate on ecological systems; nor

10   were there data about the possible uptake of perchlorate into agricultural products irrigated by

11   contaminated water. Analytical tests had been derived to detect perchlorate in water, but little

12   work had been done to extend these methods to testing plant and animal tissues or food crops for

13   perchlorate.

14   Searches of available databases revealed minimal information on the ecological effects of

15   ammonium perchlorate or any of perchlorate's other salts. Little data exist to describe

16   perchlorate's ecological effects on various soil, sediment, or aquatic receptors, including aquatic

17   vertebrates, aquatic or sediment invertebrates, and bacteria or plants. The data that were

18   available suggested effects on thyroid-hormone-mediated development in the South African

19   clawed frog, *Xenopus laevis*, in the range of 50 to 100 ppm, and 1,000 ppm had been shown to

20   completely block the metamorphosis of tadpoles. Effects on development and population growth

21   also had been indicated in the freshwater lamprey at 100 ppm and the freshwater hydra at

22   350 ppm. Mortality was observed in cold-water trout (6,000 to 7,000 ppm) and *Daphnia magna*

23   (670 ppm). Effects on seed germination and growth of agricultural plants were reported at

24   10 ppm.

25   Under the auspices of the Ecological/Transport and Transformation Subcommittee of the

26   Interagency Perchlorate Steering Committee (IPSC, see Section 1.5), the U.S. Air Force (USAF)

27   Detachment 1, Human Systems Center, Brooks Air Force Base (AFB), in conjunction with EPA,

28   developed a proposal for a battery of screening-level bioassays in laboratory-reared organisms

29   representative of soil, sediment, and water column receptors, to evaluate dose-response

30   relationships. The identified tests focus on identifying gross (direct) toxicity tests whose

31   endpoints can include mortality, growth, and reproductive success. Bioassays with standard

1   protocols and general regulatory acceptance were chosen. Although these were screening-level

2   tests and provided only an indication of gross toxicity, they provided the dose-response

3   information required to make decisions about the need for a next tier of tests to be completed

4   (e.g., bioavailability, bioaccumulation, histopathology).

5   Additional new studies were recommended at the 1999 external peer review in the

6   ecotoxicology arena as well, and some additional data has become available that improves the

7   information base somewhat. Most significantly, additional data are available on effect levels in

8   aquatic animals, an aquatic plant, a terrestrial plant, and a soil invertebrate; and some of these

9   data are for chronic durations. In addition, surveys have been conducted at several sites of

10   known or suspected perchlorate contamination with analysis of environmental and biological

11   materials for perchlorate levels. While these new data have been incorporated in the current

12   revision and are described in Chapter 8, the knowledge in this arena requires that the ecological

13   assessment must still be characterized as a screening level rather than definitive. The number of

14   species is still quite low and the site surveys aimed only to describe the range of exposures at the

15   sites. The ecotoxicological review will undergo the same peer review process as the health risk

16   assessment that is described in Section 1.4.

17

18

19   **1.4   RISK CHARACTERIZATION AND REGULATORY AGENDA**

20   This section briefly describes pending regulatory activities that this evaluation and

21   characterization will likely influence. Particular emphasis is placed on the revised health risk

22   assessment and ecotoxicology assessments.

23

24   **1.4.1 U.S. Environmental Protection Agency Regulatory Plans**

25   The Safe Drinking Water Act (SDWA), enacted by Congress in 1974 and amended in 1986

26   and again in 1996 (U.S. Code, 1996), provides the basis for safeguarding public drinking water

27   systems from contaminants that pose a threat to public health. The purpose of the SDWA is to

28   protect public health by ensuring that public drinking water systems provide tap water that is safe

29   for drinking and bathing. Within EPA, the Office of Ground Water and Drinking Water

1 develops National Primary Drinking Water Regulations (NPDWR) to control the levels of

2 contaminants that may occur in public drinking water systems.

3 The 1996 amendments to the SDWA require EPA to publish a list of contaminants that are

4 not currently subject to a NPDWR and are known or anticipated to occur in public water systems.

5 This list, known as the Contaminant Candidate List (CCL), is the source of priority contaminants

6 for research, guidance development, regulatory determinations, and monitoring by the states.

7 The SDWA requires EPA to determine whether or not to regulate at least five contaminants from

8 the CCL by 2001. The CCL also must be reviewed and updated every 5 years; the next review is

9 scheduled for 2003.

10 With broad public input and consultation with the scientific community, a draft CCL was

11 published on October 6, 1997. The draft CCL specifically requested comment on whether to

12 include perchlorate on the CCL based on the limited information EPA had received on

13 perchlorate's occurrence in drinking water supplies at the time of publication. As a result of the

14 public comments and the obtainment of additional occurrence information, EPA determined that

15 sufficient information exists to raise concern over perchlorate's potential public health impact

16 and added perchlorate to the final CCL published on March 2, 1998.

17 The CCL consists of 50 chemical and 10 microbiological contaminants and is divided into

18 two categories: (1) contaminants for which sufficient information exists to begin to make

19 regulatory determinations in 2001, and (2) contaminants for which additional research and

20 occurrence information is necessary before regulatory determinations can be made. Perchlorate

21 falls into the latter category because of the need for additional research in the areas of health

22 effects, treatment technologies, analytical methods, and extent of occurrence.

23

24 **1.4.2 State Regulatory Plans**

25 The CA DHS and the California EPA Office of Environmental Health Hazard Assessment

26 (CA EPA OEHHA) reviewed the EPA risk assessment reports for perchlorate and established its

27 action level at 18 ppb based on the provisional RfD values from the EPA Superfund Technical

28 Support Center. The CA DHS advises water utilities to remove drinking water supplies from

29 service if they exceed the 18-ppb action level. If the contaminated source is not removed from

30 service because of system demands, and if drinking water provided by the utility exceeds the

31 action level, the CA DHS advises the utility to notify its customers. On August 1, 1997, the CA

1   DHS informed drinking water utilities of its intention to develop a regulation requiring

2   monitoring of perchlorate as an unregulated chemical. Legislative action to establish a state

3   drinking water standard for perchlorate by January 2000 (California Senate Bill 1033 [California

4   State Senate, 1998]) was vetoed by the governor after passage by both houses. The governor

5   supported prioritizing the regulation of perchlorate in drinking water but objected to the strict

6   time schedule required.

7   In July 2001, the CA EPA OEHHA posted a notice on its web site indicating that it was

8   initiating a risk assessment for perchlorate in connection with the development of a public health

9   goal (PHG) for a number of chemicals in drinking water (www.oehha.ca.gov/public_info/public/

10   phgannounc.html). PHGs are concentrations of chemicals in drinking water that are not

11   anticipated to produce adverse health effects following long-term exposures. These goals are

12   non-regulatory in nature but are to be used as the health basis with which to update the state

13   primary drinking water standards established by CA DHS for chemicals in drinking water subject

14   to regulation. A 45-day public comment period will be provided after posting, followed by a

15   public workshop. Scientific peer reviews are arranged through the University of California. The

16   overall process will include time for revisions, further public comment, and responses to

17   comments. The new PHGs are scheduled for publication in 2003.

18   New York, Arizona, and Texas also initially adopted the level of 18 ppb as their version of

19   advisory levels for water supply systems. Texas and Arizona health departments revised their

20   perchlorate advisory levels based on research presented in EPA's December 1998 External

21   Review Draft Toxicity Assessment. In July 1999, Texas arrived at a value of 22 ppb in drinking

22   water by calculating the exposure of a 15 kg child drinking 0.64 liter per day and using the

23   reference dose proposed in the 1998 EPA ERD document. Texas revised this value to 4 ppb in

24   October 2001 based in part on the interim ORD guidance (Noonan, 1999). Arizona derived a

25   14 ppb level in March 2000, based on a 15 kg child drinking 1 liter per day and using the

26   proposed RfD in the 1998 EPA ERD document. New York State has continued to use 18 ppb as

27   the advisory level for perchlorate in drinking water.

28   The Nevada Division of Environmental Protection (NDEP) has authority under Nevada

29   Water Pollution Control Regulations to address pollutants in soil or groundwater. The state's

30   Corrective Action Regulations direct NDEP to establish action levels for hazardous substances,

31   pollutants, or contaminants, using drinking water standards such as a maximum contaminant

1 level (MCL), health advisories, or background or protective levels (determined by IRIS or the

2 equivalent). In August 1997, Nevada determined that the action level of 18 ppb, as established

3 by EPA, would be the recommended action level for cleanup, pending a more current risk

4 assessment.

5

6

7 ## 1.5 SUMMARY

8   Perchlorate contamination is a concern for several reasons. First, there are uncertainties in

9 the toxicological database that is used to address the potential of perchlorate to produce human

10 health effects when present at low levels in drinking water. Additionally, the actual extent of

11 perchlorate occurrence in ground and surface waters and other media (soils or plant and animal

12 tissues) is unknown—a problem compounded by limits to the analytical detection method. The

13 efficacy of different treatment technologies for various water uses, including drinking and

14 irrigation, is also not well established. Finally, the nature and extent of ecological effects and

15 details about transport and transformation phenomenon in various environmental media have

16 been studied only superficially. EPA aims to more comprehensively characterize the risks to

17 human and ecological health from perchlorate contamination through the integrative approach

18 presented in Figure 1-5.

19   Thus, a number of key pieces of information and scientific advances are essential to

20 adequately characterize the risks of perchlorate contamination and to develop scientifically-based

21 management strategies that effectively mitigate the potential risks posed by perchlorate

22 contamination. Accurate characterization of exposures relies on reliable analytical detection

23 methods. The exposure estimates cannot be gauged with respect to their risk unless a robust

24 health risk estimate is available. Treatment technologies should be targeted to levels of concern

25 and tailored to the intended water use. Technology transfer is necessary so that all affected

26 parties and concerned citizens are appraised of accurate and reliable information that is

27 up-to-date with the evolving state-of-the-science. The purpose of the revised risk

28 characterizations presented in this document is to serve in this integrative approach by providing

29 more robust risk estimates than those that currently exist provisionally in order to better gauge

30 the potential human health and ecological impacts.

31
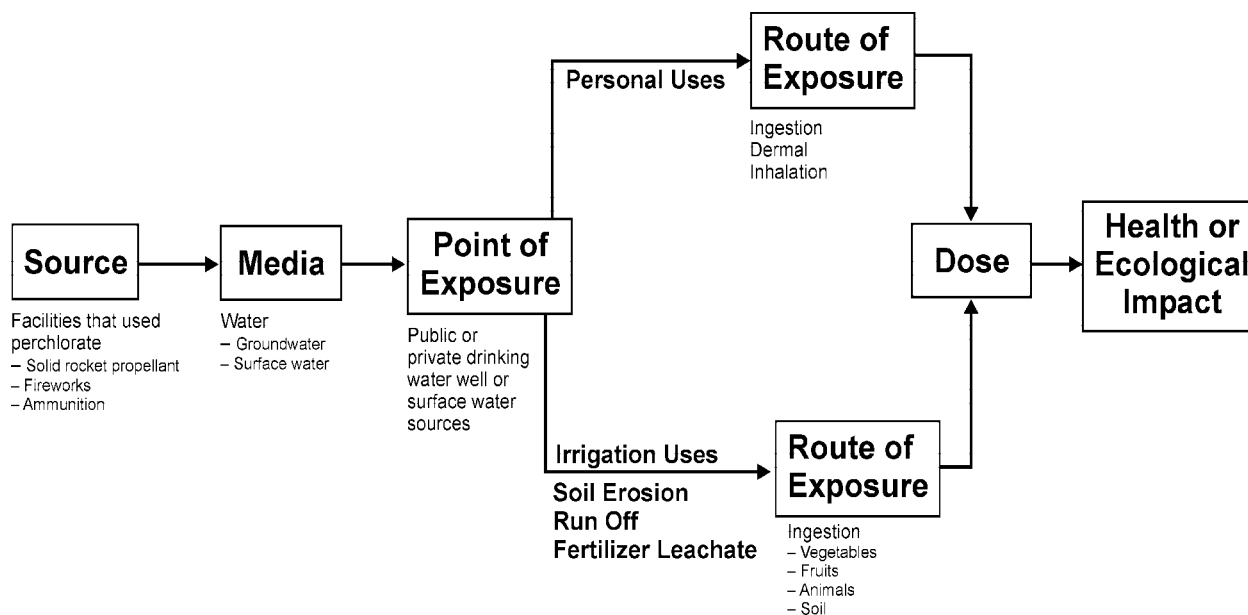
Figure 1-5. Considerations for comprehensive characterization of perchlorate contamination. (Modified from Underwood, 1998.)

1   The National Center for Environmental Assessment (NCEA) in the Office of Research and

2   Development (ORD) of EPA first evaluated the emerging information and new human

3   health/toxicity and ecotoxicity data from the testing strategy (see Chapter 3) and issued an

4   external peer review draft in December 1998. In February 1999, an external peer review

5   workshop was convened. The peer review panel endorsed the conceptual approach proposed by

6   NCEA and recommended additional studies and analyses. This revised risk characterization

7   document represents a response to those recommendations and includes data made available to

8   the EPA as of Fall 2001.

9   As with any risk assessment, incorporation of new data is an iterative process. Because of

10   regulatory schedule constraints, this assessment has gone forward with the recognition that new

11   data may warrant further revision at a future date. Data in additional analyses that are warranted

12   and which will be arriving in the period between the issuance of the external peer review draft

13   and the external peer review workshop are identified herein and may be presented at that

14   workshop.

1   Independent, external peer review of the study protocols, toxicity studies, and revised risk

2   assessment for perchlorate will be critical to ensuring that future decisions will be protective of

3   human health and that the potential for ecotoxicology is characterized appropriately. The IRIS

4   program will oversee the external peer review and has tasked a qualified contractor to manage

5   the peer review of technical issues related to the development of the human health and

6   ecotoxicology assessments, including system design, conduct of toxicity studies, statistical

7   analysis of data, designation of effect levels, selection of critical effect and uncertainty factors,

8   and risk characterization. The peer review will be conducted by a panel of technical experts

9   selected by contractors in ecotoxicology; neurotoxicology; developmental, reproductive, genetic,

10   and general toxicology; endocrinology; pathology; biostatistics; dose-response modeling; and

11   risk assessment.

12   The risk characterization assessment package, supporting studies, and study protocols for

13   the new data will be distributed to the peer review panel in advance of the peer review meeting.

14   The peer reviewers are charged with evaluating experimental protocols, performance, and results

15   for any new studies since 1999 in addition to how the data are used in this risk assessment. Peer

16   reviewers will independently review the risk assessment package and supporting studies and will

17   submit their written comments to the IRIS contractor prior to the peer review meeting. The IRIS

18   contractor will compile the peer reviewers comments and distribute them to all of the reviewers

19   prior to the meeting which will be held on March 5 and 6, 2002. Sacramento was selected as a

20   location for its accessibility to stakeholders and peer reviewers. The public will be invited to

21   attend and observe the peer review meeting and may obtain pre-meeting comments at that time.

22   Following the peer review meeting, the peer review panel will generate a report detailing their

23   comments on the reference dose package and supporting studies. NCEA then will generate a

24   responsiveness summary report that will discuss how comments made by the peer reviewers have

25   been addressed. The revised risk characterization will be issued subsequently by EPA as a final

26   IRIS assessment after Agency consensus review across offices and laboratories and a final IRIS

27   program clearance.

28   It should be noted that this assessment effort was accomplished in an expedited time frame

29   through the partnership and cooperation of a number of governmental entities. The IPSC was

30   formed in January 1998 to bring together government representatives from EPA; DoD; the

31   National Institute for Environmental Health Sciences (NIEHS); and affected state, tribal, and

1   local governments. Participation in IPSC also has been solicited from other governmental

2   entities. The purpose of the IPSC is to facilitate and coordinate accurate accounts of related

3   technological issues (occurrence, health effects, treatability, waste stream handling, analytical

4   detection, and ecological impacts) and to create information-transfer links for interagency and

5   intergovernmental activities regarding these areas of concern.

6   Figure 1-6 shows the structure of the IPSC, members of its executive committee, and

7   co-chairs of the subcommittees. Note that a subcommittee exists for each of the outstanding

8   controversial issues regarding perchlorate contamination. These are identified in the

9   comprehensive characterization framework presented in Figure 1-5. Research to obtain

10   additional data and the development of new methods and applications is underway in these

11   human health and ecotoxicology areas, as well as in most of the others, to ensure that the state-

12   of-the-science is brought to bear in addressing the unique issues of perchlorate contamination.

13   The IPSC collaborated in 1998 with EPA ORD NCEA on a draft report to a Congressional

14   committee that assesses the state-of-the-science on the health effects of perchlorate on humans

15   and the environment and the extent of perchlorate contamination. The report also contained

16   recommendations for future research to address emerging issues (U.S. Environmental Protection

17   Agency, 1998e). This report will be finalized and sent to Congress after the IRIS file is

18   completed. Updates on activities of IPSC can be found on the EPA Office of Water (OW) web

19   site at the following address: http://www.epa.gov./ogwdw/ccl/perchlor/perchlo.html. Discussion

20   papers presented by the IPSC present additional information on the areas (e.g., analytical and

21   treatment technology) that have not been discussed in detail herein.

22

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┌─────────────────────────────────┐
│      Executive Committee        │
│   Peter Grevatt (EPA-OSWER)     │
│    Annie Jarabek (EPA-NCEA)     │
│    Kevin Mayer (EPA-Region 9)   │
│  Lt Col Dan Rogers (DoD-USAF)   │
│     Karen Wirth (EPA-OW)        │
└─────────────────────────────────┘
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Health Effects / Toxicology Annie Jarabek (EPA-NCEA) Dave Mattie (DoD-USAF)	Ecological Impacts / Transport andTransformation Randy Bruins (EPA-NCEA) Cornell Long (DoD-USAF) Steven McCutcheon (EPA-NERL) Mark Sprenger (EPA-OERR)
Treatment Technology Jeff Cornell (DoD-USAF) Wayne Praskins (EPA-Region 9) Ed Urbansky (EPA-NRMRL)	Analytical Sanwat Chaudhuri (UT DEQ) Jackson Ellington (EPA-NERL) Howard Okamoto (Cal-DHS) Steve Pia (EPA-NERL) Carey Yates (DoD-USN)
Communications Larry Glidewell (DoD-USAF) Linda Tuxen (EPA-NCEA) Debbie Schechter (EPA-Region 9)	Occurrence Kevin Mayer (EPA-Region 9) Debra Tellez (EPA-Region 6)

Figure 1-6. **Structure and membership of the executive committee, subcommittees areas, and co-chairs of IPSC. The IPSC is designed to ensure an integrated approach to addressing the perchlorate contamination challenge and to provide accurate accounts of technical issues to stakeholders. (OSWER = Office of Solid Waste and Emergency Response, NCEA = National Center for Environmental Assessment, DoD = Department of Defense, USAF = U.S. Air Force, OW = Office of Water, NERL = National Exposure Research Laboratory, OERR = Office of Emergency Response and Remediation, NRMRL = National Risk Management Research Laboratory, Cal DHS = California Department of Health Services, USN = U.S. Navy, UT DEQ = Utah Department of Environmental Quality).**

2. PHYSICOCHEMICAL CHARACTERISTICS

1

2

3

4   This chapter provides an overview of the physicochemical properties of perchlorate. These

5   are important to understanding the persistence of perchlorate in the environment and to

6   understanding how perchlorate is processed in various plants and animals. Additional

7   toxicokinetic and toxicodynamic information can be found in Chapters 3 and 6; additional data

8   on transport and transformation, including biotransport, are discussed in Chapters 8 and 9.

9   In the solid state, the perchlorate anion has been determined by X-ray diffraction to have a

10   nearly perfect tetrahedral geometry with the four oxygen atoms at the vertices and the chlorine

11   atom at the center as shown in Figure 2-1. In aqueous solution, the geometry is probably

12   perfectly tetrahedral. The average chlorine-to-oxygen bond distance is 1.42 pm (Schilt, 1979b),

13   and the oxygen-to-oxygen distance is 2.43 pm. The partial molar ionic volume is 44.5 cm^3/mol

14   at 25 °C, compared with 36.7 for iodide.
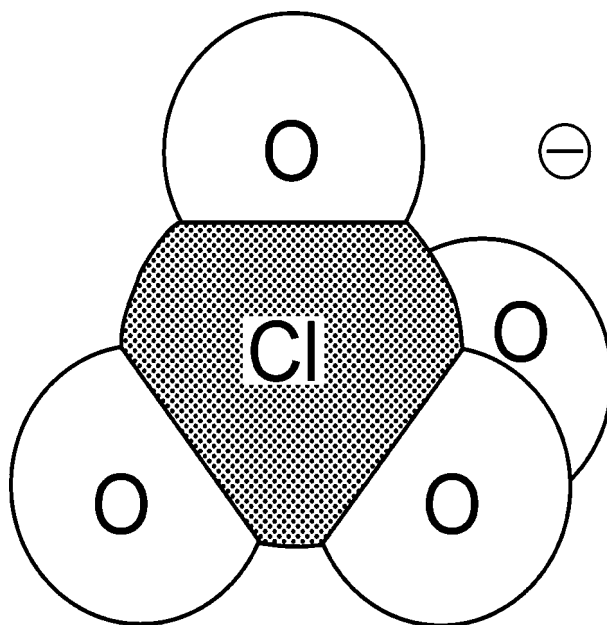
Figure 2-1. Chemical structure of perchlorate.

1   Perchlorate is widely known to be a very poor complexing agent and is used extensively as

2   a counter anion in studies of metal cation chemistry, especially in nonaquous solution (Urbansky,

3   1998). In this application, it is comparable with other noncomplexing or weakly ligating anions,

4   such as trifluoromethanesulfonate (triflate $[CF_3SO_3^-]$), tetrafluoroborate (BF_4^-), and, to a lesser

5   extent, nitrate (NO_3^-). Some exceptions are known, but are rare, such as some copper

6   compounds (Burke et al., 1982). All of these anions have a highly delocalized ($CF_3SO_3^-$, NO_3^-,

7   ClO_4^-) or sterically blocked (BF_4^-) monovalent anionic charge and large volume. The low charge

8   density reduces their affinity for cations and their extent of aquation (see Table 2-1).

9

10

**TABLE 2-1. GIBBS FREE ENERGIES OF FORMATION FOR
SELECTED ANIONS IN AQUEOUS SOLUTION (Urbansky, 1998)**

Anion	ΔG_f°, kJ Mol^{-1}
BF_4^-	$-1,490$
PO_4^{3-}	$-1,019$
SO_4^{2-}	-744
HCO_3^-	-587
OH^-	-157
Cl^-	-131
NO_3^-	-109
Br^-	-104
ClO_4^-	-8.5
ClO_3^-	-8.0

1   This low association with cations is responsible for the extremely high solubilities of perchlorate

2   salts in aqueous and nonaqueous media. As noted, the ammonium and the alkali metal salts of

3   perchlorate generally are readily soluble in water. Salts of the smaller univalent cations (i.e.,

4   ammonium [NH_4^+], lithium [Li^+], and sodium [Na^+]) are very soluble; whereas, those of the

5   larger univalent cations are less so (i.e., potassium [K^+], rubidium [Rb^+], and cesium [Cs^+]).

6   Quaternary ammonium salts are less soluble still. The outstanding example is sodium

1 perchlorate, which is extremely soluble (>8 mol dm^{-3}). Table 2-2 lists these solubilities as well

2 as other key physicochemical properties.

3

4

TABLE 2-2. PHYSICOCHEMICAL PROPERTIES OF AMMONIUM AND ALKALI METAL PERCHLORATES AT 25 °C (Schilt, 1979).

Physical Property	Magnitude of Physicochemical Property of Perchlorate					
	NH$_4$	Li	Na	K	Rb	Cs
Molecular Weight (g mol^{-1})	117.49	106.40	122.44	138.55		
Density	1.952	2.429	2.499	2.5298	2.9	3.327
Solubility (w/w %)						
Water	24.922	59.71	209.6	2.062	1.338	2.000
Methanol	6.862	182.25	51.36	0.105	0.000	0.093
Ethanol	1.907	151.76	14.71	0.012	0.009	0.011
n-Propanol	0.387	105.00	4.888	0.010	0.006	0.006
Acetone	2.260	136.52	51.745	0.155	0.095	0.150
Ethyl Acetate	0.032	95.12	9.649	0.001	0.016	0.000
Ethyl Ether	0.000	113.72	0.000	0.000	0.000	0.000
Thermochemical data						
$\Delta H_f^{''}$, kJ mol^{-1}	−290.4	−384.0	−385.7	−435.5	−434.7	−434.7
$\Delta G_f^{''}$, kJ mol^{-1}	−88.9[b]	−254[c]	−255[b]	−304	−306	−307
$\Delta S_f^{''}$, kJ mol^{-1}	186[b]	130[c]	142[b]	151	161	175
$\Delta H_{soln}^{''}$, kJ mol^{-1}	−26.6	26.1	14.7	50.6	56.8	55.6
Magnetic susceptibility ($\times 10^6$)	46.3	32.8	37.6	47.4	—	69.9
Molar refraction	17.22	—	13.58	15.27	—	—

[a]Thermochemical data converted from kcal/mol using 1,000 cal = 4.184 J.
[b]Weast (1989).
[c]Dean (1985).

1   Because of their large solubilities, the health risk assessment for perchlorate anion (ClO_4^-)

2 is appropriate for perchlorate salts, including ammonium perchlorate *[CASRN 7790-98-9]*,

3 sodium perchlorate *[CASRN 7601-89-0]*, potassium perchlorate *[CASRN 7778-74-7]*, and

1   lithium perchlorate *[CASRN 7791-03-9]*. The estimate is not appropriate to characterize the risk

2   of effects of perchloric acid ($HClO_4$) *[CASRN 7601-90-3]* because it is a strong acid, and the

3   dominant mode of toxicity is the irritating action of the hydrogen ion on skin and mucous

4   membranes.

5   Perchlorate can be a strong oxidizing agent under certain conditions as indicated by its high

6   reduction potential; therefore, the question has arisen as to whether or not it has the potential to

7   behave as an oxidant in biological systems. The thermodynamics of the halogen oxoanions and

8   oxoacids to participate in redox reactions are well understood. Under standard conditions in 1 M

9   acid, where the species is reduced to chloride, the oxidizing strength and standard reduction

10   potential ($E°$) increase as follows: $Cl_2 <$ HOCl $< HClO_2 < ClO_3^- < ClO_4^-$. The reduction

11   potentials for the oxoanions increase with increasing acidity or decreasing pH (i.e., they are

12   stronger oxidizing agents in acidic solution). Consider, for example, the reduction of

13   chlorine(VII) to chlorine(V) under both acidic and alkaline conditions. In 1.0 M H^+(aq) solution

14   (pH = 0),

15

16                $ClO_4^- + 2\ H^+ + 2\ e^- \rightarrow ClO_3^- + H_2O,$ $E° = 1.20$ V.   (2-1)

17

18   In 1.0 M OH^-(aq) solution (pH = 14),

19

20                $ClO_4^- + H_2O + 2\ e^- \rightarrow ClO_3^- + 2\ OH^-,$ $E° = 0.37$ V.   (2-2)
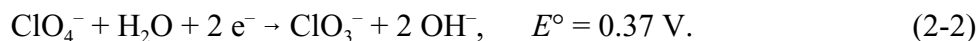
21

22   The effect of pH can be explained in terms of Le Châtelier's principle. In Reaction 2-1,

23   hydrogen ion is plentiful and acts a reactant; this drives the reaction forwards. In Reaction 2-2,

24   hydroxide ion is a product of the reaction and is already present at 1 M. This reduces the driving

25   force for this reaction to take place. The reaction is still spontaneous, as shown by the positive

26   value of $E°$; nonetheless, the driving force is considerably smaller for this case.

27   Thermodynamically, perchlorate is a stronger oxidant in the chlorine oxoanion series at the

28   extremes of the pH scale; however, such extremes are difficult to achieve in vivo (Tsui, 1998).

29   In Chapter 1, perchlorate anion was described as a nonlabile oxidant. Although the driving

30   force for reduction is very high, the activation energy required to start the process is also very

31   high. With the chlorine oxoanions, kinetic lability runs counter to the thermodynamic stability.

| 1 | That is, the most stable species, hypochlorite (ClO⁻), reacts fastest; whereas, the least stable |

1   That is, the most stable species, hypochlorite (ClO^-), reacts fastest; whereas, the least stable

2   species, perchlorate (ClO_4^-), reacts the slowest. It is important to note that the activation energy

3   required for the reduction of perchlorate to take place is a function not only of the perchlorate,

4   but also of the chemical nature of the reductant. With common reducing agents (e.g., thiosulfate,

5   sulfite, or ferrous ions), the activation energy is too high for any reaction to be observed. In fact,

6   this property (lack of lability) is exploited routinely in chemical studies where perchlorate salts

7   are used to control the ionic medium and strength, but do not themselves react.

8   An alternative way of expressing the thermodynamic driving force for a reaction is the

9   Gibbs free energy function. Although the driving force for redox reactions is often conveniently

10   expressed in terms of the potential, there are practical limitations to this approach. For example,

11   in the decomposition reaction of ammonium perchlorate in Equation 1-1, an electric potential

12   cannot be measured. The Gibbs free energy of reaction, ΔG°_{rxn}, is a measure of the energy

13   available to do work when a reaction is performed under constant pressure at standard state

14   conditions.[1] When ammonium perchlorate explodes, the gaseous products push against the

15   surrounding air and thereby perform expansion work on the atmosphere.[2] ΔG°_{rxn} specifies the

16   maximal nonexpansion mechanical work that can be obtained from a chemical reaction carried

17   out at constant temperature and pressure.[3] If the nonexpansion work is the electrical work of a

18   redox process, then an additional relationship applies (Equation 2-3), where n is the number of

19   electrons transferred; F is the Faraday constant, 96,485 C (mol e)$^{-1}$; and E° is the electric

20   potential for the reaction under standard state conditions.

21

---

[1]This is the case with reactions occurring exposed to the open air, rather than inside a sealed container. In a sealed container, where volume is constant and pressure changes, a different thermodynamic quantity, the Helmholtz free energy ΔA°_{rxn}, is used instead. The superscript circle indicates standard state conditions (i.e., solution concentrations of 1 mol dm^{-3} and gas pressures of 1 bar). All thermodynamic data are for a temperature of 298 K. All of the thermodynamic relationships herein apply at other conditions, and reference tables exist only for standard conditions. For other conditions, appropriate corrections must be made.

[2]Expansion work (W_{exp}) is significant only when a reaction has a net change in the number of gas molecules and can be calculated from the equation of state for a perfect gas: $W_{exp} = -P\Delta V = \Delta nRT$ (where P = pressure (atm), V = volume (L), n = number of moles, R = ideal gas constant (L atm k^{-1}), and T = temperature (K) and T and P are constant). For reactions occurring in the condensed phases, $W_{exp} \approx 0$.

[3]To obtain the maximal nonexpansion work, it is assumed that the process occurs reversibly so the loss of energy as heat is minimized. Although this is approximately true for an electrochemical cell, most chemical reactions do not take place under conditions that approach reversibility. For example, explosions are so irreversible because so much internal energy is lost as heat that the nonexpansion work is much smaller than ΔG°_{rxn}.

1 $$\Delta G^{\circ}_{\text{rxn}} = -w_{\text{max}} = -nFE^{\circ} \qquad (T, P \text{ constant}) \qquad\qquad (2\text{-}3)$$

2

3 The negative sign is necessary because the work done on the environment represents a loss of

4 free energy from the chemical system. Nonexpansion work includes, but is not limited to,

5 causing an electric current to flow or lifting an object against gravity. Whenever a chemical

6 reaction has the ability to do work on the surroundings, it will take place spontaneously.[4] $\Delta G^{\circ}_{\text{rxn}}$

7 is calculated as follows using Hess's law:

8

9 $$\Delta G^{\circ}_{\text{rxn}} = \Sigma\, \Delta G^{\circ}_{\text{f}} \text{ (all products)} - \Sigma\, \Delta G^{\circ}_{\text{f}} \text{ (all reactants)}. \qquad\qquad (2\text{-}4)$$

10

11 The Gibbs free energy of formation, $\Delta G^{\circ}_{\text{f}}$, is calculated for the formation of a compound from its

12 standard state as an element; consequently, $\Delta G^{\circ}_{\text{f}} = 0$ for $Cl_2(g)$ and $O_2(g)$. For Reaction 1-1,

13

14 $$\Delta G^{\circ}_{\text{rxn}} = 2\Delta G^{\circ}_{\text{f}} \, [N_2O(g)] + 8\Delta G^{\circ}_{\text{f}} \, [H_2O(g)] - 4\Delta G^{\circ}_{\text{f}} \, [NH_4ClO_4(s)]$$

15 $$= 2(104) + 8(-229) - 4(-89) \text{ kJ} = -1{,}268 \text{ kJ}. \qquad\qquad (2\text{-}5)$$

16

17 This large negative value for $\Delta G^{\circ}_{\text{rxn}}$ suggests that the decomposition of ammonium perchlorate is

18 spontaneous and has a large quantity of energy available to do work. When 4 moles (468 g) of

19 ammonium perchlorate decompose, enough energy is released to lift a 1 kg mass 130 km, heat

20 and completely boil 0.5 kg of water (starting from 25 °C) , or power a 100-W light bulb for 3.5 h.

21 Each molecule contains a large amount of potential chemical energy; however, a handful of

22 ammonium perchlorate will not spontaneously explode. The free energy is not released because

23 the reaction kinetics are too slow at room temperature—only an infinitesimal fraction of the

24 molecules possesses enough energy to reach the activation energy of the transition state at any

25 point. The activation energy for the reaction between an ammonium cation and a perchlorate

26 anion also is too great for a reaction to occur.

---

[4]Readers who have studied thermodynamics will recall that the determining factor for the spontaneity of a chemical process is a net increase in the entropy of the universe (i.e., $\Delta S^{\circ}_{\text{univ}} > 0$). It can be shown that $\Delta G^{\circ}_{\text{rxn}} = -T\Delta S^{\circ}_{\text{univ}}$; therefore, $\Delta S^{\circ}_{\text{univ}} > 0$ means $\Delta G^{\circ}_{\text{rxn}} < 0$, and $\Delta S^{\circ}_{\text{univ}} > 0$ means $\Delta G^{\circ}_{\text{rxn}} < 0$ (because $T > 0$). As a consequence of these relationships, it can be stated definitively that negative free energy available to do positive nonexpansion work is a measure of the thermodynamic spontaneity of a chemical reaction. This implies that any chemical reaction capable of performing positive nonexpansion work will occur spontaneously. Conversely, positive free energy suggests that the reverse reaction is spontaneous.

1   The distinction between thermodynamic spontaneity and kinetic lability must be

2   emphasized. A reaction with $\Delta G^\circ_{rxn} \ll 0$ and $E^\circ \gg 0$ is thermodynamically favored, but may be

3   so slow as to take virtually an infinite amount of time to occur (as is the case with most

4   perchlorate reductions). On the other hand, a reaction that occurs very quickly may have a very

5   small driving force. Reaction rates are fast when the combined internal energies of the reactants

6   closely approach the activation energy required to form the transition state. In a similar case, the

7   kinetic barrier (activation energy) explains why an open gas jet does not burst into flame until the

8   heat of a match is applied.

9   It is well established that, in aqueous solution, chlorine(I), chlorine(III), and chlorine(V)

10   species undergo their most facile reductions via nucleophilic attack at the chlorine atom rather

11   than at the oxygen atom. When oxoanions are dissolved in water, the rate of net oxygen atom

12   exchange (Equation 2-6) can be used to understand how reactions proceed:

13

14        $$OClO_n^- + H_2\text{\O} \rightleftharpoons \text{\O}ClO_n^- + H_2O, \quad \text{\O a labeled oxygen atom; } 0 \le n \le 3. \qquad (2\text{-}6)$$
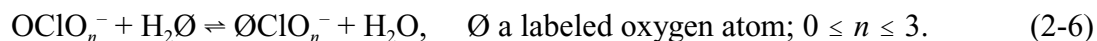
15

16   Reaction 2-6 proceeds through an associative mechanism in which the incoming water molecule

17   attacks the central chlorine atom. Consider the simplest example, hypochlorous acid, for which

18   the following mechanism is the accepted explanation (where Ø is again a labeled oxygen atom):

19
20 H_2O

21        $$HOCl + H_2\text{\O} \rightleftharpoons [HO\cdots Cl\cdots\text{\O}H_2]^\ddagger \rightleftharpoons OH^- + Cl\text{\O}H + H^+ \rightleftharpoons H_2O + Cl\text{\O}H \qquad (2\text{-}7)$$
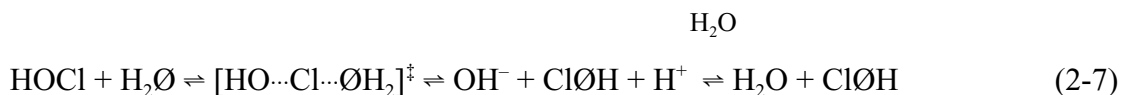
22

23   The aquated species $[HO\cdots Cl\cdots\text{\O}H_2]^\ddagger$ represents the activated complex and is the transition state

24   of Reaction 2-7; the proton is not directly transferred from the labeled water molecule to the

25   hydroxide that is part of the HOCl molecule. Rather, a proton is lost to the bulk water of the

26   solution form the activated complex, and another proton is gained. This activated complex may

27   revert back to reactants or proceed to products.[5] As the number of oxygen atoms increases, the

28   water has greater difficulty accessing the reaction site. The oxidation state of the chlorine

29   increases by 2 with each additional oxygen atom; accordingly, the chlorine becomes more and

---

[5]Note that $\Delta G^\circ_{rxn} = 0$ because the reactants and products are chemically identical. This suggests a process at equilibrium in which the forward and reverse rates are balanced.

1   more electron-poor and holds the oxygen atoms closer to share their electrons. (This factor will

2   be expanded on further when perchlorate is examined specifically.)

3   In perchlorate, which contains chlorine(VII), the central chlorine atom is sterically blocked

4   from the attack of an incoming reducing agent by the tetrahedrally oriented oxygen atoms.

5   As the oxidation state of the central chlorine atom increases, the strength of the chlorine-oxygen

6   bonds also increases. The electron-deficient chlorine(VII) draws electron density from the

7   oxygen ligands resulting in increased $O(p\pi) \rightarrow Cl(d\pi)$ back donation despite the high

8   electronegativity of the oxygen atoms. Increased O-Cl bond strength thus further complicates

9   oxoanion reduction by making oxygen-atom abstraction even more difficult.

10   Perchloric acid normally exhibits oxidizing behavior when heated and concentrated. When

11   cold and dilute, $HClO_4$ acts only as a strong Brønsted-Lowry acid with no more oxidizing

12   character than other mineral acids, such as sulfuric or hydrochloric acids. In the absence of free

13   H^+, as in vivo, a reducer or a catalyst with a lot of free potential energy would be requisite to

14   increase the rate (Tsui, 1998).

15   All observable perchlorate reductions reported in the literature are initiated via oxygen

16   atom abstraction by air-sensitive transition metal species (Urbansky, 1998). The metal cations

17   that react with perchlorate are all sensitive to atmospheric oxygen because they are strong

18   (thermodynamically) and labile (kinetically facile) reductants. None of these metal ions would

19   survive under human physiologic conditions. Certainly, any reductant capable of reacting with

20   perchlorate, such as $Ti^{III}(aq)$ (Earley et al., 2000), Ch_3ReO_2 (Abu-Omar et al., 1996), or certain

21   Re^V complexes (Abu-Omar et al., 2000) would not survive in a physiologic environment. Thus,

22   the activation energy to perchlorate reduction is so high that perchlorate cannot be expected to

23   act as an oxidant under human physiological conditions (i.e., dilute solution, moderate

24   temperatures, and nearly neutral pH). This is supported by absorption, distribution, metabolism,

25   and elimination studies that show perchlorate is excreted virtually unchanged after absorption

26   (see Chapters 3 and 6).

27   A catalyst increases the rate of chemical reactions by reducing the activation energy,

28   increasing the number of collisions, or properly orienting chemical reactants. Many catalysts

29   reduce the activation energy, but some have multiple effects. When a perchlorate ion collides

30   with a reducing agent, the two entities can recoil unaffected or they can interact. If they interact,

31   the entity they form is called an activated complex and is a transition state from which they can

1   separate or react. If they have sufficient internal energy (enough to overcome the activation

2   energy), the species will react. For perchlorate, this means an oxygen atom is transferred to the

3   reductant. If a catalyst is involved, it can act as an intermediate, removing oxygen atoms from

4   the perchlorate and transferring them to the reductant. In the case of the rhenium (V) catalysts,

5   the coordinated rhenium center accepts oxygen atoms from (and is therefore oxidized by) the

6   perchlorate. This oxidized species (now containing Re^{VII}) then transfers an oxygen atom to (and

7   is therefore reduced by) any reducing agent; however, the authors used thioethers and mercaptans

8   for this purpose (Abu-Omar et al., 2000). Of particular interest in this work was that the

9   conditions were not nearly so forcing as what is normally required for perchlorate reduction. The

10   reaction took place at roughly neutral pHs and ambient temperatures.

11   Some bacteria have catalysts (i.e, enzymes known as reductases) that allow the microbes to

12   use perchlorate as an oxidant in anaerobic metabolic pathways. Although oxygen is a stronger

13   oxidant than perchlorate, bacteria will utilize perchlorate under low-oxygen conditions. For

14   example, perchlorate-reducing monera use perchlorate reductases under conditions where

15   conventional inorganic chemistry suggests that perchlorate reduction should be imperceptibly

16   slow (Urbansky, 1998; Logan, 1998). Over the past few years, there has been a profusion of

17   work in this area, mostly slanted towards bioremediation (Coates et al., 1999, 2000; Logan, 2001;

18   Nzengung and Wang, 2000).

19   This chapter provides a brief summary of some physiochemical properties of the

20   perchlorate anion, especially the salient features that might bear on its environmental and

21   toxicological chemistry. Additional chemical issues are explored in some depth in Chapter 9 as

22   related to analysis of environmental samples. Additional chemical-specific issues as related to

23   the pharmacokinetics of perchlorate in organisms are discussed in Chapters 3 and 6.

24

3. TOXICOKINETICS/TOXICODYNAMICS AND MODE-OF-ACTION TESTING STRATEGY

This chapter explains the rationale that was the basis of the testing strategy which was designed to evaluate the potential critical targets for perchlorate and to establish a database robust enough to support a quantitative risk assessment. Aspects of the toxicokinetics and toxicodynamics of perchlorate and its interaction with the thyroid are discussed as the basis for the development of a testing strategy based on the mode of action of perchlorate. *Mode of action* is defined as a chemical's influence on molecular, cellular, and physiological functions (Federal Register, 1996; Wiltse and Dellarco, 1996). Understanding the mode of action helps to interpret the relevancy of the laboratory animal and human data to inform the most appropriate dose-response procedure (see Chapter 7).

As discussed in Chapter 2, perchlorate salts dissolve readily in water. The resultant anion is easily absorbed from the gastrointestinal tract. However, because of its high charge, neither perchlorate, nor other electrolytes applied from aqueous solution or aqueous media penetrate the skin readily (Scheuplein and Bronaugh, 1983). Uptake of inorganic ions such as perchlorate through the skin is typically less than 10% and frequently less than 1%. Exposure via inhalation of fumes or vapors is considered negligible because the vapor pressure of perchlorate salts and acids is low at room temperatures. The risk from exposure to particles would depend on the particle size distribution. Thus, the ingestion route is the major concern for the risk posed by the perchlorate contamination and is the focus of this characterization.

3.1 ABSORPTION, DISTRIBUTION, METABOLISM, AND ELIMINATION OF PERCHLORATE

Limited absorption, distribution, metabolism, and elimination (ADME) studies were in existence prior to the testing strategy discussed in Section 3.5. Although experimental studies in laboratory species and humans had been performed using radiolabeling techniques, most were at high concentrations, and the published data were expressed simply as thyroid:blood ratios of

1   radioactivity counts that provided no information on internal dose to biological tissues. Oral

2   drinking water administration, the most relevant to the contamination issue, was not the norm.

3   Time-course studies were very limited and essentially nonexistent for repeated administration.

4   More importantly, no data existed on the co-administration of iodide (I⁻) and perchlorate even

5   though data were necessary to develop a physiologically based pharmacokinetic model (Fisher,

6   1998a). The following section describes the limited pharmacokinetic information that was

7   considered when the data gap was highlighted during the development of protocols for the testing

8   strategy. The development of physiologically-based pharmacokinetic models that describe

9   ADME for perchlorate with data from the testing strategy will be discussed in Chapter 6.

10   Perchlorate appears to be eliminated rapidly, primarily in the urine (>90%), and virtually

11   unchanged from both rats (Eichler and Hackenthal, 1962) and humans (Anbar et al., 1959).

12   Durand (1938) measured urinary elimination from two human subjects who ingested 794 mg of

13   sodium perchlorate in 100 g of water. Urinary elimination accounted for 50% of the

14   administered dose within 5 hr and 95% within 48 hr. Half-lives have been reported for the rat

15   ranging from <8 hr (95% in 60 hr) to ≈20 hr (Wolff, 1998). Stanbury and Wyngaarden (1952)

16   reported that perchlorate appears in the urine within 10 to 15 min of oral dosing and that peak

17   plasma levels occur within 3 hr. Perchlorate was reported to undergo a two-phased urinary

18   elimination process in rats and calves. In rats, the first phase accounted for approximately 96%

19   of the administered dose and had a half-life of 1 to 2 hr. The second phase accounted for 4% and

20   had a half-life that ranged from 72 to 80 h. In calves, the first-phase half-life was reported to be

21   2 to 2.5 hr, and the second 23 to 27 hr (Selivanova et al., 1986, as cited in Allred, 1998). The

22   kinetics of long-term administration of perchlorate have not been characterized. The distribution

23   and metabolism of perchlorate and its relevance to potential toxicity in the thyroid will be

24   discussed in greater detail in Section 3.3 following discussions of iodine metabolism and thyroid

25   physiology in Section 3.2.

26

27   **3.1.1 Human Studies**

28   The majority of the human data on perchlorate ADME prior to the strategy was comprised

29   of the therapeutic case and clinical studies of Graves' disease patients described in Section 4.2.2.

30   These studies established the effect of perchlorate on the sodium (Na⁺)-iodide (I⁻) symporter

31   (NIS) but were of limited use in establishing quantitative dose-response relationships.

1   Anbar et al. (1959) demonstrated that perchlorate was not metabolized in humans. Four

2   patients were administered 200 mg (approximately 2.9 mg/kg using a default body weight of

3   70 kg) double-labeled $K^{36}Cl^{18}O_4$, and urine was collected 3 h after dosing. Perchlorate was found

4   to be excreted at approximately 200 μg/min in the urine. Total urine radioactivity was

5   distributed between ^{36}Cl , $^{36}Cl^{18}O_4^-$, $^{36}ClO_4^-$ and $^{36}Cl^-$ and indicated that perchlorate was excreted

6   unchanged in the urine. No human data existed with which to adequately characterize the

7   pharmacokinetics of perchlorate during steady-state, low-dose, repeated administration.

8

9   ## 3.1.2 Laboratory Animal Studies

10   Although the perchlorate discharge test has been performed in rats (Atterwill et al., 1987),

11   the procedure is very different than that used in humans and does not readily allow for

12   comparison or extrapolation. Rats are dosed intraperitoneally (ip) with 100 μL (1 μCi) $^{125}I^-$, then

13   dosed ip with potassium perchlorate at 5, 10, 25, or 50 mg/kg body weight from 1 to 6 h

14   afterwards. Results are expressed as thyroid:blood ratios, which differ from how most human

15   data are expressed. Additionally, the time points at which uptake is measured are highly

16   dissimilar to those used in human studies.

17   Anbar et al. (1959) also attempted to confirm the lack of perchlorate accumulation and lack

18   of metabolism in the thyroid in rats. White rats were injected ip with $^{36}KClO_4$, and the specific

19   activity per gram of tissue was measured at 30 min, 4 hr, and 12 hr. The activity was greatest in

20   the thyroid and peaked at 4 h. The salivary and adrenal glands also had high activity levels.

21   Rabbits also were tested; the thyroid activity levels were again the highest of any tissue and

22   peaked at 2 h. Rabbit testes had the next highest specific activities.

23   In one of the only co-administration studies, Anbar et al. (1959) simultaneously

24   administered $^{131}I^-$ and $^{36}ClO_4^-$ in equimolar concentrations. The thyroid:blood specific activity for

25   iodide was slightly higher than the ratio for perchlorate (1.80 and 1.69, respectively).

26   Halmi et al. (1956) examined iodide uptake in male Sprague-Dawley rats when active

27   transport was completely blocked via sodium perchlorate. The rats were first administered 6 mg

28   of propylthiouracil (PTU) subcutaneously to prevent iodide organification. Iodide uptake was

29   prevented by administration of 100, 200, or 400 mg sodium perchlorate with half of each dose

30   administered along with the PTU and the other half administered 45 min later with 5 to 50 μCi

31    $^{131}I^-$. The rats were sacrificed 1.0 to 1.5 h after the iodide administration. Perchlorate reduced the

1   thyroid:blood ratio from 22.7 to 0.45; radioiodide was found to account for 30% of the thyroid

2   gland volume when it entered the gland by diffusion alone. Rats sacrificed 4.0 to 4.5 h after

3   iodide administration produced similar results, indicating that equilibrium is reached prior to

4   1.0 to 1.5 h. The distribution of radioiodide in other tissues also was examined. Perchlorate did

5   not affect the organ:serum iodide ratios in the following organs: submaxillary salivary gland,

6   parotid salivary gland, pituitary gland, adrenal glands, testes, spleen, kidneys, lung, skin, or

7   diaphragm. However, perchlorate administration did affect the stomach wall:serum and gastric

8   juice:serum iodide ratios (0.36 and 0.75, respectively) compared with the ratios for controls

9   administered sodium chloride (1.45 and 15.8, respectively). This suggested a gastric iodide

10   pump subject to inhibition by perchlorate and, as will be discussed in Chapter 6, the

11   gastrointestinal tract is another tissue with NIS.

12   Goldman and Stanbury (1973) administered 0.1 μCi of the potassium salt of ^{36}Cl-labeled

13   perchlorate (K^{36}ClO$_4$) by ip injection to male Sprague-Dawley rats that had been maintained on a

14   low-iodine diet for 4.5 to 5.0 weeks prior to dosing (approximately 40 μg stable perchlorate per

15   injection). The radionucleide retention in the thyroid, expressed as percent of dose per gram of

16   tissue, was recorded at 2 h (6 rats), 4 h (5 rats), 8 h (6 rats), 24 h (6 rats), 48 h (6 rats), and 96 h

17   (5 rats). The peak was reported to appear around 4 h and then to fall to approximately 5% of this

18   peak value after approximately 96 h. An exponential function was used to estimate a half-life of

19   20 h. Urinary excretion data indicated that the disappearance rate from the plasma and thyroid

20   and the appearance rate in the urine corresponded closely although the question was raised as to

21   whether there is some curvilinearity to the urinary excretion, which may suggest limited

22   saturation. The retained dose and its standard deviation in tissues at 96 h were reported as

23   0.142 ± 0.1, 0.125 ± 0.09, 0.098 ± 0.03, 0.048 ± 0.04, and background for the thyroid, kidney,

24   spleen, liver, and brain, respectively.

25   Chow et al. (1969) compared the uptake of radiolabeled perchlorate and iodide ions with

26   stable ions in normal and thyroid-impaired rodents. Intact male Sprague-Dawley rats were

27   injected ip with 0.1, 0.2, or 5.0 meq/kg stable potassium perchlorate (14, 28, or 690 mg/kg,

28   respectively) 2 h prior to sacrifice. The specific activity of the chlorine label (^{36}Cl$^-$) was

29   25.2 μCi/mmol. Thyroid impairment was affected by pretreatment with thyroid-stimulating

30   hormone (TSH) (1 international unit [IU] TSH in 0.9% saline solution ip 18 h prior to

31   perchlorate administration), hypophysectomization (removal of the pituitary), TSH and

1   hypophysectomization, or PTU (0.1% PTU in drinking water for 2 weeks prior to perchlorate

2   administration). Perchlorate at the 0.1- and 0.2-meq/kg dose levels was found to preferentially

3   concentrate in the rat thyroid as compared to the plasma, and the concentration was related

4   inversely to dose. The high dose level did not result in the concentration of radiolabeled

5   perchlorate in the thyroid. Rats pretreated with TSH or PTU also concentrated perchlorate at the

6   lower dose levels. At the two lower levels, hypophysectomized rats were not able to concentrate

7   perchlorate compared with intact rats, but the thyroid perchlorate concentration at the high dose

8   level did not differ between intact and altered rats. In a second subset of the same study, rats

9   were exposed to 0.005, 0.01, 0.02, 0.05, or 0.10 meq/kg perchlorate (0.69, 1.4, 2.8, 6.9, or

10 14 mg/kg, respectively) under the same general conditions. The concentration of radiolabeled

11 perchlorate in the thyroid again was related inversely to perchlorate dose. Male albino guinea

12 pigs also were exposed to the same doses. The guinea pigs displayed the same relationships as

13 the rats, but concentrated more perchlorate in the thyroid compared to plasma levels.

14   Chow and Woodbury (1970) demonstrated that perchlorate is actively sequestered by the

15 thyroid gland at low doses but that the capacity of the symporter to actively sequester perchlorate

16 is exceeded at higher doses. Male Sprague-Dawley rats were functionally nephrectomized by

17 ligating the renal pedicle of both kidneys 24 h before the rats were sacrificed. Perchlorate was

18 administered as the radiolabeled potassium salt ($K^{36}ClO_4$) in solution by ip injection at 0.005,

19 0.1, or 2.0 mmol/kg stable potassium perchlorate (0.69, 14, and 280 mg/kg body weight,

20 respectively, assuming 0.266 kg body weight; actual weight 226 ± 4 g) 2 to 240 min before

21 sacrifice. A group of control rats received [^{14}C]-insulin, $^{35}SO_4^{-2}$ or $^{36}Cl^-$ 2 h prior to sacrifice to

22 determine thyroid follicle volume and intrafollicular membrane potential. Concentrations of

23 perchlorate in the thyroid and plasma were measured at 0.033, 0.067, 0.13, 0.2, 0.50, 1.0, 2.0,

24 and 4.0 h after sacrifice. Again, perchlorate was actively sequestered by the thyroid gland at the

25 low dose, but the capacity of the symporter to actively sequester perchlorate was exceeded at the

26 higher doses (e.g., the thyroid:plasma [milligrams per gram:milligrams per liter] ratios at 15 min

27 or 4 h post-dosing were 6.4, 0.69, and 0.36 or 13.8, 0.93, and 0.44 at the 0.5, 14.0, or

28 280.0 mg/kg doses, respectively). These data suggest that maximal inhibition by perchlorate of

29 active uptake of iodide probably occurs below 14 mg/kg potassium perchlorate (10.0 mg/kg as

30 perchlorate). If perchlorate-induced inhibition of active iodide uptake is substantial, iodide still

31 may enter the thyroid by diffusion, but in a smaller amount. Likewise, if inhibition of iodide

1   uptake by perchlorate is incomplete, then iodide still may be actively sequestered into the thyroid,

2   again in a smaller amount. Thus, perchlorate-induced thyroid hormone perturbations may

3   plateau in adult rats dosed with perchlorate greater than approximately 5 to 10 mg/kg of

4   perchlorate (Fisher, 1998a).

5   Wolff and Maurey (1962) demonstrated the competitive nature of the perchlorate inhibition

6   in sheep thyroid tissue slices incubated at 37 °C for 100 min. This study showed that the

7   K_m constants for anion accumulation and the K_i constants for inhibition of accumulation were

8   identical within the error of the method.

9   Eichler and Hackenthal (1962) presented perchlorate elimination data for male and female

10   Wistar rats dosed subcutaneously with 0.2, 1.0, or 6.0 of the $^{36}Cl^-$ sodium perchlorate salt

11   ($Na^{36}ClO_4$) per 100 g body weight (2, 10, or 60 mg/kg). The elimination curves showed nearly

12   linear, rapid excretion of perchlorate until 6 hr, at which time the curve slope started to decrease.

13   The rate of excretion increased with dose. The elimination rates of the different doses prior to

14   24 h were significantly different from each other but were similar after 24 h. Over 60 hr, 93.4 to

15   97.4% of the administered dose was recovered, again suggesting that perchlorate was not

16   metabolized.

17   In a recent review (Von Burg, 1995), perchlorate elimination curves in rats and calves were

18   described as biphasic in both species. For rats, 96% of administered perchlorate is eliminated

19   with a half-life of 1 to 2 hr. The second portion of the curve accounts for 4% of the dose, with

20   half-life of 72 to 80 hr. Calves have a faster overall rate of elimination, but the initial elimination

21   is slower. The first-phase half-life was 2.0 to 2.5 hr, and the second-phase half-life ranged from

22   23 to 27 hr.

23   An intravenous (iv) study performed at AFRL/HEST in Sprague-Dawley rats with

24   perchlorate to characterize its inhibition of iodide uptake supports the conclusion that there is

25   inhibition at low concentrations and there is a gradual plateau at higher concentrations (Meyer,

26   1998). Rats were dosed once by iv tail-vein injection with either 0.01, 0.1, 1.0, or 3.0 mg/kg of

27   cold (i.e., not radiolabeled) ammonium perchlorate in saline. Perchlorate was administered as

28   ammonium perchlorate, and the data are presented as milligrams perchlorate per kilogram body

29   weight. Two hours after dosing with perchlorate, the rats were dosed again by iv tail-vein

30   injection with 33 μg/kg ^{125}I dissolved in saline. Rats were sacrificed at selected times (n = 6 per

31   time point) up to 24 h. Total and free ^{125}I were measured in serum, thyroid, and urine.

1   Perchlorate serum, thyroid, tissue, and urine analyses began in January 1999 and are reported in

2   Chapter 6.   For control comparison, rats were dosed once by iv tail-vein injection with 33 μg/kg

3   nonradiolabeled iodide and ^{125}I mixed in physiologic saline. Rats (n = 6) were sacrificed at the

4   same selected time points up to 24 hr.

5   Table 3-1 shows the percent of inhibition of ^{125}I uptake as measured by bound ^{125}I in the

6   thyroid. Inhibition of ^{125}I uptake into the thyroid by perchlorate was measured by bound or free

7   [125]I in the thyroid at various time points after the single-dose of perchlorate. Because the ^{125}I was

8   administered 2 hr after dosing with ammonium perchlorate, these time points correspond to 4, 8,

9   and 11 h after dosing. The most profound inhibitory effects were found at the 1.0- and 3.0-mg

10   perchlorate/kg dose group; however, the trend for ^{125}I inhibition is evident at the 0.01- and

11   0.1-mg/kg levels (Meyer, 1998). By 24 h (26 h after dosing with perchlorate), inhibitory effects

12   on ^{125}I uptake were still observed at the 1.0- and 3.0-mg/kg dose groups.

13   Recovery of ^{125}I in urine 24 hr after dosing with [125]I (26 h after ammonium perchlorate) was

14   between 79 and 88% for control ^{125}I-dosed rats and perchlorate-dosed rats. The control ^{125}I-dosed

15   rats excreted 79.5% (SD ± 5.50) of the ^{125}I dose over the 24-hr period; whereas, the perchlorate-

16   dosed rats excreted 87% (SD ± 7.84), 86% (SD ± 4.47), 87.8 (SD ± 20.20) and 79.3 (SD ± 10.58)

17   of the ^{125}I dose in urine at the 0.01-, 0.1-, 1.0-, and 3.0-mg/kg dose levels, respectively. The

18   amount of ^{125}I in serum was elevated in the perchlorate-dosed animals compared to the control

19   ^{125}I-dosed rats for up to 6 hr in all dose groups, suggesting that thyroid function was altered by

20   perchlorate and that a transient "discharge" of organified ^{125}I occurred as reported in studies

21   summarized in Chapter 3. Free ^{125}I levels in serum were similar between perchlorate-dosed and

22   control ^{125}I-dosed rats (Meyer, 1998). These results are consistent with those of Chow et al.

23   (1969) and Chow and Woodbury (1970). The pattern for the inhibition of iodide uptake, albeit

24   only after a single dose, is strikingly similar to the patterns shown for the thyroid hormone

25   decreases. Consequently, data on the species differences (i.e., rat versus human in particular) in

26   perchlorate inhibition of the symporter will provide a basis for evaluating the degree of

27   uncertainty that should be applied when utilizing laboratory animal data as the model for humans

28   (see Chapter 7).

29   Repeated dose studies in rats (Fisher, 1998a) and in humans (Channel, 1998a) to establish

30   the kinetics of perchlorate at steady-state performed by AFRL/HEST to further characterize the

31   inhibition of iodide uptake by perchlorate are discussed in Chapter 6.

TABLE 3-1. PERCENT INHIBITION OF IODIDE UPTAKE IN THE THYROID GLAND OF SD RATS DOSED WITH PERCHLORATE (Meyer, 1998)

Time Points[a]	Dose (mg perchlorate/kg)	[Iodide] (μg/g)	Percentage of Inhibition
2 hr	Control[b]	24.4	—
	0.01	21.3	13
	0.1	18.6	24
	1	7.4	70
	3	2.99	88
6 hr	Control[b]	46.5	—
	0.01	36.7	21
	0.1	32.0	31
	1	19.2	59
	3	9.13	80
9 hr	Control[b]	55	—
	0.01	49.2	11
	0.1	39.2	29
	1	24.7	55
	3	10.0	82

[a]Time points correspond to dosing with ^{125}I and to 4, 6, and 11 hr after dosing with ammonium perchlorate.
[b]Dosed with only iodide (33 μg/kg).

3.2 IODINE METABOLISM AND THYROID PHYSIOLOGY

Iodine plays a central role in thyroid physiology as both a constituent of thyroid hormones and a regulator of thyroid gland function. Like perchlorate, iodine is absorbed efficiently from the gastrointestinal tract. Iodine in organic form is converted mostly to iodide before absorption (Cavalieri, 1997). The kidneys account for about two-thirds of the iodide cleared from plasma and more than 90% of the iodide cleared from the body. Sweat and breast milk account for various fractions of iodide loss, and fecal elimination constitutes approximately 1% of total body iodide clearance.

1   The thyroid gland concentrates iodide against an electrochemical gradient by a carrier-

2   mediated mechanism driven by adenosine triphosphate (ATP). The activation energy required

3   for perchlorate reduction is so high that it cannot act as an oxidant under physiological conditions

4   (i.e., dilute solution, moderate temperatures, and neutral pH). Plasma membrane experiments

5   indicate that the sodium cation (Na^+) and iodide cotransport are electrogenic, with a

6   thermodynamically downhill transport of approximately two Na^+ ions driving one iodide ion

7   against its electrochemical gradient into the cell. The transport is sensitive to ouabain, an

8   inhibitor of ATPase. The molecule responsible for the transport of iodide has been named the

9   *sodium (Nat)/iodide (I) symporter or NIS*. The thyroid thus has a specialized ability to

10   concentrate iodide selectively from the surroundings where the concentration is very low (10^{-8} to

11   10^{-7} M) and where the concentration of chloride ions is in the order of 0.01 to 0.1 M. The

12   transport is "active," not only by electrochemical criteria, but also by metabolic ones: it does not

13   occur in the cold, it requires oxygen, and, as mentioned, it is a function of the ATP level.

14   In addition to the thyroid, other organs that can concentrate iodide include the salivary glands,

15   gastric mucosa, choroid plexus, mammary glands, and the placenta. Iodide secreted into the

16   saliva and gastric juice is reabsorbed in the small intestine (Cavalieri, 1997).

17   Nevertheless, it is essentially only in the thyroid that the newly concentrated iodide can be

18   metabolized further to form thyroid hormone; and, only in the thyroid, does TSH regulate the

19   process. Thyroid hormones play numerous and profound roles in regulating metabolism, growth,

20   development, and maintenance of homeostasis. It is generally thought that these actions result

21   from the effects of the thyroid hormones on protein synthesis (Hill et al., 1989).

22   Figure 3-1 shows a schematic representation of thyroid hormone biosynthesis and secretion

23   in a single thyroid follicular cell. The thyroid hormones are stored as amino acid residues in

24   thyroglobulin (Tg), a protein constituting most of the colloid in the thyroid follicles. In situ, the

25   follicular cell displays functional and structural polarity: the vascular space is at the bottom, and

26   the lumen of the follicle is at the top. The striated circle straddling the basolateral membrane

27   represents the iodide transporter. The process of thyroid hormone biosynthesis is first stimulated

28   by TSH binding to the follicular cell TSH receptor and cyclic adenosine monophosphate (cAMP)

29   activation (Hard, 1998). The protein portion of Tg is synthesized on rough endoplasmic

30   reticulum (ER), and carbohydrate moieties are added by the Golgi apparatus (GA).

31   Thyroglobulin proceeds to the apical surface in secretory vesicles (small open circles) that
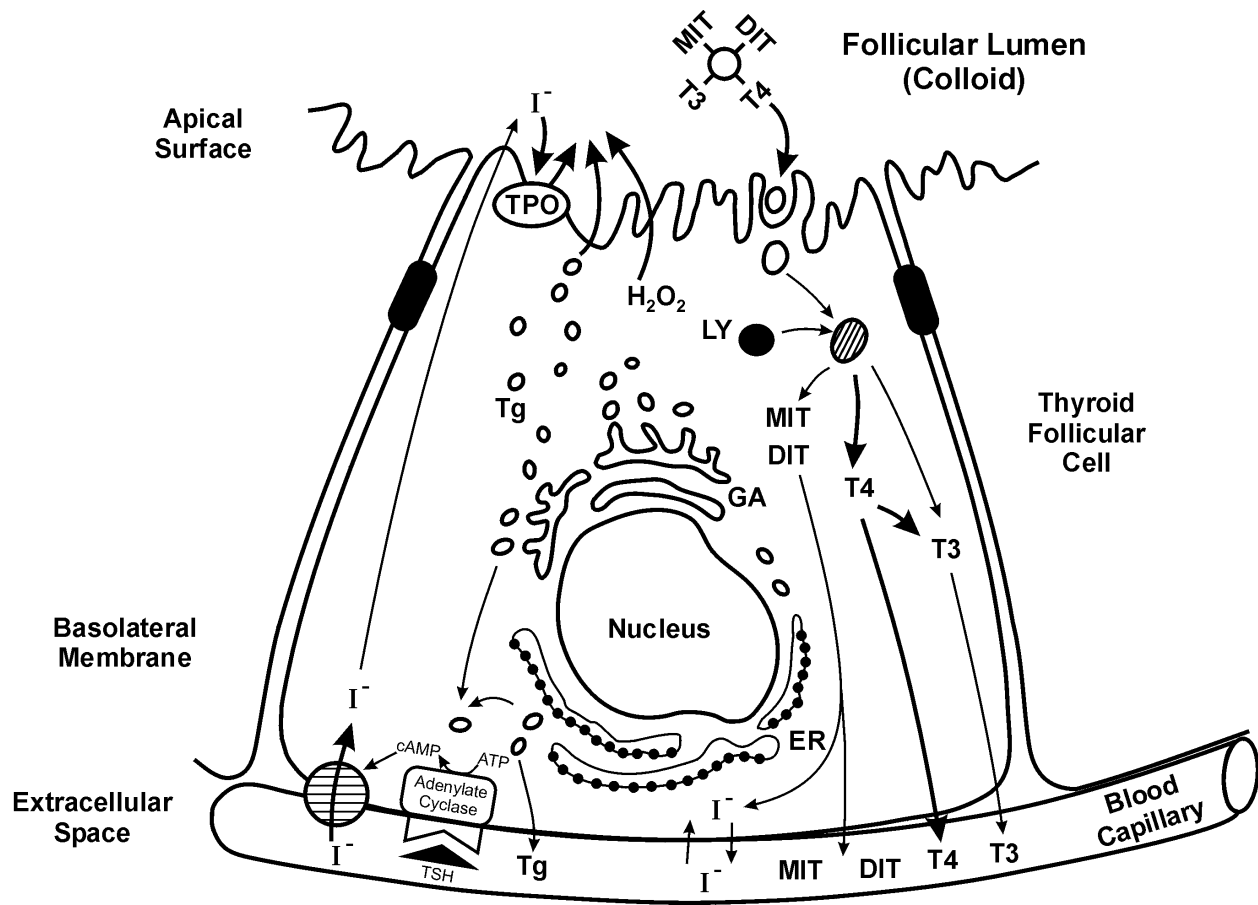
Figure 3-1. Schematic representation of thyroid hormone biosynthesis and secretion in a single thyroid follicular cell. (Modified from Hill et al., 1989; Cavalieri, 1997; and Fisher, 1996.)

1 fuse with the cell membrane and discharge their contents into the follicular lumen. Iodide enters

2 the cell by active transport, and then, at the apical surface, is oxidized by thyroid peroxidase

3 (TPO). The hydrogen-peroxide-generating system is represented by hydrogen peroxide (H_2O_2).

4 Organification occurs at or near this apical cell-colloid interface; the oxidized iodide is

5 incorporated into tyrosine residues in peptide linkage in Tg. Two iodinated tyrosyl groups couple

6 in ether linkage to form tetraiodothyronine (T4), which initially remains trapped in Tg. Hormone

7 secretion first involves pinocytosis of colloid-containing iodinated Tg (large open circle) at the

8 apical border of the follicular lumen and resolution into vesicles that fuse with lysosomes (LY,

9 dark circle). Lysosome proteolysis (striated circle) then converts Tg to amino acids, T4,

1	triiodothyronine (T3), diiodotyrosine (DIT) and monoiodotyrosine (MIT). Iodotryosine
2	dehalogenase regenerates iodide from MIT and DIT for reuse within the thyroid or release into
3	the blood, accounting for the iodide leak in the chronic state of iodine excess in certain thyroid
4	disorders. Type I iodothyronine deiodinase converts a fraction of the free T4 to T3. Both
5	hormones (T4 and T3) are released into the blood circulation by a process that is not well
6	understood. The thyroid also releases Tg, of which some is iodinated and some uniodinated as
7	newly synthesized protein.
8	Although T4 is by far the major hormone secreted by the thyroid (typically at 8 to 10 times
9	the rate of T3), T4 is considered a prohormone because about 33% of the T4 secreted undergoes
10	5'-deiodination to T3 in the peripheral tissues and T3 is about fourfold more potent than T4.
11	Another 40% undergoes deiodination of the inner ring to yield the inactive material, reverse
12	triiodothyronine (rT3), which recently has been postulated to play an inhibitory role on the
13	conversion of T4 to T3. T3 is regarded as the active hormone because it is the form that appears
14	to activate a response by nuclear DNA. Upon entering the circulation, both T4 and T3 are bound
15	and transported in strong, but not covalent, association with plasma proteins.
16	The major plasma-protein carrier in humans is thyroxine-binding globulin, a glycoprotein
17	with a very high affinity for T4 and a lower affinity for T3. In rats, the T4 and T3 are bound to
18	prealbumin (PA) or albumin with a weaker attachment. Control of the circulating concentrations
19	of these hormones is regulated primarily by a negative feedback involving three organs: (1) the
20	thyroid, which produces thyroid hormone, and (2) the pituitary gland and (3) hypothalamus,
21	which respond to and help maintain optimal T3 and T4 levels (Hill et al., 1998). Figure 3-2
22	shows the schematic for this hypothalamic-pituitary-axis and the feedback mechanisms.
23	The hypothalamus stimulates the pituitary gland through thyrotropin-releasing hormone
24	(TRH) to produce TSH, which prompts the thyroid to produce T4 and T3. Once secreted into the
25	blood, T4 and T3 are bound to plasma proteins (thyroid-binding globulin [TBG] in humans or
26	prealbumin [PA] and albumin in rats). In addition to the aforementioned conversion of T4 to T3
27	in peripheral tissues, thyroid hormone also is metabolized irreversibly in the liver by uridine
28	diphosphyl glucuronosyl transferases (UDPGTs) to either glucuronic (T4) or sulfate (mainly T3)
29	conjugates that are excreted in bile. A portion of the conjugated material is hydrolyzed in the
30	intestine, and the free hormones thus released are reabsorbed into the blood via enterohepatic
31	circulation. The remaining portion of the conjugated material is excreted in the feces.

Influences from Periphery
via Nervous System

(-)

Hypothalamus
(Paraventricular Nuclei)

TRH (+)

(-)

Pituitary
(Thyrotroph)

TSH (+)

Thyroid

Thyroglobulin

Peroxidase

I^- I → I_2 + Tyrosine → MIT DIT → T4, T3

- - - Deiodination - - -

Plasma/Blood

T4, T3 $\underset{TBG}{\overset{PA}{\rightleftarrows}}$ PP-TH

Bound to
Plasma
Proteins

I^- Trapping   Organic Binding-coupling   Proteolysis
T4, T3 Release

(Thiocyanate
Perchlorate)

(Thiourea, PTU
Sulfonamides
Methimazole
Aminotrazole)

(Iodide-
Excess)

(+)

Certain Tissues
T4 → T3

Target Tissue
(Nuclear Receptor)

**Liver
UDPGTs**
↓
T4-GLUC

**Bilary
Excretion**

Figure 3-2.   Schematic of the hypothalamic-pituitary-thyroid axis and feedback
mechanisms (PP-TH = plasma protein-thyroid hormone, PTU =
propylthiouracil, UDPGT = uridine diphosphyl glucuronosyl transferase,
T4 GLUC = T4-glucuronide conjugate). (Modified from U.S. Environmental
Protection Agency, 1998a; Hill et al., 1998; and Capen, 1997).

1   Cells in the hypothalamus and pituitary gland respond to levels of circulating T4 and T3

2   such that when thyroid production levels are high, there is a signal to reduce the output of (TRH)

3   and TSH. Similarly, when thyroid hormone levels are low, the pituitary is prompted to deliver

4   more TSH to the thyroid in order to increase the output of T4 and T3. This negative feedback

5   loop helps the body respond to varying demands for thyroid hormone and to maintain hormone

6   homeostasis. Thus circulating T4, T3, and TSH are monitored readily in experimental animals

7   and humans and so may serve as biomarkers of exposure to and indicators of the effects from

8   agents that disrupt the status of the hypothalamic-pituitary-thyroid axis (U.S. Environmental

9   Protection Agency, 1998a).

10   In the absence of thyroid-binding globulin, as in the rat and mouse, a greater fraction of

11   thyroid hormone is free of protein binding and subject to metabolism and removal from the body.

12   As a consequence, the half-life of T4 in the rat is only about 1 to 24 hr, in contrast to the 6 to

13   7 day half-life in humans. Rats compensate for the increased turnover rate by secreting more

14   TSH from the pituitary gland. Table 3-2 provides the interspecies and intraspecies differences in

15   both thyroid hormone and gland structure between rats and humans. The consequences of

16   disrupting the status of the hypothalamic-pituitary-axis will be discussed in Section 3.4.

17

18

19   ## 3.3 TOXICOKINETICS OF PERCHLORATE

20   Because of the complex anatomy of the thyroid follicle, all of the locations where

21   perchlorate inhibition is exerted remain to be established (Wolff, 1998). Perchlorate has been

22   established as a competitive inhibitor of iodide uptake across the basolateral membrane (i.e., acts

23   by the inhibition at NIS). Figure 3-3 shows a comparison of the molecular dimensions of

24   perchlorate and iodide. The following potency series was constructed for monovalent anion-

25   based inhibition of iodide transport in thyroid slices: $TcO_4^- \geq ClO_4^- > ReO_4^- > SCN^- > BF_4^- > I^- >$

26    $NO_3^- > Br^- > Cl^-$ (Wolff, 1998). However, it is not clear whether this anion sequence, measured

27   at very high concentrations, has any mechanistic relation to what occurs at low concentrations in

28   the thyroid. It is important to determine which solution properties of the anions determine this

29   sequence (e.g., crystal radius, hydrated radius, hydration enthalpy, charge density).   Strong base

30   anion-exchange resins (usually a large cation with a weak field) exhibit a marked preference for

31   ClO_4^- (e.g., compared to Cl^-); thus, it seems likely that selectivity for iodide or perchlorate in the

TABLE 3-2. INTERSPECIES AND INTRASPECIES DIFFERENCES IN THYROID STRUCTURE AND T3, T4, AND TSH HORMONES (U.S. Environmental Protection Agency, 1998a)

Parameter	Human	Rat
Thyroxine-binding globulin	Present	Essentially absent
T_4 Half-life	5 to 6 Days	0.5 to 1 Day
T_3 Half-life	1 Day	0.25 Day
T_4 Production rate/kg body weight	1 ×	10 × that in humans
TSH	1 ×	6 to 60 × that in humans
Follicular cell morphology	Low cuboidal	Cuboidal
Sex differences		
Serum TSH	$M^a = F^a$	$M \leq 2 \times F$
Cancer sensitivity	$F = 2.5 \times M$	$M > F$

[a]M = male, F = female.

Perchlorate (ClO$_4^-$):

**Max. Dimension:
(xyz) = 5.420
angstroms**

**Min. Dimension:
(xyz) = 4.724
angstroms**

Iodide (I$^-$):

**Max. Dimension:
(xyz) = 4.400
angstroms**

**Min. Dimension:
(xyz) = 4.400
angstroms**
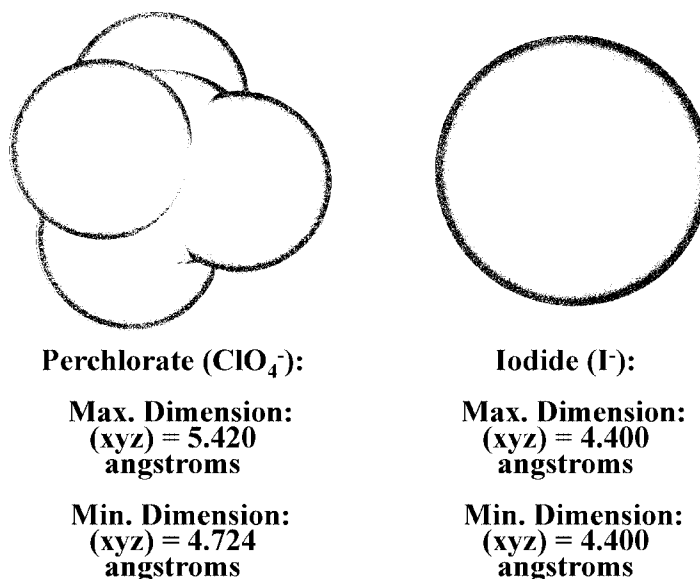
Figure 3-3. Comparison of the molecular dimensions for the perchlorate (left) and iodide (right) anions.

thyroid may be based on an anion-exchange mechanism using a large cation such as a quaternary amine (e.g., arginine) (Wolff, 1989).

Perchlorate also has been used to stimulate the efflux of iodide already stored in the follicular lumen of the gland (Atterwill et al., 1987). The exact nature of the mechanism for this effect has not been established, however. Transport of iodide out of the cell is downhill electrically, but this could be accounted for by the high concentration gradient that is established from follicular lumen (iodide stored in the colloid) to the basolateral and extracellular space. This may be the rate-limiting aspect for perchlorate efflux effect. Perchlorate added to the apical side of a polarized thyroid cell monolayer is substantially less effective than when added to the basolateral side (Wolff, 1998). Moreover, perchlorate rapidly increases the secretory response to TSH, and TSH increases iodide efflux before it increases iodide influx, suggesting that additional control points may exist.

Thus, perchlorate appears to have no effect on the iodination process itself but, rather, displaces iodide by competitive uptake at the NIS. Perchlorate is concentrated by thyroid tissue in a manner similar to iodide, but it is not significantly metabolized in the gland nor peripherally, as mentioned previously. It is not unequivocally established whether there are additional effects of perchlorate on iodide transport within the thyroid. Pharmacokinetic studies with perchlorate, both acute and particularly once steady state has been achieved, have provided some useful data with which to gain insight on this issue. The potential impacts as health endpoints of interest for human health risk assessment of this perturbation in the hypothalamic-pituitary-thyroid axis and hormone economy will be discussed in Section 3.4.

3.4 TOXICODYNAMICS OF THYROID HORMONE PERTURBATIONS

Given the established mode of action for perchlorate as the inhibition of iodide uptake at the NIS, it is important to distinguish the temporal aspects with respect to potential adverse tissue response.

3.4.1 Carcinogenic Effects

In higher organisms, when demands for more thyroid hormone are small, existing thyroid follicular cells can meet the demand. With increased need, as a result of certain chemical

1   exposures or iodine deficiency, the thyroid responds by increasing the size (hypertrophy) and

2   number (hyperplasia) of thyroid follicular cells to enhance hormone output. With continued TSH

3   stimulation, there is actual enlargement of the thyroid (goiter) and, at least in rodents, eventual

4   neoplasia of the thyroid follicular cells. Because TSH-producing pituitary cells also are

5   stimulated, they too sometimes undergo hyperplasia and neoplasia (U.S. Environmental

6   Protection Agency, 1998a; Hill et al., 1998). The EPA Assessment of Thyroid Follicular Cell

7   Tumors (U.S. Environmental Protection Agency, 1998a), as well as reviews recommended

8   therein, provides details about thyroid follicular cell carcinogenesis. Figure 3-4 shows

9   schematically the possible antithyroid effects that could influence carcinogenesis. Note that

10 effects, not only in the thyroid but also in peripheral tissues and the liver, may cause demand on

11 thyroid hormone production such that the TSH stimulation of the thyroid to produce more

12 hormone is enlisted. Table 3-3 lists mechanisms of antithyroid-mediated neoplasia in rodents.

13 The potential for an indirect effect of perchlorate has been established, but genotoxicity

14 information was required to evaluate its potential for direct effects. As will be discussed in

15 Section 3.5, a battery of genetoxicity assays was included in the testing strategy.

16   Long-term perturbations in the hypothalamic-pituitary-thyroid axis by the various

17 influences listed in Table 3-3 are more likely to predispose the laboratory rat to a higher

18 incidence of proliferative lesions (Capen, 1997). One factor that may play a role in this

19 interspecies quantitative difference in sensitivity to thyroid stimulation is the influence of protein

20 carriers of thyroid hormones in the blood (Table 3-2). Both humans and rodents have

21 nonspecific, low-affinity protein carriers of thyroid hormones (e.g., albumin). However, in

22 humans, other primates, and dogs, there is a high-affinity binding protein, thyroxine-binding

23 globulin, which binds T4 (and T3 to a lesser degree). This protein is missing in rodents and

24 lower vertebrates. As previously indicated, T4 is bound to proteins with lower affinity in the

25 rodent and is more susceptible to removal from the blood, by metabolism, and through excretion

26 than in dogs and primates.

27   In keeping with this finding, the serum half-life of T4 is much shorter in rats (less than

28 1 day) than it is in humans (5 to 9 days); this difference in T4 half-life results in a 10-fold greater

29 requirement for exogenous T4 in the rat with a nonfunctioning thyroid than in the adult human.

30 Serum T3 levels also show a species difference: the half-life in the rat is about 6 hr; whereas, it is

31 about 24 hr in humans. High thyroid hormone synthetic activity is demonstrated in thyroid
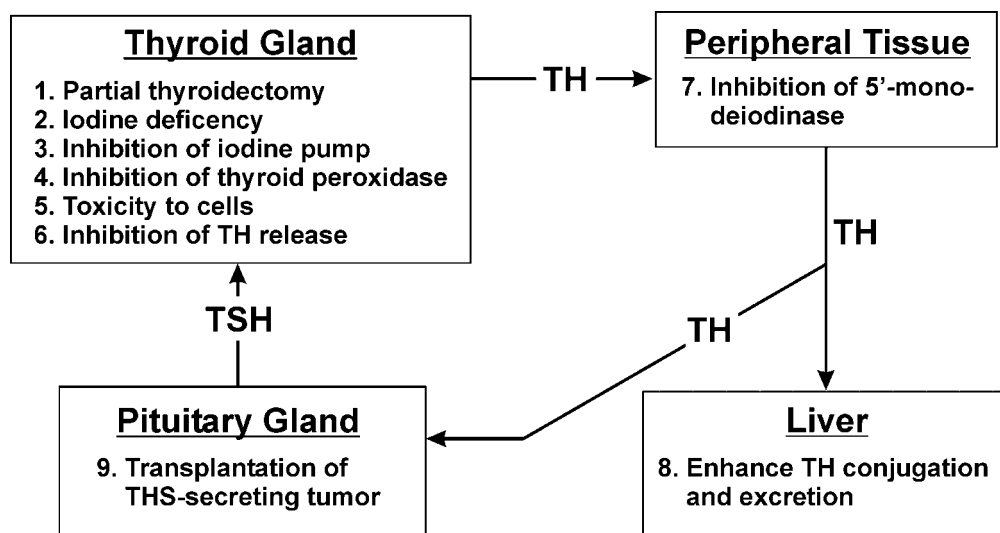
Figure 3-4. Schematic of antithyroid effects that influence thyroid carcinogenesis. (U. S. Environmental Protection Agency, 1998a; and Hill et al., 1998).

TABLE 3-3. MECHANISMS OF ANTITHYROID-MEDIATED NEOPLASIA IN RODENTS (U.S. Environmental Protection Agency, 1998a).

- **DNA Directed**
  - X rays
  - ^{131}I
  - Genotoxic chemicals

- **Indirect**
  - Partial thyroidectomy
  - Transplantation of TSH-secreting pituitary tumors
  - Iodide deficiency
  - Chemicals inhibiting iodide uptake
  - Chemicals inhibiting thyroid peroxidase
  - Chemicals inhibiting TH
  - Chemicals inhibiting conversion of T3 and T4
  - Chemical inhibiting hepatic thyroid hormone metabolism and excretion

1	follicles in rodents, where the follicles are relatively small and are surrounded by cuboidal
2	epithelium. Follicles in primates demonstrate less activity and are large with abundant colloid,
3	and follicular cells are relatively flattened (low cuboidal) (McClain, 1992).
4	The accelerated production of thyroid hormones in the rat is driven by serum TSH levels
5	that are probably about 6- to 60-fold higher than in humans. This assumes a basal TSH level in
6	rats and humans of 200 ng/mL and 5 μU/mL, respectively, and a potency of human TSH of 1.5 to
7	15 IU/mg of hormone (U.S. Environmental Protection Agency, 1998a). Thus, it appears that the
8	rodent thyroid gland is chronically stimulated by TSH levels to compensate for the increased
9	turnover of thyroid hormones. It follows that increases in TSH levels above basal levels in rats
10	could more readily move the gland towards increased growth and potential neoplastic change
11	than in humans. In addition to considerations about the influence of serum thyroid hormone
12	carrier proteins, there are differences between humans and laboratory animals in size and life
13	span and in the pharmacokinetics and pharmacodynamics of endogenous and exogenous
14	chemicals. Any comparison of thyroid carcinogenic responses across species should be
15	cognizant of all these factors.
16	A number of goitrogenic compounds, those that either interfere with thyroid hormone
17	synthesis or secretion, have been demonstrated to result in thyroid follicular cell adenomas in
18	rats. Excessive secretion of TSH alone has been reported to produce a high incidence of thyroid
19	follicular cell adenomas. The pathogenic mechanism of thyroid follicular cell tumor
20	development in rodents involves a sustained excessive stimulation of the thyroid by TSH. In the
21	multistage model of this pathogenesis, the proliferative lesions often begin as hyperplasia, may
22	proceed to the development of benign tumor (adenomas), and infrequently develop into
23	malignant tumors (Figure 3-5).
24	The precise molecular steps in the carcinogenic process leading to thyroid follicular cell
25	cancer have not been elucidated totally although significant insights into the problem have been
26	described (Farid et al., 1994; Said et al., 1994). Normal cell division in the thyroid seems to be
27	affected by an interplay among several mitogenic factors, namely TSH, insulin-like growth
28	factor-1 (IGF-1), insulin, epidermal growth factor (EGF), and possibly fibroblast growth factor
29	(FGF). Additionally, other factors, such as transforming growth factor β, certain interferons, and
30	interleukin 1, may inhibit growth.
31	

Morphologic Continuum ━▶

Normal

Hyperplasia

Adenoma

Carcinoma

Significance in Risk Assessment
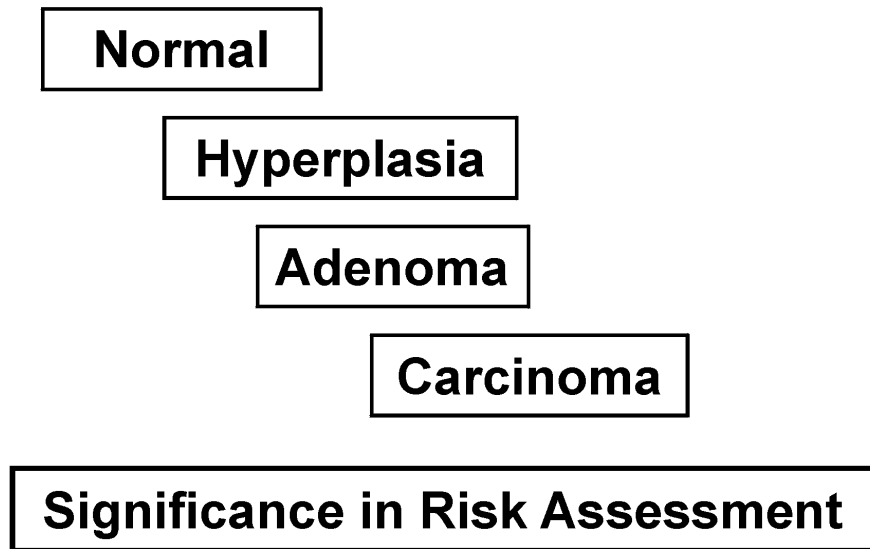
Figure 3-5. **Proliferative changes involved in the multistage characterization of thyroid follicular cell neoplasia in rodents represent a morphologic continuum. Although these lesions typically are classified as discrete entities, the overlap in morphologic features should be emphasized because only imprecise criteria to separate borderline proliferative lesions exist. Thyroid neoplasia in rodents is considered relevant to human risk assessment (U.S. Environmental Protection Agency, 1998a) but thought to be protective (Capen, 1997).**

1   Figure 3-6 shows the possible molecular events in human thyroid follicular carcinogenesis.

2   In spite of the potential qualitative similarities, there is evidence that humans may not be as

3   sensitive quantitatively to thyroid cancer development from thyroid-pituitary disruption as are

4   rodents. Rodents readily respond to reduced iodide intake with the development of cancer;

5   whereas, humans develop profound hyperplasia with "adenomatous" changes with only

6   suggestive evidence of malignancy. Even with congenital goiters from inherited blocks in

7   thyroid hormone production, only a few malignancies have been found in humans. Thus, despite

8   a common physiology in regard to the thyroid-pituitary feedback system, the role of disruption of

9   this axis in human cancer development is much less convincing. EPA has adopted the following

10   science

```
┌─────────────────────────┐
│  Thyroid follicular cells │
└─────────────────────────┘
            │  TSH, insulin, IGF-1, EGF, FGF
            ▼
┌─────────────────────────┐
│    Nodular hyperplasia    │
└─────────────────────────┘
            │  ras, gsp Chromosome aberrations
            │  of 5, 7, and 12
            ▼
┌─────────────────────────┐
│    Follicular adenoma     │
└─────────────────────────┘
            │  Loss of heterozygosity at 3p
            ▼
┌─────────────────────────┐
│    Follicular carcinoma   │
└─────────────────────────┘
            │  p53
            ▼
┌─────────────────────────┐
│    Anaplastic carcinoma   │
└─────────────────────────┘
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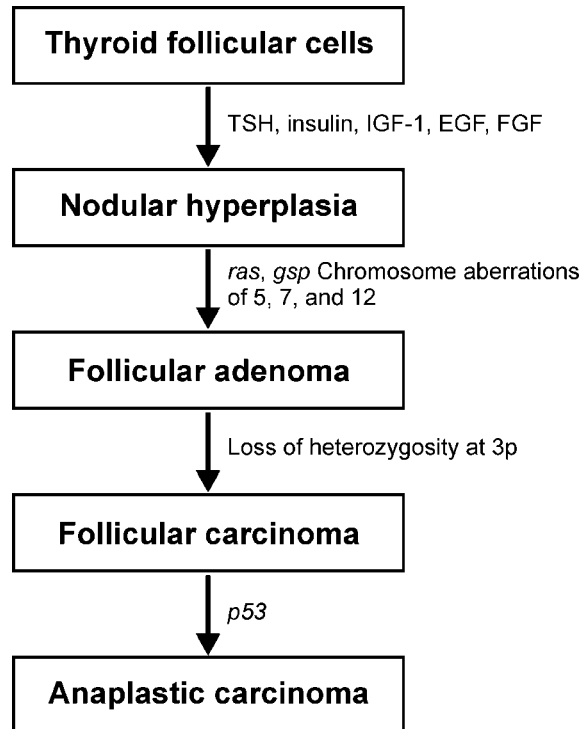
Figure 3-6. Possible molecular events in human thyroid follicular carcinogenesis (*ras* = *ras* protooncogene, *gsp* = GTP-binding protein mutation, *p*53 = *p*53 tumor suppressor gene) (U.S. Environmental Protection Agency, 1998a and Hill et al., 1998).

1   policy that recognizes the role of mode-of-action information regarding thyroid-pituitary

2   disruption and mutagenesis to potential thyroid carcinogenesis (U.S. Environmental Protection

3   Agency, 1998a).

4   • It is presumed that chemicals that produce rodent thyroid tumors may pose a carcinogenic

5   hazard for the human thyroid.

6   • In the absence of chemical-specific data, humans and rodents are presumed to be equally

7   sensitive to thyroid cancer caused by thyroid-pituitary disruption. This is a conservative

8   position when thyroid-pituitary disruption is the sole mode of action because rodents appear to

9   be more sensitive to this carcinogenic mode-of-action than are humans. When the thyroid

10   carcinogen is a mutagenic chemical, the possibility that children may be more sensitive than

11   adults needs to be evaluated on a case-by-case basis.

1   • Adverse rodent noncancer thyroid effects (e.g., thyroid enlargements) following short- and

2   long-term reductions in thyroid hormone levels are presumed to pose human noncancer health

3   hazards.

4   The new data on the antithyroid activity of perchlorate that has resulted from the testing

5   strategy will be evaluated in Chapter 7 according to criteria provided in the guidance (U.S.

6   Environmental Protection Agency, 1998a) to determine the likelihood that the chemical would

7   act indirectly, via disruption of the thyroid-pituitary axis, or directly on DNA.

8

3.4.2 Neurodevelopmental Deficits and Other Potential Adverse Effects Resulting from Thyroid Hormone Disruption

11   As expressed by the external review panel convened by Toxicology Excellence for Risk

12   Assessment (TERA) in 1997, there was concern about other potential adverse effects of

13   perchlorate-induced hypothyroidism. Humans respond as do experimental animals in regard to

14   short- and mid-term disturbances in thyroid functioning from various anti-thyroid stimuli such as

15   iodide deficiency, partial thyroidectomy (surgically or $^{131}I^-$ induced), and goitrogenic chemicals

16   such as thionamides (U.S. Environmental Protection Agency, 1998a). For instance, thyroid

17   hormone is critical to normal brain and physical development. This dependency begins in the

18   uterus and extends to 3 years of age in humans. Thus, there was concern that hypothyroidism

19   during pregnancy could result in neurodevelopmental effects.

20   The role of the placenta in thyroid hormone metabolism is shown in Figure 3-7. Although

21   the fetus is initially dependent on maternal thyroid hormone levels, the potential for disruption of

22   fetal hormone production remains once the fetal thyroid assumes this function because

23   perchlorate can cross the placenta. Disruption of circulating thyroid hormones can have

24   drastically different effects on fetuses and infants than on adults, depending on the developmental

25   stage at exposure (Table 3-4). It is important to emphasize that even transient disruption may

26   lead to permanent effects in the developing organism.

27   Chemical-induced alterations in thyroid hormone homeostasis are known to adversely

28   affect the development of many organ systems, including the nervous and reproductive systems

29   (Porterfield, 1994; Jannini et al., 1995). Severe developmental hypothyroidism caused by iodine

30   deficiencies or a congenital condition has devastating effects on fetal and postnatal development,

31   including mental deficiencies and hearing, speech, and motor deficits (Porterfield, 1994; Sher
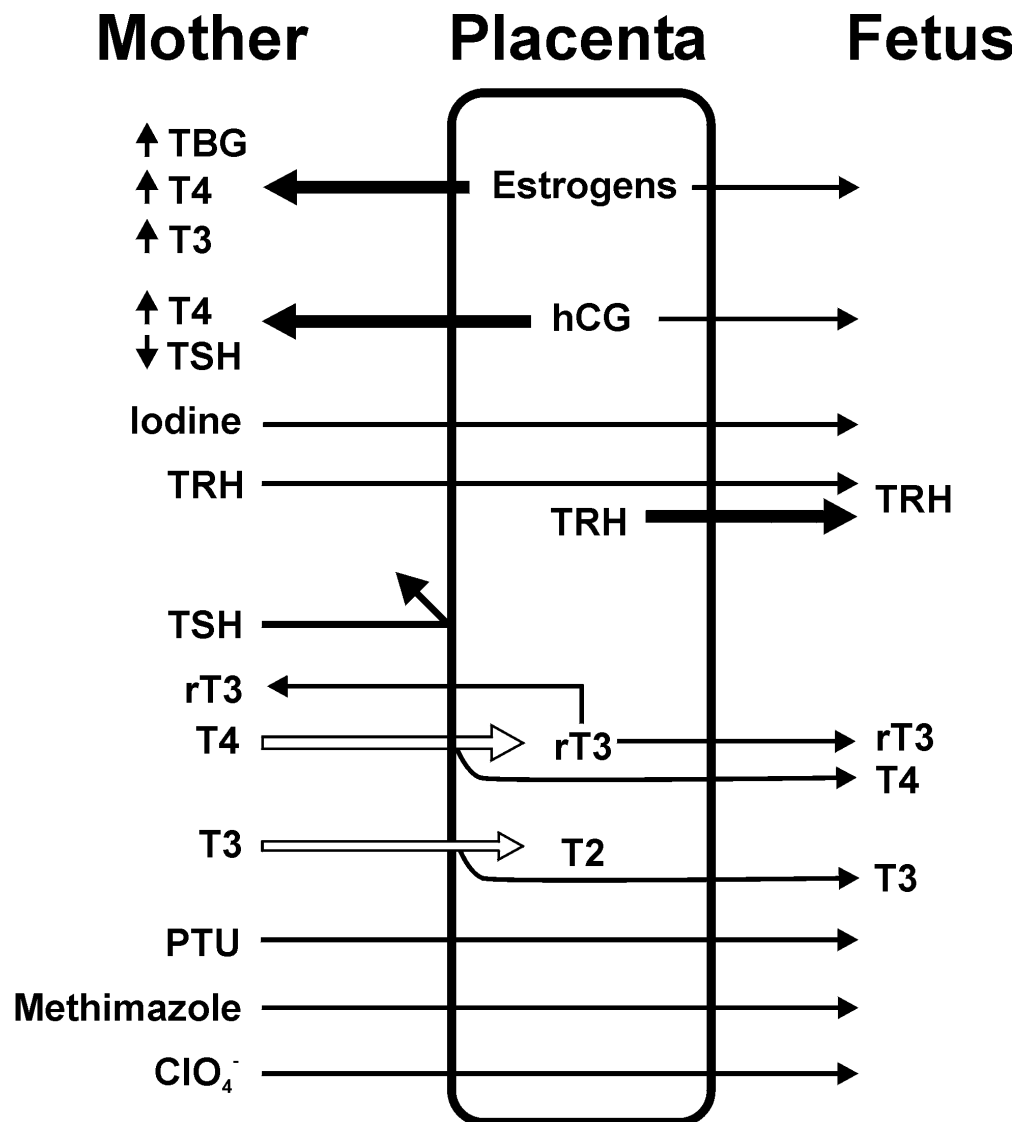
Figure 3-7. Schematic representation of the role of the placenta in thyroid hormone metabolism during human pregnancy. The placenta produces estrogens and hCG that increase maternal TBG levels and stimulate maternal thyroid hormone production, respectively. Both activities tend to increase maternal T4 and T3 concentrations and to inhibit maternal TSH secretion. Iodide and TRH readily cross the placenta, and the placenta itself synthesizes TRH. The placenta is impermeable to TSH and only partially permeable to T4 and T3. Placental Type III iodothyronine monodeiodinase enzymes degrade T4 to rT3 and T3 to 3,3'-diiodothyronine (T2). Propylthiouracil and methimazole readily cross the placenta. Given its physicochemical characteristics and similarity to iodide, perchlorate also is anticipated to cross readily. (Modified from Fisher, 1996 and Underwood, 1998).

TABLE 3-4. MAIN SYMPTOMS AND EFFECTS OF HYPOTHYROIDISM

Developmental *(Transient disruption leads to permanent effects.)*	Adult *(Transient disruption leads to transient effects.)*
• Delayed reflex ontogeny • Impaired fine motor skills • Deaf-mutism, spasticity • Gait disturbances • Mental retardation • Speech impairments	• Run down, slow, depressed • Sluggish, cold, tired • Dryness and brittleness of hair • Dry and itchy skin, constipation • Muscle cramps • Increased menstrual flow • Thyroid tumors in rodents

1 et al., 1998). It is important to emphasize that these effects are caused by a lack of thyroid

2 hormones alone, rather than by tumor development or thyroid hypertrophy/hyperplasia due to

3 increases in TSH. Thus, the important species comparison may be perchlorate's action of iodide

4 uptake inhibition at the NIS. In fact, data discussed in Chapters 5 and 6 show that the sensitivity

5 of the NIS is quite similar across species.

6   During development, thyroid hormones regulate cell proliferation, migration, and

7 differentiation. Intracellularly, THs bind to thyroid hormone receptors that interact with thyroid

8 response elements to alter expression of messenger ribonucleic acids (mRNAs) and subsequent

9 protein synthesis. The pituitary-thyroid TSH feedback loop may or may not be activated during

10 development, depending on the mechanism of action of the chemical. The adversity of

11 congenital hypothyroidism, usually less severe than endemic cretinism, can be ameliorated via

12 early postnatal thyroxine therapy. In contrast, the effects of developmental iodine deficiency can

13 not be corrected with only postnatal therapy, indicating that iodine deficiency during pregnancy

14 is the causative action (Cao et al., 1994). Clearly, xenobiotics that contribute to fetal or maternal

15 hypothyroidism or hypothyroxenemia are of concern.

16   Since the previous external peer review, studies reported in the clinical and epidemiological

17 literature have reinforced concerns for deficits in neuropsychological development related to

18 maternal thyroid deficiency. Haddow et al. (1999) showed an effect on IQ scores in children

19 (ages seven to nine) who had normal thyroid function at birth but were born to women with

20 abnormal thyrotropin levels versus children born to a matched cohort of women with normal

21 thyrotropin levels as controls. Haddow et al. (1999) concluded that even mild and probably

1   asymptomatic hypothyroidism in pregnant women can adversely affect their children's

2   subsequent performance on neuropsychological tests.

3   Pop et al. (1995) noted an average impairment of 10.5 IQ points in the offspring of mothers

4   with high thyroid peroxidase antibody (TPO-Ab) titers during pregnancy. In a later prospective

5   study these same researchers evaluated developmental indices at 3 weeks, 10 months, 1 and

6   2 years of age and demonstrated that a maternal free T4 blood level that was less than the 10^{th}

7   percentile of first trimester values (10.4 pmol/L in their study series) was associated with

8   distinctly impaired psychomotor development whether or not TSH and TPO-Abs were elevated

9   (Pop, et al., 1999). Smit et al. (2000) reported a similar relationship between free T4 and early

10 neurodevelopment of children born from treated hypothyroid women.

11   Morreale de Escobar et al. (2000) evaluated epidemiological, clinical, and basic research

12 data to ascertain if the principal factor leading to neurodevelopmental deficits in children was

13 related to maternal hypothyroidism, whether clinical or subclinical (as defined by TSH higher

14 than the 98^{th} percentile of the normal population); or if they were instead related to maternal

15 hypothyroxinemia per se (decrement in T4 without concomitant increase in TSH). These

16 researchers concluded that conditions resulting in hypothyroxinemia alone (a low for gestational

17 age circulating maternal free T4 level whether or not TSH was increased) poses an increased risk

18 for poor neuropsychological development of the fetus. T4 is the required substrate for the

19 ontogenically-regulated generation of T3 in the amounts needed for optimal brain development,

20 both temporally and spatially. Normal maternal T3 concentrations did not seem to prevent the

21 potential damage of a low T4 supply (Morreale de Escobar et al., 2000). Hypothyroxinemia

22 seems to be much more frequent in pregnant women than either clinical or subclinical

23 hypothyroidism and autoimmune thyroid disease (AITD), especially in regions where the iodine

24 intake of the pregnant woman is inadequate to meet her increased needs for T4 (Morreale de

25 Escobar et al., 2000).

26   Figure 3-8 illustrates the windows of susceptibility for insults to the brain resulting from

27 hypothyroxinemia. A similar map has been developed for rats, and time lines have begun to be

28 compared and correlated (Rice and Barone, 2000), as shown in Figure 3-9. Morreale de Escobar

29 et al. (2000) reported findings that altered early migration of cortical cells can be observed in rats

30 with severe iodine deficiency. Porterfield (2000) has also discussed the potential for
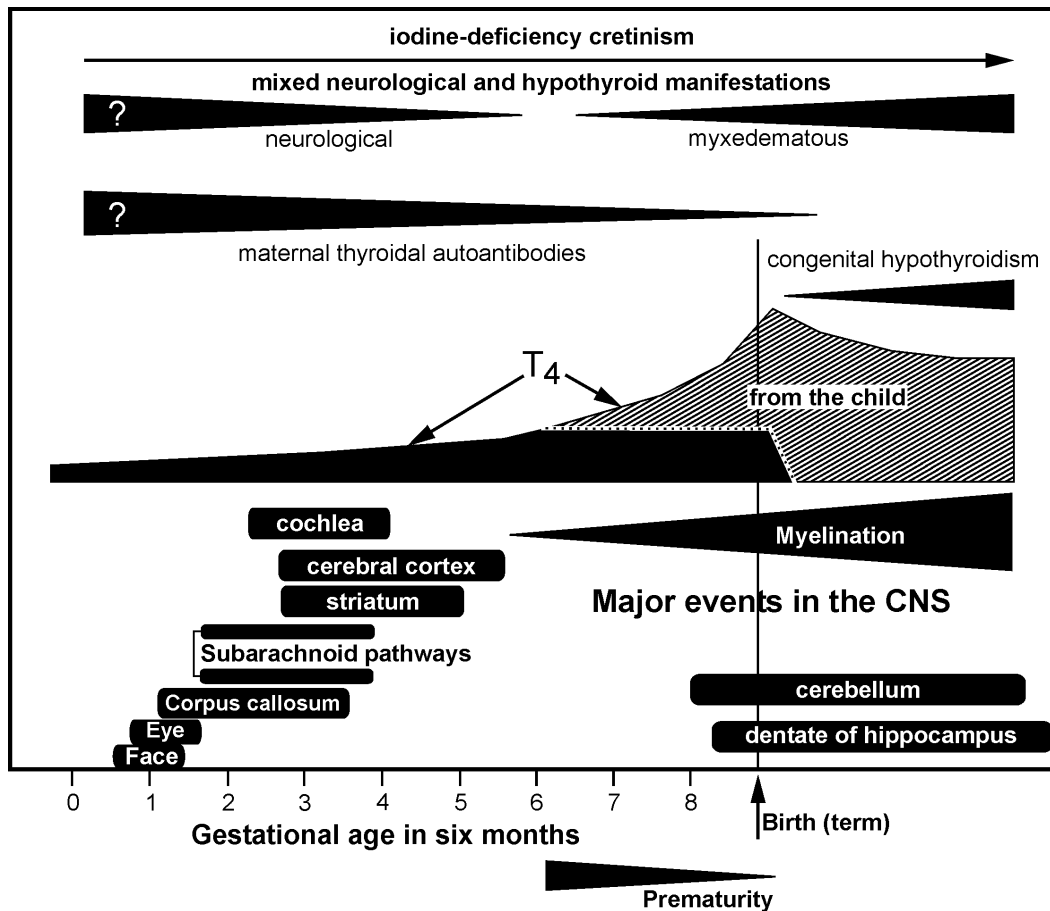
Figure 3-8. Approximate timing of major insults to the brain resulting from hypothyroxinemia, superimposed on major neurodevelopmental events in humans. Conditions resulting in early maternal hypothyroxinemia, combined to later impairment of the fetal thyroid, are the most damaging, with central nervous system (CNS) damage that is irreversible at birth. The most frequent cause is maternal iodine deficiency (ID) and the presence of maternal autoimmune thyroid disease (AITD). Unless ID is also present, the CNS damage in congenital hypothyroidism is preventable by early postnatal treatment because the normal maternal thyroxinemia has avoided damage to the brain until birth. If maternal hypothyroxinemia persists, normal maternal concentrations of T_3 do not protect the fetal brain because of its dependence on intracerebral regulation of local T_3 availability by deiodinating pathways using T_4 as a substrate. Interruption of the contribution of maternal T_4 in premature infants with an immature thyroid may also underlie their increased risk of neurodevelopmental problems, the more severe the earlier their birth. The *question mark* indicates that it is unknown whether very early CNS development, corresponding to a period when the general morphogenesis of the pros encephalon (neurolation and segmentation) is being determined, is thyroid hormone sensitive or not (Morreale de Escobar et al., 2000).

(A)

Birth

Embryonic | Fetal | Postnatal

Ovulation
Fertilization
Implantation
Functional organization
Histogenesis
Organogenesis
GD 5-6   GD 15   Adolescence
GD 8-9   GD 11   GD 21-22   PND 15   PND 35-45
Neurulation
Proliferation and migration
Differentiation and synaptogenesis
Apoptosis
Gliogenesis
Myelination

(B)

(B)	Cell proliferation	Migration of neurons	Subplate neurons	Synapse formation	Myelination							

Prenatal period (months): 0 1 2 3 4 5 6 7 8 9

Cell proliferation: Radial glia and neurons; Glia

Migration of neurons: Brain and spinal cord; External granular

Synapse formation: Marginal zone; Subplate; Hippocampus; Reticular formation; Visual cortex; Association cortex

Myelination: Somatosensory; Roots of Spinal nerves; Vestibular; Cerebellum + extrapyramidal; Reticular formation; Visual; Auditory; Pyramidal system; Association + commissure

Postnatal period (years): 1 2 3 4 5 10

Migration of neurons: Layer cerebellum

Figure 3-9. Timelines of developmental processes in the nervous system of rats (a) and humans (b). Rat timeline is compared to timing of fertilization, organogenesis, and histogenesis. Human perinatal period is scaled in months and the postnatal development is scaled in years (Rice and Barone, 2000).

1 neurotoxicity and altered brain development that may result from exposure to environmental

2 chemicals that disrupt thyroid function even on a transient basis.

3 These concerns for the potential adverse effects of perchlorate on T4 and T3, especially

4 during pregnancy, are compounded by the growing appreciation that women of childbearing age

5 have relatively low iodide intake. A January 2001 report by the National Academy of Sciences

6 (NAS) concerning the dietary reference intake of trace-mineral nutrients, including iodine,

7 indicated that less than 25% of the total population was below the estimated average requirement

8 for iodide and stressed a need to look at levels of adequacy for susceptible age groups and status

9 during pregnancy and lactation. The higher requirements during this time indicate a potential

10 susceptibility as shown in Table 3-5. The NAS also cautions against using urinary iodine as a

11 biomarker for iodine status unless the data are from 24-hour collections or are normalized against

12 creatinine. Other reports suggest that the level of iodide intake is less than a third of the range

13 recommended for pregnant women by the World Health Organization (WHO) (Caron et al.,

14 1997).

15

16

TABLE 3-5. DIETARY REFERENCE INTAKES (DRI) FOR IODIDE
(National Academy of Sciences, 2001)

Age or Status	Adequate Intake (AI) μg/day	Estimated Average Requirement (EAR) μg/day	Recommended Dietary Allowance (RDA) μg/day
0-6 months	110		
6-12 months	130		
1-3 years		65	90
4-8 years		65	90
9-13 years		73	120
14-18 years		95	150
19-15 years		95	150
51 + years		95	150
Pregnancy		160	220
Lactation		209	290

1   The prevalence of abnormal thyroid function continues to be debated and this is

2   confounded by the variable definitions of the disease state as well as the different measures of

3   thyroid function (Canaris et al., 2000). Most reports are still defined by TSH levels rather than

4   for hypothyroxinemia per se, but recent presentations suggest that TSH is a poor test to assess the

5   severity of tissue hypothyroidism (Meier et al., 2001), and recommendations in the epidemiologic

6   literature are proposing that screening of pregnant women should include the determination of

7   free T4 (Morreale de Escobar et al., 2000). Age, sex and dietary iodine levels are confounding

8   factors, although virtually all studies report higher prevalence rates for hypothyroidism

9   (as defined by increased TSH) in women with age (Canaris et al., 2000). Rates as high as 24%

10   among women older than 60 years have been reported. Suppressed TSH levels have been

11   associated with decreased bone density, increased risk of atrial fibrillation, premature atrial beats,

12   and effects on serum lipids notably elevated serum cholesterol levels.

13   Together these findings strongly suggest that a susceptible population of particular concern

14   for perchlorate exposure is pregnant women with hypothyroxinemia and that the iodine

15   deficiency represents an additional potential insult that could exacerbate the effects of perchlorate

16   toxicity. The elderly, especially women, represent another potentially susceptible population, as

17   well as people with cardiac dysfunction or risk factors such as elevated serum cholesterol.

18   As mentioned above, reproductive toxicity was also a concern as a potential effect of

19   perchlorate's mode of action. In females, thyroid hormones appear to have a role in stimulating

20   the onset of human chorionic gonadotropin (hCG) production by the placenta early in pregnancy.

21   Human chorionic gonadotropin is essential for the maintenance of pregnancy. Therefore, a

22   hypothyroid condition has potential to interfere with normal placental function and fetal

23   survival, as well as the potential to interfere with lactation. Suppression of thyroid hormone

24   secretion with radioactive iodine or goitrogens reduces milk yield in lactating animals. This

25   effect may be caused by suppression of placental lactogen production. Thyroid-releasing

26   hormone is known to play a role in prolactin release during the estrous cycle. Additionally, the

27   thyroid is necessary for the transition to the anestrus state in seasonally breeding species.

28   In summary, effects on thyroid hormone levels have roles in estrous cycle regulation, pregnancy

29   maintenance, fetal growth, and lactation.

30   In males, the primary effects of hypothyroidism appear to occur during testicular

31   development. The testis is responsive to thyroid hormones only during a limited time during the

1   perinatal and prepubertal periods. Thyroid hormone is a major regulator of seminiferous

2   epithelium development by inducing the normal differentiation of Sertoli cells, gonocytes, and

3   Leydig cells, and by limiting the proliferation of those cell types. In the hypothyroid condition,

4   those cells proliferate beyond the norm, and the steroidogenic function of the Leydig cells, on a

5   per-cell basis (but not necessarily in total), is impaired. Secretory activity of the Sertoli cells also

6   appears to be impaired. In boys, untreated hypothyroidism is associated with marked and

7   precocious testis enlargement, but low androgen activity. In a small study, hypothyroid men had

8   complaints of reduced libido that was probably related to a defective leutenizing hormone

9   response to gonadotropin-releasing hormone.

10   The inclusion of an immunological evaluation of mice exposed to perchlorate was

11   warranted because of evidence from earlier clinical studies that indicated a link between the

12   treatment of Graves' disease with perchlorates and serious hematological effects that may be

13   linked to immune mechanisms. A small number of patients undergoing perchlorate therapy have

14   been reported to develop aplastic anemia, agranulocytosis, lymphadenopathy, leukopenia, or skin

15   rashes. The antithyroid drugs propylthiouracil and methimazoles are reported to exert their

16   effects on the hematopoietic system through immune mechanisms. Because the use of these

17   antithyroid drugs by a small number of patients also resulted in sequelae similar to that of some

18   patients under perchlorate treatment, it has been postulated that perchlorate also may act via the

19   immune system.

20

21

22   **3.5   DEVELOPMENT OF A TOXICITY TESTING STRATEGY BASED**
23   **ON MODE OF ACTION**

24   Because the RfD is intended to be a lifetime dose-response estimate, the typical objective

25   of a database to support such a quantitative assessment is to evaluate a comprehensive array of

26   testing endpoints that represent various life stages during which potential effects could occur

27   (e.g., the developing fetus through adult) and for effects on reproductive capability (shown

28   schematically in Figure 3-10). As discussed in the previous sections, thyroid hormone

29   deficiencies, such as those induced by perchlorate, can affect normal metabolism, growth, and

30   development. No robust data existed prior to this time to evaluate other potential target tissues or

31   effects. There were limited data on effects caused by long-term exposures and no data with
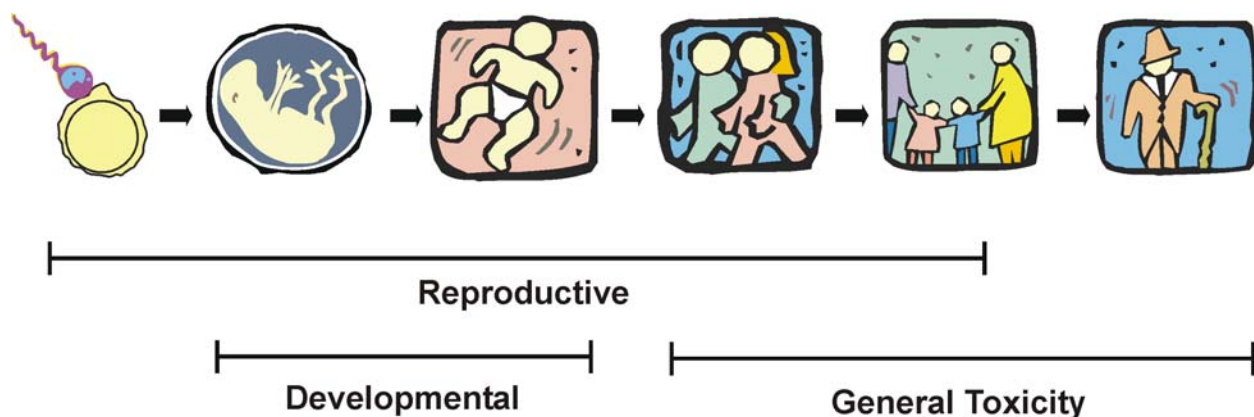
Figure 3-10. Schematic illustrating that a high confidence RfD is based on data that address all potentially critical stages over a lifetime.

1   which to evaluate the effects of perchlorate in potentially susceptible populations such as in

2   developing fetuses, nor were there data on the effects of perchlorate on the reproductive capacity

3   of male or female laboratory animals. Table 3-6 shows the minimum database for derivation of

4   an RfD with low confidence (a 90-day bioassay) and the rationale for other tests typically

5   included to bolster the confidence in the derivation–the same suite of tests that has been

6   discussed for perchlorate. These data typically also reduce the uncertainty for which uncertainty

7   factors are applied (see Table 3-7), either because the absence of data on a suspected endpoint

8   (e.g., developmental toxicity) has been addressed or because mechanistic data provide insight on

9   the relevance of the laboratory animal model, including the magnitude of interspecies and

10   intrahuman variability in toxicokinetics and toxicodynamics. Any individual chemical database

11   may fall in between this range of high and low certainty, depending on the quality of the

12   individual studies and whether the dose response for suspected endpoints is characterized well.

13   The objective of the testing strategy was to provide a comprehensive database that

14   described the mode-of-action-based pathogenesis in quantitative terms so that the resultant

15   estimate could be more predictive and ultimately support the development of a robust RfD

16   estimate that reduced the uncertainties inherent in the provisional, presumably protective values

17   (see Figure 3-11).

18

TABLE 3-6. MINIMUM DATABASE FOR DERIVATION OF AN ORAL REFERENCE DOSE

Mammalian Database[a]	Confidence	Comments
Two chronic oral bioassays in different species One two-generation reproductive study Two developmental toxicity studies in different species	High[b]	Minimum database for high confidence
One subchronic oral bioassay	Low	Minimum database for estimation of an RfD

[a]Rationale is to use different species to evaluate variability in species sensitivity unless a particular laboratory animal model is more appropriate.
[b]Rationale is to address all potentially critical life stages.

TABLE 3-7. FACTORS FOR UNCERTAINTIES IN APPLIED EXTRAPOLATIONS USED TO DERIVE REFERENCE DOSES[a]

10_H	–	Human to sensitive human
10_A	–	Experimental animal to human
10_S	–	Subchronic to chronic duration
10_L	–	LOAEL(HEE)[a] to NOAEL(HEE)[a]
10_D	–	Incomplete to complete database
MF	–	Modifying factor. Professional assessment of scientific uncertainties of the study and database not explicitly addressed above. Default for the MF is 1.0 (e.g., applied for small sample size or poor exposure characterization).

[a]HEE = human equivalent exposure.

1   As illustrated in Figure 3-11, it is ultimately desirable to have a comprehensive

2   biologically-based dose-response model that incorporates the mechanistic determinants of

3   chemical disposition, toxicant-target interactions, and tissue responses integrated into an overall

4   quantitative model of the pathogenesis (Jarabek, 1995a). Because the internal tissue dose of the

5   chemical or its toxic moiety in a target tissue is not always proportional to the applied dose of a

6   compound, emphasis has been placed on the need to distinguish clearly between the exposure

7   concentration and the dose to critical target tissues. Consequently, the term "exposure-dose-

8   response" has been recommended as more accurate and comprehensive (Andersen et al., 1992).

9   This expression refers, not only to the determination of the quantitative relationship between

10   exposure concentrations and target tissue dose, but also to the relationship between tissue dose

11   and the observed or expected responses in laboratory animals and humans. The process of
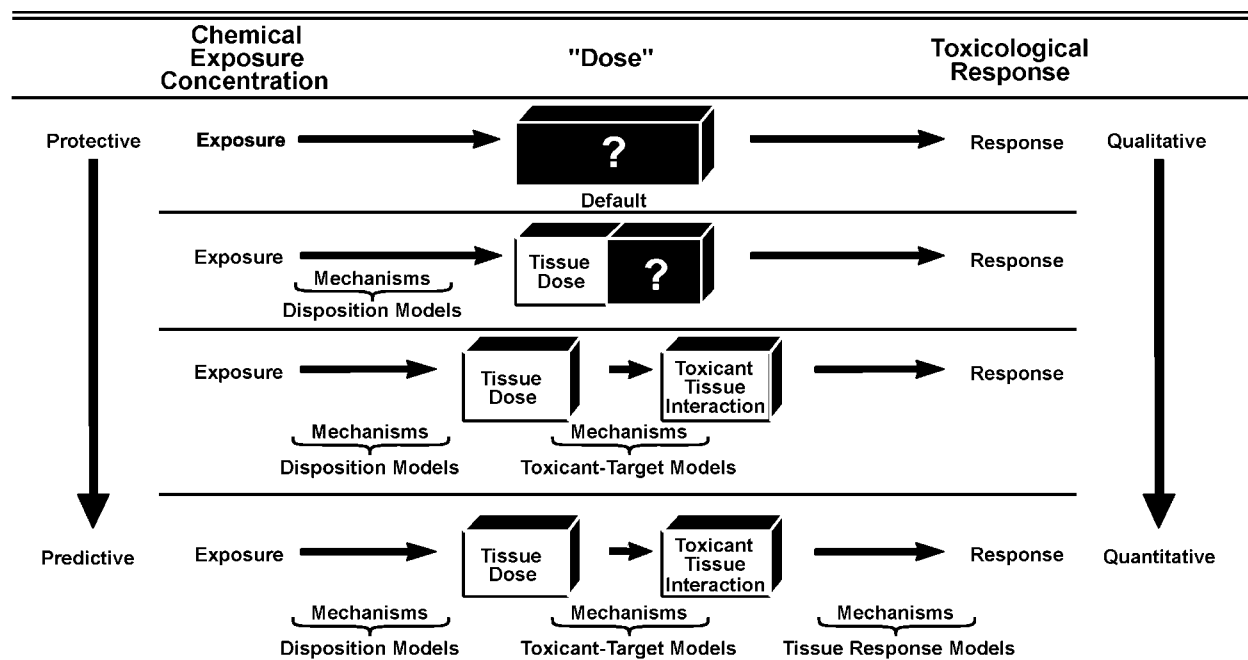
Figure 3-11. Schematic characterization of comprehensive exposure-dose-response continuum and the evolution of protective to predictive dose-response estimates (U.S. Environmental Protection Agency, 1994 and Jarabek 1995b).

1   determining the exposure-dose-response continuum is achieved by linking the mechanisms or

2   critical biological factors that regulate the occurrence of a particular process and the nature of the

3   interrelationships among these factors. This can be especially important for interspecies

4   extrapolation and to understanding intrahuman variability.

5   Dose-response estimates based on characterization of the exposure-dose-response

6   continuum at the rudimentary ("black box") level necessarily incorporate large uncertainty

7   factors to ensure that the estimates are protective in the presence of substantial data gaps. With

8   each progressive level, incorporation and integration of mechanistic determinants allow

9   elucidation of the exposure-dose-response continuum and, depending on the knowledge of model

10   parameters and fidelity to the biological system, a more accurate characterization of the

11   pathogenesis process (Jarabek, 1995a). Because of the increase in accuracy of the

12   characterization with each progressive level, dose-response estimates also progress from more

13   protective to factually-based (predictive).

1   Eight new studies were recommended as part of the original testing strategy after the May

2   1997 external peer review to provide such a comprehensive array of endpoints. These studies are

3   described below along with the role they were anticipated to play in informing the revised health

4   risk assessment (see Table 3-8).

5

6

TABLE 3-8. PERCHLORATE PEER REVIEW RECOMMENDED STUDIES SUMMARY

Study	Description	Potential Use in Assessment
90-Day subchronic bioassay + TH[a] + reproductivity + genotoxicity + recovery	Tests for other target tissues; evaluates effect on TH in young adult rats; reproductive parameters added; mouse micronuclei and a recovery group	Minimum database for RfD dose-response for TH in young adult rats; additional information on others; may allow decrease in uncertainty factor (UF) for database deficiencies
Developmental neurotoxicity + TH	Evaluates nervous system in fetal and postnatal rats; TH in does (P0-generation) and pups (F1-generation)	Potentially critical effect; comparison of developmental versus adult effects on TH
Developmental study + TH	Evaluates birth defects in rabbits; TH in does at end of gestation	Potentially critical effect; data in second species for TH effects; may reduce UF for database deficiencies
Two-Generation reproductive toxicity + TH	Evaluates fertility of adult rats and toxicity in offspring over two generations; TH in parents (F0-generation) and offspring (F1- and F2-generations)	Potentially critical effect; may reduce UF for database deficiencies
ADME studies	Characterize absorption, distribution, metabolism, and elimination in rats and humans; iodine inhibition and perchlorate kinetics and hormone homeostasis	Interspecies extrapolation
Mechanistic studies	Evaluate mechanism of TH response and sensitivity in rats and humans	Interspecies extrapolation; determine susceptible subpopulation
Genotoxicity assays	Test for toxicity to DNA	Mode-of-action information for thyroid neoplasia; may reduce UF for database deficiencies
Immunotoxicity	Evaluates immune system structure and function	Potentially critical effect; may reduce UF for database deficiencies

[a]Thyroid hormones (T4 and T3); Thyroid stimulating hormone (TSH), a pituitary hormone, was also assayed in those studies.

1   **(1) 90-Day Subchronic Oral Bioassay Study.** This study was considered the minimum data

2   requirement for derivation of an oral RfD. The study aimed to identify other target tissues,

3   to test young adult rats, and to provide data on the effect of repeated exposure to perchlorate

4   on thyroid hormone levels. The 30-day recovery phase, i.e., evaluation of the thyroid status

5   30 days after perchlorate was stopped, would provide data necessary to characterize its

6   anti-thyroid effects with respect to carcinogenicity (U.S. Environmental Protection Agency,

7   1998a). These data were collected to allow reduction of the uncertainty factor applied for

8   database deficiencies.

9

10   **(2) Developmental Neurotoxicity Study.** This study was designed to evaluate the potential for

11   developmental neurotoxicity of perchlorate by assessing functional and morphological

12   endpoints in offspring from the mother exposed during pregnancy and lactation.

13   Neurotoxicity endpoints were likely to be a critical effect, and the developing organism a

14   sensitive subpopulation. It was hoped that these data would allow reduction of the

15   uncertainty factors applied for intrahuman variability and database deficiencies.

16

17   **(3) Segment II Developmental Study.** This study was conducted to evaluate the potential for

18   perchlorate to cause birth defects in rabbits and to evaluate a potentially critical effect and

19   subpopulation. This study also was conducted to provide data on the thyroid hormone

20   effects in a second species (in addition to rats). These data might allow reduction of the

21   uncertainty factor applied for database deficiencies.

22

23   **(4) Two-Generation Reproductive Toxicity Study.** This study was designed to evaluate the

24   potential for perchlorate to cause deficits in reproductive performance in adult rats and for

25   toxicity in the young offspring. The primary goal of this study was to identify a potentially

26   critical effect and to allow for reduction of the uncertainty factor applied for database

27   deficiencies.

28

29   **(5) Absorption, Distribution, Metabolism, and Elimination Studies.** These ADME studies

30   aimed to understand the pharmacokinetics (i.e., how perchlorate is absorbed, distributed,

31   metabolized, and excreted) of perchlorate in test animals and humans. These data were to

1   provide information to support construction of quantitative extrapolation of dose across

2   species (e.g., rat to human).

3

4   **(6) Perchlorate Mechanism Studies.** These studies provided a link to the pharmacokinetic

5   studies and were conducted via a comparison of existing literature and of new *in vitro* and

6   *in vivo* data that evaluated the effects of perchlorate on the iodide uptake mechanism across

7   species to aid in the quantitative extrapolation of dose.

8

9   **(7) Genotoxicity Assays.** These studies evaluated the potential for carcinogenicity by

10   evaluating mutations and toxic effects on DNA. These data were useful to determining

11   whether the benign thyroid tumors were likely to be a result of the proposed threshold

12   pathogenesis process.

13

14   **(8) Immunotoxicity Studies.** These studies were planned to evaluate the potential for

15   perchlorate to disrupt immune function and identify a potentially critical effect. These data

16   would help to reduce the uncertainty factor applied for database deficiencies. Because

17   concern was raised for these potential adverse effects based on the previous clinical

18   experience with treatment of Graves' disease patients, these studies were considered

19   necessary to a comprehensive database for perchlorate.

20

21   In the 1998 external review draft (U.S. Environmental Protection Agency, 1998d), a model

22   based on mapping the events of the mode of action for perchlorate was proposed as shown in

23   Figure 3-12. The key event was identified as the inhibition of iodide uptake at the NIS, followed

24   by decreases in thyroid hormones and increases in TSH. Both the potential neurodevelopmental

25   and neoplastic sequelae of this perturbation in thyroid hormone economy were proposed as

26   downstream adverse health outcomes. The conceptual model was endorsed by the external peer

27   review panel in 1999 (Research Triangle Institute, 1999), and additional studies were

28   recommended to reevaluate indications of developmental and neurodevelopmental in rats for

29   effects observed in the 1998 database. Delineating the continuum of histopathological changes

30   in the thyroid was also recommended. The results of all the studies in the testing strategy (both

31
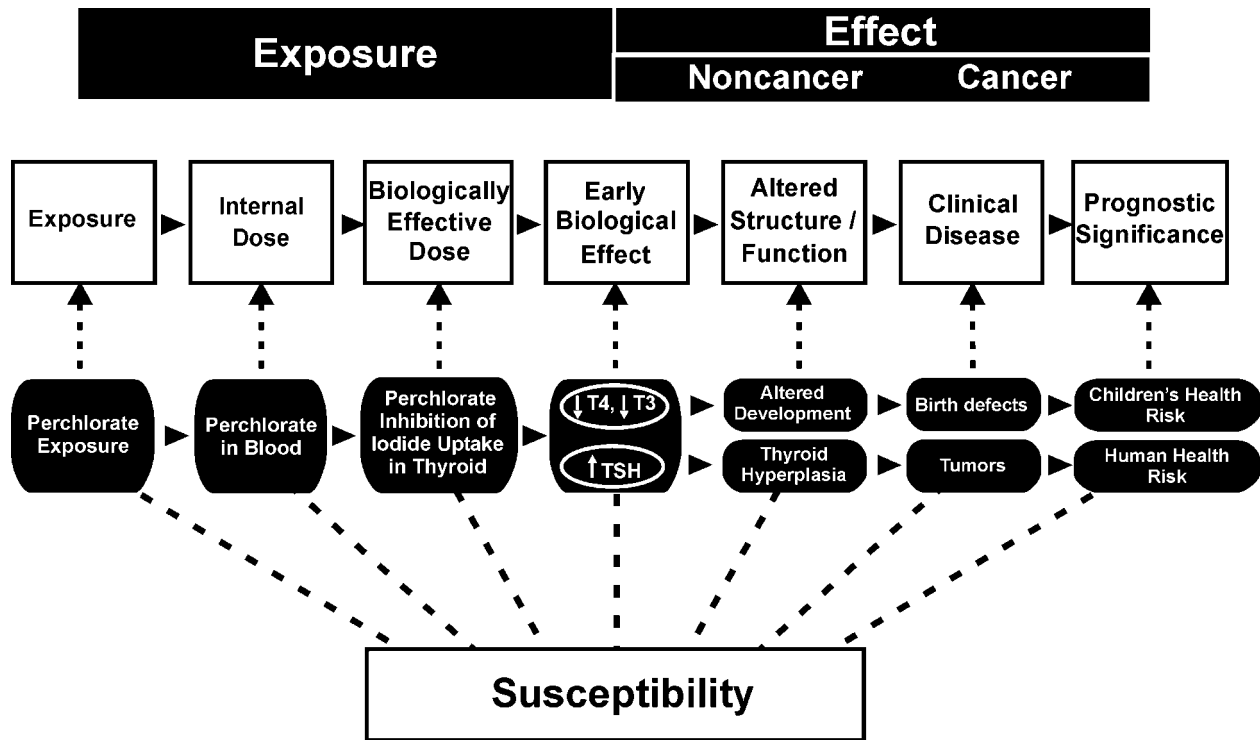
Figure 3-12. **Mode-of-action model for perchlorate toxicity proposed by the U.S. EPA (U.S. Environmental Protection Agency, 1998d). Schematic shows the exposure-dose-response continuum considered in the context of biomarkers (classified as measures of exposure, effect, and susceptibility) and level of organization at which toxicity is observed (U.S. Environmental Protection Agency, 1994; Schulte, 1989). The model maps the toxicity of perchlorate on this basis by establishing casual linkage or prognostic correlations of precursor lesions.**

1   "old " 1998 and "new" 2001), as well as additional studies now available in the literature, will be

2   reported together with EPA's interpretation and evaluation in Chapter 5.

4. HUMAN HEALTH EFFECTS DATA

The available data on the human health effects of perchlorate exposures are limited. Until the emerging concern regarding environmental contamination, the majority of the studies were clinical reports on patients treated with potassium perchlorate for Graves' disease. The non-EPA, independent peer review held in March 1997 (Toxicology Excellence for Risk Assessment, 1998a) concluded that the experimental design limitations of the studies prior to that time precluded their use in quantitative dose-response assessment. The CA DHS also determined in 1997 that there were major limitations on the human studies. Nevertheless, the studies were useful in hazard identification and supported the conceptual model for the mode of action of perchlorate available at the time as described in Chapter 3.

Since the external peer review of the previous 1998 external review draft held in 1999 by the U.S. Environmental Protection Agency (Research Triangle Institute, 1999), some ecological studies have been performed that have addressed the limitations in the human data with some success. Two occupational populations with inhalation exposure to perchlorate were also studied, and some additional clinical studies in healthy adults performed. On December 14, 2001, after internal peer review of this document, the Agency articulated its interim policy on the use of third-party studies submitted by regulated entities (U.S. Environmental Protection Agency, 2001c). For these purposes, EPA is considering "third party studies" as studies that have not been conducted or funded by a federal agency pursuant to regulations that protect human subjects. Under the interim policy, the Agency will not consider or rely on any such human studies (third-party studies involving deliberate exposure of human subjects when used to identify or quantify toxic endpoints such as those submitted to establish a NOAEL or NOEL for systemic toxicity of pesticides) in its regulatory decision making, whether previously or newly submitted. Some of the clinical studies contained in this database fall in this category of studies not to be considered. However, the scientific and technical strengths and weaknesses of these studies were described before this Agency policy was articulated. Therefore, because of the scientific shortcomings of these studies, they will not be used as "principal studies" in the derivation of a RfD. The ethical issues surrounding the conduct of these studies or their use for

1 regulatory purposes in light of the Agency's interim policy will not be discussed in this

2 document. The Agency is requesting that the National Academy of Sciences conduct an

3 expeditious review of the complex scientific and ethical issues posed by EPA's possible use of

4 third-party studies which intentionally dose human subjects with toxicants to identify or quantify

5 their effects.

6

7

4.1 EPIDEMIOLOGICAL DATA

9 To be informative to quantitative dose-response analysis for risk assessment applications,

10 epidemiological studies must pose research questions that are based on appropriate physiological

11 issues relevant to the mode of action for the chemical and its toxic effect. In some contexts, a

12 sufficient specification may take relatively simple form. For example, with occupational cancer,

13 the generally assumed underlying mechanisms lead to a simple test: does exposure to a substance

14 or mixture specified as a dependent parameter, X, at time t_1 increase the incidence of specific

15 cancers at time $t_2 > t_1 + a$, where a >0 is some lag time. The relation of risk at t_2 to the history of

16 prior exposure may be a complex one, but almost always, risk is an increasing function of

17 exposure at various time intervals, $X_{(it)}$. This test may require controlling for confounding

18 factors, which is usually not difficult when relevant detailed information is available.

19 In contrast, determining the effect of an environmental exposure on a regulated system

20 could be more of a challenge. Thus, cancers whose risk depends on endocrine status introduce

21 increased complexity. Environmental perturbations of physiological systems that have inherent

22 variability over time and are imbedded in control networks that function to minimize disruption

23 make it a challenge to determine which endpoints to measure. Cross-sectional assessments

24 during chronic exposures may capture variability in some regulated biological parameters while

25 other parameters will tend to stabilize at "normal" levels despite substantial environmental

26 impact on production and function. In such instances it can be difficult to distinguish alterations

27 due to the xenobiotic from the variation that occurs in response to other environmental factors.

28 Short-term fluctuations in exposure often have no effect independent of cumulative dose for

29 chronic diseases such as lung cancer or other respiratory diseases but may be important for

30 endocrine system functions that affect neurodevelopmental, hyperplastic, neoplastic, immune, or

31 autoimmune events (Park, 2001).

1   The effect of the perchlorate anion on the hypothalamic-pituitary-thyroid feedback system

2   is an example of a regulated system that is potentially difficult to characterize. Important effects

3   may be evident as shifts in average levels of measurable factors, but more important effects may

4   involve alterations in transient responses to demands on the regulated system (Park, 2001).

5   Multiple covariates that may influence potential perchlorate health effects include iodine

6   availability, age, gender, ethnicity, health status, diet, and possibly social class. For neonates, the

7   birth process itself stimulates an endocrine cascade with the amplitudes of endpoint variation

8   depending on birth weight, gestational age, age at sampling (in hours), and possibly

9   environmental temperature. Post-partum developmental risk factors for the neonate and growing

10   child include perchlorate exposure via lactation or consumption of contaminated water.

11   Individual perchlorate exposures are difficult to measure or even estimate in population-

12   based studies. This makes their usefulness to quantitative dose-response analysis limited,

13   particularly if confounding variables are not controlled and small population sizes are evaluated.

14   The few population-based studies from geographic areas that have experienced perchlorate

15   contamination offer little help beyond indicating that clinical thyroid disease is not greatly

16   increased in populations with sustained drinking water contamination as high as 15 μg/L in the

17   past. However, most of the studies have principally evaluated thyroid function or hormone status

18   and have not evaluated neurodevelopmental or other deficits in children or adults resulting from

19   perturbed thyroid function over sustained periods of exposure.

20

21   ## 4.1.1 Ecological Studies

22   Rockette and Arena (1983) reviewed death certificates for workers known to have been

23   exposed to perchloric acid, magnesium perchlorate, and other chemicals in a U.S. chemical plant.

24   Because the workers had received multiple chemical exposures, the authors could not associate

25   an elevated death rate for a particular time period or work area and a specific chemical.

26   The Environmental Health Investigations Branch within the CA DHS, under a cooperative

27   agreement with ATSDR, conducted health assessment activities and consultations on the

28   Aerojet-General Corporation Superfund site in Sacramento County, CA (California Department

29   of Health Services, 1997; 1998a,b,c,d,e). A preliminary health review (California Department of

30   Health Services, 1997) analyzed several statewide databases for possible perchlorate-related

31   outcomes during the suspected years of contamination within the zip codes most likely exposed.

1   In California, thyroid hormone levels in newborns are measured and kept on file by the Genetic

2   Disease Branch of the Centers for Disease Control and Prevention. Data for the period 1985

3   through 1996 from relevant zip codes was assessed for a total of 11,814 thyroid hormone screens.

4   Although an extrapolation of the statewide rate predicted there would be 3.76 cases of

5   hypothyroidism, four cases were observed. In the non-exposed areas, six cases of

6   hypothyroidism were found although 6.41 cases were predicted. These data suggested no

7   association between residence in the potentially-exposed zip codes and neonatal hypothyroidism.

8   The TSH levels (ascertained only in neonates with initially low T4 levels) in the potentially-

9   exposed areas were statistically significantly lower than those in the nonexposed areas. The

10   database also was evaluated for diagnosis of goiter among the first five reported hospitalized

11   individuals residing in the zip code of most likely contamination from the years 1991 to 1995.

12   Because there are so many diseases or conditions that can produce a goiter other than perchlorate

13   ingestion, and because the database can not differentiate this aspect, it was concluded that these

14   data would not be useful in determining the prevalence of thyroid enlargement due to perchlorate

15   in the affected water district. The same zip code also was evaluated for agranulocytosis or

16   aplastic anemia as one of the top five diagnoses for the years 1991 to 1995. There were a total of

17   76 cases in 5 years, less than the statewide rate of 41.6 per year. The rate for aplastic anemia was

18   3.8 hospitalizations per 100,000 individuals per year, a rate higher than the statewide rate of 2.2.

19   However, all but one of the hospitalizations also had an additional diagnosis of cancer with

20   chemotherapy or radiation treatment; these treatments are likely explanations for this

21   observation; acquired immunodeficiency syndrome (AIDS) may be another. The registry also

22   was searched for cases of childhood leukemia (either acute lymphocytic leukemia or acute

23   myelogenous leukemia). Again, the rate for the potentially exposed zip code was less than the

24   corresponding rate for California.

25   The CA DHS concluded that the data on goiter, agranulocytosis, and aplastic anemia did

26   not indicate an increase in incidence; however, these data do not provide definitive causitive

27   information because other likely causes for these conditions existed. Increases in the incidence

28   of decreased neonatal thyroid hormone levels, hypothyroidism, or childhood leukemia rates were

29   not observed. The CA DHS noted that the major limitation on studies of this nature is that

30   imposed by the absence of good exposure estimates and the absence of data on transport and

31   transformation models which would provide dose reconstruction for the affected population. It is

1 unclear when the contaminated plume entered the drinking water supply; consequently, the time

2 period analyzed may have been too broad. Improving this exposure information was one of the

3 recommendations made in the report to Congress regarding perchlorate (U.S. Environmental

4 Protection Agency, 1998e). Finally, that perchlorate is not specific for producing thyroid

5 dysfunction or hematological abnormalities makes assessing these outcome surveys difficult.

6 Table 4-1 shows the approximate prevalence of these disorders in the neonatal period

7 (1:30,000 to 1:100,000), and suggests that studies with large numbers of subjects may be

8 necessary to detect subtle effects.

9   Based on these results, the CA DHS investigated several other water service areas for

10 exposure (California Department of Health Services, 1998a,b,c,d,e) and ascertained that

11 complete exposure pathways to perchlorate contaminated water existed in several areas. These

12 studies reinforced the need for this document which attempts to properly characterize the risk

13 posed by perchlorate contamination by providing better exposure estimates and a revised health

14 risk estimate.

15   Since the 1999 external peer review, eight new population studies have been performed.

16 One of these studies has examined effects in the general population (Li et al., 2001), another in

17 school-age children (Crump et al., 2000), and six have been devoted to evaluating neonatal

18 endocrine status in areas with contaminated drinking water (Crump et al., 2000; Lamm et al.,

19 1999; Li et al., 2000a,b; Brechner et al., 2000; Schwartz, 2001). In each study, the critical

20 covariates were captured with varying degrees of success and only one study (Schwartz, 2001)

21 offers a convincing description of neonatal perchlorate effects (Park, 2001).

22   In a study of the general population, Li et al. (2001) investigated physician-generated

23 medical insurance claims for thyroid problems in a Medicaid insured population in Nevada,

24 comparing all counties that were known not to have perchlorate contaminated drinking water

25 with the one county that had contamination at approximately 10 μg/L. This was a study of

26 period-prevalence, i.e., the proportion of the population that had claims for thyroid-related

27 disorders anytime during a two-year period. Incident cases could not be identified within this

28 database. Thyroid patients were defined as having one or more of the following diagnoses of

29 thyroid disease according to the International Classification of Diseases, 9[th] Revision (ICD-9):

30 (1) simple and unspecified goiter (ICD-9 Code 240); (2) non-toxic nodular goiter (ICD-9 Code

31 241); (3) thyrotoxicosis with or without goiter (ICD-9 Code 242); (4) congenital hypothyroidism

TABLE 4-1. THYROID DISORDERS AND THEIR APPROXIMATE PREVALENCES IN THE HUMAN NEONATAL PERIOD (Fisher, 1996).

Thyroid Dysgenesis	1:4000
Agenesis	
Hypogenesis	
Ectopia	
Thyroid Dyshormonogenesis	1:30,000
TSH unresponsiveness	
Iodide trapping defect	
Organification defect	
Defect in thyroglobulin	
Iodotyrosine deiodinase deficiency	
Hypothalamic-Pituitary Hypothyroidism	1:100,000
Hypothalamic-pituitary anomaly	
Panhypopituitarism	
Isolated TSH deficiency	
Thyroid hormone resistance	
Transient Hypothyroidism	1:40,000
Drug induced	
Maternal antibody induced	
Idiopathic	

1   (ICD-9 Code 243); (5) acquired hypothyroidism (ICD-9 Code 244); (6) thyroiditis (ICD-9 Code

2   245); (7) other disorders of the thyroid (ICD-9 Code 246) and (8) malignant neoplasms of the

3   thyroid gland (ICD-9 Code 193). Two of these disorders have very low prevalence: congenital

4   hypothyroidism (0.01%) and thyroid cancer (0.02%).

5   Comparisons were made between the exposed county, which includes Las Vegas, and

6   (a) an unexposed county with a similar large city (Reno), and (b) all other counties (unexposed).

7   There were no statistically significant period-prevalence rate differences between the exposed

8   county and the two categories of comparison counties; however, the differences between the

9   comparison county groups themselves were quite large, indicating that either important

10   confounding risk factors were not controlled or estimates were unstable due to the small numbers

11   of cases in the comparison counties. For acquired hypothyroidism, prevalences (%) in the two

12   categories of unexposed counties were significantly different (Reno: 1.17 [95% CI = 1.05 to

13   1.30, using a normal approximation to the Poisson distribution for number of cases] and other

1   counties: 1.44 [95% CI = 1.29 to 1.59]). Age, gender, ethnicity, iodine intake, and other

2   important risk factors were unavailable in this database and there could have been differential

3   under- or over-diagnosis in this Medicaid population. Interestingly, when comparing the two

4   counties with large urban centers and restricting focus to the 6 (out of 8) more prevalent

5   outcomes (total n=3069), all 6 showed elevated (but not individually significant) rate ratios for

6   the exposed county, ranging from 1.01 to 1.89. While these findings appear to rule out a large

7   perchlorate-related excess (i.e., greater than two-fold) for some thyroid disorders such as

8   acquired hypothyroidism (appearing as routine medical insurance claims), the study had a

9   statistical power of less than 0.5 to detect a 50% excess for several specific thyroid disorders

10   (i.e., the observed relative rises exceeded 1.50 but were not statistically significant).

11   Unfortunately, owing to potentially overwhelming confounding (e.g., related to age, gender,

12   ethnicity, or iodine intake) or because of small numbers of cases in the comparison counties, little

13   else can be concluded from this study.

14   The Crump et al. (2000) study of school children (mean age 7.3 years) in three Chilean

15   cities permitted comparisons on effects of drinking water with widely varying perchlorate

16   content: 0, 5, and 100 ppb. A total of 162 school-age children were studied, 127 of whom had

17   lifelong residence in their respective cities. Controlling for age, gender, and urinary iodine,

18   a highly significant trend of increasing T4 levels—the opposite to the expected direction for

19   effects on T4 from perchlorate—was observed with increasing perchlorate content in the water.

20   The city with the highest concentrations (100 ppb) had a significant five-fold excess in family

21   history of thyroid-related problems. Children in all three cities had elevated goiter prevalence,

22   but it was highest in the city with intermediate concentrations (5 ppb) which was believed to also

23   have iodine deficiencies. A variable introduction of iodized salt in earlier years may have

24   affected these observations. It is not known what role boiling drinking water may have played or

25   how the microbiological quality of drinking water varied across the cities studied. Ethnic and

26   socioeconomic attributes were thought to be similar across the three groups of children but were

27   not controlled for in the analysis. Whether ambient indoor and outdoor temperatures may have

28   played a role in thyroid functional status was not investigated. It would appear that uncontrolled

29   confounding effects, particularly from environmental or other factors, make it difficult to

30   interpret the observed effects of drinking water contaminated with perchlorate at levels as low as

31   5 ppb on thyroid function in this study. Controlling for urinary iodine in the analyses would

1   better address whether iodine deficiency differences across the three cities studied may have

2   distorted the association of T4 changes with perchlorate exposure. The paradoxical trend

3   observed in this study remains unexplained.

4   Crump et al. (2000) also studied newborns screened for hypothyroidism by a heel-stick

5   blood sample between February 1996 and January 1999 in the same three Chilean cities.

6   A systematic laboratory error gamma counter contamination occurred between December 1, 1997

7   and June 30, 1998 which caused TSH to be reported very low (0.1 μU/mL) for a high proportion

8   (29.1%) of the blood samples analyzed. The error was reported to be limited to this 7-month

9   period and to have affected a similar proportion of samples from each of the three cities. All data

10   obtained during the 7-month period in question were excluded, leaving 9,784 neonatal records

11   for analysis. Analysis revealed a statistically significant decline in TSH (log-transformed) with

12   increasing city-perchlorate levels, a trend opposite to that hypothesized. The analysis was

13   adjusted for gender and age at screening as categorical variables in days but covariates lacking

14   included iodine intake (known to be low in one city), ethnicity, and birth weight. The ages at

15   screening differed across the three cities studied; the median ages were 3, 4, and 6 for the

16   unexposed, low, and high perchlorate studies, respectively. Other important environmental

17   factors may have played a role such as ambient temperatures, caloric intake, and social class.

18   This paradoxical finding parallels the similar result in school age children in the same Chilean

19   population discussed above, and remains unexplained.

20   Lamm et al. (1999) examined rates of congenital hypothyroidism in 7 counties of California

21   and Nevada with perchlorate contaminated drinking water. This outcome is defined as a result of

22   a mandatory screening program at birth that involves a preliminary T4 determination followed by

23   a TSH assay in a prescribed subset with low T4. Age at screen is not considered in this

24   procedure for selecting candidates for TSH determination and screening age distributions by

25   county were not reported. County-specific levels of perchlorate contamination were unavailable.

26   Rates for the California births were adjusted only for Hispanic ethnicity, observed to be a risk

27   factor in this and other studies (Brechner et al., 2000; Schwartz, 2001). The county rate ratios for

28   congenital hypothyroidism ranged from 0.6 to 1.1 relative to the statewide expected rates and

29   were not statistically significant for all exposed counties combined, the rate ratio was 1.03 (95%

30   CI = 0.90 to 1.16). Expected rates based on the non-exposed counties of the two states were not

31   used. Had only non-exposed counties been used for comparison (given that the exposed counties

1   comprise a substantial fraction but assuming it is less than half of the state's population) the

2   resulting rate ratios for the exposed counties would have been 1% or higher. Most critically

3   lacking in the analysis was classification on age at time of blood sample for the screening test.

4   Birth weight and further detail on ethnicity and other risk factors were also unavailable.

5   Therefore, it is likely that uncontrolled confounding has played a role in this study, making it

6   difficult to interpret and allows for some role of perchlorate in the almost two-fold observed

7   variation in risk of neonatal hypothyroidism across counties.

8   Li et al. (2000a) compared the mean monthly T4 levels derived from mandatory screening

9   of all newborns in Las Vegas (exposed) and Reno (unexposed), controlling for birth weight

10   (within the restricted range 2.5-4.5 kg) and for age at sample (days 1, 2 or 3 versus 4), for the

11   period April 1998 through June 1999. Statistical differences in the mean birth weight and mean

12   age at time of sample were noted for the Las Vegas (n = 17,308) and Reno (n = 5,882) newborns.

13   The exposure variable was based on monthly measurements made on Las Vegas finished water

14   by the Southern Nevada Water Authority using IC with a detection limit of 4 ppb. The source of

15   the Las Vegas water supply, Lake Mead, is known to have thermal stratification that causes

16   seasonal variation in drinking water perchlorate content. The water supply in Reno comes from

17   the mountains via Lake Tahoe, the Truckee River, and local wells. Tests of these water sources

18   for Reno were reported to detect no perchlorate (data not shown nor was it specified if these

19   measurements were made monthly). A highly significant period or seasonal effect was observed

20   for both cities (perhaps suggesting an ambient temperature effect), but no difference was

21   observed between cities during the period of exposure (7 out of the 15 months of observation

22   when perchlorate content was high in Las Vegas drinking water). Highly significant age effects

23   were observed, but the dependence of these age effects on exposure (i.e., an exposure interaction)

24   was not examined. For reasons that are obscure, T4 levels reported in this study were

25   considerably higher than those reported by others (17 versus 7-10 μg/dL). The restriction on

26   birth weight would be inappropriate if birth weight were an intervening variable (i.e., itself

27   affected by thyroid changes resulting from perchlorate exposure). Regressions on first trimester

28   and 9-month cumulative exposures using monthly perchlorate levels and grouping birth

29   outcomes by month in Las Vegas and Reno revealed no trends for T4 differences between the

30   two cities although more powerful analyses could have been performed using individual

31   observations. This study suggests that clinically significant individual neonatal T4 differences

1   have not resulted from current perchlorate exposures although the possibility of important

2   variation with exposure conditional on neonatal age was not examined.

3   In a parallel study design, Li et al. (2000b) studied TSH levels in Las Vegas and Reno

4   newborns over an eleven-month period from December 1998 to October 1999. Las Vegas water

5   had measurable perchlorate levels in 8 of the 11 months. The perchlorate exposure measures and

6   assumptions were the same as in Li et al. (2000a). TSH levels were determined on screening

7   samples that were below the 10th percentile on T4 in each daily batch of samples collected

8   throughout the state, selected without regard to age at screening. TSH levels from the two cities

9   for birth weights restricted to 2.5-4.5 kg were compared adjusting for gender and age at screen

10   (days 2-7 versus 8-30). Births whose screening sample was taken on the first day were excluded

11   because those TSH levels were considered unstable. The study did not report whether the age at

12   screen distribution differed between the two cities. Ethnicity and other risk factors were not

13   available. Using a log-transformed TSH level to facilitate statistical testing, they found no

14   difference in TSH levels between the two populations (a very small negative effect was estimated

15   for TSH with exposure), however, the log transformation may have suppressed important

16   differences at the high end of the TSH distribution and the analysis was not restricted to the

17   8 months when exposure differed between the two cities. Examination of an exposure with age

18   interaction was not reported. Excluding births screened on the first day may have further

19   obscured differences arising from perchlorate exposure, differences that pertain to thyroid

20   responsiveness. This study suggests that TSH levels in newborns after the first day did not differ

21   substantially between two cities with and without perchlorate contamination of drinking water as

22   estimated by monthly measurements.

23   Brechner et al. (2000) studied TSH levels in Arizona newborns assayed over a three-year

24   period between October 1994 and the end of December 1997 in the Arizona Newborn Screening

25   Program. In this program, total T4 is assayed in daily batches of specimens received from all

26   over the state. TSH is measured in selected samples, representing approximately 10% of the

27   samples with the lowest T4 levels from each batch. TSH levels were compared between two

28   cites, Flagstaff and Yuma, representing areas of nonexposure and exposure to perchlorate. Zip

29   codes were used to determine that Yuma was the only area with essentially all of its drinking

30   water supplied by the Colorado River below Lake Mead. Exposure data were not available for

31   the period between 1994 and 1997. Measurements made by the U.S. Environmental Protection

1   Agency Region 9 laboratory in August 1999 reported perchlorate levels at 6 ppb in both raw and

2   finished water for Yuma and not detectable in Flagstaff water. Because the water processing

3   facilities have not changed in either city and perchlorate is known to persist for long periods,

4   Brechner and colleagues presumed that comparable differences between the perchlorate levels in

5   the two cities existed during the period of analysis. Controlling for age at screen (days 0, 1-4,

6   5+) and Hispanic ethnicity, these investigators found a statistically significant elevation in TSH

7   for the exposed population in Yuma (crude TSH: 19.9 versus 13.4 mU/L; adjusted TSH effect

8   not reported). However, the age-at-screening distributions differed considerably between these

9   two cities presenting a possibility for some residual confounding on age. In Yuma (exposed)

10   5.9% of newborns were screened in the first 24 hours when TSH levels peak (mean TSH =

11   30 mU/L), compared with 2.4% of Flagstaff newborns (mean TSH = 23 mU/L). Thirty-one

12   percent of Yuma births were screened at day 6 compared with 46% of Flagstaff births.

13   Additionally, because of this negative association between age and exposure, the analysis of

14   variance procedure employed had the potential for bias arising from colinearity. The age and

15   exposure effect estimates would be jointly affected: overestimating exposure and

16   underestimating age effects, or visa versa. Other factors not controlled included gender and birth

17   weight. This study offers positive support for an association of increased neonatal TSH with

18   perchlorate exposures; however, similar to other studies on this question, it has some unresolved

19   methodological issues, most notably the strong association between age at screen and perchlorate

20   exposure.

21   There is a subtle form of bias in the Brechner and other studies where TSH was determined

22   on a low - T4 percentile subset of all births that mixes on a daily basis ages at screen for samples

23   from all over the state. Bloods with low T4 are selected, but the T4 distribution depends on age.

24   Births with screen ages that usually have higher T4 (typically after 24 hr) are less likely to be

25   selected for TSH determination; conversely, at ages under 24 hr, births are more likely to be

26   selected. Both summary and age-specific TSH comparisons would be unbiased with respect to

27   exposure effects only if the same age at screen distributions were obtained in both the exposed

28   and unexposed populations. The effect of this bias on estimation of overall perchlorate exposure

29   effects is difficult to predict, depending in part on how perchlorate exposure affects T4 as well as

30   on its effects on TSH, and on how sampling age varied with exposure status. It is conceivable

31   that this bias could explain some of the elevated TSH in perchlorate-exposed neonates of the

1   Brechner et al. (2000) study, but the same sampling bias was potentially present in the Li et al.

2   (2000b) study that found no effect. The latter study, however, excluded neonatal blood samples

3   taken during the first 24 hours. That is the period when the strongest perchlorate-related

4   differences were observed in the Brechner et al. (2000) study.

5   Schwartz (2001) analyzed both T4 and TSH levels for all California newborns screened in

6   1996, making use of detailed covariate information on age, birth weight, ethnicity and birth

7   multiplicity. Perchlorate exposure was assigned using the mothers' postal zip codes that were

8   linked to state water testing data on all drinking water sources. These estimates of perchlorate

9   levels were ultimately collapsed into four exposure categories: 0, 1-2, 3-12, 13+ ppb. This level

10   of exposure detail far exceeded that of any other studies, very likely resulting in the least

11   exposure misclassifications.

12   An analysis of covariance (ANCOVA) model was used in this analysis. The ANCOVA

13   model is a multiple linear regression model that can simultaneously estimate effects for levels

14   categorical variables like gender as well as for continuous variables like age or birth weight.

15   Controlling for age at screening (6-hour increments up to 48 hours), gender, single versus

16   multiple birth, birth weight (in 5 levels), and ethnicity (20 categories), a highly statistically

17   significant declining trend was observed for T4 with the four perchlorate exposure levels (0,

18   -9.7, -11.2, -18.2). T4 levels in this model declined with age (relative to its final level after

19   48 hours) until about 18 hrs (-50 mg/dL below final level) and then increased over the next 30

20   hours (to 36 mg/dL above final level) before assuming its final level after 48 hours. For TSH

21   (log-transformed), there was a significant increasing trend with perchlorate exposure (0, 0.029,

22   0.03, 0.128), and the TSH level followed a more rapid time course increasing immediately after

23   birth, then declining to a final level by 24 hours. Substantial birth weight, gender, ethnicity and

24   birth multiplicity effects were observed for T4, and smaller effects were observed for TSH.

25   The models specified in this study tested for uniform additive exposure effects for T4 and

26   TSH across all covariate categories, including baseline shifts. Another issue of considerable

27   physiological interest would have been whether the amplitudes of the T4 and TSH surges

28   depended on perchlorate exposure with baseline levels relatively unaffected, which could be

29   tested by evaluating an interaction between age and exposure. An examination of interaction was

30   not reported. The bias in TSH measurements introduced by the T4-triggered sample selection

31   described above for other effects studies would also affect the Schwartz study. This bias would

1   not affect inferences on exposure effects if the age at screen distribution were similar across the

2   four exposure levels. These distributions were not reported in the Schwartz study.

3   The Schwartz study also modeled the effect of two screening performance criteria on the

4   same set of predictors: (a) "presumptive positive criterion" and (b) a positive finding of

5   congenital hypothyroidism. Not surprisingly, these models did not predict the standard screening

6   outcomes well because the screening algorithm does not take into account the several very

7   important predictors identified in this study. Rather, finding a presumptive positive is based

8   entirely on T4 without regard to age at screen, birth weight, etc. Similarly, identifying a case of

9   congenital hypothyroidism is based only on T4-triggered sample selection and subsequent TSH

10   determination (>25 μU/ml).

11   The Schwartz study is by far the most convincing of the neonatal studies, being based on

12   the most elaborate exposure assignment and the most detailed collection of covariate information

13   pertaining to neonatal thyroid function. It is unlikely that bias arising from the TSH sampling

14   could produce such a consistent TSH exposure response and would play no role in the stronger

15   (based on narrower confidence intervals for the parameter estimates) exposure response observed

16   for T4.

17

18   **4.1.2 Occupational Studies**

19   There are two publications investigating workers in ammonium perchlorate production

20   (Gibbs et al., 1998; Lamm et al., 1999). The route of exposure for each was by inhalation to

21   perchlorate dust, introducing a considerable uncertainty in dose-response analysis especially due

22   to poor characterization of particle size distribution. Both studies were also cross-sectional in

23   design and, therefore, subject to survivor bias in that workers experiencing adverse effects could

24   have left employment. This issue was not addressed in either study. It would have been

25   particularly noteworthy had any former employee no longer in the cohort experienced thyroid

26   disorders, aplastic anemia, or related hematological disorders, each of which have been reported

27   in settings where perchlorate is used for short periods at higher doses in the treatment of disease

28   (Lawrence et al., 1999). The airborne exposures that were characterized corresponded to daily

29   doses on the order of 20 to 50 mg and possibly higher as the air-sampling methods excluded

30   large particulate (> 50 μm) that could add considerable mass to the daily inhaled or ingested

1   dose. In the study that investigated this (Lamm et al., 1999), the daily absorbed dose based on

2   urinary perchlorate actually exceeded the inhaled dose.

3   There was no clear evidence for any perchlorate effect on thyroid function, as defined by

4   the investigators, in these two cross-sectional occupational studies. However, historical exposure

5   classification was limited in one study and absent in the other. Former employees were lost to

6   follow-up, and neither study controlled for potential confounding arising from body mass,

7   environmental temperatures, or socioeconomic status. There was no measurement of thyroid

8   iodine status or of any index thyroid dynamic responsiveness that conceivably could be altered

9   even though steady-state TSH and T4 levels appear to be in the normal range. Because of the

10   cross-sectional design and measured endpoints, the studies did not evaluate the dynamics of

11   hypothalamic-pituitary-thyroid feedback that are likely important in target populations such as

12   hypothyroxinemic pregnant women and their fetuses.

13   Gibbs et al. (1998) performed a case control occupational epidemiology study to evaluate

14   thyroid function and standard clinical blood test parameters of liver, kidney, and bone marrow

15   function in employees exposed to ammonium perchlorate airborne dust at a production facility

16   and an associated cross-blending facility. Exposure estimates were based on multiple samples

17   (average sample number = 17) for eight homogenous exposure groups defined by similar job

18   activities: control, maintenance/foreman, and six discrete operator job categories. Personal

19   breathing zone samples (n = 119) were used for the work categories and full-shift area samples

20   were used for the control group (n = 19). The control exposure was not zero but was several

21   orders of magnitude below any exposure category. In their 1997 analyses, when ammonium was

22   quantified using National Institute for Occupational Safety and Health Method 6016 which had a

23   minimum reporting limit of 0.017 mg/m^3, concentrations in a large number of the samples were

24   reported as undetectable. The 1998 analyses were performed using the modified EPA 300.0

25   methodology that determines perchlorate using ion chromatography and has a reporting limit of

26   approximately 0.00004 mg/m^3.

27   Effects were examined in either a single-shift design (pre- and post-shift parameter

28   measurements) or working lifetime design based on medical surveillance data that included

29   thyroid examination since 1996 (blood tests, physical exam, and history since 1994). Dose was

30   reconstructed based on personnel records for job type and area samples.

1   Despite the lack of particle size distribution data, an inhaled "dose" was calculated for a

2   single shift as (Gibbs et al., 1998):

$$\left(\begin{array}{c}\text{respiratory} \\ \text{rate}\end{array}\right) \times \left(\begin{array}{c}\text{inhalation} \\ \text{concentration}\end{array}\right) \times \left(\begin{array}{c}\text{exposure} \\ \text{duration}\end{array}\right) \times \left(\begin{array}{c}\text{fraction} \\ \text{absorbed}\end{array}\right). \qquad (4\text{-}1)$$

3

4   Working lifetime exposure estimates were calculated as:

5

$$\sum (\text{mean group exposure}) \times (\text{years in exposure group}) \times 2{,}000, \qquad (4\text{-}2)$$

6

7   where the 2,000 was an average of the number of hours worked yearly based on typical overtime

8   rates at the facilities.

9   Daily respiratory rates of 0.0165 m^3/kg-hr and 0.0068 m^3/kg-hr were estimated for "active"

10   and "sedentary" workers, respectively, based on Beals et al. (1996). These estimates are slightly

11   lower than the default EPA respiratory rates and are moderately lower than those recommended

12   by the International Commission on Radiological Protection in its recent human respiratory tract

13   model (International Commission on Radiological Protection, 1994). Average body weights of

14   the workers were larger than the typical default body weights. Because current practice usually

15   scales ventilation rate based on body weight, higher ventilation rates were expected.

16   The absence of particle size diameter and distribution data is a significant limitation of the

17   study because this data is required to assess the potential inhalability of the ammonium

18   perchlorate aerosol. Data from another production facility indicate the majority of particles are

19   200 μm (Hancock, 1998). Particles larger than 30 μm are typically not inhalable by humans

20   (U.S. Environmental Protection Agency, 1996b). Further, there was no mention of face volume

21   performance of the personal samplers using 5-μm filters although this is an important

22   consideration in dusty environments when the particles under investigation have a large diameter.

23   This consideration is especially important here because the filter cassettes were changed when

24   respirators were used. Even if a 5-μm particle diameter could be assumed, the inhaled "dose"

25   calculation should have included an adjustment for inhalability and deposition efficiency to

1   calculate the deposition fraction, approximately 0.3 at 5 μm (U.S. Environmental Protection

2   Agency, 1996b).

3   The assumption about the solubility of the inhaled particles is also problematic because this

4   parameter is particle-diameter dependent. The particle diameter dictates the location

5   (extrathoracic, tracheobronchial, pulmonary) where the particle deposits and the local milieu and

6   clearance vary with location also influence solubility (U.S. Environmental Protection Agency,

7   1996b; Snipes et al., 1997). The solubility of cesium chloride (CsCl) in beagles was used to

8   estimate a fraction absorbed. Although CsCl and NH_4ClO_4 may have similar solubilities,

9   additional uncertainty is introduced because the CsCl particle diameter or inhalability function

10   for the beagles was not taken into account; and the hydroscopicity, which influences the initial

11   deposition site, may not be the same. The assumptions about dose could have been validated

12   with a mass balance approach. For example, perchlorate could have been measured in the blood

13   when samples were taken for thyroid hormone analyses. Additionally, urine samples could have

14   been monitored for perchlorate because it is excreted in the urine. These additional

15   measurements would have afforded some confidence that the inhaled dose estimates were

16   reasonable.

17   Standard clinical thyroid profiles included a total serum T4, triiodothyronine resin uptake,

18   and TSH. Bone marrow function was evaluated during medical surveillance examinations with

19   standard tests from the complete blood count which included hemoglobin, hematocrit, red blood

20   cell count, mean corpuscular volume, white blood cell count, and platelet count. Standard serum

21   chemistries were used to assess kidney (serum creatinine level and blood urea nitrogen) and liver

22   (serum glutamyl pyruvic transaminase [SGPT], serum glutamyl oxaloacetic transaminase

23   [SGOT], g-glutamyl transpeptidase [GGTP], and alkaline phosphatase) functions.

24   Dependent variables for the single-shift study were the cross-shift change in measures of

25   thyroid function. Explanatory variables included race, gender, age, hours awake prior to the

26   pre-shift test, number of hours slept during the most recent period prior to the test, time of day,

27   and shift length. Dependent variables for the working lifetime included measures of thyroid,

28   bone marrow, liver, and kidney functions. For the thyroid tests, an additional explanatory

29   variable was used to indicate whether the measurement was from a routine physical in 1996 or

30   from a pre-shift or a post-shift examination in 1997 or 1998. The dose variables were group

31   (control, low dose, or high dose) and estimated cumulative exposure. The dose group

1   designation was an arbitrary stratification of <8 mg/kg-day and >8 mg/kg-day. Multiple

2   regression was used to analyze the relationship between effect measures and explanatory

3   variables. A sequential approach was used to determine whether a dependent variable should be

4   log-transformed and whether any outliers (defined as a value corresponding to a residual larger in

5   absolute value than three standard deviations) should be eliminated from an analysis.

6   Estimated doses for the single shift-study ranged from 0.0002 to 0.436 mg/kg-day with a

7   mean of 0.036 mg/kg-day and median of 0.013 mg/kg-day. The dose estimate was not a

8   significant predictor of thyroid function parameters measured in 83 control (65 male, 18 female)

9   or 18 exposed (15 male, 3 female) individuals. Working lifetime exposure estimates ranged from

10   0.5 to 7.0 (mean 3.5) mg/kg for the low-dose group and from 8.0 to 88.0 (mean 38.0) mg/kg for

11   the high-dose group. The duration of exposure ranged from 1 to 27 years (mean 8.3).

12   No significant correlations were detected in any measures of thyroid, bone marrow, liver, or

13   kidney function; however, significant gender and race differences were apparent in the clinical

14   tests of bone marrow, liver, and kidney functions. Females were slightly lower in hemoglobin,

15   hematocrit, SGPT, GGTP, and creatinine than males; black workers were slightly lower than

16   whites in hemoglobin and hematocrit and slightly higher in creatinine.

17   The only significant finding ($p = 0.01$) was that cross-shift TSH changes were greater for

18   those who worked a 12-h shift than for those who worked 8-h shifts, accounting for a

19   0.45 urinary IU/mL increase across the shift. This was attributed to the influence of circadian

20   changes in serum TSH. However, the TSH increase (10%) across a single work shift in an

21   exposed group ($n = 18$) compared to an unexposed group ($n=83$) was observed in groups that

22   together comprised less than half of employees eligible for study. Comparison of workers in

23   three groups (unexposed, low and high cumulative exposure) resulted in consistent patterns for

24   all thyroid parameters in which the unexposed group had values intermediate between those of

25   the low and the high cumulative dose groups. This suggests that important confounding was

26   present (i.e., that the comparison group, which apparently included office workers, differed from

27   the exposed groups on other important risk factors) as well. For thyroid (TSH) and liver

28   outcomes (SGOT, GGPT, SGPT), there were subtle indications of exposure effects: the standard

29   deviation increased substantially in the high dose group, as did the average values but not the

30   percentiles up to the 75[th], suggesting that a small subgroup had undergone a considerable upward

1   excursion. Statistical tests (regression analysis) of these effects were severely limited by the

2   apparent confounding that affected baseline levels.

3   In the second study of ammonium perchlorate workers, Lamm et al. (1999) assembled a

4   comparison group at the same facility from an unrelated process thought to have low exposure to

5   inhaled perchlorate.   Workers were classified using presumptive exposure based on visible dust

6   generated. Pre- and post-shift urine samples were collected to measure urinary perchlorate,

7   iodine and creatinine levels. Post-shift blood samples were analyzed for complete blood count

8   (CBC), hemoglobin, hematocrit and additional red cell parameters (mean corpuscular volume,

9   mean corpuscular hemoglobin, and mean corpuscular concentration). A clinical chemistry panel

10   was also run on post-shift serum samples. Thyroid parameters included TSH, free T4, T4, T3,

11   thyroid hormone binding ratio, thyroid peroxidase antibodies, and clinical examination. Urinary

12   perchlorate measurements were used to calculate a post-shift level of perchlorate (mg) per g of

13   creatinine as an excretion dose, D:

14

$$D = k[E_i - 0.354\ E_0]/0.646. \tag{4-3}$$

15

16   The right hand term in brackets is the post-shift adjusted level in mg perchlorate per gram of

17   creatinine. Perchlorate absorption was calculated as a time-weighted average exposure using an

18   assumption that the percent absorbed which is excreted is 95%. The human adult creatinine

19   excretion rate was then used to link perchlorate excretion rate in terms of creatinine to rates in

20   terms of time, so that the exposure dose was then calculated as:

21

22   12 hours x 60 minutes / hour x 0.001 g/mg x 1 mg creatinine/min x [post-shift]/0.646.   (4-4)

23

24   While particle size distribution data were collected, this information was not utilized in the

25   analyses. Inhalation exposure was instead categorized into either "total" or "respirable". While

26   these categories correlated with each other to a good degree (r = 0.82), perchlorate absorption

27   (mg/shift) did not correlate as well to respirable (r = 0.45) as it did to total particles (r = 0.54).

28   The comparison group had current absorbed doses equal to 20% of the low perchlorate-exposed

29   group and 3% of the high exposed group even though the inhaled dose of the comparison group

30   was 4% of that of the low dose and 0.02% of the high dose group. This suggests that there was

1 considerable exposure misclassification, arising perhaps from general environmental

2 contamination at the work site or in clothing. In one subject, urinary perchlorate increased over a

3 12 hr period during which there was thought to be no exposure. No significant associations were

4 observed between perchlorate exposure and thyroid parameters; however, measures of

5 cumulative exposure were not considered. Suggestions of increasing trends for T3, T4, and

6 maximum-T3 were not statistically significant but were based on small numbers (numbers of

7 workers in exposure groups: 21 for the unexposed versus 14, 8, and 15 in the low, medium, and

8 high exposure groups).

9

10

11 ## 4.2 CLINICAL STUDIES

12 The historical clinical data on perchlorate have been predominantly case reports of patients

13 whose results would be confounded either with thyroid disease or other pharmaceutical agents.

14 A few more recent studies have begun to evaluate thyroid function in healthy volunteers. This

15 section will discuss the available data on thyroid function from several clinical studies. A more

16 formal development of the pharmacokinetic data in humans is presented in Chapter 6.

17

18 ### 4.2.1 Studies in Healthy Human Subjects

19 Few data are available to demonstrate the effects of perchlorate on healthy individuals and

20 issues of ethics are likely to preclude clinical evaluation in sensitive populations such as pregnant

21 women. Exposure duration to perchlorate is typically short, from a few days to 4 weeks.

22 Burgi et al. (1974) examined the effects of perchlorate on the secretion of endogenous

23 iodine by the normal human thyroid gland. Five healthy volunteers (3 males, 2 females;

24 ages 24 to 27 years) received tracers of ^{125}I-iodide and ^{131}I-thyroxine for 17 days, followed by

25 600 mg/day perchlorate (9.7 mg/kg-day, based on actual reported average body weight of

26 61.8 kg) for 8 days. Urine and serum were analyzed for ^{125}I and ^{131}I to determine if perchlorate

27 can cause the discharge of endogenous, as well as exogenous iodide, from the thyroid. Results

28 show that this dose of perchlorate also was sufficient to completely block iodide uptake by the

29 thyroid. In addition, perchlorate caused a 65% increase in excretion of nonthyroxine iodide over

30 background. The authors attributed this increase to additional secretion of endogenous iodide

1   from the thyroid.   Treatment with carbimazole plus perchlorate caused an additional increase in

2   the secretion of nonthyroxine iodide, suggesting that perchlorate causes only a partial release of

3   endogenous iodide. This study suggests a Lowest-Observed-Adverse-Effect-Level (LOAEL) of

4   9.7 mg/kg-day for thyroid effects in healthy patients.

5   Brabant et al. (1992) administered potassium perchlorate to five healthy male volunteers

6   (age 25 to 28 years) to study changes in TSH concentration and release in response to a decrease

7   in iodine supply to the thyroid. During the first 4 weeks of the study, the volunteers were given

8   200 μg/day iodine. After iodine supplementation was discontinued, the volunteers were

9   administered 900 mg/day of potassium perchlorate orally for 4 weeks to induce a state of iodine

10   depletion. At the end of the 4-week perchlorate treatment, levels of thyroid hormones were

11   measured. Although perchlorate treatment had no effect on thyroid volume or levels of

12   triiodothyronine (T3) and thyroxine (T4), intrathyroidal iodide concentration and serum levels of

13   TSH were decreased significantly, and serum levels of thyroglobulin were nearly doubled. The

14   authors speculate that the decrease of TSH, which is the opposite of the expected response, may

15   be an early adaptive mechanism to the iodine deficiency induced by perchlorate. They suggest

16   that, early in iodide deficiency, the thyroid becomes more sensitive to TSH creating a feedback

17   mechanism that decreases TSH levels. Only as iodide deficiency becomes more prolonged do

18   TSH levels increase. This study defined a LOAEL of 13 mg/kg-day for thyroid effects. In a

19   follow-up study, Brabant (1994) repeated the earlier studies with perchlorate treatment lasting

20   longer than 4 weeks. As a result of the longer treatment, thyroid volumes increased in all

21   subjects although TSH levels did not increase.

22   Lawrence et al. (2000) performed a 14-day clinical study with nine euthyroid volunteers

23   (ages 22 to 30 years). Each subject was enrolled after a normal complete physical exam that

24   included a thyroid exam. Blood was obtained for baseline measurement of thyroid function tests,

25   TPO antibodies, CBC, and routine blood chemistries. A spot urine was obtained for routine

26   urinalysis. All baseline tests were normal.

27   Ten mg of perchlorate as potassium perchlorate was dissolved in 1-L bottles of spring

28   water. Each subject was instructed to consume the 1-L bottle intermittently during waking hours.

29   Assuming a body weight of 70 kg, this dosage is equivalent to 0.14 mg/kg-day. Blood specimens

30   were drawn between 8:00 and 9:00 a.m and 24-hour urine samples were obtained on days 7 and

31   14 during exposure and then again after another 14 days after perchlorate was discontinued.

1 Thyroid function was assessed by assays for TSH, free thyroxine index (FTI), total T3, (TT3) and

2 T4. Blood chemistries and CBC were also measured. Baseline thyroid radioactive iodine uptake

3 (RAIU) was measured using ^{123}I at 4, 8, and 24 hours after the ingestion of 150 μCi ^{123}I.

4   As reported by the authors, statistical analysis for the thyroid RAIU was carried out by

5 analysis of variance (ANOVA) with post hoc pairwise comparisons using Tukey's method. The

6 outcome measure variable was log-transformed to achieve greater homoscedasticity and a more

7 Gaussian distribution. Serial analyses were done: a three-factor ANOVA with factors as patient,

8 treatment, and time and a set of two-factor ANOVAs, one for each of the three times. The

9 analogous mixed-model ANOVAs were also run with subject as a random effect to confirm that

10 repeated measures among the subjects did not affect the results. Statistical analyses of the

11 thyroid function tests and urine and serum perchlorate and iodine values were carried out by

12 ANOVA and Student Newman Keuls (SNK).

13   Urine and serum perchlorate levels at baseline and during and after ingestion of the daily

14 10 mg perchlorate dose are presented in Table 4-2. Perchlorate levels returned to baseline after

15 the two week recovery period. There was also no significant changes in urinary iodine excretion

16 during, or 2 weeks after stopping the perchlorate administration as shown in Table 4-3. The

17 authors note that the iodide ingestion of the volunteers was not controlled in the diet and were

18 variable. It may also be worthwhile to note that the urinary iodine values are relatively high (see

19 Chapter 3), indicating a potential protective status in these subjects for the inhibition of the NIS

20 by perchlorate.

21

22

TABLE 4-2. URINE AND SERUM PERCHLORATE (ClO$_4^-$) VALUES BEFORE, DURING, AND AFTER THE INGESTION OF 10 mg OF ClO$_4^-$ DAILY FOR 14 DAYS (Lawrence et al., 2000)

Time	Urine Perchlorate[a] (mg/24 hr)	Serum Perchlorate[a] (μg/mL)
Baseline	< 0.5	0
7 Days ClO$_4^-$	7.7 ± 0.8[a]	0.61 ± 0.02
14 Days ClO$_4^-$	7.5 ± 1.0	0.59 ± 0.02
14 Days After ClO$_4^-$	< 0.5	0

[a]Mean ± SE.

TABLE 4-3. URINE AND SERUM IODINE VALUES BEFORE, DURING, AND AFTER THE INGESTION OF 10 mg OF ClO_4^- DAILY FOR 14 DAYS (Lawrence et al., 2000)

Time	Urine Iodine[a] (μg/24 hr)	Serum Iodine[a] (μg/dL)
Baseline	254 ± 69	6.5 ± 0.42[a]
7 Days ClO_4^-	233 ± 49	6.2 ± 0.34
14 Days ClO_4^-	385 ± 123	6.4 ± 0.37
14 Days After ClO_4^-	208 ± 42	6.3 ± 0.57

[a]Mean $\pm$ SE.

1   A highly significant decrease in the ^{123}I thyroid RAIU with respect to baseline

2   measurements at all three time points was noted (Table 4-4), 34%, 39%, and 41% at 4, 8, and

3   24 hours. The decrease averaged over all three time points was 38%. Two weeks after

4   perchlorate was discontinued, the thyroid RAIU values were significantly higher at all three time

5   points (average increase of 25%), indicating a rebound that may represent upregulation of the

6   NIS. The time course of the iodine inhibition could not be calculated since the subjects drank the

7   dose ad libitum over the day and there was evidence that the full 10 mg/day dose was not

8   achieved for at least some subjects because the average daily urinary excretion of perchlorate was

9   7.6 for the 2-week course of perchlorate administrations. There was a corresponding increase in

10   urinary iodide excretion during dosing followed by a drop below baseline during rebound. T3

11   levels were observed to rise throughout the 28-day trial (trend not tested).

12   In a subsequent study reported as a letter to the editor by these same investigators,

13   Lawrence et al. (2001) used nine healthy male volunteers and a dose of 3 mg/day (.04 mg/kg-day

14   assuming 70 kg body weight) and again observed decreased RAIU. The mean 8-hour decrease

15   from baseline was reported to be at 10% and at 24-hours to be 10.3%. Neither were significant

16   based on Tukey paired t-test (data not shown). The RAIU after stopping the perchlorate

17   ingestion for 14 days rebounded as in the first study and was reported to be an increase of 22% at

18   8 hours and 18% at 24 hours (p < 0.02). It is worthwhile to note when evaluating these results

19   that these data (Lawrence et al., 2000; 2001) were evaluated for use in the physiologially-based

20   pharmacokinetic (PBPK) models described in Chapter 6, but the data were excluded due to the

TABLE 4-4. THYROID ^{123}I UPTAKES BEFORE, DURING, AND AFTER THE INGESTION OF 10 mg ClO$_4^-$ DAILY FOR 14 DAYS (Lawrence et al., 2000)

Time	Thyroid ^{123}I Uptake[a] (% Dose)		
	Baseline	14 days on ClO$_4^-$	14 days after ClO$_4^-$
4 Hours	12.5 ± 1.3	8.2 ± 0.7[b]	16.6 ± 2.4[c]
8 Hours	17.3 ± 1.9	10.6 ± 1.0[b]	21.9 ± 2.8[c]
24 Hours	23.6 ± 2.6	14.0 ± 1.6[b]	27.1 ± 3.3[d]

[a]mean ± S.E.
[b]$p < 0.01$ vs. baseline and after ClO$_4^-$.
[c]$p < 0.01$ vs. baseline.
[d]$p < 0.05$ vs. baseline.

1   lack of availability of all records to the QA/QC process and unresolved issues regarding sample

2   sequences. Variability of serum and urine perchlorate results, potentially due to the unstructured

3   drinking water regimen (Merrill, 2001a,b) was noted. Serum levels from the 0.04 mg/kg-day

4   dose group ranged from non-detect to 495 mg/L on days when the subjects were supposed to

5   have consumed perchlorate. Given this variability and the unknown consequence of a 10%

6   change in thyroid RAIU of a small sample of healthy euthyroid individuals to potentially

7   hypothyroid or hypothyroxinemic pregnant women, it would be difficult to designate this effect

8   as a No-Observed-Adverse-Effect-Level (NOAEL) with any confidence.

9   Greer et al. (2000) described a third study of RAIU in healthy euthyroid subjects in an

10   abstract. Perchlorate was dissolved in 400 ml of drinking water at one of three doses to twenty-

11   four euthyroid volunteers (4 males and 4 non-pregnant females per dose; 18 to 57 years old).

12   The subjects were instructed to drink 100 ml at 4 set times throughout the day for 14 days.

13   Measurement of 8- and 24-hour RAIU was performed prior to perchlorate ingestion (baseline),

14   on exposures days 2 and 14, and on post-exposure Day 15. Expressed as a percentage of baseline

15   (mean ± S.E.), the abstract reports 24-hour RAIU values for the 0.02, 0.1 and 0.5 mg/kg-day dose

16   groups as: 83 ± 5.6, 59 ± 3.5 and 31 ± 2.6 on exposure day 2; 85 ± 5.6, 57 ± 4.7, and 34 ± 4.5

17   on exposure day 14; and 111 ± 5.1, 96 ± 12, and 108 ± 12 on post-exposure Day 15. These

18   correspond to RAIU inhibition values expressed as % of baseline (where "−" indicates inhibition

19   relative to baseline) for the 0.02, 0.1 and 0.5 mg/kg-day dose groups of −17, −41, and −69 on

1 exposure Day 2; −15, −43, and −66 on exposure Day 14; and +11, −4, and +8 on post-exposure

2 Day 15. The authors report no difference between males and females and that a linear log-dose

3 relationship was observed with the regression slopes indistinguishable between the 8- and

4 24-hour measurements (data not shown).

5   In other unpublished data provided in Merrill (2001a; Attachment #7) these same

6 investigators tested seven euthyroid subjects (six non-pregnant females and one male) at a dose

7 of 0.007 mg/kg-day. Expressed as a percent of baseline, the average 8- and 24-hour RAIU

8 inhibition values measured on exposure Day 14 were −6.2 and −1.8%. The inhibition values

9 ranged from −38.6% to +27.9% of baseline at the 8-hour time point and −26.7 to +39% of

10 baseline at the 24-hour time point. The range for the post-exposure Day 14 RAIU inhibition

11 values was −19.3 to +45% of baseline. No measurements were made on Day 2 when the RAIU

12 inhibition would have been greater. There was no RAIU inhibition measured on post-exposure

13 Day 15. In the Greer et al. (2000) abstract, the authors estimate the no effect level at

14 0.007 mg/kg-day.

15   In order to evaluate whether the 0.007 mg/kg-day dose had a sufficient sample size to

16 detect a difference of the observed magnitude as in the other doses tested, the EPA calculated the

17 power of the usual t-test for the 14-day exposure data. A log transform of the ratio of the

18 individual values at Day 14 to their baseline values was based on the non-central t distribution.

19 The power at the 0.007 mg/kg-day dose was low (0.1) compared to the other doses: 0.95, 0.998,

20 and 0.999 at 0.02, 0.1, and 0.5 mg/kg-day.

21   The EPA has also been made aware of another human clinical study being performed at

22 Loma Linda and funded by Lockheed Martin (Beck, 2001). The study is not yet completed

23 because the objective sample size for each dose group has not yet been attained. Human

24 euthyroid volunteers (male and non-pregnant females) have been dosed with perchlorate in gel

25 caps at 0.007, 0.014, and 09.04 mg/kg-day. Measurements were made at baseline, 3-months,

26 6-months, and after recovery from exposure for RAIU, T3, T4, and TSH levels. These dosages

27 are the same as already tested so the added value to the human database, especially with respect

28 to the now prominent concern for neurodevelopmental effects secondary to hypothyroxinemia or

29 even transient decrements in T4, is not readily apparent. The additional data may potentially

30 reduce the variability and low power due to the small sample sizes of the previous studies if

31 sufficiently comparable in design.

4.2.2 Studies in Patients with Graves' Disease

Potassium perchlorate had been used to treat Graves' disease in humans; consequently, most of the prior data on perchlorate effects on humans are in patients with this disease. Graves' disease is an autoimmune disorder which causes patients to carry immunoglobulins in their blood that bind to TSH receptors on thyroid cells and act like TSH to stimulate DNA synthesis and cell divisions, leading to a hyperthyroid state. Symptoms of the disease include increased synthesis and secretion of iodide-containing hormones into the blood by the thyroid gland, thyroid gland enlargement, increased basal metabolism, and weight loss. Perchlorate inhibits the excessive synthesis and secretion of thyroid hormones by inhibiting the uptake of iodide into the thyroid and causes an efflux (discharge) of accumulated iodide in the gland.

Stanbury and Wyngaarden (1952) evaluated therapeutic perchlorate use in patients (n = 8, although reporting of exact numbers for various aspects [e.g., different dose levels] of the study is sketchy) with Graves' disease and found that perchlorate caused the discharge of iodide accumulated in the thyroid and blocked the uptake of iodide into the thyroid. Within 30 min of administration, a single dose of 100 mg potassium perchlorate caused the nearly complete release ($\approx$80%) of ^{131}I from the thyroids of Graves' disease patients previously treated with tracer amounts of ^{131}I and 1-methyl-2-mercaptoimidazole (MMIA). MMIA was given to cause accumulation of ^{131}I in the thyroid because MMIA prevents the oxidation of iodide ion to iodine and its attachment to tyrosyl groups (see Chapter 3). A single dose of 10 mg perchlorate appeared to cause a ~50% release of accumulated iodine. The authors reported that perchlorate doses as low as 3 mg caused detectable, but incomplete, release of iodide from the thyroid (although quantitative data for doses less than 10 mg were not presented). In addition, Stanbury and Wyngaarden (1952) reported that the uptake of tracer levels of ^{131}I into the thyroid glands of two patients with Graves' disease was markedly inhibited for as long as 6 hr when 100 mg of potassium perchlorate was given orally 1 h prior to administration of the tracer. Beyond 6 h, uptake of ^{131}I recommenced. Inhibition of iodide uptake also occurred in three patients without MMIA treatment. The authors stated that no toxic effects were encountered in any patients who were given, in more than three doses, a total not exceeding 600 mg potassium perchlorate. This

1   study was used to identify a LOAEL of 1.4 mg/kg-day[1] for complete release of iodine from the

2   thyroid for the RfD reviewed in March 1997 (Toxicology Excellence for Risk Assessment,

3   1997). Because it was not clear what degree of iodide efflux constitutes an adverse effect, a

4   NOAEL was not designated for this study. An expert peer review panel later determined this

5   study was inadequate for RfD derivation (Toxicology Excellence for Risk Assessment, 1998b).

6   Godley and Standbury (1954) report using potassium perchlorate to treat 24 patients with

7   Graves' disease. Patients were treated with 600 to 1,200 mg/day (typically 200 mg every 8 h)

8   for at least 11 weeks with a few patients treated as long as 45 to 52 weeks. A decrease in iodide

9   uptake was observed. Five patients became euthyroid after continuous administration for

10   28 weeks. Two patients developed gastrointestinal problems that were assumed to result from

11   perchlorate treatment. In one of these patients, these effects occurred at 600 mg/day, but the dose

12   that the other patient received is not specified. Other side effects of antithyroid agents, such

13   hematological changes, liver damage, and skin rash, were not observed. This study suggested a

14   LOAEL of 9 mg/kg-day in humans for short-term exposures.

15   Crooks and Wayne (1960) observed one case of skin rash and three cases of nausea (12%)

16   among 35 patients treated with 600 mg/day (9 mg/kg-day) and 165 patients given 1,000 mg/day

17   (14 mg/kg-day). All patients had diffuse goiters and exophthalmos, classic signs of Graves'

18   disease. In another group of 10 patients given 1,500 mg/day (21 mg/kg-day) and 40 patients

19   given 2,000 mg/day (29 mg/kg-day), five cases of skin rash, two cases of nausea, and one case of

20   agranulocytosis occurred (16%). Leukocyte counts returned to normal in the patient with the

21   agranulocytosis when perchlorate treatment was stopped. The length of treatment was unclear

22   but generally appears to have been less than 8 weeks although it appears that one patient was

23   monitored for 22 weeks. The authors report that the "time to cure" Graves' disease using

24   perchlorate is approximately 9 weeks. The authors also report that 1 of 12 infants born of

25   mothers given 600 to 1,000 mg/day was born with a very slightly enlarged thyroid that returned

26   to normal size in 6 weeks; no other abnormalities were noted. This study suggested a LOAEL

27   between 9 and 14 mg/kg-day.

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[1]Unless otherwise indicated, for human studies in which the actual body weight of the subjects was not
reported, the dose in milligrams per kilogram per day was calculated assuming a body weight of 70 kg. Thus, a dose
of 100 mg/day ÷ 70 kg is 1.4 mg/kg-day.

1   Morgans and Trotter (1960) reported that 3% of 180 patients treated with 400 to

2   1,000 mg/day (6 to 14 mg/kg-day) potassium perchlorate and 18% of 67 patients treated with

3   1,200 to 2,000 mg/day (17 to 29 mg/kg-day) displayed a variety of adverse reactions that

4   included skin rash, sore throat, gastrointestinal irritation, and lymphadenopathy. Reactions

5   occurred within 2 to 3 weeks of drug administration. This study suggested a LOAEL between

6   6 and 14 mg/kg-day.

7   Connell (1981) reported a case study of a single 72-year-old female Graves' disease patient

8   who was treated with 200 mg/day (3 mg/kg-day) potassium perchlorate for 22 years without any

9   indication of adverse side effects. Thyrotoxicosis recurred 4 weeks after stopping potassium

10   perchlorate administration, suggesting that this dose level provided sufficient clinical control of

11   the hyperthyroidism. The study also suggested that the adverse reactions seen at higher doses

12   may not occur at lower doses, even after long-term treatment.

13

14   **4.2.2.1 Hematological Effects**

15   Between 1961 and 1966, the occurrence of severe hematological side effects in patients

16   receiving long-term potassium perchlorate treatment for Graves' disease led to a decreased use of

17   potassium perchlorate as a therapeutic agent. Several authors (Hobson, 1961; Johnson and

18   Moore, 1961; Fawcett and Clarke, 1961; Krevans et al., 1962; Gjemdal, 1963) report case studies

19   in which a single patient suffered fatal aplastic anemia after treatment doses ranging from 6 to

20   14 mg/kg-day. The duration of treatment ranged from 3 mo (Johnson and Moore, 1961) to 8 mo

21   (Hobson, 1961). In all cases, patients were started at the high end of the treatment range for a

22   period of time and then were reduced to the lower end of the treatment range after the appearance

23   of side effects. In two cases (Hobson, 1961; Gjemdal, 1963), patients had co-exposures to other

24   drugs. Other case reports are available that report nonfatal agranulocytosis in patients treated

25   with 14mg/kg-day for 12 days (Southwell and Randall, 1960) or 3 mo (Sunar, 1963). Barzilai

26   and Sheinfeld (1966) report that 11% of 76 patients developed leukopenia or other unspecified

27   side effects after treatment with 1,000 mg/day (14 mg/kg-day) for a little as 2 mo. Within this

28   group, there was one case of fatal aplastic anemia and one case of fatal agranulocytosis.

29   These studies suggest that doses in the range of 6 to 14 mg/kg-day may represent a frank

30   effect level in patients with Graves' disease although there were questions as to whether these

31   effects were caused by the disease itself, whether there was some contamination, or whether the

effects occurred only at high doses. A review by Wenzel and Lente (1984) concluded that the "severe adverse reactions, such as agranulocytosis, were likely to occur only when large doses of more than 1,000 mg potassium perchlorate were administered." There is no information to suggest that humans without Graves' disease would have a similar reaction to perchlorate.

Antithyroid drugs appear to exert their effects on the hematopoietic system through an immune mechanism. Wing and Fantus (1987) reviewed the adverse effects of two antithyroid drugs, propylthiouracil and methimazole, and concluded that most reactions were related to the immunologic effects of these drugs. They noted that skin rash and granulocytopenia were among the most commonly reported adverse effects of these drugs. Less commonly reported effects include aplastic anemia, leukopenia, and antibodies to insulin and glucagon. In fact, Wing and Fantus (1987) recommend that patients be instructed to report skin rash immediately, as this may be an early sign of adverse immune reaction caused by the antithyroid drugs. Although these authors did not include perchlorate in their investigation, the similarity of the effects seen after perchlorate treatment—including rash, leukopenia, agranulocytosis, and aplastic anemia— suggest that perchlorate also may act in a similar fashion to induce an immune effect.

There is a tight functional connectivity between the immune and endocrine systems which is mediated, at least in part, by shared receptors and mediators among the systems (Kammuller, 1995). Thus, although the mechanism of perchlorate action on the hematopoietic system is not known, it is likely to be an immune reaction. Although it is possible that perchlorate may cause hematological effects in healthy humans, it appears that Graves' disease patients are likely to be more sensitive to this type of immune-induced adverse effect than are healthy people. The increased sensitivity to immunologic function in Graves' disease patients arises because of the underlying abnormal immunologic function in Graves' disease. Immunoreactivity to antithyroid drugs is another expression of the compromised immune system in these patients (Wall et al., 1984; Wing and Fantus, 1987).

4.3 SUMMARY OF CONCLUSIONS REGARDING HUMAN HEALTH EFFECTS STUDIES

The recent human studies support the established effect of perchlorate at the NIS. Using these data as the basis for quantitative dose-response assessment is more difficult. Of the five

population studies investigating the effects of perchlorate exposures on TSH levels in newborns (Lamm et al., 1999; Li et al. 2000b; Crump et al., 2000; Brechner et al., 2000; Schwartz, 2001), the Brechner et al. (2000) study had a somewhat better exposure classification owing to a more narrow, but still ecological, geographical focus (two small cities) and Schwartz had a relatively detailed exposure classification down to the level of zip codes. Only these two studies had positive findings in newborns. The restriction of birth weight in Li et al. (2000b) could have reduced study sensitivity if thyroid endpoints in non-normal birth weights are especially effected by perchlorate. The strong dependence of thyroid endpoints on birth weight observed in several studies raises the possibility that birth weight itself could be an intervening variable in perchlorate effects. That is, perchlorate exposure may affect birth weight. This would be a testable hypothesis in several of the studies. If birth weight were an intervening variable, birth weight restriction in the Li et al. (2000a,b) studies or controlling for birth weight as a confounder in the Li et al. (2000a,b), Brechner et al. (2000) and Schwartz (20001) studies may have resulted in an underestimation of perchlorate exposure effects.

In the one study that reported age-specific perchlorate exposure effects on TSH (Brechner et al., 2000), the largest effect was in the first 24 hours after birth. This observed exposure-age interaction was not statistically evaluated. The study with the strongest findings (Schwartz, 2001) actually focused only on the first 2 days after birth. Therefore, excluding day-one screened births as in the Li et al. (2000b) study may severely reduce or eliminate the ability to detect a perchlorate effect.

The well-known TSH surge at birth is thought to represent a response to temperature change (Schwartz, 2001). This suggests that ambient temperatures – prenatal and perinatal – might be important determinants of thyroid endpoints. The strong period/seasonal effect observed in the Li et al. (2000b) study supports this temperature conjecture and the unexpected trends across Chilean cities in the Crump et al. (2000) and variations across U.S. counties in the Lamm et al. (1999) and Schwartz (2001) investigations could also be related to temperature.

It should also be noted that all of the studies in this review examined endpoints that may be insensitive to the consequences of altered thyroid function. No detailed models of thyroid dynamic response were postulated with subsequent analysis of relevant endpoints that would reliably detect the specific perchlorate- or environmentally-induced defects. Nonetheless, one study examining neonatal thyroid status in the first five days found a perchlorate effect that was

1   greatest in the first 24 hours and that rapidly declined over the next two days, suggesting

2   alteration of thyroid response to the birth event. The issue of iodine depletion in exposed

3   populations was not directly evaluated although experimental evidence of short-term depletion in

4   adults at high doses was observed.

5   All of the observational field studies utilized "ecological" exposure rather than individual-

6   specific dose measurements; the relative specificity of the dose metric varied widely from

7   "exposed/not exposed", to an average concentration in drinking water for a given zip code. The

8   occupational studies used air sampling to estimate homogeneous exposure groups. Nevertheless,

9   there was evidence of perchlorate effects on neonatal thyroid status, with the studies by Brechner

10   et al. (2000) and Schwartz (2001) contributing the most compelling observations, and iodine

11   depletion was observed experimentally. The presence of exposure misclassification and

12   potentially serious confounding in many of the studies makes interpretation difficult and allows

13   for the possibility of missed effects even at the level of current thyroid function (e.g., steady state

14   levels of TSH or T4). The full implications of these findings are unclear; however, they should

15   be taken seriously, especially in populations already at risk for thyroid deficiency. These

16   considerations are summarized in Table 4-5.

17   The present review differs from a recent summary co-authored by two major participants in

18   industry-funded perchlorate research (Soldin et al., 2001). That review argues that there is now

19   sufficient evidence to recommend safe levels for regulatory purposes. The authors see no

20   immediate need for refinement of the physiological issues underlying the existing epidemiologic

21   study designs or for new initiatives in evaluating such issues in human populations. Potentially

22   important aspects of the mode-of-action for perchlorate not well addressed in the available

23   human studies include: (1) short-term effects of variable exposure during pregnancy, for

24   example, on critical neurodevelopmental effects; (2) the effects of iodine depletion on the T4 or

25   TSH surge response at birth, i.e., whether the effect of perchlorate on fetal thyroid status depends

26   additionally on prior cumulative exposure; (3) the equilibration of this regulated system under

27   chronic exposure and the masking of potential deficiency states such that steady-state T4 or TSH

28   levels appear normal despite substantial impact on production and function; and (4) the special

29   situation of populations or individuals with inadequate iodine intake where thyroid

30   responsiveness may be compromised.

1   The recent clinical data (Lawrence et al., 2000; 2001; Greer et al., 2000) may be more

2   useful in helping to characterize the potential effects on thyroid function if the mode of action

3   framework is superimposed on the interpretation of the data (i.e., that prevention of significant

4   iodide inhibition would preclude adverse neurodevelopmental and neoplastic sequelae).

5   However, given the current controversy in evaluating thyroid status, particularly in pregnant

6   women, it is difficult to ascertain the degree of iodide inhibition to designate as adverse. Further,

7   there is considerable uncertainty associated with using small sample sizes of euthyroid

8   individuals as the basis of such a determination, so that the use of a factor to account for this in a

9   risk derivation would be warranted, particularly when the variability as noted is considered and

10   the range of inhibition of iodide uptake at levels suggested to be "No-Observed-Adverse-Effect-

11   Levels" include values as great as 38.6% below baseline. A discussion considering these human

12   clinical data in comparison to the laboratory animal toxicological study results can be found in

13   Section 7.1.5.1.

TABLE 4-5. SUMMARY OF HUMAN POPULATION STUDIES (Park, 2001)

	Publication	Study Population	ClO$_4^-$ Source and Levels	Duration	Outcomes studied	Findings	Problems/Comment
1	Gibbs JP, Ahmad R, Crump KS, et al JOEM 1998; 40:1072-1082. *Evaluation of a population with occupational exposure to airborne ammonium perchlorate for possible acute or chronic effects on thyroid function.*	Kerr-McGee workers in voluntary medical surveillance 1994-98; 170 out of 254 did survey; 130 did single shift evaluation	Airborne exposure to AP in 8 homogenous exposure groups: 0.04-627 μm/m^3 using closed face cassettes	1 day 1-27 yr.	T3U, T4, FTI, TSH, liver, kidney and hematol fcn T4: 7.5 μg/dL TSH: 2.0 μIU/ml	Indication of increase in TSH over work shift: 2.2 -> 2.5. In workforce, T4 declines and TSH increases from low to high exposure but also from low exposure to unexposed; see inconsistent TSH trends using two lab groups; for both thy and liv outc, SDs increased in high dose group: for thy and liv fcn, averages for low vs high AP very different but %iles up to 75th are not. Implies big excursion at high exposure end.	Possibly half of eligibles did not participate in shift study; possibly confounded by shift duration. Did not evaluate ITR. Suggestion of inappropriate unexposed comparison group. In this steady state and cross-sectional population, difficult to assess thyroid regulatory status. SDs suggest heterogeneity of effect. **Indications of chronic effects.**
2	Lamm SH, Braverman LE, Li FX, et al. JOEM 1999; 41:248-260. *Thyroid health status of ammonium perchlorate workers: a Cross-sectional occupational health study.*	American Pacific workers: 37 AP and 21 azide workers: full feasible participation; all from same site with same other work attributes	Airborne exposure in 3 AP groups based on visible dust level; total and respirable AP by individual closed-face samplers 10-11 hrs on subset from each exp group; levels: total dust (mg/day): .01, .34, 6.57, 59.4; resp fraction (mg/day): .02, .09, .60, 8.6	1 day n=58; 6 days n=2	Urine AP, T3, T4, FTI, TSH, THBR, and hematologic fcn T4: 7.0 pg/dL TSH: 2.6 JLrU/InI	18% of total airborne Mb is respirable (range *8-25);* urinary excretion of P shows much higher absorbed dose in unexposed workers than expected from air samples: (mg): .88. 4.0, 10.9, 33.6 (assuming 8 hr halflife). Thy, hematol by current exp group: no association (T3, T4); absorb dose greatly exceeds resp total inhaled dose [F51. See aberrant clearance in 1 of 2 6-day subjects fF2]. Authors conclude no AP health effects.	Some misclassification apparent among exposure groups based on absorbed dose; non-inhalable contribution may constitute important deficit in air sampling results. Steady-state, cross-sectional population difficult to interpret. Thy, hematol results based on current, non-cumulative AP exposure are uninterpretable for chronic effects. Possible increasing trend for max(T3) with exposure group.
3	Lawrence JE, Lamm SH, Braverman LE. J Endocrinol Invest 1999; 22:405-407. *The use of perchlorate for the prevention of thyrotoxicosis in patients given iodine rich contrast agents.*	Radiocontrast patient series	Therapeutic high oral doses (1000 mg) in day prior to contrast agent	1 day	Misc. thyroid parameters —	Recommend in high risk patients (low iodide areas and elderly) a combination of perchlorate and contrast agent.	Not relevant to and uninformative on chronic exposure effects in adults and acute effects in infants.

TABLE 4-5 (cont'd). SUMMARY OF HUMAN POPULATION STUDIES (Park, 2001)

	Publication	Study Population	ClO_4^- Source and Levels	Duration	Outcomes studied	Findings	Problems/Comment
4	Li FX, Squartsoff L, Lamm SH. JOEM 2001; 43:630-634, *Prevalence of thyroid diseases in Nevada counties with respect to perchlorate in drinking water.*	Medicaid population at risk for thyroid disease in Nevada in 1997-98.	Perchlorate in drinking water in one county (P= 8.9-11.6 μg/L) versus all others	Lifetime	ICD 240-246; ICD 193: thy cancer	Exposed county (Clark) with Las Vegas compared to another county with a city (Reno/Washoe) as well as with all other counties. No significant excesses found for exposed county for the 8 outcomes studied. Actually, the comparison counties (one with a city, and all others) for all important outcomes differed more between them than with the exposed county. For the 6 more prevalent outcomes (n=3069) the exposed county had higher rates than the unexposed (Washoe) county.	Based on period-prevalence rates. Two outcomes with small numbers are not informative: congenital hypothyroidism (n=22) and thyroid cancer (n=44). **The difference in the comparison counties suggests that uncontrolled confounders or uncertain estimates are affecting this analysis and that the study is uninterpretable for all but large effects.** Confounders might include age, gender, body mass, diet, iodine intake, ethnicity, occupational exposures.
5	Crump C, Michaud P, Tellez R et al. and Crump KS, Gibbs JP. JOEM 2000; 42:603-612. *Does perchlorate in drinking water affect thyroid function in newborns or school-age children?*	School children from 1 or 2 schools in three cities in Chile (n=53,49,60 in 0, low and high P cities); all newborns 2/96-1/99 in same cities (n=8888,468,428)	Geological Na-P in drinking water (0, 5.5, 111.6 μg/L)	Recent and lifetime for 6-8 yr-olds; gestation	T3, T4, free T4, FTI, TSH, hematol, liver, kidney, prev:goiter, prev:family H_x thy disease T4:10.0 μg/dL TSH: 3.0 μIU/mL	Did comparisons across cities. Urinary I/creatine low in city-2 lifetime residents: (1,092, 862, 963); goiter high in city-2 recent residents: (17.7, 26.5, 23.3%) and high in city-3 lifetime residents: (22.2, 19.5, 26.0 based on 8, 8, 13 cases); family H_x of the disease high in city-3: OR=4.9 (11.1, 9.8, 30.0); highly significant increase in T4 with increased P (1.25, 1.34, 1.50). Highly significant decrease in log (TSH+1) in newborns in city-3–high P (.91, .91, .66) [T9], which is in the unexpected direction. There was a diverse age-at-screen distribution across cities.	Dietary, ethnic, birthwt, SES confounders of thy fcn uncontrolled; observe trends in unexpected directions; suggesting confounding. Unknown if some Chileans boil drinking water. **Significant paradoxical effects indicate uncontrolled confounding and inappropriate thy fcn model in relation to P in this population.** Possible role of ambient temperatures.
6	Lawrence JE, Lamm SH, Pino S, Richman K, Braverman LE. Thyroid 2000; 10:659-663. *The effect of short-term low-dose perchlorate on various aspects of thyroid function.*	9 healthy, male volunteers K-perchlorate – 10mg/day	Potassium perchlorate 10 mg/day	14 days	T3, T4, FTI, TSH, THBR, RAIU, liver, hematology T4: 7.0 μg/dL TSH: 1.0 μIU/mL	Assumed identical P doses. Upward trend for T3 at BL, 7-, 14-, and 28-days (136, 140, 151, 157; trend not tested). See depressed I-uptake at 14 days (40%) with rebound at 28 days; non-24 hour urinary- and serum-I was unchanged throughout. Authors conclude: no thyroid impact because of large I-storage.	Hematol, liver test results clinically "normal" but no data presented. **Inappropriate assessment: clinical rather than epidemiological**. T3 effect not addressed; dietary I not controlled or reported. **Suggests long term iodine depletion.**

TABLE 4-5 (cont'd). SUMMARY OF HUMAN POPULATION STUDIES (Park, 2001)

	Publication	Study Population	ClO₄⁻ Source and Levels	Duration	Outcomes studied	Findings	Problems/Comment
7	Lawrence JE, Lamm S, Braverman LE. Thyroid 2001. 11:295 (letter) *Low dose perchlorate (3 mg daily) and thyroid function.*	8 healthy volunteers	Potassium perchlorate 3 mg/day	14 days	T3, T4, FTI, TSH, THBR, RAIU, liver, hematol	No signif changes (data not presented) except for depressed I-uptake at 14 days (10%) with significant rebound (22%) at 28 days;	**Implies some I depletion over 2 weeks at 3 mg/day** (seen by other investigators at 1.4 mg/day).
8	Lamm SH, Doemland M. JOEM 1999; 41:409-411. *Has perchlorate in drinking water increased the rate of congenital hypothyroidism?*	Newborns in CA and NV in 1996-97 in 7 counties	Perchlorate in drinking water: 4-16 μg/L	Gestation	Congenital hypothyroid-ism based on neonatal screen (expected= $35/10^5$) —	Compared counties. Hispanic-adjusted prevalence ratios by county: 0.6 (n=8) to 1.1 (n=136); none statistically significant.	No county-specific levels of P; no individual consumption. Should have used other CA and NV counties for expected rates. **Identification of cases is limited by screening procedure that dues not consider age at screen, ethnicity and birthweight.** Unable to address transient developmental sequelae.
9	Li Z, Li FX, Byrd D, et al. and Lamm. JOEM 2000; 42:200-205. *Neonatal thyroxine level and perchlorate in drinking water.*	Newborns in Reno (n=5,882)and Las Vegas (n=17,308) NV 4/98 – 6/99 with birthwt 2.5-4.5kg and age at screen < 5 days and non ICU	Perchlorate in drinking water of Las Vegas: 0 up to 15 μg/L, measured monthly	Gestation	T4 T4:17.0 μg/dL	Compared cities. Significant period effect (seasonal) (ΔT4=.60) when adj for birthweight (.85/kg), age at screen (day 1,2,3 vs. 4: −1.275, .408,. 758) and gender (.727). No city * period interaction implies no P effect. Age * exposure interaction not investigated. Did regressions on monthly means (T4, cum.P); also, used 10 percentile T4 as an outcome–no effect. See jump in T4 at newborn return visits in days 2-4.	**These T4 levels are much higher than in other neonate studies (7-10).** Birthweight may be intervening variable: P causing reduced birthwt via impaired thy fcn. Loss of power in regressions using monthly means instead of individual obs. Early return visits have selection bias: reason for early return.
10	Li FX, Byrd DM, Deyhle GM et al. and Lamm. Teratology 2000; 62:429-431. *Neonatal thyroid-stimulating hormone level and perchlorate in drinking water.*	Newborns in Reno and Las Vegas NV 12/98 – 10/99 with birthwt 2.5-4.5 kg	Perchlorate in drinking water of Las Vegas: 0 up to 15 μg/L, measured monthly	Gestation	TSH TSH: 10.0 μIU/mL	Compared cities. TSH levels, adjusted for gender and age at screen (2-7 vs. 8-30): no difference for LV vs. Reno.	TSH log transformation for variance stabilization could suppress TSH differences in the high range; inadequate control for age at screen (LV vs Reno), ethnicity and birthwt (2.5-4.5 kg); birthwt may be intervening variable. TSH levels may not be relevant vs T4. **Insensitive to developmental issues and short-term time variability of P exposure.**

TABLE 4-5 (cont'd). SUMMARY OF HUMAN POPULATION STUDIES (Park, 2001)

	Publication	Study Population	ClO₄⁻ Source and Levels	Duration	Outcomes studied	Findings	Problems/Comment
11	Brechner RJ, Parkhurst GD, Humble WO et al. JOEM 2000; 42:777-782. *Ammonium perchlorate contamination of Colorado River drinking water is associated with abnormal thyroid function in newborns in Arizona.*	Newborns 10/94-12/97 in two Arizona cities whose T4 screen was below state-wide daily 10%ile	Perchlorate in drinking water <16 μg/L TSH: 13.4 μIU/mL	Gestation	TSH	Compared cities. TSH higher in newborns from exposed city (median: 19.9 vs 13.4); age at screen distribution very different between two cities: exposed screened sooner. Stratifying on age at screen (0, 1-4, 5+ days) and Hispanicity, see signif increase (p=.017); adj effect not reported.	TSH levels (13-20) higher than reported for other newborns (7-10).] Selection on T4 level is problematic due to strong age dependence of T4 surge at birth thus causing variable percentile discrimination with age (8-40% were screened depending on age). This effect could increase TSH of the exposed city relative to unexposed city but the effect of the bias is difficult to predict. Uncontrolled other confounding e.g., birthwt, gest. age, iodine intake, SES.
12	Schwartz J.Dissertation, UC Berkeley, 2001. *Gestational exposure to perchlorate is associated with measures of decreased thyroid function in a population of California neonates.*	99% of California newborns screened for thy disease in 1996	Perchlorate in drinking water classified in 3 levels and assigned by zip code: 1-2,3-12, 13+ μg/L	Gestation	T4, TSH, presumptive positive; congenital hypothyroidm T4: 160 mg/dL TSH: 7.6 μIU/mL	Compared across four levels of estimated exposure. Has detailed covariates: birthweight, age at screen in hours, ethnicity in 20 groups; birth multiplicity; ANCOVA model with extensive control of most confounders finds highly significant decrease in T4 (mean=166) with P level (0, -9.7, -11.2, -18.2) and large effects for birthweight (-72 for birthweight 1500-2500), age (-50 for hours 7-18) and ethnic groups (-10 to -30); see initial T4 fall followed by surge by 12 hours and stays elevated until 36 hours; initial onset of TSH surge unresolvable in time; stays elevated till 18 hours. Significant P effect on TSH (0, .029, .03, .128) but birthweight effects models (-.09 for <1.5 kg). Model for presumptive positives shows strong age at screen and ethnicity effects; for congenital hypothyroidism, insignificant effect.	[T4 is reported at levels 10,000-fold higher than in other studies.] presumptive positive criterion not clear (all at or below 9 mg/dl plus lowest 5% immediately above 9 mg/dl?). NO P-ITR reported, e.g., P * age (especially on surge amplitude), P * birthweight; possible selection bias in identification of TSH subjects. Age at screen was not included in logistic regression model of congenital hypothyroidemia. **This study presents strong evidence of perchlorate health effects in neonates from drinking water contamination with perchlorate.**

TABLE 4-5 (cont'd). SUMMARY OF HUMAN POPULATION STUDIES (Park, 2001)

	Publication	Study Population	ClO$_4^-$ Source and Levels	Duration	Outcomes studied	Findings	Problems/Comment
13	Soldin OP, Braverman LE, Lamm SH. Therapeutic Drug Monitoring 2001; 23:316-331. *Perchlorate clinical pharmacology and human health: a review.*	Review of animal and human evidence				This review, co-authored by two major participants in industry funded perchlorate research, argues that there is now sufficient evidence to recommend safe levels for regulatory purposes, i.e., at this time there is no need for further refinement of the physiological issues underlying the existing epidemiologic study designs or for new initiatives in evaluating such issues in human populations.	Not considered in this review are issues such as (1) short term effects of variable exposure during pregnancy, (2) the effects of maternal iodine depletion on T4 or TSH surge response at birth, (3) the equilibration of this system under chronic exposure and the masking of potential deficiency states, and (4) the special situation of populations with inadequate iodine intake.

I = iodine; P = perchlorate; AP = ammonium perchlorate; exp = exposure; thy = thyroid; liv = liver; hematol = hematologic; ITR = interaction; outc = outcomes; SD = standard deviation; H$_x$ = history; [Tn] = table in paper; [Fn] = figure in paper.

5. TOXICOLOGICAL EFFECTS IN LABORATORY ANIMAL STUDIES

This chapter provides a review of the relevant laboratory animal toxicity data for quantitative dose-response analysis of the toxic effects of perchlorate exposure. Evidence that both the neoplastic and non-neoplastic effects of perchlorate derive from its anti-thyroid effects at the sodium (Na^+)-iodine (I^-) symporter (NIS) should be appreciated. Studies completed before the initiation of the perchlorate testing strategy described in Chapter 3 are included here, but the major emphasis is on these newer studies given their contemporary design and integrated approach to evaluating perchlorate's mode of action. This introduction provides a brief review of the status of issues after the previous external peer review and a summary of studies recommended and performed since that time. In response to the 1999 external peer review, the EPA committed to a second external peer review to address these recommendations and to evaluate the data from new analyses and studies (Noonan, 1999).

At the external peer review in February 1999, it was noted by the EPA that the thyroid histopathology that had made a significant contribution to the risk assessment had never undergone an independent peer review by a second pathologist in any of the studies. In addition, these studies had been performed at several different laboratories with several different study pathologists using different lesion grading systems. The external peer review panel agreed that these inconsistencies between study reports made it difficult to compare studies and could contribute to variability in the resultant dose-response estimate (Research Triangle Institute, 1999).

In response, the National Center for Environmental Assessment (NCEA) committed to a Pathology Working Group (PWG) process in collaboration with the NIEHS. The purpose of the independent peer review and PWG was to decrease variability in response across the studies by providing a common nomenclature for lesions and a consistent pathology review. Determination of No-Observed-Adverse-Effect-Levels (NOAELs) or designation of adversity was not the objective of this review. NCEA asked Dr. Douglas C. Wolf in the EPA's National Health and Environmental Effects Research Laboratory (NHEERL) to conduct the requisite independent

1   peer review (second pathology review) using one consistent lesion grading system on the

2   materials. Dr. Wolf was chosen because he had not been involved in any of the work performed

3   with ammonium perchlorate and because he had developed a thyroid grading scheme (Hooth

4   et al., 2001) to analyze a similar thyroid response in rodents exposed to sodium chlorate that

5   would be useful to the perchlorate review.

6   After the initial pathology review of 100% of the thyroid slides by Dr. Wolf, Dr. Peter

7   Mann of Experimental Pathology Laboratories, Inc. (EPL), reviewed 100% of the slides for

8   quality assurance/quality control (QA/QC) and consistency. Subsequent to this QA/QC review

9   of the independent peer review, a NIEHS-sponsored PWG of 5 experienced veterinary

10   pathologists was conducted on a subset of the slides. Recommendations of that PWG

11   (Experimental Pathology Laboratories, 2000) were then incorporated into the final report on the

12   independent review of 100% of the slides conducted and reported by Dr. Wolf (Wolf, 2000).

13   Both of these reports were made available almost immediately to the public on the NCEA

14   website. During subsequent analyses it was appreciated that the slides provided for the

15   two-generation study (Argus Research Laboratories, Inc. 1999) were from animals not on test

16   and some of the mean severity scores were miscalculated. These minor changes are provided in

17   Wolf (2001).

18   The thyroid slides that underwent the PWG review included materials from the following

19   studies: Argus Research Laboratories, Inc. (1998a,b,c); Caldwell, et al. (1995); Keil et al.

20   (1998); and Springborn Laboratories, Inc. (1998). It should be noted that the two-generation

21   reproduction study performed by Argus Research Laboratories (1999) was completed at the time

22   of the PWG, and the review included all final thyroid tissue slides despite its listing in the PWG

23   and Wolf (2000; 2001) reports as 1998c. The newest study, that of Argus Research Laboratories,

24   Inc. (2001) described below in Section 5.3.3, was also performed with the new nomenclature and

25   grading system. The study pathologist had been a member of the PWG; therefore, the pathology

26   results can be considered consistent with the results of Wolf (2000, 2001). However, a second

27   independent review of the pathology in that study has not been performed.

28   All analyses performed on thyroid histopathology in this revised risk assessment rely on

29   either the PWG data (Wolf, 2000; 2001) or the new 2001 study (Argus Research Laboratories,

30   Inc., 2001). The revised benchmark dose (BMD) analyses for thyroid colloid depletion,

31   hypertrophy, and hyperplasia diagnosed in the studies reviewed by the PWG are presented in

1   Table 5-1 (Geller, 2001a). Figures 5-1 and 5-2 present these estimates and their distributions

2   graphically in comparison to the previous 1998 assessment values. It is worthwhile to note that

3   while hyperplasia occurs at slightly higher concentrations in the analysis of the overall data array,

4   there is considerable overlap with the distributions of the other two thyroid histopathology

5   indices (colloid depletion and hypertrophy). This overlap is especially evident when evaluating

6   BMD or benchmark dose lower confidence level (BMDL) values within individual studies.

7   The potential for variability due to inconsistent handling of the radioimmunoassay (RIA)

8   kits used for serum thyroid and pituitary hormone levels was also noted at the external peer

9   review (Research Triangle Institute, 1999). In response, the Air Force Research Laboratory

10   (AFRL) conducted a study to compare serum thyroid hormone and TSH data obtained by RAI

11   procedures for three different research laboratories that participated in perchlorate toxicity

12   studies involving hormone analysis (Narayanan, 2000). The purpose was to statistically

13   investigate the reproducibility (i.e., variability across laboratories) and the repeatability (i.e.,

14   variability within a laboratory) of the hormone measurements expressed as counts per minute

15   (CPM). RIA kits from the same batch number and with the same expiration date were used for

16   all the hormone measurements for all the standard and unknown samples. For unknown samples,

17   six rat serum samples plus six samples obtained from different species (dog, guinea pig, rabbit

18   and mouse) were used. Assays were performed using the RIA kits according to the

19   manufacturers' recommended procedures and each laboratories' standard operating procedures.

20   Reproducibility limits (RL) for each sample and for each hormone were determined. The

21   RL was defined as approximately 95% of all pairs of means from the same hormone and same

22   sample; different laboratories should differ in absolute value by less than the RL. The difference

23   in means between any two laboratories is a normally distributed random variable with a mean of

24   zero. The range ± RL is then the middle 95% for this distribution (i.e., 2.5% in each tail). The

25   reproducibility varied for each hormone with T3 showing the best reproducibility and TSH the

26   least. Three replicates ensured a more reproducible sample even when repeatability was not as

27   consistent. The results suggest that the variability in the RIA determination should be considered

28   when determining effect levels.

29   It was also recommended at the external peer review, by the biostatistician Dr. Joseph

30   Haseman, that different approaches to the thyroid and pituitary hormone analyses be explored

31   (Research Triangle Institute, 1999). EPA complied with this request and developed two new

TABLE 5-1. BENCHMARK DOSE (BMD)[a] AND BENCHMARK DOSE LOWER CONFIDENCE LIMIT (BMDL)[a] ESTIMATES CALCULATED FROM THE WOLF (2000, 2001) THYROID HISTOPATHOLOGY DATA (Geller, 2001a)

Study Name, Time Point Wolf (2000; 2001) Table Number	Ammonium perchlorate dose levels test (mg/kg-day)	Colloid Depletion				Hypertrophy				Hyperplasia			
		BMD	BMDL	$\chi^{2\,b}$	Exp[c]	BMD	BMDL	$\chi^{2\,b}$	Exp[c]	BMD	BMDL	$\chi^{2\,b}$	Exp[c]
1. Caldwell Tbls. 1 and 2	0, 1.25, 5, 12.5, 25, 50, 125, 250	13.29	0.72	0.97	4.37		Not done[d]			35.29	0.78	0.20	0.88
2. Subchonic, 14-day Tbls. 3 and 6	0, 0.01, 0.05, 0.2, 1.0, 10.0	2.55	0.28	0.20	0.74	0.75	0.017	0.54	0.78		NOE[e]		
3. Subchronic, 90-day Tbls. 4 and 7	0, 0.01, 0.05, 0.2, 1.0, 10.0	0.13	0.03	0.70	0.50	0.21	0.008	0.74	0.55	8.36	2.09	1.00	7.87
4. Subchronic, 120-day Tbls. 5 and 8	0, 0.05, 1.0, 10.0		NOE				NOE				NOE		
5. Neurobehav., F0 Fem Tbl. 9	0, 0.1, 1, 3, 10		NOE				NOE				NOE		
6. Neurobehav., PND5 Tbls. 10 and 11	0, 0.1, 1, 3, 10	0.45 0.53	0.009 0.33	0.46 0.67[f]	0.94 1.0	0.92 1.27	0.24 0.88	.024 0.26[f]	0.81 1.0	15.18 11.02	1.86 3.62	0.70 0.32[f]	0.36 1.0
7. Neurobehav., adult Tbls. 12 and 13	0, 0.1, 1, 3, 10	0.72	0.029	0.23	0.89	3.48	NC	0.72	0.29		NOE		
8. 2-gen., P1 Tbls. 14 and 15	0, 0.3, 3, 30	1.97	0.11	0.68	3.84		Poor fit[g]			7.89	2.44	0.41	0.72
9. 2-gen., P2 Tbls. 16 and 17	0, 0.3, 3, 30	2.16	0.90	0.06	1.16	0.99	0.15	0.67	0.70	4.62	0.0004	0.14	0.31
10. 2-gen., F1-weanling Tbls. 18 and 19	0, 0.3, 3, 30	2.51	0.80	0.17	1.2	0.21	0.057	0.40	0.79	2.74	0.66	0.85	0.52
11. 2-gen., F2-weanling Tbls. 20 and 21	0, 0.3, 3, 30		Poor fit			1.19	0.32	0.25	0.52		NOE		
BMDL Range: Rat Studies		**0.009 - 0.90**				**0.008 - 0.74**				**0.0004 - 3.62**			
12. Dev tox., rabbit dams Tbl. 22	0, 0.1, 1, 10, 30, 100	0.12	0.008	0.19	0.36		Poor fit			1.53	0.42	0.13	0.61
13. Immunotox. Mice, combined studies Tbl. 23	0, 0.1, 1, 3, 30	26.07	5.15	1.00	7.88	1.62	0.97	0.58	0.84	24.92	4.48	1.00	7.86

[a] Units of mg/kg-day.

[b] χ^2 p-value.

[c] Exponent in Weibull model fit not restrained to ≥ 1.0 unless indicated.

[d] Not done: Because of non-routine staining, cytological characteristics were not adequate to make determination of hypertrophy on these samples (Wolf, personal communication).

[e] No observed effect (NOE): Either no incidence of endpoint noted in animals tested or no notable difference between dosed and controls.

[f] Exponent in Weibull model fit restrained to ≥ 1.

[g] Poor fit: $p < 0.05$ for χ^2 test.
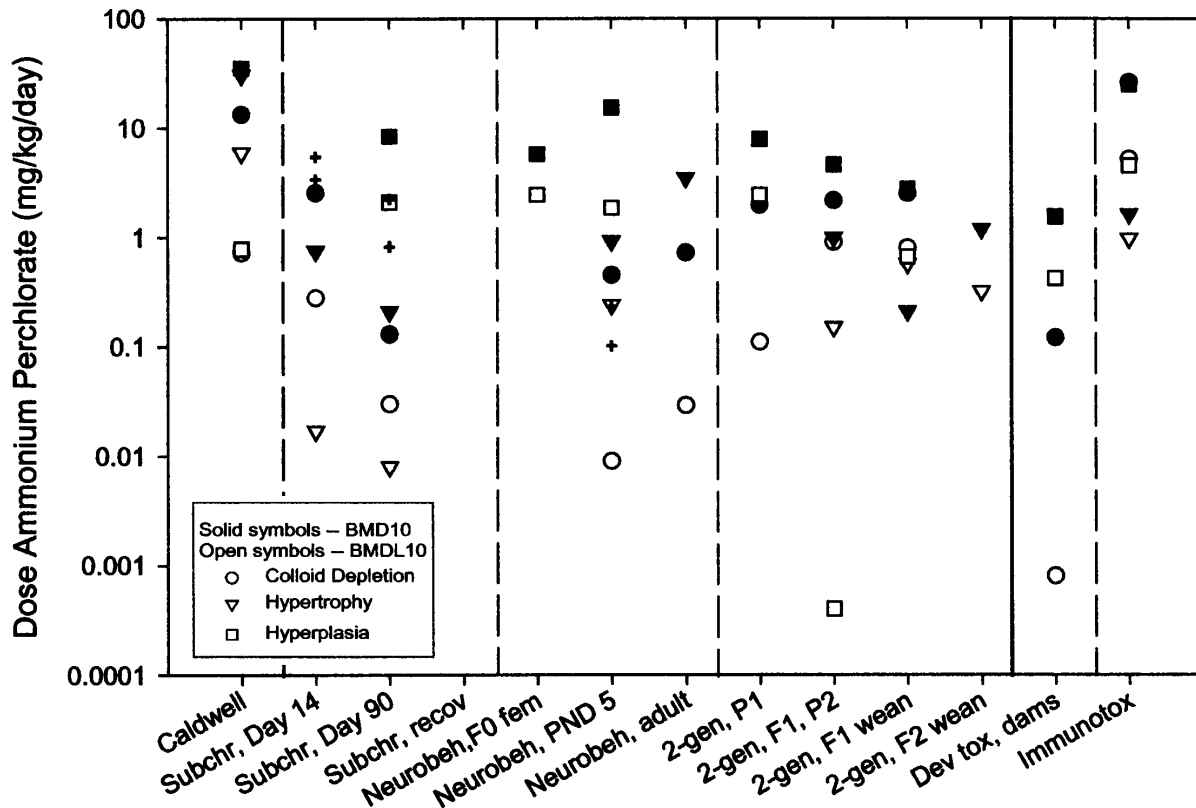
Figure 5-1. Benchmark dose (BMD) and benchmark dose lower limit (BMDL) estimates recalculated for thyroid histopathology based on 2000 Pathology Working Group review (Wolf, 2000; 2001). Data on incidence of colloid depletion, thyroid hypertrophy and thyroid hyperplasia were submitted to the EPA for the perchlorate risk characterization. Values used are presented in Table 5-1 (Geller, 2001a). Greater value represents the BMD and lesser value represents the BMDL. The + denotes BMD and BMDL from previous EPA risk characterization (U.S. Environmental Protection Agency, 1998d; Geller, 1998a). Values to the left of the vertical solid line are from the rat studies. Values to the right are from the developmental study in rabbits (Argus Research Laboratories, 1998c) and the mouse immunotoxicity studies (Keil et al., 1998). Study denoted by "Caldwell" refers to Caldwell et al. (1995); "Subchronic" to Springborn Laboratories, Inc. (1998); "Neurobeh" to the 1998 developmental neurobehavioral study (Argus Research Laboratories, 1998a); and "2-gen" to the completed 2-generation reproductive toxicity study in rats (Argus Research Laboratories, 1999).
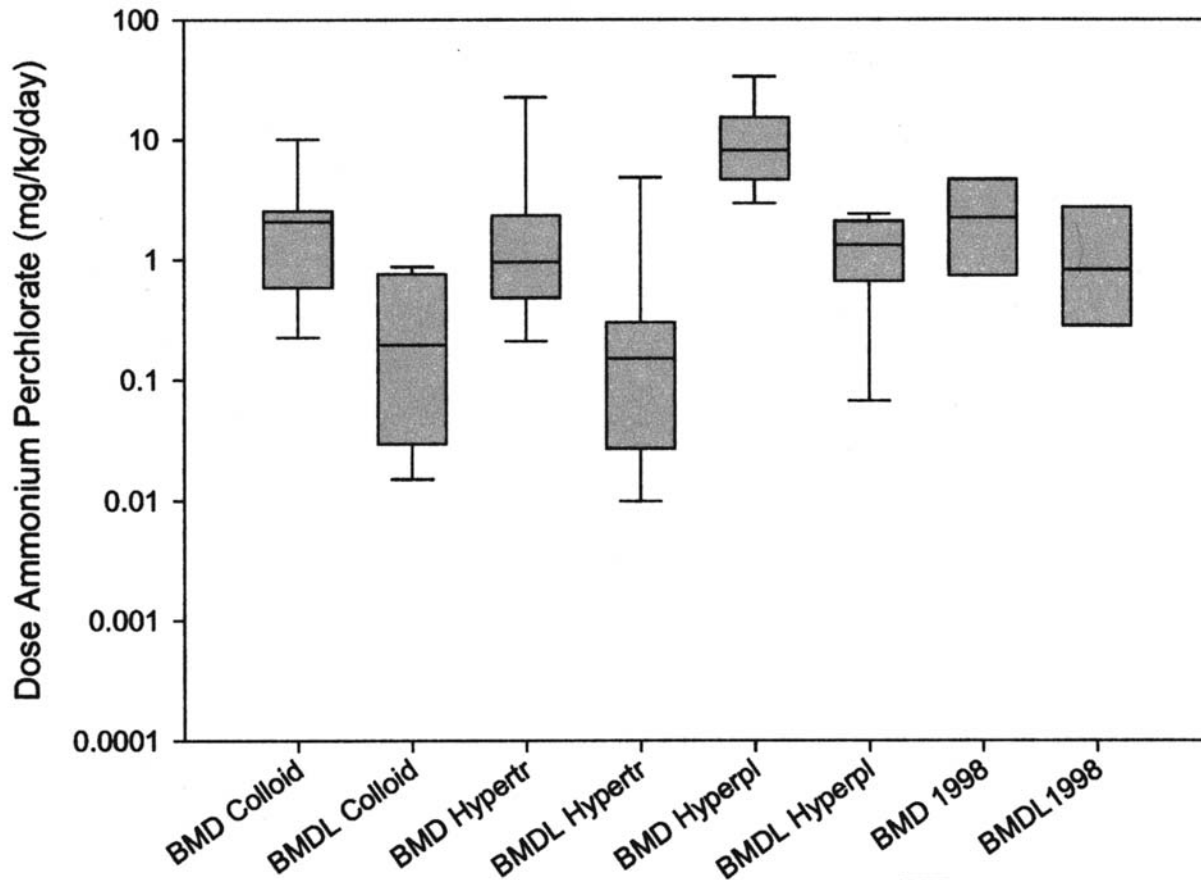
Figure 5-2. Distribution of BMD and BMDL estimates shown by "box and whisker" plots of colloid depletion (colloid), hypertrophy (hyptry), and hyperplasia (hyppls) from rat studies recalculated for thyroid histopathology based on 2000 Pathology Working Group review (Wolf, 2000; 2001). Values are presented in Table 5-1. Study #4 was excluded since it was a 30-day recovery experiment and #5 was excluded due to lack of monotonicity. The boundary of the box closest to zero indicates the 25th percentile, a line within the box denotes the median, and the boundary of the box farthest from zero indicates the 75th percentile. Whiskers above and below the box indicate the 90th and 10th percentiles. The two rightmost boxes plot values from the combined rat studies from the 1998 EPA risk characterization (U.S. Environmental Protection Agency, 1998d; Geller, 1998a).

1   approaches to the analyses that address these comments (Crofton and Marcus, 2001; Marcus,

2   2001; Crofton, 2001a). All thyroid and pituitary hormone analyses presented will utilize these

3   new approaches. The reanalyses of the hormone data for the previous set of studies can be found

4   in Table 5-2.

5   Finally, a number of additional new toxicology studies were recommended by the EPA and

6   the external review panel in 1999. These included a study of the developmental effects of

7   perchlorate (Section 5.4.3); a re-evaluation of the effects of perchlorate on neurodevelopmental

8   motor activity (Section 5.3.2); refinement of the evaluation of immunotoxicity concerns with a

9   repeat of the sheep red blood cell (SRBC) response using the established plaque-forming cell

10 (PFC) assay for humoral effects and an additional test for contact hypersensitivity (Section 5.6);

11 and what has become known as the "Effects Study" (Section 5.3.3). The objective of the

12 "Effects Study" (Argus Research Laboratories, Inc., 2001) was to reevaluate brain morphometry

13 effects and to evaluate thyroid histopathology and thyroid and pituitary hormones at various

14 stages of development, including during gestation and post-natal days 5, 10 and 22.

15

16

17 **5.1   CHRONIC STUDIES AND GENOTOXICITY ASSAYS**

18   This section discusses the data establishing perchlorate as a carcinogen. A few long-term

19 studies at comparatively high doses performed before the 1997 perchlorate testing strategy

20 showed that perchlorate causes thyroid tumors. These studies are discussed in Section 5.1.1. In

21 order to invoke the conceptual mode-of-action framework for the anti-thyroid effects of

22 perchlorate causing thyroid neoplasia via a non-genotoxic mechanism, the testing strategy had to

23 determine whether or not perchlorate acts directly with DNA. This evidence is discussed in

24 Section 5.2.2. The completed genotoxicity data were presented at the 1999 external peer review

25 as Attachment A to the February 1, 1999 submission provided by NCEA to the peer review panel

26 (Zeiger, 1999a,b; Dellarco, 1999; BioReliance, 1999; Moore, 1999). Dr. David Brusick, the

27 genetic toxicologist on the previous external peer review panel, agreed with the EPA conclusions

28 (Research Triangle Institute, 1999) that perchlorate's ability to cause thyroid tumors was not

29 likely to be via a directly genotoxic mechanism.

30   It should be noted that perchlorate exposure also caused a statistically-significant increase

31 in tumors at the 30 mg/kg-day dose in the F1-generation pups of the two-generation rat

TABLE 5-2. A COMPARISON OF NOAELs AND LOAELs FROM THE ORIGINAL 1998 ANALYSES AND THE 2001 RE-ANALYSES FOR HORMONE AND MORPHOMETRY ON THYROID FOLLICULAR LUMEN SIZE
(Crofton and Marcus, 2001; Marcus, 2001; Crofton, 2001a)

Species/Study	Time Point, Age (Doses, mg/kg-day)	Endpoint	Sex	Original Analyses		Re-Analyses[a,b]	
				NOAEL	LOAEL	NOAEL	LOAEL
Rat 14-Day (Caldwell et al., 1995)	14-Day *(males - 0.0, 0.11, 0.44, 1.11, 2.26, 4.32, 11.44, 22.16) (females - 0.0, 0.12, 0.47, 1.23, 3.06, 4.91, 11.47, 24.86)*	T3	M	0.11	0.44	0.11	0.44
			F	—	0.11	—	0.12
		T4	M	—	0.11	—	0.11
			F		0.12		0.12
		TSH	M	0.44	1.11	0.44	1.11
			F	0.12	0.47	—	**0.12**
		hTg	M	—	0.11	—	0.11
			F	—	0.12	—	0.12
		rT3	M	0.44	1.11	**0.11**	**0.44**
			F	0.47	1.23	**0.12**	**0.47**
Rat Subchronic Study (Springborn, 1998)	14-Day *(0, 0.01, 0.05, 0.2, 1.0, 10.0)*	T3	M	—	0.01	—	0.01
			F	10.0	—	10.0	—
		T4	M	1.0	10.0	—	**0.05**
			F				
		TSH	M	0.05	0.2	**0.01**	**0.05**
			F	0.01	0.05	—	**0.01**

TABLE 5-2 (cont'd). A COMPARISON OF NOAELs AND LOAELs FROM THE ORIGINAL 1998 ANALYSES AND THE 2001 RE-ANALYSES FOR HORMONE AND MORPHOMETRY ON THYROID FOLLICULAR LUMEN SIZE (Crofton and Marcus, 2001; Marcus, 2001; Crofton, 2001a)

Species/Study	Time Point, Age *(Doses, mg/kg-day)*	Endpoint	Sex	Original Analyses		Re-Analyses[a,b]	
				NOAEL	LOAEL	NOAEL	LOAEL
Rat Subchronic Study (Springborn, 1998) (cont'd)	90-Day *(0, 0.01, 0.05, 0.2, 1.0, 10.0)*	T3	M	—	0.01	—	0.01
			F	—	0.01	—	0.01
		T4	M	—	0.01	—	0.01
			F			—	0.01
		TSH	M	0.05	0.2	0.05	0.2
			F				
Rat Subchronic Study (Springborn, 1998)	120-Day *(0, 0.05, 1.0, 10.0)*	T3	M	1.0	10.0	1.0	10.0
			F				
		T4	M	—	0.05	**—**	**0.05**
			F			**1.0**	**10.0**
		TSH	M	10.0	—	—	**0.05**
			F				
Rat Developmental Neurotoxicity Study (Argus, 1998a)	PND5 *(0, 0.1, 1.0, 3.0, 10.0)*	Lumen size	M	1	3	**0.3**	**3**
			F				
	PND90 *(0, 0.1, 1.0, 3.0, 10.0)*	Lumen size	M	Data not available for original analyses		**10**	—
			F			**10**	—
	PND5 *(0, 0.1, 1.0, 3.0, 10.0)*	T4		1.0	3.0	**0.1**	**1.0**
		T3		0.1	1.0	0.1	1.0
		TSH		3.0	10.0	3.0	10.0
	PND90	T4, T3, and TSH		No data available			

TABLE 5-2 (cont'd). A COMPARISON OF NOAELs AND LOAELs FROM THE ORIGINAL 1998 ANALYSES AND THE 2001 RE-ANALYSES FOR HORMONE AND MORPHOMETRY ON THYROID FOLLICULAR LUMEN SIZE (Crofton and Marcus, 2001; Marcus, 2001; Crofton, 2001a)

Species/Study	Time Point, Age (Doses, mg/kg-day)	Endpoint	Sex	Original Analyses NOAEL	Original Analyses LOAEL	Re-Analyses[a,b] NOAEL	Re-Analyses[a,b] LOAEL
Mouse Hormone and Immunotoxicity (Keil et al., 1998)	14-Day (0.0, 0.1, 1.0, 3.0, 30)	T4	M	3.0	30.0	—	**0.1**
		T3	M	Data not available at time of 1998 analysis		—	**0.1**[c]
		TSH	M	No data			
	90-Day (0.0, 0.1, 1.0, 3.0, 30)	T4	M	0.1	3.0	—	0.1[c]
		T3	M	Data not available at time of 1998 analysis		—	**0.1**[d]
		TSH	M	30.0	—	—	**0.1**[d]
	120-Day (0.0, 0.1, 1.0, 3.0, 30)	T4	M	30.0	—	30.0	—
		T3	M	Data not available at time of 1998 analysis		30.0	—
		TSH	M	30.0	—	—	—
Rabbit Developmental Toxicity (Argus, 1998b)	Gestation Day 28 (0.0, 0.1, 1.0, 10.0, 30.0, 100.0)	T4	F	0.1	1.0	0.1	1.0
		T3	F	100	—	100	—
		TSH	F	100	—	100	—

[a]Bold indicates where 2001 analyses is different than 1998 analyses.
[b]Results from the liberal and conservative statistical approaches were the same.
[c]No dose response - 0.1 and 1.0 differ from control; 0.3 and 30.0 do not differ from control.
[d]No dose response - 0.1 and 1.0 differ from control; 0.3 and 30.0 do not differ from control.

1   reproductive study (Argus Research Laboratories, 1999). These pups were used as the parents of

2   the second generation (F2) pups in the study. When these F1 animals were sacrificed after only

3   19-weeks, tumors were observed (Wolf, 2000). The type was the expected benign thyroid

4   adenoma consistent with the anti-thyroid effect at the NIS (iodine uptake inhibition) with thyroid

5   hormone disruption followed by TSH upregulation. The early onset at 19 weeks is remarkable

6   and suggests the potential for in utero imprinting, a phenomenon beginning to be appreciated

7   with other endocrine-disrupting compounds (Prins et al., 2001; Phillips et al., 1998; Seckl, 1997).

8   These tumor results will be discussed in Section 5.5.

9

10   5.1.1 Cancer Studies

11   Kessler and Krüskemper (1966) provided potassium perchlorate in drinking water at a

12   concentration of 0 or 1% to male Wistar rats for 2 years. Body weights and thyroid weights were

13   reported for groups of 6 to 8 rats sacrificed after 0, 40, 120, 220, and 730 days of treatment, and

14   thyroid glands from the animals were examined histologically. Using body weight data provided

15   in the report to calculate a time-weighted average body weight of 0.336 kg and using an

16   estimated water consumption of 0.045 L/day (calculated with the allometric equation

17   recommended by U.S. Environmental Protection Agency [1987]), a dose of 1,339 mg/kg-day can

18   be derived. Body weights of control and treated animals were comparable throughout the

19   experiment. In contrast, thyroid weights, both relative and absolute, were increased markedly in

20   treated rats compared to controls at each examination interval. Histological examination of

21   thyroids from treated rats at 40 days revealed follicular cell hyperplasia. The authors

22   characterized these changes as typical for a thyroid gland stimulated by TSH during a relatively

23   short period of time. After 200 days of perchlorate treatment, diffusely degenerative changes

24   with fibrosis and increased colloid were observed. The authors commented that the course of the

25   histological changes in the thyroid was similar to that produced by long-term administration of

26   thiouracil, another antithyroid agent. The authors further reported that 4 of 11 rats treated with

27   potassium perchlorate for 2 years developed benign tumors of the thyroid gland and that

28   20 untreated Wistar control rats displayed no thyroid gland tumors. The 1,339 mg/kg-day dose

29   suggested a free-standing LOAEL because no other doses were tested.

30   Pajer and Kališnik (1991) administered 0 or 1.2% sodium perchlorate in drinking water to

31   groups of 36 female BALB/c mice (12/group) for up to 46 weeks. Eight or 12 weeks after the

1	beginning of the experiment, one group of treated and control mice were totally irradiated with
2	0.8 Gy on 5 consecutive days at a dose rate of 1.45 Gy/min so that each mouse received a total of
3	4 Gy. Assuming a body weight of 0.0353 kg and a water consumption rate of 0.0063 L/day (U.S.
4	Environmental Protection Agency, 1987), a dose of 2,147 mg/kg-day can be calculated. Thirty
5	animals died during the experimental period; however, details about the cause of death were not
6	provided. Forty-two animals were sacrificed at 46 weeks for histological examination of the
7	thyroid and pituitary gland. No other tissues were examined. Obvious treatment-related
8	histological changes were observed in the thyroid and pituitary gland, including thyroid follicular
9	cell carcinoma. Immunoperoxidase staining of pituitary thyrotropic cells and antihuman TSH
10	serum provided qualitative evidence of increased TSH production in the pituitary gland.
11	Perchlorate treatment was associated with an increased total volume of the thyroid and of the
12	distal parts of the anterior pituitary gland (adenohypophysis). In addition, increased average
13	volume and numbers of epithelial, thyrotropic, and parafollicular cells were observed. Irradiation
14	appeared to enhance the effects of perchlorate treatment. This study suggested a free-standing
15	LOAEL of 2,147 mg/kg-day for thyroid effects.
16	

5.1.2 Genotoxicity Assays

18	ManTech Environmental Technology, Inc. (1998) performed a battery of three genotoxicity
19	assays (*Salmonella typhimurium*/microsome mutagenesis assay [Ames assay], the mouse
20	lymphoma cell mutagenesis assay [L5178Y-TK test], and the *in vivo* mouse bone marrow
21	micronucleus induction assay) with ammonium perchlorate to help determine its potential for
22	various interactions with DNA and to gain insight into its possible carcinogenicity. To confirm
23	the findings of ManTech Environmental Technology, Inc., the EPA requested that the National
24	Toxicology Program (NTP) also evaluate ammonium perchlorate in the Ames assay and the
25	mouse bone marrow micronucleus test (Zeiger, 1999a). The sponsor (PSG) also had the mouse
26	lymphoma assay repeated (BioReliance, 1999).
27	Ammonium perchlorate was evaluated in the Ames assay (*Salmonella typhimurium*/
28	microsome mutagenesis assay), which is a well-defined assay for detection of mutagens.
29	It measures the reversion from a histidine-independent state (his⁻) induced by chemicals that
30	cause base-pair changes or frameshift mutations in the genome of the organism (i.e., it measures
31	for point mutations [e.g., substitution, addition, or deletion of one or a few DNA base pairs

1   within a gene]). In this assay, bacteria are exposed to the test chemical with and without a

2   metabolic activation system (Arochlor 1254-induced rat liver S9 with co-factors).

3   The mutagenicity is evaluated by the increase in the number of revertant colonies. The L5178Y

4   mouse-lymphoma assay is another short-term *in vitro* assay to detect both point mutations and

5   structural chromosomal changes. The *in vivo* mammalian micronucleus test detects the damage

6   of chromosomes or of the mitotic apparatus caused by a clastogenic chemical in bone marrow

7   cells (polychromatic erythrocyte [PCE] stem cells) of treated animals. Micronuclei are believed

8   to be formed from chromosomes or chromosome fragments left behind during anaphase of

9   mitosis. The induction of micronuclei indicates changes in either chromosome structure or

10   number in bone marrow cells. ManTech Environmental Technology, Inc. (1998) performed this

11   assay in Swiss-CD-1 mice and the NTP used B6C3F1 mice (Zieger, 1999a). The micronucleus

12   assay also was performed as part of the 90-day bioassay in Spraque-Dawley rats (Springborn

13   Laboratories, Inc., 1998). This is considered an adequate series of tests to determine the

14   mutagenic and clastogenic (chromosomal breaking) potential of an agent. It should be noted that

15   perchlorate is not likely to be mutagenic, given its physical and chemical properties (i.e., it is

16   simply an anion). Although perchlorate is an oxidizing agent, it is not expected to produce

17   oxidative DNA damage because of the kinetic considerations discussed in Chapter 2.

18

19   **5.1.2.1 *In Vitro* Assays**

20   Ammonium perchlorate was not found to be mutagenic in the *Salmonella typhimurium*

21   (Ames assay) with and without Arochlor 1254-induced rat liver S9 activation by two separate

22   laboratories (ManTech Environmental Technology, Inc., 1998; Zeiger, 1999a). In the ManTech

23   study, ammonium perchlorate was dissolved in distilled water and tested at five concentrations

24   (5,000, 2,500, 1,250, 625, and 312.5 μg/plate) in tester strains TA98, TA100, TA1535, and

25   TA1537, with and without Arochlor 1254-induced rat liver S9 using the plate incorporation

26   assay. Although this study was regarded as adequate, the EPA requested that the Ames assay be

27   repeated by the NTP to confirm the negative findings and to include additional tester strains (i.e.,

28   TA102, and TA104) that are able to detect a variety of oxidative mutagens. Therefore, the NTP

29   evaluated ammonium perchlorate in the Salmonella/Ames assay in tester strains TA98, TA100,

30   TA1535, TA97, TA102, and TA104 (Zeiger, 1999b). Ammonium perchlorate was dissolved in

31   distilled water and tested using the preincubation procedure at doses of 10,000, 3,333, 1,000,

1	333, and 100 μg/plate, with and without metabolic activation from Arochlor-induced rat and
2	hamster livers. Ammonium perchlorate was neither toxic nor mutagenic under the conditions of
3	the NTP assay.
4	The L5178Y/$tk^{+/-}$ mouse lymphoma assay also was used to evaluate the mutagenic and
5	chromosomal breaking potential of ammonium perchlorate in vitro. Ammonium perchlorate was
6	reported to be negative both in the absence and presence of rat Arochlor-induced S9 liver
7	activation (ManTech Environmental Technology, Inc., 1998). Ammonium perchlorate was
8	evaluated at 5.0, 2.5, 0.5, 0.25, 0.05, and 0.025 mg/mL without S9 activation, and at 2.5, 0.5,
9	0.25, 0.05, and 0.025 mg/mL with S9 activation. Although a small increase in mutation
10	frequency was found in the absence of S9 activation at 2.5 mg/mL, which appeared to be
11	statistically significant (p < 0.05) by the two-tail Student's t-test, a repeat assay found no increase
12	in mutation frequency at this concentration compared with controls. Therefore, ammonium
13	perchlorate is considered to be negative in the absence of S9 activation. Confidence in the
14	negative findings without S9 activation is reinforced by the wide range of ammonium perchlorate
15	concentrations evaluated. Although ammonium perchlorate also was reported as negative in the
16	presence of S9 activation, the response of the positive control, 3-methyl cholanthrene (MCA), in
17	the actual experiment was too low (182.6×10^{-6}) to be acceptable. The highest dose of
18	ammonium perchlorate produced a mutation frequency of 194×10^{-6}. The MCA at 2.5 μg/mL
19	should induce a mutation frequency of 300 to 350×10^{-6} or higher. Such a low positive control
20	response weakens the confidence for the negative finding with S9 activation. In addition, the
21	cloning efficiencies for the S9 test appear to be too high (143%), further reducing the confidence
22	in a negative finding. Therefore, only the assays on ammonium perchlorate without S9 are
23	considered unequivocally to be negative. Although perchlorate is not expected to be metabolized
24	to a mutagenic intermediate, these S9 data were not of sufficient quality to support a
25	negative-response conclusion.
26	Because of the problems described above, the sponsor (PSG) had the mouse lymphoma
27	assay repeated. In this recent mouse lymphoma assay, ammonium perchlorate was evaluated at
28	concentrations of 1000, 2000, 3000, 4000, and 5000 μm/ml without and with Arochlor
29	1254-induced rat liver S9 activation (BioReliance, 1999). No increase in mutant frequencies
30	were found after treatment with perchlorate. The data were judged to be of sufficient quality to
31	determine perchlorate to be nonmutagenic both with and without S9 activation. Although the

1   background mutant frequency was low, particularly in the S9 experiment, the data set still is

2   considered to be very good overall, as well as internally consistent. The problems that were

3   observed in the data generated by the first laboratory (ManTech Environmental Technology, Inc.,

4   1998) were not present in the data form the BioReliance (1999) study.

5

6   **5.1.2.2**   *In Vivo* **Assays**

7   The potential for ammonium perchlorate to induce micronuclei was evaluated in mice and

8   rats. Ammonium perchlorate was administered by drinking water gavage for 3 consecutive days

9   to Swiss CD-1 mice (5 females and 5 males per dose group) at 1,000, 500, 250, 125, and

10   62.5 mg/kg-day (ManTech Environmental Technology, Inc., 1998). Twenty-four hours after the

11   last dose, the mice were sacrificed, and the frequency of micronucleated cells were evaluated by

12   counting 1,000 PCEs per animal. The assay was conducted in accordance with existing EPA

13   Federal Insecticide, Fungicide, and Rodenticide Act/Toxic Substances Control Act

14   (FIFRA/TSCA) testing guidelines. No increase in the frequency of micronuclei were found for

15   any dose group. There is some uncertainty whether a maximum tolerated dose (MTD) was

16   reached in this study. The study authors reported that at 2,000 mg/kg, 4 out of 6 animals died

17   after one dosing of ammonium perchlorate. Typically, the assay is performed at 85% of the

18   MTD, and the 1,000 mg/kg-day represents approximately 50% of the LD_{50}. There was no

19   indication of toxicity to the bone marrow cells because the polychromatic erythrocyte to

20   normochromatic erythrocyte (PCE/NCE) ratio was not different from negative controls.

21   Furthermore, the study authors did not report any indication of clinical signs of toxicity in the

22   highest dose group. Despite a rebuttal submitted by Dourson (1998) on behalf of the sponsor

23   (PSG), EPA remained concerned because of the importance of this test in the overall

24   determination of the approach to be taken for the carcinogenicity assessment (i.e., to rule out

25   direct genotoxicity).

26   The NTP agreed to expedite and repeat this test in response to an EPA request. The assay

27   was performed by ip injection to ensure the greatest delivery to the bone marrow. Male B6C3F1

28   mice were treated with 125, 250, 500, 1,000, 1,500, and 2,000 mg/kg ammonium perchlorate in

29   buffered saline, plus solvent and positive (cyclophosphamide) controls. Note that this study uses

30   two dose groups higher than those used in the previous study (i.e., 1,500 and 2,000 mg/kg).

31   Furthermore, the use of ip injection as the route of administration would result in a direct

1   delivery of the compound to the bone marrow cells versus delivery from drinking water gavage.

2   Five mice per group were injected daily for 3 consecutive days and were sacrificed 24 h after the

3   last injection; 2,000 PCEs were scored per animal for micronuclei. All animals in the 1,500- and

4   2,000-mg/kg groups died after the first ip injection, and 4/5 animals died in the 1,000-mg/kg

5   group after the second ip injection. No increases in percent PCE were observed in any of the

6   remaining test groups (125, 250, and 500 mg/kg). No bone marrow toxicity was seen as

7   indicated by the percent of PCE (Zeiger, 1999a,b). These results are interpreted to be consistent

8   with those of the ManTech Environmental Technology, Inc. (1998) study that used gavage

9   drinking water administration, and confirm that perchlorate does not induce micronuclei in

10   rodents.

11   The 90-day subchronic bioassay using Spraque-Dawley rats also evaluated micronuclei

12   induction (Springborn Laboratories, Inc., 1998). The frequency of micronuclei induction was

13   examined in both the males and females after the 90-day sacrifice in the 10-mg/kg-day dose

14   group of ammonium perchlorate administered by drinking water. Although there was no

15   induction of micronuclei at this dose, 10 mg/kg-day does not appear to reach a MTD because

16   there were no overt signs of toxicity. However, the definition of MTD may be somewhat moot,

17   given the changes in thyroid hormone economy and histopathology seen in the thyroids at that

18   dose. There was significant reduction in the PCE/NCE ratio (i.e, an indicator of toxicity to the

19   bone marrow cells).

20

21   **5.1.2.3   Summary of Genotoxicity Battery Results**

22   Negative results were reported in all genotoxicity assays conducted on ammonium

23   perchlorate when evaluated by two independent laboratories. Ammonium perchlorate was not

24   mutagenic in the Ames assay (with or without S9 activation). Negative results were also found

25   in the mouse lymphoma gene mutation assay without and with S9 activation. Ammonium

26   perchlorate did not induce chromosomal anomalies when evaluated for micronuclei induction in

27   the bone marrow of mice when administered via drinking water gavage or ip injection.

28   No increases in micronuclei were found in Spraque-Dawley rats when evaluated from the 90-day

29   study at the highest dose, which produced both thyroid hormone perturbations and follicular cell

30   hyperplasia.

1   In conclusion, ammonium perchlorate does not have the potential to be mutagenic or

2   clastogenic. The *in vitro* and *in vivo* studies discussed above provide support for that conclusion.

3   Therefore, mutagenicity is not considered a possible mode of carcinogenic action for this

4   chemical.

5

6

5.2 GENERAL TOXICITY: SHORT-TERM AND SUBCHRONIC TESTING

9   The majority of the data on perchlorate toxicity available from previous studies or as a

10   result of the current perchlorate testing strategy involved either short-term or subchronic

11   exposures and are presented in this section. As discussed in Chapter 3, the testing strategy

12   included targeted studies to evaluate different endpoints, e.g., developmental neurotoxicity

13   (Section 5.3), developmental studies (Section 5.4) reproductive studies (Section 5.5) and

14   immunotoxicity assays (Section 5.6). The rationale behind the 90-day study (Section 5.2) with

15   satellite examination of thyroid and pituitary hormones and a 30-day recovery period was to

16   evaluate anti-thyroid effects as possible precursor lesions. If a NOAEL could be established for

17   these precursor lesions, it was thought that a two-year bioassay would not be required. This

18   assumption is now more tenuous due to the tumors observed in the F1-generation at 19 weeks.

19   The integration of these results with the available human data to arrive at a risk assessment will

20   be discussed in Chapter 7.

21

5.2.1 Historical Data

23   Mannisto et al. (1979) measured serum levels of TSH, T3, and T4 by RIA in groups of 5 to

24   6 male Sprague-Dawley rats weighing 180 to 220 g that were exposed to potassium perchlorate

25   in their drinking water at concentrations of 0, 10, 50, 100, or 500 mg/L for 4 days. Potassium

26   perchlorate doses of 0, 1.5, 7.6, 15.3, or 76.3 mg/kg-day, respectively, were calculated assuming

27   a body weight of 0.2 kg and a water consumption rate of 0.0305 L/day (U.S. Environmental

28   Protection Agency, 1987). Perchlorate produced statistically significant increases in serum TSH

29   levels and decreases in serum T3 and T4 levels. Significant changes in all three parameters were

30   measured in the 100 and 500 mg/L (15.3 and 76.3 mg/kg-day, respectively) dose groups. In the

31   50 mg/L (7.6 mg/kg-day) dose group, levels of T3 and T4 were decreased significantly; TSH

1 levels were increased slightly, but the increase was not significant. At the low dose, T3, T4, and

2 TSH levels were unchanged from controls. This study suggested a NOAEL of 1.5 mg/kg-day

3 and a LOAEL of 7.6 mg/kg-day for short-term exposures to potassium perchlorate.

4   Shigan (1963) administered 190 mg/kg-day of potassium perchlorate in water to rabbits and

5 white rats (number, sex, and strain not identified) for 3 mo. The author did not indicate whether

6 the compound was administered in drinking water or by gavage with water. The animals were

7 examined for cardiac function; liver function, based on changes in serum proteins; immune

8 function, based on leukocyte phagocytosis; and adrenal function. Perchlorate at the dose tested

9 caused a change in the electrocardiogram and a decrease in serum proteins, indicating a

10 disruption of the glycogen-forming function of the liver. Shigan (1963) did not indicate whether

11 these changes were observed in both rabbits and rats. Perchlorate had no effect in the remaining

12 tests. This study suggested a LOAEL of 190 mg/kg-day although the study translation is reported

13 incompletely, limiting its usefulness for risk assessment.

14   In a second set of experiments, Shigan (1963) also treated rabbits and white rats (number,

15 sex, and strain not identified) with 0, 0.25, 2.0, and 40 mg/kg-day of potassium perchlorate for

16 9 mo. The medium for dosing was not reported. The animals were examined for cardiac and

17 liver function, for conditioned reflexes, and for uptake and discharge of iodide by the thyroid. In

18 the two highest dose groups, there was a statistically significant increase in the amount of iodide

19 excreted from the thyroid; this increase was not observed in the 0.25-mg/kg-day dose group. The

20 study does not indicate if the effect was seen in one or both species tested. This study suggested

21 a NOAEL of 0.25 mg/kg-day and a LOAEL of 2 mg/kg-day for thyroid effects.

22   Hiasa et al. (1987) measured serum levels of T3, T4, and TSH by radioimmunassay in

23 groups of 20 male Wistar rats administered 0 or 1,000 ppm potassium perchlorate in the diet for

24 20 weeks. Assuming a body weight of 0.34 kg (the average final body weight of rats treated with

25 perchlorate) and a food consumption rate of 27.4 g/day (U.S. Environmental Protection Agency,

26 1987), an estimated dose of 80.7 mg/kg-day was calculated. Absolute and relative thyroid

27 weights were significantly increased compared to controls in perchlorate-treated rats. No effects

28 were seen on liver weights. The T4 levels decreased slightly, but the decrease was

29 not statistically significant. The T3 levels were unchanged compared to controls. The TSH

30 levels were increased statistically significantly compared to controls. Histological examination

1   of the thyroid revealed diffused small follicles in perchlorate-treated rats and one case of

2   follicular hyperplasia. Thus, the 80.7-mg/kg-day dose could be considered a LOAEL.

3   Gauss (1972) fed female NMRI mice a diet containing 0 or 1% potassium perchlorate for

4   up to 160 days. Mice were between 50 and 60 days old at the beginning of treatment and

5   weighed between 19 and 28 g (average, 23.23 g). During the first 2 mo of treatment, body

6   weights increased about 12%; body weight data for longer treatment periods were not reported.

7   Assuming a body weight of 23 g and a food consumption value of 4.625 g/day (U.S.

8   Environmental Protection Agency, 1987), a dose of 2,011 mg/kg-day was calculated. Thyroid

9   glands were examined histologically at 10- to 20-day intervals throughout the 160-day study

10   period. Thyroid and nuclei volumes and height of epithelial follicles were increased in treated

11   mice throughout the treatment period compared to controls. The histological examinations

12   showed a progressive change in the histological appearance of the thyroid of treated mice,

13   beginning with colloid loss, nuclei volume expansion, and rising epithelium height, followed by

14   the appearance of hypertrophy and hyperplasia of the thyroid parenchyma. At later stages of the

15   treatment period, hyperplastic follicles, areas of adenomatic tissue, adenoma complexes, and

16   secreting cystadenomas were observed; however, no progression to malignancy was apparent.

17   The 2,011 mg/kg-day dose suggested a free-standing LOAEL because no other doses were tested.

18

19   ## 5.2.2 Caldwell et al. (1995) 14-Day Study

20   Caldwell et al. (1995) administered ammonium perchlorate in drinking water at

21   concentrations of 0, 1.25, 5.0, 12.5, 25, 50, 125, or 250 mg/L to Sprague-Dawley rats

22   (6/sex/group) for 14 days. The actual dose administered to each animal was calculated by

23   multiplying the concentration of ammonium perchlorate administered in the drinking water by

24   each rat's average water consumption over the 14-day period and dividing this number by each

25   animal's average body weight for the same period, resulting in doses (male/female) of 0,

26   0.11/0.12, 0.44/0.47, 1.11/1.23, 2.26/3.06, 4.32/4.91, 11.44/11.47, and 22.16/24.86 mg/kg-day,

27   respectively (Caldwell et al., 1995). Caution must be used when interpreting these reports

28   because the conversion is sometimes not included (e.g., the Channel [1998b] consultative letter

29   reports results in units of the test concentrations rather than the dose converted to milligrams per

30   kilogram per day). Thyroids were weighed, histopathology and morphometry performed, and

31   thyroid hormone levels were measured with a radioimmune assay technique.

1   The consultative letter of Channel (1998b) provides results and comments on a

2   histopathological analysis of the rat thyroids from the Caldwell et al. (1995) 14-day study that

3   was performed by the Air Force Research Laboratory/Human Effectiveness Directorate

4   (AFRL/HEST) and never officially published (Eggers, 1996, as cited in Channel, 1998b).

5   As part of the previous assessment, EPA requested from the AFRL/HEST the previously

6   unpublished histopathology data from the 14-day oral dosing study performed by Caldwell et al.

7   (1995). The histopathology was discussed in the paper on the study design (Caldwell and Mattie,

8   1995) but had not been published in either Caldwell et al. (1995) or King (1995). The

9   histopathology data discussed herein were provided in a consultative letter from the AFRL/HEST

10   (Channel, 1998b). The EPA also performed a reanalysis of the thyroid hormone data (T4, T3,

11   rT3, TSH, and thyroglobulin [hTg]) found in the Caldwell et al. (1995) and King (1995) reports

12   (Crofton, 1998a). Because these individual data were supplied only electronically on Microsoft

13   Excel® spreadsheets and not submitted formally to EPA, Crofton, (1998a) represents the official

14   publication of these data. These histopathology data and reanalyses of effect levels using the

15   PWG results and new hormone analyses are found in the following sections.

16

17   **5.2.2.1   Thyroid Histology Data**

18   Channel (1998a) submitted that the incidence of thyroid follicular cell hypertrophy

19   determined by standard histology was significantly different from control at a lower dose

20   (0.44 0.47 mg/kg-day) than for the incidence of decrease in follicular lumen size (2.26

21   3.06 mg/kg-day), but the statistics indicate a NOAEL at 0.11 0.12 mg/kg-day. However, the

22   documentation of the statistics was not provided, and Eggers (1996) apparently combined both

23   sexes for the analyses. It is recommended in the report (Channel, 1998a), and EPA concurred,

24   that a re-analysis was warranted for a number of reasons. First, there was a gender-by-treatment

25   interaction observed in the thyroid hormone analyses (see Section 5.2.2.2). Secondly, there was

26   an apparent dose trend, despite the limited sample size, in the incidence of response: male and

27   female combined was 7/12, 6/11, 11/12, 10/12, 12/12, 12/12, 12/12, and 12/12; male only was

28   3/6, 4/6, 5/6, 5/6, 6/6, 6/6, 6/6, and 6/6; and female only was 4/6, 2/5, 6/6, 5/6, 6/6, 6/6, 6/6, and

29   6/6 for the 0, 0.1, 1.0, 5.0, 10, 20, 50, and 100 mg/kg-day groups, respectively. Finally, the

30   analysis did not combine severity and incidence data for the decrease in lumen size, but the mean

31   severity scores alone were statistically significant from control above the 0.44/0.47 mg/kg-day

1   group. A separate computerized morphometric analysis of follicular lumen size was performed

2   by AFRL/HEST for the 0, 0.11/0.12, 1.11/1.23, 4.32/4.91, and 22.16/24.86 mg/kg-day groups,

3   and a statistically significant difference in the incidence of decrease in lumen size was evident in

4   the males at the 1.11 mg/kg-day dose and, in females, at the 4.91 mg/kg-day dose; however, the

5   gender-by-treatment effect was not taken into account. Relative thyroid weights were

6   significantly increased in 11.44/11.47 and the 22.16/24.86 mg/kg-day dose groups compared to

7   controls.

8   Results of the PWG analysis can be found in Wolf (2000; 2001; Tables 1 and 2). Female

9   rats appeared to be slightly more sensitive in this study with a NOAEL designated at 1.23 mg/kg-

10 day; whereas, in males it was somewhat difficult to ascertain. This may be due to the difficulty

11 that the PWG had in reading the slides from this study due to the non-routine staining method

12 (periodic acid shift [PAS] reaction with a green counterstain) as noted in Wolf (2000). BMD

13 analysis (Table 5-1) for the combined female and male data results in BMDL values for a 10%

14 increase in incidence at 0.72 mg/kg-day for colloid depletion and 0.78 mg/kg-day for hyperplasia.

15 The difficulty noted above with the staining for this study was most prominent in evaluating

16 hypertrophy (Wolf, personal communication), so that these estimates were not calculated.

17 Re-analysis of the morphometry on thyroid follicular lumen size identified a NOAEL at the

18 0.44/0.47 mg/kg-day dose.

19

20 **5.2.2.2   Thyroid and Pituitary Hormone Analyses**

21   The thyroid and pituitary hormone data were reanalyzed using five two-way analysis of

22 variance (ANOVA) tests, one each for all of the hormones (Crofton, 1998a). Data from

23 dependent measures (T3, T4, rT3, TSH, and hTg) were subjected to separate two-way ANOVAs,

24 with gender (male and female), and treatment (dose) as independent, between-subject variables.

25 Step-down ANOVA tests were conducted as indicated by significant interactions and discussed

26 in Crofton and Marcus (2001) and Marcus (2001). Mean contrasts were performed using

27 Duncan's Multiple Range Test. Results of these reanalyses are similar to those stated in the

28 Caldwell et al. (1995) and King (1995) reports with some notable exceptions. Figure 5-3 shows

29 the dose-dependent effects on T3, T4, and TSH.

30   There was a significant gender-by-treatment interaction on total serum T3, and subsequent

31 step-down ANOVA tests showed significant treatment effects for both genders. Figure 5-3(A)
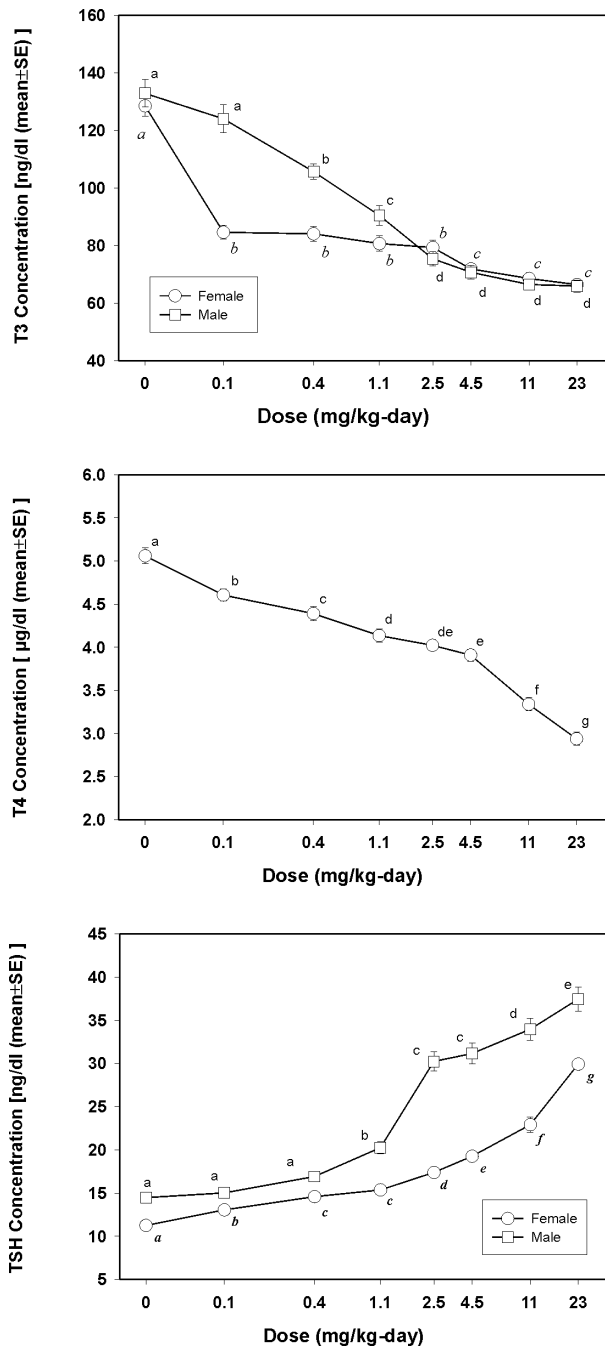
Figure 5-3. **Effects in the Caldwell et al. (1995) study of 14-day drinking water administration of ammonium perchlorate to SD rats on serum total T3 (A), T4 (B), and TSH (C) concentrations (ng/mL; mean ± SE) as recalculated in Table 5-2 (Crofton and Marcus, 2001). Means with different letters were significantly different (p < 0.05). Data of Channel (1998b) and Crofton (1998a). Daily dose was estimated from water consumption data.**

1   illustrates dose-dependent decreases in T3 for both genders while females were slightly more

2   sensitive compared to males. The overall gender-by-treatment interaction was not significant for

3   T4, but there was a significant main effect of treatment (Figure 5-3(B)). Perchlorate also

4   decreased T4 in a dose-dependent manner. There was a significant gender-by-treatment

5   interaction on total serum TSH, and subsequent step-down ANOVA tests showed significant

6   treatment effects for both genders. Dose-dependent increases in TSH were observed for both

7   genders; however, females were slightly more sensitive compared to males.

8   The Caldwell et al. (1995) study is the only one in which an additional thyroid hormone,

9   rT3, and hTg were assayed (Tg in rats was assayed with a human RIA kit, thus the notation "h").

10   There was no significant gender-by-treatment interaction for rT3. Figure 5-4(A) clearly indicates

11   that perchlorate increases rT3 in a dose-dependent manner. There was a significant gender-by-

12   treatment interaction on hTg, and subsequent step-down ANOVA tests showed significant

13   treatment effects for both genders. Figure 5-4(B) illustrates the dose-dependent increases in hTg

14   for both genders. Both genders were equally sensitive, with males exhibiting a slightly greater

15   response to the lowest dosage.

16   Perchlorate exposure decreased circulating T3 and T4 and increased TSH. This report also

17   provides evidence that rT3, formed mostly in extrathyroidal tissues, was increased by this

18   exposure. Thyroglobulin also was increased. The NOAELs and LOAELs are summarized in

19   Table 5-2. A NOAEL for TSH was observed in males only at 0.44 mg/kg-day and at 0.11/0.12

20   for rT3. Note that free-standing LOAELs (i.e., effects at the lowest dosage tested) were found at

21   0.11/0.12 mg/kg-day for T3 in females, for T4 and hTg in both sexes, and for TSH in females.

22

5.2.3 The 90-Day Testing Strategy Bioassay in Rats

24   The 90-day study that was part of the testing strategy consisted of oral administration of

25   ammonium perchlorate via drinking water to male and female Sprague-Dawley rats at doses of

26   0, 0.01, 0.05, 0.2, 1.0, and 10 mg/kg-day (Springborn Laboratories, Inc., 1998). This study has

27   also been reported in the literature (Siglin et al., 2000), but because that manuscript did not use

28   the thyroid histopathology as reported by the PWG (Wolf, 2000) it will not be discussed further

29   in this document. A 14-day sacrifice also was included in the study for comparison with the

30   Caldwell et al. (1995) study of that same duration. Ten rats/sex/dose were used, and an

31   additional 10 rats/sex/dose were sacrificed after the 30-day recovery period following cessation
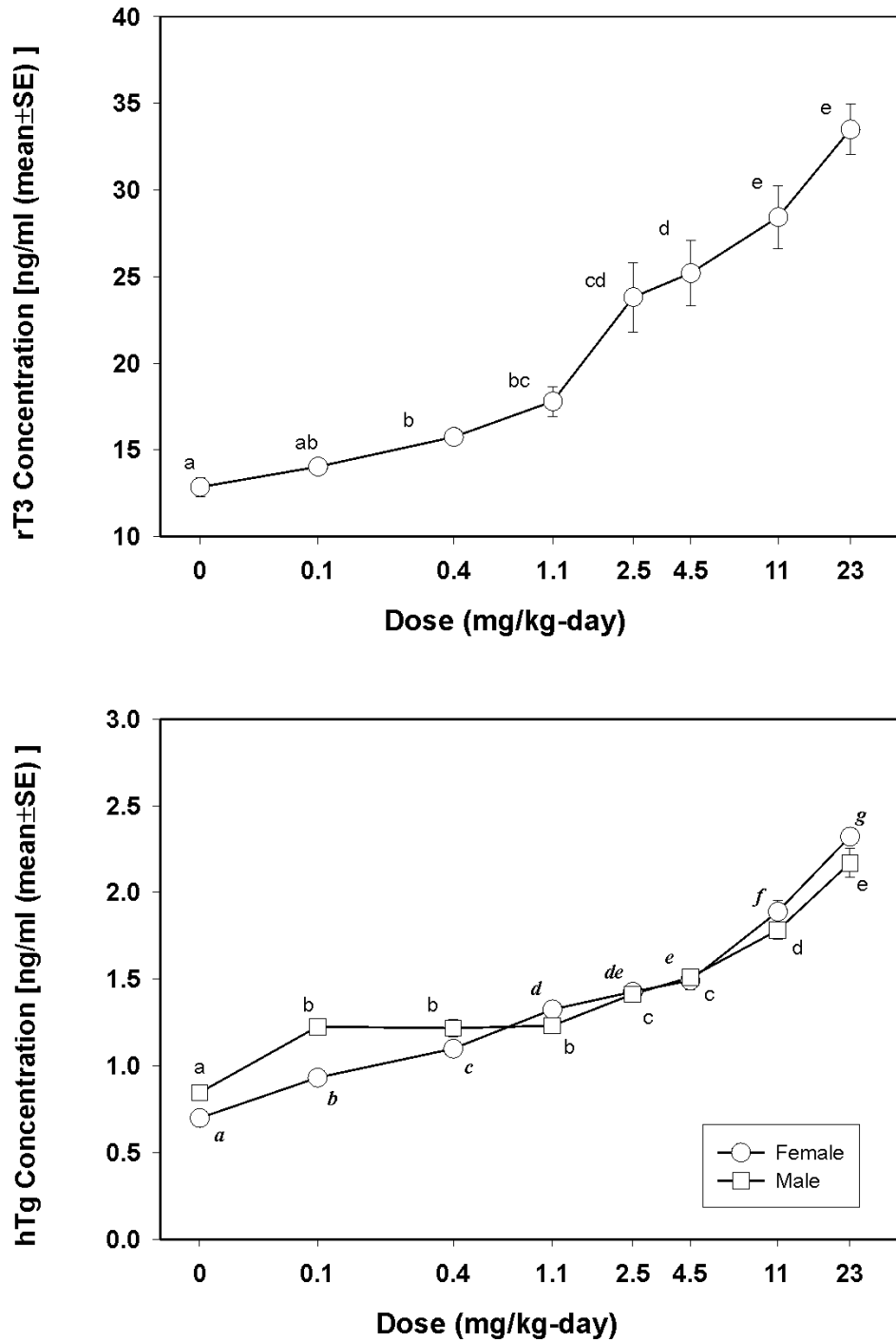
Figure 5-4. Effects in the Caldwell et al. (1995) study of 14-day drinking water administration of ammonium perchlorate to SD rats on serum rT3 (A) and hTg (B) concentrations (ng/mL; mean ± SE) as recalculated in Table 5-2 (Crofton and Marcus, 2001). Data of Channel (1998b) and Crofton (1998a). Means with different letters were significantly different (p < 0.05). Daily dose was estimated from water consumption data.

1 of the 90-day exposure at doses of 0, 0.05, 1.0, and 10 mg/kg-day to evaluate reversibility of any

2 observed lesions.

3   The stock solution of the test article was diluted with reverse osmosis (RO) water and

4 prepared fresh five times during the study (at least once every 5 weeks). Stability analyses were

5 performed by the sponsor (AFRL/HEST) and showed that ammonium perchlorate solutions were

6 stable for 109 days (Tsui et al., 1998). The sponsor also confirmed that the stock and dosing

7 solutions were within an acceptable concentration range (Springborn Laboratories, Inc., 1998;

8 Appendix B). Control drinking water solutions were analyzed by the sponsor to confirm no

9 contamination of detectable nitrate, an ion that could cause possible interference to estimating the

10 dose of test article. Dosing solutions were prepared fresh for each week, and the administered

11 concentrations were adjusted based on measured body weights and water intake.

12   The parameters evaluated included clinical observations, body and organ weights, food and

13 water consumption, hematology, clinical chemistry, ophthalmology, and gross necropsy.

14 Histopathology was performed on all tissues from the control and high-dose groups. The liver,

15 kidneys, lungs, thyroid/parathyroid, and gross lesions from all intermediate dose groups and for

16 the recovery groups also were examined microscopically. Evaluation of additional reproductive

17 parameters, i.e., estrous cyclicity in females and sperm motility and morphology in males, also

18 was performed. Thyroid hormone analyses were performed at the 14-, 90-, and 120-day

19 sacrifices. Only the 0, 0.05, 1.0 and 10.0 mg/kg-day groups were continued until the 120-day

20 time point. All hormone and tissue collection was balanced over time-of-day to control for the

21 circadian rhythms of hormones.

22

23 **5.2.3.1   General Toxicity, Thyroid Histopathology Results, and Satellite**
24 **Reproductive Assay**

25   There were no clinical signs of toxicity observed during the treatment or recovery periods.

26 All rats survived to scheduled sacrifice except one female rat in the 0.05 mg/kg-day group that

27 was found dead during the recovery period. However, this death was considered unrelated to

28 treatment because no deaths occurred in any of the higher dose groups, and the histopathologic

29 evaluation for cause of death was inconclusive. No statistically significant or remarkable

30 findings were observed among the groups with respect to clinical observations, body weights,

31 food or water consumption, ophthalmology, hematology, or clinical chemistry.   Miscellaneous

1   lesions that occurred with equal incidence and severity in all dose groups and controls included

2   extramedullary hematopoiesis in the livers, inflammation in the lungs, minimal nephropathy in

3   the kidneys and inflammation of the heart. Because none of these lesions demonstrated a dose

4   response, and some are commonly seen in young rats, they were not considered treatment-related.

5   The only treatment-related lesions observed at gross necropsy were reddened thyroids, attributed

6   to minimal congestion of the blood vessels.

7   Absolute thyroid weight and thyroid weight relative to both final body weight and brain

8   weight were increased significantly in males of the 10 mg/kg-day dose group after 14 and

9   90 days of treatment and in females at the 10 mg/kg-day dose group after 90 days indicating

10   LOAEL at 10 and a NOAEL at 1 mg/kg-day. These thyroid weight measures were comparable to

11   control values in both males and females of the 10 mg/kg-day group at the end of the 30-day

12   recovery period. Histopathology was evaluated on Days 14, 90, and 30 postexposure (120 days).

13   The corresponding PWG review results can be found in Wolf (2000; 2001, Tables 3 through 8).

14   Male rats appeared to be slightly more sensitive, exhibiting follicular cell hyperplasia by Day 14

15   and not recovering fully for any of the thyroid histopathological indices by 30 days post

16   exposure. On Day 14, females showed decreased colloid and follicular cell hypertrophy at

17   10 mg/kg-day. Males also showed a significant increase in these two thyroid response measures

18   at this dose but also exhibited changes at lower doses and in addition showed hyperplasia.

19   By 90 days, all three response measures (colloid depletion, follicular cell hypertrophy, and

20   follicular cell hyperplasia) in both sexes were significant at 10.0 mg/kg-day, again indicating a

21   LOAEL at 10 and a NOAEL at 1 mg/kg-day. Recovery of the thyroid histopathological changes

22   was essentially complete by 30 days post-exposure although the males did have some indication

23   of residual toxicity.

24   The BMD analyses for these data are found in Table 5-1 and Figures 5-1 and 5-2. Data for

25   females and males were combined. The BMDL for colloid depletion and hypertrophy at 14 days

26   were 0.28 and 0.017 mg/kg-day, with no estimate for hyperplasia. By 90-days, the BMDL values

27   decreased for colloid depletion and hypertrophy to 0.03 and 0.008 mg/kg-day. The BMDL value

28   for hyperplasia was 2.09 mg/kg-day. No observed effect was estimated for the 120 day value.

29   Estrous cyclicity was evaluated for 3 weeks prior to sacrifice in all females of the 90- and

30   120-day termination groups by examining daily vaginal smears. The number and percentage of

31   females cycling and the mean cycle length were determined for each group. There is an apparent

1	dose-related response for the absolute number and proportion of females with an abnormal
2	estrous cycle (defined as less than 3 or more than 5 days). The number and percentage of
3	females with at least one abnormal cycle in those females cycling was 1/10 (10%), 1/10 (10%),
4	5/9 (56%), 6/9 (67%), 0/8 (0%), and 0/10 (0%) at the 0, 0.01, 0.05, 0.2, 1.0, and 10-mg/kg-day
5	doses. The proportion began to increase at the 0.05 mg/kg-day dose level, peaked at the
6	0.2 mg/kg-day dose level, and then declined at the two higher doses. This suggests the
7	possibility of an inverted U-shaped dose-response pattern. Examination of the 120-day data
8	(after 30-day recovery) also revealed changes in cyclicity with 1/5 (20%), 1/7 (14%), 1/6 (16%),
9	and 4/6 (67%) females not cycling in the 0.0, 0.05, 1.0, and 10-mg/kg-day groups, respectively.
10	Because the number of rats in the add-on groups (n = 10) did not provide the level of statistical
11	power that would be desired, this indication of an effect in a study with limited power was of
12	concern in 1998, but the results of the two-generation reproductive study completed in 1999 did
13	not indicate any effects on this endpoint (Section 5.5.1).
14	Sperm samples were obtained from all male rats terminated after 90 or 120 days for
15	evaluation of sperm count, concentration, motility, and morphology. The mean percentage of
16	normal sperm was calculated for each group. There were no treatment-related effects on sperm
17	parameters noted although again the number tested is small. The effects on the percentage of
18	normal sperm appear to be artifacts because of a single outlier in each of the two groups with
19	lower means. These occurred at different dose levels in the exposure versus recovery phases.
20	

21 **5.2.3.2   Thyroid and Pituitary Hormone Analyses**

22	The assays for T4, T3, and TSH were performed using RIA kits according to the
23	manufacturer's standard procedures. Assay kits from the same batch number and with the same
24	expiration date were used for each animal termination period (Study Days 14, 90, or 120).
25	Samples and standards were run in triplicate. The Springborn Laboratories report included an
26	appendix (Springborn Laboratories, Inc., 1998; Appendix I) containing the results of these
27	thyroid hormone assays. The Springborn report used a series of individual ANOVA tests to
28	determine main effects of treatment for all three hormones in both genders and at three time
29	points during the study (Day 14, Day 90, and Day 120 a [30-day recovery time]). As part of its
30	1998 assessment, EPA reanalyzed these thyroid hormone data using three-way ANOVA tests,
31	one for each of the three hormones, to allow for a statistical comparison of the interaction

1   between gender, time, and treatment (Crofton, 1998b). The Crofton (1998b) analysis also

2   contains a printout of all of the individual animal data, an omission from Springborn

3   Laboratories, Inc. (1998). As suggested in the external peer review (Research Triangle Institute,

4   1999), EPA reanalyzed these data from each hormone at each time point (Day 14, Day 90, and

5   Day 120) with two-way ANOVA tests. Gender and treatment (dose) were used as independent

6   between-subject variables. Dependent variables were T3, T4, and TSH. Step-down ANOVA

7   tests were conducted as indicated by significant interactions (Crofton and Marcus, 2001; Marcus,

8   2001). Mean contrasts were performed using Duncan's Multiple Range Test.

9   Results of the EPA reanalyses, shown in Table 5-2 and illustrated in Figures 5-5 through

10   5-7, are similar to those stated in the contract report (Springborn Laboratories, Inc., 1998) with a

11   few notable exceptions. First, there is only a marginal interaction between gender and treatment,

12   resulting from a slight difference in magnitude of effects between genders. However, no

13   differences in LOAELs between genders were observed (with minor exceptions likely caused by

14   small changes in variance between groups, which are probably not biologically significant [see

15   below]). Results of the analyses for each thyroid hormone and TSH are discussed individually.

16   There were significant day-by-gender-by-treatment interactions for T3 on Day 14 and

17   Day 90. Therefore, separate ANOVA tests were conducted on each gender to test for a main

18   effect of treatment. Lack of a significant gender-by-treatment interaction on the 120-day data led

19   to one subsequent ANOVA to test for a main effect of treatment. Data from Day 14 revealed a

20   LOAEL of 0.01 mg/kg-day for males (see Figure 5-5). There was a NOAEL of 10 mg/kg-day for

21   T3 in females. The low potency of perchlorate on T3 in females at the 14-day time point may be

22   artifactual. Not plotted on the figure for Day 14 are all the available data from control female

23   rats from this laboratory, including the Day 90 and Day 120 time points, and the data from two

24   other studies. These historical data show that the group mean for females in Figure 5-5 for the

25   14-day time point may be artificially low relative to some of the other data from the AFRL/HEST

26   laboratory. Thus, the biological significance of this gender-dependent effect of perchlorate after

27   14-days of exposure is suspect. Consistent with this conclusion is the significant dose-dependent

28   decrease in T3 concentrations in female rats exposed to 0.125 to 250 mg/kg-day perchlorate in a

29   previous 14-day exposure study by this same laboratory (Caldwell et al., 1995). The LOAEL for

30   effects on T3 for both males and females was 0.01 on Day 90. The NOAEL for effects on T3 at
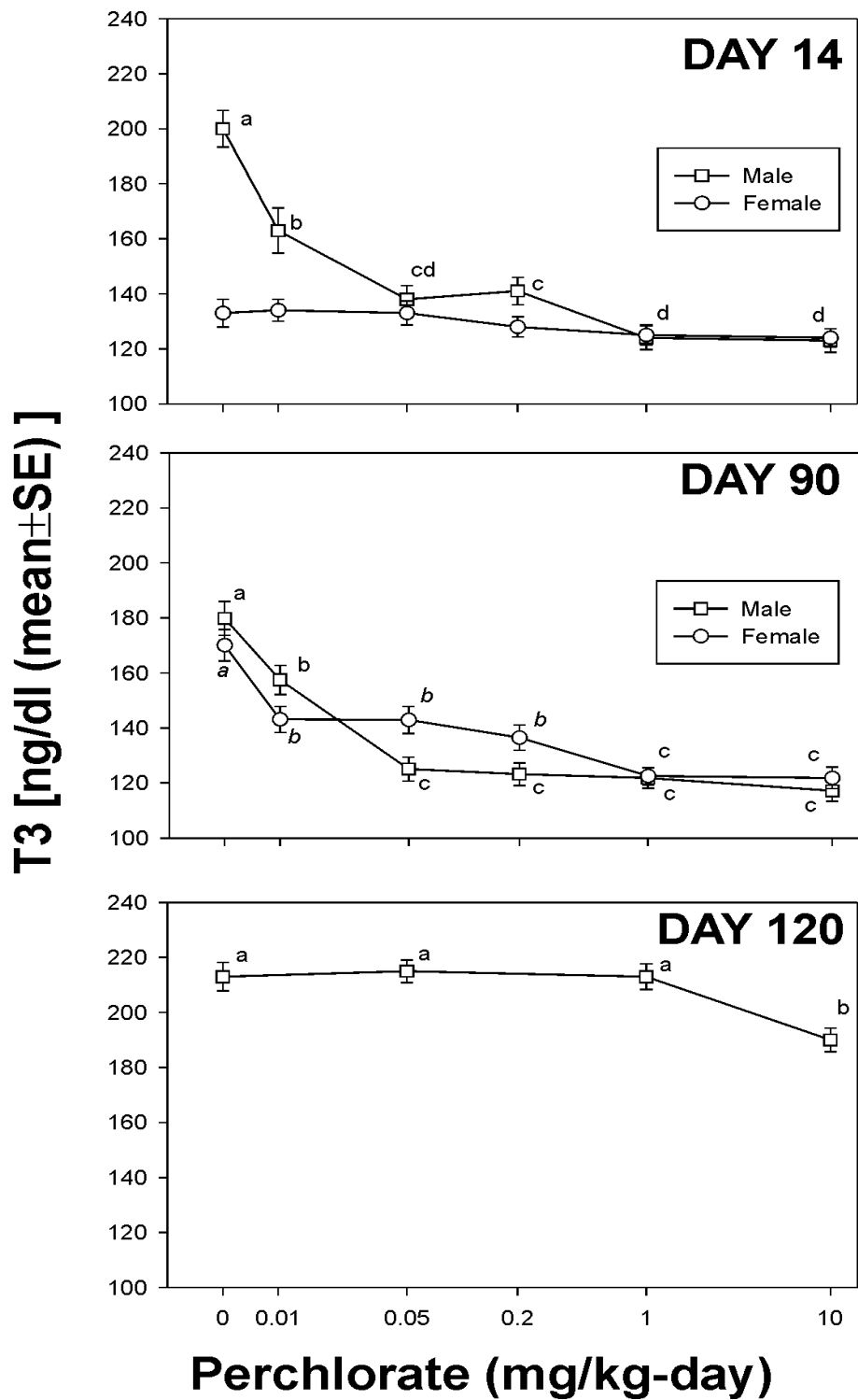
Figure 5-5. Effects from 90-day drinking water administration of ammonium perchlorate to SD rats on serum total T3 concentrations as recalculated in Table 5-2 (Crofton and Marcus, 2001). Means with different letters were statistically different ($p < 0.05$). The 120-day time point is 30 days after cessation of exposure.
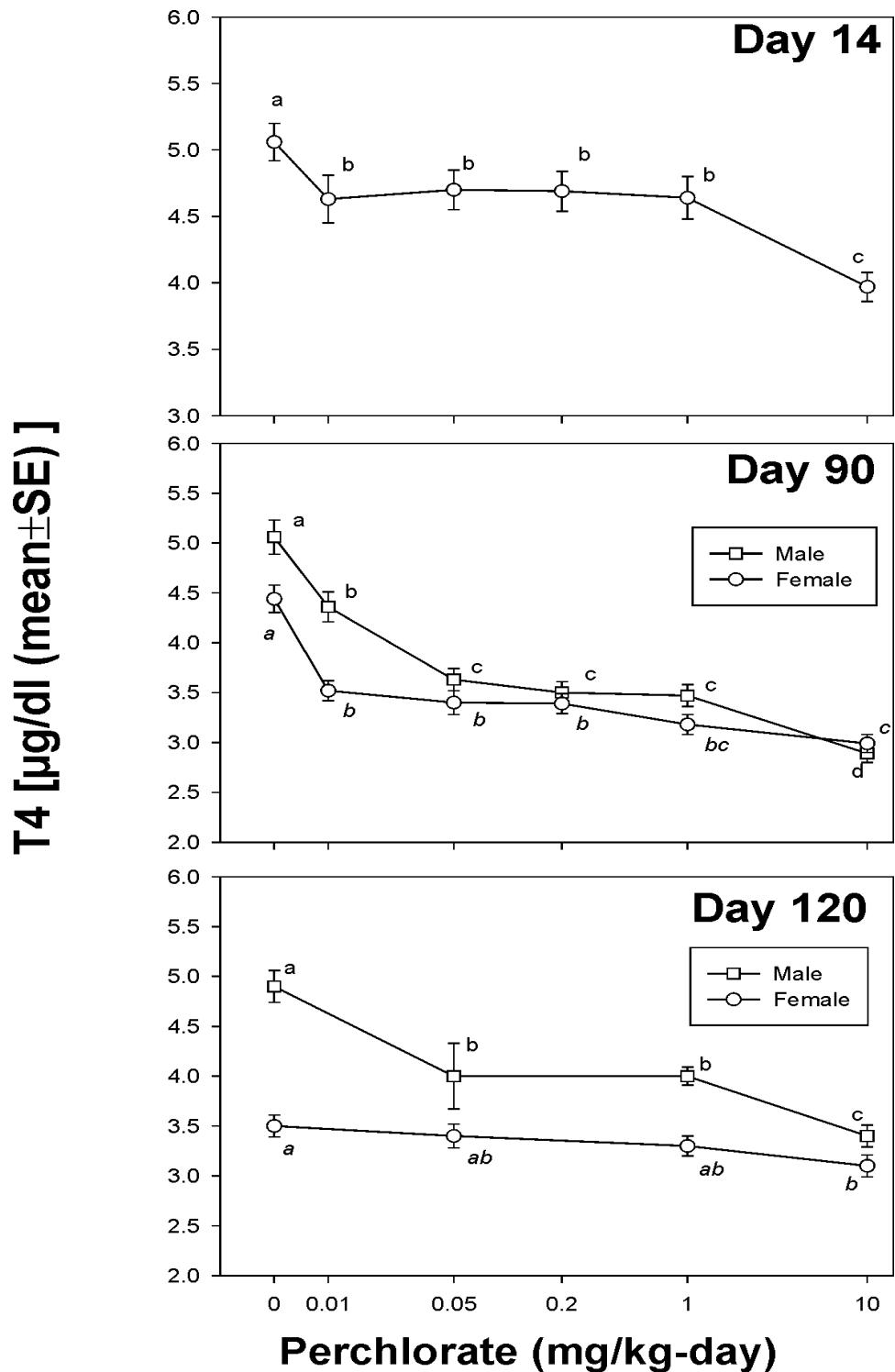
Figure 5-6. Effects from 90-day drinking water administration of ammonium perchlorate to SD rats on serum total T4 concentrations as recalculated in Table 5-2 (Crofton and Marcus, 2001). Means with different letters were significantly different (p < 0.05). The 120-day time point is 30 days after cessation of exposure.
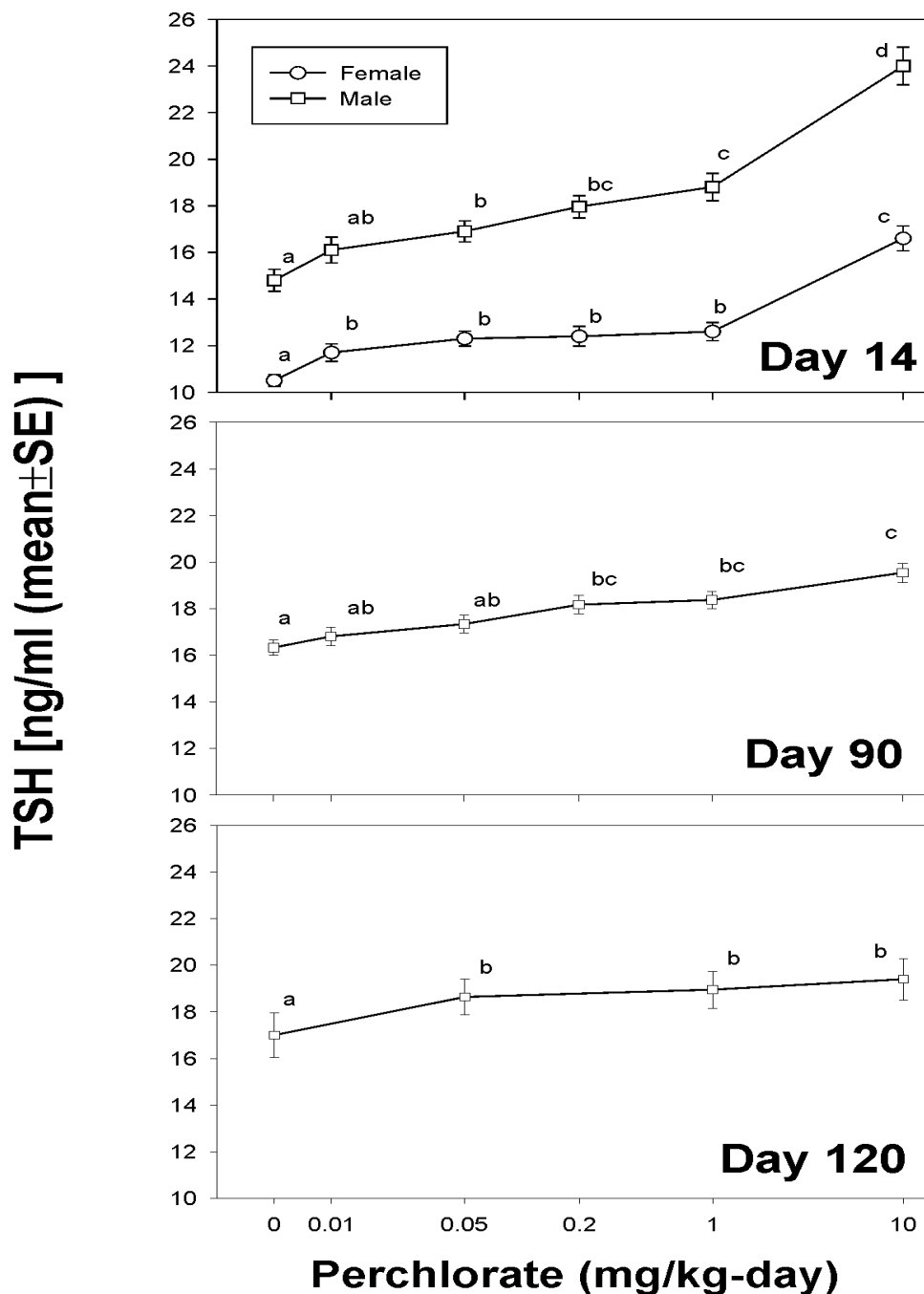
Figure 5-7. Effects from 90-day drinking water administration of ammonium perchlorate to SD rats on serum total TSH as recalculated in Table 5-2 (Crofton and Marcus, 2001). Data of Springborn Laboratories, Inc. (1998). A main gender-by-treatment interaction was observed for Day 14, but not Days 90 and 120; therefore, data are presented separately for males and females on Day 14 and collapsed across gender for Days 90 and 120. Means with different letters were significantly different ($p < 0.05$). The 120-day time point is 30 days after cessation of exposure.

1   Day 120 was 10 mg/kg-day, indicative of a recovery of T3 concentrations after cessation of

2   treatment.

3   There were significant day-by-treatment interactions for effects on T4 at the 90- and

4   120-day time points but not at the 14-day time point. Mean contrast tests for Day 14 data

5   revealed a free-standing LOAEL of 0.01 mg/kg-day for effects on T4 in both sexes. The

6   0.01 mg/kg-day dosage was also a free-standing LOAEL on Day 90 for effects on T4 in both

7   sexes. Analysis of the data from the 30-day recovery period (the Day 120 time point) revealed a

8   free-standing LOAEL of 0.05 mg/kg-day in males and a NOAEL of 1.0 mg/kg-day in females for

9   effects on T4.

10   There was a significant day-by-gender-by-treatment interaction for TSH only on Day 14.

11   Therefore, separate ANOVA tests were conducted on each gender to test for a main effect of

12   treatment for the Day 14 time point. Lack of a significant gender-by-treatment interaction for the

13   90- and 120-day data led to subsequent one-way ANOVA tests at each time point to test for a

14   main effect of treatment. Perchlorate caused a dose-dependent increase in TSH that was apparent

15   at the Day 14 and Day 90 time points (see Figure 5-7). The NOAEL for effects on TSH at

16   Day 14 data was 0.01 mg/kg-day in the males. The 0.01 mg/kg-day dose was a free-standing

17   LOAEL in the females. This small difference between males and females likely is caused by

18   small changes in variance between groups rather than by a biologically significant difference (the

19   absolute increase relative to the control mean in the 0.05-mg/kg-day female group is actually

20   smaller than the same comparison in the males). The TSH concentrations did not recover to

21   control values 30 days after cessation of treatment with a free-standing LOAEL at 0.05 mg/kg-

22   day in both sexes.

23   The data demonstrate a dose- and time-dependent effect of perchlorate on thyroid hormones

24   and TSH. There was no LOAEL established in this data set due to multiple effects at the lowest

25   dose of 0.01 mg/kg-day. There was some evidence of recovery at the Day 120-evaluation

26   (30 days after cessation of treatment). The NOAEL for effects on T3 increased to 1.0 mg/kg-day.

27   However, the omission of the 0.01 mg/kg-day dose group at the 120-day time point make it

28   difficult to conclude about a recovery of effects on T4 and TSH.

29

30

1 ## 5.3 DEVELOPMENTAL NEUROTOXICITY STUDIES

2 Concern for potential neurodevelopmental sequelae was warranted given the established

3 mode of action for perchlorate, and the original 1997 testing strategy included a developmental

4 neurotoxicity study (Argus Research Laboratories, Inc., 1998a). Results of that study raised

5 additional issues and concerns so that the external peer review convened in 1999 recommended

6 additional testing. This section describes results of the available studies that tested

7 neurodevelopmental indices per se. The 1998 neurodevelopmental study is reviewed in

8 Section 5.3.1. Results of the new study on motor activity are reviewed in Section 5.3.2. The

9 "Effects Study" repeated the study of brain morphometry as a measure of neurodevelopmental

10 toxicity and is reviewed in Section 5.3.3.

11

12 ### 5.3.1 The 1998 Developmental Neurotoxicity Study

13 The neurobehavioral developmental study of ammonium perchlorate that was part of the

14 original 1997 testing strategy was performed by drinking water administration in Sprague-

15 Dawley rats (Argus Research Laboratories, Inc., 1998a). A schematic of this study design is

16 provided as Figure A-1 (Appendix A) of this document to aid understanding of terminology and

17 the protocol. It should be noted that Argus Laboratories identifies the day of birth as PND1;

18 therefore, the age of PND10 and PND22 actually correspond to PND9 and PND21 in this study.

19 The description of the study design will use the Argus nomenclature in order to readily compare

20 with the contract report. Subsequent supplemental data submittals and additional analyses

21 pertaining to this were requested by EPA and provided by Argus Laboratories study (York,

22 1998a,b,c,d,e).

23 Female rats (25/dosage group) were administered target doses of 0, 0.1, 1.0, 3.0, and

24 10 mg/kg-day by continual access to ammonium perchlorate in nonchlorinated RO deionized

25 water beginning on gestation day zero (GD0) and ending at scheduled sacrifice. Test substance

26 concentrations were evaluated weekly, based on actual water consumption levels recorded the

27 previous week and adjusted as necessary to more closely achieve the target dose levels. Test

28 solutions were prepared weekly. The stability of the stock solution and that concentrations

29 agreed well with nominal concentrations were determined by AFRL/HEST (Argus Research

1   Laboratories, Inc., 1998a; Appendix J). Feed and water consumption were recorded daily during

2   exposure.

3   After acclimation for 14 days, virgin female rats were cohabited with breeder male rats

4   (one male rat per female rat) for a maximum of 7 days. Female rats with spermatozoa observed

5   in a vaginal smear or a copulatory plug observed in situ were considered to be at GD0. The

6   F0-generation dams were examined at approximately the same time each day during the exposure

7   period for signs of maternal behavior, autonomic dysfunction, abnormal postures, abnormal

8   movements or behavior patterns, and unusual appearance. Pregnancy outcome measures

9   evaluated at birth included pregnancy rate, duration of gestation, number of implantation sites,

10   gestation index (number with live pups/number pregnant), number of pups/litter, sex ratio of

11   pups, and viability and lactation indices. Maternal body weight was recorded on GD0, daily

12   during the exposure period, weekly during the post-weaning period, and at sacrifice. The same

13   set of signs as examined during exposure were evaluated on a weekly basis during post-weaning.

14   Thyroids from all F0-generation rats were weighed and evaluated histologically. Five dams per

15   group were selected for sacrifice and blood collection on post-natal day 10 (PND10) from those

16   with no surviving pups or with litters of less than eight pups. Thyroid and pituitary hormone

17   analyses (T3, T4, and TSH) were done on the blood (see Section 5.3.1.3). All dams not selected

18   for continued observation were sacrificed on PND22.

19   Pups (F1-generation) were counted and clinical signs were recorded once daily during

20   pre-and post-weaning. Body weight was recorded on PNDs 1, 5, 8, 12, 14, 18, and 22 and then

21   weekly during post-weaning. Feed consumption values were recorded weekly during

22   post-weaning. Pups that appeared stillborn and those that died before initial examination on

23   PND1 were examined for vital status, and the gross lesions were preserved. Pups that were not

24   selected for continued observation were sacrificed and necropsied on PND5. Blood was sampled

25   for thyroid and pituitary hormone analysis, and the thyroids were examined histologically. The

26   F1-generation pups not selected for continued observation on PND10 (n = 102) were sacrificed

27   and examined for gross lesions. Post-weaning pups that were selected for continued observation

28   were given ammonium perchlorate in RO deionized water with chlorine (added at a maximum of

29   1.2 ppm as a bacteriostat).

30   Other pups (F1-generation) were assigned to four different subsets for additional

31   evaluations. The first male and female pup (1/sex/dose; total of 97 male and 100 female pups)

1   were assigned randomly to Subset 1 for brain weight and neurohistological examination

2   (including morphometric measurements). All pups were selected for fixed brain weights on

3   PND12; 6/sex/dose (total of 30 male and 30 female pups) were selected for neurohistological

4   examination. The second male and female pup (1/sex/dose; total of 100 male and 100 female

5   pups) were assigned randomly to Subset 2 for passive avoidance testing on PNDs 23 to 25 and

6   PNDs 30 to 32; water maze testing on PNDs 59 to 63 and PNDs 66 to70; and scheduled sacrifice

7   at PNDs 90 to 92, with blood collection for thyroid and pituitary hormone analysis. The third

8   male and female pup (1/sex/dose; total of 100 male and 100 female pups) were assigned

9   randomly to Subset 3 for motor activity evaluation on PNDs 14, 18, 22, and 59; auditory startle

10   habituation on PNDs 23 and 60; and scheduled sacrifice on PNDs 67 to 69. The fourth male and

11   female pup (1/sex/dose; total of 100 male and 100 female pups) were assigned randomly to

12   Subset 4 for regional brain weight evaluation on PNDs 81 to 86 (6/sex/dose; total of 30 male and

13   30 female rats) and neurohistological examination on PNDs 82 to 85 (6/sex/dose; total of

14   30 male and 30 female rats). Female pups also were evaluated for the age of vaginal patency

15   beginning on PND28, and male pups were evaluated for the age of preputial separation beginning

16   on PND39. A few of these measurements inadvertently went unrecorded, but the laboratory

17   asserted that this did not affect the results because a sufficient amount of data on other rats was

18   recorded.

19

20   **5.3.1.1   Results of General Toxicity Measures, Neurohistology, and Morphology**

21   Results in the dams (F0-Generation) revealed no treatment-related effects on food or water

22   consumption (Argus Research Laboratories, Inc., 1998a; Appendix B, Tables B7 through B14),

23   mortality (Appendix B, Tables B2 and B18), clinical signs (Appendix B, Table B2), necropsy

24   (Appendix B, Table B18), body weight (Appendixes A and B, Figure A1 and Tables B3 through

25   B6), or pregnancy outcome measures (Appendix B, Tables B15 through B16). Effects on thyroid

26   weight, histopathology, and thyroid and pituitary hormone analyses will be discussed below in

27   Sections 5.3.1.2 and 5.3.1.3.

28   Results in the pups (F1-generation) revealed no treatment-related effects on feed

29   consumption (Argus Research Laboratories, Inc., 1998a; Appendix C, Tables C18 and C19),

30   mortality (Appendix C, Tables C1 and C2), clinical signs (Appendix C, Tables C1 and C2), body

31   weight (Appendixes A and C, Figures A2 and A3 and Tables C3 through C6), or sexual

1   development landmarks (Appendix C, Table C11). No treatment-related effects were observed

2   on mortality, brain weight, or body weight in the pups of Subset 1 at PND12 (Argus Research

3   Laboratories, Inc., 1998a; Tables D1 and D2), Subset 2 at PNDs 90 to 92 (Tables E3 and E4), or

4   Subset 3 at PNDs 67 to 69 (Tables F5 and F6). Results of the neurobehavioral tests from

5   Subsets 2 and 3 will be discussed in Section 5.3.1.4.

6   In the Subset 1 subgroup subjected to neurohistological examination (the F1 pups sacrificed

7   on PND12), morphometric analyses revealed a 23.4% increase in the size of the corpus callosum

8   in females and a 30.2% increase in males (not significant) at the high dose (10 mg/kg-day).

9   Slight decreases in brain weight also were noted at the highest dose in females. In Subset 4 (the

10 F1 pups sacrificed on PND82), there was a continued effect on the size of the corpus callosum

11 (20.9% increase) in males, but no effect in females at the highest dose. There was also a 3.4%

12 increase in the brain weight in males and increases in the size of the frontal cortex (9.2%) and the

13 caudate putamen (10.2%). The EPA concluded that the effects may be significant and that

14 analyses of the next lower dose (or, at least, historical control data for the affected endpoints)

15 were warranted and requested additional analyses from the sponsor (PSG). York (1998d)

16 responded with morphometry analyses of the next lower dose (3.0 mg/kg-day) of the Subset 1

17 F1 pups at PND12. The new analysis noted, in addition to previous findings, a statistically

18 significant increase in the anterior/posterior cerebellum size, a statistically significant decrease in

19 the caudate putamen for the F1 PND12 female pups, and a statistical significant decrease in the

20 hippocampal gyrus size for the F1 PND12 male pups. These effects were not considered

21 treatment-related by the Primedica/Argus pathologist because they were not dose dependent.

22   A preliminary reanalysis by EPA (Crofton, 1998c) of the control, 3- and 10-mg/kg-day

23 groups (York 1998d) was restricted to the corpus callosum because this was the area with the

24 largest effect. The analysis revealed no interaction of gender and treatment; however, there was a

25 significant effect of treatment ($F[2,30] = 7.65$, $p < 0.0021$). There was a significant increase in

26 the size of the corpus callosum only in the 10-mg/kg-day group. Group means were 288, 278,

27 and 366 μm for the controls and 3- and 10-mg/kg-day groups, respectively. Incorporation of

28 historical control data from both PND10 and PND12 (mean for controls = 264 μm for PNDs 10

29 and 265 μm for PND12; York, 1998a) supports the conclusion that the control values for corpus

30 callosum size in the data set are within the "normal" range (York 1998a; see also Argus Research

31 Laboratories, Inc., 1998a).

1   EPA did not agree with the argument put forth by Argus Research Laboratories, Inc.

2   (1998a) that these effects were "not suggestive of a neurotoxic effect" because of "an unknown

3   biological significance." EPA considers a 27% increase in the size of any brain region to be a

4   potentially adverse effect (U.S. Environmental Protection Agency, 1998e), and designated

5   10 mg/kg-day as the LOAEL and the NOAEL at 3 mg/kg-day for these changes in brain

6   histology. No additional evaluation of the brains from the neurohistological examination of

7   Subset 4 pups (PND82 to PND85) were ever submitted to EPA although it was suggested again

8   that the next lower dose group be analyzed because of the significant increases in brain weights

9   and in the frontal cortex and corpus callosum measurements for the males in the high-dose group.

10   Additional analyses of the brain morphometry were provided by the EPA at the 1999

11   external peer review (Geller, 1999a) that corroborated the preliminary finding of Crofton

12   (1998c). The data were analyzed using a 2-way ANOVA, with dose and sex as independent

13   variables. To correct for multiple comparisons, the acceptable alpha for significance (for all

14   interaction main effects) was corrected to 0.016 (alpha of 0.05 divided by the square root of the

15   number of ANOVA tests).

16   Significant effects of dose were found in corpus collosum, hippoacampal gyrus, anterior

17   and posterior cerebellum, and caudate putamen. An effect of sex was also found in caudate

18   putamen. The effect on corpus callosum was confirmed and showed an increase in size at the

19   10 mg/kg-day dose. Hippocampal gyrus (12% less than control) and caudate putamen (7.3% less

20   than control) showed a decrease in size at the 3 mg/kg-day dose, with no significant difference

21   between control and high dose, yielding a U-shaped dose response. The anterior and posterior

22   cerebellum showed a significant increase in size at the 3 mg/kg-day group (13%).

23   Because of concern for this effect voiced at the 1999 external peer review, the blocks of

24   brain tissue were evaluated to determine if they could be refaced and additional sections

25   evaluated. It was determined that the remaining materials were of insufficient quality for

26   additional sectioning and histological evaluation (Harry, 2001). As an alternative, brain

27   morphometry measurements were included in the "Effects Study", described below in

28   Section 5.3.3, to determine if the alteration in brain morphometry could be repeated.

29

30

31

5.3.1.2   Evaluation of Thyroid Histopathology

Appendix O of the Argus Research Laboratories, Inc. (1998a) neurodevelopmental study presents thyroid histopathology data provided by the sponsor (AFRL/HEST). Note that the data analyzed by EPA in the 1998 document for PND5 F1-generation rat pups are from the final report for the PND5 time point (Channel, 1998c). Channel (1998c) reported that the decrease in follicular lumen area in these pups at PND90 to PND92 showed no significant differences between dose groups and controls for either females or males based on t-test or Mann-Whitney Rank Sum Test (M-W RST). These data suggest a recovery from the effects observed in the thyroids of the pups at PND5.

The report also contained measurements, performed by Dr. William Baker of AFRL/HEST, of both follicular epithelial cell height and the follicular lumen diameter. These data were subsequently formally transmitted to EPA by consultative letter (Channel, 1998c) in Microsoft Excel® spreadsheets. For the final morphometric study (Channel, 1998c), the arbitrary decision based on ease of detection of this region in digitized images was made by Dr. William Baker to focus on only a lumen area measurement because of time constraints (Jarabek, 1998). The mean follicular lumen area represents the mean area of all follicular lumens measured from the three histological sections sampled from each rat and is expressed in microns. In the opinion of Dr. Charles Capen of Ohio State University (Crofton, 1998d), the measurement of follicular height is usually more sensitive than those of follicle diameter and lumen area. In support of this opinion, data collected by Dr. Baker (Argus Research Laboratories, Inc., 1998a; Appendix O) demonstrated significant increases in males rats in the incidence of follicular epithelial cell hypertrophy at doses much lower than those doses that increased the incidence of decreased lumen area. The difference observed between standard histopathology as originally reported by Argus Research Laboratories, Inc. (1998a) and the thyroid morphometry performed by Dr. Baker was analyzed extensively by the EPA in its 1998 assessment. The results indicated that the morphometry performed on lumen size was a less sensitive measure of thyroid histopathology. The analyses of the thyroid morphometry are retained in this reassessment; whereas, the PWG review results will be presented below for the histopathology.

Data from the dependent measure (follicle lumen size) based on the morphometric analyses (Channel, 1998c) were available for pups sacrificed at ages PND5 and PND90. These data were reanalyzed by EPA (Crofton and Marcus, 2001). Because there was only one block of animals at

1 PND90 compared to two blocks of data at PND5, and because the slides for PND90 were

2 processed at a much later time, the data for the two ages were analyzed separately. Data from

3 PND5 pups were subjected to three-way ANOVA tests with gender, treatment (dose), and block

4 (two separate analyses of separate blocks of data) as independent between-subjects variables.

5 Data from PND90 were subjects to a two-way ANOVA with gender and treatment (dose) as

6 independent between-subjects variables. Step-down ANOVA tests were conducted as indicated

7 by significant interactions and recalculated by Crofton and Marcus (2001) and Marcus (2001).

8 Mean contrasts were performed using Duncan's Multiple Range Test. Note that in the Crofton

9 and Marcus (2001) memorandum the 0.1 mg/kg-day dose is incorrectly labeled as 0.3 mg/kg-day.

10 There was a significant main effect of treatment on lumen size for all doses at PND5, resulting in

11 a free-standing LOAEL of 0.1 mg/kg-day. The data are plotted in Figure 5-8. There was no

12 significant effect of perchlorate on lumen size at PND90.

13   The thyroid histopathology as reviewed and reported by the PWG can be found in Wolf

14 (2001; Tables 9 through 13). This report includes corrections for slides sent to EPA that

15 contained animals with autolysis and those necropsied at different times than indicated for the

16 study protocol or to exclude dams that did not have litters.

17   The F0 generation dams (Wolf, 2001: Table 9) exhibited decreased colloid and increases

18 in both hypertrophy and hyperplasia. A clear dose-response was not evident, however, with the

19 possible exception of colloid depletion at levels above 0.1 mg/kg-day.

20   Thyroid histopathology in the pups on PND4 (Wolf, 2001: Tables 10 and 11) was more

21 pronounced, with colloid depletion and increases in hypertrophy at 0.1 and 3 mg/kg-day.

22 Hyperplasia appeared to be effected at 3 mg/kg-day. The BMD analyses presented in Table 5-1

23 support these levels with BMDL estimates for colloid depletion at 0.33, increased hypertrophy at

24 0.88, and increased hyperplasia at 3.62 mg/kg-day. These results were obtained with a

25 constrained model, but an adequate fit is obtained by fitting the model without restricting the

26 exponent on dose to be ≥ 1 and results in a BMDL for pups on PND4 in this study at 0.009 for

27 colloid depletion (Geller, 2001a).

28   The argument for the lack of biological plausibility of unrestricted functions is based on

29 cancer modeling theory from the early 1960s (Mantel and Bryan, 1961) that attempted to derive a

30 default procedure for modeling tumor data at the time when cancer was thought to be a one-stage

31 process and many bioassays used only 1 dose and control. Given the increased sophistication of
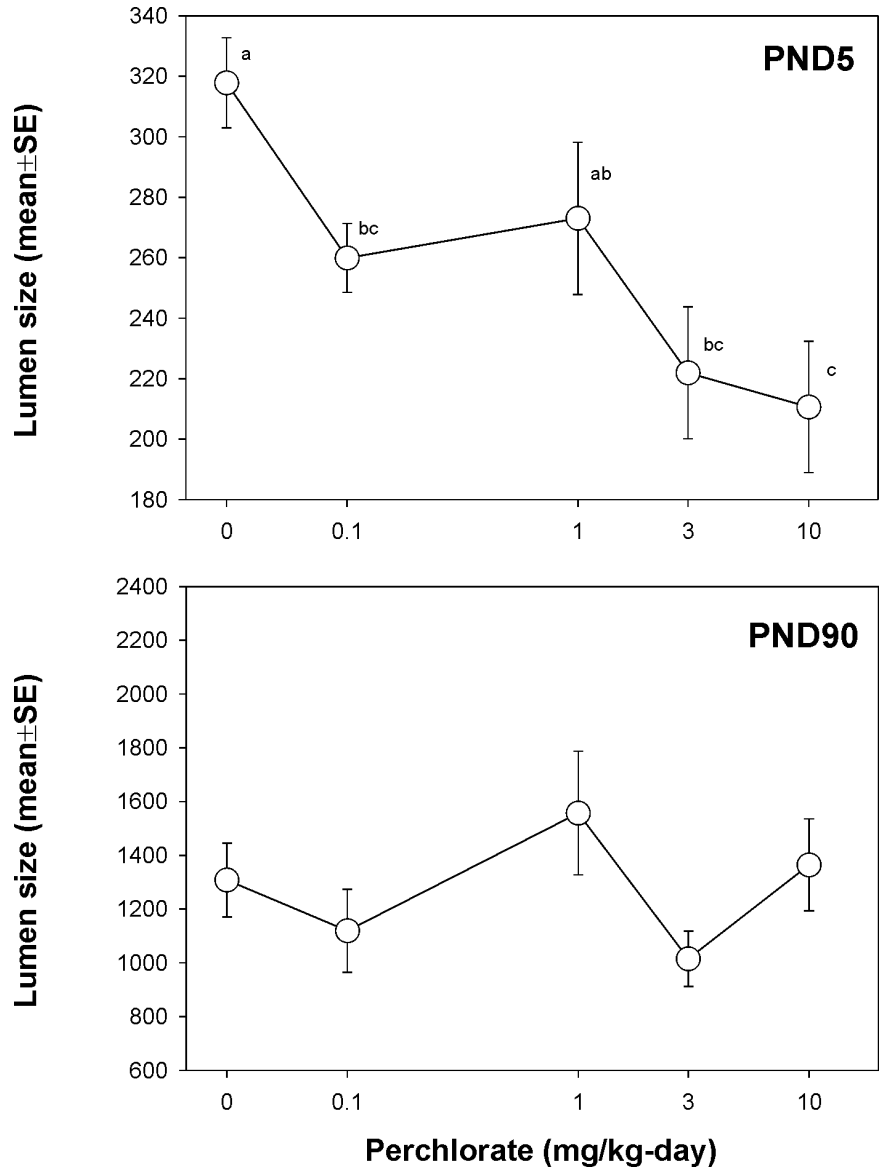
Figure 5-8. Effects from maternal drinking water administration of ammonium perchlorate to SD rats on thyroid gland follicular lumen size in F1-generation offspring on PND5 as recalculated in Crofton and Marcus (2001). Data of Channel (1998c) and Argus Research Laboratories, Inc. (1998a). Means with different letters were significantly different (p < 0.05). Daily dose was estimated from water consumption data.

1   contemporary bioassays and the level of organization at which effects are now being identified

2   (i.e., precursor events at the cellular and molecular levels), Hasselblad et al. (1995) have argued

3   that restricting the slopes of fits to the data prioritizes mathematical convenience over fitting the

4   data. The thyroid hormone data show exquisite sensitivity to very low doses of perchlorate. This

5   suggests that models fit with nonsupralinear slopes and lower doses need to be tested. It is

6   interesting to note that PWG results for colloid depletion are very similar to the 1998 EPA

7   analysis on the previous histopathological read by Argus Laboratories, Inc. (1998a) for

8   hypertrophy/hyperplasia that resulted in a BMDL of 0.1 mg/kg-day.

9   Histopathology in the animals from PND90 and PND92 (Wolf, 2001: Tables 12 and 13)

10   indicated variable effects on colloid depletion, hypertrophy, and hyperplasia. As indicated in

11   Table 5-1, a BMDL was only calculated with confidence for colloid depletion with a resultant

12   estimate of 0.03 mg/kg-day.

13   Evaluation of the histopathology in this study indicate that the pups are the most sensitive

14   with a BMDL between 0.009 and 0.33 mg/kg-day.

15

5.3.1.3   Thyroid and Pituitary Hormone Analyses

17   Serum was collected and thyroid hormone analyses performed as part of the

18   neurodevelopmental study (Argus Research Laboratories, Inc., 1998a; Crofton, 1998f)). The

19   following is a statistical analysis of the thyroid and pituitary hormone data (T4, T3, and TSH)

20   found in that report (Crofton and Marcus, 2001). At the time of this assessment, individual

21   animal data were available from both the F1-generation pups (male and female samples were

22   pooled for each litter) on PND5 and the F0 generation (parents) on post-partum Day 10 (PP10).

23   Only the F1 data were reanalyzed because of the very limited (n = 2 to 5/group) data for the

24   parental F0 PP10 group.

25   All data were supplied in Microsoft Excel® spreadsheets via E-mail by Dr. David Mattie

26   (AFRL/HEST). Data for dependent measures (T4, T3, and TSH) were subjected to separate one-

27   way ANOVA tests. Treatment (dose) was used as the independent, between-subjects variable.

28   Mean contrasts were performed using Duncan's Multiple Range Test.

29   There were significant main effects of treatment for all the hormones. The data are plotted

30   in Figure 5-9. Results of these reanalyses are similar to those stated in the report (Argus

31   Research Laboratories, Inc., 1998a). There was a significant decrease in both T3 and T4, as well
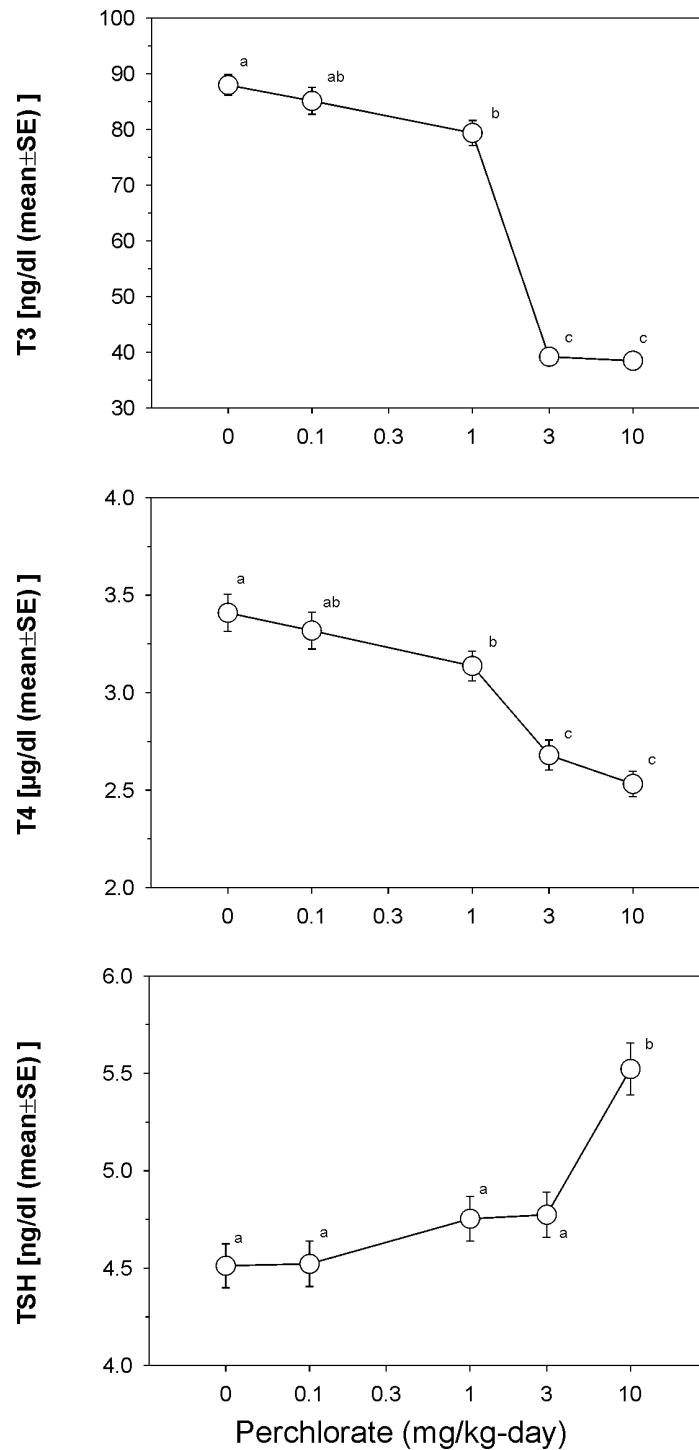
Figure 5-9. **Effects from maternal drinking water administration of ammonium perchlorate to SD rat F1-generation pups on serum total T3 (A), T4 (B) and TSH (C) concentrations (ng/dL; mean ± SE) as recalculated in Table 5-2 (Crofton and Marcus, 2001). Data of Argus Research Laboratories, Inc. (1998a). Means with different letters were significantly different (p<0.05). Daily dose was estimated from water consumption data.**

as the expected increase in TSH. The NOAEL for the effects of perchlorate on T3, T4, and TSH are 0.1, 0.1, and 3.0 mg/kg-day, respectively. These results are consistent with the known mechanism-of-action of perchlorate (inhibition of thyroid hormones). The increased TSH is likely a result of the activation of the pituitary-thyroid feedback mechanism.

5.3.1.4 Behavioral Evaluations

The 1998 EPA review of the behavioral evaluations performed on Subset 3 pups agreed with the Argus Research Laboratories, Inc. (1998a) report with one exception regarding an increase in motor activity in male rats on PND14 that no perchlorate-induced changes were detected in any of the other behavioral indices (i.e., passive avoidance, water maze, auditory startle). The EPA disagreed with the Argus Research Laboratories, Inc. (1998a) report and subsequent submissions (York, 1998a,b,c,d,e) with regard to the significance of the motor activity changes.

The data originally were analyzed using two separate three-way ANOVA tests (age, treatment, and habituation block), one for each gender (Argus Research Laboratories, Inc., 1998a). This analysis demonstrated a significant decrease in the amount of habituation in the two highest dose groups on PND14 in the male pups. There were no changes detected at any other ages (i.e., PND18, PND22, PND59). On initial review by EPA, it was recommended to the sponsor (PSG) that an additional analysis of the data be conducted using gender as a within-subject variable, or alternatively, to use a nested design with gender nested under litter (see Holson and Pearce [1992] and Cox [1994], for a review of statistical methods used in developmental studies and the importance of using litter as the unit of measure). The EPA also questioned why the method or statistics did not detect significance for the dose-dependent increase in total session counts that amounted to a 95% increase over controls in the highest dosage group (see Figure 5-10). The response from Argus Laboratory (York, 1998b) included a new analysis in which gender was used as a between-subjects variable. No interactions with, or main effects of, treatment were found in this analysis.

EPA remained concerned that Argus Research Laboratory and the sponsor (PSG) failed to respond adequately to the request for an explanation of why the analysis failed to detect significance in the PND14 motor activity for the male rats. Figure 5-10 illustrates the clear dose-dependent increase in two different measurements of motor activity: (1) time-spent-in-
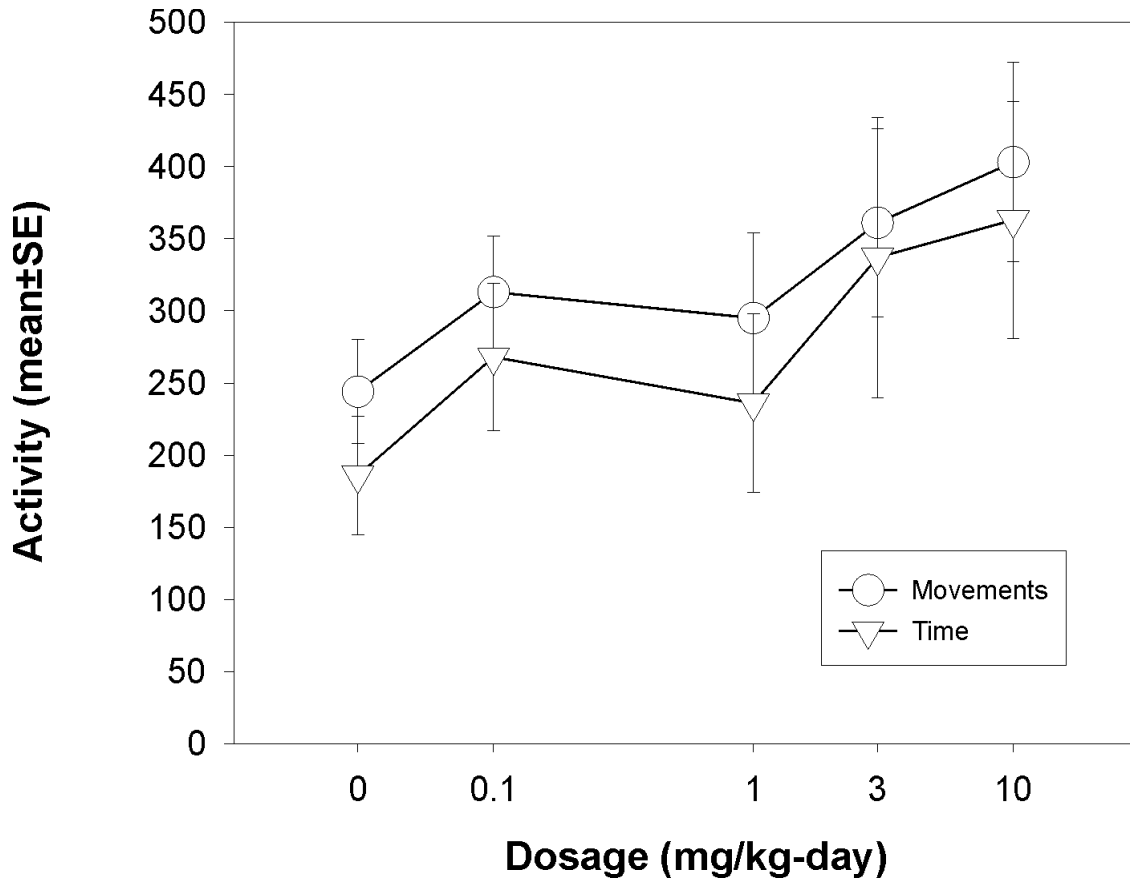
Figure 5-10. The effects of developmental exposure to perchlorate on motor activity in male rats on PND14. Data of Argus Research Laboratories, Inc. (1998a). The dose-dependent increases in both number of movements and time spent in movement were not statistically different, even though the increases were substantial at the higher dosages.

1   movement ("time") and (2) total number of movements ("movements"). The time variable

2   increased over 95% at the highest dose relative to controls (group means of 363 and 186,

3   respectively). The number-of-movements variable increased approximately 65% relative to

4   controls. Expert opinion of EPA neurotoxicologists was sought, and it was their opinion that

5   increases in motor activity over 50%, especially in developing animals, were clearly of concern

6   from a biological perspective (Crofton et. al., 1998). The critical issue for evaluation of these

7   motor activity data was how to resolve the difference between what is a clearly a biologically

8   significant alteration in behavior with a lack of statistical significance. In an attempt to resolve

9   the issue, EPA also requested positive control data from the testing laboratory for this device that

1   was not provided in the original report, as well as any available historical control data. York

2   (1998a) replied with a number of positive control studies and a limited amount of historical

3   control data from PND14 pups.

4   The positive control data were requested to help understand the sensitivity of the device in

5   detecting increases in motor activity (i.e., what is the smallest increase in motor activity that has

6   been detected by this device). Unfortunately, the positive control data were of limited use in

7   interpreting the sensitivity of the device. The submission (York, 1998a) contained data from

8   experiments with amphetamine and triadimefon in adult rats. The smallest increase in activity

9   that was induced by either chemical was a 109% increase relative to controls. Although these

10   effects were statistically significant, they are greater than the effects produced by the highest

11   dosage of perchlorate in the PND14 pups. There were also positive control data from

12   chlorpromazine-treated animals that showed significant decreases ($\geq$32%) in activity. However,

13   ability to detect decreases does not necessarily translate to the detection of increases.

14   The historical control data from PND14 rats were requested to help understand the

15   variability normally found in control animals. Unfortunately, the historical control data

16   submitted were only useful in that the data raised more suspicion that the degree of experimental

17   control over this behavior by the testing facility was inadequate. For the time data, the control

18   mean for the perchlorate data set was 186 sec. For the three relevant historical control data sets,

19   the means were 1026, 965, and 458 sec. Either the lab had very little control over the behavior,

20   or the data were from a different test apparatus or from a different usage of the same apparatus.

21   In any case, the data were of no use in helping EPA determine the historical profile of control

22   animal behavior in this test apparatus.

23   In lieu of the absence of useful positive control and historical control data, EPA was left

24   with the issue of ascertaining statistical versus biological significance. There were a number of

25   reasons for the lack of statistical significance. The first reason was the extremely large within-

26   group variability exemplified by coefficients of variation (CV) greater than 100%. It was the

27   opinion of Crofton et al. (1998) that this was likely caused by the inability of the testing

28   laboratory to gain adequate control over the behavior being tested. This large variability results

29   in very little statistical power and increases the potential for Type II errors. Normally, an

30   increase in sample size (by additional testing) allows for adequate power to refute or support the

31   conclusion of an effect. Given the CVs of about 100%, simple power calculations (see Cohen,

1987) for detecting a 40% change in one group out of five results in needed group sizes of about 70 to 90 animals per group. The second reason was that the effect, a 95% increase, while rather large from a biological perspective, occurs in only one gender on only 1 day out of 4 test days. The large variability coupled with the complicated design (treatment, age, gender, and block) would tend to mask anything other than extremely large effects. This conclusion is consistent with the content of a phone conversation (Crofton, 1998g) with Dr. Simon Mats. Dr. Mats was the statistician from the contract laboratory (Primedica/Argus) who conducted the revised statistical analysis of these data. Lastly, the effect seen in the males on PND14 may indeed be a Type I error and may not be found again if this experiment was repeated.

The assignment of biological significance to the effect seen was supported by both the underlying mode of action of perchlorate and the effects of other chemical and physical insults on the motor activity of post-natal rats. The hypothesis that a thyrotoxic chemical would induce a delay in any aspect of nervous system development is highly plausible. A delay in the onset of habituation would be evidenced by an increase in overall counts, as well as a decrease in the rate of a habituation (Ruppert et al., 1985a,b). This delay could be quite transient. Other agents that interfere with thyroid hormones during development are known to induce delays of a few days magnitude in developmental landmarks such as eye opening (Goldey et al., 1995a,b). This is the type of effect seen on PND14 in the Argus Research Laboratories, Inc. (1998a) report.

Developmental exposure to numerous hypothyroid-inducing agents (e.g., propylthiouracil, methimazole) are known to result in delays in the ontogeny in many behaviors (cf., Comer and Norton, 1982; Goldey et al., 1995a,b; Schneider and Golden, 1986; Tamasy et al., 1986), including the development of habituation. However, effects of these chemicals on total motor activity counts vary from increased to decreased, depending on the chemical and age of testing. Rice (2000) has noted parallels between the features of attention deficit hyperactivity disorder (ADHD) and the behavior of monkeys exposed to polychlorinated biphenyls (PCBs). The mechanism for the gender-dependent nature of the effect of perchlorate also remains to be determined. In addition, there are numerous reports from the literature that support the biological significance of a 40 to 50% increase in motor activity in postnatal rats (cf., Campbell et al., 1969; Ruppert et al., 1985a,b).

In summary, EPA maintained that the increase in activity should be considered biologically significant until additional data could be marshaled to suggest or prove otherwise. The

1   inadequacy of standard parametric statistics to detect a significant difference suggested that

2   alternative analyses should be used on these data, such as the benchmark approach. This type of

3   statistical approach may be useful because of the inverse relationship between the data variability

4   and the benchmark dose (BMD). The BMDL estimates were calculated for data on the

5   movement (number of movements) and time (time spent moving) measures from the motor

6   activity test from PND14 pups. These data were fit by a linear function with fairly shallow slope,

7   yielding BMD estimates for movement and time of 1.94 and 1.33 mg/kg-day and BMDL

8   estimates of 1.04 and 0.66 mg/kg-day, respectively. These BMD and BMDL estimates could

9   serve as estimates of LOAEL and NOAEL for this data set. The estimates are in accord with

10   doses with activity values that may have emerged as significantly different from control had the

11   data set not had its unusually high variability. These BMD analyses bring the motor activity

12   NOAEL more within the range of the T3 and T4 NOAEL and below that for TSH.

13

14   ## 5.3.2 Motor Activity Study (Bekkedal et al., 2000)

15   In response to recommendations at the 1999 peer review for an additional study, the United

16   States Navy (USN) performed a study that included evaluation of motor activity in Sprague

17   Dawley rats of both sexes (Bekkedal et al., 2000). Female Sprague-Dawley rats were dosed with

18   ammonium perchlorate for two weeks at 0, 0.1, 1.0, 3.0 or 10.0 mg/kg-day prior to mating with

19   the breeder males and through PND10. PND1 was counted as the day when the first pup was

20   observed in the cage. All pups within a litter were weighed on PND5 when the litters were

21   culled to eight pups of 4 males and 4 females or as close as possible to that combination. Pups

22   and dams from any litters with less than 8 pups were eliminated. On PND14, one male and one

23   female were randomly selected from each litter to be used in the motor activity testing. These

24   same animals were tested on PND14, PND18 and PND22. Nine different measures of motor

25   activity were automatically recorded using Opto-Varimex activity meters at ten minute intervals.

26   The measures included: frequency and time of ambulatory movements, frequency and time of

27   sterotypic movements, frequency of movements in the horizontal plane, distance traveled in the

28   horizontal plane, frequency of rears, total number of horizontal movements made while in the

29   rearing position (vertical plane movements), and time spent resting.

30   Bekkedal et al. (2000) analyzed each of the nine measures of motor activity separately

31   using a univariate repeated-measures ANOVA. The between subjects variable was perchlorate

1   dose, with 5 levels. The three within-subject variable were sex (2 levels), age (3 levels), and time

2   block (9 levels). Due to violation of the sphericity assumption, the Greenhouse-Geisser test was

3   employed with a fiducial limit set at p < 0.05. No statistically significant differences were found

4   for the main effect of perchlorate exposure for any of the 9 measures nor any reliable interactions

5   related to dose. The authors do note, however, a general pattern of dose-dependent changes in

6   the later sessions (90-minute). They also note that this pattern, as in the previous Argus

7   Laboratories, Inc. (1998a), suggest that exposed pups have a slightly slower rate of habituation

8   and thus maintain a higher level of activity as compared to untreated pups. Additional follow-up

9   tests were suggested.

10

11   **5.3.2.1   EPA and NIEHS Statistical Analyses of Motor Activity Effects**

12   Because EPA was concerned about effect on motor activity in the original study and it

13   appeared that a similar pattern of effects was again emerging in the study repeated by Bekkedal

14   et al. (2000), EPA requested that NIEHS perform a statistical evaluation that could formally

15   integrate the various measures together as well as statistically compare the two studies with each

16   other (Dunson, 2001a). A Bayesian hierarchical model (Gelfand et al., 1990) was chosen to

17   assess the weight of evidence of a dose-response trend in motor activity. A linear mixed-effects

18   regression model (Laird and Ware, 1982) related dose, sex, age, habituation time and a

19   habituation time x dose interaction term to the expected number of ambulatory movements, with

20   an animal-specific intercept included to account for within-animal dependency. To complete a

21   Bayesian specification of the model, a vague (or uninformative) but proper prior distributions for

22   each of the unknown parameters was chosen. In particular, the prior for the parameters that

23   related dose to motor activity was centered on a value corresponding to the null hypothesis of no

24   effect of perchlorate. The model was fit using BUGS, a widely-used software package for

25   Bayesian analyses (Gilks et al., 1994).

26   The analyses were conducted under a variety of different choices of prior variance for the

27   dose parameters and prior means and variances for the other parameters in the model. The dose

28   level associated with a 10% increase in the number of ambulatory movements by inverse

29   estimation (refer to Appendix A in Dunson, 2001a). The choice of 10% as the benchmark level

30   is consistent with standard practice for dichotomous outcomes.   The 5% level often used for

1   continuous outcomes was judged to be too low for measuring a biologically significant increase

2   in motor activity. Conclusions were consistent across the analyses.

3   As noted by Bekkedal et al. (2000), the effect of ammonium perchlorate on the number of

4   ambulatory movements was found to increase significantly with habituation time (posterior

5   probability = 0.98). In the first habituation interval there was modest evidence of an increase in

6   motor activity with dose (posterior probability = 0.79), while in the final interval there was clear

7   evidence of an increase in motor activity with dose (posterior probability > 0.99). The posterior

8   density for the expected increase in the logarithm of the number of ambulatory movements at the

9   final habituation time per unit (mg/kg-day) increase in dose of ammonium perchlorate is plotted

10   in Figure 5-11 for the USN study (Bekkedal et al., 2000). The posterior density is centered on a

11   positive slope and assigns low probability to a negative slope, suggesting a clear increase in

12   motor activity with dose. The dose estimated to increase the mean number of ambulatory

13   movements at the final habituation time by 10% is 1.62 with a 95% credible interval of (0.90,

14   7.87). There was no evidence of an interaction between age and dose, nor of any effect of

15   gender.

16   The previous study of Argus Laboratories, Inc. (1998a) was also analyzed in this fashion

17   and results were very similar (Figure 5-11). In contrast to the Bekkedal et al. (2000) study,

18   dosage began at the first day of gestation and continued through parturition and up to lactation

19   day 10 (PND10).   Dams were dosed at 0, 0.1, 1.0, 3.0 and 10.0 mg/kg-day. Movements of each

20   pup were monitored by a passive infrared sensor. Each test session was 90 minutes in duration.

21   The number and time spent in movement was tabulated at each five-minute interval. In order to

22   be comparable with the USN analysis, every two of the five-minute intervals were combined into

23   a ten-minute interval. However, the Bekkedal et al. (2000) study did not have data for PND59,

24   so the results are not entirely comparable. Again, there was evidence of an increase in the effect

25   of ammonium perchlorate on motor activity at the later habituation times (posterior probability =

26   0.93). In the first habituation interval there was no evidence of an increase in motor activity with

27   dose (posterior probability = 0.58), while in the final interval there was moderate evidence

28   (posterior probability = 0.94). The dose estimated to increase the average of ambulatory

29   movements in the final habituation time by 10% is 4.60 with a credible interval of (2.18,

30   infinity). This interval was wider than the interval observed in the Bekkedal et al. (2000) study;
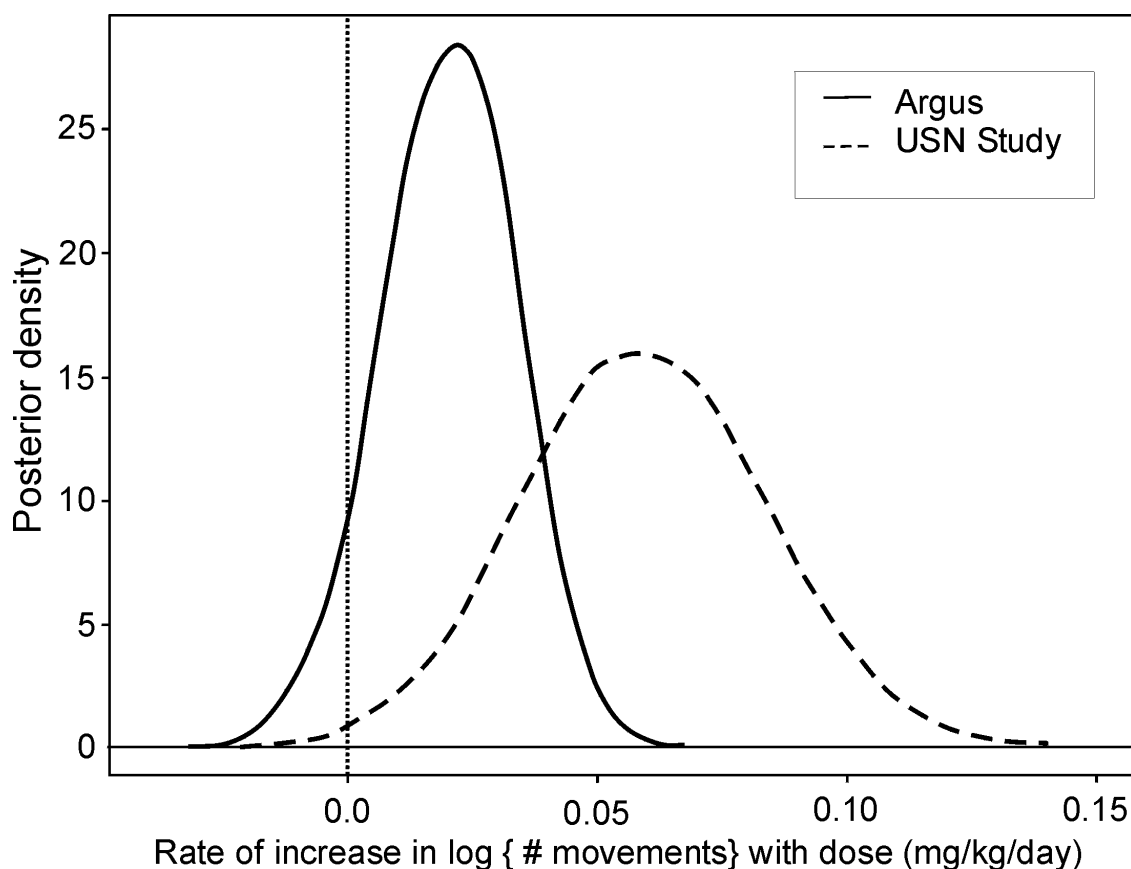
Figure 5-11. **Bayesian estimates of the posterior densities for the expected increase in the logarithm of the number of ambulatory movements at the final habituation time per unit dose (mg/kg-day) increase of ammonium perchlorate (Dunson, 2001a). A separate analysis for the Argus Research Laboratories, Inc. (1998a) and United States Navy (Bekkedal et al., 2000) was performed.**

1   possibly due to greater variability in the Argus data as noted in 1998 by EPA. This result is

2   slightly higher than the BMD analysis (Section 5.3.1.4) estimate of 1.04 mg/kg-day.

3   One of the advantages of Bayesian analysis is that it provides for formal combination of

4   data from different studies. To perform a combined analysis of data from the USN Study

5   (Bekkedal et al., 2000) and the Argus (1998) study, a modification of the model described above

6   was used (Dunson, 2001a). The number of ambulatory movements was first standardized by

7   subtracting the overall mean and dividing by the standard deviation. A linear mixed-effects

8   regression model that incorporated distinct baseline parameters (i.e., intercept, age-effects,

1   habituation time effects, error variances) for the two studies was then fit, assuming common

2   slope parameters. This approach allowed the different studies to have distinct baseline

3   parameters, including aging effects.

4   Figure 5-12 shows the posterior density from the combined analysis of the Argus Research

5   Laboratories, Inc. (1998a) study and the Bekkedal et al. (2000) study. In this combined analysis,

6   the posterior probability of an increase in motor activity with dose was 0.99. For rats that

7   averages 34.09 ambulatory movements at the final habituation time in the absence of exposure

8   (the average value in the Argus study), the estimated dose needed to increase this average by

9   10% is 3.33 [95% credible interval = (1.91,12.78)].

10
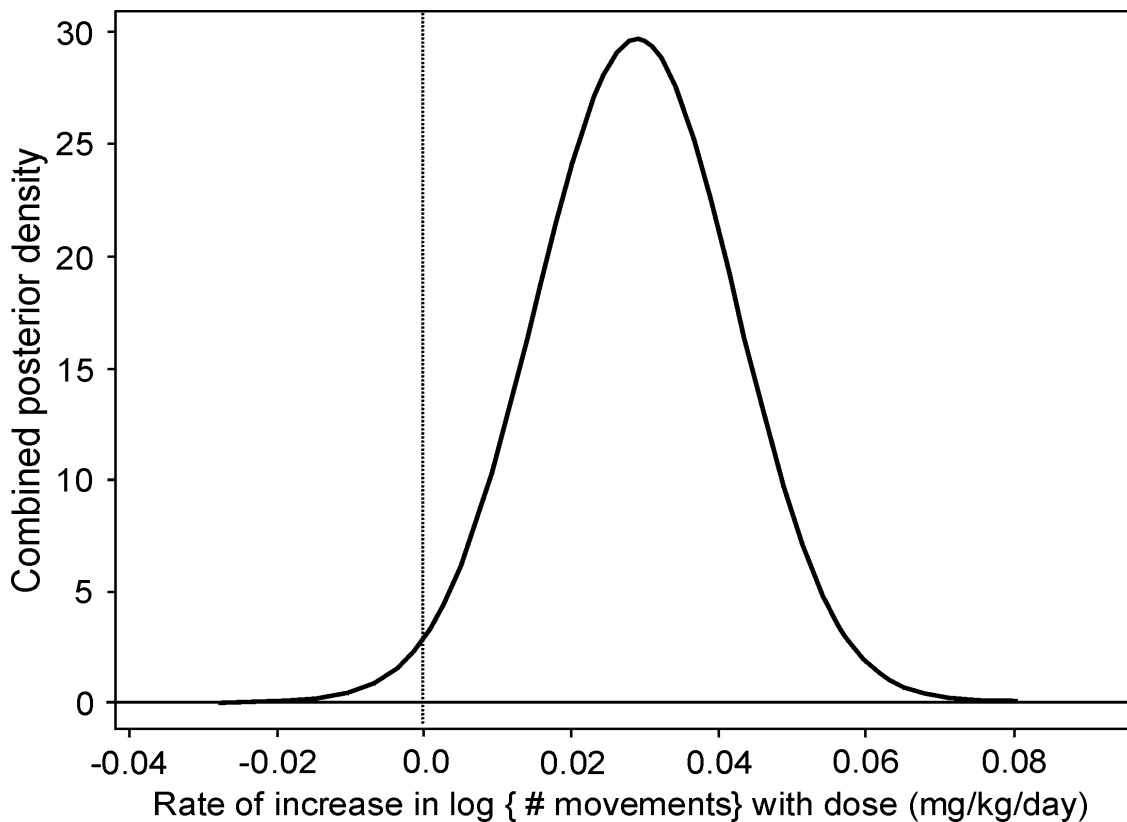
Figure 5-12. **Bayesian estimate of the posterior density for the expected increase in the logarithm of the number of ambulatory movements at the final habituation time per unit dose (mg/kg-day) increase of ammonium perchlorate for the combined data from the two studies of motor activity effects shown in Figure 5-12 (Dunson, 2001a).**

1   There was evidence of an increasing dose-response trend in motor activity in both the

2   Argus Research Laboratories, Inc. (1998a) and Bekkedal et al. (2000) studies, although the effect

3   in the Argus study was less pronounced, likely due to the variability in the data previously noted.

4   Given this, it is remarkable that the two studies showed such similar results. The Bayesian

5   analysis can be applied to risk assessment in an analogous fashion to the benchmark dose

6   analysis (Hasselblad and Jarabek, 1996). The lower limit on the estimated dose corresponding to

7   a 10% increase in motor activity relative to control can be used as a surrogate for the NOAEL for

8   the point of departure for reference dose derivation. For the Argus Research study, the lower

9   limit of the 95% credible interval for the dose was 2.18, while for the Bekkedal et al. (2000)

10   study the corresponding estimate was 0.90. In the combined analysis, the lower limit was 1.91.

11   Because of the variability in the Argus Research Laboratories, Inc. (1998a) study, a NOAEL that

12   relied on the Bekkedal et al. (2000) was chosen at 1.0 mg/kg-day to represent effects on motor

13   activity from these combined data.

14

5.3.3 The 2001 "Effects Study"

16   The Argus Research Laboratories, Inc. (2001) study was performed in response to

17   recommendations made at the 1999 external peer review (Research Triangle Institute, 1999) for

18   additional analyses of the thyroid and brain effects during gestation and post-natal days. Because

19   Argus Laboratories identified the day of birth as PND1, the age nomenclature of PND5, PND10,

20   and PND22 (Argus, 2001) is off by one day as referenced by EPA definition. These ages are

21   therefore referred to as PND4, PND9, and PND21.

22   It should be noted that exposure in this study started two weeks prior to the start of

23   cohabitation. The rationale was to ensure a hypothyroid state, but given the response of the rat

24   system to perturbation, it is more likely that this resulted in the dams already compensating for

25   the effect of perchlorate prior to pregnancy by upregulation of the NIS, making comparison with

26   the 1998 developmental neurotoxicity study (Section 5.3.1) more difficult.

27   The thyroid and brain from one male and one female pup per litter were selected for

28   histological and morphometric evaluation, with one set evaluated on PND4, PND9, and PND21.

29

30

1   **5.3.3.1   Results of General Toxicity Measures**

2   There were no remarkable clinical or necropsy observations. Average body weights and

3   body weight changes for female rats were comparable among the five exposure groups through

4   the pre-cohabitation and gestation periods. Body weight gains for female rats in the 1.0 and

5   30.0 mg/kg-day target dosage groups were significantly increased on PND12 to PND15

6   compared to the carrier group. These increases were not considered treatment-related because

7   they were a singular occurrence and were transient.

8

9   **5.3.3.2   Evaluation of Thyroid Histopathology**

10   The thyroid histopathology in this study was evaluated using the same scoring system as

11   developed for the PWG review and was performed by one of the pathologists who served on the

12   PWG. A second read of these slides has not occurred. The data will be discussed individually

13   for each of the time points. Benchmark dose analyses conducted by EPA will be presented in

14   Section 5.3.3.2.1.

15   Absolute thyroid weights were increased significantly in the 30.0 mg/kg-day group in the

16   dams on GD21 and decreased colloid; increased hypertrophy and increased hyperplasia were also

17   noted at this dose. Thyroid weights were not collected for fetuses on GD21, but colloid depletion

18   was noted in both male and female fetuses at both the 1.0 and 30.0 mg/kg-day doses.

19   Thyroid weight in pups was measured on PND4, and the absolute weight was significantly

20   effected at 30 mg/kg-day, suggesting a NOAEL at 1.0 mg/kg-day. Histopathology was evident at

21   lower doses, suggesting a NOAEL at 0.1 for colloid depletion; however, no real dose-related

22   trend in either hypertrophy or hyperplasia was evident.

23   Thyroid weight in dams on PND9 continued to be effected significantly at 30 mg/kg-day,

24   with histopathology noted at lower doses. The pups on PND9 were more sensitive than the

25   dams, exhibiting statistically increased absolute thyroid weights at 0.1 mg/kg-day and higher

26   doses and suggesting a NOAEL at 0.01 mg/kg-day. A dose-related trend in histopathology in

27   this same range of doses was noted in the pups, especially for colloid depletion.

28   Thyroid weight in dams on PND21 remained significantly effected at 30 mg/kg-day, with a

29   clear dose-related trend in colloid depletion, hypertrophy and hyperplasia. All three

30   histopathological indices were increased at 30 mg/kg-day, and hyperplasia was also significantly

31   increased at the 1 mg/kg-day dose. It is interesting to note that hyperplasia was more sensitive

1 than both hypertrophy and colloid depletion in the dams at this time point, perhaps indicating a

2 system coming into the chronic phase of compensation described in Chapter 6.

3 Pups on PND21 also continued to exhibit increased thyroid weights at both 1 and

4 30 mg/kg-day (females only at 1.0 mg/kg-day). Colloid depletion was clearly significant at

5 30 mg/kg-day, and hyperplasia was noted although not designated as significant. Despite the

6 assertion by Argus Research Laboratories, Inc. (2001) that there was no dose-related trend in

7 hyperplasia, a BMD analysis indicated otherwise (see below). Hypertrophy was not noted, again

8 indicating an overlap among the three diagnostic indices of thyroid effects used by the PWG.

9 Benchmark dose analyses performed by EPA are presented in Table 5-3 (Geller, 2001b).

10 A benchmark response level of a 10% increase in incidence over controls, i.e., BMD10 and

11 BMDL10, was adopted for all studies. Data were fit with a log-logistic function constrained such

12 that the slope was ≥ 1.

13

14 **5.3.3.2.1 Benchmark Dose Analyses of Thyroid Histopathology**

15 BMDL values in the dams on GD21 were 1.01, 1.19, and 8.51 mg/kg-day for colloid

16 depletion, hypertrophy, and hyperplasia. By PND9, these values decreased to 0.13, 1.01, and

17 0.92 mg/kg-day. Similar values for dams on PND21 were 0.62, 1.24, and 0.99 mg/kg-day for

18 colloid depletion, hypertrophy, and hyperplasia. Of note is the overlap between the estimates for

19 hypertrophy and hyperplasia.

20 The effects of ammonium perchlorate on the pups' thyroid glands are largely limited to

21 colloid depletion. The dams show additional dose-related effects on thyroid histopathology that

22 were evaluated as thyroid hypertrophy and hyperplasia. The low incidence of these latter two

23 endpoints in pups may be related to the duration of exposure compared to the dams and the adult

24 rats examined in earlier studies (Geller, 2001a). Alternatively, hyperplasia and hypertrophy may

25 be have been difficult to detect in the smaller thyroid glands from the young pups.

26 The BMDL10 is lowest in the GD21 pups and is estimated at 0.12 mg/kg-day for the male

27 and female pups combined, or for male pups alone, and for female pups alone at 0.04 mg/kg-day.

28 The BMDL10 increases with age (Figure 5-13), suggesting that the thyroid gland may be most

29 susceptible to the effects of perchlorate during gestation or at the time of parturition (Geller,

30 2001b). This is likely due to the double effects of perchlorate inhibition of thyroid function in

TABLE 5-3. BENCHMARK DOSE (BMD)[a] AND BENCHMARK DOSE LOWER CONFIDENCE LIMIT (BMDL)[a] ESTIMATES FROM THYROID HISTOPATHOLOGY IN THE "EFFECTS STUDY"
(Argus Laboratories, Inc., 2001; Geller, 2001b)

Study Population "Effects" Study (Argus, 2001)	Colloid Depletion				Hypertrophy				Hyperplasia			
	BMD	BMDL	$\chi^{2\ b}$	Exp[c]	BMD	BMDL	$\chi^{2\ b}$	Exp[c]	BMD	BMDL	$\chi^{2\ b}$	Exp[c]
GD 21 Dams	5.10	1.01	1.00	17.90	15.46	1.19	1.00	6.25	28.54	8.51	1.0	5.03
GD 21 Male pups	0.69	0.12	1.00	8.82	NOE[4]	NOE	NOE	NOE	NOE	NOE	NOE	NOE
GD 21 Female pups	0.18	0.04	0.60	2.08	NOE	NOE	NOE	NOE	NOE	NOE	NOE	NOE
GD 21 M + F pups	0.65	0.12	0.16	7.80	NOE	NOE	NOE	NOE	NOE	NOE	NOE	NOE
PND4 Male pups	0.88	0.29	0.12	7.37	NOE	NOE	NOE	NOE	NOE	NOE	NOE	NOE
PND4 Female pups	0.82	0.18	0.12	7.78	NOE	NOE	NOE	NOE	NOE	NOE	NOE	NOE
PND4 M + F pups	0.84	0.33	0.02	7.50	NOE	NOE	NOE	NOE	NOE	NOE	NOE	NOE
PND9 Dams	0.62	0.13	0.59	2.65	2.65	1.01	0.22	17.86	2.24	0.92	0.49	1.0
PND9 Male pups	1.29	0.71	0.59	6.40	NOE	NOE	NOE	NOE	NOE	NOE	NOE	NOE
PND9 Female pups	0.33	0.13	0.61	1.30	NOE	NOE	NOE	NOE	NOE	NOE	NOE	NOE
PND9 M + F pups	0.93	0.48	0.36	3.77	NOE	NOE	NOE	NOE	NOE	NOE	NOE	NOE
PND21 Dams	1.21	0.62	0.34	4.90	15.60	1.24	1.0	6.34	3.59	0.99	0.66	1.0
PND21 Male pups	17.33	1.36	1.0	5.85	NOE	NOE	NOE	NOE	26.97	5.45	0.58	5.06
PND21 Female pups	16.42	1.24	1.00	5.94	NOE	NOE	NOE	NOE	NOE	NOE	NOE	NOE
PND21 M + F pups	17.32	2.17	1.0	5.92	NOE	NOE	NOE	NOE	54.17	13.70	0.24	1.0

[a] Units of mg/kg-day.
[b] χ^2 goodness of fit criterion
[c] Exponent in log-logistic function restricted to be ≥ 1.0.
[d] NOE = No observed effect.
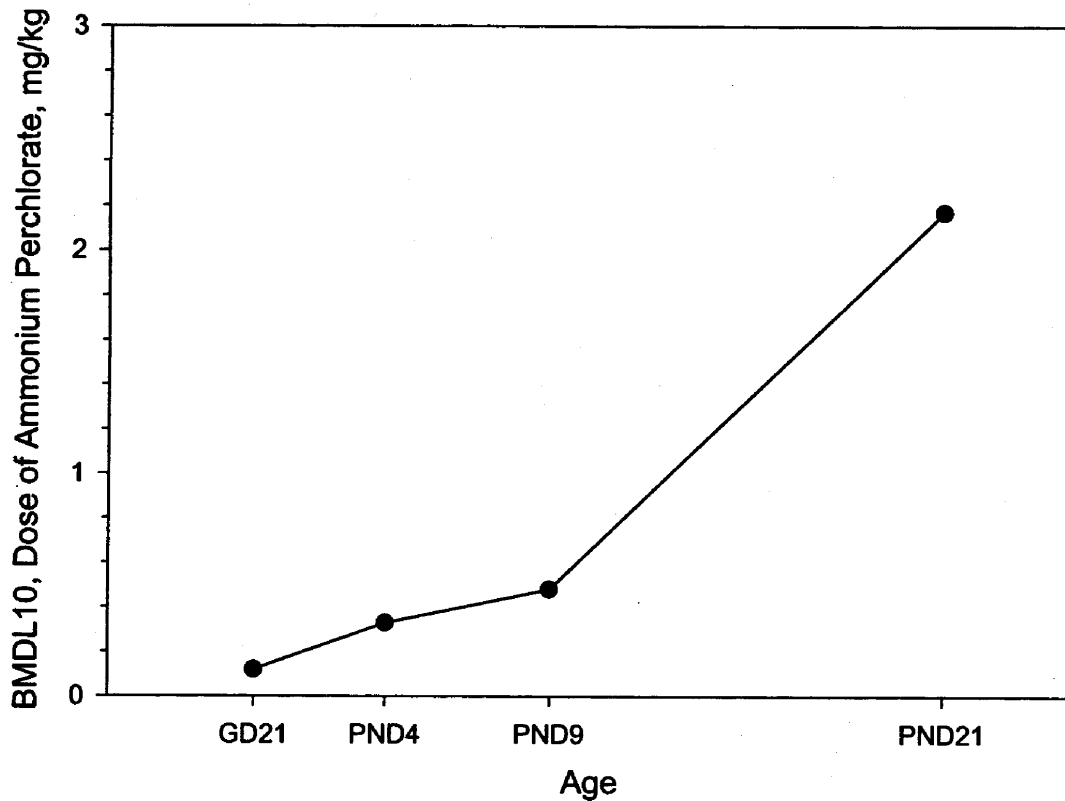
Figure 5-13. Lower confidence limit on the dose of ammonium perchlorate in drinking
water that produced a 10% increase in the incidence of colloid depletion in
the thyroid gland as a function of post-natal age of rat pups. Data of Argus
Laboratories, Inc. (2001). Male and female data combined (Geller, 2001b).

1   the pup and the lack of protection of the pup by the dam because of her own compromised

2   thyroid function. After 21 days of post-natal exposure, the male pups also show follicular cell

3   hyperplasia.

4   The BMD and BMDL estimates of 0.84 and 0.33 mg/kg-day for the PND4 male and female

5   pups in this study (Table 5-3) do corroborate the BMD and BMDL for colloid depletion for the

6   PND4 pups from the 1998 Neurobehavioral Developmental study of 0.53 and 0.33 mg/kg-day

7   (Table 5-1). However, it should be noted that an unrestricted model also fits those data

8   adequately and results in a BMD and BMDL estimate of 0.45 and 0.009 mg/kg-day, suggesting

9   variability in those analyses (Geller, 2001b). Again, the lower estimates based on the 1998 data

10   at this time point (PND4) may be due to differences in the dosing of the dams between the two

11   studies.

1   The BMD and BMDL estimates of 17.32 and 2.17 mg/kg-day for the PND21 male and

2   female pups in this "Effects Study" (Table 5-3) are somewhat higher than the previous 1999

3   two-generation reproductive toxicity study estimates of 2.51 and 0.80 mg/kg-day (Table 5-1).

4   However, comparison of the results of the two-generation reproductive toxicity study to the

5   current results may be difficult because of differences in the spacing of doses tested.

6

7   **5.3.3.3   Thyroid and Pituitary Hormone Analyses**

8   Thyroid (T3 and T4) and pituitary (TSH) hormones were also analyzed in the "Effects

9   Study" at various time points. Thyroid hormones and TSH were evaluated in the dams and fetus

10   on GD21, in the dams on PND10 and PND22, and in neonates on PND5, PND10, and PND22

11   (corresponding to PND4, PND9, and PND21 according to EPA nomenclature as explained

12   earlier). Table 5-4 presents the results of ANOVA analyses performed by EPA (Crofton, 2001b).

13   Maternal serum measures of the hormones were subjected to separate two-way ANOVA.

14   Treatment (dose) and age (GD21 or PND5, PND10 or PND22) were the independent between-

15   subjects variables. Two separate approaches were used to address the offspring data due to

16   differences in experimental design. The data from GD21, PND5 and PND10 were obtained from

17   litter-pooled samples due to the small volumes of blood and no gender analyses were possible.

18   These data were subjected to separate two-way ANOVA with age (GD21, PND5, or PND10) and

19   treatment (dose) as between-subjects variables. Blood samples from PND22 were not pooled so

20   that the data from this age were subjected to separate two-way ANOVA with gender and

21   treatment (dose) as independent variables. Mean contrasts were performed using Duncan's

22   Multiple range test. Significant two-way ANOVA were followed by step-down one-way

23   ANOVA to determine the main effects of treatment. If the interaction term was not significant,

24   then the model was refit if main effects were found. A reduced model was then fitted to the data

25   retaining only the main effects found significant previously, described as the "liberal" approach

26   in Crofton and Marcus (2001) and Marcus (2001).

27   EPA benchmark dose analyses (Geller, 2001c) of these results will also be discussed. The

28   benchmark estimates were generated using the Bench Mark Dose Software version 1.30, and fit a

29   Hill equation constrained such that the exponent on dose was ≥ 1.0 (Geller, 2001c). The BMDL

30   estimates indicate that the thyroid and pituitary hormones are exquisitely sensitive to the effects

31   of perchlorate.

TABLE 5-4. NOAELs AND LOAELs FOR EFFECTS ON THYROID AND PITUITARY HORMONES FROM THE ARGUS 2001 "EFFECTS STUDY" (Crofton, 2001b)

Generation	Hormone	Age	Sex	Effect Level Designation	
				NOAEL	LOAEL
Dams	T3	GD21	F		
		PND10	F	1.0	30.0
		PND22	F		
	T4	GD21	F	—	0.01
		PND10	F	0.1	1
		PND22	F	1.0	30.0
	TSH	GD21	F	—	0.01
		PND10	F	—	0.01
		PND22	F	0.01	0.1
Fetus and Offspring	T3	GD21	Pooled		
		PND5	Pooled	—	0.01
		PND10	Pooled		
		PND22	F	0.01	1.0
			M		
	T4	GD21	Pooled		
		PND5	Pooled	0.01	0.1
		PND10	Pooled		
		PND22	F	no significant effects	
			M	—	0.01
	TSH	GD21	Pooled	0.1	1.0
		PND5	Pooled	no significant effects	
		PND10	Pooled	—	0.01
		PND22	F	0.01	0.1
			M	—	0.01

[a]Dosages of 0, 0.01, 0.1, 1.0, and 30 mg/kg-day.

5.3.3.3.1 Maternal Hormone Analyses

Exposure to perchlorate produced significant decreases in thyroid hormones and an increase in TSH in the dams at the various ages tested. For effects on maternal T3, there was no age-by-treatment interaction and the NOAEL at all time points was 1.0 mg/kg-day. There was a significant age-by-treatment interaction for effects on maternal T4. Step-down analyses resulted in a LOAEL at 0.01, 1.0 and 30.0 mg/kg-day at GD21, PND9 and PND21. The 0.01 mg/kg-day level is a LOAEL for the dams at GD21. There was also a significant age-by-treatment interaction for the effects on maternal TSH. Step-down analyses resulted in a LOAEL at 0.01, 0.01 and 0.1 mg/kg-day at GD21, PND9 and PND21. As for the effects on T4, there was no NOAEL at GD21 for the effects on TSH. There was no NOAEL for the effects on TSH at PND9 as well. These effects on T4 and TSH at GD21 are consistent with the Argus Laboratories Inc. (2001) analyses. Benchmark dose analyses resulted in BMD estimates of 1.63, 0.006 and 2.38 mg/kg-day for the effects on T3, T4, and TSH at GD21. BMDL estimates were only calculable for T4 in the dams and resulted in an estimate of 0.004 mg/kg-day. Benchmark dose calculations were not performed for the dams on PND9. At PND21, a BMDL estimate was calculable only for TSH in the dams with a resultant estimate of 0.53 mg/kg-day.

5.3.3.3.2 Fetal and Neonatal Hormone Analyses

Maternal exposure to perchlorate resulted in hypothyroidism in the offspring. There were significant dose-related decreases in thyroid hormones and increases in TSH at all time points evaluated.

There were no age-by-treatment interactions for the effects on T3 at any age tested. The LOAEL for GD21, and post-natal days 4 and 9 was 0.01 mg/kg-day. This value is lower than that reported in the Argus Laboratories, Inc. (2001) analyses. The specified benchmark dose analysis were not computable for T3 at PND4 or PND21. There was no significant gender-by-treatment interaction for the effects on T3. The NOAEL for effects on T3 at PND21 was 0.1 mg/kg-day. A BMDL was calculable only for the male pups and resulted in an estimate of 0.13 mg/kg-day.

There were also no age-by-treatment for the effects on T4. The LOAEL was 0.1 mg/kg-day and the NOAEL was 0.01 mg/kg-day for GD21 and PND4 and PND9. At PND21, there was a significant gender-by-treatment interaction for the effects on T4. There was no NOAEL

1   established for the male pups and 0.01 mg/kg-day was a LOAEL, whereas 0.01 was suggested as

2   a NOAEL in the Argus Laboratories, Inc. (2001) analyses. The females did not show significant

3   effects in either the EPA or Argus Laboratories, Inc. (2001) analyses. BMDL estimates were

4   extremely sensitive for changes in T4 at PND21 in the males with a BMD and BMDL at

5   0.001 and 2.86 x 10^{-7} mg/kg-day. Benchmark analyses did not converge for the data from the

6   female pups alone or for the combined data.

7   There was a significant age-by-treatment interaction for the effects on TSH. Step-down

8   analyses revealed a NOAEL at 0.1 mg/kg-day for GD21. There was no significant effect on TSH

9   at PND5, but then no NOAEL on PND9 with a LOAEL at 0.01 mg/kg-day. The LOAEL was

10   also 0.01 mg/kg-day in male pups at PND21. The females were slightly less sensitive as

11   suggested by the significant gender-by-treatment interaction. The NOAEL in female pups on

12   PND21 was 0.01 mg/kg-day. Benchmark analyses on the combined data resulted in a BMD and

13   BMDL of 0.06 and 0.02 mg/kg-day for the effects on TSH.

14

15   **5.3.3.4   Brain Morphometry Effects**

16   Due to the deficiencies of the remaining tissue blocks from the previous developmental

17   neurotoxicity study (Argus Research Laboratories, Inc. 1998a), it was determined that the

18   recommendation of the external peer review panel to evaluate more sections could not be

19   accomplished unless a new study was performed (Harry, 2001). Thus, one major objective of the

20   Argus Laboratories, Inc. (2001) "Effects Study" was replication of brain morphometric

21   measurements in order to address concerns raised by the US EPA, the NIEHS, and the external

22   peer review panel regarding results observed in the 1998 developmental neurotoxicity study

23   (Argus, Protocol Number 1613-002, 1998a; U.S. Environmental Protection Agency, 1998d). The

24   purpose was to evaluate, under more rigorous experimental conditions and according to the EPA

25   developmental neurotoxicity guidelines (U.S. Environmental Protection Agency, 1998b),

26   whether the effect in the corpus callosum identified by the EPA in the previous assessment

27   (Section 5.3.1) would be replicated.

28   In addition, another objective was to identify effects that may occur in other brain regions.

29   Details with respect to the rationale motivating the experimental design can be found in Harry

30   (2001). A brief summary of important points will be provided here, but the reader is referred to

31   Harry (2001) for specifics on this protocol and to other review articles (Garman et al., 2001;

1   Adams et al., 2000; Rice and Barone, 2000; U. S. Environmental Protection Agency, 1998b,g,h)

2   for a fuller appreciation of the state-of-the-science supporting the use of these measures as

3   developmental neurotoxicity indices in risk assessment. The use of the rodent and not a non-

4   human primate was based on the degree of difficulty and the ethical issues involved with

5   conducting such screening studies in addition to the need to replicate previous findings. The

6   work, to document the process of normal development and alterations in the rat cited in these

7   reviews, supports the use of rodent models for determining potential adverse effects on the

8   developing brain.

9   It should be noted that Argus Laboratories identifies the day of birth as PND1; therefore,

10   the age nomenclature as recommended in the EPA guidelines for PND10 and PND22 actually

11   corresponds to PND9 and PND21 in this study. Likewise, in the previous 1998 Argus Research

12   Laboratories, Inc. Study (Section 5.3.1), the morphometry performed on PND12 was actually

13   done on PND11. While the actual ages were slightly different between the two studies, the

14   concept of capturing an active process of development with brain morphometry remains in effect

15   (Harry, 2001).

16   The motivation for evaluation of brain morphometry was based on the fact that the

17   formation and maturation of the nervous system is critically dependent upon both a temporal and

18   spatial organization pattern (U.S. EPA, 1998b; Harry, 2001). Within this framework, an

19   interdependency between the various cell types in the brain and a precise spatial relationship of

20   one cell type to one cell type another has been demonstrated. During this time, the developing

21   system is undergoing rapid maturation of organizational and regulatory processes. Thus, the

22   disruption of the developmental profile of one cell type may significantly influence critical events

23   in later development, resulting in an alteration of the normal formation of the brain and its

24   functional connections. Many toxic agents have been shown to interfere with one or more of the

25   developmental processes of the brain (i.e., cell division of neuronal and glia precursor cells, cell

26   interaction with the immediate environment through surface receptors or cell adhesion

27   molecules, regulation of cytoskeletal processes that control proliferation and migration, cell-cell

28   interactions that underlie synaptogenesis, development of the cerebral circulation and the blood-

29   brain barrier, myelination, and programmed cell death). Such perturbations may not be evident

30   by standard histological assessments as often there is little, if any, evidence of cell death. Rather

1   what is seen is a delay or disruption in the normal development and maturation of specific neural

2   regions (Harry, 2001).

3   Immersion fixation was the tissue processing method of choice and was both recommended

4   and agreed upon by both the EPA and the PSG for the study. While the tissue fixation method of

5   choice in adult rodents is via cardiac perfusion, even this procedure is not without problems that

6   can compromise tissue integrity. It has been documented that immersion fixation artifacts can

7   influence histological and morphometric evaluations of adult brains; however, a less than optimal

8   cardiac perfusion can also result in morphological artifacts. For the younger animal, there is less

9   of a consensus on the proper manner of fixation. With the decreasing size and blood volume of

10 the younger animal (PND4 and PND9) used in the protocol, the difficulty of ensuring a good

11 fixation via cardiac perfusion is significantly increased over that in the adult. Further, because

12 comparisons were to be made between the 1998 and the 2001 study, consistency in method of

13 fixation was considered to be a critically important variable to maintain as constant across

14 studies.

15   Following the review of the previous developmental neurotoxicity study (Argus Research

16 Laboratories, Inc., 1998a), and in considering design considerations for the subsequent study, the

17 plane of cut for the brain was discussed (Garman, 2001a,b). While sagittal sections for analysis

18 were recommended for some aspects of morphometric analysis, coronal sections were ultimately

19 adopted since comparisons were to be made between the 1998 and the 2001 study. This final

20 design of the study also adhered to the EPA developmental neurotoxicity testing guidelines that

21 call for coronal sections (U.S. Environmental Protection Agency, 1998g, h). It was originally

22 recommended by the NIEHS that measurements of the corpus callosum in coronal sections

23 should not be conducted at the midline due to possible edema artifacts that can occur from the

24 close proximity of the ventricle. Three sites were recommended for measurement that would

25 have been consistent with the evaluation conducted by NIEHS on the sections from the Argus

26 Research Laboratories, Inc. (1998a) study (Section 5.3.1). It was agreed upon in the final design

27 meeting with PSG contractors that, given the time constraints and need for comparison to the

28 1998 study, one measurement per hemisphere would be recorded at the same site as used in this

29 previous study (Garman, 2001a,b). This was a site just off of the midline of the two

30 hemispheres.

1   Finally, a question raised in the PSG-contracted review (Toxicology Excellence for Risk

2   Assessment, 2001) with regard to age of sampling as it relates to myelin formation should be

3   addressed. The process of myelination is a "developmental landmark" for the maturation of the

4   brain, that is initiated upon the presence of the axon and continues over an extended period of

5   time. It is a structure that matures over time with the accumulation of protein and structural

6   lamella. One major period of myelin protein and lipid synthesis occurs approximately between

7   PND19 and PND35. Thus, while examination at PND21 would not capture the final

8   accumulation of myelin, it would capture events occurring at a time during which myelin

9   processing and lamella wrapping of the axon is actively occurring. Therefore, this may represent

10   a period of critical development of the myelin sheath. Examination of animals with a mature

11   myelin sheath (e.g., ages greater than PND40) may offer information regarding whether any of

12   the changes seen at earlier time points represent a permanent structural alteration. The majority

13   of studies that have examined myelin development and/or alterations in this developmental

14   process have employed biochemical, molecular, as well as, morphological evaluations to make

15   such determinations regarding delay or hypomyelination. From such studies, the time most

16   appropriate for examination appears to be between the ages of PND15 and PND35. Thus,

17   examination of the corpus callosum at PND9 is probably at the limit of early development for an

18   evaluation of the myelin sheath. However, it should be noted again that this study was intended

19   to determine if the effects seen previously (Argus Research Laboratories, Inc., 1998a) could be

20   repeated. Effects in the corpus callosum in that previous study occurred at the early (PND11)

21   and remained at the late (PND82) time points. Brain weight and the size of the frontal cortex and

22   caudate putamen also were effected at the PND82 sacrifice (Section 5.3.1.1).

23   In addition, the development of the axonal pathways connecting the two hemispheres via

24   the corpus callosum also continues to develop during this time period. While the study design

25   allowed for the collection of tissue at PND4, it is felt that any measurements recorded at such age

26   would be very limited in their contribution to the interpretation of the currently available data set.

27   In addition, given the variability of the plane of cut and the difficulty in examining brains of

28   young animals, EPA and NIEHS agree that examination of the corpus callosum in younger

29   animals (the remaining materials available for PND4) would present an even greater problem.

30   Figure 5-14 illustrates where the section levels were taken for the brain morphometry

31   measurements and shows the anatomical landmarks on the ventral and dorsal surfaces of the
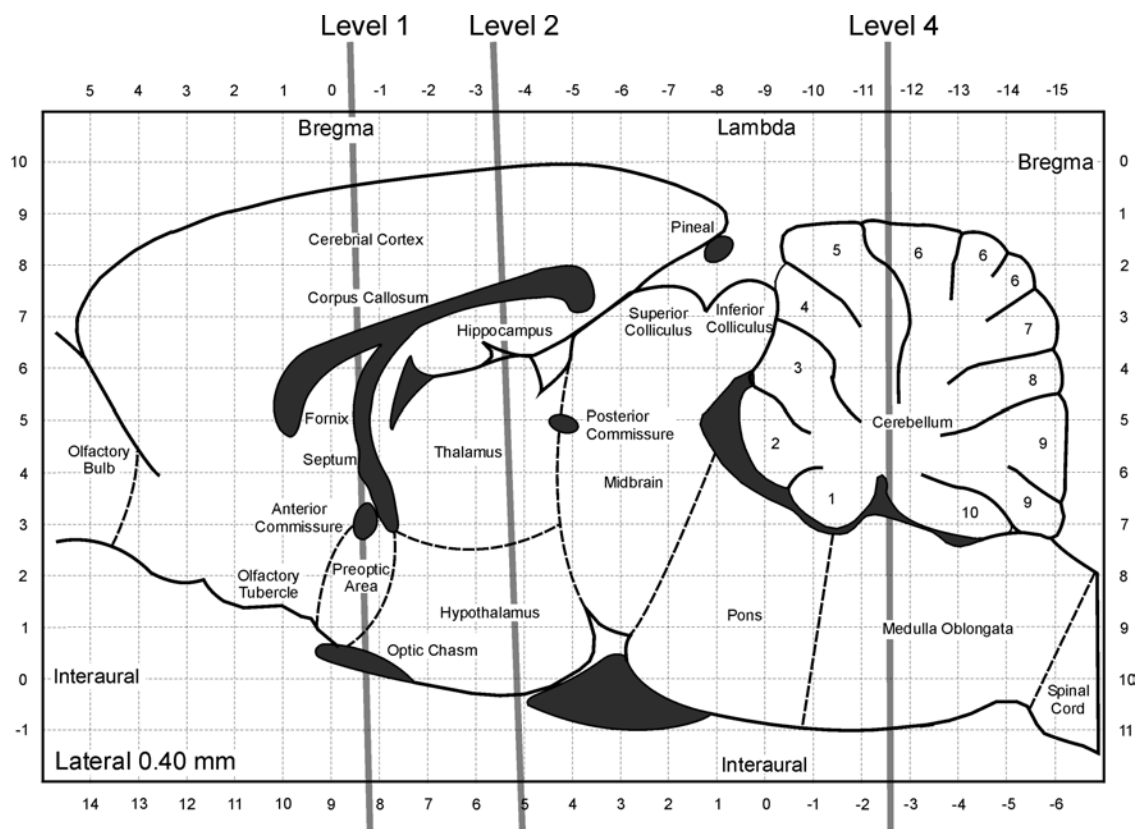
Figure 5-14. **Topograph of the approximate anatomical landmarks on the ventral and dorsal surfaces of the brain used for making the morphometry measurements (Garman, 2001c). The topograph provided is for an adult brain, but the same landmarks are used for PND9 and PND21 brains although the sections at these two other ages would differ due to the rapid growth during this period.**

1   brain. The veterinary pathologist who performed the work has noted that while the landmarks

2   were the same for both the PND9 and PND21 brains, it must be appreciated that the sections

3   from one age versus the other would not look precisely similar (Garman, 2001c) due to the fact

4   that the brain is rapidly growing at this time.

5   Overall, the images of the brain sections from the PND9 and PND21 time points

6   demonstrated that the processing of the brain was adequate for conducting limited morphometric

7   measurements as outlined in the protocol. As mentioned by the PSG-contracted reviewers

8   (Toxicology Excellence for Risk Assessment, 2001) and stated in the study and additional reports

9   (Argus Research Laboratories, Inc., 2001; Consultants in Veterinary Pathology, 2001; Garman,

1   2001d), there was a greater degree of variation in the PND9 sections than in the PND21 brain

2   sections (Harry, 2001). Many sections in the PND9 brains also showed signs of disruption or

3   damage that may have compromised the measurements. For these reasons the EPA relied upon

4   the PND21 measurements, despite corroborating effects from the materials at PND9.

5   There were no significant effects of treatment or sex on brain weight, anterior-posterior

6   cerebrum length, or anterior-posterior cerebellar size at either age tested. As discussed in the

7   Argus Research Laboratories, Inc. (2001) report, statistical analyses consisted of Students' t-test

8   comparisons between the control and the corresponding group of each sex at each separate dose

9   level. For example, PND9 male control striatum measurements were compared to measurements

10 for the PND9 male 30 mg/kg-day dose group, then PND9 male control striatum measurements

11 were compared to the PND9 1 mg/kg-day-dose group. These analyses were run separately for

12 both sexes and ages and all brain areas, right and left sides. The Argus Laboratories, Inc. (2001)

13 analyses found a large number of significant effects on brain morphometry at doses of 0.1 and

14 0.01 mg/kg-day ammonium perchlorate in drinking water.

15   Guidelines on the assessment of neurotoxicity (U.S. Environmental Protection Agency,

16 1998b) specify that alterations in brain structure should be considered adverse and relevant to

17 human health risk assessment. Alterations in brain structure are consistent with the mode-of-

18 action for perchlorate, i.e., transient decrements in T4 and T3 during development can result in

19 neurodevelopmental effects. The significant findings reported in the Argus Laboratories, Inc.

20 (2001) report strongly argue, therefore, that adverse effects of ammonium perchlorate are present

21 at the lowest dose tested and that this data set contains only LOAELs, no NOAELs.

22   While the analysis in the Argus report was provocative, the number of t-tests run increases

23 the risk of introducing Type I error into this analysis. To address this, a more conservative

24 multivariate analysis, profile analysis (Johnson and Wichern, 1988; Tabachnick and Fidell,

25 2001), was run by the EPA (Geller, 2001d). Profile analysis is more conservative than the

26 analysis described above because a multiple analysis of variance (MANOVA) takes into account

27 any correlations between the independent variables; whereas, the multiple t-tests assume

28 complete independence. This analysis also reduced the number of main effects tests by nesting

29 gender within litter and by constructing a vector composed of all of the morphometric data from

30 each animal, then comparing these vectors. The approach is explained in more detail below.

31

5.3.3.4.1 Description of EPA Profile Analysis of Brain Morphometry Effects

1

2   When a series of measurements are made from a single animal, i.e., within-subjects

3   measurements, they can be used to build a profile or vector of scores across the measurement

4   variables. Profile analysis makes between-groups comparisons using a vector composed of all of

5   the (within-subject) measurements taken from each animal. Its primary test, for parallelism of

6   the vectors, establishes whether the pattern of results between treatment groups is the same or

7   different. It is a much more rigorous and conservative test, requiring that all of the measurements

8   (i.e., all brain regions) show a dependence on dose with the same pattern. This determination

9   also allowed examination of the entire set of data without an *a priori* expectation of effect in one

10   brain region or another or the direction of the effect (i.e., decrease or increase). While there is

11   indication that certain areas of the brain are likely susceptible to the effects on thyroid hormones

12   of perchlorate (e.g., Madeira et al., 1991, 1992, 1993), and the previous study performed by

13   Argus indicated that the corpus callosum was affected (U.S. Environmental Protection Agency,

14   1998d; Crofton, 1998c), definitive gestational windows for specific brain areas are unknown.

15   Profile analysis determines whether there were dose-related changes in the pattern of brain

16   growth, i.e., brain growth in one region relative to another while precluding prior expectations

17   about specific areas of the brain or the direction and magnitude of these changes.

18   The profile analysis was run on the data from the PND9 and PND21 animals separately

19   with gender nested within litter (PROC GLM, SAS Institute, Inc, Cary, NC). The data were

20   provided in electronic form from Argus Research Laboratories, Inc. (2001) and in an additional

21   report (Garman, 2001d). Profile analysis requires data from each endpoint for each animal. Data

22   from individual brain regions, both right and left sides, were missing from 8 animals in the PND9

23   cohort and 3 animals in the PND21 cohort, eliminating these animals from the analysis (Geller,

24   2001d: Table 1). If a sex by treatment interaction was found, separate analyses were run on

25   males and females. Treatment effects within a brain region were examined with univariate

26   analyses of variance with gender nested within litter. Dunnett's two-tailed t-test was used to

27   compare each dose group to controls at $\alpha = 0.05$ for step-down tests of treatment effects within a

28   brain region as guided by the overall (univariate) treatment or sex by treatment effects.

29   Right and left side measures of the same brain structures were examined with profile

30   analyses (whole set of data) and repeated measures analyses of variance (univariate analysis on

31   each brain region). While there was no *a priori* reason to expect other than a bilateral effect, the

1   presence of this kind of bias could reflect either anisometries in brain regions (i.e., lateralization)

2   or sectioning that was not perfectly perpendicular to the anterior-posterior axis of the brain and

3   that would have resulted in sampling brain regions at different depths on right and left side.

4   These analyses, together with examination of the images of the brain sections (Harry, 2001)

5   demonstrated some systematic variability in the sectioning resulting in differences in right versus

6   left measurements in different brain regions. The magnitude of the variability was small and not

7   always in the same direction, even within a brain region (varying with the dose group sampled).

8   The small magnitude of difference relative to the dose-related changes found in this study, the

9   fact that different brain regions varied in their laterality bias in different directions, and that

10   different dose groups varied in different directions all argue for simply averaging the right and

11   left brain region measurements for each animal rather than tailoring different analyses for

12   different brain regions. In addition, averaging could help to reduce variability in the data due to

13   sampling only one histological section/brain region/animal. Therefore, data from right and left

14   sides of the brain were averaged before the analysis of dose effects. Where data were missing

15   from only one side of the brain, the existing measurement was used for the analysis.

16   Two additional analyses were run with adjustments to the raw morphometry data in

17   response to suggestions made by reviewers hired by the PSG (Toxicology Excellence for Risk

18   Assessment, 2001) designed to subtract variability due to variation in brain size and focus on

19   changes in the sizes of brain areas relative to one another. As suggested by the PSG review, one

20   analysis was run dividing all of the linear dimensions through by the post-fixation brain weight

21   from each brain. However, EPA and NIEHS note that there are little historical data for

22   normalizing data with post-fixation brain weight (Harry, 2001) and that fixation results in the

23   loss of any evidence of hydration-related changes such as edema or other swelling.

24   The second additional analysis was suggested by the NIEHS and also adjusted for brain

25   size using the anterior-posterior (a-p) measurements of cerebrum and cerebellum and the full

26   width measure of hippocampus to adjust the linear dimensions. In this analysis, frontal, parietal,

27   and corpus collosum dimensions were divided by a-p cerebrum size; dentate, CA1, and CA3

28   were divided by hippocampal width; and the cerebellar linear measurement was divided by the

29   a-p cerebellum measurement. Hippocampus, a-p cerebrum, and a-p cerebellum were not

30   included in the analysis as separate measures. The striatum and external germinal layer

31   measurements were not adjusted by these other linear dimensions.

1   An additional two analyses were run on the PND21 data. These analyses omitted (1) the

2   posterior corpus callosum measurement, or (2) the posterior corpus collosum and all

3   hippocampal measures; i.e., all measures that came from the Level II section since there was

4   some indication that there may have been a systematic difference in the plane of sectioning with

5   dose (Harry, 2001).

6

7   **5.3.3.4.2 Results of EPA Profile Analysis of Brain Morphometry Effects**

8   The brain morphometry profiles were not parallel across treatment groups for PND9 pups

9   (Geller, 2001d: Table 2). The absence of parallel profiles obviates further analysis for equal

10   profiles. This means that the effects of developmental dosing with ammonium perchlorate were

11   different on different brain regions. Planned contrasts show that the 0.01 and 1.0 mg/kg-day

12   doses were significantly different than controls (Geller, 2001d: Table 2A). Adjusting for brain

13   weight had little effect on these results (Geller, 2001d: Table 2B), though the adjustment for the

14   linear size of the different brain regions made the effect at the highest dose (30 mg/kg-day) also

15   significantly different from control (Geller, 2001d: Table 2C).

16   The brain morphometry profiles were also not parallel across treatment groups for the

17   PND21 pups (Geller, 2001d: Table 2A). Contrasts between each of the dose groups and controls

18   showed that the controls differed from all other dose groups at better than $p < 0.0001$, including

19   at the lowest dose used, 0.01 mg/kg-day ammonium perchlorate in drinking water. The absence

20   of parallel profiles obviates further analysis for equal profiles. The analysis adjusting for brain

21   weight or regional size yielded similar, highly significant effects (Geller, 2001d: Tables 2B, 2C).

22   Sex by dose interactions were significant in the parallel profiles analysis of the raw data and with

23   the data adjusted by brain region size. The parallel profile MANOVA remained significant at

24   $p < 0.0001$ in the overall and contrast tests with the posterior corpus callosum or posterior corpus

25   callosum and all hippocampal measurements (i.e., all measurements taken at section Level II

26   removed from the analysis) decreasing concern for confounding introduced by potential bias in

27   sectioning at this level suggested for the males (Harry, 2001).

28   The profile analysis was done using the raw (right-left averaged) data values. Because the

29   brain structures measured yield a range of measurements varying 10-fold, it is difficult to plot the

30   raw data vector in a meaningful way in order to see the differences driving the findings of

31   significant differences between dose groups. Figure 5-15 plots the (unadjusted) region-by-region
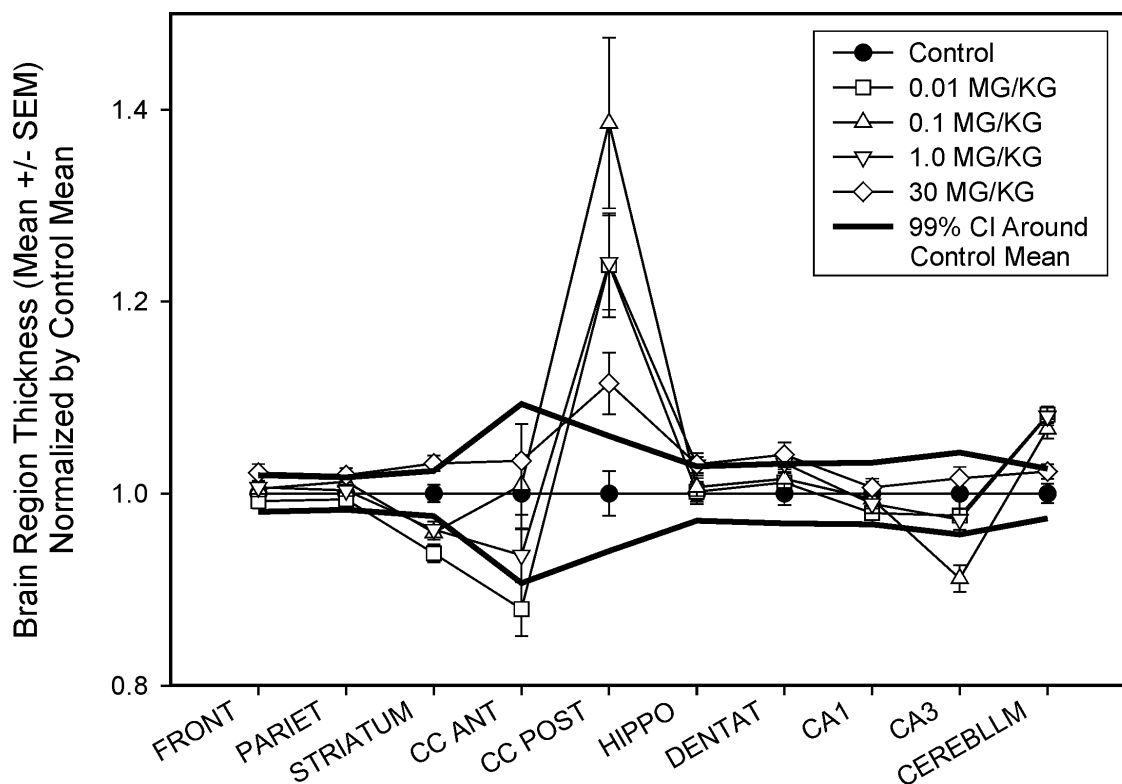
Figure 5-15. Profile analysis of brain morphometry measurements for PND21 rat pup brain regions. The male and female data on linear thickness measurements were combined and normalized by the control mean of each region. The control data are represented by the horizontal line at 1.0. Profile analysis determines whether the vectors of measurements from each treatment group differ from each other and control in a dose-dependent fashion. The heavy line represents the ± 99% confidence interval around the mean control values. Note that while this plot uses the normalized data to more easily illustrate the data vectors, the actual analysis was performed using raw data values (Geller, 2001d). A similar analysis showed effects in PND9 brains (data not shown).

1   size of each brain structure normalized by the mean size of that brain structure in the controls,

2   male, and female combined for the PND21 pup data. The control group is therefore represented

3   by a horizontal line at 1.0 with associated variability. The other dose groups differ from this

4   horizontal line to different extents, and the parallel profiles analysis tests, in essence, whether

5   these departures make the other dose groups significantly "non-horizontal". Note that the

1   analysis was not done on the normalized data; the control values were divided through to aid in

2   visualizing the data vectors used in this analysis. The 99% confidence intervals around the

3   control means represent an envelope inside of which comparable values ± standard error of the

4   mean (SEM) are not significantly different from controls.

5

6   **5.3.3.4.2.1   Univariate analyses of brain morphometry**

7   While the main reason to use profile analysis was to benefit from the power it brings to an

8   analysis by its conservative constraint that requires the entire vector of measurements depend on

9   dose with a consistent pattern, univariate analyses also were evaluated to gain insights into

10 effects on specific regions.

11   **PND9 brains**. Univariate tests yielded significant effects of treatment with ammonium

12 perchlorate in the frontal and parietal regions of the cerebral cortex, the striatum, region CA1 of

13 the hippocampus, the corpus callosum, and the external germinal layer of PND9 pup brains

14 (Geller, 2001d: Table 3A). There is an increase in size at the 1.0 mg/kg-day dose in the frontal,

15 parietal, and striatum measurements, and decreases in size in CA1 and the external germinal

16 layer. There were also treatment-by-sex interactions in the corpus callosum and CA1 regions

17 (Geller, 2001d: Table 3A). Both of these brain regions showed a treatment-related decrease in

18 linear extent in females while showing an increase in size in males. While most of the changes

19 in linear extent measured in the sampled brain regions were ±5 to 11%, the male corpus callosum

20 was increased 23% at both the 0.1 and 1.0 mg/kg-day doses.

21   The adjustment for brain size reduced the significance of treatment effects in the striatum,

22 CA1, and external germinal layer (Geller, 2001d: Table 3A, center). The analysis using

23 adjustment for regional size (Geller, 2001d: Table 3A, right) was nearly identical to the raw data

24 analysis, with the addition of significant effects being noted on cerebellum.

25   A comparison of the profile analysis and the analysis presented in Argus Research

26 Laboratories, Inc. (2001) shows similar results were obtained on the PND9 brain morphometry

27 with one exception. Both analyses found an increase in linear extent of frontal, parietal, and

28 striatum at 1.0 mg/kg-day ammonium perchlorate and in the corpus callosum at the 0.1 and

29 1.0 mg/kg-day dose, with the corpus callosum increase limited to males. There was a decrease in

30 the linear extent of the striatum at 0.1 mg/kg-day dose and decreases in the size of region CA1 of

1   females at the 0.01, 0.1, and 1.0 mg/kg-day doses. The Argus Laboratories, Inc. (2001) analysis

2   did not detect a significant difference in female CA1 at the 0.01 mg/kg-day dose.

3   A post-hoc analysis of the plane of cut of the PND9 brain sections suggested that the

4   0.1 and 1.0 mg/kg-day dose groups were sectioned at a different depth than were the other dose

5   groups (Harry, 2001). This likely contributed to the small but significant increase in size of the

6   frontal, parietal, and striatum sections in the 1.0 mg/kg-day dose groups and may have

7   contributed to the large increase in size of the anterior corpus callosum seen in the PND9 males.

8   **PND21 brains**. The striatum, cerebellum, and corpus callosum II (posterior sample) all

9   showed significant changes with the lowest administered dose of ammonium perchlorate, 0.01

10 mg/kg-day (Geller, 2001d: Table 3B, left). The striatum was significantly reduced in size at all

11 but the highest dose. Region CA3 of the hippocampus similarly showed a U-shaped dose

12 response. The cerebellum and the posterior corpus callosum increased in size with dose in an

13 inverted U-shape. There were sex-by-treatment interactions in striatum and frontal cortex such

14 that the female rats showed a stronger dose-related decrease in linear measurement than males.

15 Both males and females show a complex dose response in the anterior corpus callosum

16 measurement. As in the PND9 animals, the changes in linear extent were generally in the $\pm$ 5 to

17 11% range with the exception of the posterior portion of the corpus callosum, which showed an

18 increase in size of 24% in the 0.01 and 1.0 mg/kg dose groups, and a 39% increase in the

19 0.1 mg/kg dose group.

20   The adjustments for brain size had little effect on the region by region results at PND21

21 (Geller, 2001d: Table 3B, center, right). Dividing through by the a-p or hippocampal

22 measurements resulted in additional significant dose effects noted on CA1 and a sex by dose

23 effect on cerebellum.

24   The Argus Research Laboratories Inc. (2001) and current EPA analyses agreed. Both

25 analyses found a significant decrease in size of the striatum at 0.01, 0.1, and 1.0 mg/kg doses and

26 increases in size of the corpus callosum II (posterior) and cerebellum at the same doses. Both

27 analyses noted the decrease in size of CA3 at the 0.1 mg/kg dose, the decreased anterior corpus

28 callosum in females at 0.01 mg/kg, and the increased size of the frontal region in males at 0.1 and

29 30 mg/kg.

30

31

5.3.3.4.3 Conclusions of EPA Brain Morphometry Analyses of Brain Morphometry Effects

There were significant differences in brain morphometry due to treatment with ammonium perchlorate at both PND9 and PND21 in this study. Tables 2 and 3 in Geller (2001d) enumerate strong effects of developmental exposure to ammonium perchlorate on brain morphometry considered across all regions tested and in the analysis of individual brain regions. These effects were present at PND9 and PND21, with the latter age group showing stronger effects. Many of these effects represent an increase or decrease of ± 10% in the size of a brain region, similar to the range of morphometric alteration noted in a recent study of fetal alcohol syndrome (Bookstein, et al., 2001). The corpus callosum showed a notable increase of 24% or more in linear extent at PND21 in the 0.01, 0.1, and 1.0 mg/kg ammonium perchlorate dosing groups. Adjusting the raw morphometric determinations by either brain weight or measurements of larger brain areas (i.e., cerebrum, cerebellum, and hippocampus) had no strong effect on the results of the analysis.

The significant differences in the parallel-profiles test demonstrate exposure-related changes in relative growth of different brain areas even at the lowest administered dose (Geller, 2001d: Table 2). Univariate analyses to further investigate these effects showed effects on a number of different brain regions at both ages tested. The most sensitive endpoints were the linear dimensions of the striatum, corpus callosum, and cerebellum at the 0.01 mg/kg-day dose when males and females were considered together at PND21. Thus, these analyses ultimately agree with those submitted in Argus Laboratories, Inc. (2001): exposure to 0.01 mg/kg-day ammonium perchlorate during gestational and post-partum (weanling) development resulted in measurable changes in brain structures.

The increase in the size of the corpus callosum in this study replicates that seen in the previous morphometric analysis of rats developmentally exposed to ammonium perchlorate (U.S. Environmental Protection Agency, 1998d, Crofton, 1998c). This is notable given the differences between the two studies. The previous data were obtained from tissues from rats aged PND11 rather than PND9 and PND21, and dose spacing included high doses of 3 and 10 mg/kg rather than 1 and 30 mg/kg as in this study. Fewer animals were used in the previous study (6/dose/sex) than in the current study (approximately 15/dose/sex), and litter identity was considered in the current analysis. It also has been noted by Garman (2001c), a principal investigator with

1   established experience in performing brain morphometry on a substantial number of studies, that

2   such a treatment-related pattern has not been observed in other studies.

3   It should be noted that changes in thyroid hormone levels effect different brain regions

4   differently during development. For example, developmental hypothyroidism prolongs the

5   expansion of the external granular layer and increases fissure formation in the cerebellum

6   (Lauder, et. al., 1974). Different brain regions show an inverted U or U-shape dose response;

7   this is not uncommon in biological systems as compensatory or other mechanisms may be

8   triggered at high doses.

9   Fixation artifacts are not a concern in the study because all brains were fixed and embedded

10   at the same time. In addition, dose-related effects were seen as both increases and decreases in

11   brain region size. EPA concludes from this that whatever artifacts may be present were not large

12   enough to obviate alterations of the magnitude observed. There is some concern over sectioning

13   artifacts because the brains from the different dose groups were sectioned at different intervals

14   after sacrifice (Argus Research Laboratories, Inc., 2001) and post-hoc analysis of the brain

15   sections did reveal some systematic differences in the PND9 animals and in a limited sample of

16   sections examined from the PND21 animals (Harry, 2001). Additional sectioning is being

17   performed by EPA to address whether the anterior to posterior bias selection suggested in the

18   males (Harry, 2001) is a true confounder because normative data for brain measurements at these

19   ages are not available. These new data will be made available to the external peer review panel

20   as soon as possible. Because the analyses conducted without sections from this level still

21   resulted in a significant effect at the 0.01 mg/kg-day dose and the dose-related changes noted in

22   this study have not been noted in other studies with tissue sampler treated similarly (Garman,

23   2001c), this concern is somewhat mitigated. Certainly to be protective of public health, these

24   effects should be viewed as adverse until additional data either confirm or contradict that

25   conclusion.

26   In summary, two different analyses of the brain morphometry data from the 2001 "Effects

27   Study" (Argus Research Laboratories, Inc., 2001) yielded significant effects (i.e., alteration of

28   brain structures) of developmental exposure to ammonium perchlorate in drinking water at doses

29   of 0.01 mg/kg-day and higher in a mammalian (rat) model of neurodevelopment. These

30   alterations included a 23-39% increase in the size of the corpus callosum over controls in the

31   progeny of dams dosed with 0.01 to 1.0 mg/kg of ammonium perchlorate in drinking water.

1   Alteration of brain structures in a laboratory animal model is considered to be an adverse

2   neurotoxic effect (U.S. Environmental Protection Agency, 1998b). One of the analyses used a

3   series of t-tests; the other a more conservative multivariate analysis employing a nested model

4   profile analysis followed by univariate analysis of specific brain regions. The latter method is

5   more likely to be considered a valid analytic method because it better incorporates the design

6   elements of the study and reduces the likelihood of Type I statistical error. These effects on brain

7   morphometry dictate a designation of 0.01 mg/kg-day as a LOAEL.

8

9

5.4   DEVELOPMENTAL STUDIES

11   The 1997 testing strategy included a developmental study in rabbits to evaluate both a

12   potential critical effect and to characterize the toxic effects of perchlorate in a species other than

13   rats. Testing guidance for developmental toxicity typically requires data in two different species.

14   A new study of developmental toxicity in rats was recommended at the 1999 external peer

15   review. This section reviews the historical data on the developmental effects of perchlorate

16   (5.4.1), the 1998 study in rabbits (5.4.2), and the new 2000 study in rats (5.4.3).

17

5.4.1 Historical Studies

19   Brown-Grant (1966) examined the effects of perchlorate on implantation and pregnancy

20   outcome in Wistar rats. Potassium perchlorate or potassium chloride (control) was administered

21   at 1.0% (w/v) in drinking water from GD2 through GD8. The daily calculated intake rates were

22   237 and 371 mg/rat for potassium perchlorate and potassium chloride, respectively. Rats were

23   administered methythiouracil 45 min before injection of 5 μCi sodium radioiodide (^{131}I) and

24   sacrificed 2 h later. Rats clearly not pregnant were sacrificed on Day 20; whereas, pregnant rats

25   were allowed to deliver prior to sacrifice. Pregnancy was successful in 7/11 control rats and

26   8/11 perchlorate-treated rats. Among nonpregnant animals, implantation sites were not found.

27   Litter size, number of pups, and pregnancy were not affected.

28   In the same study, false pregnancy was induced by mating females with vasectomized

29   males. Females were dosed as before on GD2 through GD8 to 0.25 or 1.0% potassium

30   perchlorate or potassium chloride (control). These doses correspond to 63 and 246 mg potassium

1	perchlorate/rat and 82 and 308 mg potassium chloride per rat, respectively. Deciduoma
2	formation was induced through traumatizing one uterine horn while under anesthesia. Rats
3	exposed to the 0.25% dose were traumatized on GD3 and sacrificed on GD7. Trauma and
4	sacrifice occurred on GD4 and GD8, respectively, in the 1.0%-dose group. Methylthiouracil and
5	sodium radioiodide (^{131}I) were administered prior to sacrifice as before. Deciduoma formation
6	was not different between dosed and control rats. Thyroid weights were increased significantly
7	in the rats of the 1.0% potassium perchlorate-dose group.
8	A related study was performed by Brown-Grant and Sherwood (1971). Wistar rats were
9	mated shortly post-partum, and the present litter was culled to nine. The dams were then
10	administered 0.1% potassium iodide or 1.0% potassium chloride, potassium perchlorate, or
11	potassium iodide in the drinking water until sacrifice. The average daily intake of potassium
12	perchlorate and potassium chloride was 615 and 655 mg/rat, respectively; calculated daily doses
13	were approximately 2,440 and 2,660 mg/kg body weight. The litters were sacrificed on GD9 or
14	GD10. The dams then were sacrificed on GD12 or GD13, allowing time for the new blastocysts
15	to implant. Potassium perchlorate again did not affect blastocyst ability to survive prior to
16	implantation or implantation rate after lactation ceased. Relative thyroid weights of the dams and
17	litters were increased significantly compared with potassium-chloride-dosed controls. The high
18	dose of potassium iodide (average daily intake of 234 mg/rat [approximately 1,150 mg/kg]) was
19	maternally toxic.
20	All dams were sacrificed on Day 12 or 13 and examined for the number of implantation
21	sites. There was 100% incidence of dams with implantation sites for all groups except the
22	perchlorate-treated group in which only 70% of the dams had implantation sites. The number of
23	implantation sites per dam was comparable for all groups. Thyroid weights in the perchlorate-
24	treated dams appeared to be increased compared with the chloride- or iodide-treated dams. Also,
25	thyroid weights of the offspring of perchlorate-treated dams were increased compared with
26	offspring from iodide-treated dams. The authors concluded that treatment with potassium
27	perchlorate had no significant effect on blastocyst survival or the ability to implant under
28	conditions delaying implantation (i.e., concurrent lactation).
29	Postel (1957) reported administration of 1% potassium perchlorate in drinking water to
30	pregnant guinea pigs (n=16) and a control group (n = 3) receiving a diet of 0.48 μg iodine per
31	gram. Dosing with perchlorate during GD21 through GD48 produced enlarged thyroids in the

1 fetuses compared to the thyroids of control fetuses. In contrast, perchlorate treatment did not

2 have any effect on the thyroids in dams. Enlarged fetal thyroids also occurred when perchlorate

3 treatment was accompanied by daily subcutaneous treatment with T3 doses as high as

4 32 μg/kg/day. From water intake and body weight data, the author calculated an average daily

5 dose to the dams of 740 mg/kg-day. The fetuses were not examined for other developmental

6 effects. This study suggested a free-standing LOAEL of 740 mg/kg-day for fetal thyroid

7 enlargement because no other doses were tested. In a separate experiment to test effects on adult

8 guinea pigs, 0 or 1% potassium perchlorate was administered to nonpregnant female guinea pigs

9 for 30, 60, or 90 days. Thyroid enlargement and hyperplasia were apparent in treated animals

10 after 60 or 90 days of treatment.

11   Similar results in rabbits were described by Lampe et al. (1967). Dams were dosed with

12 100 mg potassium perchlorate/kg by weight daily, mixed with feed. Dosing occurred from

13 conception through GD21 or GD28. Maternal thyroid weights in treated animals were three

14 times higher than control thyroids; fetal thyroids were nearly four times the control weights. The

15 number of epithelial cells were increased, and the amount of colloid decreased in treated animals.

16 The relative volume of the stroma, the supporting matrix, was increased because of the reduced

17 follicle sizes. Likewise, maternal thyroids showed decreased luminal size and increased

18 epithelial cells. The authors asserted that these results demonstrated that the placenta is

19 permeable to perchlorate. Because fetal thyroids were more enlarged relative to maternal thyroid

20 glands, the fetal thyroid system is independent of the maternal regulatory system and more

21 sensitive to changes in iodine availability.

22

23 ## 5.4.2 Segment II Developmental Toxicity Study in Rabbits

24   A developmental toxicity study was performed in New Zealand White (Hra:[NZW]SPF)

25 rabbits as part of the overall perchlorate testing strategy (Argus Research Laboratories, Inc.,

26 1998c). This study has also appeared in the literature (York et al., 2001a); however, because that

27 manuscript did not use the PWG review of thyroid histopathology and its conclusions on other

28 endpoints are the same as the contract report, the manuscript will not be discussed further in this

29 document. To aid understanding of terminology and the protocol, a schematic of the study

30 design is provided in Figure A-3 of Appendix A to this document. The study design meets the

31 requirements of the 1998 EPA Office of Pollution Prevention and Toxic Substances (OPPTS)

1 870.3700 guideline. A deviation from the use of double staining was noted in Appendix D of the

2 Argus report, but EPA determined that this should not have had an effect on the overall outcome

3 of this study.

4   The dose groups tested were 0, 0.1, 1.0, 10, 30, and 100 mg/kg-day of ammonium

5 perchlorate in RO water provided by continual access on presumed GD6 to GD28. Each group

6 was comprised of 25 time-mated does assigned on a randomized basis stratified by weight.

7 Doses were selected on the basis of a dose range-finding study (Argus Research Laboratories,

8 Inc., 1998d) in which thyroid histopathology was evident in the does at 20, 50, and 100 mg/kg-

9 day; thyroid hormone levels (T3, T4, and TSH) in the does were reduced at all doses; and three

10 malformed fetuses from three litters in the 20-mg/kg-day group were observed upon gross

11 external examination. EPA was concerned about these pilot study results, particularly because

12 the original target doses of 0.1 and 10 mg/kg-day were changed on GD13 to 50 and 100 mg/kg-

13 day based on the lack of clinical toxicity at these doses. The fact that these were the doses at

14 which effects were observed, together with the fact that a low number of animals (n = 5) was

15 used in this range-finding study caused EPA to counsel the sponsor (PSG) to examine an

16 expanded range of doses in the definitive study. The dose groups chosen for the definitive

17 developmental study were thus aimed to bracket the dose levels in the range-finding study and to

18 go below the doses causing thyroid hormone perturbations and above those associated with the

19 fetal malformations.

20   Dosing solutions of ammonium perchlorate were prepared at least weekly from stock

21 solution, and the results of the concentration analyses were within acceptable ranges. Stability of

22 solutions was assumed based on determinations by AFRL/HEST for the 90-day bioassay as

23 discussed in Section 5.2.3. Rabbits were observed for viability at least twice daily, and body

24 weight, food and water consumption, clinical observations, deaths, abortions, and premature

25 deliveries were evaluated daily. On GD29, rabbits were terminated and cesarean sections were

26 performed. Blood samples from the does were taken for evaluation of thyroid and pituitary

27 hormones (T3, T4, and TSH). Gross necropsy was performed on the thoracic, abdominal, and

28 pelvic viscera of each doe. Parameters evaluated in the does included pregnancy status, gravid

29 uterine weight, number of corpora lutea in each ovary, number and distribution of implantations,

30 early and late resorptions, and live and dead fetuses. The thyroids/parathyroids were evaluated

31 histologically. Weight, gross external alterations, sex, in situ brain status (in one-half of the

1 fetuses in each litter), brain histology (in the other one-half of all fetuses in each litter), cavitated

2 organs, and skeletal and cartilaginous alterations were examined in the fetuses. No

3 measurements of thyroid structure or function were made in the fetuses.

4

5 **5.4.2.1   Results of Maternal Examinations and Thyroid Histopathology**

6 Two does in the 1.0-mg/kg-day group aborted either dead pups or had late resorptions on

7 GD28. Both of these abortions were considered unrelated to treatment because the incidences

8 were not dose-dependent and were consistent with historical control data for rabbits in that

9 laboratory (Argus Research Laboratories, Inc., 1998c; Appendix J). One doe in the 100-mg/kg-

10 day group delivered prematurely on GD27 (normal delivery in rabbits occurs on GD31), but it

11 was assumed that this rabbit had been identified and shipped incorrectly by the supplier because

12 the pups appeared to be full-term (i.e., they had fur and were nursing). There were no treatment-

13 related effects on maternal clinical signs, body weight, body weight change, gravid uterine

14 weight, or food and water consumption. It is interesting to note that there were decreases (not

15 statistically significant) in several of these endpoints, at the 1.0-mg/kg-day group–the same at

16 which the abortions occurred–as did one adverse necropsy observation of a mottled liver.

17 However, none of these responses showed a dose-response with the current treatment regimen,

18 and none were out of the range of normal occurrence.

19 The only remarkable histopathology in the does was observed in the thyroids. There was

20 an apparent dose-related but not statistically significant decrease in thyroid weight). The

21 histopathology in the dams as reviewed by the PWG can be found in Wolf (2000; 2001,

22 Table 22). There was a clear dose-response for colloid depletion, hypertrophy, and hyperplasia,

23 indicating that another species has conserved the hypothalamic-pituitary-thyroid feedback

24 regulation. All three indices appeared to be significantly increased at 1.0 mg/kg-day and above.

25 Benchmark dose analyses resulted in BMDL estimates of 0.008 for colloid depletion and 0.42 for

26 hyperplasia. A poor fit prevented BMDL estimation for hypertrophy.

27

28 **5.4.2.2   Developmental Endpoints**

29 There were no treatment-related effects on gross external endpoints (Argus Research

30 Laboratories, Inc.,1998c, Table 16). With regard to soft tissue anomalies (Argus Research

31 Laboratories, Inc.,1998c, Table 17), there were several occurrences of lung lobe and gallbladder

1   absence, but their incidence was not treatment related. The statistically significant decrease in

2   folded retina was attributed to an artifact of tissue processing. There were no treatment-related

3   effects in skeletal or ossification alterations (Argus Research Laboratories, Inc.,1998c, Tables 18

4   and 19), and no indication of an increased incidence of the more apical endpoint (i.e., any

5   skeletal change). The fetal NOAEL thus is identified as greater than 100 mg/kg-day for embryo-

6   fetal developmental toxicity, other than that which may have occurred in the thyroid.

7

8   **5.4.2.3   Maternal Thyroid and Pituitary Hormone Analyses**

9   The thyroid and pituitary hormone (T3, T4, and TSH) analyses were performed by

10   AniLytics, Inc., for the does in the developmental rabbit study (Argus Research Laboratories,

11   Inc.,1998c). Assays for T3 and T4 were performed using RIA kits according to manufacturer's

12   standard procedures. Assay kits from the same batch number and with the same expiration date

13   were used for the T3 and T4 measurements for each rabbit. The TSH assay was a

14   double-antibody, RIA procedure developed for rabbits and performed by AniLytics, Inc. The

15   analyses discussed in the Argus Research Laboratories, Inc. (1998c) report contain data from

16   both pregnant and nonpregnant rabbits, with both groups combined in the analyses. Because of

17   the known effects of pregnancy on thyroid hormones, EPA decided to reanalyze separately the

18   data from the pregnant and nonpregnant animals. However, EPA determined that the analyses

19   for nonpregnant animals were not useful because of the very limited number of subjects per

20   group (final number of does: n = 3, 1, 0, 1, 1, and 1 nonpregnant does/group, and n = 22, 24, 25,

21   24, and 23 pregnant does/group for the 0.0, 0.1, 1.0, 10, 30, and 100 mg/kg-day groups,

22   respectively). Therefore, EPA conducted reanalyses for these two groups separately (Crofton,

23   1998h). All data were taken from Appendix I of the report (Argus Research Laboratories,

24   Inc.,1998c). The analyses used the pregnancy status data subsequently submitted (York, 1998e).

25   Data from dependent measures (T3, T4, and TSH) were subjected to separate one-way ANOVA

26   tests with treatment (dose) as the independent between-subjects variable as calculated in Crofton

27   and Marcus (2001) and Marcus (2001). Mean contrasts were performed using Duncan's

28   Multiple Range Test.

29   The main effect of treatment was not significant for T3. The T3 data are plotted in

30   Figure 5-16A. There was a main effect of treatment and a significant difference between group

31   means for the control versus 1.0, 10, 30, and 100 mg/kg-day groups on T4. These data are
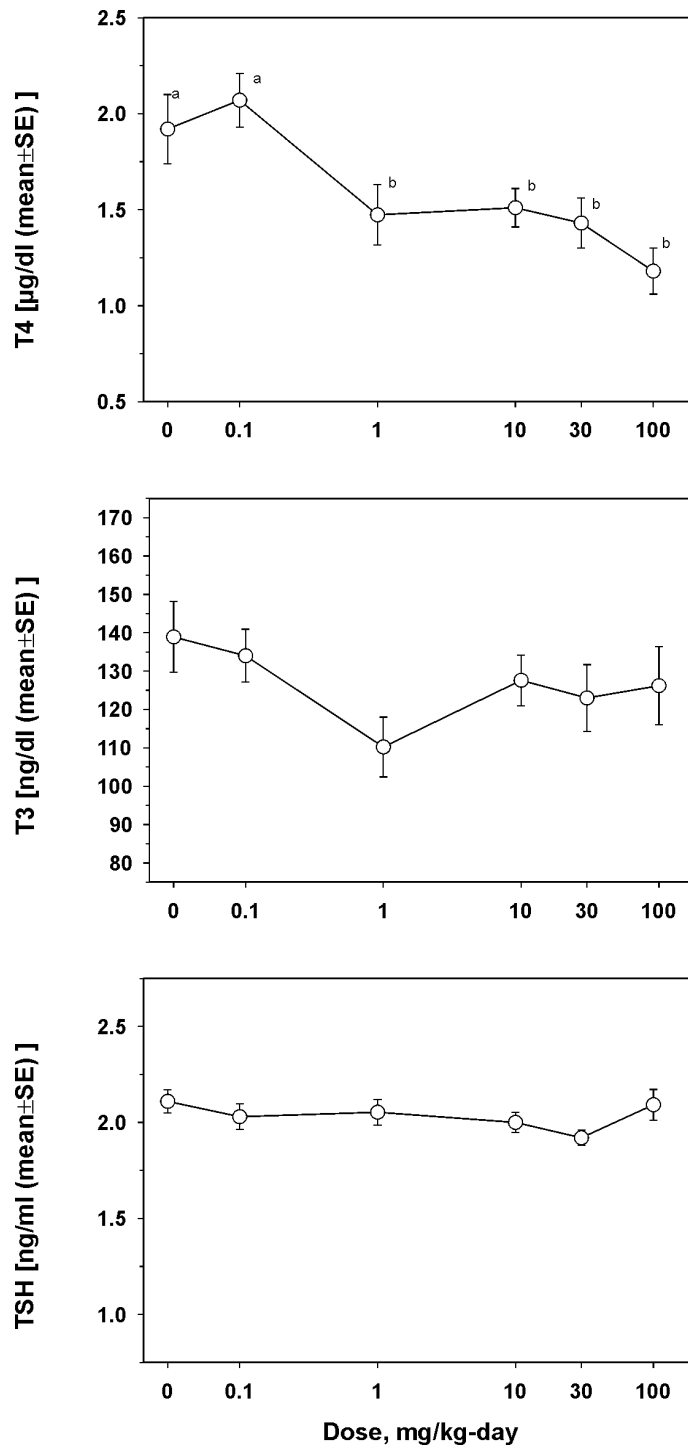
Figure 5-16. Effects from ammonium perchlorate in drinking water administration in pregnant New Zealand rabbits during GD6 to GD28 on T3 (A), T4 (B) and TSH (C) concentrations (ng/dL; mean ± SE) as recalculated in Table 5-2 (Crofton and Marcus, 2001). Data of Argus Research Laboratories, Inc. (1998c). Means with different letters were significantly different (p<0.05). Daily dose was estimated from water consumption data.

1   plotted in Figure 5-16B. The main effect of treatment was not significant for TSH

2   (Figure 5-16C). Results of these EPA reanalyses are different from those stated in the report.

3   The report (Argus Research Laboratories, Inc., 1998c) states that the NOAEL for T4 was

4   10 mg/kg-day. The current EPA analyses excluding nonpregnant animals, demonstrate a

5   NOAEL at 0.1 mg/kg-day for T4. There was no statistical significance of any dose on T3 or

6   TSH.

7   The lack of effect of any dose of perchlorate on T3 and TSH is difficult to explain. One

8   must note that these data are from rabbits (the majority of other data are from rats) and that the

9   data were collected 1 day prior to birth from the maternal compartment (whereas, all other data

10   were collected in adults or from postnatal day time points). In a previous study in guinea pigs

11   (Postel, 1957), enlarged thyroids were found in fetuses; whereas, there was no change in maternal

12   weight or histology. Lampe et al. (1967) demonstrated a larger effect on fetal thyroid weight

13   compared to maternal thyroid weights during late gestational exposure to perchlorate in rabbits.

14   These data warrant caution when comparing effects of perchlorate in the maternal with the

15   fetal/post-natal compartments.

16

5.4.3 Segment II Developmental Study in Rats

18   As recommended at the 1999 external peer review, a developmental study in addition to the

19   one in rabbits was performed in rats (Argus Research Laboratories, Inc., 2000). The EPA review

20   (Kimmel, 2000) was first performed on the audited final report (June 2000) and then on

21   clarifications provided by the principal investigator (York, 2000) that do not appear in the final

22   report.

23   Rats were given continuous access to target dosages of 0.01, 0.1, 1.0, and 30 mg/kg-day

24   ammonium perchlorate in deionized drinking water beginning at least 15 days before

25   cohabitation and continuing through the day of sacrifice. Each dosage group was comprised of

26   24 females, assigned on a random basis, stratified by weight. There were no maternal deaths.

27   Of these females, 20 were selected for evaluation; of these, 19, 19, 17, 20, and 20 were pregnant

28   in the 0, 0.01, 0.1, 1.0, and 30 mg/kg-day groups. The EPA OPPTS 870.3700 testing guidelines

29   recommend 20 pregnant animals per group at necropsy so that the power of the study to detect an

30   exposure-related response was somewhat lower.

1   All rats were sacrificed on day 21 of presumed gestation (GD21), and a gross necropsy of

2   the thoracic, abdominal, and pelvic visera was performed. Gravid uterine weights were recorded,

3   and the uterus then excised and examined for pregnancy, number and distribution of

4   implantations, live and dead fetuses, and early and late resorptions. The number of corpora lutea

5   in each ovary was recorded. Placentae were examined for abnormalities (size, color or shape).

6   Each fetus was identified, weighed and examined for sex and gross external alterations.

7   Approximately one-half of the fetuses in each litter were examined for soft tissue alterations.

8   The heads of these fetuses were examined by free-hand sectioning. The remaining fetuses in

9   each litter were examined for skeletal alterations and cartilage development.

10

11   **5.4.3.1   Results of Maternal Examinations**

12   Three dams in the 30 mg/kg-day group showed an increase in localized alopecia that was

13   statistically significant and was observed over 9-11 days during mid-late gestation. EPA feels

14   that this should be considered biologically significant and exposure-related despite the claim by

15   Argus Research Laboratories, Inc. (2000) and the study director (York, 2000) that such incidence

16   is within the range observed historically at their testing facility.

17   There were no other maternal parameters that were clearly supportive of exposure-related

18   effects. There was a statistically significant increase in corrected maternal body weight gain over

19   gestation in the 0.1 and 30.0 mg/kg-day groups, and an increase (not statistically significant) in

20   the 1.0 mg/kg-day group. There was also a reduction, again not statistically significant, in gravid

21   uterine weight in three of the four exposure groups. These latter changes may be associated with

22   reduced number of implants in the exposed groups (see below).

23

24   **5.4.3.2   Developmental Endpoints**

25   The Argus Research Laboratories, Inc. (2000) report (Table B17) did not record

26   preimplantation loss as an endpoint. EPA notes that there is an increase in this parameter over

27   control (12%) at each dose level: 0.01 (18%), 0.1 (20%), 1.0 (16%), and 30.0 (25%) mg/kg-day.

28   Whether this is statistically significant or biologically significant is unclear; although a decrease

29   in live fetuses in three of the four exposure groups that was significant at the highest dosage was

30   reported. Given the reduced power of this study to detect an effect, consideration was paid to

31   this finding. The lack of an effect on live fetuses at the 1.0 mg/kg-day level is not clear, and

1   these results by themselves are insufficient to establish an effect level below 30 mg/kg-day. EPA

2   recommends that preimplantation loss and embryo/fetal viability should be evaluated in any other

3   study reports on this chemical.

4   Ossification sites per litter for sternal centers and forelimb phalanges were significantly

5   reduced at 30 mg/kg-day, but Argus Laboratories, Inc. (2000) dismissed them as "reversible

6   developmental delays." EPA disagrees and contends that developmental delays, be they

7   permanent or reversible, are not to be discounted as potential indicators of developmental

8   toxicity. EPA additionally had some concern over the staining technique used for cartilage

9   (Kimmel, 2000) which was not accepted by Argus Research Laboratories, Inc. (York, 2000) as an

10 issue.

11

5.4.3.3 Conclusions Regarding Developmental Toxicity in Rats

13   Based on the review of the maternal and fetal data, EPA concludes that there are signs of

14 maternal and developmental toxicity at the 30.0 mg/kg-day level suggesting it as a LOAEL with

15 a NOAEL then at 3.0 mg/kg-day. While none of the results were so clear that a definitive

16 assessment can be made, the suggestive results are important to consider in light of the overall

17 data base and mode of action for the toxicity of perchlorate.

18

19

5.5 TWO-GENERATION REPRODUCTIVE TOXICITY STUDY

21   The 1997 recommendation to characterize the potential perchlorate toxicity on reproductive

22 parameters in a two-generation study was completed in 1999 (Argus Research Laboratories, Inc.,

23 1999). This study has also been reported in the literature (York et al., 2001b), but since that

24 manuscript did not use the PWG review of thyroid histopathology and its conclusions on other

25 endpoints are the same as in the contract report, the manuscript will not be discussed further in

26 this document. A schematic of the study design is provided as Figure A-2 of this document

27 (Appendix A) to aid understanding of terminology and the protocol.

28   The target doses (30 rats/sex/group) were 0, 0.3, 3.0, and 30 mg/kg-day of ammonium

29 perchlorate in RO water provided by continual access. Concentrations were adjusted based upon

30 actual water consumption and body weights recorded the previous week. Dosing solutions of

31 ammonium perchlorate were prepared weekly, and the results of concentration analyses were

1   within acceptable ranges (±10%) with one exception in the 3.0-mg/kg-day target group on May 5,

2   1998 (15.8%). The stock solution was prepared at least once, but the exact number of times was

3   not reported. Stability of solutions was assumed based on determinations by AFRL/HEST for

4   the 90-day bioassay, as discussed in Section 5.2.3.

5   On arrival, Spraque-Dawley rats were assigned randomly to individual housing, and

6   consecutive order was used to assign the P1 generation rats to cohabitation (one male rat per

7   female rat). The cohabitation period lasted a maximum of 14 days. Females with spermatozoa

8   observed in a vaginal smear or with a copulatory plug observed in situ were considered to be at

9   GD0 and assigned to individual housing. Estrous cycling was evaluated daily by examination of

10 vaginal cytology beginning 21 days before the scheduled cohabitation period and continuing until

11 GD0. The rats were observed for viability at least twice each day of the study and daily for

12 clinical signs. Body weights were recorded weekly during acclimation, on the first day of

13 dosage, weekly thereafter, and at scheduled sacrifice. Feed consumption and water consumption

14 values were recorded at least three times per week. Females were evaluated for duration of

15 gestation (GD0 to the day the first pup was delivered). Day 1 of lactation (LD1, post-partum)

16 was defined as the day of birth and was the first day on which all pups in a litter were weighed

17 individually. Maternal behavior was observed on LD1, 4, 7, 14, and 21. Rats that did not deliver

18 a litter were sacrificed on GD25 and examined for pregnancy status. Each litter was evaluated

19 for litter size (live and dead pups versus live pups only) and pup viability at least twice each day

20 of the 21-day post-partum period, and pups were counted daily. Deviations from expected

21 nursing behavior also were recorded. All F1-generation rats were weaned at the same age based

22 on observed growth and viability at LD21, unless required to be extended to LD28.

23   At the end of the 21-day post-partum period, all surviving P1 rats were sacrificed. Gross

24 necropsy was performed on all animals, and all gross lesions were examined histologically.

25 Organ weights were obtained for the thyroid, adrenal glands, brain, epididymides, heart, kidneys,

26 liver, ovaries, pituitary, prostate, seminal vesicles, spleen, and testes. The thyroids and

27 parathyroids were submitted for histopathological examination. Histopathology of other organs

28 was performed for the control and high-dose groups. Blood was collected for determination of

29 hormone levels (T3, T4, and TSH). Portions of the epididymides were used either for evaluation

30 of sperm count or motility. The left testis was homogenized after weighing for analysis of

31 spermatid concentration (spermatids per gram of tissue).

1   Pups not selected for continued evaluation in the study also were sacrificed on LD21.

2   Blood was pooled by sex per litter for analysis of T3, T4, and TSH. At least 3 pups/sex/litter

3   were necropsied and examined for gross lesions, including a single cross-section of the head at

4   the level of the frontal-parietal suture and examination of the head for apparent hydrocephaly.

5   Brain, thymus, spleen, and thyroid/parathyroid organ weights were obtained prior to fixation.

6   The adrenal glands, thyroid/parathyroid, kidneys, and liver were retained in formalin.

7

8   ## 5.5.1 General Toxicity Results and Evaluation of Reproductive Parameters

9   There was a statistically significant decrease in water consumption by males, but not by

10 females. The decrease with males and a smaller decrease with females were sufficiently small

11 that they are not considered to be biologically significant (Argus Research Laboratories, Inc.,

12 1999; Tables B5 and B6). There was a significant increase in ovarian weight at the 0.3-mg/kg-

13 day dose level only (Argus Research Laboratories, Inc., 1999; Table C26). There also was

14 slightly increased (not statistically significant) pituitary weight in females at the 0.3- and

15 3.0-mg/kg-day dose levels.

16 The fertility results are potentially of concern, but the statistical analyses did not show any

17 significant differences between groups for any of the tested parameters (Argus Research

18 Laboratories, Inc., 1999; Table C21 through C23). However, at 0.3 mg/kg-day, there were four

19 pairs that did not mate compared with one or two pairs in the other groups. Also at

20 0.3 mg/kg-day, there were three females that showed at least one signal of persistent diestrus and

21 one with persistent estrus (Argus Research Laboratories, Inc., 1999; Table C40). Incidences

22 were lower in all other groups. Only one of those females did not have evidence of mating, but

23 there were also four females that did not have evidence of mating in the 0.3 mg/kg-day group.

24 When mating and conception failures are combined, pregnancy rates were 28/30, 22/30, 26/30,

25 and 24/30 for the 0-, 0.3-, 3.0-, and 30-mg/kg-day groups, respectively. Of the females that were

26 pregnant, litter size was slightly lower at the 3.0- and 30-mg/kg-day dose levels, with values of

27 15.0, 14.9, 14.1, and 14.0 with increasing dose level. A similar trend was seen in the number of

28 implantation sites (15.8, 15.8, 15.0, and 15.0). None of these results were statistically significant

29 for the P1 generation, and the effect was not seen in the F1 generation. Consequently, this was

30 not considered a significant finding (Clegg, 1999; Rogers, 2000). Note should be made that

31 female intake of perchlorate during the last week of gestation was higher (Argus Research

1 Laboratories, Inc., 1999; Table C1). Additionally, in many of the perchlorate intake and feed

2 consumption summary data, observations were reported for a low numbers of rats, apparently

3 because of spillage.

4 In the F1 matings, all three perchlorate-dosed groups had a slightly higher fertility index

5 than did the vehicle controls, but this appears to be due to a control value that was low (Clegg,

6 1999; Rogers, 2000). These findings, the high dosage level of 30 mg/kg-day is designated as a

7 NOAEL for reproductive parameters (Rogers, 2000), a finding that is consistent with the

8 preliminary evaluation presented by EPA in 1999 (Clegg, 1999).

9

5.5.2 Evaluation of Thyroid Histology

11 The histopathology from the completed Argus Laboratories, Inc. (1999) two-generation

12 reproductive study was limited to the thyroid gland and can be found in Wolf (2001; Tables 14

13 through 21). In addition to the precursor lesion data (colloid depletion, hypertrophy, hyperplasia)

14 discussed in Section 5.5.2.1, Wolf (2000) noted that two animals from the high dose group (30

15 mg/kg-day) in the F1 generation (second parental generation, P2) in the study had adenomas and

16 one of these animals had two adenomas for a total of three. Although statistically significant

17 decreases in colloid were reported at both the 3.0 and 30.0 mg/kg-day dose levels (Argus, 1999),

18 none of the rats in the other groups (0, 0.3, 3.0 mg/kg-day) developed thyroid follicular cell

19 adenomas (0/30, 0/30, 0/30, respectively). These animals were dosed from conception to 19

20 weeks of age (adult male F1 rats). The tumors were considered to be treatment related (Wolf,

21 2000). Compared to the background incidence of thyroid follicular cell adenomas in male F344

22 rats after 2 years on study at 38/3419 from 67 NTP studies or 1.1% incidence at the 2-year end

23 sacrifice date, this study showed an incidence of 2/30 or 6.7% at 19 weeks. The tumors that

24 occurred in the F1 generation male rat pups at 19 weeks were considered particularly remarkable

25 (Wolf, 2000), and the EPA asked the NIEHS to review this incidence in context with the data

26 from the National Testing Program (NTP). The finding is especially of concern since three of the

27 F1 males in this high dosage group died of unknown causes (Rogers, 2000). This NIEHS

28 analysis of the tumor incidence is described below (Dunson, 2001b) in Section 5.5.2.2.

29

30

31

5.5.2.1 Thyroid Weight, Colloid Depletion, Hypertrophy, and Hyperplasia

Absolute thyroid weight was increased significantly in the P1 males at the 3.0- and 30-mg/kg-day dose levels. An increase was significant in females at 30 mg/kg-day. A significant increase in thyroid weight relative to both body weight and brain weight also occurred at 30 mg in both sexes (Argus Research Laboratories, Inc., 1999; Tables B11 through B13 and C26 through C28). The histopathology for the P1 generation as reported by the PWG can be found in Wolf (2000; 2001, Tables 14 and 15). All three indices (colloid depletion, hypertrophy, and hyperplasia) were present with a clear suggestion of an increase in females for colloid depletion and hypertrophy at 3 and 30 mg/kg-day that supported the thyroid weight changes. Hyperplasia was more prominent at 30 mg/kg-day. Benchmark dose analyses using the male and female data for the P1 generation combined (Table 5-1; Geller, 2001a) result in BMDL estimates of 0.11 mg/kg-day for colloid depletion and 2.44 mg/kg-day for hyperplasia. The data for hypertrophy resulted in inadequate model fit.

The F1-generation (second parental, P2 generation) rats also exhibited all three thyroid histopathological indices in a dose-related fashion with 3 and 30 mg/kg-day as effect levels (Wolf, 2000; 2001, Tables 16 and 17). Benchmark dose analyses (Table 5-1; Geller, 2001a) using the male and female data combined for the P2 generation estimate 0.90, 0.15, and 0.0004 mg/kg-day as the BMDL for colloid depletion, hypertrophy, and hyperplasia. Of note is the dramatic overlap between colloid depletion and hypertrophy in this generation. It was the males in these rats, exposed in utero and then sacrificed at 19 weeks, that showed the 3 adenomas.

The F1-generation weanling rat data are presented in Tables 18 and 19 (Wolf, 2000; 2001) and also exhibit the three thyroid histopathology indices increased at 3 and 30 mg/kg-day. Benchmark dose analyses (Table 5-1; Geller, 2001a) using the male and female data combined result in BMDL estimates of 0.80, 0.057, and 0.66 mg/kg-day for colloid depletion, hypertrophy, and hyperplasia. Again, the overlap among indices is present.

Data for the second weanling generation (F2) rats are presented in Wolf (2000, 2001; Tables 20 and 21). Decreased colloid and hypertrophy remain increased at 3 and 30 mg/kg-day, but hyperplasia was not remarkable. Benchmark dose analyses (Table 5-1; Geller, 2001a) only provided adequate fit to the hypertrophy data and resulted in a BMDL of 0.32 mg/kg-day.

1   Across the generations, this study results in a range of BMDL estimates (mg/kg-day) for

2   colloid depletion of 0.11 to 0.90, for hypertrophy of 0.057 to 0.32, and for hyperplasia of

3   0.0004 to 2.44. Of note is the low BMDL value for hyperplasia (0.0004 mg/kg-day) in the P2

4   generation, the same animals that exhibited tumors.

5

6   **5.5.2.2   Bayesian Analysis of Tumor Incidence**

7   In order to properly interpret the results from a given toxicological study, it is often

8   necessary to consider the data in light of additional information from outside of the study such as

9   the variability and average level of response for positive and negative controls in past studies that

10   are similar to the current study. It is also necessary to account for confounding effects that an

11   exposure may have on variables that are associated with the outcome of interest. For example, it

12   is important to adjust for animal survival to avoid bias in analyses of animal tumorigenicity

13   (McKnight and Crowley, 1984) and reproductive toxicity (Dunson and Perreault, 2001).

14   Typically, expert knowledge and information from related studies are accounted for only

15   informally in the interpretation of a statistically significant or non-significant result. However,

16   there are clear advantages to formally incorporating such extra information into the statistical

17   analysis because it can be very difficult to interpret statistical significance when some aspect of

18   the data is inconsistent with outside information (e.g., the control response is higher or lower

19   than typically seen in related studies). In addition, the formal incorporation of outside

20   information can improve sensitivity and limit bias when assessing toxicological effects. The

21   advantages of including historical control data, in particular, has been well documented in the

22   toxicological and statistical literature (Dunson and Dinse, 2001; Haseman, Huff, and Boorman,

23   1984; Ibrahim, Ryan and Chen, 1998; Tarone, 1982).

24   Although frequentist (i.e., non-Bayesian) hypothesis tests can sometimes incorporate

25   historical control data (see, for example, Tarone, 1982), outside information can be incorporated

26   more naturally and flexibly within a Bayesian analysis. In Bayesian analyses, the unknown

27   parameters in a statistical model are assigned prior probability distributions quantifying

28   uncertainty prior to observing data from a current study. For example, based on experience with

29   an assay system, a toxicologist may be 95% certain that the average level of response among

30   vehicle control animals is between bounds A and B with C being the most likely value. This

31   information can be formally incorporated into a Bayesian analysis through a prior distribution,

1 for a parameter measuring expected control response, which is centered on *C* and assigns 95%

2 probability to values between *A* and *B*. Alternatively, the prior distribution can be estimated

3 using data or summary statistics for control animals in historical studies if such information is

4 available (Ibrahim, Ryan and Chen, 1998; Dunson and Dinse, 2001). For parameters about

5 which little is known, noninformative or vague prior distributions that assign equal prior

6 probability to a wide range of plausible values can be chosen.

7   Bayesian inferences about toxicological effects can be based on the posterior distribution

8 for the parameters in the statistical model. The posterior distribution, which quantifies the

9 current state of knowledge about the unknown quantities in the statistical model, is obtained by

10 updating the prior distribution with the information in the data from the current study using

11 Bayes theorem (refer to Gelman et al., 1995 for an overview). One can use the posterior

12 distribution as a basis for conclusions about effects of interest by using posterior means, 95%

13 credible intervals, and posterior probabilities as Bayesian alternatives to the maximum likelihood

14 estimates, 95% confidence intervals, and p-values used in frequentist analyses. For example, as

15 an alternative to a p-value, one could calculate the posterior probability of an increase in the

16 proportion of animals with an adverse response in a treated group relative to the control.

17 Bayesian approaches have been developed for a wide variety of toxicological applications,

18 including risk assessment (e.g., Hill, 1996; Hasselblad and Jarabek, 1996), toxicokinetic

19 modeling (e.g., Bernillon and Bois, 2000), and analysis of skin papilloma data (Dunson et al.,

20 2000).

21   Without incorporating historical data on spontaneous neoplasms in Sprague-Dawley rats,

22 the difference between 0/30 in the vehicle control and 2/30 in the 30 mg/kg-day group is

23 non-significant by standard tests (e.g., Fisher's exact). However, the reported historical control

24 incidence of thyroid follicular adenomas for male Sprague-Dawley rats in two-year studies is

25 approximately 3-4% (Chandra et al., 1992; McMartin et al., 1992), suggesting that these tumors

26 should be extremely rare among 19-week old animals in the absence of a treatment effect.

27 Without formally incorporating this historical information into the statistical analysis through a

28 prior distribution, it is very difficult to assess the weight of evidence in favor of a treatment-

29 related increase in thyroid follicular adenoma incidence. A Bayesian approach was used to

30 assess the effect of ammonium perchlorate in drinking water on thyroid follicular cell adenoma

1 incidence in male Sprague-Dawley rats from the two-generation study (Argus Research

2 Laboratories, Inc., 1999).

3

5.5.2.2.1 Choosing prior distributions based on historical controls

5 The proportion of control male Sprague-Dawley rats developing thyroid follicular cell

6 adenomas in two-year carcinogenicity studies has been reported in the literature. Chandra et al.

7 (1992) reported a rate of 48/1340 (3.6%), and McMartin et al. (1992) reported a rate of 23/583

8 (3.9%). In order to incorporate this historical control data into our analysis of the effect of

9 ammonium perchlorate on thyroid incidence at 19 weeks of age, we follow a Bayesian approach.

10 The historical data can be summarized using a Beta (71,1852) prior distribution for the

11 probability of a male Sprague-Dawley rat developing a thyroid follicular cell adenoma (in the

12 absence of treatment with a test agent) by the time of natural death or sacrifice at two years. The

13 Beta prior is the standard choice for a prior distribution on a probability (c.f., Dunson and

14 Tindall, 2000 and Gelman et al., 1996 for further discussion of the Beta prior). The values

15 71 and 1923 are simply the numbers of control male Sprague-Dawley that did and that did not

16 develop thyroid follicular cell adenomas, respectively, from the Chandra et al. (1992) and

17 McMartin et al. (1992) articles.

18 To account for the fact that the Argus (1999) study recorded thyroid incidence at 19 weeks

19 and not at the time of natural death or at sacrifice at two years, a prior distribution for the ratio of

20 the probability of thyroid follicular cell adenomas at 19 weeks to the lifetime probability in a

21 two-year study was chosen. Portier, Hedges, and Hoel (1986) suggest that the probability of a

22 control male Fischer 344/N rat developing a thyroid follicular cell adenoma increases

23 approximately in proportion to age$^{4.78}$. Based on this estimate and on the average survival time

24 for male Fischer 344/N rats in the NTP historical control database (95.2 weeks), the prior

25 expectation for the ratio is $(19/95.2)^{4.78} = 5e-04$. Allowing for a high degree of uncertainty in this

26 prior expectation due to uncertainty in the Portier, Hedges, and Hoel (1986) estimate and in

27 extrapolation from Fischer 344/N rats to Sprague-Dawley rats, a Beta (0.11, 2.6) for the ratio was

28 chosen. This prior has median 5e-04 and 95% interval (0,0.379).

29

30

31

1 **5.5.2.2.2 Results of NIEHS analysis**

2   Using the prior described in the previous subsection and "updating" the prior with control

3 data from the Argus study (i.e., 0 tumors out of 30 control male rats), the NIEHS analysis

4 estimated that a control rat has a 0.15% chance of developing a thyroid follicular cell adenoma by

5 19 weeks (Dunson, 2001b). In addition, had perchlorate had no effect on the incidence of thyroid

6 follicular cell adenomas, the probability of observing two or more rats with these tumors out of

7 30 would be approximately 0.005. Thus, the data strongly support the hypothesis that

8 ammonium perchlorate in the drinking water at 30 mg/kg-day causes an increase in the incidence

9 of thyroid follicular cell adenomas.

10

11 **5.5.2.2.3 Summary of NIEHS analysis of tumor data**

12   Incorporating historical control data in a Bayesian analysis, a significant increase in thyroid

13 follicular cell adenoma incidence at 19 weeks in male Sprague-Dawley rats exposed to 30 mg/kg-

14 day relative to controls was found (Dunson, 2001b). There was no evidence of an increase at low

15 dose levels. This finding raises concern for in utero imprinting (i.e., that pups exposed in utero

16 are subsequently more susceptible to thyroid hormone perturbation during post-natal

17 development and adulthood), a phenomenon that is now appreciated in the endocrine disruption

18 arena (Prins et al., 2001; Phillips et al., 1998; Seckl, 1997).

19

20 ## 5.5.3 Thyroid and Pituitary Hormone Analyses

21   Thyroid and pituitary hormones were assayed in the P1-generation (both sexes), the

22 F1-generation adults, the F1-generation pups (PND21) and the F2-generation pups.

23   In the P1-generation, there was an unexpected and unexplained increase in T3 levels.

24 Effects on T4 and TSH were as expected, with a significant decrease in T4 and increase in TSH

25 at the 30 mg/kg-day level.

26   An anomalous increase in T3 was also reported in the F1-generation adults. Significant

27 ($p \leq 0.01$) decreases in T4 of the F1-generation adult males occurred at the high dosage but

28 increases ($p \leq 0.05$) at the mid-doses are unexplained; TSH in the adult males was significantly

29 increased ($p \leq 0.01$) at the 30 mg/kg-day level. Similar results were reported for the

30 F1-generation adult females.

1   In the F1-generation pups, the only statistically significant effects was an unexpected

2   decrease in TSH at the mid doses in the males and an increase in the females at the lowest.

3   Similarly seemingly spurious results were observed for the F2-generation pups.

4   Geller (1999b) presented the EPA analysis of thyroid hormones for this study for the

5   P1-and F1-generation using separate repeated-measures ANOVAs with treatment as the

6   independent between-subjects variable and sex as a within-litter repeated-measures variable.

7   Mean contrasts were performed using Tukey's Studentized Range (HSD) test. In order to correct

8   for multiple comparisons, the alpha for significance (for all interaction main effect tests) was

9   adjusted to 0.029 (alpha of 0.05 divided by the square root of the number of ANOVA tests).

10   In the P1-generation rats, there was a significant dose effect and dose by sex interaction for

11   both T4 and TSH. A NOAEL was identified for males only for T4 and TSH at a dose of

12   3.0 mg/kg-day.

13   In the F1-generation (weanling pups) on PND21, the contract laboratories reported a

14   decrease in TSH and an increase in T4. This effect was discounted by Argus Research

15   Laboratories, Inc. (1999) because the decrease was not dosage-dependent and because TSH

16   would be expected to increase and T4 to decrease. EPA found similar results with its analyses,

17   noting that the significant dose effect on female T4 data was due to an elevated level in the

18   0.3 mg/kg-day group relative to the high dose and also noting that the results were inconsistent

19   with the mode of action for perchlorate (Geller, 1999b).

20   A significant increase in TSH was found in the adult F1 (P2 generation) rats at 30 mg/kg-

21   day; a finding consistent with the tumors observed at this dosage, but T4 and T3 appeared to

22   have increased in a dose-dependent fashion. Again the reason for this disparity is not clear.

23

24

25   **5.6   IMMUNOTOXICITY STUDIES**

26   As discussed in Chapter 3, immunotoxicity studies were included in the perchlorate testing

27   strategy due to indications in humans and laboratory animals that perchlorate may affect immune

28   and hematological function. For example, a study by Weetman et al. (1984) that appeared as a

29   Letter to the Editor in *The Lancet*, investigated the effect of potassium perchlorate on human

30   T- and B-cell responses to mitogens *in vitro*. Perchlorate at concentrations of 0, 0.01, 0.1, and

31   10 mmol/L (1.17 g/L) were tested in cultures of human peripheral blood lymphocytes. IgG and

1   IgM in culture superhatants were measured by ELISA after culture for 10 days with pokeweed

2   mitogen (PWM). Perchlorate at 0.1 to 10 mmol/L resulted in inhibition of PWM-induced LgG

3   production at 10 mmol perchlorate/L inhibited IgM production. Lymphocyte proliferation as

4   measured by ^{3}H-thymidine incorporation was reduced by 33 to 35% in cultures from five of six

5   individuals in the presence of the T-lymphocyte mitogen phytohemagglutinin (PHA). Weetman

6   et al. (1984) concluded that perchlorate had significant immunosuppressive activity on

7   lymphocytes at pharmacologically-relevant concentrations in the absence of cytotoxicity, the

8   latter of which was assessed by ethidium bromide/acridine orange fluorescence. Unfortunately,

9   no details were provided as to when viability was determined during the 10 days of lymphocyte

10   culture with perchlorate and PWM. While these and other results were not sufficient to infer that

11   perchlorate was immunosuppressive or had other immunotoxic effects, there was uncertainty

12   with respect to its potential to do so. It was not known whether this could be a direct effect of

13   perchlorate but could plausibly also be due to its anti-thyroid effects.

14   An array of 14- and 90-day experiments, to evaluate the effects of drinking water

15   administration of ammonium perchlorate on immunotoxicological and hematological parameters

16   were performed using female B6C3F1 (Keil et al., 1998; Kiel et al., 1999; BRT-Burleson

17   Research Technologies, Inc., 2000a,b,c,) or CBA/J Hsd mice (BRT-Burleson Research

18   Technologies, Inc., 2000a,b,c). Parameters also were evaluated 30 days after one 90-day study to

19   assess the reversibility on several observed effects. The mouse was chosen for these studies

20   because it is the typical experimental species for immunotoxicological studies. In addition, data

21   were collected on thyroid and pituitary hormones and thyroid histology to provide additional

22   insight on interspecies variability by comparison with results of the rabbit and rat studies

23   included in the testing strategy. The mice (8 to 10 weeks of age) were acclimated for 1 week

24   prior to initiation of any study. Ammonium perchlorate was obtained from the sponsor

25   (AFRL/HEST), and different lots were used for each of the major study groups (i.e., Keil, et al.,

26   1998; Keil et al., 1999; BRT-Burleson Research Technologies, Inc., 2000a,b,c,). Primary stock

27   solutions were prepared approximately every 1 to 2 months, and dosing solutions were prepared

28   weekly. In the Keil et al. (1998) studies, there was an indication of a trend that mice exposed at

29   the 30 mg/kg-day level consumed slightly less water on a weekly basis ($\approx$3 mL/week less than

30   control). Consequently, differences were noted in the actual exposure for the high-dose group in

31   the 14-day studies. This difference was not as marked in the 90-day studies. Concentration of

1   dosing solutions was verified by the sponsor (AFRL/HEST; data not shown). The one apparent

2   disparity in dose level (0.1 mg/kg-day; experiment not specified) was rectified after

3   reexamination of calculations (data not shown) (Keil et al., 1998). The mice were exposed to

4   levels of 0, 0.1, 1.0, 3.0, or 30 mg/kg-day in the Keil et al., (1998, 1999) studies; while in the

5   BRT-Burleson Research Technologies, Inc. (2000a,b,c) studies, the mice were exposed to levels

6   of 0, 0.02, 0.06, 0.2, 2.0 or 50 mg/kg-day. The doses were established based on the mean body

7   weight for each treatment group per week. Each dose group generally consisted of 6 to 10 mice,

8   with the exception that some control groups in the BRT-Burleson Research Technologies, Inc.

9   (2000a,b,c) studies had a group size of 20.

10   A number of 14-day experiments were conducted. In Experiments "C", "G", "I", "J", "T",

11   and "K" (Keil et al., 1998), the mice were sacrificed at Day 14; and body weight, organ weight

12   and cellularity (thymus, spleen, liver, and kidney), a number of immunotoxicology and

13   hematological parameters, thyroid histology, and thyroid and pituitary hormone levels were

14   measured. These data are summarized in Tables 3, 6, 9, 12, 14, 16, 18, and 21 of the "Final

15   Report" (Keil et al., 1999). In Experiments "U" and "V" (Keil et al., 1998), mice were

16   challenged with sublethal amounts (2,300 or 2,700 colony-forming units [CFU]) of *Listeria*

17   *monocytogenes* on Day 7 and then sacrificed on Day 14. The spleens were removed for a

18   delayed-type hypersensitivity (DTH) assay (Keil et al. 1999: Table 31). In experiments "H",

19   "F", and "M" (Keil et al., 1998), mice were challenged with P815 tumor cells by ip injection.

20   At the 14-day terminal sacrifice, spleens were removed for the cytotoxic T lymphocyte (CTL)

21   activity assay (Keil et al., 1999: Table 23).

22   A series of 90-day experiments also were conducted. In Experiments "A", "D", and "N"

23   (Keil et al., 1998), mice were sacrificed after 90 days; and body weight, organ weight and

24   cellularity (bone marrow, thymus, spleen, liver, and kidney), a number of immunotoxicology and

25   hematological parameters, thyroid histology, and thyroid and pituitary hormone levels were

26   measured (Keil et al., 1999: Tables 4, 7, 10, 13, 15, 17, 19, 20, and 22). In Experiments "B" and

27   "E" (Keil et al., 1998), these same endpoints were measured after a 30-day recovery period (Keil

28   et al., 1999: Tables 5, 8, 11, and 22,). In Experiment "P" (Keil et al., 1998), mice were

29   challenged with P815 tumor cells by ip injection on Day 76. Spleens were removed at terminal

30   sacrifice for the CTL activity assay (Keil et al., 1999: Table 24).

1   Two host resistance models, one a bacteria and the other a tumor, were used in 90-day

2   experiments. Mice in Experiment "L" (Keil et al., 1998) were challenged with *Listeria*

3   *monocytogenes* by iv injection. At terminal (90-day) sacrifice, spleens and livers were removed

4   and cultured for *L. monocytogenes* growth. Unfortunately, the challenge concentration (i.e.,

5   5360 CFU) of bacteria used was excessive, thereby prohibiting enumeration of the bacteria in the

6   spleens of these mice. A second 90-day *L. monocytogenes*-challenge experiment (Keil et al.,

7   1999) was subsequently undertaken using a lower (i.e., 2700 CFU) challenge concentration (see

8   Keil et al., 1999: Table 34). For the tumor model, in Experiments "Q" and "O" (Keil et al.,

9   1998), mice were challenged with B16F10 tumor cells by iv injection on Day 76. At the 90-day

10   sacrifice, the lungs were removed, and the number of tumor nodules in both lungs were

11   enumerated (Keil et al. 1999: Table 33).

12   The IgM and IgG antibody responses to sheep red blood cells (SRBCs) of mice exposed to

13   ammonium perchlorate for 90 days and the IgM anti-SRBC response of mice exposed for 14 days

14   was determined using an enzyme linked immunosorbent assay (ELISA) (figures on page 59, Keil

15   et al., 1999). Based on EPA comments in 1998 and external peer review recommendation

16   (Research Triangle Institute, 1999), a second contract was let to determine the antibody response

17   to SRBCs using the more traditional antibody plaque-forming cell (PFC) assay (BRT-Burleson

18   Research Technologies, Inc., 2000a,b,c). Unlike the ELISA, which measures SRBC-specific

19   IgM antibody in serum, the PFC assay quantifies the number of plasma cells in the spleen which

20   produce SRBC-specific IgM. The potent immunosuppressant cyclophosphamide (CP) was used

21   as a positive control in these latter studies. In both the 14- and 90-day studies, mice were

22   immunized iv with SRBCs 4 days prior to assay. The positive control mice were injected ip with

23   15 mg/kg-day CP on the last 4 days of dosing prior to assay.

24   Concern about potential effects of ammonium perchlorate on contact hypersensitivity, also

25   raised at the 1999 external peer review, were addressed in studies performed by Burleson et al.

26   (2000). Eight-week-old female CBA/J Hsd mice that had been acclimated one week prior to

27   dosing were exposed to 0, 0.02, 0.06, 0.2, 2.0, or 50.0 mg/kg-day for 14 or 90 days. The contact

28   sensitizer, 2,4-dinitrochlorobenzene (DNCB), was applied to the surface of both ears on days 9,

29   10 and 11 in the 14-day study, and on days 92, 93, and 94 in the 90-day study. Mice were

30   assayed using the local lymph node assay (LLNA) on day 14 and 97 for the 14-day and 90-day

1   studies respectively. A CP-positive control group was included in each study, with

2   administration of 15 mg/kg-day CP ip for 5 consecutive days prior to assay.

3   Data from the Keil et al. (1998, 1999) studies were analyzed as follows. Initially, analysis

4   of variance was performed using Tukey's multicomparison ($p < 0.05$) for the various parameters

5   measured. A Fisher's multicomparison test was used in previous interim reports but not in the

6   final one. The previous analyses reported effects. Consequent to criticisms of the analyses

7   performed, as stated in the previous external review Draft Toxicological Review Document on

8   Perchlorate (U.S. Environmental Protection Agency, 1998d), and reinforced by the comments of

9   Dr. Kimber White at the previous external peer review (Research Triangle Institute, 1999), these

10   and new data (i.e., the 14-day antibody response to SRBCs and the 90-day host resistance to

11   *L. monocytogenes*) were analyzed as indicated in the "Final Report" (Keil et al., 1999). That is,

12   data were combined from two or three experiments and evaluated by the Kolmogorv-Smirnov

13   test for normality and Bartlett's test for homogeneity of variance. If data displayed a normal

14   distribution and equal variance, two-way ANOVA, with experiments and treatments as factors,

15   was performed. Tukey's pairwise comparison was performed to determine differences ($p<0.05$)

16   between control and treatments if no interaction was identified due to combining multiple

17   experiments. If a significant interaction was identified in the ANOVA, data from each

18   experiment were analyzed using one-way ANOVA and Tukey's pair wise analysis. The Kruskal-

19   Wallis test was used if data were not normally distributed or variances were not equal; and if

20   significant, the Mann-Whitney test was employed to determine differences ($p<0.05$) between

21   control and treatments.

22   The results of the BRT-Burleson Research Technologies, Inc. (2000a,b,c) studies were

23   analyzed as follows. Data from each treatment group were compared by first performing a

24   Bartlett's Chi-Square test for variance of homogeneity. If found to be non-significant, ANOVA

25   was employed using dose. If significant, then Dunnett's *t*-test was performed, with $p<0.05$ being

26   significant. On the other hand, if Bartlett's Chi-square was significant, the non-parametric

27   Kruskal-Wallis test was performed, which if significant was followed by a Jonckheere's-Terpera

28   test for dose-dependent trends. The parametric ANOVA and the non-parametric extended

29   Cochran-Mantel-Haenszel test were performed to determine whether the data could be pooled.

30   Results for the general toxicity and organ weight measures will be discussed in

31   Section 5.6.1. Thyroid histopathology evaluations will be reported in Section 5.6.2, and analyses

1   of T3, T4, and TSH in Section 5.6.3. Results for the immunotoxicological and hematological

2   parameters are discussed in Sections 5.6.4 and 5.6.5. A summary of the results and their

3   potential significance is presented in Section 5.6.6.

4

5.6.1 Results for General Toxicity, Organ Weight, and Cellularity Measures

6   There were no effects observed on body, thymus, spleen, liver, or kidney weights in the

7   14-, 90-, or 120-day studies (Keil et al., 1999: Tables 6-8). Earlier interim reports indicated

8   considerable variability in the splenic and thymic cellularity of ammonium perchlorate-exposed

9   mice. This variability was due, in large part, to technical errors. Recognizing this, the contractor

10   performed additional studies (i.e., "on at least two or more occasions") in which "no significant

11   changes in cellularity were observed." (Keil et al., 1999). As such, in the "Final Report" no

12   consistent alteration in splenic or thymic cellularity was observed in the 14–, 90-, or 120-day

13   studies (Keil et al., 1999:   Tables 9-11), nor in splenic lymphocyte CD4/CD8 subsets (Keil et al.,

14   1999: Tables 14 and 15). With the exception that CD4-CD8+ thymic lymphocytes were

15   increased in mice exposed to 0.1- and 1.0-mg/kg-day doses in the 14-day experiment, there were

16   no other alterations in thymocyte subsets observed in the 14- or 90-day studies (Keil et al., 1999:

17   Table 12).   Furthermore, there were no alterations in the number of peritoneal macrophages

18   obtained from mice exposed to any doses of ammonium perchlorate in the 14-, 90-, and 120-day

19   studies (Keil et al., 1999: Tables 9-11), nor in bone marrow cellularity in the 14- and 90-day

20   studies (Keil et al., 1999: Tables 9 and 10). Due to the absence of effects in the latter studies, no

21   120-day study was performed.

22

5.6.2 Evaluation of Thyroid Histology

24   Thyroid histopathology evaluation was performed for two experiments (A and D) in the

25   Keil et al. (1998) study and eventually published in the final report (Keil et al., 1999). These data

26   were transmitted by Warren (1999), and a preliminary review by EPA was presented at the 1999

27   external peer review (Jarabek, 1999). The materials were provided to the PWG review, and the

28   results are found in Wolf (2000, 2001; Table 23). These results corroborate the preliminary

29   analyses that showed decreased colloid, follicular hypertrophy and hyperplasia to occur at the

30   30 mg/kg-day dose. Congestion in the intrafollicular capillaries and the nuclear to cytoplasmic

1 ratio of the follicular cells were not recorded by the PWG but were both noted in the Warren

2 (1999) report at 30 mg/kg-day (Jarabek, 1999). Hypertrophy was additionally observed in the

3 lower doses of experiment "A", and the reason for the disparity between the two studies is

4 unclear. These results support the assertion that the hypothalamic-pituitary-thyroid feedback

5 regulatory mechanism is conserved across species (rats, rabbits, mice and humans) and suggest a

6 NOAEL of 3 mg/kg-day in this strain of mouse.

7

8 ### 5.6.3 Thyroid and Pituitary Hormone Analyses

9   The report (Keil et al., 1998) contains thyroid hormone and thyrotrophin (TSH) data from

10 14- and 90-day exposures to ammonium perchlorate in B6C3F1 mice. The following is a

11 statistical analysis of the thyroid and pituitary hormone data (T4 and TSH) found in that report.

12 There were no data for T3 reported in the original study submitted to EPA (Keil et al., 1998).

13 The EPA reanalyzed the data that were supplied in Excel® spreadsheets to EPA by Dr. Deborah

14 Keil, and the data are published therein (Crofton, 1998i). Subsequent submission of additional

15 data files also containing data for T3 were included in reanalyses (Crofton, 2001a). Data for

16 dependent measures (T4 and TSH) were subjected to separate analyses. The T4 and TSH data

17 were analyzed with a two-way ANOVA, with duration (14, 90, and 120 days) and treatment

18 (dose) as the independent between-subjects variables as reanalyzed by Crofton and Marcus

19 (2001) and Crofton (2001a) as reported in Table 5-2. Mean contrasts were performed using

20 Duncan's Multiple Range Test.

21   Results of these EPA reanalyses are different from those stated in the Keil et al. (1998)

22 report. The EPA reanalysis of the T3 data (Crofton, 2001a) found main effects of time and

23 treatment, but no time-by-treatment interaction. Mean contrast testing showed a LOAEL of

24 0.1 mg/kg-day; however, the dose-related decrease was not linear. The 0.1 and 3.0 mg/kg-day

25 doses differed from controls but the 1.0 and 30.0 mg/kg-day doses did not. There was a

26 significant time-by-treatment interaction for T4. After 14 days of exposure there was no effect

27 with a NOAEL at 30 mg/kg-day; whereas, after 90 days of exposure the LOAEL was 0.1 mg/kg-

28 day. T4 recovered after 30 days postexposure. There was no effect of perchlorate on TSH

29 concentration contrary to the changes in histopathology discussed in Section 5.6.2.

30

1	These effects are of interest in that they demonstrate effects in mice comparable in nature to
2	that in rats and indicate that the hypothalamic-pituitary-thyroid feedback system is conserved
3	across species.
4	

5.6.4 Results of Immune Function Assays

6	No consistent alteration in CTL activity was observed in three 14-day studies ("M", "H",
7	and "F", Keil et al., 1998). No effects were observed on CTL activity in Experiments "M" and
8	"H". However, in Experiment "F", increases in CTL activity were observed at the 0.1-mg/kg-day
9	ammonium perchlorate dose for effector to target cell (E:T) ratios of 100:1, 30:1, and 10:1, and,
10	at the 1- and 3-mg/kg-day doses, for an E:T ratio of 10:1. In a 90-day study ("P", Keil et al.,
11	1998) there were no alterations in CTL activity at any dosages or E:T ratios. The variability and
12	inconsistencies observed in the early interim reports were ascribed to "technical issues" that were
13	consequently "corrected". In fact, the data presented in Tables 23 and 24 (Keil et al., 1999)
14	which includes data for dexamethasone, a potent immunosuppressant and positive control,
15	indicates that there were no effects of ammonium perchlorate AP exposure on CTL activity.
16	There was also no consistent alteration in the DTH response, as measured by the
17	lymphoproliferation (LP) of splenic lymphocytes from *L. monocytogenes*-challenged mice
18	incubated with soluble *Listeria* antigen (SLA) in two 14-day studies ("U" and "V", Keil et al.,
19	1998). The LP response was increased only in cultured splenic lymphocytes from mice in the
20	30-mg/kg-day group stimulated with 0.1 μg/mL SLA in Experiment "U" and in cultures of
21	splenic lymphocytes from mice in the 3-mg/kg-day group stimulated with 8 μg/mL SLA in
22	Experiment "V" (Keil et al., 1998). The "Results Summary and Status" page of Keil et al. (1998)
23	indicates that a 90-day DTH study was planned. These 90-day data and a summary of the 14-day
24	data are presented in Tables 32 and 31 respectively, of the "Final Report" (Keil et al., 1999). The
25	data indicated an enhanced LP response in mice dosed at 30-mg/kg-day in both the 14-day and
26	90-day studies.
27	No alteration in splenic natural killer (NK) cell activity was observed in two 14-day studies
28	("G" and "T", Keil et al., 1998). The 14-day Experiment "T" data are presented in a table;
29	however, the raw data and statistics for this study were not found in the submission. Inconsistent
30	results were obtained in two 90-day studies ("D" and "N", Keil et al., 1998) in which NK cell
31	activity was increased at the 30-mg/kg-day ammonium perchlorate in Experiment "N"; however,

1   no effects were observed at any doses in Experiment "D". A similar increase in NK cell activity

2   at the 30-mg/kg-day dose was observed in the 120-day Experiment "E" (see also the data in

3   Tables 21-22, Keil et al., 1999, in which the positive control dexamethasone was employed).

4   The lack of any change in the number of B16F10 tumor nodules in the lungs of mice from the

5   90-day "Q" study (Keil et al., 1998; see also Table 33, Keil et al., 1999), particularly at

6   30 mg/kg-day, suggests that the increased NK-activity does not reflect a significant biological

7   effect (see below). The EPA notes that there is a good deal of variation in NK activity data for

8   the controls in the 14-day "G" study, the 90-day "D" and "N" studies, and the 120-day "E" study,

9   which were 34, 6.4, 13.6, and 18.4 lytic units/10^7 splenic lymphocytes, respectively. Also, the

10   14-day "G" study was not included in Table 33 (Keil et al., 1999).

11   Decreased *in vitro* phagocytosis of *L. monocytogenes* was observed at 3 and 30 mg/kg-day

12   of ammonium perchlorate in the 14-day "C" and 90-day "A" studies (Keil et al., 1998). In the

13   90-day "N" study, macrophage phagocytosis was decreased in all dose groups. However, in the

14   14-day "G" and 90-day "D" studies and in two 120-day studies ("B" and "E"), no effect on

15   macrophage phagocytosis was observed (Keil et al., 1998). In the "Final Report" (Keil et al.,

16   1999), these alterations were confirmed (i.e., decreased phagocytosis at 1.0 and 30.0 mg/kg-day

17   in the 14-day study (Keil et al., 1999: Table 27) and decreased phagocytosis at 0.1, 1.0, 3.0, and

18   30.0 mg/kg-day in the 90-day study (Keil et al., 1999: Table 28). However, after a 30 day

19   recovery period (i.e., 120-day study, Keil et al., 1999: Table 29) phagocytic function was

20   comparable across control and treated mice. These data suggest that ammonium perchlorate

21   suppresses the phagocytic capacity of peritoneal macrophages *in vitro*, but that this suppression

22   may be reversed after a 30-day recovery period. Criticism of the use of an *in vitro* rather than an

23   *in vivo* assessment of macrophage function was raised in the 1998 EPA ERD document and at

24   the 1999 external peer review by Dr. Kimber White (Research Triangle Institute, 1999).

25   No consistent alteration in peritoneal macrophage nitrite production was observed in 14-,

26   90-, and 120-day studies. Increased nitrite production from macrophages cultured with interferon

27   (IFN) occurred at doses of 3 and 30 mg/kg-day and from macrophages cultured with IFN and

28   lipopolysaccharide for the 30-mg/kg-day dose in the 90-day "D" study (Keil et al., 1998). Also,

29   increased nitrite production from macrophages cultured with IFN was observed at 3 mg/kg-day in

30   the 90-day "N" study (Keil et al., 1998). An increase in nitrite production for macrophages

31   cultured with IFN or LPS alone also occurred for the 30-mg/kg-day group in the 120-day "B"

1   study (Keil et al., 1998). These data suggest a "trend" toward increased nitrite production at the

2   higher doses of ammonium perchlorate.

3   A subsequent analysis of these data, as presented in Tables 25 and 26 (Keil et al., 1999),

4   demonstrates "no significant difference in nitrite production of peritoneal macrophages" (Keil

5   et al., 1999).

6   A 90-day study ("L", Keil et al., 1998) was performed to determine if exposure of mice to

7   ammonium perchlorate results in alterations in resistance to infection with *L. monocytogenes*.

8   A trend toward increased resistance was suggested by the data; however, technical difficulties

9   were encountered. For example, there was variability in the number of *L. monocytogenes* CFU/g

10   liver recovered from control mice. It was not possible to enumerate the number of CFU/g spleen

11   in mice due to the high concentration of bacteria injected and also to an inadequate dilution of

12   spleen cell suspensions. In a subsequent 90-day study, mice were challenged with a lower

13   concentration of bacteria such that both the CFU/g liver and spleen could be determined. These

14   results, presented in Table 34 (Keil et. al., 1999), indicate that ammonium perchlorate exposure

15   does not alter resistance to infectious challenge to *L. monocytogenes*.

16   No effects were observed in an initial 90-day B6F10 tumor challenge host-resistance model

17   experiment ("Q", Keil et al., 1998). Another 90-day B6F10 tumor challenge experiment (i.e.,

18   "O") was performed, and the combined results of these two experiments are presented in

19   Table 33 (Keil et al., 1999). These data indicate that there were no differences in the number of

20   tumors present in the lungs of ammonium perchlorate-exposed mice compared with control mice.

21   Two separate groups of studies examining the effect that ammonium perchlorate has on the

22   antibody response to SRBCs were performed by independent contractors (Keil et al, 1999;

23   BRT-Burleson Research Technologies, Inc, 2000a,b,c). Initial studies were performed by Keil

24   et al. (1999), in which the IgM and IgG antibody responses were determined using ELISAs.

25   As indicated in the figures on page 59 (Keil et al., 1999), no change in the IgM levels in a 14-day

26   study, nor in the IgM and IgG levels in a 90-day study, was observed between control and any

27   ammonium perchlorate treated mice .

28   In the second set of studies, the anti-IgM SRBC PFC assay was employed (BRT-Burleson

29   Research Technologies, Inc, 2000a,b,c), using CP as a positive immunosuppressant control.

30   In the 14-day study there were no differences in the PFC response between control and treated

31   mice when expressed either as the number of PFC/spleen or PFC/10^6 spleen cells (BRT-Burleson

1   Research Technologies, Inc, 2000a,b,c: Figures 3 and 4). On the other hand, in the 90-day study

2   the PFC response was increased in the 2.0 and 50.0 mg/kg-day groups when expressed as the

3   PFC/spleen and increased only in the 50.0 mg/kg-day group expressed as PFC/10^6 spleen cells

4   (BRT-Burleson Research Technologies, Inc., 2000a,b,c: Figures 5 and 6). This disparity was not

5   due to any difference in splenic cellularity between the control and treated mice. In both the

6   14- and 90-day studies, CP significantly inhibited the PFC response, expressed either as

7   PFC/spleen or PFC/10^6 spleen cells compared to the controls.

8   The results of the effect that 14- and 90-day exposure to ammonium perchlorate has on the

9   development of a contact hypersensitivity response to DNCB, as determined by the LLNA,

10   indicate that an ammonium perchlorate dose as low as 0.06 mg/kg-day enhances this response.

11   In the 14-day study, the LLNA was increased at doses of 0.06, 0.2, and 50.0 mg/kg-day, but not

12   2.0 mg/kg-day (BRT-Research Technologies, Inc., 2000a,b,c: Figure 8). The results of the

13   90-day study were somewhat different in that, while the LLNA was enhanced at 0.06 and

14   0.2 mg/kg-day, it was suppressed at 50 mg/kg-day (BRT-Research Technologies, Inc., 2000a,b,c:

15   Figure 9). Another disparity between these two studies was that while CP suppressed the LLNA

16   in the 14-day study, it did not suppress this response in the 90-day study.

17

18   **5.6.5 Results for Evaluations of Hematological Parameters**

19   There were no differences observed between control and dosed mice in 14- or 90-day

20   experiments for erythrocyte cell count, hemoglobin, hematocrit, mean corpuscular volume, mean

21   corpuscular hemoglobin, and mean corpuscular hemoglobin concentration; nor in leukocyte

22   differential counts of neutrophils, monocytes, and lymphocytes. Because of the absence of

23   effects in these studies, no 120-day study was performed. No effects were observed in a single

24   14-day study (Experiment "T", Keil et al., 1998) on platelet counts. An increase in the

25   percentage of reticulocytes was observed in the peripheral blood of mice exposed to 3 mg/kg-day

26   of ammonium perchlorate in a 90-day study ("N", Keil et al., 1998). No other reticulocyte data

27   are available because of "the minimal availability of blood obtained from each mouse" in other

28   studies (Keil et al., 1998). In a subsequent 14-day study, there were no changes in the

29   hematological parameters examined between control and ammonium-perchlorate-treated mice

30   (Keil, et al., 1999: Table 16).

1	 No consistent alteration in the bone marrow stem cell assay was observed. An increase in

2	the number of colony-forming units was observed in bone marrow cell cultures from mice dosed

3	at 30 mg/kg-day in a 14-day study ("K", Keil et al., 1998). However, there was no effect of

4	ammonium perchlorate exposure on the stem cell assay in a 90-day study ("D", Keil et al., 1998).

5	In a subsequent 90-day study, while no alteration in the stem cell assay was observed between

6	control and ammonium perchlorate-treated mice, exposure to the positive control dexamethasone

7	resulted in suppression of the stem cell assay (Keil, et al., 1999: Table 20).

8

5.6.6 Results Summary

10	 The results of the various studies of immue function are summarized in Table 5-5.

11	Although innate (i.e., macrophage and NK cell function) and cell-mediated (i.e., cytotoxic

12	T lymphocytes [CTL], CD4, and CD8) immune functions were evaluated in the initial studies by

13	Keil et al, (1998), EPA noted that humoral immunity (i.e., B cells and antibody response) was not

14	(Smialowicz, 1999). The EPA suggested strongly that the antibody response to SRBCs is one of

15	the most commonly effected functional parameters in animals exposed to chemical

16	immunosuppressants (Luster et al., 1988). In fact, it is one of the assays required by EPA for test

17	rules. The EPA also requested that an additional 90-day *L. monocytogenes* host-resistance study

18	be undertaken consequent to technical problems associated with the initial 90-day study (Keil

19	et al., 1998). As such, the EPA felt that these data would provide a more comprehensive

20	evaluation of the potential for immunosuppression by ammonium perchlorate. In addition, the

21	EPA requested that thyroid histology and thyroid and pituitary hormone data be obtained in order

22	to provide additional insights on interspecies variability for this effect.

23	 Consequently, the sponsor and investigators, Keil et al. (1998), agreed to perform these

24	assays, the results of which are presented in the "Final Report" (Keil et al., 1999).

25	 Subsequent to receipt of the results of the antibody response to SRBCs (Keil et al., 1999),

26	in which antibody titers were expressed as the O.D. 50 or midpoint titer, rather than the more

27	conventional titer to achieve a 0.5 O.D., a second request to determine the potential effects of

28	ammonium perchlorate on the response to SRBCs was issued. In this same solicitation, the EPA

29	also requested that a sensitization test be performed. The results of these studies are found in

30	BRT-Burleson Research Technologies, Inc. (2000a, b, c).

TABLE 5-5. SUMMARY OF IMMUNOTOXICITY TEST RESULTS

Series/Strain/Sex (Study)	Exposures Period and Doses (mg/kg/d)	Endpoint	NOAEL/LOAEL Designations
Mouse/B6C3F1/Female (Keil et al., 1998;1999)	14-days 0, 0.1, 1.0, 3.0, or 30	Weights: body, thymus, spleen, liver, kidney	None
		Cellularity: spleen, thymus, bone marrow	None
		Splenic CD4CD8 cells	None
		NK cell activity/B16F10 tumor challenge	None/Not Done
		CTL to P815 cells (*in vitro*)	Increased at 0.1, 1.0 and 3.0; no effect in subsequent "corrected" study.
		L. monocytogenes challenge	Not Done
		DTH to *L. monocytogenes* antigen	Increased at 30. NOAEL = 3.0, LOAEL = 30
		Macrophage phagocytosis (*in vitro*)	Decreased at 1.0 and 30. NOAEL = 0.1, LOAEL = 1.0
		Macrophage nitrate (*in vitro* + IFN or LPS)	None
		IgM ELISA to SRBCs	None
	90-days 0, 0.1, 1.0, 3.0, or 30	Weights: body, thymus, spleen, liver, kidney	None
		Cellularity: spleen, thymus, bone marrow	None
		Splenic CD4CD8 cells	None
Mouse/B6C3F1/Female (Kiel et al., 1998; 1999)	90-days 0, 0.1, 1.0, 3.0, or 30	NK cell activity/B16F10 tumor challenge	Increase NK activity at 30 in one experiment and at 30 in 120-day study: NOAEL = 3, LOAEL = 30. No effect on B16F10 tumor challenge.
		CTL to P815 cells (*in vitro*)	None
		L. monocytogenes challenge	None
		DTH to *L. monocytogenes* antigen	Increase at 30. NOAEL = 3.0, LOAEL = 30
		Macrophage phagocytosis (*in vitro*)	Decreased at 0.1, 1.0, 3.0 and 30, LOAEL = 0.1

TABLE 5-5 (cont'd). SUMMARY OF IMMUNOTOXICITY TEST RESULTS

Series/Strain/Sex (Study)	Exposures Period and Doses (mg/kg/d)	Endpoint	NOAEL/LOAEL Designations
		Macrophage nitrate (*in vitro* + IFN or LPS)	None
		IgM ELISA to SRBCs	None
Mouse/CBA/JHsd/Female (BRT-Burleson Research Technologies, Inc., 2000a,b,c)	14-days 0, 0.02, 0.06, 0.2, 2.0, or 50	anti-SRBC PFC/10^6 cells	None
		anti-PFC/spleen	None
		LLNA to DNCB	Increased at 0.06, 0.2, and 50, but not at 2.0. NOAEL = .02, LOAEL = 0.06
	90-days 0, 0.02, 0.06, 0.2, 2.0, or 50	anti-SRBC PFC/10^6 cells	Increased at 50. NOAEL = 2.0, LOAEL = 50
		anti-PFC/spleen	Increased at 2.0 and 50. NOAEL = 0.2, LOAEL = 2.0
		LLNA to DNCB	Increased at 0.06 and 0.2, but not at 2.0; decreased at 50. NOAEL = 0.02, LOAEL = 0.06

NK = natural killer; CTL = cytotoxic lymphocyte; DTH = delayed type hypersensitivity; IFN = interferon; SRBC = sheep red blood cell; PFC = plaque forming colony; LLNA = local lymph node assay; DNCB = 2,4-Dinitrochlorobenzene.

1   The three immune function parameters altered by ammonium perchlorate exposure were

2   the following: suppression of *in vitro* peritoneal macrophage phagocytosis of *L. monocytogenes*,

3   enhancement of the PFC response to SRBCs, and enhancement of the LLNA to DNCB. These

4   results are summarized and discussed below.

5   Decreased *in vitro* phagocytosis of *L. monocytogenes* by peritoneal macrophages obtained

6   from mice dosed for 14 days at 1- or 3- and 30-mg ammonium perchlorate/kg-day was observed

7   (Keil et al, 1998, 1999). In mice exposed for 90-days, phagocytosis was decreased in all dosage

8   groups (Keil, 1998, 1999). However, in the 120-day (i.e., 90-day ammonium perchlorate

9   exposure followed by 30-day recovery) studies, no effect on macrophage phagocytosis of

10   *L. monocytogenes* was observed (Keil et al., 1998, 1999). Taken together, these data suggest that

1   ammonium perchlorate suppresses the *in vitro* phagocytic capacity of peritoneal macrophages,

2   but that this suppression is reversed after a 30 day recovery period.

3   This decrease in macrophage phagocytic activity could be expected to be reflected in the

4   results of the *L. monocytogenes* infectivity data because, along with other immune system

5   components, macrophages play a pivotal role in resistance to infection by this bacterium.

6   For example, the pathogenesis of *L. monocytogenes* is associated with its ability to grow within

7   mononuclear phagocytes. Complement (C') plays an important role in *L. monocytogenes*

8   infections, as demonstrated by the fact that C'-deficient mice have impaired host resistance to

9   this bacterium. This impairment in C'-deficient mice is caused by the absence of macrophage-

10   associated C'. The T-lymphocytes also play a major role in defense against *L. monocytogenes*

11   because complete elimination of bacteria from infected tissue is accomplished by macrophages

12   activated by T-cell dependent mechanisms.

13   However, the *L. monocytogenes* host-resistance studies indicate that ammonium

14   perchlorate exposure of mice does not alter the ability to combat this bacterial infection. With

15   the exception that clearance of *L. monocytogenes* from the liver of mice given a 5360 CFU

16   challenge following dosing at 3.0 mg AP/kg/day for 90 days was reduced, no other effect was

17   observed (Keil et a., 1999: Table 43).   These data imply that while in vitro phagocytosis by

18   peritoneal macrophages of this bacterium was reduced following ammonium perchlorate

19   exposure, the ability of macrophages from other *in situ* sites (e.g., spleen, liver) to clear

20   *L. monocytogenes* was not altered.

21   Exposure of mice to 2.0 or 50.0 mg ammonium perchlorate/kg/day for 90, but not 14, days

22   resulted in enhancement of the antibody response to SRBCs as determined by the PFC assay

23   (BRI-Burleson Research Technologies, Inc., 2000a,b,c). In both the 14- and 90-day studies, the

24   PFC response was suppressed by dosing mice with the immunosuppressive positive control CP.

25   The PFC assay is routinely used for identifying chemicals that are immunosuppressive. The

26   reason why the highest dose(s) of ammonium perchlorate, given over 90 days, enhanced this

27   response is not known. It is possible that under these dosing conditions ammonium perchlorate

28   may have an adjuvant-like or enhancing effect on the antibody response to SRBCs. The ELISA

29   data for mice exposed to up to 30.0 mg ammonium perchlorate/kg/day, for 14 or 90 days (Keil

30   et al., 1999), do not corroborate this enhanced response to SRBCs as determined by the PFC

1   assay. However, taken together, the PFC and ELISA data indicate that ammonium perchlorate

2   does not suppress the immune response to SRBCs.

3   The LLNA is an accepted approach for identifying chemicals with the potential of causing

4   dermal contact hypersensitivity (CHS) reactions in humans. In this assay the test substance,

5   2,4-dinitrochlorobenzene (DNCB) was topically applied on three consecutive days to both ears of

6   the mouse. Two days later the mice were injected iv with radioactive uridine (e.g., [125]IUDR).

7   Five hours later, the lymph nodes draining the ears, referred to as the "auricular" lymph nodes,

8   were removed and [125]IUDR incorporation by the lymph node cells determined. Since the nodes

9   draining the ear (i.e., "auricular" nodes) have no standard anatomical nomenclature, experience

10   in identifying these nodes as well as appropriate and consistent excision of these nodes from

11   control and test mice is critical. The LLNA evaluates the induction phase of the CHS reaction by

12   assessing the influx of lymphoid cells and the differential argumentation of lymphocyte

13   proliferation elicited by exposure to the test chemical relative to that of a vehicle control.

14   The data from BRT-Burleson Research Technologies, Inc. (2000a,b,c) report that exposure

15   to ammonium perchlorate enhances/exacerbates the LLNA response to DNCB at doses of 0.06,

16   and 0.2 mg/kg/d in both the 14- and 90-day. While a dose of 50.0 mg/kg-day for 14 days also

17   enhanced this response, a dose of 2.0 mg/kg-day did not. Similarly, a dose of 2.0 mg/kg-day in

18   the 90-day study did not enhance the LLNA response to DNCB. In contrast to the 14-day study,

19   exposure of mice to 50.0 mg ammonium perchlorate/kg/day in the 90-day study resulted in

20   suppression of the LLNA response. In the 90-day study, the positive control CP did not suppress

21   the LLNA response to DNCB. The failure of CP to suppress this response in the 90-day vs.

22   14-day study is disquieting because CP was administered similarly (i.e., 15 mg/kg-day for

23   5 consecutive days prior to the LLNA) in both studies. The only difference between these two

24   studies was that the mice in the 90-day study were 11 weeks older. This difference in age,

25   however, should not influence the ability of CP to suppress this response. The fact that CP did

26   not suppress the LLNA response in the 90-day study calls into question the performance of this

27   and perhaps the 14-day study.

28   Application of the LLNA for identification of chemicals that are contact sensitizers

29   routinely involves a demonstration of a dose-related increase in the LLNA using, at a minimum,

30   three increasing concentrations of the test agent. Neither the 14- nor 90-day ammonium

31   perchlorate LLNA data demonstrate a dose-response relationship, which would be expected if

1   ammonium perchlorate was acting additively or synergistically with DNCB to increase the

2   LLNA response. While higher concentrations of a contact sensitizing agent will increase the

3   LLNA response, there is no information in the literature that indicates such an increase results in

4   a more serious or potentially detrimental effect on the host. Consequently, the physiologic

5   significance of the observed increase in the LLNA response to DNCB in ammonium perchlorate-

6   exposed mice is unknown. This is unlike the situation with immunosuppressive agents where

7   suppression of specific immune function(s) can be linked to a biological detrimental effect (i.e.,

8   decreased host resistence to an infectious agent or tumor).

9   It is interesting to note that there are published reports in which non-sensitizing agents have

10   been employed to improve the sensitivity of the LLNA to detect sensitizers. For example,

11   Vitamin A acetate dietary supplementation enhances the detection of weak sensitizers, and at low

12   concentrations of moderate sensitizers, assessed by the LLNA. The mechanism(s) for this

13   increased detection of contact sensitizers is not known. However, topically applied retinol causes

14   epidermal hyperplasia which may lead to increased numbers of antigen-presenting cells in the

15   epidermis. Retinoids also up regulate the sensitization phase of DTH induction through direct or

16   indirect stimulation of T cells. Non-sensitized mice, fed a diet supplemented with retinol,

17   display somewhat higher LLNA responses compared to control mice on a normal diet. This

18   suggests that dietary retinol itself causes cellular infiltration and/or proliferation in the absence of

19   a contact sensitizer as measured by the LLNA. It may be that ammonium perchlorate, in the

20   absence of DNCB, has the capability of raising the baseline LLNA response compared to water

21   control mice. Unfortunately, there were no negative controls in the Burleson et al. (2000)

22   studies. Appropriate negative controls would have included the following: (1) ammonium

23   perchlorate-dosed non-sensitized mice; (2) ammonium perchlorate-dosed and ammonium

24   perchlorate-challenged mice; and (3) water control mice dermally exposed to ammonium

25   perchlorate on the ear pinna. Another group of appropriate and informative studies would

26   involve ammonium perchlorate-dosed mice that would be challenged with a series of low to

27   moderate concentrations of DNCB, for comparison with the current LLNA "optimal DNCB"

28   response concentration data.

29   Enhancement of the LLNA to DNCB in mice exposed to 0.06 mg ammonium

30   perchlorate/kg-day for 14 or 90 days represents the Lowest Observed Effect Level (LOEL) for all

31   of the immune function tests performed. While this is the LOEL it is unknown if this is the

1 Lowest-Observed-Adverse-Effect- Level (LOAEL) because it is not clear that enhancement of

2 the LLNA is a physiologically relevant adverse effect. Studies are needed to determine if

3 ammonium perchlorate itself is a contact sensitizer as determined by the LLNA, as described

4 above, and whether the degree of the LLNA response to ammonium perchlorate itself or to a

5 known contact sensitizer can be linked to a quantifiable adverse outcome.

6   It is important to note that clinical studies in the 1960s reported that some patients suffering

7 from Graves' disease and treated with potassium perchlorate presented with agranulocytosis

8 and/or skin rashes. While the studies reported by Keil et al. (1998, 1999) indicated that there

9 were no alterations in the proportion of peripheral blood leukocytes of mice dosed with

10 ammonium perchlorate for 14- or 90-days, the work of BRI-Burleson Research Technologies,

11 Inc. (2000a,b,c) suggests that ammonium perchlorate appears to exacerbate the contact

12 sensitizing potential of the known skin sensitizer DNCB. However, due to the uncertainties

13 associated with any attempt to extrapolate from the incomplete database of the mouse LLNA

14 performed by BRI-Burleson Research Technologies, Inc. (2000a,b,c) to the clinical observations

15 of skin rash and agranulocytosis in Graves' disease patients treated with potassium perchlorate,

16 an uncertainty factor based on deficiencies in the database is recommended to be applied to this

17 risk assessment.

18

CHAPTER 6. CONSTRUCTION OF PBPK MODELS TO ADDRESS PERCHLORATE'S MODE-OF-ACTION

1
2
3
4

5   The purpose of this chapter is to describe the progress that has been made in developing

6   physiologically-based pharmacokinetic (PBPK) models to aid interspecies extrapolation of

7   effects observed in the toxicity studies. The models describe perchlorate and iodide kinetics in

8   rats and humans. Because of the complex challenge posed in arriving at a representation of the

9   regulation system for hypothalamic-pituitary-thyroid feedback, the modeling effort was not able

10   to satisfactorily develop models that linked the observed effects of perchlorate inhibition of

11   iodine uptake at the NIS with the resultant hormone perturbations and available toxicological

12   information in the proposed mode-of-action framework.

13   Because of their potential role in the risk assessment and regulatory applications, the EPA

14   required that all human clinical data utilized in this modeling effort undergo a quality

15   assurance/quality check (QA/QC). The QA/QC report is presented in Merrill (2001a,b). These

16   QA/QC data represent the most contemporary, comprehensive, and consistent set of human

17   pharmacokinetic data available for perchlorate.

18   The PBPK models discussed herein (Merrill, 2001c,d; Clewell, 2001a,b) were developed

19   by the AFRL/HEST to provide more accurate descriptions of the kinetics of iodide and

20   perchlorate with respect to perchlorate's inhibition of iodide uptake at the NIS and their serum

21   and tissue time courses as well as to aid evaluation of subsequent perturbations in thyroid

22   hormones and TSH. A general discussion of the model development for the various PBPK

23   model structures of perchlorate distribution will be provided in this chapter to aid appreciation of

24   their attributes and applications. Because of the mode of action for perchlorate, an accurate

25   description of iodide kinetics is critical to the description of perchlorate effects on iodide uptake

26   at the NIS so that each of these models also includes iodide-specific parameters and accounts for

27   iodide disposition.

28   A similar model was developed for each of the various life stages of importance to

29   interspecies extrapolation of the laboratory animal data: adult, pregnant rat and fetus, and the

30   lactating rat and neonate. The adult male rat model was developed using data from the ADME

1   studies in the perchlorate testing strategy, together with experimental data and parameter values

2   available in the existing literature. The subsequent model structures for the human and various

3   life stages of the rat were similarly developed based largely on the adult male rat structure

4   through scaling and optimization of parameters to available data.

5   It should be noted that the original motivation for including human studies (as discussed in

6   Chapter 3) in the perchlorate testing strategy was to support such interspecies extrapolation and

7   not to derive NOAEL estimates for thyroid effects in the human population. As discussed in

8   Chapter 4, the EPA feels that both the observational epidemiological and the human clinical

9   studies have significant scientific and technical limitations that preclude their use as the basis for

10   a quantitative dose-response assessment. In addition, some of the clinical studies contained in

11   this database fall in the category of studies not to be considered under EPA's Dec. 14, 2001

12   interim policy on the use of third-party human studies (U.S. Environmental Protection Agency,

13   2001c). However, the scientific and technical strengths and weaknesses of these studies were

14   described before this Agency policy was articulated. Therefore, because of the scientific

15   shortcomings of these studies, they will not be used as "principal studies" in the derivation of

16   an RfD. The clinical study subject attributes (euthyroid adults) and study design issues (sample

17   size, RAIU time points, etc.) made these data less reliable than the laboratory animal toxicological

18   data to ascertain effect levels for the basis of an RfD derivation. Models of perchlorate distribution

19   for human pregnancy and lactation have not been developed.

20   More detailed discussion can be found for each model structure in the accompanying

21   references provided for each in the sections that follow. The adult male rat and human model

22   (Merrill, 2001c,d) will be discussed in Section 6.2. Section 6.3 discusses the pregnant dam and

23   fetal rat PBPK model (Clewell, 2001a), and the lactating dam and neonate model (Clewell,

24   2001b) is discussed in Section 6.4. The purpose of providing these model descriptions and a

25   discussion of the data used to develop and validate their structures is to provide the external peer

26   reviewers an opportunity to critically evaluate the model structures, the use of the data in model

27   development or validation exercises, and the model applications.

28   The simultaneous ordinary differential equations used in the proposed PBPK models to

29   simulate radioiodide and perchlorate distribution were written and solved using advanced

30   continuous simulation language (ACSL) software (AEqis Technologies, Austin, TX).

31

1   **6.1 MODE-OF-ACTION FRAMEWORK AND UNDERLYING**
2   **MODELING APPROACH**

3   The mode-of-action model proposal by the EPA for the previous perchlorate assessment

4   and discussed in Chapter 3 served as the conceptual construct for the development of the PBPK

5   models. Shown again in Figure 6-1, the model lays out the biomarkers of exposure and effect in

6   a continuum from ingestion of perchlorate in drinking water and uptake into the blood, the key

7   event of iodide uptake inhibition at the NIS in the thyroid gland, and subsequent effects on

8   thyroid hormone economy leading to neurodevelopmental and neoplastic sequelae.

9
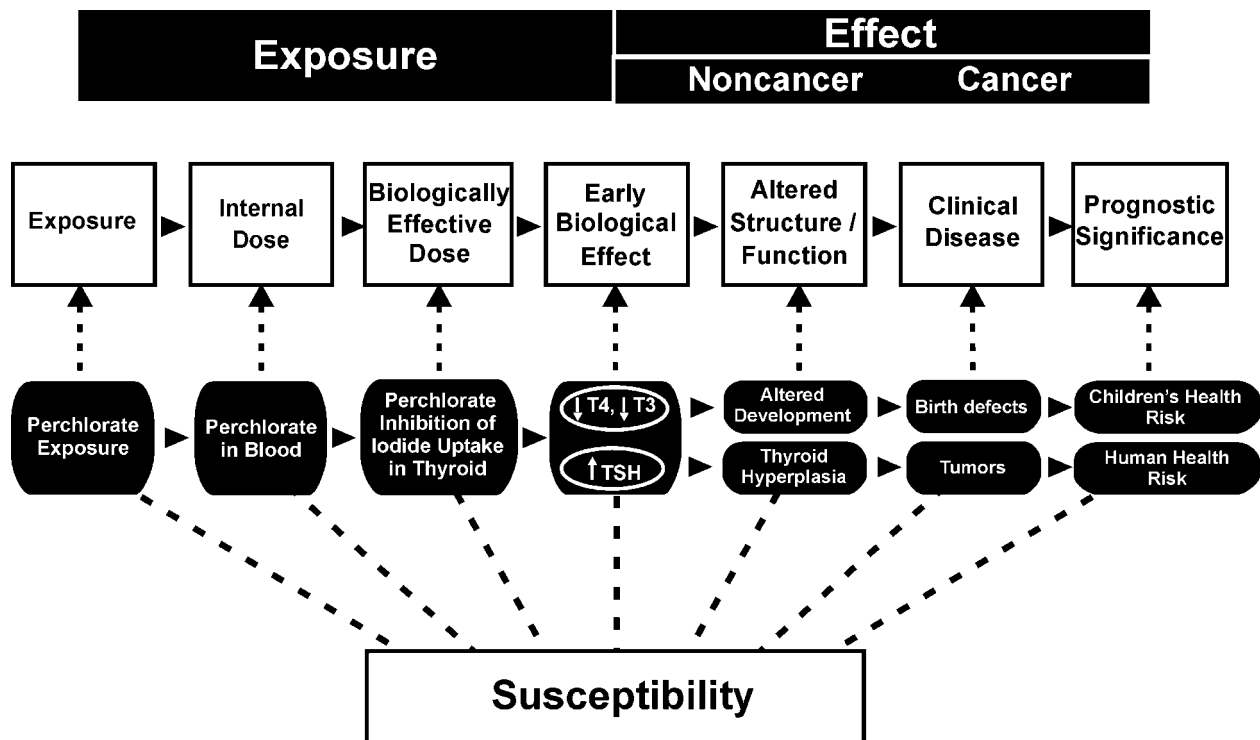
Figure 6-1. Mode-of-action model for perchlorate toxicity proposed by the U.S. EPA (U.S. Environmental Protection Agency, 1998d). Schematic shows the exposure-dose-response continuum considered in the context of biomarkers (classified as measures of exposure, effect, and susceptibility) and level of organization at which toxicity is observed (U.S. Environmental Protection Agency, 1994; Schulte, 1989). The model maps the toxicity of perchlorate on this basis by establishing casual linkage or prognostic correlations of precursor lesions.

1   The temporal pathological and serum hormone changes that accompany exposure to

2   perchlorate corresponding to this continuum are represented in Figure 6-2. The inhibition of

3   iodide uptake at the NIS results in a transient decrease in serum T4 and T3. This transient phase

4   of thyroid hormone deficit is of concern during pregnancy and development due to the critical

5   role that these hormones play in preventing adverse neurodevelopmental sequelae as described in

6   Chapters 3 and 5. The hypothalamic-pituitary-thyroid feedback system is designed to regulate

7   the circulating levels of thyroid hormone and will respond to the thyroid hormone decreases by

8   upregulating TSH production in order to stimulate the thyroid to increase its production of

9   thyroid hormones to compensate. Represented as the "chronic phase" in Figure 6-2, the

10   upregulation of TSH would bring the system back into apparent homeostasis. As depicted in the

11   figure, however, this apparent homeostasis may actually represent subclinical disease in that the

12   system is only maintaining homeostasis by upregulation and can be considered a stressed system

13   with respect to its ability to compensate for additional insults caused by other chemicals or

14   diseases that might impact the thyroid. Further, it should be emphasized that recent

15   epidemiological investigations have indicated concern about decrements in T4, i.e., thyroxinemia

16   without concomitant upregulation of TSH that would constitute hypothyroidism (Morreal de

17   Escobar, 2000; Haddow et al., 1999; Pop et al., 1999).

18   In order to adequately characterize the transient phase of events, evaluation of the initial

19   effect of perchlorate at the NIS is necessary. This can be accomplished by determining

20   perchlorate inhibition with radioactive iodide uptake (RAIU) studies. The timing and route of

21   administration are important considerations in evaluating these types of studies. Studies of

22   RAIU that occur during the chronic phase, such as longer-term studies of hormones, offer little

23   insight to the critical decrements in T4 that may occur during the transient phase due to iodine

24   inhibition. Likewise, longer-term studies of hormones often represent the upregulated system

25   and may not be especially informative.

26

27   **6.1.1 Parallelogram Approach to Interspecies Extrapolation**

28   PBPK models have proven to be very useful tools for performing interspecies extrapolation

29   of dose for applications in risk analysis. Interspecies extrapolation is often necessary because, as

30   in this case of perchlorate, critical effects at levels of organization below that of the population

31   (e.g., thyroid histopathology or brain morphometry) can not be evaluated easily or ethically in
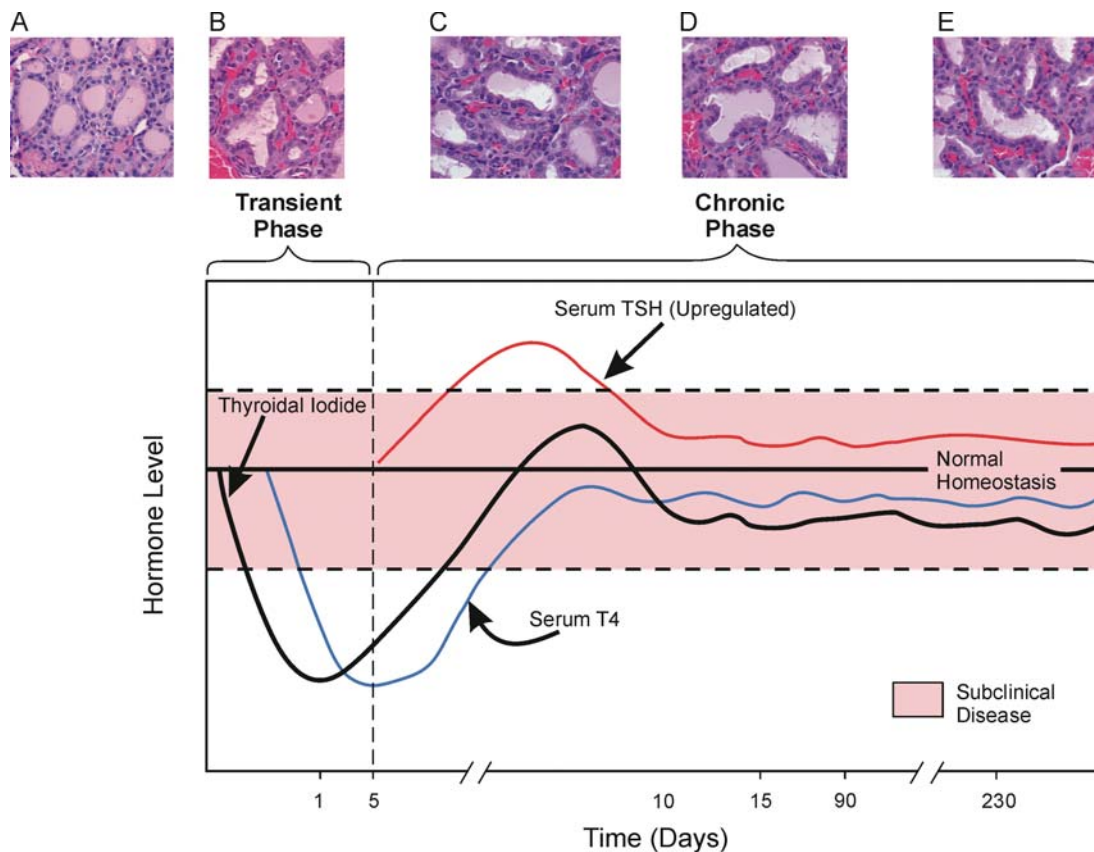
Figure 6-2. Schematic of thyroid and pituitary hormone levels with associated pathology after acute versus chronic dosing with perchlorate. The transient phase is represented by decreases in thyroidal iodide due to the inhibition by perchlorate at the NIS with subsequent drop in T4. The transient drops in T4 can lead to permanent neurodevelopmental sequelae. Once TSH is upregulated via the hypothalamic-pituitary-thyroid feedback, T4 appears to be in normal homeostasis but actually can represent subclinical or undiagnosed disease (hypothyroxinemia). The upregulation of TSH can result in neoplasia. Normal thyroid tissue is represented in Panel A. Panel B shows lace-like colloid depletion which is more pronounced in subsequent panels C, D and E. Panels D and E represent hypertrophy and hyperplasia.

1 humans. A basic tenet of molecular epidemiology is that these precursor lesions are often more

2 closely related to the exposure than are the traditional outcome measures of morbidity and

3 mortality (U.S. Environmental Protection Agency, 1994).

4   A parallelogram approach as shown in Figure 6-3 is used to predict the dose-response

5 relationship for humans based on the dose-response in laboratory animals. Because these critical

Human
Effective Dose

Human
Health Effect

Rat
Effective Dose

Rat
Adverse Effect
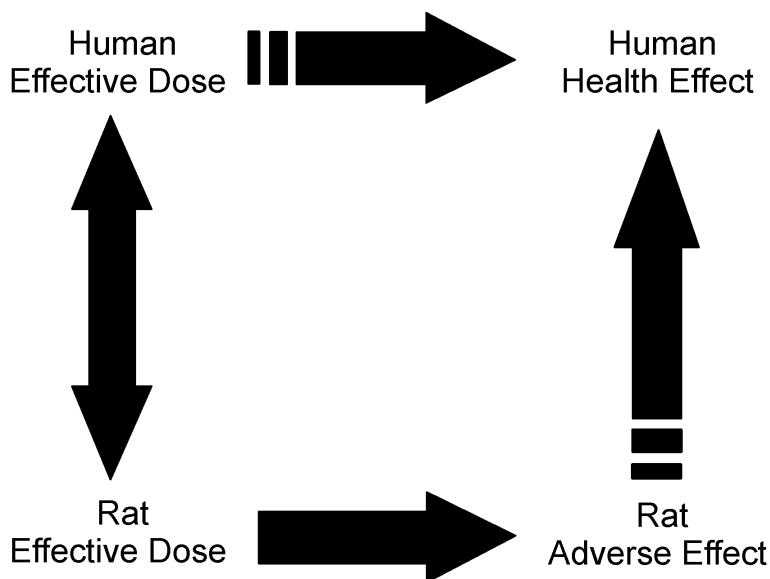
Figure 6-3. Schematic of parallelogram approach used for interspecies extrapolation (U.S. Environmental Protection Agency, 1994). Dose and adverse effect in rat can be used to predict human effective dose and response.

1   effects cannot be accurately measured in humans, the dose associated with an observed critical

2   effect in the laboratory animal is scaled to the human by adjusting the PBPK model with human

3   physiological parameters and variables. The human model is typically constructed by

4   allometrically scaling some parameters in the laboratory animal model based on body weight, and

5   some parameters such as partition coefficients can be measured *in vitro*. An administered dose

6   associated with the critical effect is determined based on an appropriate internal dose metric.

7   The internal dose is scaled to an equivalent exposure (HEE) in humans by exercising the human

8   model with human parameters and exposure assumptions. Thus, the HEE represents the human

9   exposure that would result in the same amount of internal dose metric in a human as that which

10   caused the effect in the laboratory animal.

11   The dose-response relationship is considered to be the same as that in the laboratory animal

12   as the default or more biologically-based models may contain additional parameters that also

13   account for species-specific determinants of toxicant-target interaction. Figure 6-4 illustrates the

14   use of the laboratory animal and human PBPK models to arrive at the HEE. Simulations used to

**Rat
PBPK Model**

Laboratory
Animal
Exposure
(mg/kg-day)

Rat
Effective
Dose Metric
(mg/kg-day)

Human
Equivalent
Exposure
(mg/kg-day)

Human
Effective
Dose Metric
(mg/kg-day)
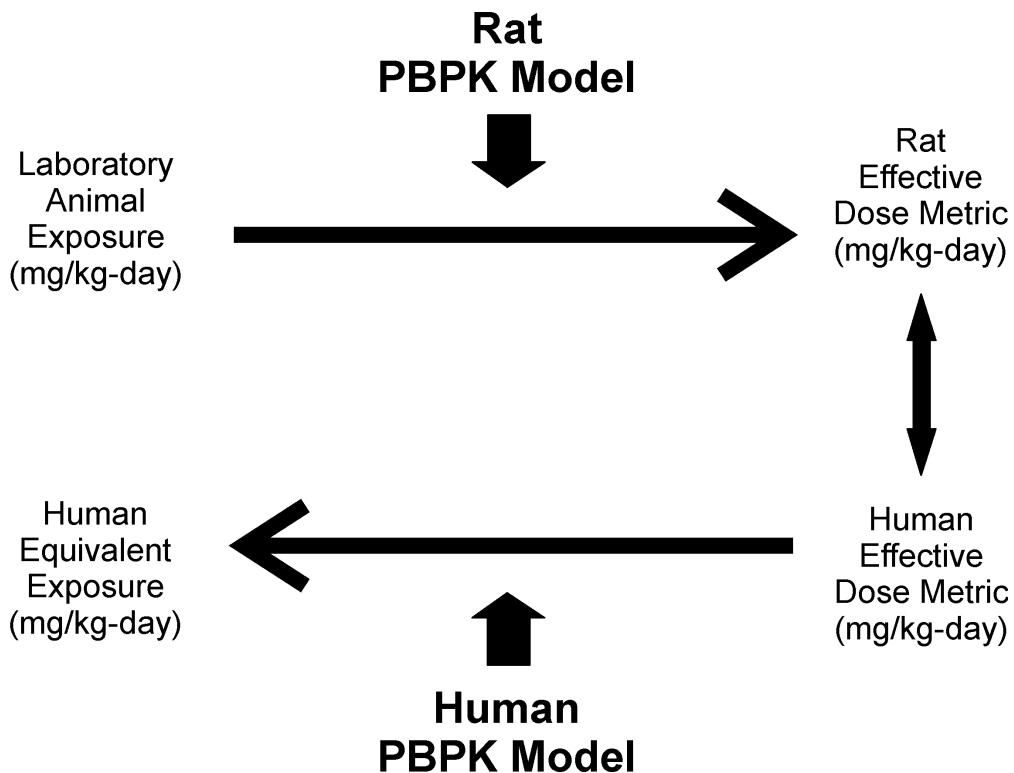
**Human
PBPK Model**

Figure 6-4. Illustration of how human equivalent exposure (HEE) is calculated using PBPK models. An effective internal dose associated with a critical health effect at an administered dose (mg/kg-day) is calculated by simulating the experimental exposure regimen (e.g., 5 days/week) for a relevant metric (e.g., area under the curve in blood, [AUCB]). The human PBPK model is then used to simulate an exposure that achieves the same effective internal dose metric level using human parameters.

1 arrive at HEE for different internal dose metrics and a sensitivity analysis of the adult model

2 structure will be discussed in Section 6.5.

3   The parallelogram approach has also been used to predict effective doses for structurally

4 related chemicals (Jarabek et al., 1994). Disposition of one chemical associated with an effect

5 can be predicted for another after appropriate adjustments for chemical structure and activity are

6 made. In the case of these models, it should be appreciated that the accurate modeling of iodide

7 in addition to that of perchlorate represents such a validation.

8

9

1   **6.1.2 Extending the Parallelogram Approach to Various Experimental**
2   **Life Stages**

3   Because effects at various life stages (adult, pregnant dam, fetus, lactating dam, and

4   neonate) were evaluated in the perchlorate laboratory animal studies, the parallelogram approach

5   had to be extended as shown in Figure 6-5. There are no human models of perchlorate

6   disposition for pregnant women, lactating women, fetuses, or children, so the relationships to the

7   adult human HEE had to rely on the relationships determined in the laboratory animal species.

8   This approach assumes that the relationships, expressed as ratios between one life stage and

9   another, will be comparable in humans.

10

11
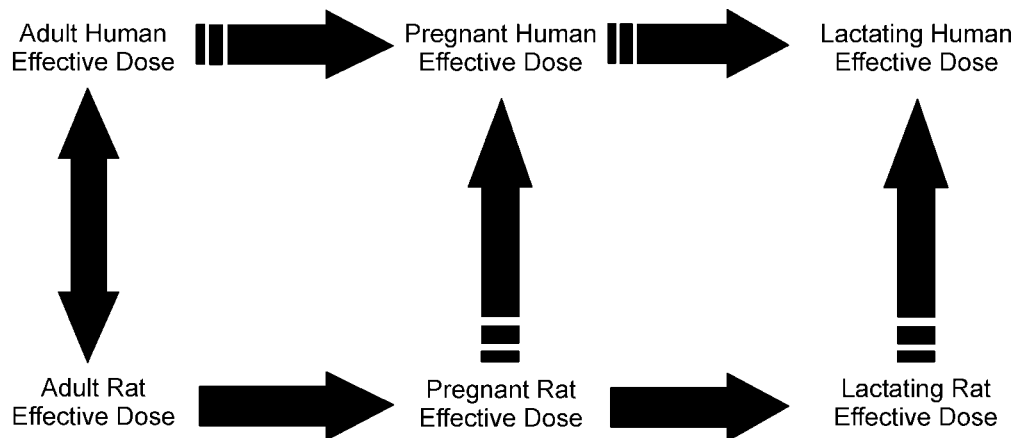
**Figure 6-5. Schematic of extended parallelogram approach used for perchlorate due to
            effects at different life stages. Doses in the pregnant rat and fetus are related
            back to the adult male rat, likewise, the effects in lactating rats and neonates.**

1   The various PBPK models are used to predict equivalent effective doses at the various

2   administered doses used in the experiments; e.g., 1.0 mg/kg-day ammonium perchlorate given in

3   drinking water to both the adult male rat and the pregnant dam. Each PBPK model is exercised

4   (adult rat and pregnant rat) to predict the amount of internal dose metric achieved at each life

5   stage. The ratio of the effective internal dose metrics of the life stage in question is then used to

1   adjust the HEE based on the adult male rat. For example, the HEE for the pregnant dam would

2   be found by adjusting the HEE for the adult male rat by the ratio of the male rat:pregnant rat as:

3

$$\text{Pregnant HEE (mg/kg-day)} =$$
$$\text{Adult male rat HEE (mg/kg-day)} \times \frac{\text{Adult male rat internal effective dose metric}}{\text{Pregnant rat internal effective dose metric}} \quad (6\text{-}1)$$

4

5   This ratio is unitless and accounts for the differences between the two life stages in

6   question in an analogous fashion to the dosimetric adjustment factor (DAF) used in the EPA's

7   inhalation reference concentration methods to extrapolate respiratory tract doses in different

8   regions of the laboratory animal to human equivalent concentrations (U.S. Environmental

9   Protection Agency, 1994). The same ratio approach is used to extend the model predictions to

10   HEE estimates for the fetus, lactating dam, and neonate. Development of the ratios for two

11   internal effective dose metrics, perchlorate area-under-curve (AUC) concentrations in serum and

12   iodide uptake inhibition, will be discussed in Section 6.5.

13

14

6.2 ADULT RAT AND HUMAN MODEL STRUCTURES

16   Because the same model structure is used to describe perchlorate and iodide disposition

17   (absorption, distribution, and elimination) for both the adult male rat and human, this section will

18   describe the development of both of these models together. Data supporting development and

19   validation of the structures will be summarized in this section while additional detail, including

20   some of the governing equations, can be found in the consultative letters from the AFRL/HEST

21   (Merrill, 2001c,d).

22   As discussed in Chapter 2, the perchlorate anion (ClO_4^-) is very similar in ionic size, shape,

23   and charge to that of iodide (I^-). These shared properties allow perchlorate to interfere with the

24   first stage of thyroid hormone synthesis by competitively inhibiting the active transfer of iodide

25   into the thyroid by the sodium (Na^+)-iodide (I^-) symporter or NIS. The NIS is a protein that

26   resides in the basolateral membrane of thyroid epithelial cells (Spitweg et al., 2000). NIS

27   simultaneously transports both sodium and iodide ions from extracellular plasma into the thyroid

28   epithelial cell via an active process. Energy is provided by the electrochemical gradient across

1   the cell membrane. The low intracellular concentration of sodium is maintained by sodium-

2   potassium pumps (Ajjan et al., 1998). The kinetics of perchlorate and iodide anions differ

3   mainly in that iodide is organified in the thyroid (thyroid hormone production); whereas,

4   perchlorate is thought to be unreactive and eventually diffuses from the thyroid into systemic

5   circulation.

6   The proposed PBPK model structure for the adult male rat (Merrill, 2001c) and human

7   (Merrill, 2001d) describes active uptake of iodide and perchlorate in gastric juice, thyroid, and

8   skin, and competitive inhibition of iodide uptake by perchlorate in NIS-containing tissues, as

9   well as venous equilibration with slowly and richly perfused tissues as shown in Figure 6-6.

10   Tissues that exhibited evidence of sodium iodide symporter and were found to concentrate either

11   anion were depicted as compartments with nonlinear uptake (Merrill, 2001c,d). Tissues with

12   active uptake include the thyroid, skin, and gastric mucosa (Wolff, 1998; Chow et al., 1969;

13   Kotani et al., 1998). Although other tissues have been known to sequester iodide and similar

14   anions (e.g., salivary glands, choroid plexus, ovaries, mammary glands, placenta) (Brown-Grant,

15   1961, Honour et al., 1952; Spitzweg et al., 1998), the iodide and perchlorate pools of these

16   tissues was expected to be too small to significantly affect plasma levels. These tissues were

17   lumped with slowly and richly perfused tissues.

18   The model also includes separate compartments for plasma, kidney, liver, and fat. These

19   compartments do not maintain concentrations greater than the plasma at steady state, and

20   therefore, were not described with terms for active uptake. The rapid urinary clearance of

21   perchlorate (Yu, 2000) mandated the inclusion of a kidney compartment in the model. A liver

22   compartment was also utilized due to its significant impact on iodide homeostasis. The majority

23   of extrathyroidal deiodination takes place within the liver. Fat was primarily added as an

24   exclusionary compartment. Due to its significant percentage of body weight, the skin represents

25   an important pool for slow iodide turnover.

26   The modelers at AFRL/HEST found that a separate skin compartment was necessary.

27   Experiments performed with radioiodide in rats resulted in skin:serum iodide ratios of close to

28   one (Yu, 2000). Other researchers have reported higher ratios in rats, but results have not been

29   consistent. Similar observations during dialysis with pertechnate of slow uptake and retention in

30   human skin was observed by Hays and Green (1973) and the skin was therefore maintained as a

31   separate compartment in the model. The skin contains two sub-compartments representing the
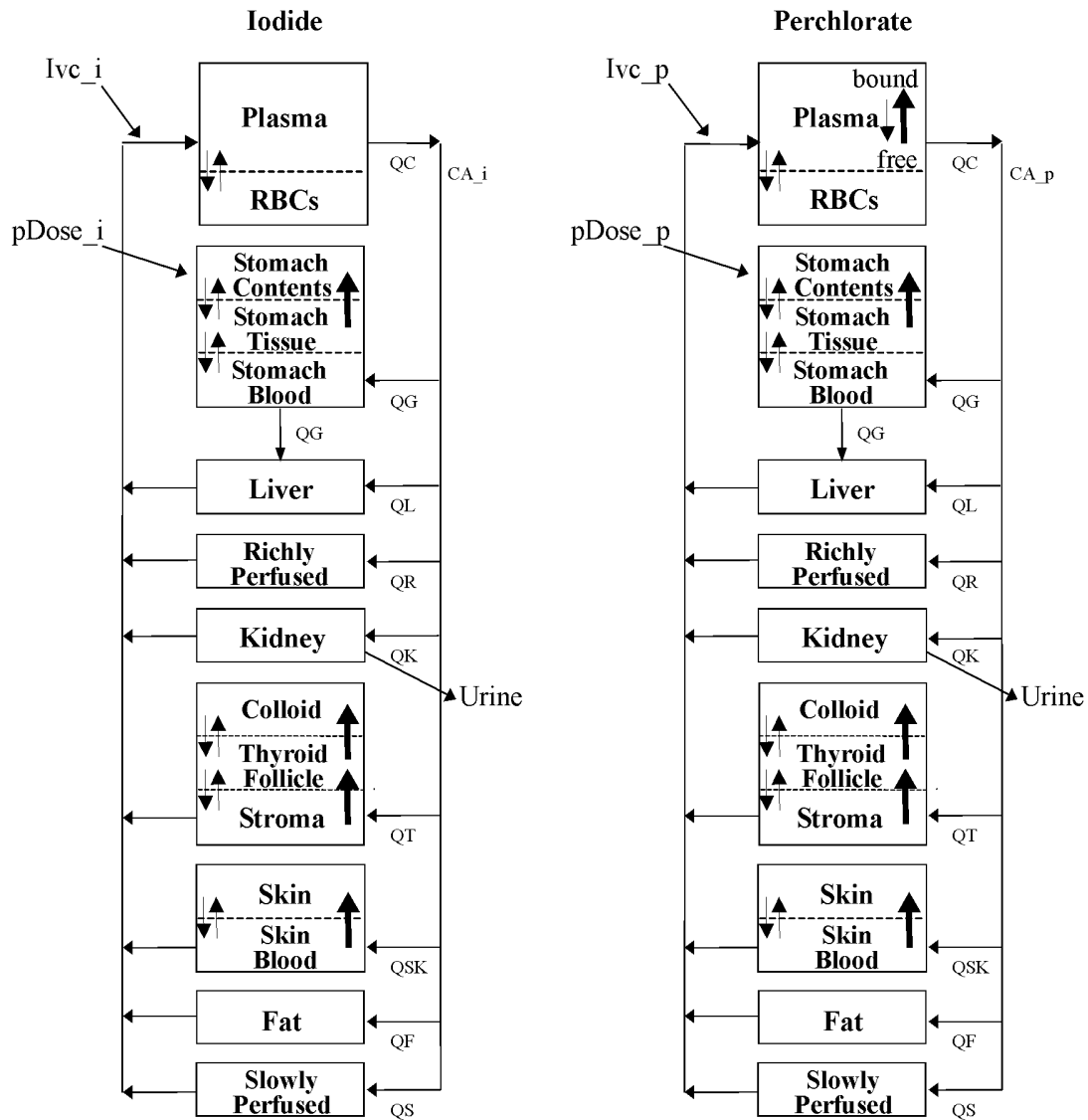
Figure 6-6. Schematic for the adult male rat and human PBPK models of perchlorate and iodide distribution (Merrill, 2001c,d). Bold arrows indicate active uptake (except for plasma binding) at NIS sites in thyroid, gut, and skin. Plasma binding was also described with Michaelis-Menten terms for the association of perchlorate anion to binding sites with first-order clearance rates for dissociation. Small arrows indicate passive diffusion. Boxes represent specific compartments in the model structure. The thyroid consists of the stroma, the follicle, and the colloid; and the stomach consists of the capillary bed, stomach wall, and stomach contents. The skin contains two subcompartments: the capillary bed and skin tissue. Permeability area cross products and partition coefficients were used to describe the first-order movement of the perchlorate (ClO_4^-) and iodide (I^-) anions into deeper subcompartments.

1   capillary bed and the skin tissue. The thyroid and stomach consist of three sub-compartments:

2   the stroma, the follicle, and the colloid in the thyroid and the capillary bed, stomach wall, and

3   contents in the case of the stomach.

4   Active uptake into the thyroid colloid, stomach contents, and skin were described using

5   Michaelis-Menten kinetics for nonlinear processes (Figure 6-6, bold arrows). Permeability area

6   cross products and partition coefficients were used to describe the first order movement of the

7   anions (ClO_4^- and I^-) between the capillary bed, tissue, and inner (deep) compartments

8   (Figure 6-6, small arrows) that results from the inherent electrochemical gradient within the

9   tissues. Passive diffusion through the kidney, liver, and fat compartments were described with

10   partitions and blood flows. Plasma binding of perchlorate was described with Michaelis-Menten

11   terms for the association of the perchlorate anions to plasma binding sites and a first order

12   clearance rate for the dissociation. First-order clearance rates from the kidney were also used to

13   describe urinary clearance of the anions.

14   The blood compartment differs between the perchlorate and iodide models. The

15   perchlorate blood compartment is composed of plasma and plasma proteins to simulate binding.

16   Plasma binding was required to simulate serum perchlorate concentrations at lower doses.

17   Iodinated hormones bind to plasma proteins, but free iodide apparently does not. Therefore, a

18   single compartment for plasma iodide was used. The free anions in plasma are available for

19   diffusion and active uptake into tissues.

20   The presence of NIS is an indicator of active uptake for iodide. NIS is highly expressed in

21   thyroid epithelial cells. Lower levels of expression have been detected in the mammary gland,

22   salivary gland, skin, stomach, and colon (Ajjan et al., 1998; Spitzweg et al., 1998). However,

23   only the thyroid has been found to organify iodide (Ajjan et al., 1998). The most important

24   regulator of symporter gene and protein expression is thyroid-stimulating hormone (TSH). This

25   is also the case for other important thyroid proteins such as thyroglobulin and thyroid peroxidase

26   (Spitzweg et al., 1998).

27   The parameters used in the adult male rat and human model for the various compartments

28   are provided in Table 6-1. The parameters were based on literature values or fitted to data using

29   the model as described in the table. It is important to note that the model structure for both

30   species is the same. The difference, per typical for PBPK models, is that there are species- and

31   chemical-specific parameters for each. For example, the volume of the thyroid (as percent of

TABLE 6-1. PHYSIOLOGICAL PARAMETERS FOR THE ADULT MALE RAT AND HUMAN PBPK MODELS (Merrill, 2001c,d)

Physiological Parameters

Tissue Volumes	Male Rat	Source	Human	Source
Body Weight BW (kg)	0.3	Measured (rat specific)	~70.0	Subject-specific
Slowly Perfused VSc (%BW)	74.6	Brown et al., 1997	65.1	Brown et al., 1997
Richly Perfused VRc (%BW)	11.0	Brown et al., 1997	12.4	Brown et al., 1997
Fat VFc (%BW)	7.4	Brown et al., 1997	♂ 21.0 ♀ 2.7	Brown et al., 1997
Kidney VKc (%BW)	1.7	Brown et al., 1997	0.44	Brown et al., 1997
Liver VLc (%BW)	5.5	Brown et al., 1997	2.6	Brown et al., 1997
Stomach Tissue VGc (%BW)	0.54	In house male rat ClO_4^- kinetics (Yu et al., 2000)	1.7	Brown et al., 1997
Gastric Juice VGJc (%BW)	1.68	In house male rat ClO_4^- kinetics (Yu et al., 2000)	0.071	Licht and Deen, 1988
Stomach Blood VGBc (%VG)	4.1	Altman & Dittmer, 1971b	4.1	Altman & Dittmer, 1971a
Skin Tissue VSkc (%BW)	19.0	Brown et al., 1997	3.7	Brown et al.,1997
Skin Blood VSkBc (%VSk)	2.0	Brown et al., 1997	8.0	Brown et al.,1997
Thyroid Vttotc (%BW)	0.0077	Malendowicz, 1977	0.03	Yokoyama et al., 1986
Thyroid Follicle VTc (%Vttot)	59.9	Malendowicz, 1977	57.3	Brown et al.,1986
Thyroid Colloid VDTc (%VTtot)	24.4	Malendowicz, 1977	15.0	Brown et al.,1986
Thyroid Blood VTBc (%VTtot)	15.7	Malendowicz, 1977	27.6	Brown et al.,1986
Plasma Vplasc (%BW)	4.1	Brown et al., 1997, Altman & Dittmer, 1971a	4.4	Marieb, 1992; Altman & Dittmer, 1971b
Red Blood Cells VRBCc (%BW)	3.3	Brown et al., 1997, Altman & Dittmer, 1971a	3.5	Marieb, 1992; Altman & Dittmer, 1971b
Adjusted Slowly Perfused VS (L)	0.138	Calculated from model	28.0	Calculated from model
Adjusted Richly Perfused VR (L)	0.01	Calculated from model	5.34	Calculated from model

TABLE 6-1 (cont'd). PHYSIOLOGICAL PARAMETERS FOR THE ADULT MALE RAT AND HUMAN PBPK MODELS (Merrill, 2001c,d)

Physiological Parameters Tissue Volumes	Male Rat	Source	Human	Source
Blood Flows				
Cardiac Output QCc (L/hr-kg)	14.0	Brown et al, 1997, Hanwell & Linzell, 1973	16.5	Brown et al., 1997; Hanwell & Linzell, 1973
Slowly Perfused QSc (%QC)	24.0	Brown et al., 1997	5.2	Brown et al., 1997
Richly Perfused QRc (%QC)	76.0	Brown et al., 1997	17.5	Brown et al., 1997
Fat QFc (%QC)	6.9	Brown et al., 1997	22.0	Brown et al., 1997
Kidney QKc (%QC)	14.0	Brown et al., 1997	1.0	Leggett & Williams, 1995; Malik et al., 1976
Liver QLc (%QC)	17.0	Brown et al., 1997	1.6	Brown et al., 1997
Stomach QGc (%QC)	1.61	Malik et al., 1976	13.0	Calculated, using 24% QC as flow to all slowly perfused tissues (Brown et al., 1997)
Skin QSkc (%QC)	5.8	Brown et al., 1997	33.0	Calculated, using 76% QC as flow to all richly perfused tissues (Brown et al., 1997)
Thyroid QTc (%QC)	1.6	Brown et al., 1997		
Adjusted Slowly Perfused QS (%QC)	11.3	Calculated from model		
Adjusted Richly Perfused QR (%QC)	41.8	Calculated from model		

body weight), the maximum capacity of thyroid iodide or perchlorate uptake, and plasma binding of perchlorate. The chemical-specific parameter for each model for both perchlorate and iodide are provided in Table 6-2.

In order to simulate the daily dosing regimen of the drinking water experiment, the rats were assumed to drink at constant rate for 12 of the 24 hours per day (1800 to 0600 hours). A pulse function in ACSL was used to introduce drinking water to the gastrointestinal (GI) compartment of the rat for the first 12 hours of each 24-hour period and to stop dosing while the rat was presumably sleeping. Intravenous (iv) dosing was introduced into the venous blood compartment of the model. Intraperitoneal (ip) injection was introduced into the model in the same manner as the iv dosing.

6.2.1 Data and Methods

This section summarizes the AFRL/HEST data and data available in the literature that were used for model development. Details on experimental methods, including protocol design, exposure regimen, chemical source and purity, animals (housing, feeding, surgical procedures, etc), and the analytical methods for measurement of RAIU; of perchlorate in plasma, urine and tissues; and of thyroid hormones and TSH can be found in the associated consultative letters from AFRL/HEST (Merrill, 2001c,d; Yu, 2000, 2001, 2002; Yu et al., 2000).

6.2.1.1 Studies in Laboratory Rats

The studies performed at AFRL/HEST included both "acute" iv experiments to measure radiolabled iodide or perchlorate as well as measurements of the same after drinking water administration. These two different regimens provided a better characterization of the transient ("acute") and chronic behavior necessary for an accurate description of the disposition of the anions. Adult male Sprague-Dawley rats (330 ± 35 g; n = 6 rats per group) that were purchased from Charles River Laboratory (Raleigh, NC) were used in the experiments.

In these experiments, the term total iodine includes bound iodine plus fee inorganic iodide. Carrier doses included tracer doses of carrier free radiolabled iodide ($^{125}I^-$) along with non-radiolabeled iodide. Free $^{125}I^-$ radioactivity was determined by subtracting the bound from total measurements (Merrill, 2001c; Yu, 2000, 2001, 2002; Yu et al., 2000).

TABLE 6-2. CHEMICAL-SPECIFIC PARAMETERS FOR THE ADULT MALE RAT AND HUMAN PBPK MODELS (Merrill, 2001c,d)[a]

Partition Coefficients (unitless)	Male Rat			Human		
	Perchlorate	Iodide	Source	Perchlorate	Iodide	Source
Slowly Perfused/Plasma PS_	0.31	0.21	Yu et al., 2000; Halmi et al., 1956	0.31	0.21	Halmi et al., 1956;Yu et al., 2000
Richly Perfused/Plasma PR_	0.56	0.40	Yu *et al.*, 2000; Halmi *et al.*, 1956	0.56	0.40	Halmi *et al.*, 1956;Yu *et al.*, 2000
Fat/Plasma PF_	0.05	0.05	Pena et al., 1976	0.05	0.05	Pena et al., 1976
Kidney/Plasma PK_	0.99	1.09	Perlman et al., 1941	0.99	1.09	Perlman et al., 1941; Yu et al., 2000
Liver/Plasma PL_	0.56	0.44	Perlman et al., 1941	0.56	0.44	Perlman et al., 1941; Yu et al., 2000
Gastric Tissue/Gastric Blood PG_	1.80	1.40	Yu et al., 2000; Yu, 2000	1.80	0.50	Yu et al., 2000;Yu, 2000
Gastric Juice/Gastric Tissue PGJ_	2.30	3.00	Yu et al., 2000; Yu, 2000	2.30	3.50	Yu et al., 2000; Yu, 2000
Skin Tissue/Skin Blood PSk_	1.15	0.70	Yu, 2000, Perlman et al., 1941	1.15	0.70	Perlman et al., 1941; Yu, 2000
Thyroid Tissue/Thyroid Blood PT_	0.13	0.15	Chow & Woodbury (1970)	0.13	0.15	Chow & Woodbury (1970)
Thyroid Lumen/Thyroid Tissue PDT_	7.00	7.00	Chow & Woodbury (1970)	7.00	7.00	Chow & Woodbury (1970)
Red Blood Cells/Plasma	0.80	1.00	Yu et al., 2000; Rall et al., 1950	0.80	1.00	Rall et al., 1950; Yu et al., 2000
Max Capacity, Vmaxc (ng/hr-kg)						
Thyroid Colloid Vmaxc_DT	1.0E+04	4.0E+07	Fitted	2.5E+5	1.0E+8	Fitted
Thyroid Follicle Vmaxc_T	2.2E+03	5.5E+04	Fitted	5.0E+4	~1.5E+5	Fitted
Skin Vmaxc_S	6.2E+05	5.0E+05	Fitted	1.0E+6	7.0E+5	Fitted
Gut Vmaxc_G	3.0E+05	1.0E+06	Fitted	1.0E+5	9.0E+5	Fitted
Plasma Binding Vmaxc_Bp	9.5E+03	—	Fitted	5.0E+2	—	Fitted

TABLE 6-2 (cont'd). CHEMICAL-SPECIFIC PARAMETERS FOR THE ADULT MALE RAT AND HUMAN PBPK MODELS (Merrill, 2001c,d)[a]

Partition Coefficients (unitless)	Male Rat			Human		
	Perchlorate	Iodide	Source	Perchlorate	Iodide	Source
Affinity Constants, Km (ng/L)						
Thyroid Lumen Km_DT	1.0E+08	1.0E+09	Golstein et al., 1992	1.0E+8	1.0E+9	Golstein et al., 1992
Thyroid Km_T	2.5E+05	4.0E+06	Gluzman & Niepomniszcze, 1983; Wolff, 1998	1.8E+5	4.0E+6	Gluzman & Niepomniszcze, 1983; Wolff, 1998
Skin Km_S	2.0E+05	4.0E+06	Gluzman & Niepomniszcze, 1983; Wolff, 1998	2.0E+5	4.0E+6	Gluzman & Niepomniszcze, 1983; Wolff 1998
Gut Km_G	2.0E+05	4.0E+06	Gluzman & Niepomniszcze, 1983; Wolff, 1998	2.0E+5	4.0E+6	Gluzman & Niepomniszcze, 1983; Wolff, 1998
Plasma binding Km_B	1.1E+04	—	Fitted	1.8E+4	—	Fitted
Permeability Area Cross Products (L/hr-kg)						
Gastric Blood to Gastric Tissue PAGc_	0.80	0.10	Fitted	0.6	0.2	Fitted
Gastric Tissue to Gastric Juice PAGJc_	0.80	0.10	Fitted	0.8	2.0	Fitted
Skin Blood to Skin Tissue PASkc_	1.0	0.10	Fitted	1.0	0.06	Fitted
Plasma to Red Blood Cells PARBCc_	0.10	1.00	Fitted	1.0	1.0	Fitted
Follicle to thyroid blood PATc_	4.0E-05	1.0E-04	Fitted	1.0E-4	1.0E-4	Fitted
Lumen to Thyroid Follicle PADTc_	0.01	1.0E-04	Fitted	0.01	1.0E-4	Fitted
Clearance Values (L/hr-kg)						
Urinary excretion CLUc_	0.07	0.05	Fitted	0.126	0.1	Fitted
Plasma unbinding Clunbc_	0.1	—	Fitted	0.025	—	Fitted

[a]All parameters listed are notated in the model by either an i (for iodide) or p (for perchlorate) following an underscore in the parameter name (e.g., PR_i, PR_p, Vmaxc_Ti, Vmaxc_Tp, etc.).

6.2.1.1.1 Acute iv Experiments in Rats

Radiolabeled iodide ($^{125}I^-$) kinetics. Male rats were administered a single iv tail-vein injection with physiological saline (control group) or 33 mg/kg $^{125}I^-$ (with carrier) in physiological saline. Rats were euthanized by CO_2 asphyxiation at 5, 15, and 30 minutes (min), 1, 2, 6, 9, 24, 32, 48, and 96 hours (hr) post dosing to collect thyroid and blood from the vena cava. Rats for the 24 hour time point were placed individually in metabolism cages to collect urine.

In an additional study, male rats were intravenously dosed with 33 mg/kg $^{125}I^-$ (with carrier) and euthanized at 0.5, 2 and 6 hours post dosing. Total, bound, and free $^{125}I^-$ were analyzed in thyroid and serum, and total $^{125}I^-$ was measured in skin and gastric contents (Yu, 2001).

Radiolabeled $^{36}ClO_4^-$ kinetics. Naïve adult male rats (300 ± 20 g) were dosed once by iv tail-vein injection with 3.3 mg/kg radiolabeled perchlorate. Due to the low specific activity, a smaller dosing level could not be achieved. Each rat received less than 6 μCi. Rats were euthanized by CO_2 asphyxiation at 0.5, 6, 12, 24, 32, and 48 hours after dosing. The thyroid, intestinal tract, intestinal tract contents, muscle, skin, liver, kidney, spleen, bladder, plasma, and red blood cells were harvested from the rats and stored at -20°C until analysis of $^{36}ClO_4^-$. Rats for 12, 24, 32, and 48 hours time points were placed individually in metabolism cages for urine collection. Metabolism cages were washed with 500 mL de-ionized water. Urine and cage wash samples were stored under the same conditions until analysis.

$^{125}I^-$ Kinetics and Inhibition from Acute iv Dosing with ClO_4^-. Rats were injected with one of five doses of perchlorate (0.0, 0.01, 0.1, 1.0, and 3.0 mg/kg). At 2 hours post dosing, they were challenged with $^{125}I^-$ with carrier (33 mg/kg) by intravenous injection and euthanized at 5, 15, and 30 min, 1, 2, 6, 9, and 24 hours post dosing of iodide. This corresponds to 2.08, 2.25, 2.5, 3, 4, 8, 11, and 26 hours, respectively, after dosing with perchlorate. Blood and thyroid were harvested from all time point groups; urine was collected from rats in the 24 hours dose group. Perchlorate and iodide levels were determined in the thyroid, serum and urine.

In an additional study, three rats were intravenously dosed with 0.0, 0.1, and 1.0 mg/kg perchlorate and challenged two hours later with 33 mg/kg $^{125}I^-$. Rats were euthanized at 15 min, 1, 2, and 4 hours after they were dosed with iodide. Levels of perchlorate and $^{125}I^-$ were determined in thyroid, serum, skin and gastric contents (Yu, 2001).

6.2.1.1.2 Drinking Water Studies in Rats

Three drinking water studies (1, 5, and 14 days) were performed with target perchlorate concentrations of 0.0, 1.0, 3.0, 10.0, and 30.0 mg/kg-day with adult male rats continually exposed via drinking water. At the end of day 1, 5, or 14, rats (n=6 per group) were challenged once with 33 mg/kg $^{125}I^-$ with carrier and euthanized at 2 hours post iodide dosing. Blood and thyroid gland were collected for ClO_4^- and $^{125}I^-$ analyses in serum. For the 10 and 30 mg/kg dose groups, perchlorate was measured in serum and thyroid on day 5; however, the iodide inhibition study for these dose groups was conducted on Day 14.

6.2.1.2 Human Studies

The data used in development of the Merrill (2001d) human model were obtained from Hays and Solomon (1965) or recent data, both published and unpublished, that underwent the QA/QC check described in the introduction of this chapter (Merrill, 2001a,b). These data included the published and unpublished data from a human study of drinking water exposure to perchlorate that measured RAIU in the thyroid (Greer et al., 2000).

Data supporting model validation were obtained from another unpublished drinking water study conducted under contract to AFRL/HEST by Drs. Holger Leitolf and Georg Brabant of the Medizinische Hoschschule, Hanover, Germany. Urinary perchlorate clearance data by Eichler (1929), Kamm and Drescher (1973), and Durand (1938) were also used to validate model predictions.

6.2.1.2.1 Human Iodide Kinetic Data (Hays and Solomon, 1965)

A comprehensive human kinetic study on early iodide distribution was reported in 1965 by Hays and Solomon. The authors studied the effect of gastrointestinal cycling on iodide kinetics in nine healthy males after an iv dose of 10 μCi radiolabeled iodide ($^{131}I^-$), approximately 3.44×10^{-3} ng $^{131}I^-$/kg body weight. Frequent measurements of radioiodide uptake in the thyroid, gastric secretions, plasma, and cumulative urine samples were taken during the three hours following injection. Gastric secretions were collected using a nasogastric tube with constant suction while the subjects remained in a resting position (only standing to urinate). Saliva was not collected separately and therefore pooled, to some extent, with gastric juices. To account for the removal of gastric iodide from circulation and to determine its impact on free iodide

1 distribution, the authors ran a control session on the same subjects without aspirating gastric

2 secretions. Aspirated gastric secretions accounted for 23% of the $^{131}I^-$ administered.

3

4 **6.2.1.2.2 Perchlorate Kinetics and Inhibition of Thyroid Iodide Uptake (Greer et al., 2000)**

5 **Perchlorate data**. As described in Chapter 4, Greer et al. (2000) recently studied the

6 effects of repeated low level exposure to perchlorate on humans. Subjects received 0.5, 0.1,

7 0.02, or 0.007 mg/kg-day perchlorate in drinking water over a two week period. Each dose group

8 consisted of eight healthy volunteers (four males and four females) with no signs or symptoms of

9 thyroid disorders (euthyroid). The daily dose was dissolved in 400 mL water and divided into

10 four 100 mL servings that were ingested at approximately 0800, 1200, 1600, and 2000 hours.

11 Baseline serum and urine samples were collected before the first perchlorate treatment.

12 During perchlorate exposure, serum samples were collected at the following approximate times:

13 day 1 at 1200 and 1600, day 2 at 0800, 1200, and 1700, day 3 at 0900, day 4 at 0800 and 1200,

14 day 8 between 0800 and 0900 and day 14 at 0800 and 1700. Serum samples were also collected

15 on post-exposure days 1, 2, 3, and 14. Twenty-four hour urine collections were taken on

16 exposure days 1, 2, 14 and post-exposure days 1 through 3. Serum and 24-hour urine samples

17 from the study were provided to AFRL/HEST compliments of Dr. Monte Greer of Oregon

18 Health Science University (OHSU), Portland, OR, and Dr. Gay Goodman of Intertox, Seattle,

19 WA. The samples were analyzed for perchlorate at the Operational Toxicology Branch, Human

20 Effectiveness Directorate at the Air Force Research Laboratory (AFRL/HEST), Wright Patterson

21 Air Force Base (WPAFB), OH, using the analytical methods described in Merrill (2001d).

22 **Iodide Inhibition Data**. Eight and 24 hour thyroid $^{123}I^-$ uptakes (radioiodine uptake or

23 RAIU) were measured one to two days prior to perchlorate treatment (baseline) on days 2 and

24 14 of perchlorate exposure and 14 days after perchlorate exposure was discontinued. A gelatin

25 capsule containing 100 mCi of $^{123}I^-$ was administered orally at 0800, before the first perchlorate

26 solution for that day was drunk. Thyroid scans were then taken 8 and 24 hours later.

27 **Thyroid and Pituitary Hormone Data**. The serum samples were also analyzed for TSH,

28 T4, T3, and free T4 at OHSU. However, these hormone data were not used in the PBPK model

29 described below. Statistical analysis of the data is described in Attachment 2 of Merrill (2001d).

30 In summary, there was little effect of perchlorate on levels of T4, free T4, or T3. TSH

31 decreased significantly from baseline by Exposure Day 3. On Post-Exposure Day 1, the TSH

1 levels of the subjects in the 0.5 mg/kg-day group had decreased by an average of 35% from

2 baseline (ranging from 17% to 52%). Therefore, it appears that TSH was dropping while

3 inhibition remained the same. It is possible that there is an increase in thyroid sensitivity to TSH

4 as an early response to inhibition (Brabant et al., 1992). This increased sensitivity (possibly an

5 increase affinity of the TSH receptor) could possibly decrease circulating TSH levels while T4

6 has not decreased sufficiently yet to stimulate the hypothalamus to increased TRH secretions.

7 After perchlorate was discontinued, between Post-Exposure Days 1 and 15, the mean TSH level

8 increased significantly over baseline (23% greater than baseline), with TSH of one subject

9 remaining below baseline. The drop in TSH during perchlorate exposure and the rise above

10 baseline measurements after perchlorate seem counter-intuitive to the TSH regulation expected

11 but may be part of a rebound phenomenon as the NIS begins to upregulate.

12 In addition, the data by Greer et al. (2000) showed an increase in radioiodide uptake in

13 excess of baseline measurements 14 days after perchlorate exposure. An increase in radioiodide

14 uptake is expected due to the rise in TSH mentioned above. This rebound effect has been noted

15 in other human inhibition studies (using both iodide and perchlorate as inhibitors). Saxena et al.

16 (1962) evaluated the prophylactic doses of iodide required to suppress thyroid uptake of $^{131}I^-$ in

17 euthyroid mentally defective children. They found a minimal effective oral dose of 1500 to

18 2000 μg iodide per square meter of body surface per day was required to completely suppress

19 $^{131}I^-$ uptake. Within a week after iodide administration was stopped, a rebound of uptake was

20 noted. In some instances these uptakes were even higher in subsequent weeks.

21

22 **6.2.1.2.3 Supporting Kinetic Studies**

23 Both urine and serum perchlorate concentrations for a validation exercise were provided

24 from a recent unpublished study by Drs. Brabant and Leitolf of Medizinische Hochschule,

25 Hanover, Germany. In their study, seven healthy males ingested 12.0 mg/kg perchlorate

26 dissolved in 1 liter of drinking water every day for two weeks. The daily perchlorate dose was

27 divided equally in three portions and ingested three times per day (approximately between 0600

28 and 0800, 1100 and 1300 and 1800, and 2000 hours). Blood specimens were collected on days 1,

29 7, and 14 of perchlorate treatment and on the two mornings after perchlorate administration was

30 discontinued. Samples were analyzed for perchlorate at AFRL/HEST.

1   Three published studies reported cumulative urine concentrations collected from healthy

2   males after receiving a high oral dose of perchlorate (Durand, 1938; Kamm and Drescher, 1973;

3   Eichler, 1929). Oral doses administered in these studies were 784 mg $NaClO_4$ (635 mg ClO_4^-;

4   Durand, 1938); 1000 mg $NaClO_4$ (765 mg ClO_4^-; Kamm and Drescher, 1973), and 2000 mg

5    $KClO_4$ (1400 mg ClO_4^-; Eichler, 1929). The studies did not report serum perchlorate levels but

6   could be used to validate the model.

7   Stanbury and Wyngaarden (1952) measured radioiodide uptake in a patient with Grave's

8   disease. The patient received a tracer dose of $^{131}I^-$ as a control before perchlorate dosing and

9   again one hour after administration of 100 mg $KClO_4$. Thyroid scans of radioiodide uptake were

10   performed both after the control and perchlorate sessions to determine the level of inhibition.

11

12   ## 6.2.2 Adult Male Rat Model Development

13   This section summarizes some key features necessary to the development of the adult male

14   rat model structure and shows results of predictions made with simulations against experimental

15   data used to parameterize and validate the model.

16

17   ### 6.2.2.1 Physiologic Parameters and Tissue Partition Coefficients

18   The adult male rat volumes and blood flows were obtained from the literature or the

19   AFRL/HEST studies as described in Table 6-1. Allometric scaling was used to account for

20   parameter differences due differences in body weights between rats and humans. Because no

21   steady-state values from infusion studies were available, the partition coefficients for iodide and

22   perchlorate were estimated from the various studies listed in Table 6-2. The liver:serum and

23   muscle:serum ratios of 0.56 and 0.31 were obtained in the AFRL/HEST radiolabled perchlorate

24   ($^{36}ClO_4^-$) iv study described above. The liver:serum partition value was used to represent

25   partitioning to the liver and richly perfused compartments and the muscle:serum value to

26   represent the slowly perfused compartment.

27   For compartments with nonlinear uptake of the anions, effective partition coefficients were

28   used that represented either approximate tissue:serum concentration ratios or electrical potential

29   gradients. Chow and Woodbury (1970) measured electrochemical potentials within the thyroid

30   stroma, follicular membrane, and colloid at three different doses of perchlorate. The measured

31   difference in electrical potential between the thyroid stroma and follicle was interpreted by

1	Merrill (2001c) as an effective partition coefficient for the perchlorate and iodide anions,
2	hindering the entry of negatively charged ions into the follicle. The equal and opposite potential
3	from the follicle to the colloid enhances passage of negatively charged species into the colloid
4	and indicates an effective partition coefficient of greater than one. The equivalence between
5	electrical potential differences ϕ_i - ϕ_f and effective partition coefficients for the thyroid
6	subcompartments (stroma:follicle and follicle:colloid) were estimated in the manner of Kotyk
7	and Janacek (1977) based on the Chow and Woodbury (1970) data as described in Merrill
8	(2001c).
9	
10	**6.2.2.2 Chemical-Specific Parameters**
11	The various active transport processes, tissue permeabilities, and clearance rates (excretion)
12	are described in PBPK models for each species on a chemical-specific basis. This section
13	outlines how the values for perchlorate and iodide used in the adult male rat model were derived.
14	The values can be found in Table 6-2 and details on derivation in Merrill (2001c).
15	
16	**6.2.2.2.1 Affinity Constants and Maximum Velocities for Active Transport Processes**
17	Kinetic values for the saturable (Michaelis-Menten) active uptake process of perchlorate,
18	the affinity constant and maximum velocity capacity (Km_p and Vmaxc_p), were not available
19	in the literature nor were they determined experimentally at AFRL/HEST. Only the affinity of
20	iodide for NIS was available in the literature. The Merrill (2001c) adult rat model uses a
21	Michaelis-Menten affinity constant (Km) value of 4.0×10^6 ng/L to describe the affinity of iodide
22	(Km_i) across compartments involving active transport by NIS (e.g., in the thyroid and gastric
23	juices). This was based on the mean value of 3.96×10^6 ng/L for iodide derived by Gluzman and
24	Niepomniszcze (1983) from thyroid slices of 5 normal individuals. The thyroid slices were
25	incubated with several medium iodide concentrations. The experimentally determined Km
26	values for iodide are similar across species (Gluzman and Niepomniszcze, 1983) and across
27	different tissues (Wolff, 1998). This average literature value was therefore used for iodide in
28	tissues described with active uptake.
29	The values for perchlorate affinity were originally assumed to be the same as those for the
30	Km of iodide, due to the similar mechanism in which the two anions are transported into the
31	tissues. Thus, the iodide values were adjusted for the difference in mass to give an estimated

1   value for the affinity of perchlorate. The molar equivalent of iodide's Km for perchlorate is

2   3.1×10^6 ng/L. However, these values were not adequate for use in the models. Several

3   literature sources suggest that perchlorate may have a significantly higher affinity for NIS than

4   iodide. In his 1963 paper (Wolff and Maurey, 1963) and his 1998 review, Wolff concluded that

5   perchlorate has a greater affinity than iodide for the NIS. This assumption was based upon his

6   work with iodide, perchlorate, and several other anions actively sequestered in the thyroid.

7   Wolff measured the Km of a few of the anions and inhibition constants (Ki's) for several ions,

8   including perchlorate. As noted in Chapter 2, Wolff found that the relative potency of inhibition

9   by the various anions could be described with the following series: $TcO_4^->ClO_4^->ReO_4^->SCN^-$

10 $>BF_4^- > I^->NO_3^->Br^->Cl^-$. Wolff reported that the measured Km values for several of these

11 inhibiting anions were not the same as those measured for iodide. In fact, measured values for

12 Km and Ki for several of the inhibiting anions revealed that affinity increased with increased

13 inhibitory potency.

14   Several studies suggest perchlorate is a more potent inhibitor than iodide. In the rat

15 thyroid, Wyngaarden et al. (1952) have shown that perchlorate was a more powerful inhibitor of

16 the iodide trap than thiocyanate. Halmi and Stuelke (1959) showed that perchlorate was ten

17 times as effective as iodide in depressing tissue to blood ratios in the rat thyroid and gut.

18 Similarly, Harden et al. (1968) compared human saliva to plasma radioiodide concentration

19 ratios after equimolar doses of perchlorate and iodide. The saliva:plasma iodide ratios during

20 resting conditions were approximately seven times lower after a molar equivalent dose of

21 perchlorate versus iodide. Lazarus et al. (1974) also demonstrated that perchlorate was taken up

22 to greater extent in mice salivary glands than iodide. These studies, in addition to the work of

23 Chow et al. (1969), support the use of a lower Km for perchlorate uptake in the tissues with

24 sodium iodide symporter. Based on this information, a value of 2.5×10^5 ng/L for the thyroid

25 (Km_Tp) and 2.0×10^5 ng/L for skin (Km_Sp) or gut (Km_Gp), approximately 10 times lower

26 than that of iodide, was estimated by Merrill (2001c,d) to represent perchlorate's affinity for

27 transport by the NIS.

28   The apical follicular membrane (between the thyroid follicle and colloid) also exhibits a

29 selective iodide uptake mechanism. Golstein et al. (1992) measured a Km value of

30 approximately 4.0×10^9 ng/L for the transport of iodide between the thyroid follicle and colloid

31 (Km_DTp) in bovine thyroid. This iodide channel also appears to be very sensitive to

1   perchlorate inhibition and shares a similar permeability to perchlorate as to iodide. The ability of

2   perchlorate to inhibit iodide uptake at the apical follicular membrane suggests that the Km of

3   perchlorate at the apical follicular membrane (Km_Dtp) is also lower than that of iodide. Model

4   simulations of thyroid inhibition supported a value of 1.0×10^8 ng/L, approximately ten times

5   less than that of iodide.

6   Whereas the Km is similar across tissues containing NIS, the maximum velocity term

7   (Vmaxc) does vary between tissues and species (Wolff, 1998), being lower in humans than other

8   species (Gluzman and Niepomniszcze, 1983; Wolff and Maurey, 1961). Maximum velocities or

9   capacities (Vmaxc) were not found in the literature and were estimated for a given compartment

10   by fitting the simulation to the data at varying doses.

11

12   **6.2.2.2.1 Effective Partitions, Permeability Area Cross Products and Clearance Values**

13   Permeability area cross products and partition coefficients were used to describe diffusion

14   limited uptake in tissues requiring subcompartments. The permeability area values in the Merrill

15   (2001c) model were fitted by setting the partition coefficients to the literature values in

16   Table 6-2. Fitted clearance values were used to describe first-order urinary excretion rates and

17   reversible plasma binding to serum. Equations for these representations are provided in Merrill

18   (2001c).

19

20   **6.2.2.3 Adult Male Rat Model Simulation Results and Validation**

21   The simulations shown in this section result from exercising the model with the

22   physiological and chemical-specific parameters provided in Tables 6-1 and 6-2. Figure 6-7

23   illustrates the model predictions versus data time course for the iv radiolabeled perchlorate study

24   described in Section 6.2.1.1.1. The model produced good simulations for the trend of the data

25   but slightly over predicts the thyroid concentrations at later time points (Panel A). Model

26   predictions fit the data well for perchlorate concentrations in the serum (Panel B) and kidney

27   (Panel C), as well as the amount excreted in the urine (Panel D). Other tissue concentrations not

28   shown herein also were predicted well by the model (Merrill, 2001c).

29   Figure 6-8 shows that plasma binding of perchlorate was necessary to provide adequate

30   model predictions. Thyroid, serum, and urine were collected from the iv studies described in

31   Section 6.2.1.1.1 using cold (i.e., not radiolabeled) perchlorate at 0.01, 0.1, 1.0, and 3.0 mg/kg.
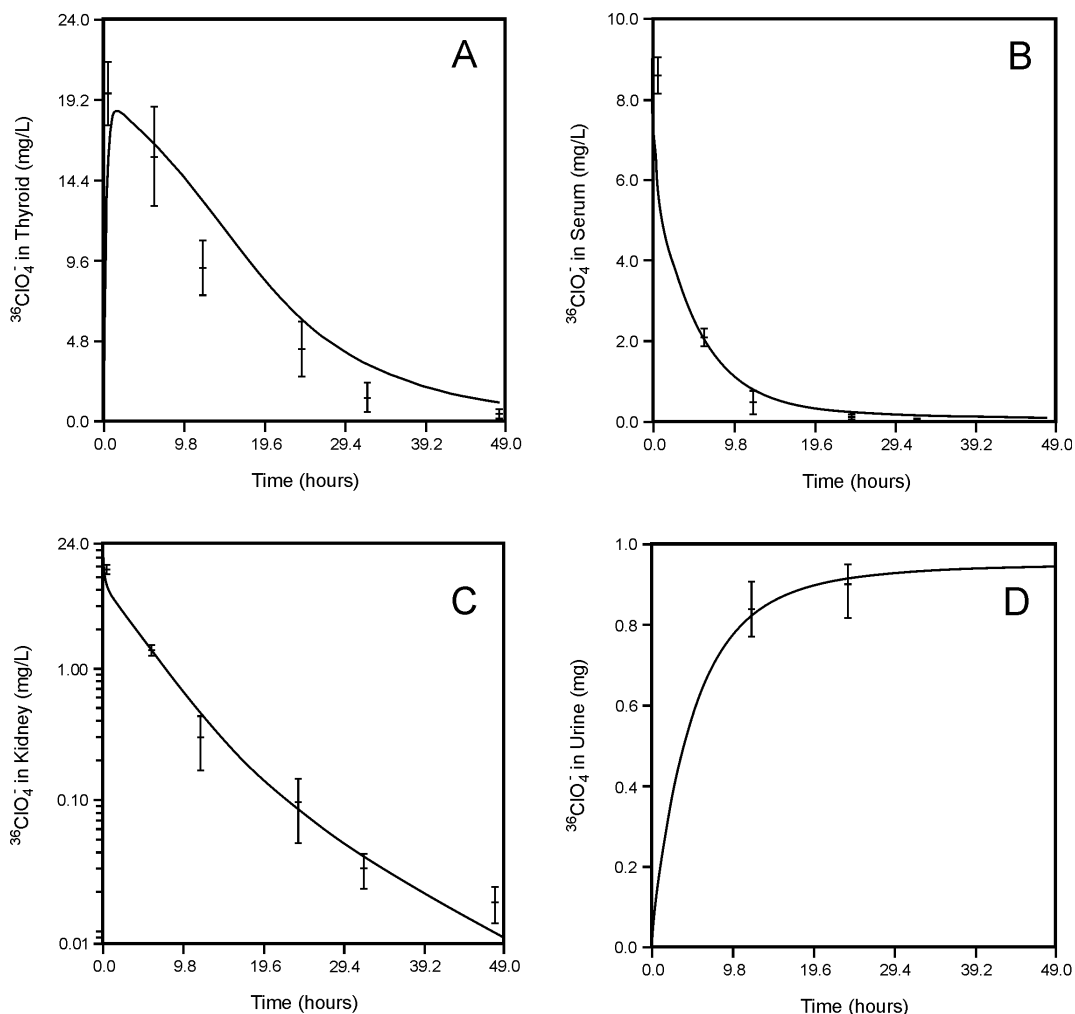
Figure 6-7. **Adult male rat PBPK model predictions after an acute iv dosing with radiolabeled perchlorate ($^{36}ClO_4^-$). Panels A and B show model predictions (lines) versus data time course (mean ± SD) of labeled perchlorate (mg/L) in the thyroid and serum. Panel C shows model predictions versus data time course of labeled perchlorate (mg/L) in the kidney. Panel D shows cumulative excretion (mg) of labeled perchlorate in the urine (Merrill, 2001c).**

1   Model predictions without plasma binding (Panel A, left) resulted in an underestimation of

2   serum perchlorate concentrations at the 1 mg/kg-day dosage level and below. Low serum

3   predictions suggested either greater uptake into other tissues or protein binding. To provide

4   better estimates of perchlorate serum concentrations at the 0.01 and 0.1 mg/kg doses, Merrill

5   (2001c) added protein binding to the venous blood compartment of the model. An affinity
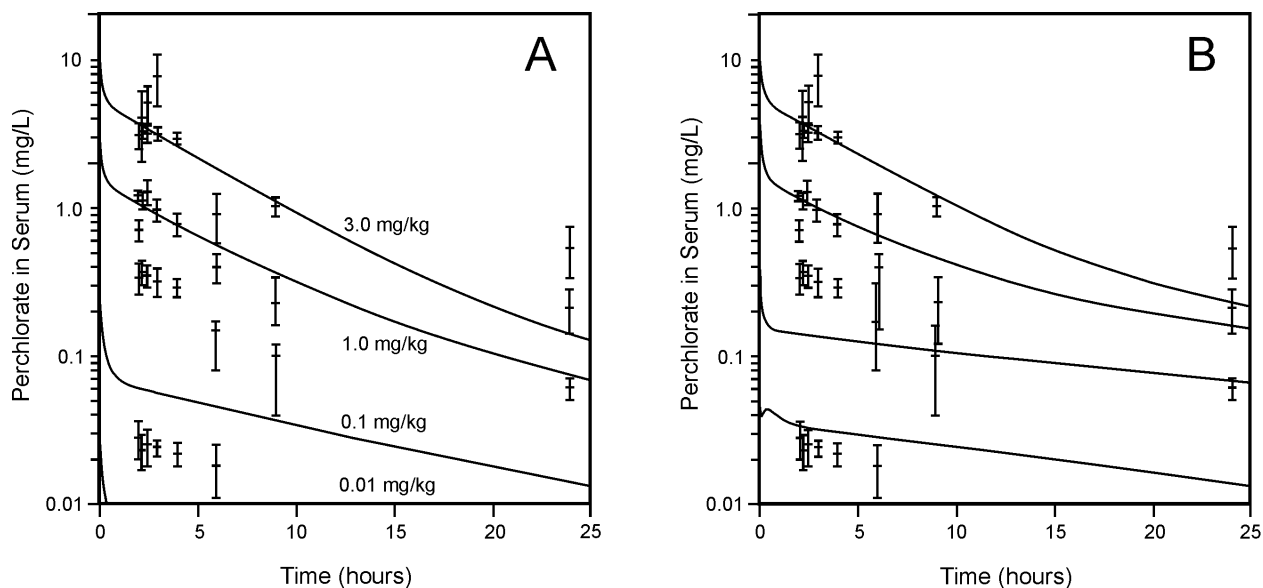
Figure 6-8. **Simulations illustrating the necessity of including plasma binding in the adult male rat PBPK model structure (Merrill, 2001c). Model predictions (lines) versus data time course (mean ± SD) of perchlorate concentration (mg/L) in serum after doses of 3.0, 1.0, 0.1 and 0.01 mg/kg-day are shown in Panel A without and in Panel B with plasma binding. Only part of the simulation for the 0.01 dose in Panel A can be seen in the lower left corner. Data of Yu (2000).**

1   constant for this binding of perchlorate in the blood (Km_Bp) of 1.1E6 ng/L and a maximum

2   velocity capacity for this blood binding (Vmaxc_Bp) of 9.3E3 ng/h/kg was fitted to serum levels

3   from doses ranging 0.01 to 3.0 mg/kg (Panel B, right). The model underpredicts serum

4   perchlorate from the 0.1 mg/kg dose group; but it fits serum at 0.01 mg/kg and cumulative urine

5   across the doses. Interestingly, the urinary excretion at 0.01 mg/kg was lower than the other

6   doses, accounting for elevated serum concentrations. Mean 24 hour urinary excretions (± SD) of

7   perchlorate were approximately 97% (± 2), 72% (± 1), 87% (± 17), and 91% (± 13) of the

8   administered iv dose for the 0.01, 0.1, 1.0, and 3.0 mg/kg dose groups, respectively.

9   The literature discussed in Chapter 3 and in Merrill (2001c) suggests that serum albumin is

10   the major binding protein; however, it does not confirm that albumin is the only binding site.

11   Merrill (2001c) notes that no studies were found that evaluated whether perchlorate or similar

12   anions bind to thyroglobulin. However, Yamada (1967) studied the effects of perchlorate and

1   other anions on T4 metabolism and noted a significant decrease in serum protein-bound iodide

2   (PBI) in thyroidectomized T4-maintained perchlorate-fed rats. In a 1968 in vitro study, Yamada

3   and Jones reported that T4 was displaced from plasma protein as indicated by an uptake of T4 by

4   muscle in the presence of plasma taken from perchlorate-fed rats. This suggested, but did not

5   demonstrate directly, that perchlorate interferes with T4 binding with plasma proteins.

6   Pertechnetate is known to bind to plasma proteins. Hays and Green (1973) studied the

7   blocking of pertechnetate binding with human serum proteins by other anions. Perchlorate was

8   found to be one of the most effective, while iodide was ineffective. In dialysis studies, inorganic

9   iodide did not bind to plasma proteins. The pertechnetate binding appeared to be reversible in

10   serum.

11   Simulations of thyroid perchlorate concentrations and of the amount of perchlorate excreted

12   in the urine from the four dose groups are shown in Figure 6-9. It was noted that the thyroid

13   concentrations resulting from the 3.0 mg/kg cold perchlorate study were slightly higher than

14   those from the radiolabeled perchlorate ($^{36}ClO_4^-$) study at 3.3 mg/kg (Figures 6-9A and 6-7A,

15   respectively). This may reflect the analytical differences in measuring cold versus radiolabeled

16   perchlorate. The model slightly underpredicts the thyroid concentrations at 3.0 mg/kg, based on

17   the cold perchlorate data (Figure 6-9A), and slightly overpredicts the $^{36}ClO_4^-$ thyroid

18   concentration at 3.3 mg/kg (Figure 6-7A).

19   The model is able to adequately predict data from studies that were not used in the

20   development process. Figure 6-10 shows the model predictions versus the data of Chow and

21   Woodbury (1970) and Eichler (1929). Model predictions fit the data well for radiolabeled

22   perchlorate concentration in the thyroid (A); whereas, the serum (B) is underpredicted. Merrill

23   (2001c) notes the difference and provides some plausible explanations. The rats in the Chow and

24   Woodbury (1970) study were functionally nephrectomized by ligating the renal pedicle of both

25   kidneys and given the radiolabeled perchlorate ip. Analytical differences between AFRL/HEST

26   and Chow and Woodbury could exist, and it is also possible that the nephrectomization creates

27   physiological changes that can not be accounted for sufficiently by "turning off" urinary

28   excretion in the model simulations. One hypothesis is that saturation in NIS-containing tissues

29   occurs to a lesser extent as a result of increased extracellular sodium cation (Na^+) and possibly

30   other competitive anions when renal clearance is blocked, thereby increasing the arterial

31   radiolabeled perchlorate. While the underprediction in serum would suggest the need for an
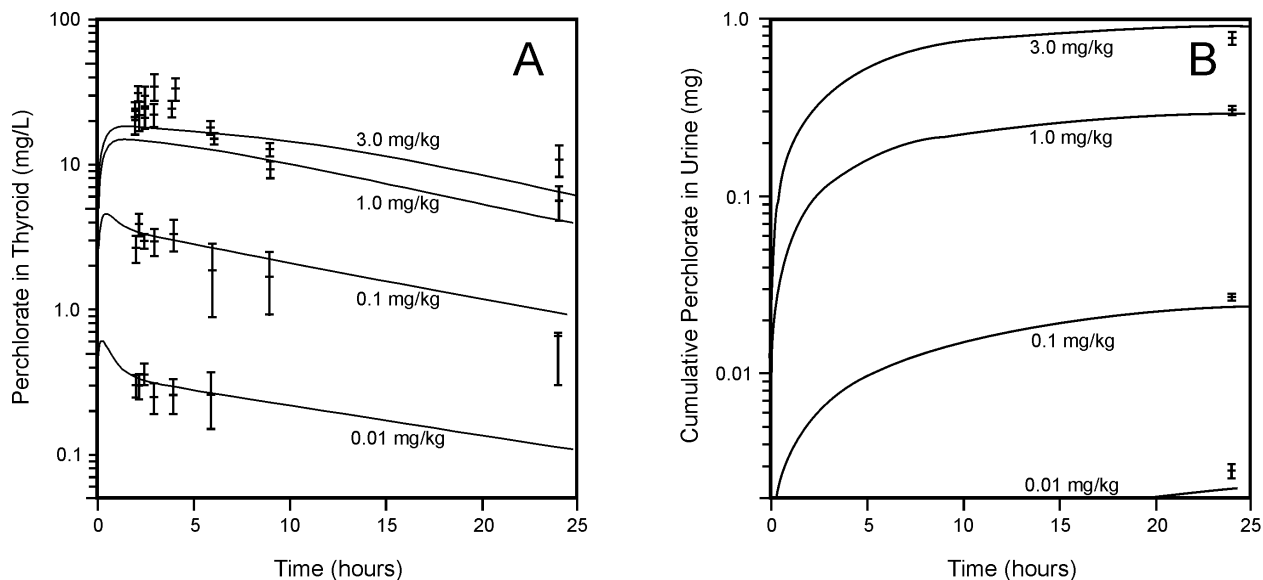
Figure 6-9. Adult male rat PBPK model predictions (lines) versus data time course (mean ± SD) of perchlorate concentrations in the thyroid (mg/L) in Panel A or cumulative excreted perchlorate in the urine (mg) in Panel B (Merrill, 2001c). Male rats were dosed iv with 3.0, 1.0, 0.1 or 0.01 mg/kg-day perchlorate (Yu, 2000).

1   increased binding constant for perchlorate, this was not consistent with the data from

2   AFRL/HEST for studies at lower doses (Merrill, 2001c). Panel C in Figure 6-10 shows the

3   model predictions versus the data of Eichler (1929) for cumulative perchlorate excreted in the

4   urine. These rats were given perchlorate subcutaneously (sc) at doses of 1.6, 8.0, and 49 mg/kg.

5   The adult male rat model (Merrill, 2001c) is also able to predict iodide distribution.

6   Figure 6-11 shows the model predictions versus a time course for radiolabeled iodide data from

7   the AFRL/HEST experiments outlined in Section 6.2.1.1.1. Adequate fit is demonstrated for

8   both the thyroid and serum concentrations at doses of radiolabeled iodide differing by an order of

9   magnitude (0.033 and 0.33 mg/kg).

10   Figure 6-12 demonstrates the fit of the model simulations of perchlorate thyroid

11   concentration (mg/L) after drinking water exposures to perchlorate. The model was coded to

12   simulate oral dosing for 12 hours per day, assuming that rats drink fairly continuously during

13   their waking hours. The same perchlorate parameters used to describe the "acute" (iv) kinetics

14   also adequately described serum concentrations from these "chronic" drinking water exposures
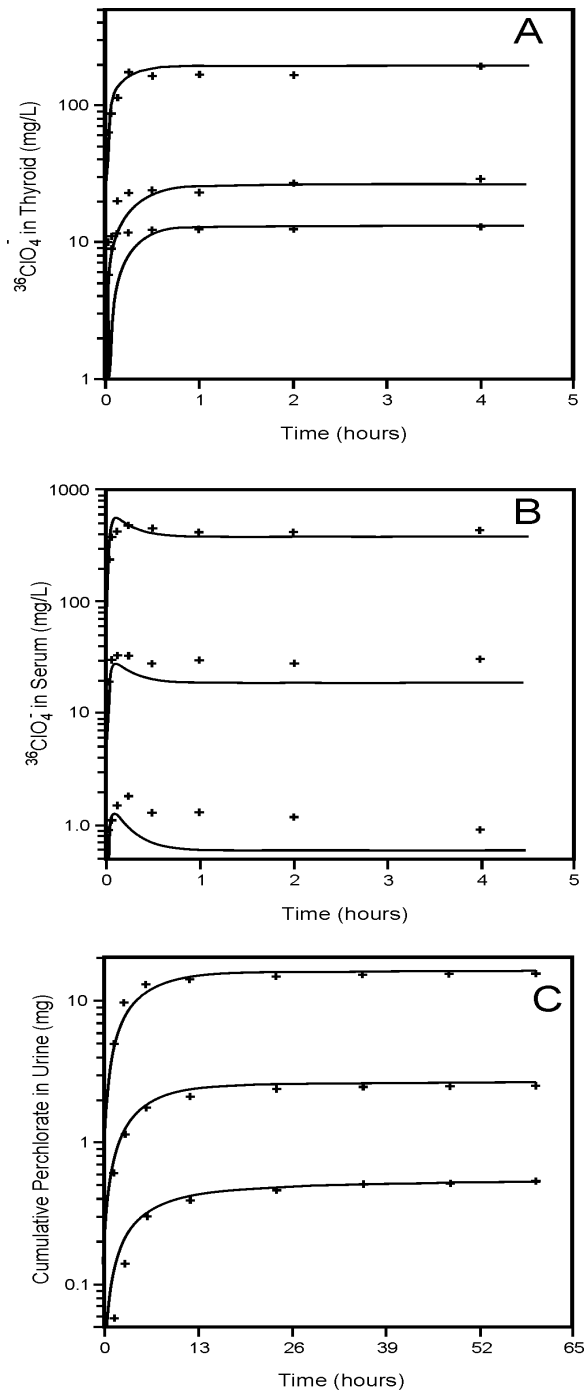
Figure 6-10. Validation for male rat PBPK model of perchlorate disposition (Merrill, 2001c). Model predictions (lines) versus data time course for concentrations (mg/L) in the thyroid (A) and serum (B) for ip administration in rats of 200, 10, and 0.5 mg/kg $^{36}ClO_4^-$ (data of Chow and Woodbury, 1970). Panel C shows model predictions (lines) and data time course for cumulative perchlorate in the urine (mg) of male rats after subcutaneous doses of 1.6, 8.0, and 49 mg/kg (data of Eichler, 1929).
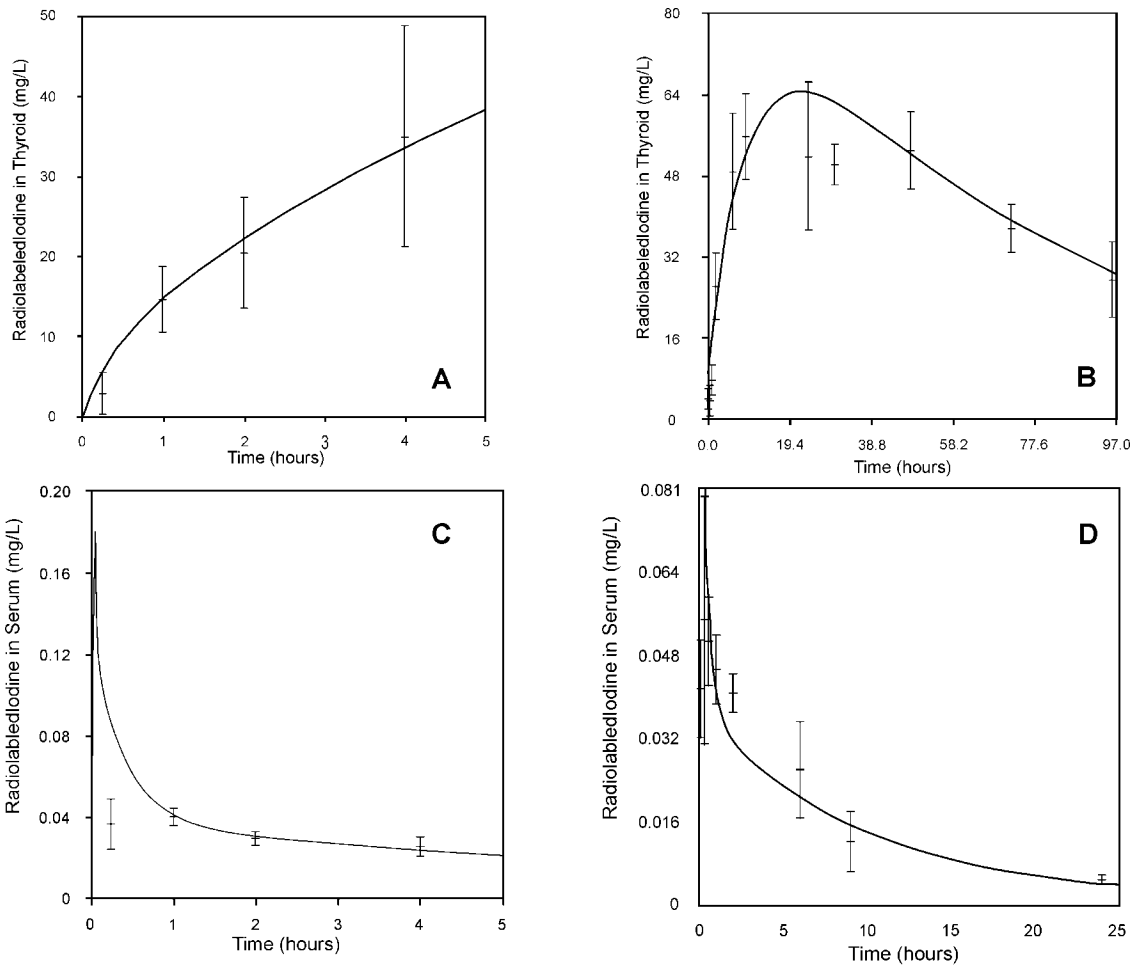
Figure 6-11. **Male rat PBPK model (Merrill, 2001c) predictions (lines) versus data time course (mean ± SD) of iodide concentrations (mg/L) at two doses of $^{125}I^-$ with carrier, 0.033 mg/kg or 0.33 mg/kg, in the thyroid (A) or (B) and in the serum (C) or (D). Data of Yu (2001).**

1   (data shown in Merrill, 2001c) but failed to predict thyroid concentrations from the 3.0 mg/kg-

2   day dose and higher. TSH in these same studies was increased during drinking water exposure

3   across all doses so that Merrill (2001c) accounted for the TSH-induced upregulation in the NIS

4   by fitting an increased effective thyroid follicle:stroma partition coefficient (PT_p) at these

5   higher doses. Merrill (2001c) noted that TSH is not expected to increase NIS in tissues other than

6   the thyroid (Brown-Grant, 1961) and that these simulations agree. Given the small size of the

7   thyroid, its upregulation would not decrease serum concentrations significantly. This explains
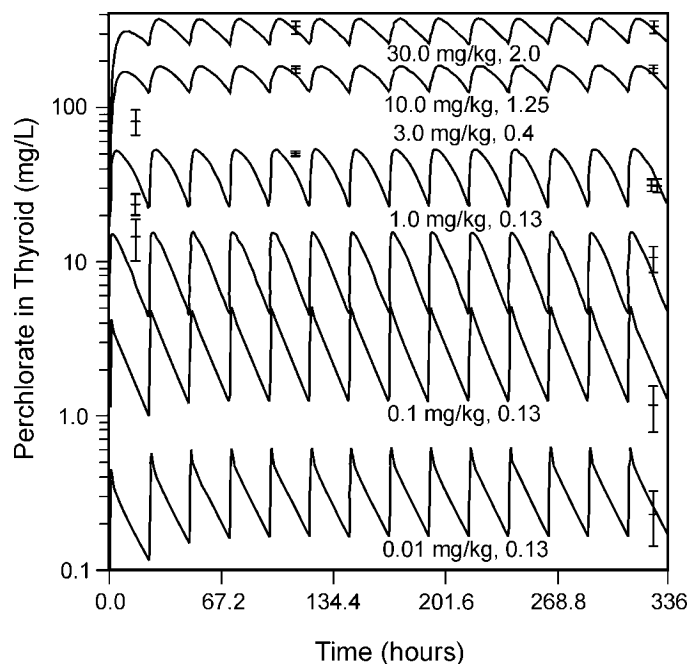
Figure 6-12. **Male rat PBPK model predictions (lines) versus data time course (mean ± SD) of thyroid perchlorate concentrations (mg/L) in male rats during ingestion of 30, 10, 3.0, 1.0, 0.1, or 0.01 mg/kg-day in drinking water for 14 days (Merrill, 2001c). Data across the doses were fit by increasing the thyroid follicle:stroma effective partitioning for perchlorate (PT_p) from 0.13 to 0.4, 1.25, and 2.0 at the 3, 10, and 30 mg/kg-day doses.**

1   why the model successfully predicted serum perchlorate concentrations across drinking water

2   doses with the same parameters used to describe acute exposures and why it could not predict

3   thyroid concentrations above 3 mg/kg-day.

4   It could be expected that other parameters (e.g., follicle size and follicular Vmaxc) would

5   also increase with TSH stimulation. There is an increase in percent of thyroid volume attributed

6   to the follicle cells (Conde et al., 1991; Ginda et al., 2000), total protein, RNA and DNA content,

7   and the incorporation of labeled amino acids into protein (Pisarev and Kleiman de Pisarev,

8   1980). However, Merrill (2001c) notes that adequate predictions could be achieved by adjusting

9   additional parameters; although, without incorporation of regulation by the hypothalamic-

10   pituitary-thyroid axis, such adjustments provide little additional insight.

11   The ability of the adult male rat model to predict iodide uptake inhibition in the thyroid is

12   demonstrated in Figure 6-13 for a single iv dose of perchlorate (right) or for a 14-day drinking
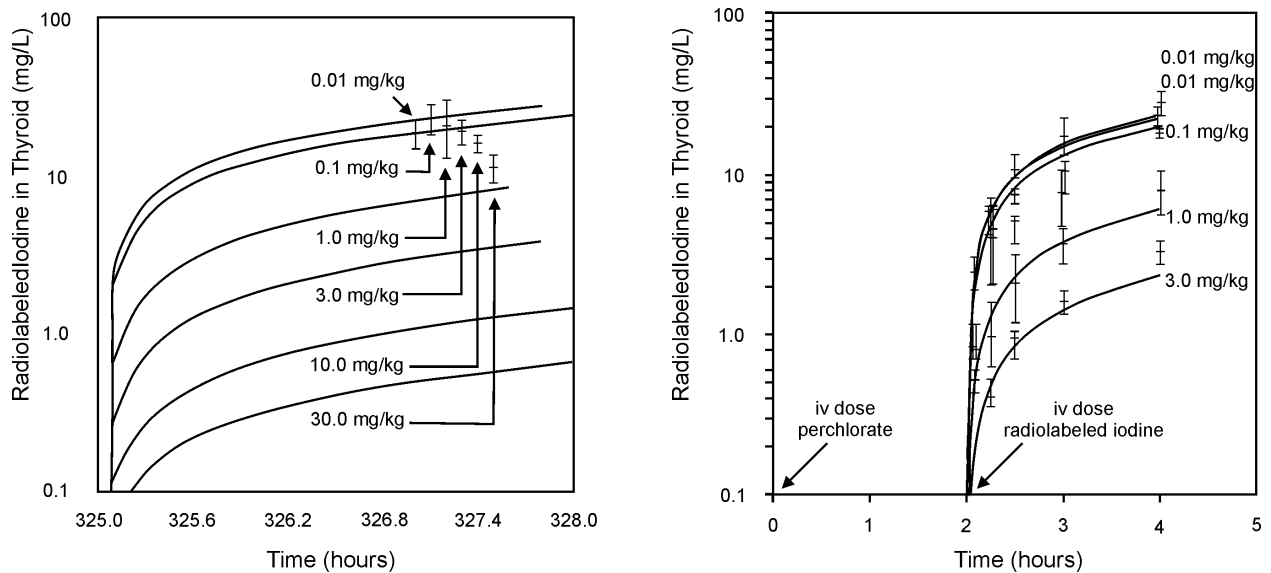
Figure 6-13. **Male rat PBPK model predictions (lines) versus data time course (mean ± SD) of iodide uptake inhibition in male rats administered perchlorate either by a single iv dose (right) or in drinking water for 14 days (left), followed by an iv dose of 33 ug/kg [125]I⁻ with carrier (Merrill, 2001c). Perchlorate doses were 3.0, 1.0, 0.1, and 0.01 mg/kg-day. Inhibition at the 0.01 and 0.1 mg/kg-day doses overlaps for the iv dose (right).**

1   water exposure (left). Perchlorate-induced inhibition of [125]I⁻ uptake in the thyroid was 13, 24, 70,

2   and 88% at 2 hours and 11, 29, 55, and 82% at 9 hours after iv dosing with [125]I⁻ with carrier for

3   the 0.01, 0.1, 1.0, and 3.0 mg/kg dose groups. Good simulations were achieved across doses.

4   However, at 3.0 mg/kg, the model slightly overpredicts inhibition 6 hrs after the perchlorate dose

5   (4 hours after [125]I⁻ administration). TSH was measured from the highest dose level (3.0 mg/kg)

6   between 8 and 48 hours post dosing and was found to increase between 8 and 12 hrs. It is

7   possible that TSH was already elevated at 6 hrs, allowing upregulation of the thyroid to

8   compensate for inhibition at that time point, which the model would not predict. Yu (2000)

9   provides greater details on hormone fluctuations resulting from the AFRL/HEST experiments.

10   With respect to iodide inhibition after 14 days of drinking water exposure to perchlorate at

11   0.01, 0.1, 1.0, 3.0, 10.0, and 30.0 mg/kg-day (Figure 6-13, left), the model overpredicts inhibition

12   at the 1.0 mg/kg-day dosage and greater. TSH-induced upregulation of the thyroid compensates

13   for competitive inhibition, resulting in little or no inhibition of radioiodide uptake on Day 14 of

1 exposure in all dose groups except 30 mg/kg-day. In all treated groups, TSH levels were already

2 increased after the first day. Serum T4 initially decreased in all dose groups except the

3 0.01 mg/kg-day group. By day 14, T4 levels had increased to control values in the 0.1 and

4 1.0 mg/kg-day dose groups. FT4 increased in all dose groups on day 1, returned to normal values

5 by day 5, and were significantly elevated across all dose groups by day 14 (except the 0.1 mg/kg-

6 day group).

7

6.2.3 Human Model Development

9   The adult human PBPK model (Merrill, 2001d) was developed concurrently with that for

10 the adult male rat (Merrill, 2001c) and updates the preliminary structure provided to EPA

11 (Merrill, 2000). Much of the early development was based upon generalizations from previous

12 AFRL/HEST work on perchlorate (Fisher, 1998a; 2000) and the work of Hays and Wegner

13 (1965) describing iodide kinetics. As discussed above and shown in Figure 6-1, a nearly

14 identical model structure to that of the adult male rat was used for the adult human. The human

15 physiological parameters will of course be different as these should be species-specific. This

16 section will only highlight notable differences in parameter sources between the two models.

17

6.2.3.1 Physiologic Parameters and Tissue Partition Coefficients

19   Human tissue volumes and blood flows were obtained from the literature as shown in

20 Table 6-1. Merrill (2001d) notes that considerable variability was reported for some parameters.

21 For example, blood flow to the gastrointestinal (GI) tract can increase ten-fold in response to

22 enhanced functional activity (secretion and digestion) (Granger et al., 1985). Blood flows used

23 in the model represent estimates of resting values. Human data on the volume of the gut

24 capillary bed (VGBc) were not found in the published literature. Therefore, Merrill (2001d) used

25 a value derived from rat stomach data (Altman and Dittmer, 1971a) for the volume of the

26 gastrointestinal blood (VGBc) in the human model.

27   Thyroid volume was obtained from ultrasound measurements on 57 healthy volunteers with

28 no thyroid disorders (37 to 74 years of age) in a study conducted by Yokoyama et al. (1986). The

29 mean thyroid volume was 13.4 ± 4.1 mL and mean thyroid volume to body weight ratio was

30 0.251 ± 0.074 mL/kg (mean $\pm$ SD), approximately 0.03% of body weight. Yokoyama et al.

31 (1986) found a positive correlation between thyroid volume and both body weight and age, with

1   weight having the most pronounced influence. The percent of total thyroid volume attributed to

2   the thyroid follicular epithelium, colloid, and stroma were estimated from histometric

3   measurements of patients at necropsy by Brown et al. (1986). Their findings on the histological

4   features of thyroids of men and women showed overlapping distributions without evidence of a

5   significant difference between sexes. However, a significant sex difference in total fat mass is

6   reported in humans, with women having approximately 10% more fat than men (Brown et al.,

7   1997). Based on these data, Merrill (2001d) used a gender-specific value for this parameter.

8

9   **6.2.3.2 Chemical-Specific Parameters**

10   The various active transport processes, tissue permeabilities, and clearance rates (excretion)

11   are described in PBPK models for each species on a chemical-specific basis. This section

12   outlines how the values for perchlorate and iodide used in the human model were derived. The

13   values can be found in Table 6-2, and the details on derivation are in Merrill (2001d).

14

15   **6.2.3.2.1 Affinity Constants and Maximum Velocities**

16   The Michaelis-Menten affinity constant (Km) estimates for perchlorate and iodide in the

17   various tissues with active transport were developed in the human in an analogous fashion to that

18   in the rat, as described above in Section 6.2.2.2., based on Golstein et al. (1992), Gluzman and

19   Niepomniszcze (1983), and Wolff (1998). The maximum velocity capacity (Vmaxc) values were

20   estimated for the various compartments by fitting the simulations to available data at various

21   doses (Merrill, 2001d).

22

23   **6.2.3.2.2 Effective Partitions, Permeability Area Cross Products, and Clearance Values**

24   Permeability area cross products and clearance values for perchlorate and iodide were

25   developed by fitting to literature values in an analogous fashion to that for the rat described in

26   Section 6.2.2.3 (Merrill, 2001d).

27

28   **6.2.3.3 Adult Human Model Parameterization and Validation**

29   The human PBPK model for iodide was developed based on the data of Hays and Solomon

30   (1965) described in Section 6.2.1.2.1. Model predictions versus the data are shown in

31   Figure 6-14 for iodide concentrations (ng/L) in the serum (A), thyroid (B), and gastric juice (C);
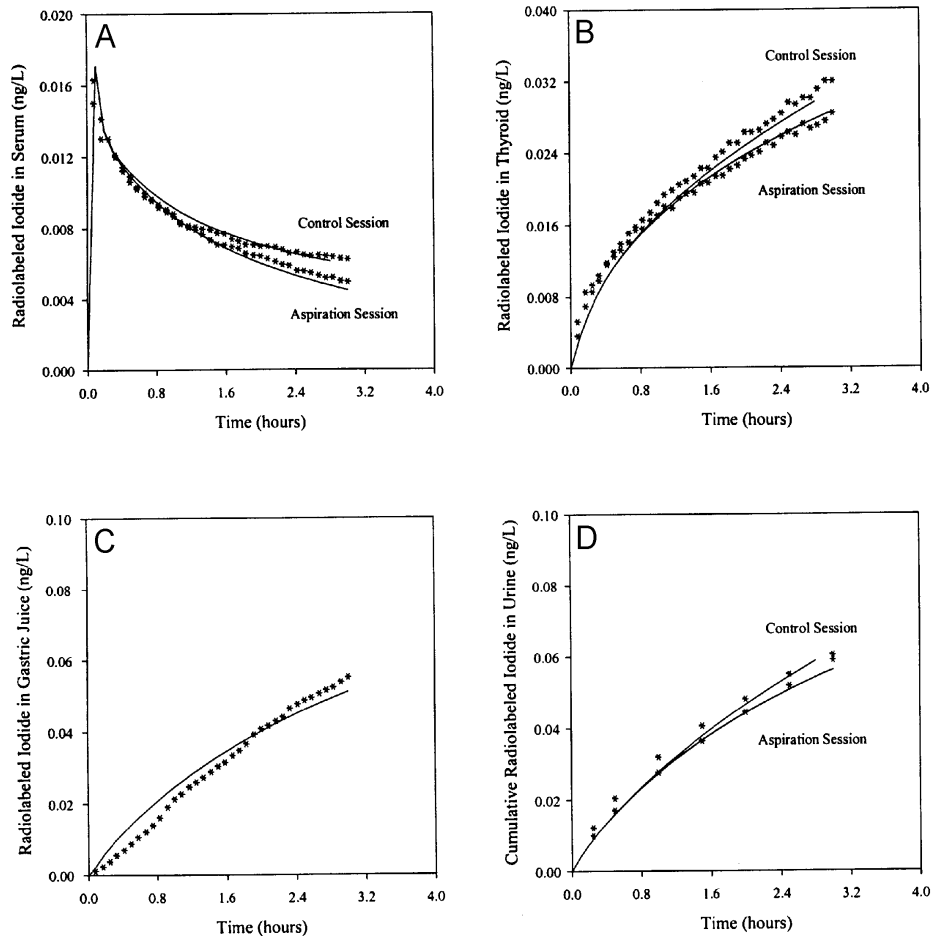
Figure 6-14. Human PBPK model (Merrill, 2001d) predictions (lines) versus mean $^{131}I^-$ concentration (mg/L) time course (asterisks) in serum (A), thyroid (B), gastric juice (C), and urine (D). Data of Hays and Solomon (1965) are for nine healthy males dosed with 10 μCi $^{131}I^-$ (approximately 3.44 ng/kg).

1   cumulative iodide excreted in the urine (ng) is shown in D. In this study, aspirated gastric juice

2   accounted for an average of 23% of the iv dose within 3 hours after iv injection with radiolabeled

3   iodide ($^{131}I^-$) (Merrill, 2001d). Simulation of the gastric juice removed during the aspiration

4   session (Figure 6-14, C) required mathematically removing the amount of $^{131}I^-$ reabsorbed by the

5   stomach wall. This was accomplished by adjusting the rate of reabsorption of $^{131}I^-$ from gastric

6   juice to gastric tissue during the aspiration session as described in Merrill (2001d). The Vmaxc

7   values for the gut and thyroid were then obtained by fitting values of $^{131}I^-$ uptake into gastric juice

1 from the aspiration session (lower lines in Figures 6-14; B and C). The urinary clearance value

2 was fitted to simulate both cumulative urine content and serum iodide concentration from the

3 aspiration session data (lower lines in Figures 6-14; A and D). Once parameters were established

4 using the aspiration session, the rate of change in the gastric juice and partitioning back into the

5 gastric juice from the systemic circulation was fitted to predict the corresponding increase in

6 $^{131}I^-$ in plasma, thyroid, and urine seen in the control session versus the aspiration session (upper

7 lines in Figures 6-14; A, B and D).

8 Figure 6-15 illustrates that, as for the adult male rat model, plasma binding of perchlorate

9 was necessary to fit the serum concentration data of the 14-day study by Greer et al. (2000). The

10 model indicates that humans have a lower binding capacity for perchlorate than rats.

11 For example, the Vmaxc value for perchlorate is 9.310^3 ng/hr-kg in the male rat versus 5.0×10^2

12 ng/hr-kg in the human. Merrill (2001d) noted that while the effect of the plasma binding is

13 subtle at 0.5 mg/kg-day dose, including the plasma binding improved the fit for uptake and

14 clearance at the 0.1 and 0.02 mg/kg-day dosage levels.
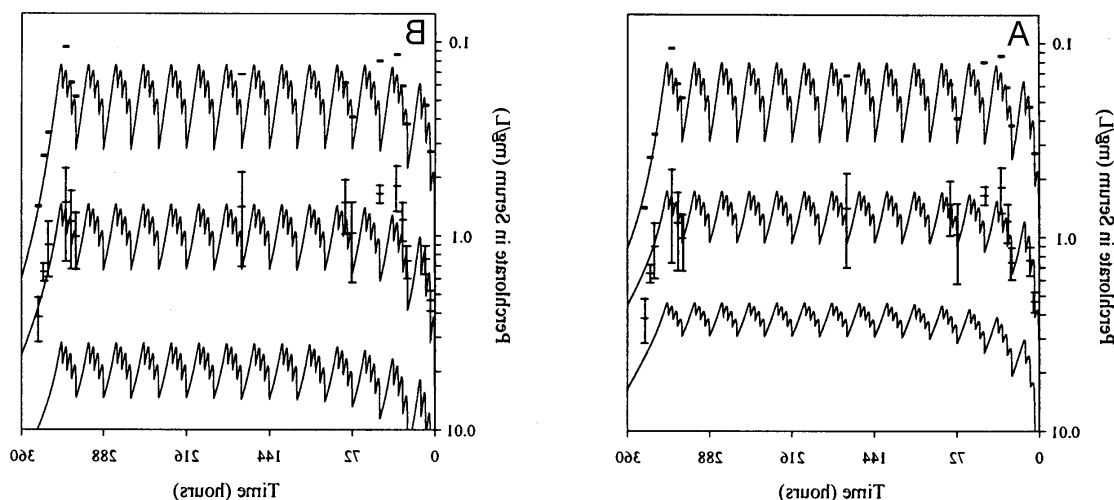
Figure 6-15. Simulations illustrating the necessity of including plasma binding in the human PBPK model structure (Merrill, 2001d). Model predictions (lines) versus data time course (mean ± SD) are shown with (A) and without (B) plasma binding for serum concentrations (mg/L) from 4 male subjects dosed with perchlorate at 0.5, 0.1, or 0.02 mg/kg-day for 14 days (data of Greer et al., 2000).

1   Cumulative urinary perchlorate excretion (mg) predictions versus the data (mean ± SD) at

2   each dosage level are shown in Figure 6-16. Merrill (2001d) also simulated serum concentration

3   (mg/L) and cumulative urinary perchlorate levels (mg) for each individual in the 0.5, 0.1, and

4   0.02 mg/kg-day dose groups of the Greer et al. (2000) study. An average value for urinary

5   clearance of perchlorate (ClUc_p) of 0.126 L/hr-kg (± 0.050) was calculated from the

6   individually fitted values. Figures 6-17 and 6-18 show a representative plot of model prediction

7   versus individual subject data at the 0.5 and 0.1 mg/kg-day dosage. Additional plots provided in

8   Merrill (2001d) provide an appreciation for the high degree of variability in the data.

9

10
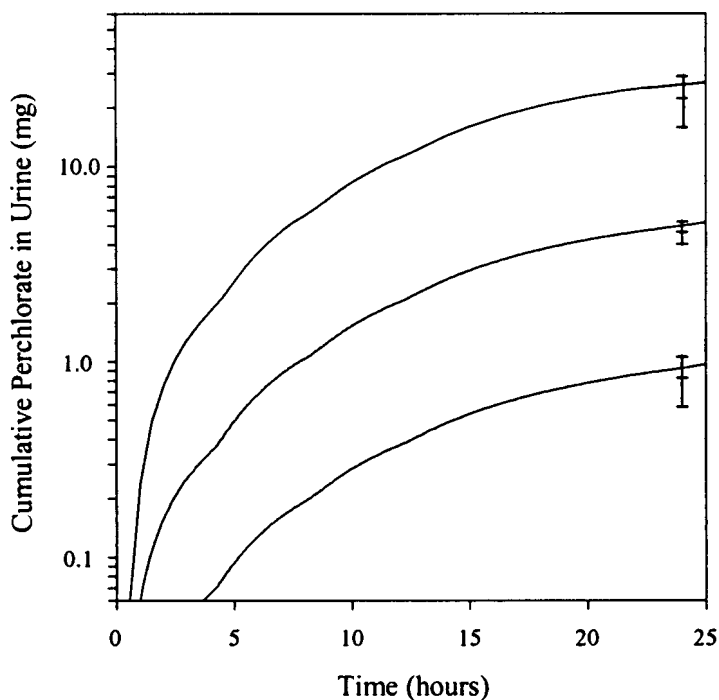
Figure 6-16.   **Human PBPK model predictions (lines) versus data (mean ± SD) of the observed cumulative urine excretion (mg) in male subjects dosed with perchlorate 0.5, 0.1, or 0.02 mg/kg-day for 14 days. Model of Merrill (2001d) and data of Greer et al. (2000).**

1   Serum perchlorate levels were not available for the 0.02 mg/kg-day dose group, but

2   cumulative urinary excretion amounts (mg) for this group were fitted using the average
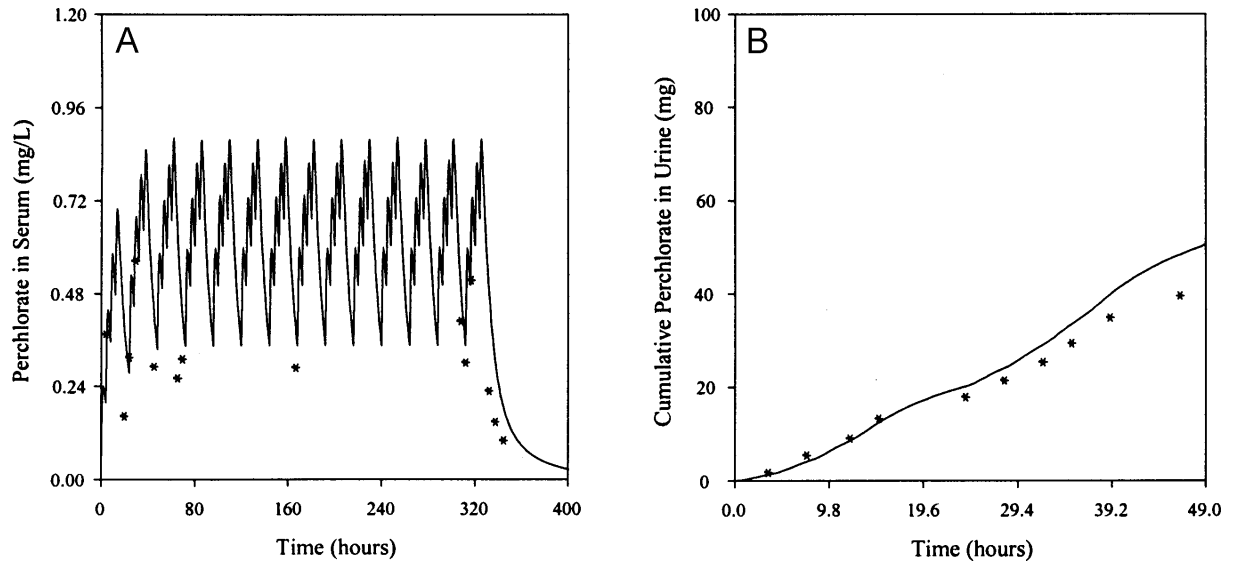
Figure 6-17. Human PBPK model predictions (lines) versus data of one subject's serum perchlorate concentration (mg/L) shown in (A) and corresponding 48-hour cumulative urine perchlorate (mg) shown in (B). Subject consumed 0.5 mg/kg-day perchlorate in drinking water, 4 times per day, for 14 days. Model predictions for the individual obtained by using study average value of all subjects for urinary clearance of perchlorate (ClUc_p). Model of Merrill (2001d) and data of Greer et al. (2000).
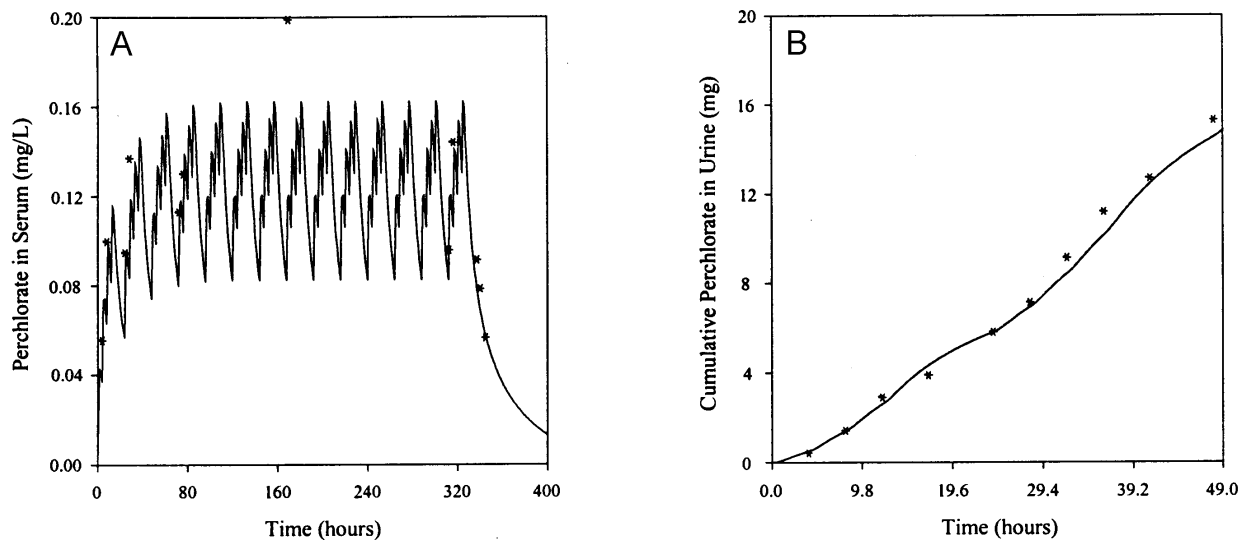
Figure 6-18. Human PBPK model predictions (lines) versus data of one subject's serum perchlorate concentration (mg/L) shown in (A) and corresponding 48-hour cumulative urine perchlorate (mg) shown in (B). Subject consumed 0.1 mg/kg-day perchlorate in drinking water, 4 times per day, for 14 days. Model predictions for the individual obtained by using study average value of all subjects for urinary clearance of perchlorate (ClUc_p). Model of Merrill (2001d) and data of Greer et al. (2000).

1   perchlorate urinary clearance (ClUC_p) value of 0.126 L/hr-kg calculated from the individual fits

2   for the 0.1 and 0.5 mg/kg-day groups. Figure 6-19 shows the model predictions versus 48-hour

3   cumulative urine perchlorate (mg) for two different subjects.

4

5
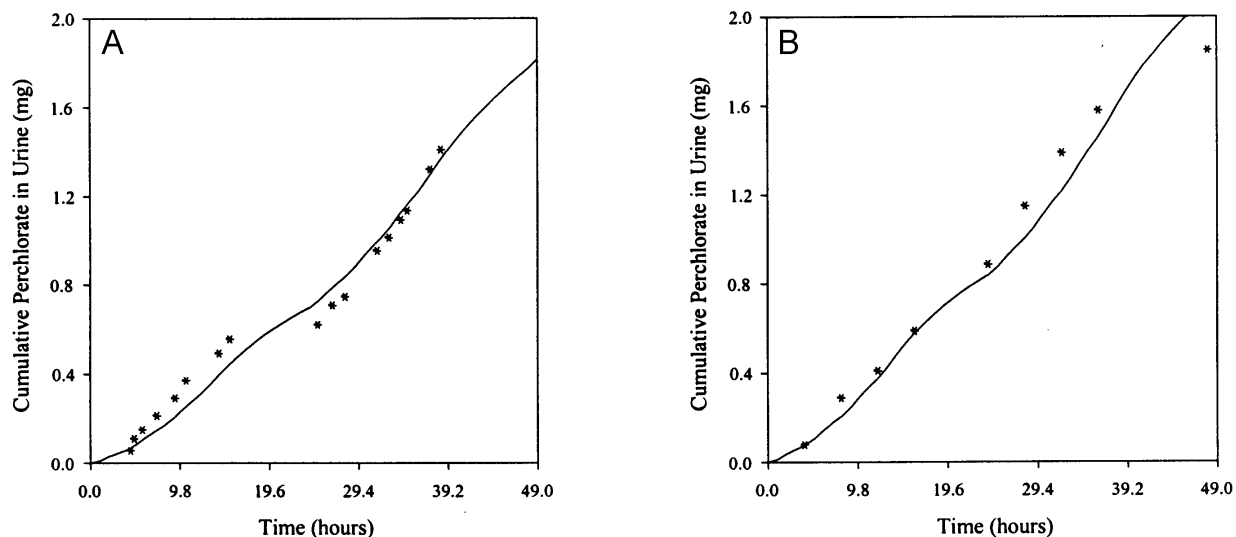
Figure 6-19. Human PBPK model predictions (lines) versus data of 48-hour cumulative urine perchlorate (mg) shown for two different subjects. Subject consumed 0.02 mg/kg-day perchlorate in drinking water, 4 times per day, for 14 days. Model predictions for the individual obtained by using study average value of all subjects for urinary clearance of perchlorate (ClUc_p). Model of Merrill (2001d) and data of Greer et al. (2000).

1   Due to its small size, variations in the thyroid parameters have little effect on serum

2   concentrations of both iodide and perchlorate. As described for Figure 6-14, Merrill (2001d)

3   estimated parameters for iodide disposition, including those of the thyroid, from fits to the data

4   of Hays and Solomon (1965). Using these same iodide parameters, baseline thyroid RAIU

5   measurements performed by Greer et al. (2000) were fit with the model by adjusting the Vmaxc

6   for the thyroid follicular epithelium (Vmaxc_Ti). Figures 6-20, 6-21, 6-22, and 6-23 illustrate

7   the model predictions of thyroid RAIU versus data for subjects in the 0.5, 0.1, 0.02, and

8   0.007 mg/kg-day dosage groups, using either the individual's Vmaxc_Ti (left) or an average

9   value (right). The average Vmaxc_Ti (1.5×10^5 ng/hr-kg) was obtained from fitting baseline
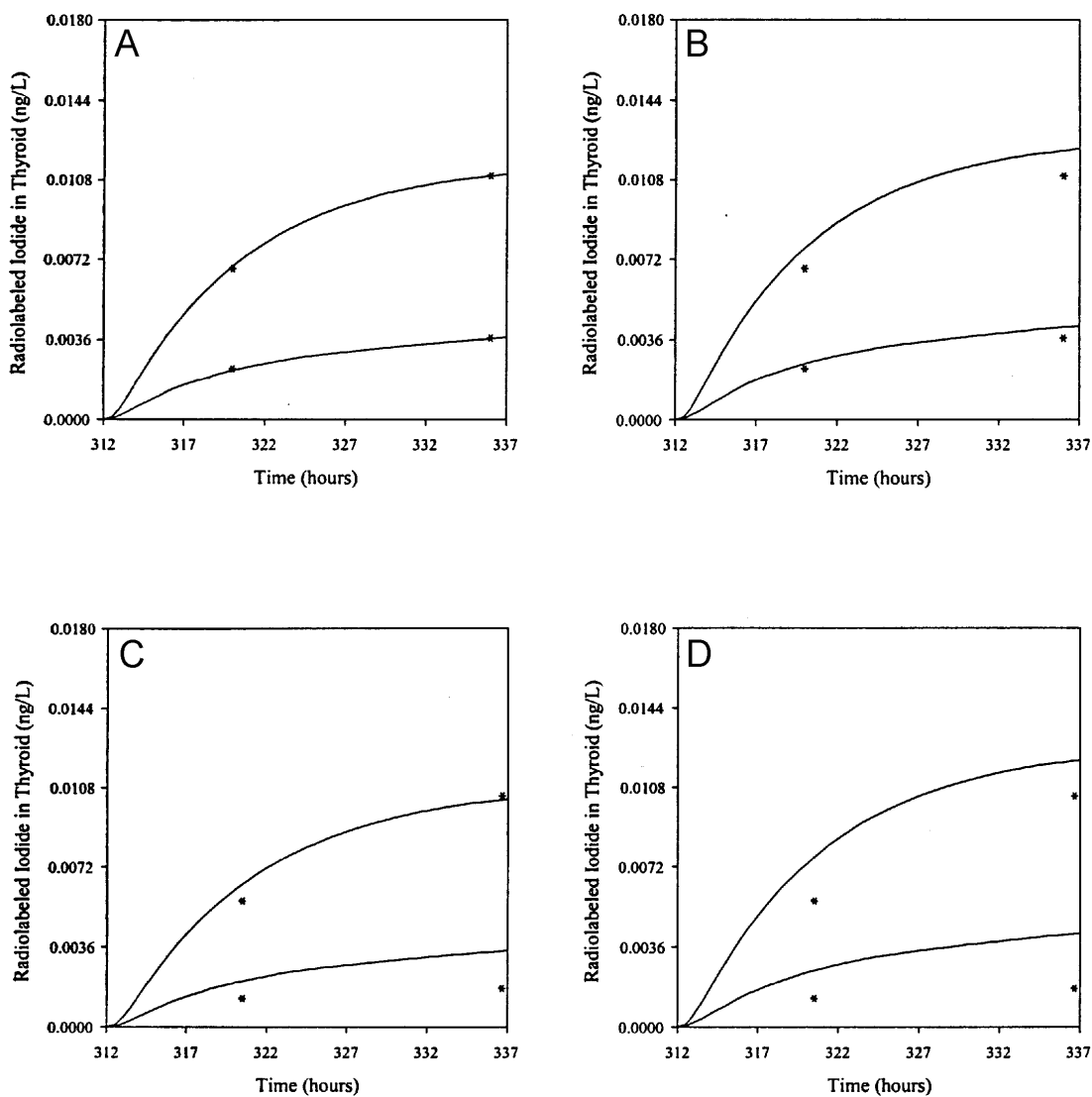
Figure 6-20. **Human PBPK model predictions (lines) versus data (asterisks) for thyroid RAIU (ng/L) on day 14 of perchlorate exposure at 0.5 mg/kg-day for a healthy female (top panel) and male (bottom panel). Prediction on left for female (A) obtained by using individually fitted maximum capacity (ng/hr-kg) for active transport of iodide into the thyroid follicular epithelium (Vmaxc_Ti) of 1.3×10^5 and on right (B) by using an average Vmaxc_Ti. Prediction on left for male (C) obtained by using individually fitted Vmaxc_Ti of 1.24×10^5 and on right (D) by using an average Vmaxc_Ti of 1.5×10^5. Model of Merrill (2001d) and data of Greer et al. (2000).**
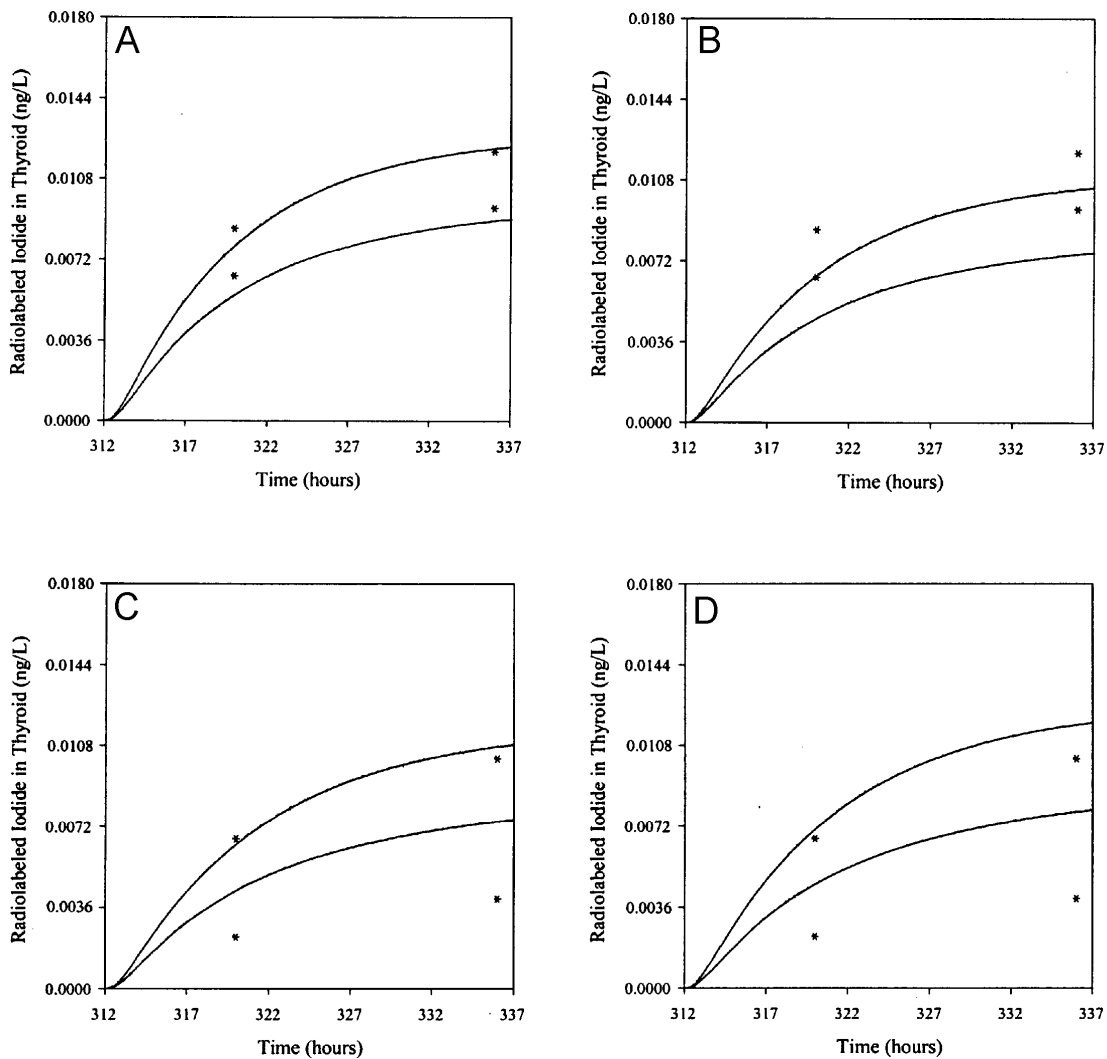
Figure 6-21. Human PBPK model predictions (lines) versus data (asterisks) for thyroid RAIU (ng/L) on day 14 of perchlorate exposure at 0.1 mg/kg-day for a healthy female (top panel) and male (bottom panel). Prediction on left for female (A) obtained by using individually fitted maximum capacity (ng/hr-kg) for active transport of iodide into the thyroid follicular epithelium (Vmaxc_Ti) of 1.65×10^5 and on right (B) by using an average Vmaxc_Ti. Prediction on left for male (C) obtained by using individually fitted Vmaxc_Ti of 1.2×10^5 and on right (D) by using an average Vmaxc_Ti of 1.5×10^5. Model of Merrill (2001d) and data of Greer et al. (2000).
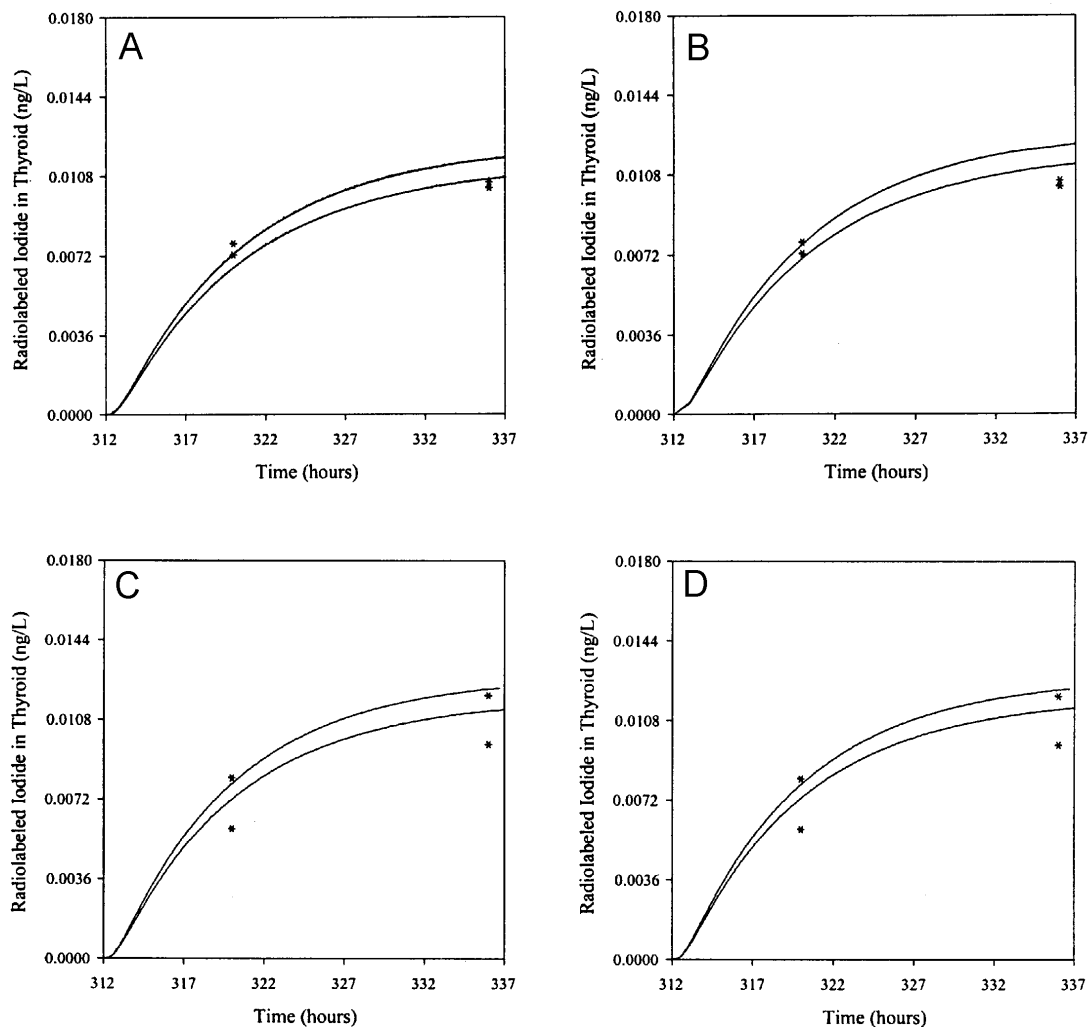
Figure 6-22. **Human PBPK model predictions (lines) versus data (asterisks) for thyroid RAIU (ng/L) on day 14 of perchlorate exposure at 0.02 mg/kg-day for a healthy female (top panel) and male (bottom panel). Prediction on left for female (A) obtained by using individually fitted maximum capacity (ng/hr-kg) for active transport of iodide into the thyroid follicular epithelium (Vmaxc_Ti) of 1.4×10^5 and on right (B) by using an average Vmaxc_Ti. Prediction on left for male (C) obtained by using individually fitted Vmaxc_Ti of 1.5×10^5 and on right (D) by using an average Vmaxc_Ti of 1.5×10^5. Model of Merrill (2001d) and data of Greer et al. (2000).**
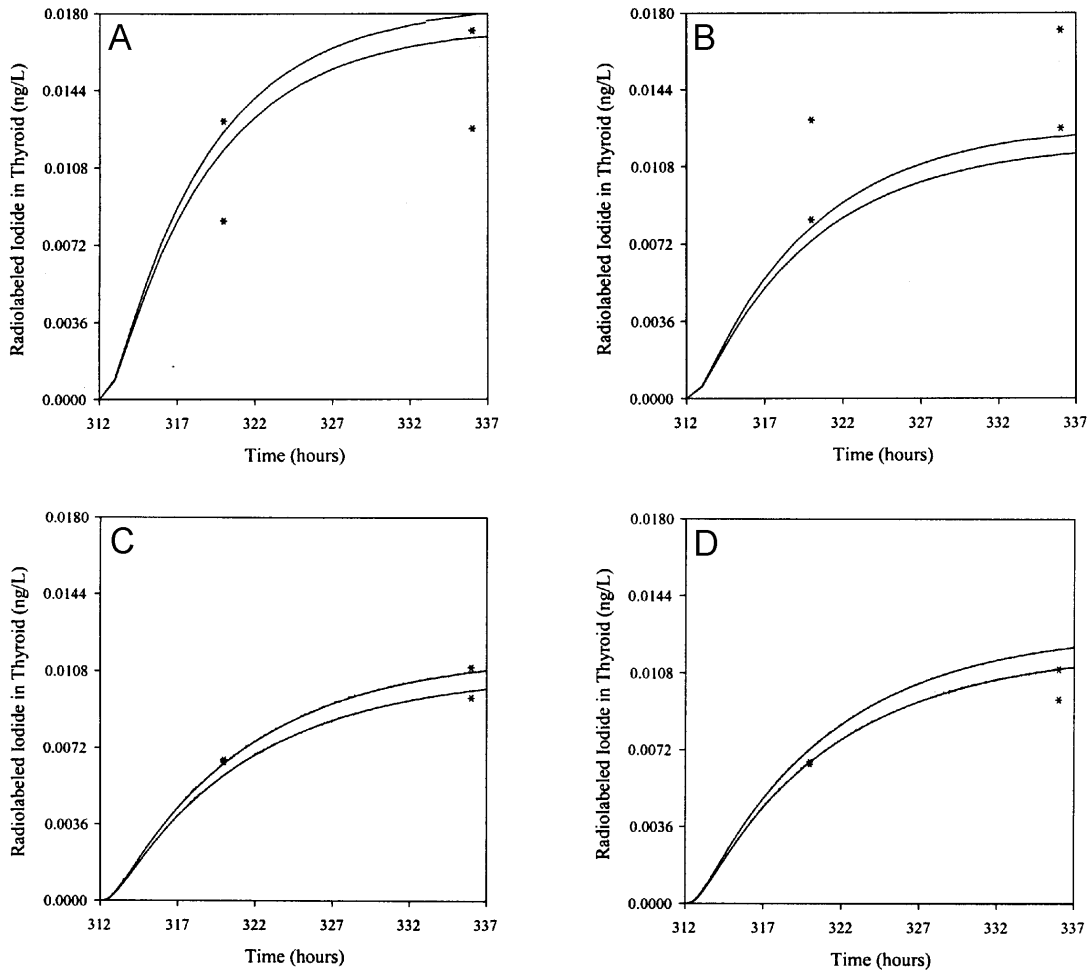
Figure 6-23. **Human PBPK model predictions (lines) versus data (asterisks) for thyroid RAIU (ng/L) on day 14 of perchlorate exposure at 0.007 mg/kg-day for a healthy female (top panel) and male (bottom panel). Prediction on left for female (A) obtained by using individually fitted maximum capacity (ng/hr-kg) for active transport of iodide into the thyroid follicular epithelium (Vmaxc_Ti) of 2.8×10^5 and on right (B) by using an average Vmaxc_Ti. Prediction on left for male (C) obtained by using individually fitted Vmaxc_Ti of 1.24×10^5 and on right (D) by using an average Vmaxc_Ti of 1.35×10^5. Model of Merrill (2001d) and data of Greer et al. (2000).**

1 radioiodide uptake measurements provided by Greer et al. (2000) across doses (see Merrill,

2 2001d; Table 3). Merrill (2001d) hypothesized that the large variability in Vmaxc_Ti, ranging

3 from 5.0×10^4 to 5.0×10^5 ng/hr-kg, may be attributed to variability in endogenous iodide levels,

4 as dietary iodide was not controlled. Merrill (2001d) estimated these values from best visual fits

5 of baseline 8- and 24-hour thyroid RAIU data. Inhibition data restricted to each time point (i.e.,

6 8- versus 24-hour time points) and from higher dose groups would be useful to test the

7 robustness of the model to predict inhibition of uptake of iodide in the thyroid.

8   The ability of the human model to predict data from other independent experiments not

9 used to develop the model is illustrated in Figure 6-24. The model adequately predicts

10 cumulative perchlorate in urine (mg) reported in three published studies using therapeutic

11 perchlorate dose levels (Merrill, 2001d). Oral doses administered in these studies were

12 approximately 9.07 mg/kg (Durand, 1938), 9.56 mg/kg (Kamm and Drescher, 1973), and

13 20 mg/kg (Eichler, 1929). It is worth noting that the previously determined urinary clearance

14 value (ClUc_p) of 0.126 L/hr-kg was used with all validation data and that an adequate fit was

15 observed.

16   The ability of the model to predict cumulative perchlorate in urine from three different

17 studies at three different doses with the same set of parameters, established from the studies by

18 Hays and Solomon (1965) and Greer et al. (2000), demonstrates the usefulness of the model and

19 provides validation for the model structure and the physiological and chemical parameters used.

20   The model also predicts serum perchlorate concentrations at 12 mg/kg-day from an

21 unpublished study performed by Dr. Georg Brabant at the Medizinische Hochschule, Hanover,

22 Germany (Figure 6-25). Subjects received 12 mg/kg-day perchlorate in drinking water near meal

23 times. Variability in the observed serum measurements is believed to reflect variability in the

24 dosing regimen, as the experimental protocol was less fixed than that used in Greer et al. (2000).

25 Again the usefulness of the model is demonstrated by its ability to successfully predict serum

26 concentrations from a dose 24 times higher than the high dose used to establish perchlorate

27 parameters (0.5 mg/kg-day).

28   The model is also able to successfully predict the thyroidal iodide uptake in a subject from

29 the Stanbury and Wyngaarden (1952) study with patients with Grave's disease. The maximum

30 velocity capacity in the follicular epithelium (Vmaxc_Ti) had to be increased to 5.0E6 ng/hr-kg,

31 a factor of ten times higher than in normal subjects, in order to achieve this fit (upper line in
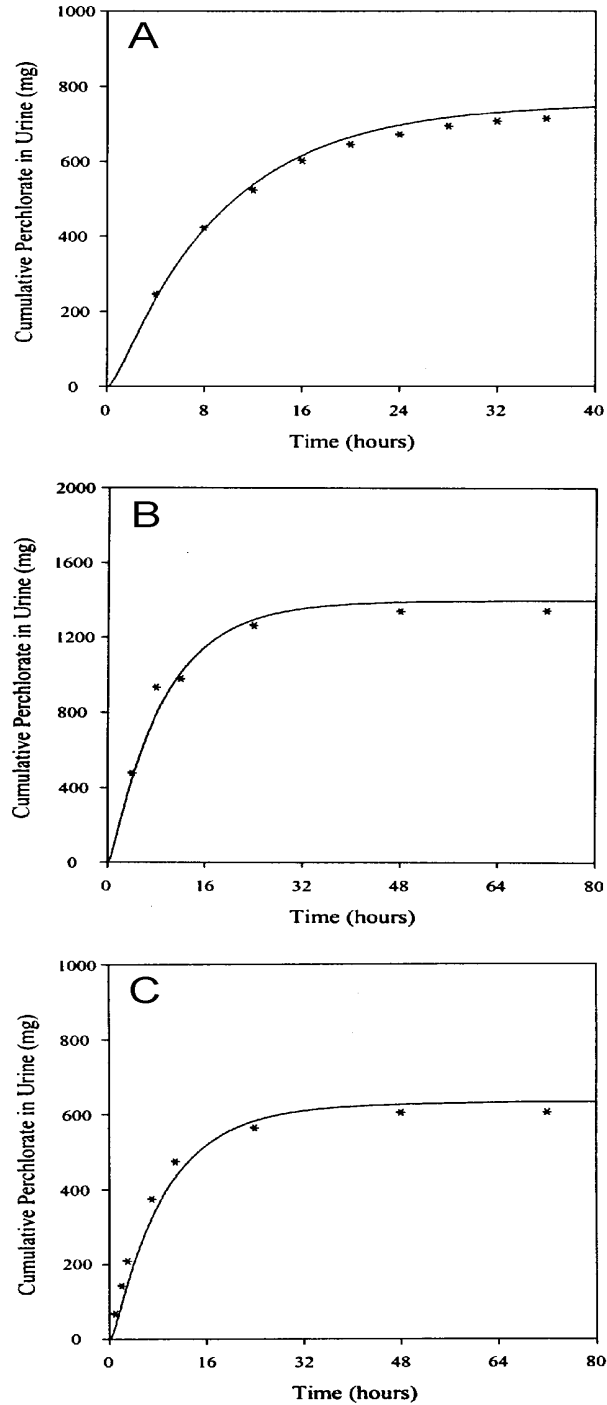
Figure 6-24. Validation for human PBPK model (Merrill, 2001d). Model predictions (lines) versus data (asterisks) for cumulative perchlorate excretion in urine (mg) in a healthy male after an oral dose of 9.56 mg (A), 20 mg (B) or 9.07 mg (C). Data are from three different studies. Data of Kamm and Drescher (1973) for (A), Eichler (1929) for (B) and Durand (1938) for (C).
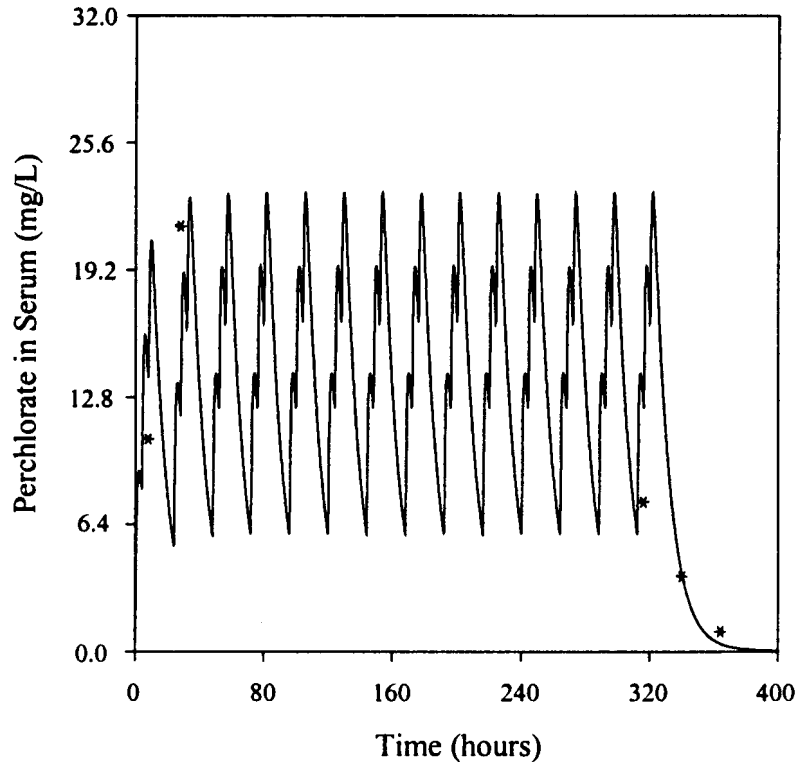
Figure 6-25. Validation for human PBPK model (Merrill, 2001d). Model predictions (lines) versus data (asterisks) for serum perchlorate concentrations (mg/L) in 5 subjects received 12 mg/kg-day in drinking water (data of Brabant and Letiolf, 2000 as cited in Merrill, 2001d). Subjects were instructed to ingest the solution 3 times/day for 14 days. Serum samples were collected 2 hours after the first dose, after 12 pm on day two, the morning of day 14 and post exposure days 1 and 2. Usefulness of the model is demonstrated by its ability to successfully predict serum concentrations at a dose 24 times higher than the dose used to develop parameters in the model.

1   Figure 6-26). This increase in Vmaxc_Ti is supported in the literature, as Gluzman and

2   Niepomniszcze (1983) measured elevated Vmaxc(s) in thyroid specimens from subjects with

3   Grave's disease. However, the model underpredicts the degree of inhibition caused by

4   perchlorate in this subject (Figure 6-26, lower line). It would appear that the increased inhibition

5   could be attributed to a lower Km value. However, Gluzman and Niepomniszcze (1983) noted

6   that the Km did not differ greatly between thyroid specimens from hyperthyroid subjects and
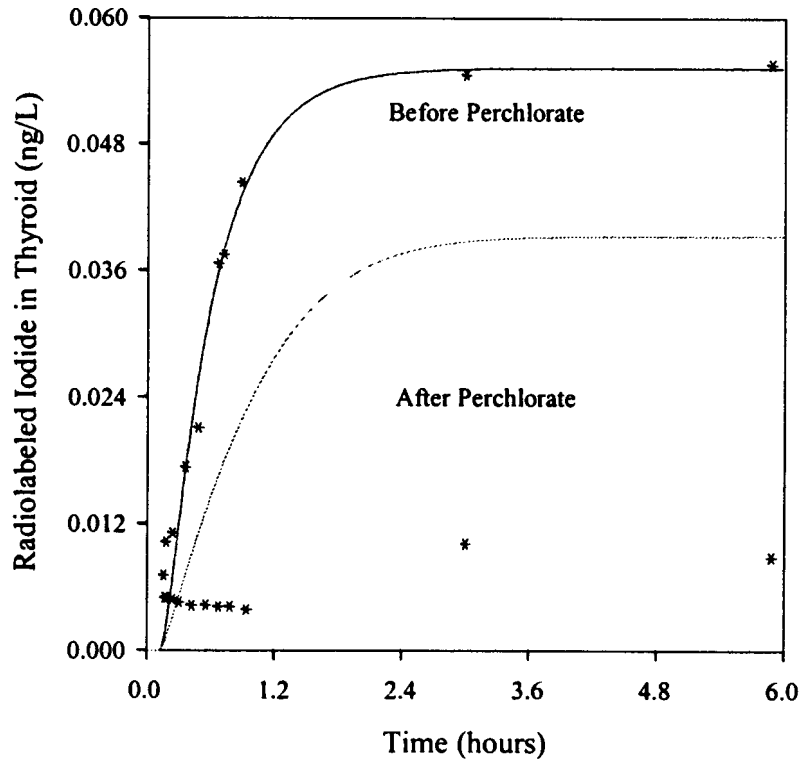
Figure 6-26. Validation for human PBPK model (Merrill, 2001d). Model predictions (lines) versus data (asterisks) for RAIU in the thyroid ($^{131}I^-$ ng/L) of a male with Graves' disease after an iv dose of 10 μCi $^{131}I^-$ before and after a 100 mg dose of potassium perchlorate. Data of Stanbury and Wyngaarden (1952).

1   normal subject. This suggests that the increased inhibition by perchlorate seen in Grave's disease
2   may be attributed to a mechanism other than NIS affinity (Merrill, 2001d).
3
4   **6.2.4 Summary**
5   The proposed model structures for the adult male rat (Merrill, 2001c) and adult human
6   (Merrill, 2001d) have been shown to adequately describe both perchlorate and iodide disposition
7   by demonstrating good correspondence between predicted tissue compartment concentrations
8   and measured values in the thyroid, serum, red blood cells, urine, liver, muscle, skin, and
9   stomach in the rat and by adequately predicting serum concentrations and cumulative urine after
10 drinking water exposure to perchlorate spanning four orders of magnitude (0.02 to 12.0 mg/kg-
11 day) in the human. Serum perchlorate levels for human subjects were not available at

1   0.02 mg/kg-day; however, the model did predict the cumulative urine from that dose group

2   (Figure 6-19).

3   The model structure of the thyroid requires three compartments (stroma, follicle, and

4   colloid) to quantify rapid organification in the gland. Differences in model parameters between

5   iodide and perchlorate indicate that iodide kinetics are very similar to perchlorate kinetics, but

6   cannot be applied directly. The main differences involve the saturable kinetics of the thyroid,

7   skin, and stomach, with perchlorate exhibiting higher Vmaxc's except in the skin. Because

8   organification of iodide occurs in both the thyroid follicle and colloid, their respective Vmaxc's

9   are over 1,000 and 10 times higher than those for perchlorate, which is discharged unchanged.

10   Perchlorate affinity for the symporters into the follicle and colloid were approximately an order

11   of magnitude greater (lower Km) than those of iodide.

12   The thyroid perchlorate concentrations from high drinking water exposures in the rat were

13   fitted by increasing the effective follicle:stroma partition coefficient (PT_p) to account for TSH

14   stimulation and upregulation of NIS. Since these values were not supported by additional data,

15   thyroid concentrations may not be as reliable. Further, the toxic effects of perchlorate are most

16   likely due to secondary effects on thyroid hormones due to its action at the NIS.

17   The model, however, could simulate serum concentrations from drinking water exposures

18   using parameters established from the acute data. The thyroid, given it's small size, would not be

19   expected to significantly alter serum concentrations, even during hyperstimulation. Although

20   TSH has not been shown to increase the NIS in other tissues, NIS-containing tissues were not

21   obtained from the AFRL/HEST studies to support this.

22   The models support plasma protein binding of perchlorate in both species; a saturable term

23   is required to simulate serum concentrations at lower doses. It is possible that perchlorate

24   competes with thyroxine for the same binding sites of plasma proteins, as the work of Yamada

25   and Jones (1968) suggests. Urinary clearance values of 0.05 L/hr for iodide and 0.07 L/hr for

26   perchlorate were used across data sets in the rats, and average urinary clearance values were

27   found to be 0.1 L/hr-kg for iodide and 0.126 L/hr-kg in humans. Excretion constants were

28   highest among the 0.1 mg/kg-day group. With the urinary excretion rates fitted to cumulative

29   urine data, the model tends to slightly underestimate serum perchlorate levels at repeated low

30   doses. Elevated serum concentrations may indicate plasma binding of perchlorate. Yamada and

31   Jones (1967) studied effects of different anions on plasma binding to thyroxine and noted that

some of the thyroxine had been displaced after perchlorate was introduced. Thus, it is possible that perchlorate competes with thyroxine for the same binding sites of plasma proteins (Merrill, 2001c,d; Clewell, 2001a).

While there are limited data suggesting iodide and perchlorate uptake through the skin, the models and the kinetic studies required this assumption in the models for both rats and humans. Without the skin compartment, the models overestimated circulating plasma inorganic iodide and perchlorate in both species. Due to its large size, skin appears to be an important pool for slow turnover of these anions. Brown-Grant (1961) noted that the uptake of iodide was higher in the male rat and pup than in the female. The findings of Merrill (2001c) agree, with the rat model requiring a higher Vmaxc in the skin for the male rat than that reported for the pregnant rat (Clewell, 2001a) discussed in the next section. Cutaneous uptake of iodide and perchlorate in mice and rats has been reported (Brown-Grant and Pethes, 1959; Zeghal et al., 1995). The lack of reported iodide in human skin from clinical radioiodide scans may be due to the difficulty in differentiating it from background radioactivity.

Merrill (2001d) notes that GI clearance of iodide is rapid and plays an important role in radioiodide conservation. Further, Merrill (2001d) suggests that the appearance of time-course radioiodine in stomach contents of any species is complicated by the fact that it reflects more than sequestration of radioiodide by NIS. Its appearance also reflects radioiodide contributed through the gradual accumulation of iodide in saliva that is swallowed involuntarily throughout the study. Several studies that examined sequestration of these anions in digestive juices have all shown high variability in the concentrations measured over time (Honour et al., 1952; Hays and Solomon, 1965; Merrill, 2001d). There is a tendency for the gastric juice to plasma ratio to be low when the rate of secretion of juice is high (Honour et al., 1952). Fluctuations in the secretion rate are probably the most important factor in determining the pattern of the concentration ratios in individuals. Therefore, variability in stomach or GI tract parameters between models is expected. However, the early rise in the gastric juice:plasma ratio mentioned earlier is a constant feature across these data sets, whether or not an attempt was made to eliminate contamination of gastric juices by dietary contents or saliva. The human model successfully predicted this same trend.

Merrill (2001d) also noted dietary iodine and endogenous inorganic iodide levels to be clearly important in modeling iodide and perchlorate kinetics, because excessive iodide levels

1	cause the ion to inhibit its own uptake. Plasma inorganic iodide (PII) is rarely reported in the
2	literature due to analytical difficulties, and it was not available in any of the studies presented in
3	this paper. While measurements of tracer radioiodide can be fitted to predict transfer rates, its
4	use is limited when attempting to predict the saturation of nonlinear compartments, such as the
5	thyroid that are dependent upon the existing amount of iodide already present. Subsequent
6	modeling efforts on predicting subsequent effects of iodide inhibition on thyroid hormone
7	synthesis and regulation in humans will require the capability of the model to predict PII.
8	
9	

6.3 PREGNANT RAT AND FETAL MODEL STRUCTURE

11   This section describes the model developed by AFRL/HEST in response to concerns about

12   interspecies extrapolation of effects due to perchlorate exposure during gestation (Clewell,

13   2001a). The model predicts the distribution of perchlorate within the pregnant and fetal rat

14   through gestation and at birth and predicts the short-term effect of acute perchlorate exposure on

15   iodide kinetics, including iodide uptake into the maternal thyroid. The general model structure

16   relied on the adult male rat model (Merrill, 2001c) described in Section 6.2 and approaches to

17   gestational growth of the dam and fetus were based on the work of O'Flaherty et al. (1992) and

18   Fisher et al. (1989) with weak acids.

19   The model structure is shown in Figure 6-27. Table 6-3 provides the physiological

20   parameters for the pregnant rat and fetus PBPK models. Table 6-4 provides the perchlorate-

21   specific parameters, and Table 6-5 provides the iodide-specific parameters for each.

22   The compartments shared with the adult male rat were developed as described in

23   Section 6.2. The pregnant rat model also includes a mammary gland and placenta compartment.

24   The mammary gland consists of two subcompartments that represent the capillary bed and the

25   tissue. The mammary gland has been shown to concentrate both perchlorate and iodide during

26   lactation. However, the mammary NIS is regulated by hormones produced during lactation and

27   has been found to increase at the onset of lactation (Tazebay et al., 2000). This concentrating

28   mechanism does not appear to be as established during pregnancy. Studies reported by Yu

29   (2000) showed mammary gland:plasma ratios of less than one for perchlorate. However,

30   mammary gland perchlorate levels are slowly built up and remain high well into the clearance

31   phase of the serum. This behavior suggested a very slow diffusion between the mammary gland
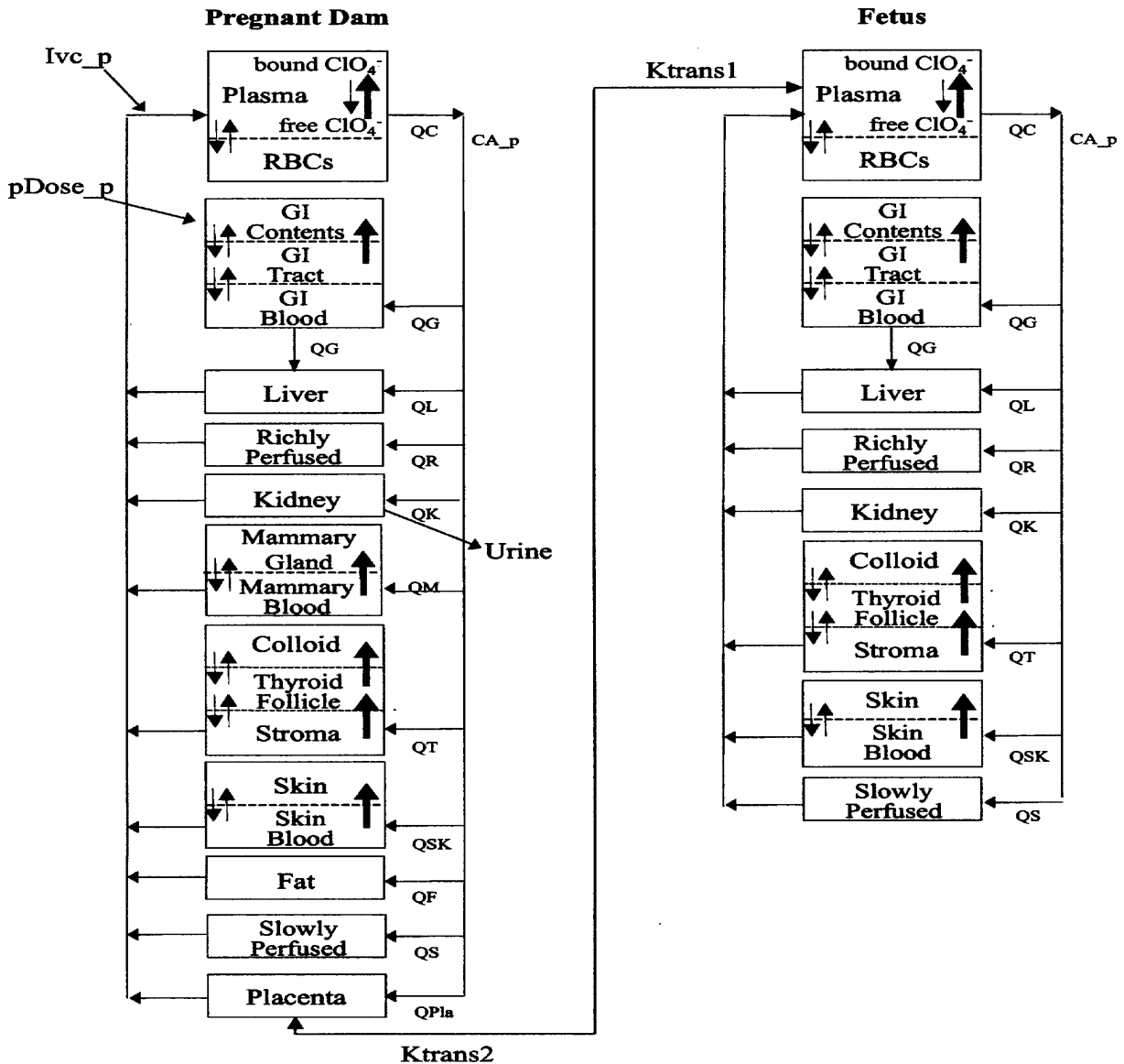
Figure 6-27. Schematic for the pregnant dam and fetal rat PBPK model of perchlorate and iodide distribution (Clewell, 2001a). Bold arrows indicate (except for plasma binding) active uptake at NIS sites into the thyroid, GI contents, and skin. Plasma binding was also described with Michaelis-Menten terms for the association of perchlorate anion to binding sites with first-order clearance rates for dissociation. Small arrows indicate passive diffusion. Boxes represent specific compartments in the model structure. The thyroid consists of the stroma, the follicle, and the colloid; and the stomach consists of the capillary bed, GI wall, and contents. The skin and mammary gland each contain two subcompartments representing the capillary bed and tissue. Permeability area cross products and partition coefficients were used to describe the first-order movement of the perchlorate (ClO_4^-) and iodide (I^-) anions into deeper subcompartments. Placental-fetal transfer and urinary clearance were represented by first order clearance rates.

TABLE 6-3. PHYSIOLOGICAL PARAMETERS FOR THE PREGNANT RAT AND FETUS PBPK MODEL (Clewell, 2001a)

Physiological Parameters	Pregnancy		
Tissue Volumes (%BW)	**Dam**	**Fetus**	**Source**
Body Weight *BW* and *V1fet* (kg)	0.280 - 0.361	0.0 - .0045	O'Flaherty et al., 1992
Slowly Perfused VSc (%BW)	74.6	74.6	Brown et al., 1997
Richly Perfused VRc (%BW)	11	11	Brown et al., 1997
Fat *VFc* (%BW)	10.0 - 11.0	0.0	Naismith et al., 1982
Kidney *VKc* (%BW)	1.7	1.7	Brown et al., 1997
Liver *VLc* (%BW)	3.4	3.4	Brown et al., 1997
GI Tract *VGc* (%BW)	3.60	3.60	Brown et al., 1997
GI Contents *VGJc* (%BW)	7.20	7.20	Yu et al., 2000
GI Blood *VGBc* (%VG)	2.9	2.9	Altman and Dittmer, 1971
Skin Tissue *VSkc* (%BW)	19.0	19.0	Brown et al., 1997
Skin Blood *VSkBc* (%VSk)	2.0	2.0	Brown et al., 1997
Thyroid Total *VTtotc* (%BW)	0.0105	0.0234	Malendowicz and Bednarek, 1986; Florsheim et al., 1966
Thyroid Follicle *VTc* (%BW)	45.9	61.4	Malendowicz and Bednarek, 1986; Conde et al., 1991
Thyroid Colloid *VDTc* (%BW)	45	18.3	Malendowicz and Bednarek, 1986; Conde et al., 1991
Thyroid Blood *VTBc* (%VT)	9.1	20.3	Malendowicz and Bednarek, 1986; Conde et al., 1991
Plasma *VPlasc* (%BW)	4.7	4.7	Brown et al., 1997; Altman and Dittmer, 1971
Red Blood Cells *VRBCc* (%BW)	2.74	2.74	Brown et al., 1997; Altman and Dittmer, 1971
Placenta *VPl ac* (%BW)	0.0 - 2.57	—	O'Flaherty et al., 1992
Mammary Tissue *VMc* (%BW)	1.0 - 5.5	—	Knight et al., 1984; O'Flaherty et al., 1992
Blood Flows (%QC)			
Cardiac Output *QCc* (L/hr-kg)	14	14.0	Buelke-Sam, 1982a & b; O'Flaherty et al., 1992
Slowly Perfused *QSc* (%QC)	24.0	24.0	Brown et al., 1997
Richly Perfused *QRc* (%QC)	76.0	76.0	Brown et al., 1997
Fat *QFc* (%QC)	7 - 8.1	0.0	Brown et al., 1997
Kidney *QKc* (%QC)	14.0	14.0	Brown et al., 1997
Liver *QLc* (%QC)	18.0	18.0	Brown et al., 1997
GI *QGc* (%QC)	13.60	13.60	Brown et al., 1997
Thyroid *QTc* (%QC)	1.6	1.6	Brown et al., 1997
Mammary *QMc* (%QC)	0.2 - 1.2	—	Hanwell and Linzell, 1973
Placenta *QPlc* (%QC)	0.0 - 12.3	—	O'Flaherty et al., 1992

TABLE 6-4. PERCHLORATE-SPECIFIC PARAMETERS FOR THE PREGNANT RAT AND FETUS PBPK MODEL (Clewell, 2001a)[a]

Pregnancy Parameters	Perchlorate Values		
Partition Coefficients (unitless)	Dam	Fetus	Source
Slowly Perfused/Plasma PS_	0.31	0.31	Yu et al., 2000
Rapidly Perfused/Plasma PR_	0.56	0.56	Yu et al., 2000
Fat/Plasma PF_	0.05	—	Pena et al., 1976
Kidney/Plasma PK_	0.99	0.99	Yu et al., 2000
Liver/Plasma PL_	0.56	0.56	Yu et al., 2000
Gastric Tissue/Gastric Blood PG_	0.50	1.80	Yu et al., 2000
GI Contents/GI Tissue PGJ_	1.30	2.30	Yu, 2000
Skin Tissue/Skin Blood PSk_	1.15	1.15	Yu, 2000
Thyroid Tissue/Thyroid Blood PT_	0.13 / 2.25	0.13 / 2.25	Chow and Woodbury, 1970[b]
Thyroid Lumen/Thyroid Tissue PDT_	7.00	7.00	Chow and Woodbury, 1970
Red Blood Cells/Plasma	0.73	0.73	Yu et al., 2000
Placenta/ Plasma PPL_	0.56	—	Assume same as richly perfused
Mammary/Plasma PMam_p	0.66	—	Anbar et al., 1959
Max Capacity, Vmaxc (ng/hr-kg)			
Thyroid Follicle Vmaxc_T	1.80E+03	1.80E+03	Fitted[c]
Thyroid Colloid Vmaxc_DT	1.00E+04	1.00E+04	Fitted[c]
Skin Vmaxc_S	6.00E+05	4.00E+05	Fitted
Gut Vmaxc_G	8.00E+05	8.00E+05	Fitted
Mammary Gland Vmaxc_M	3.90E+04	---	Molar equivalent to Vmaxc_M*i*
Plasma Binding Vmaxc_Bp	5.00E+03	1.50E+03	Fitted
Affinity Constants, Km (mg/L)			
Thyroid Follicle Km_T	1.00E+05	1.00E+05	Wolff, 1998
Thyroid Colloid Km_DT	1.00E+08	1.00E+08	Golstein et al., 1992; Wolff, 1998
Skin Km_S	1.00E+05	1.00E+05	Wolff, 1998
Gut Km_G	1.00E+05	1.00E+05	Wolff, 1998
Mammary Gland	1.00E+5	—	Wolff, 1998
Plasma Binding Km_Bp	1.00E+05	1.00E+05	Fitted
Permeability Area Cross Products, (L/hr-kg)			
GI Blood to GI Tissue PAGc_	1.00	1.00	Fitted
GI Tissue to GI Contents PAGJc_	1.00	1.00	Fitted
Thyroid Blood to Thyroid Tissue PATc_	4.0E-5 / 6.0E-4	4.0E-5 / 6.0E-4	Fitted[b]
Thyroid Tissue to Thyroid Lumen PADTc_	0.01	0.01	Fitted
Skin Blood to Skin Tissue PASkc_	1.00	1.00	Fitted
Plasma to Red Blood Cells PRBCc_	1.00	1.00	Fitted
Clearance Values, (L/hr-kg)			
Urinary Excretion CLUc_	0.07	—	Yu et al., 2000
Transfer from Placenta to Fetus Cltrans1c_	0.10	0.10	Yu, 2000
Transfer from Fetus to Placenta Cltrans2c_	0.19	0.19	Yu, 2000
Dissociation from Plasma Binding Sites Clunbc_p	0.034	0.010	Yu, 2000

[a]All parameters listed are notated in the model by either an *i* (for iodide) or *p* (for perchlorate) following an underscore in the parameter name (e.g., PR_*i*, PR_*p*, Vmaxc_T*i*, etc.)
[b]Parameters with two values indicate acute and drinking water parameters, respectively.
[c]Fetus was given maternal values for Vmax (scaled by fetal body weight) in the absence of data.

TABLE 6-5. IODIDE-SPECIFIC PARAMETERS FOR THE PREGNANT RAT AND FETUS PBPK MODEL (Clewell, 2001a)[a]

Pregnancy Parameters	Iodide Values		
Partition Coefficients (unitless)	**Dam**	**Fetus**	**Iodide Source**
Slowly Perfused/Plasma PS_	0.21	0.21	Halmi et al., 1956
Rapidly Perfused/Plasma PR_	0.40	0.40	Halmi et al., 1956
Fat/Plasma PF_	0.05	—	Pena et al., 1976
Kidney/Plasma PK_	1.09	1.09	Yu et al., 2000
Liver/Plasma PL_	0.44	0.44	Yu et al., 2000
GI Tissue/GI Blood PG_	1.0	1.0	Yu, 2000
GI Contents/GI Tissue PGJ_	2.0	2.0	Yu, 2000
Skin Tissue/Skin Blood PSk_	0.70	0.70	Perlman et al., 1941
Thyroid Tissue/Thyroid Blood PT_	0.15	0.15	Chow and Woodbury, 1970
Thyroid Lumen/Thyroid Tissue PDT_	7.00	7.00	Chow and Woodbury, 1970
Red Blood Cells/Plasma	1.00	1.00	Yu et al., 2000
Placenta/Plasma PPL_	0.99	—	Unpublished GD20 data
Mammary/Plasma PMam_p	0.66	—	Anbar et al., 1959 (for ClO_4^-)
Max Capacity, Vmaxc (ng/hr-kg)			
Thyroid Follicle Vmaxc_T	4.00E+04	0.0 – 7.5E+04	Fitted
Thyroid Colloid Vmaxc_DT	6.00E+07	6.00E+07	Fitted
Skin Vmaxc_S	6.00E+04	3.00E+05	Fitted
Gut Vmaxc_G	1.00E+06	2.00E+05	Fitted
Mammary Gland Vmaxc_M	5.00E+04	—	Fitted
Affinity Constants, Km (mg/L)			
Thyroid Follicle Km_T	4.00E+06	4.00E+06	Gluzman and Niepomniszcze, 1983
Thyroid Colloid Km_DT	1.00E+09	1.00E+09	Golstein et al., 1992
Skin Km_S	4.00E+06	4.00E+06	Gluzman and Niepomniszcze, 1983
Gut Km_G	4.00E+06	4.00E+06	Gluzman and Niepomniszcze, 1983
Mammary Gland Km_M	4.00E+06	—	Gluzman and Niepomniszcze, 1983
Permeability Area Cross Products, (L/hr-kg)			
GI Blood to GI Tissue PAGc_	0.80	0.10	Fitted
GI Tissue to GI Contents PAGJc_	0.60	0.30	Fitted
Thyroid Blood to Thyroid Tissue PATc_	1.000E-04	1.000E-04	Fitted
Thyroid Tissue to Thyroid Lumen PADTc_	1.00E-04	1.00E -04	Fitted
Skin Blood to Skin Tissue PASkc_	0.10	0.02	Fitted
Plasma to Red Blood Cells PRBCc_	1.00	1.00	Fitted
Clearance Values, (L/hr-kg)			
Urinary excretion CLUc_	0.03	—	Fitted
Transfer from Placenta to Fetus Cltrans1c_	0.06	0.06	Unpublished GD 20 Iodide iv Data
Transfer from Fetus to Placenta Cltrans2c_	0.12	0.12	Unpublished GD 20 Iodide iv Data

[a]All parameters listed are notated in the model by either an *i* (for iodide) or *p* (for perchlorate) following an underscore in the parameter name (e.g., PR_*i*, PR_*p*, Vmaxc_T*i*, etc.)

1   and blood, so that Clewell (2001a) described the two-subpartment mammary gland with both

2   diffusion of iodide and active uptake by the NIS.

3   Although it has been suggested that the placenta may contain the capability for active

4   uptake in the rat, AFRL/HEST data did not indicate placenta:plasma levels greater than one for

5   perchlorate or iodide (Yu, 2000), and unpublished iodide time course data indicate that the

6   behavior of iodide in the placenta closely mirrors that of the plasma (Clewell, 2001a). Thus, the

7   placenta was simulated with a single, flow-limited compartment.

8   Partitioning into the mammary, placenta, and other diffusion-limited compartments was

9   based on effective partitioning. This effective partitioning is probably very similar to that in the

10 thyroid where an electrochemical gradient is responsible for allowing the ClO_4^- anion to move

11 between the serum and the tissue (Chow and Woodbury, 1970). Urinary clearance and placental-

12 fetal transfer of the anions were represented by first order clearance rates.

13   The structure of the fetal perchlorate model is similar to that of the pregnant rat, with the

14 exception of the mammary gland and placenta compartments. In order to simplify the model, all

15 of the fetuses from a single litter were combined in the structure of the model, essentially

16 viewing the individual fetuses as one entity, or one large fetus. The dose to the fetus is based on

17 the transfer of perchlorate from the maternal placenta to the serum of the fetus, rather than

18 through direct exposure to the drinking water. Though a kidney is included in the fetal model,

19 urinary excretion is not used to identify the loss of perchlorate for the fetus. Since the ability to

20 produce urine is not well developed until after parturition, the loss from the fetus is described as

21 first order clearance from the fetal arterial blood to the placenta (Clewell, 2001a).

22   The pregnancy model attempts to describe perchlorate distribution in a highly dynamic

23 system. In addition to total body weight changes in the dam and fetus, maternal mammary tissue

24 and blood flow, cardiac output, fractional body fat, placenta and fetus body weight, and fractional

25 body fat are also changing with respect to time. Growth equations, based on O'Flaherty et al.

26 (1992) were used to account for these changes (Clewell, 2001a). All tissue volume and blood

27 flow values were adjusted with respect to the changing parameters.

28

29 ## 6.3.1 Data and Methods

30   This section summarizes the data that Clewell (2001a) used for development and validation

31 of the pregnant and fetal rat model structures. Details on experimental methods, including:

1   protocol design, exposure regimen, chemical source and purity, animals (housing, feeding,

2   surgical procedures, etc.), and the analytical methods can be found in the consultative letter and

3   associated reports from AFRL/HEST or cited papers therein.

4

5   **6.3.1.1 AFRL/HEST Experiments in Laboratory Rats**

6   These studies are described in the consultative letters and reports of Clewell (2001a),

7   Yu (2000, 2001, 2002) and Yu et al. (2000).

8

9   **6.3.1.1.1 Drinking Water Study**

10   Perchlorate drinking water experiments used in model development were performed at

11   AFRL/HEST and described in detail in the report Yu (2000). Pregnant dams of the Sprague-

12   Dawley strain were exposed to drinking water treated with perchlorate from gestational day (GD)

13   2 through 20, at perchlorate doses of 0.0, 0.01, 0.1, 1.0 and 10.0 mg/kg-day. GD0 was

14   determined by the presence of a vaginal plug. Both dams and fetuses were sacrificed on GD20

15   and maternal and fetal serum analyzed for free and total thyroxine (fT4 and tT4), triiodothyronine

16   (T3), and TSH. Maternal serum, thyroid, skin, GI contents, placenta, and amniotic fluid were

17   analyzed for perchlorate at all of the above doses. Fetal serum, skin and GI tract were also

18   analyzed for perchlorate at all of the doses. Two hours before sacrifice, the dams were given iv

19   doses of 33 mg/kg radiolabeled iodide ($^{125}I^-$) with carrier. Tissue concentrations of iodide were

20   measured in order to determine the inhibition in the various tissues after long-term exposure to

21   perchlorate.

22

23   **6.3.1.1.2 Preliminary Iodide Kinetics Study**

24   A preliminary study of radiolabeled ($^{125}I^-$) kinetics was performed by AFRL/HEST in which

25   timed-pregnant dams of the Sprague-Dawley strain were exposed via tail-vein injection to a

26   tracer dose (average dose = 2.19 ng/kg body weight) of the radiolabeled anion on GD20. Dams

27   (n=6) were sacrificed at 0.5, 2, 4, and 8 hours post-dosing. Maternal serum, thyroids, skin, GI

28   contents, placenta and mammary gland tissue, as well as fetal serum, skin, and GI tract were

29   collected and analyzed for iodide content at each time point. Serum was pooled for all fetuses

30   within a litter, due to limited sample volume. Fetal skin and GI tract were analyzed individually.

31

1   **6.3.1.1.3 Iodide Inhibition Kinetics Study**

2   A more in-depth study was performed by AFRL/HEST, in which Sprague-Dawley

3   timed-pregnant dams were given 1.0 mg/kg body weight perchlorate via tail-vein injection on

4   GD20; control rats were given saline. The perchlorate or saline dose was followed two hours

5   post dosing with a tail-vein injection of carrier free $^{125}I^-$ at an average dose of 1.87 ng/kg BW.

6   Dams (n=6) were sacrificed after 0.5, 1, 2, 4, 8, 12, and 24 hours. Maternal serum, thyroids,

7   skin, GI contents, placenta, mammary gland tissue, and fetal serum, skin, and GI tract were

8   collected and analyzed for iodide content at each time point. Serum was again pooled for all

9   fetuses within a litter. Fetal skin and GI tract were analyzed individually. At this time, only the

10   maternal serum, maternal thyroids and fetal serum from this study were available for use with the

11   model. Clewell (2001a) states that further validation of the model structure will be performed at

12   a later time with the remaining data, but no further work has been provided to the EPA.

13   Additional data were provided by Yu (2002).

14

15   **6.3.1.2 Data Published in the Literature**

16   Data available in the literature used in a validation exercise of the model are described

17   briefly in this section.

18

19   **6.3.1.2.1 Versloot et al., 1997**

20   Versloot and coauthors measured $^{125}I^-$ as percent of dose in maternal and fetal thyroid,

21   mammary gland, placenta, and fetal carcass without the thyroid. Pregnant Wistar rats (body

22   weight [BW] = 300 ± 5 g) were given an injection of 10 μCi carrier free $^{125}I^-$ into the right vena

23   jugularis on GD19. Measurements of the maternal thyroid were taken at 4 and 24 hours post

24   dosing. Mammary gland, placenta, fetal thyroid, and fetal carcass minus the thyroid were taken

25   only 24 hours post dosing.

26

27   **6.3.1.2.2 Sztanyik and Turai 1988**

28   Sztanyik and Turai measured the uptake of iodide into the placenta and fetal whole body

29   24 hours post dosing. Five groups of CFY albino rats (BW = 200 to 250 g) were dosed ip with

30   370 kBq (0.081 ng) carrier free radiolabeled iodide ($^{131}I^-$) on GDs 17, 18, 19, 20, and 22.

31   Although this is a different strain of rat, the GD20 fetal weights (average BW = 4.088 g) compare

1 favorably with those seen on GD20 in the Sprague-Dawley fetus. As a result, Clewell (2001a)

2 used the GD20 time point as a means of validating GD20 parameters for iodide across different

3 data sets and doses. Placental and whole body fetal ^{131}I were measured in a well-type

4 scintillation detector.

5

6 **6.3.1.2.3 Feldman et al., 1961**

7 Feldman and coauthors measured the uptake of iodide into the fetal thyroid and rest of body

8 carcass on GDs 16, 17, 18, and 19 in pregnant female Holtzman rats. A single subcutaneous

9 injection was given to the dam, containing 50 μCi of ^{131}I on each of the days mentioned above.

10 Fetal thyroid and carcasses were measured at 24 hours post dosing.

11

12 ## 6.3.2 Pregnant Rat and Fetus Model Development

13 This section summarizes only the key features that were different than the adult male rat

14 model previously described in Section 6.2.

15

16 **6.3.2.1 Physiological Parameters and Tissue Partition Coefficients**

17 Maternal parameters were scaled allometrically based on body weight as previously

18 described for the male rat. Fetal values were scaled in the same manner as the maternal

19 parameters. However, since the model actually represents several fetuses, it was necessary to

20 first scale the values for the individual fetus and then adjust for the total number of fetuses in the

21 litter (Clewell, 2001a).

22 Clewell (2001a) based the physiological description of the maternal and fetal rat during

23 gestation on O'Flaherty et al. (1992). However, growth descriptions, body weights, and organ

24 descriptions were optimized for use within this particular model structure. The model is able to

25 account for differences in gestation time, pup birth weight, and litter size between experiments

26 and strains of rats. Growth equations and parameters that change over time were described with

27 mathematical descriptions of available literature and experimental data. Details and equations

28 are provided in the consultative letter (Clewell, 2001a).

29

30

31

6.3.2.1.1 Maternal Tissues

The body weight of the dam is known to change significantly throughout the relatively short gestation time in the rat (21 days). However, the traditional approach utilizing allometric scaling to describe tissue growth in relation to the change in body weight is not a sufficient description for the changes taking place during pregnancy. As opposed to the typical growth scenario, organs and tissues cannot be assumed to increase at the same rate in this dynamic system (Clewell, 2001a). The placenta, fetal volume, and mammary tissue grow at an accelerated rate in comparison to the other organs. These require additional descriptions for their growth beyond the previously described allometric scaling by body weight.

Since the growth of the other tissues is negligible in comparison to the change in the placenta, mammary gland, fat and fetal volume, Clewell (2001a) described the total change in the maternal body weight as simply the change in these four volumes added to the initial (pre-pregnancy) body weight (BW_{init}). All other maternal organs were assumed to remain constant and were scaled allometrically relative to the initial body weight (see Table 6-5).

Mammary tissue growth during gestation was described by Knight and Peaker (1982). Based on this work, Clewell (2001a) described mammary tissue growth as a linear process during which the mammary gland reaches a maximum volume for gestation on GD21 of 4.6% of the maternal body weight.

Clewell (2001a) also described the growth of maternal fat as a linear process throughout gestation based on the work of Naismith et al. (1982). Naismith reported a 40% increase in body fat throughout gestation. Thus, in the model a linear equation was employed to describe a 40% increase in body fat during the length of gestation with an initial (non-pregnant) value of 7.0% body weight for Sprague-Dawley rats (Brown et al., 1997).

Placental volume was described in the model as a sum of three stages of growth, based on the data of Buelke-Sam et al. (1982a), Sikov and Thomas (1970), and the mathematical description of data provided in O'Flaherty et al. (1992). The placenta volume is negligible during gestational days 1 through 5. Individual yolk sac placenta enter a stage of rapid growth between days 6 and 10 of gestation, and was described by an equation that accounted for yolk sac placenta, the total volume of the placenta during this time period, and the number of fetuses present. Placental growth during gestational days 6 through 10 is defined solely by this equation. Total placenta volume changes during gestational days 10 through 21 (parturition) were defined

1   by two separate processes: the exponential decline in yolk sac volume and the increase in

2   chorioallantoic placenta (Clewell, 2001a).

3   O'Flaherty et al. (1992) also described the growth of the uterus and liver during gestation.

4   However, Clewell (2001a) did not include a specific description of growth in these organs

5   because the liver is not believed to have a major role in perchlorate kinetics. Further, because the

6   iodide model does not describe deiodination, the description of liver growth was deemed

7   unnecessary. The use of a uterine compartment was also not included in the Clewell (2001a)

8   model due to the lack of available perchlorate and iodide data. The uterus was considered to be

9   part of the lumped richly perfused tissue. EPA agrees that adding a description of liver growth

10   would only bring additional complexity to the model structure without providing a real benefit to

11   the description of perchlorate and total iodide kinetics and that the uterine compartment would be

12   purely hypothetical and could not be validated without pertinent data.

13

14   **6.3.2.1.2 Maternal Blood Flow**

15   Clewell (2001a) described temporal changes in maternal cardiac output during gestation as

16   the sum of the initial cardiac output, given in Brown et al. (1997) for a non-pregnant rat, and the

17   change in blood flow to the placenta, mammary, and fat tissues. The approach of O'Flaherty

18   et al. (1992) to changing blood flows was utilized in placental, mammary, and fat blood flows.

19   The fraction of cardiac output to the mammary gland and fat tissues are described as proportional

20   to the change in volume of the tissue. The change in blood flow to the yolk sac placenta is

21   approximately proportional to the change in volume of the yolk sac. However, the blood flow to

22   the chorioallantoic placenta increases at a faster rate than the change in volume, so three different

23   equations were used to describe the blood flow for each different stage of placental growth (GD1

24   to GD6, GD7 to GD10, GD11 to GD12, and GD13 to GD21).

25

26   **6.3.2.1.3 Fetal Tissues**

27   A three stage description of fetal growth was also described in O'Flaherty et al. (1992) in

28   order to mathematically reproduce data obtained from Beaton et al. (1954), Sikov and Thomas

29   (1970), Goedbloed (1972), and Buelke-Sam et al. (1982a). Because data are not available for

30   fetal volume between gestational days 1 through 11, an exponential growth curve was used as a

31   reasonable approximation of fetal growth and was fit to the first available data for fetal volume

1   (Clewell, 2001a). The second stage of growth describes a slower increase in fetal volume,

2   beginning on GD11, based on the same data. Clewell (2001a) described the third stage of fetal

3   growth as a linear increase between days 18 and the day of parturition. The equation is

4   dependent on the weight of the pup at the time of birth so that the model can account for the

5   differences in birth weight encountered when simulating different data sets.

6   Individual fetal organ weights were assumed to increase linearly with respect to change in

7   fetal body weight and were therefore scaled allometrically to account for changes in tissue

8   volumes. Values for tissue volumes were taken from the literature and from experimental data

9   for the fetus when available. However, most volumes were taken from adult rat data and scaled

10   allometrically for the fetus due to the lack of tissue data in fetuses.

11   Florsheim et al. (1966) measured thyroid and body weight of the rat fetus and pup from

12   GD18 through PND22 and reported a linear relationship between the thyroid weight and body

13   weight throughout the time period. The value given for the thyroid of the fetus in %fetal body

14   weight for GD19 was used in the Clewell (2001a) model. On the other hand, the physiology of

15   the developing thyroid was found by Conde et al. (1991) to change significantly between birth

16   and PND120. Conde reported a decrease in follicle volume from 61.4% to 37.2% of the total

17   volume of the thyroid from birth to 120 days. An increase in colloid volume from 18.3% of the

18   total thyroid volume at birth to 32.5% at 120 days was also reported. In the absence of

19   histometric data in the fetal thyroid, the follicle, colloid, and stroma volumes for the fetus were

20   described using the thyroid fractions measured at birth. The value for thyroid stroma was

21   calculated within the model by subtracting the colloid and follicle volumes from the total thyroid

22   volume.

23   The fetal body fat content was assumed to be zero in the Clewell (2001a) model. This

24   assumption is reasonable in light of the data given in Naismith et al. (1982). Naismith et al.

25   (1982) measured values for the body fat of PND2 and 16 rat pups, corresponding to 0.16% and

26   3.7% of the body weight. Given that body fat quickly increases in the neonatal period, it is not

27   unreasonable to assume that body fat in the fetus is negligible. The volume is certainly not large

28   enough to interfere with iodide or perchlorate kinetics. All other parameters were scaled

29   allometrically by fetal weight from the adult male rat. The male rat physiological parameters

30   were used rather than female parameters for several reasons. First, the male rat pups have been

31   shown to be more sensitive to perturbation of hormone homeostasis by perchlorate, and therefore

are considered the sensitive endpoint (Yu, 2000). Additionally, Clewell (2001a) asserts that
sufficient evidence was not found to indicate that physiological parameters between male and
female rats were present in the fetus.

6.3.2.1.4 Fetal Blood Flow

Fetal blood flow was assumed to operate independently from the mother. The transfer of
the chemical was accomplished via diffusion between the placenta and fetal blood. Therefore,
the fetal cardiac output and blood flow to organs (as % cardiac output) were scaled allometrically
from the male rat values relative to the fetal volume.

6.2.2.2 Chemical-Specific Parameters

The various active transport processes, tissue permeabilities, and clearance rates (excretion)
are described in PBPK models for each species on a chemical-specific basis. This section
outlines how the values for perchlorate and iodide used in the pregnant and fetal rat model were
derived. The values can be found in Tables 6-4 and 6-5; details on the derivation can be found in
Clewell (2001a).

6.3.2.2.1 Affinity Constants and Maximum Velocities for Active Uptake Processes

These were developed as described previously for the adult male rat model (Merrill, 2001c)
in Section 6.2. The chemical specific parameters were kept the same in male, female, neonatal
and fetal rats, and humans whenever possible. However, it was necessary to change a few of the
parameters, including the maximum velocities (Vmaxc's) in the Clewell (2001a) model for
pregnant rat and fetus. The Km values were similar between tissues and between female and
male rat and human models. However, the maximum velocity or capacity differs between tissues
(Wolff and Maurey, 1961). Since Vmaxc values for perchlorate were not given in literature, the
values were estimated with the model. In order to determine Vmaxc using the model, the
simulation for the tissue of interest was compared to various data sets with several different
perchlorate dose levels. The value for Vmaxc within a given compartment was then determined
by the best fit of the simulation to the data.

6.3.2.2.2 Effective Partitioning Permeability Area Cross Products and Clearance Values

These were developed as described previously for the adult male rat model (Merrill, 2001c) in Section 6.2. The value of 0.05 was used to represent the partitioning of perchlorate into the fat for the pregnant rat and fetus (Clewell, 2001a). This value was based on the data of Pena et al. (1976) who measured tissue:blood ratios in the laying hen after intra-muscular dosing with either a single injection of 10 μCi or 3 sequential doses of 10 μCi radiolabeled perchlorate. Although the hen is a very different species, several other tissues were reported to have values comparable to those found by Yu (2000) and Yu et al. (2000) in the male and female rat (0.3 vs. 0.31 in muscle, 0.1 vs. 0.1 in brain, 0.8 vs. 0.99 in the kidney). Clewell (2001a) noted that the use of this value is supported by the fact that the polarity of the perchlorate anion would severely limit the movement of perchlorate into fatty lipophilic tissue. Anbar et al. (1959) measured the mammary gland:blood ratios in the rat four hours after ip injection of radiolabeled perchlorate (100 mg $KClO_4$), and they reported an effective partition of 0.66 for the rat mammary gland. This value is in general agreement with that chosen by Clewell (2001a).

Maternal and fetal skin were described using the value Perlman et al. (1941) determined after a sc tracer dose of iodide for the partition coefficient in this compartment. Iodide partition coefficients were calculated from the tissue:blood ratios measured during the clearance phase of iodide data in the tissue of interest. The preliminary iodide kinetics study described in the supporting experiments was utilized for the determination of the placenta partition coefficients. For example, values for the GI tract and its contents were determined from the clearance portion of the iodide kinetic study in the adult male rat (Yu et al., 2000).

For all tissues in which a clearance was described (urinary clearance, transfer between placenta and fetal serum, and dissociation of perchlorate from the binding sites), a clearance value was determined. Since perchlorate is quickly excreted in urine and binding has little effect on serum levels at high doses, the simulation for the 10 mg/kg-day dose group was primarily dependent on the urinary clearance value (ClUc_p). The urinary clearance value for perchlorate was therefore based on the fit of the model to the serum data at the high dose. Iodide is incorporated into many of the constituents in plasma. However, it is not bound to the plasma proteins (i.e., albumin) in the same manner as perchlorate. Additionally, the iodide model is currently simplified to account for the distribution of total iodine. Therefore, the urinary clearance value (ClUc_i) was determined primarily by fitting the model simulation to the iodide

1   serum data, as blood levels were more dependent on excretion than on the amount of iodide in

2   other tissues. The clearance of both iodide and perchlorate between the fetal serum and maternal

3   placenta were based on the fit of the model simulation to the fetal and maternal blood levels and

4   to the placenta concentration.

5

6   **6.3.2.3 Pregnant Rat and Fetus Model Parameterization and Validation**

7   This section summarizes how Clewell (2001a) used the various data sets to parameterize

8   the model and the validation exercises performed.

9

10   **6.3.2.3.1 Perchlorate Model Parameterization**

11   Clewell (2001a) performed model parameterization for perchlorate using the data obtained

12   from the AFRL/HEST drinking water studies on GD20. Optimized kinetic parameters (Vmax

13   and permeability area values) were determined by fitting the model simulation to the

14   experimental data. As for the adult male rat and human, it was necessary to account for the

15   serum binding of perchlorate in order to adequately describe the blood perchlorate concentrations

16   at the lower doses (0.01 and 0.1 mg/kg-day). Figure 6-28 illustrates the importance of binding in

17   the model simulations of both maternal (A) and fetal (B) serum at 0.01 (left) versus the 10.0

18   (right) mg/kg-day dose. Binding does not have a noticeable effect on the plasma concentrations

19   in the highest dose. However, as the perchlorate dose decreases, the effect of binding is more

20   pronounced. Therefore, at lower levels, a larger percent of the injected dose will be bound.

21   As the amount consumed is increased, the binding process is saturated and eventually the amount

22   of perchlorate that is bound is negligible in contrast to the large amount of free perchlorate in the

23   plasma. This is to be expected because the number of binding sites is limited.

24   Figure 6-29 shows the fit of the model to the maternal serum (left) and thyroid (right)

25   perchlorate concentration (mg/L) in the dam on GD20. Since saturation of the symporter occurs

26   between the 1.0 and 10.0 mg/kg-day dose groups, the influence of Vmaxc in the tissues was

27   primarily in the 0.01 to 1.0 mg/kg-day doses. Thus, the fit of the model simulation to the data in

28   the lower three doses was used to determine the values for Vmaxc in the tissues. On the other

29   hand, the Vmaxc did not have a significant effect on the highest dose. The model fits to the

30   10 mg/kg-day dose group were primarily affected by the partition coefficients and permeability

31   area values. Clewell (2001a) obtained the permeability area values in the tissues by fitting the
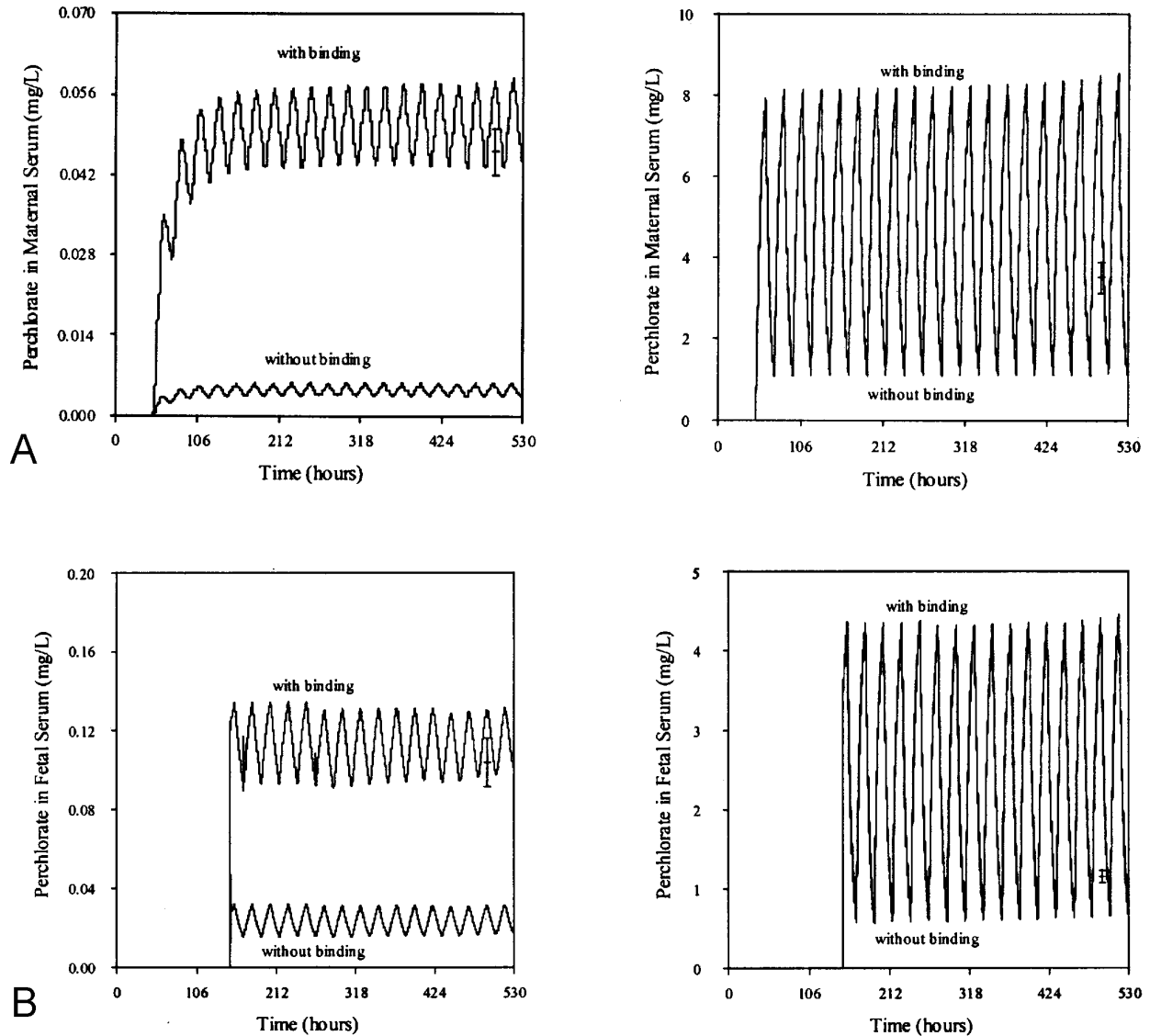
Figure 6-28. Simulations illustrating the necessity of including plasma binding in the
pregnant dam and fetal rat PBPK model structure (Clewell, 2001a). Model
predictions (lines) versus data time course (mean ± SD) are shown with and
without plasma binding for maternal (A) and fetal (B) serum concentrations
(mg/L) at two different doses, 0.01 mg/kg-day (left) and 10.0 mg/kg-day
(right).
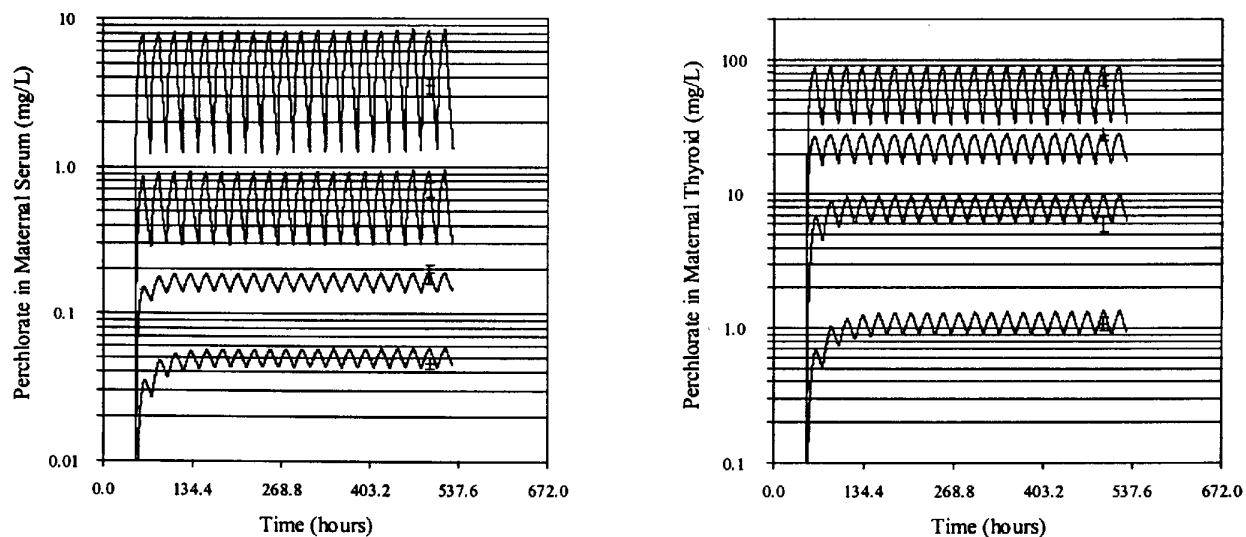
Figure 6-29. Pregnant dam and fetal rat PBPK model predictions (lines) versus data time course (mean ± SD) of perchlorate concentrations (mg/L) in maternal serum (left) and thyroid (right) on GD20 (Clewell, 2001a). Pregnant rats were dosed in drinking water with 10.0, 1.0, 0.1, or 0.01 mg/kg-day perchlorate. Data of Yu (2000).

1 highest dose to the 10 mg/kg-day data in the tissues. Maternal placenta, mammary gland, and GI

2 tract concentrations were available at the 10 mg/kg dose only. These tissues were used to verify

3 the applicability of the assigned partition coefficients to the model. Since mammary glands were

4 not available for the 0.01 through 1.0 mg/kg-day dose groups, it was not possible to fit the

5 Vmaxc value to data at which the symporter has a significant effect. Therefore, the Vmaxc in the

6 mammary gland was assigned the molar equivalent of the iodide Vmaxc. This is probably a

7 reasonable value in the non-lactating gland. Clewell (2001a) provides additional figures that

8 demonstrate the fit of the model to the GI tract, mammary glands, and placenta in the pregnant

9 dam.

10   Fewer data were available for perchlorate distribution in the fetus than in the dam due to

11 the experimental difficulty involved in sampling the small fetal tissues. Figure 6-30 depicts the

12 model simulation of the fetal serum concentration (mg/L) compared to the data obtained in the

13 drinking water study. Fetal serum and skin were pooled by litter. Fits to additional

14 compartments are provided in Clewell (2001a).
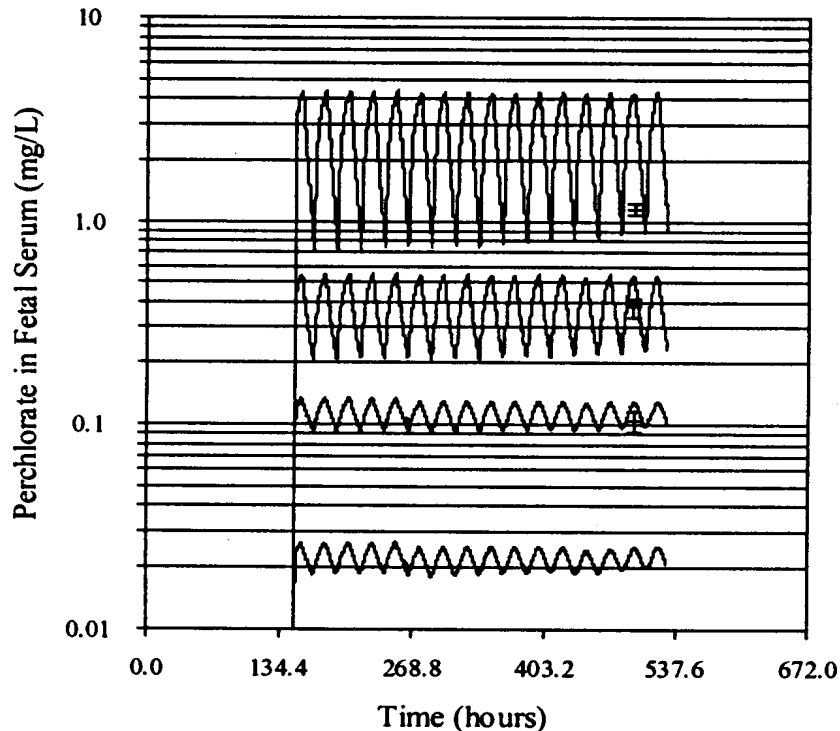
Figure 6-30. **Pregnant dam and fetal rat PBPK model predictions (lines) versus data time course (mean ± SD) of perchlorate concentrations (mg/L) in pooled fetal serum on GD20 (Clewell, 2001a). Pregnant rats were dosed in drinking water with 10.0, 1.0, 0.1, or 0.01 mg/kg-day perchlorate. Data of Yu (2000).**

6.3.2.3.2 Iodide Model Parameterization

Development of the iodide model was performed by fitting the model to the kinetic data in the tissues of the dam and fetus from the preliminary iodide study. Only the values for Vmaxc and permeability area needed to be fit with the model. The clearance value for urinary excretion was determined by fitting the maternal serum prediction to the above data while keeping good fits in the other tissues, such as the maternal skin and gut and the fetal skin. Permeability area values were adjusted to describe the behavior of the iodide data, where varying the permeability area values toward 1.0 L/hr-kg generally increased the rate at which uptake and clearance in a particular tissue occurred; and decreasing permeability area slowed the uptake and clearance. Figure 6-31 shows the model simulation of the iv injection of 2.19 ng/kg $^{125}I^{-}$ on GD20 versus the experimental data for the maternal iodide concentrations in serum (top left), thyroid (top right), mammary gland (bottom left) and placenta (bottom right). The data are described well by the
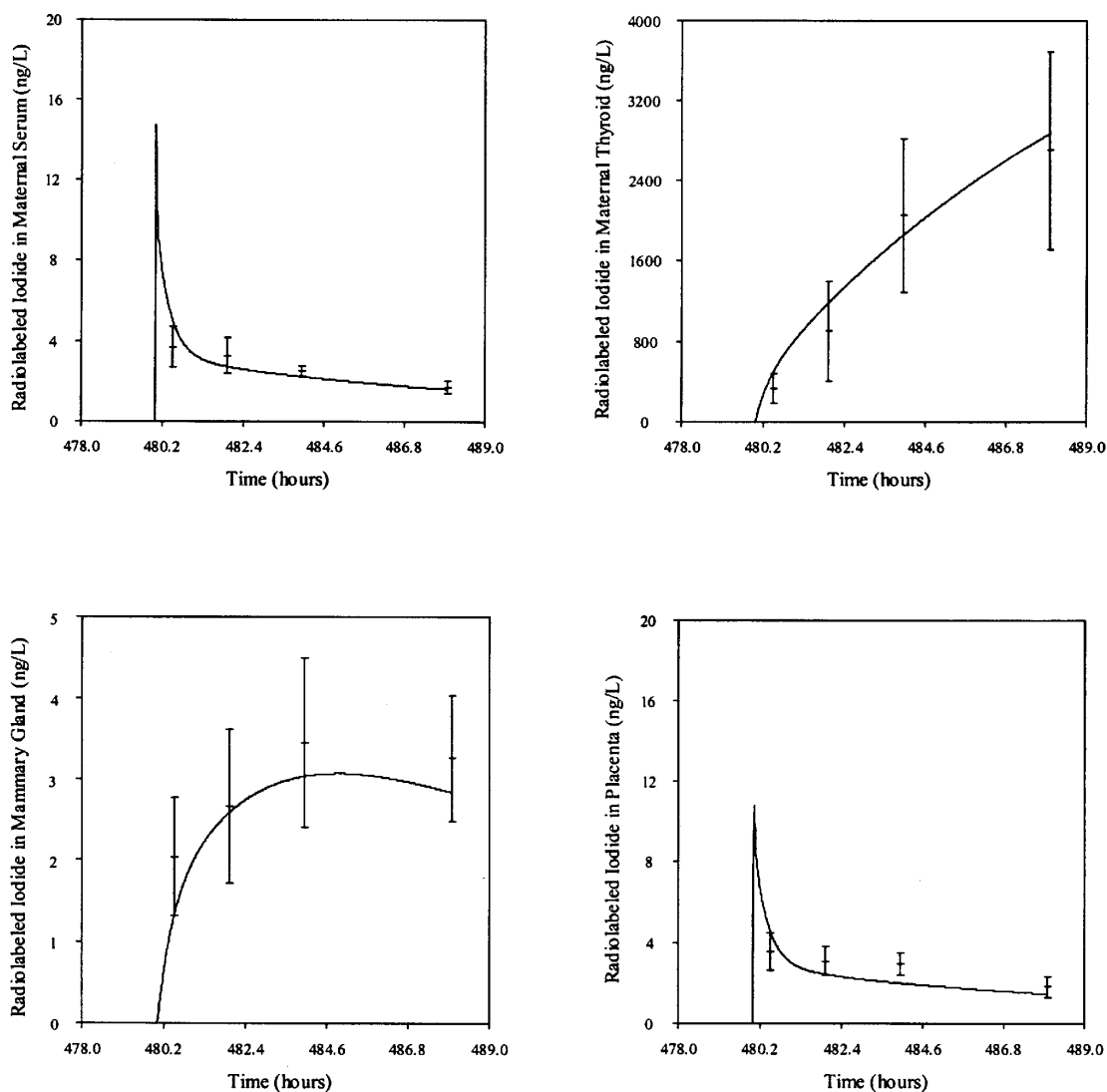
Figure 6-31. Pregnant dam and fetal rat PBPK model predictions (lines) versus data time course (mean ± SD) of $^{125}I^-$ radiolabeled iodide concentrations (ng/L) in maternal serum (top left), thyroid (top right), mammary gland (bottom left), and placenta (bottom right) on GD20 (Clewell, 2001a) Pregnant rats were dosed by iv injection with 2.19 ng/kg $^{125}I^-$ on GD20. Data of Yu (2002).

1   model. The behavior of these mammary tissue data indicate that iodide is maintained in the

2   mammary gland well into the clearance phase of the serum. In order to simulate this behavior, it

3   was necessary to use a low permeability area value (0.02 L/hr-kg) in the mammary gland

1 (Clewell, 2001a). The mammary:plasma ratios of greater than one were fit with the Vmaxc for

2 mammary NIS.

3   Clearance values for the transfer of iodide between the placenta and fetal blood were

4 determined by optimizing the fit of the fetal serum to the data points while maintaining the fit of

5 the simulations of the maternal blood and fetal tissue data. Figure 6-32 shows the model

6 simulation versus the fetal data in the preliminary iodide time course study for radiolabeled

7 iodide in fetal serum (ng/L). Clewell (2001a) shows additional simulations for fetal skin and

8 fetal GI tract.

9

10
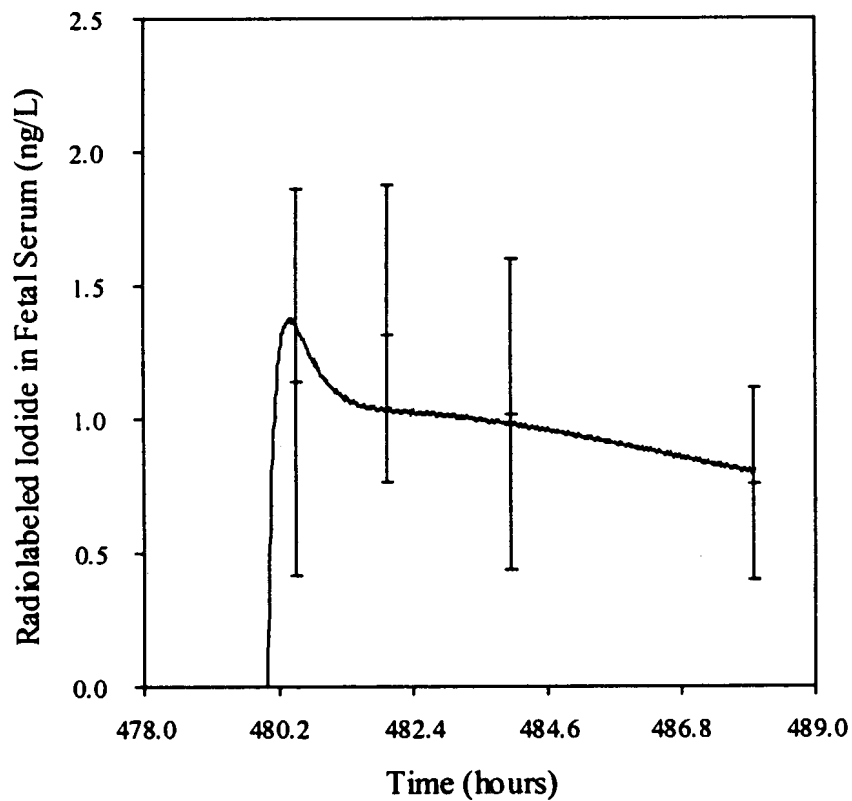
Figure 6-32. Pregnant dam and fetal rat PBPK model predictions (lines) versus data time course (mean ± SD) of [125]I⁻ radiolabeled iodide concentrations (ng/L) in fetal serum on GD20 after an iv injection to the dam with 2.19 ng/kg [125]I⁻ (Clewell, 2001a). Data of Yu (2002).

1   The data of Feldman et al. (1961) were used by Clewell (2001a) to determine the values for

2   maximum velocity of iodide uptake in the fetal thyroid. An exponential function was fit to the

3   experimental values and time points where time was gestation in hours. This equation was then

4   used in the model to account for the increasing ability of the fetal thyroid to incorporate iodide.

5   Iodide levels were negligible on GD16 but increased dramatically from GD17 to GD19 (see

6   Clewell, 2001a; Figure 25).

7

8   ### 6.3.3 Model Validation

9   The Clewell (2001a) model predictions for the inhibition of iodide uptake into the thyroid

10   and the resulting effect on the maternal and fetal serum was validated against the data collected

11   by AFRL/HEST during the inhibition study on GD20. The kinetic parameters derived from the

12   perchlorate drinking water and preliminary iodide data sets were used in the model. Because the

13   inhibition study was performed with an acute perchlorate dose, it was necessary to make some

14   slight changes in the parameters describing thyroid perchlorate kinetics. The long-term exposure

15   to perchlorate in the drinking water studies (18 days) that were used to determine the perchlorate

16   parameters is sufficient to induce up-regulation in the thyroid (Yu, 2000). Thus, it was

17   determined that the thyroid parameters in the dam at this point would be different from those

18   seen in an acute situation. The only parameters altered in order to model the acute perchlorate

19   were the partition coefficient (from 2.25 to 0.13) and permeability area value (from 6.0E-4 to

20   4.0E-5) into the thyroid at the basolateral membrane (thyroid follicle). The value for the

21   partitioning into the follicle in a naive thyroid was calculated as described previously from Chow

22   and Woodbury (1970). The permeability area value in the naive thyroid follicle was determined

23   with the lactation model, which is described in another consultative letter describing model

24   development for the lactating rat (Clewell, 2001b).

25   The model simulation was fit to the available kinetic data in the thyroid while keeping all

26   other thyroid parameters identical to those in the pregnancy model. Figure 6-33 illustrates the

27   model prediction of thyroidal iodide uptake with and without perchlorate inhibition, utilizing

28   these pre-set parameters. The model prediction of inhibition in the thyroid gland at 0.5, 1., 2, 4,

29   8, 12, and 24 hours after dosing with iodine shows an excellent fit to the data. The use of

30   parameters derived from the drinking water perchlorate data for acute iodide uptake kinetics is

31   well supported by the inhibition of iodide because inhibition is highly dependent on the
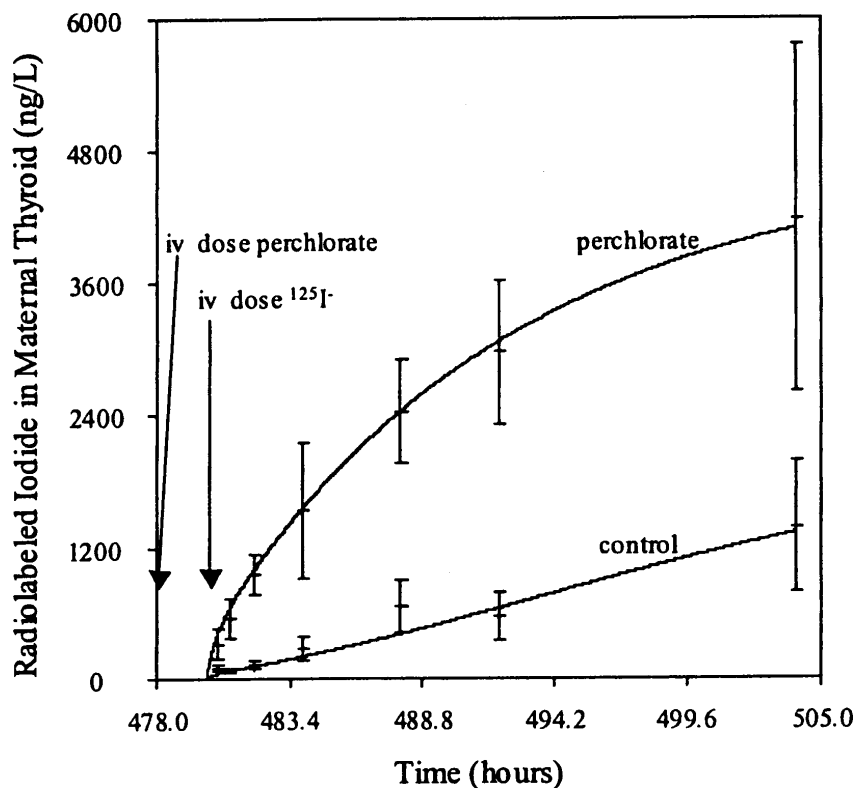
Figure 6-33. Validation for pregnant dam and fetal rat PBPK model (Clewell, 2001a). Model predictions (lines) versus data time course (mean ± SD) of ^{125}I$^-$ radiolabeled iodide concentrations (ng/L) in maternal thyroid with and without 1.0 mg/kg perchlorate administered by iv injection to the dam 2 hours prior to an iv injection with 1.87 ng/kg ^{125}I$^-$ (Clewell, 2001a). The top simulation represents the control thyroid and the lower indicates the inhibited thyroid. Data of Yu (2000, 2002).

1 perchlorate concentration in the thyroid and the perchlorate affinity constants in the apical and

2 basolateral membranes of the thyroid. Figure 6-34 illustrates the effect of perchlorate thyroid

3 inhibition on the maternal (top) and fetal (bottom) blood iodide levels. Significant differences

4 were found in the maternal serum iodide concentrations collected at the 1, 4, and 24 hour time

5 points. Fetal serum, however, did not show any significant differences in the total serum iodide

6 between the control and inhibited groups. Additional statistical analysis of these data are

7 provided as Attachment #2 in Clewell (2001a).

8   Clewell (2001a) performed a model simulation of data presented by Versloot et al. (1997)

9 in order to test the ability of the model to predict diverse data sets collected under different
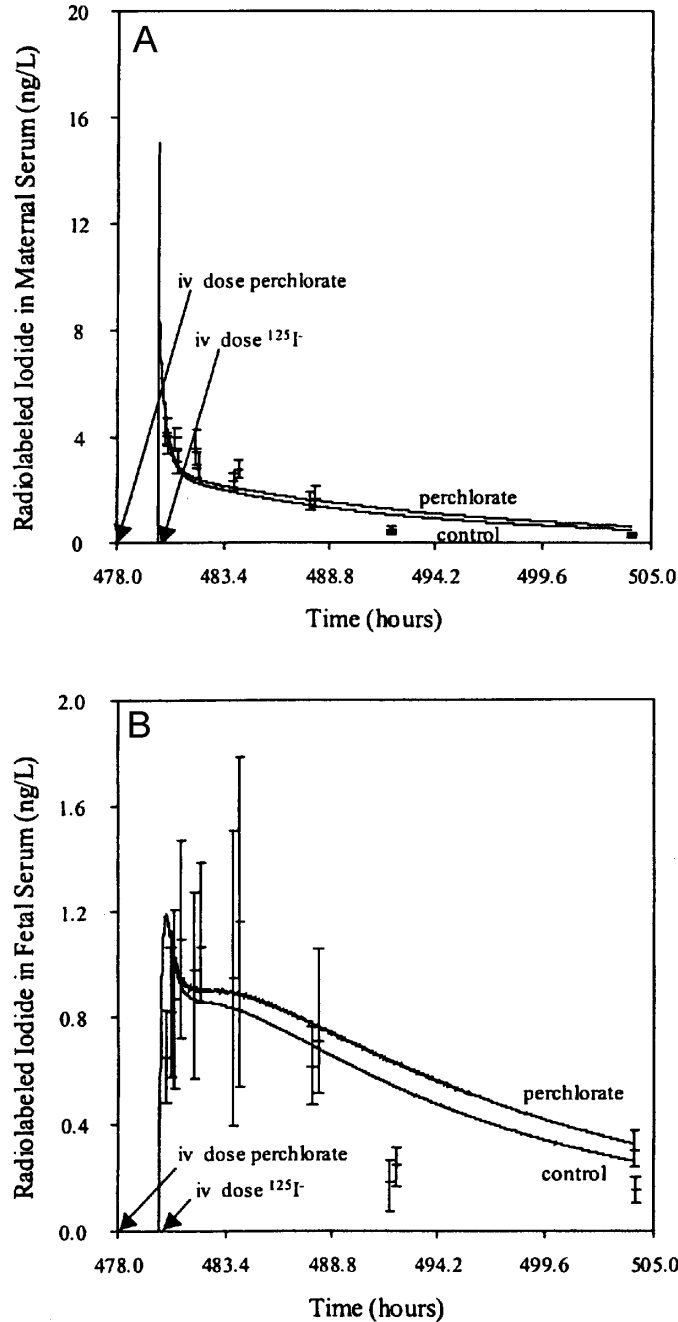
Figure 6-34. Validation for pregnant dam and fetal rat PBPK model (Clewell, 2001a).
Model predictions (lines) versus data time course (mean ± SD) of $^{125}I^-$
radiolabeled iodide concentrations (ng/L) in maternal (A) and fetal (B) serum
with and without a 1.0 mg/kg perchlorate dose administered by iv injection to
the dam 2 hours prior to an iv injection with 1.87 ng/kg $^{125}I^-$ (Clewell, 2001a).
The top simulations in each represents the serum during thyroid inhibition
and the lower represents the control serum. Data of Yu (2000, 2002).

1   conditions. This data set provided an additional time point for the iodide model validation

2   (GD19). Dams were exposed by iv injection to 10 µCi (1.74 ng/kg) carrier-free radiolabeled

3   iodide (^{125}I$^-$) on GD19. Figure 6-35 shows the model predictions versus data (mean ± SD) for the

4   amount (ng) of iodide taken up in maternal thyroid (A), mammary gland (B), and placenta (C), or

5   fetal thyroid (D). The model is able to accurately describe these tissues of interest and fits other

6   compartments (data shown in Clewell, 2001a) within a two-fold factor without changing any

7   parameters. This illustrates its predictive power and usefulness to the extrapolations required.
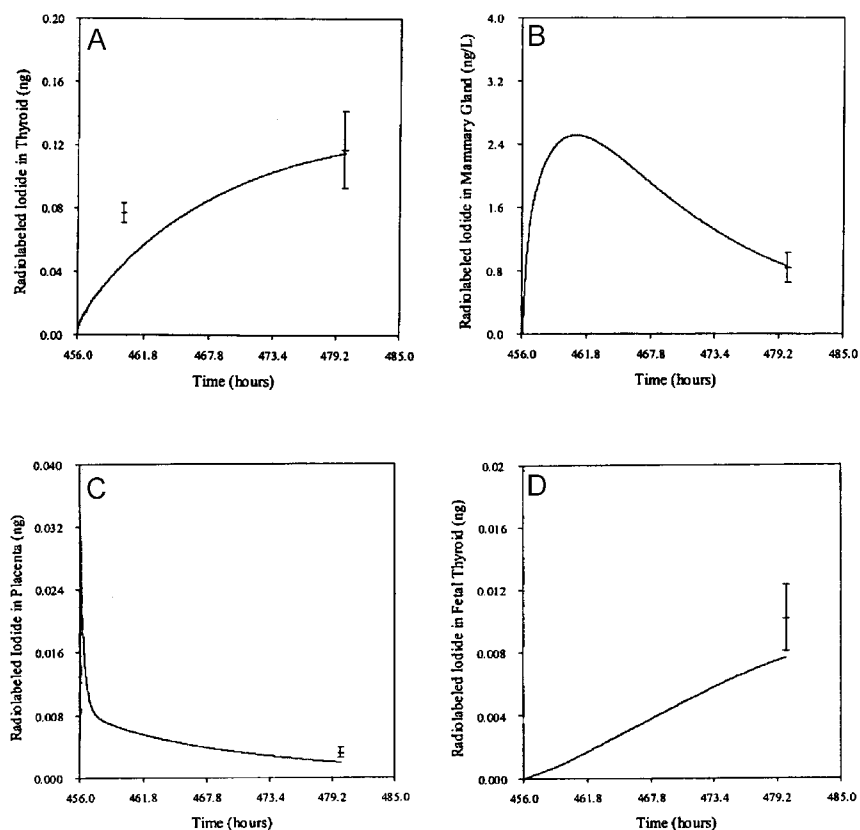
Figure 6-35. Validation for pregnant dam and fetal rat PBPK model (Clewell, 2001a). Model predictions (lines) versus data time course (mean ± SD) of total ^{125}I$^-$ radiolabeled iodide in the maternal thyroid (A), mammary gland (B), placenta (C), or fetal thyroid (D) at 24 hours afer exposure to the dam by iv injection of 10 μCi (1.74 ng/kg carrier-free) ^{125}I$^-$ in GD19 dams. Data of Versloot et al. (1997).

1   Model predictions were also shown to be in good agreement with another unrelated data

2   set, that of Sztanyik and Turai (1988), who measured carrier-free radiolabeled iodide (^{131}I$^-$) in

3   GD20 dams and in the total (whole body) fetuses afer an iv injection (Clewell, 2001a). This

4   validation illustrated adequate model fit to another time point and radioactive species of iodide.

5   The model was additionally validated against AFRL/HEST data for dams and fetuses after

6   administration of radiolabeled iodide (^{125}I$^-$) with carrier at doses four orders of magnitude greater

7   than that used to parameterize the model (33000 ng/kg versus 2.19 ng/kg). These validation

8   simulations are shown in Clewell (2001a).

9   As a final validation exercise, the model was used to predict radiolabeled iodide uptake

10 inhibition after perchlorate exposures in drinking water for 18 days at 0.0, 0.01, 1.0, and 10

11 mg/kg-day (Yu, 2000). It was apparent that even at the lowest dose, the hormonal system had

12 experienced a perturbation and was attempting to compensate for the interruption caused by the

13 perchlorate exposure (Clewell, 2001a). Maternal T4 decreased in a dose-dependent manner,

14 while TSH increased. The maternal total T4 and TSH changes were statistically significant at all

15 doses. Free T4 was significantly increased at the 0.1, 1.0, and 1.0 mg/kg-day doses and total T3

16 was significantly decreased at the 1.0 and 10.0 mg/kg-day doses. The fetus appeared to follow

17 the same trends as those seen in the dam. However, only the 1.0 and 10.0 mg/kg-day dose

18 groups show significant decreases in total T4 and the 0.01, 1.0, and 10.0 mg/kg-day doses

19 resulted in significantly increases in fetal free T4 and TSH. No significant decrease was seen in

20 fetal T3. The statistical analysis of the hormone data is provided as Attachment #3 in Clewell

21 (2001a).

22   From the perspective of iodide kinetics, these hormone changes are important indicators of

23 thyroid up-regulation. When TSH is increased, the thyroid is stimulated to increase iodide

24 uptake. It is evident, then, that after exposure to perchlorate in drinking water for 18 days, the

25 thyroid of the pregnant dam has experienced both inhibition and up-regulation and has

26 successfully compensated for the competition of perchlorate for binding sites of NIS. Therefore,

27 it is not surprising that no inhibition was reported on GD20. It is not that the inhibition is not

28 taking place, but rather that the system has compensated for the effect.

29   None of the models is currently equipped with the capability to account for up-regulation of

30 the thyroid. Therefore, when a simulation of the inhibition is performed with the model, the

31 concentration of iodide is under-predicted in a perchlorate-dose dependent manner (Clewell,

1 2001a). Figure 6-36 shows the model prediction of iodide in the thyroid of the dam at drinking

2 water doses of 0.0, 0.1, 1.0, and 10.0 mg/kg-day. The Vmaxc for iodide was decreased to

3 2.5×10^4 to fit the mean from the control data with the control simulation in order to make the

4 comparison of the inhibition data and simulations clearer. All experimental data were actually

5 taken two hours post dosing. However, the data points were separated slightly by time on the

6 plot in order to make them more visible. The prediction of thyroid perchlorate levels from this

7 same study can be seen in Figure 6-29 (right).

8

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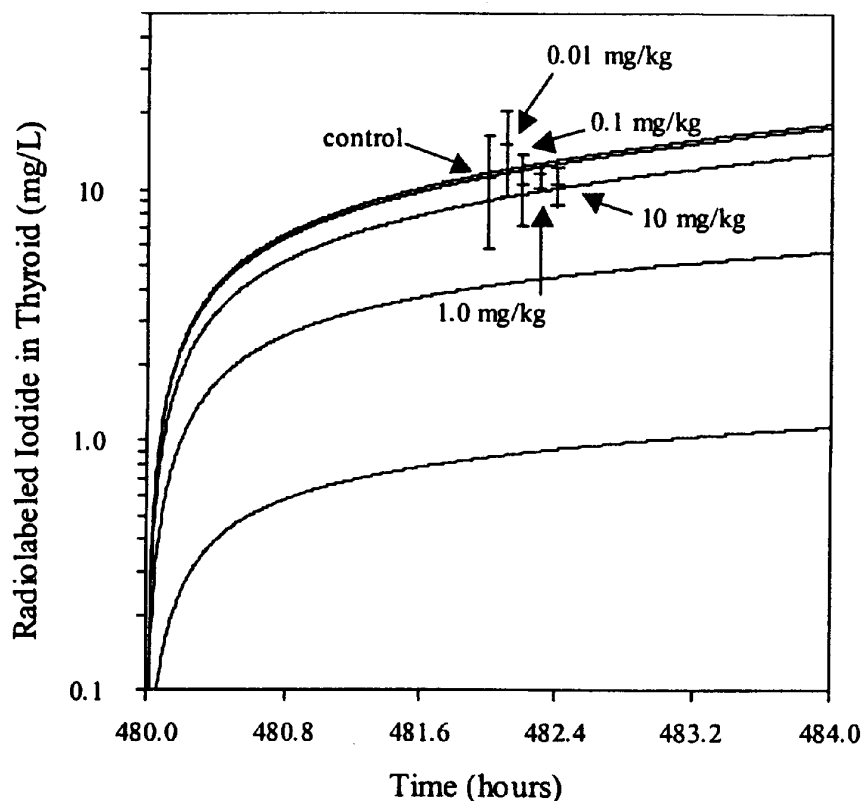
**Figure 6-36. Validation for pregnant dam and fetal rat PBPK model (Clewell, 2001a).
Model predictions (lines) versus data time course (mean ± SD) of radiolabeled
iodide in the maternal thyroid of the dam at doses of perchlorate in drinking
water at 0.0, 0.01, 1.0, and 10.0 mg/kg-day for 18 days. Data of Yu (2000).**

6.3.4 Summary

The proposed model for the pregnant rat and fetus developed by Clewell (2001a) appears to adequately describe perchlorate and iodide distribution in a highly dynamic, changing system, by accounting for growth with age-specific functions. The model predicts the transfer of perchlorate to the fetus and is also able to describe the uptake into fetal tissues of interest, such as the serum and thyroid. Fetal and dam tissues were predicted well by fitting data that spans three orders of magnitude (i.e., 0.01 to 10.0 mg/kg-day).

In addition to the requisite compartments for pregnancy (mammary gland, placenta, fetus), some differences exist that affect the kinetics of both perchlorate and iodide. The thyroidal maximum capacities are lower in the pregnant dam than in the male rat. Model parameterization in the male rat indicated the need for Vmax values for uptake into the follicle of the thyroid of 2.2×10^3 L/hr-kgr for perchlorate and 5.5×10^4 L/hr-kg for iodide, while the gestation model required values of 1.8×10^3 L/hr-kg and 4.0×10^4 L/hr-kg for the same parameters. This difference is supported in the literature. Versloot et al. (1997) suggest that the pregnant rat may have a lowered reserve of iodide in the thyroid toward the end of pregnancy, causing increased activity in the thyroid. The increased response of the pregnant rat was also seen in the studies performed by Yu (2000) and Yu et al. (2000) that reported a greater than average inhibition in the thyroid of the pregnant dam than in the male rat at the same perchlorate dose (78% vs. 70% over 8 hours). The skin of the pregnant dam also required a smaller value for Vmaxc than the male rat. This is supported by the work of Brown-Grant and Petes (1959), which reported higher levels of iodide in the skin male rats than in female rats. Skin, therefore, appears to be a more important iodide reserve in the male rat than the female. It is reassuring that the model is able to account for the majority of differences in the uptake, distribution, and excretion between the male and pregnant female by incorporating known differences in physiology.

Clewell (2001a) notes that at this time the amount of data concerning perchlorate kinetics in the pregnant rat is very limited. Although perchlorate has been used extensively in literature to study the thyroidal uptake of iodide, it has not been commonly used in rat gestation studies. As such, the perchlorate model was limited to utilizing the drinking water studies for parameterization. However, acute kinetic data were available for perchlorate in the lactating dam and were utilized in the development of the rat lactation model (Clewell, 2001b; see Section 6.4). This system is similar to that of the pregnant dam. Consequently, it was possible to simulate the

1   perchlorate kinetics of the dam with the same general model structure, changing only the

2   physiological parameters. Therefore, it seemed reasonable to use the acute perchlorate

3   parameters from the lactation model. The use of the described parameters for acute perchlorate

4   kinetics is also supported by the ability of the model to predict inhibition in the pregnant dam.

5   Clewell (2001a) discusses that acute perchlorate kinetic data to further verify the model are

6   currently being analyzed by AFRL/HEST, and these were provided to the EPA too late for

7   evaluation (Yu, 2002). In these studies, tissues were collected from pregnant dams and fetuses at

8   various time points after iv injections of perchlorate. The use of these data in the modeling effort

9   may be described in draft manuscripts provided to the external peer review.

10   The kinetic behavior of iodide was also accurately simulated with a range of doses that

11   spans nearly five orders of magnitude (0.36 to 33,000 ng/kg). The active sequestration of iodide

12   in maternal and fetal tissues and the transfer of iodide between mother and fetus was described

13   kinetically with the model, and data have been simulated at a variety of doses and at various time

14   points up to 24 hours post exposure. The fact that the model was able to simulate data from

15   other laboratories under a variety of different conditions attests to the validity of the model

16   structure and its applicability to other studies. The ability of the model to predict iodide was

17   indicative of the usefulness of the model for predictive purposes. It was possible to predict

18   inhibition out to 24 hours while simulating the serum and thyroid perchlorate and iodide levels

19   with satisfactory accuracy. This provides support for the chosen model structure, as well as

20   validation for the physiological and chemical descriptions used.

21   Clewell (2001a) notes that the inability of the model to respond to this auto-regulation

22   presents a considerable need for further model development since drinking water scenarios

23   would allow time for the hypothalamic-pituitary-feedback system to upregulate. Given that the

24   temporal windows of developmental susceptibility are not well established across species, this

25   issue may have to wait for further fundamental neurodevelopmental research.

26   The EPA has also become aware of a recent human biokinetic model for iodine and

27   radionuclides at various ages (fetus, children, mothers) that may provide some additional

28   information with which to validate the iodide kinetic components of the proposed models from

29   AFRL/HEST scientists (International Commission on Radiological Protection, 2001, 1989).

30

31

6.4 LACTATING AND NEONATAL RAT MODEL STRUCTURE

This section describes the model developed by AFRL/HEST in response to concerns about interspecies extrapolation of effects observed in laboratory rats immediately after parturition up to about PND22 (Clewell, 2001b) and updates the preliminary structure provided to EPA (Clewell, 2000). The model predicts the distribution of perchlorate within the lactating dam and neonatal rat during these first few weeks of life, and also predicts the short-term effect of acute perchlorate exposure on iodide kinetics, including iodide uptake in the maternal thyroid.

Concern regarding the kinetics of perchlorate in lactating dams and neonates was motivated by the knowledge that the mammary gland is another tissue with active transport via the NIS, as described in Section 6.3. Perchlorate can thus competitively inhibit the uptake of iodide into the mammary gland in a manner reminiscent of the thyroid, and reduce the amount of available iodide to the infant. Studies utilizing radiolabeled iodide in lactating rats have shown perchlorate to be an effective inhibitor of iodide secretion of into breast milk (Potter et al., 1959, Brown-Grant, 1961). The fact that perchlorate not only inhibits the uptake of iodide, but is also taken up itself into the mammary tissue by way of the NIS, results in an additional risk to the neonate. The perchlorate is then concentrated in the milk and transferred to the litter through suckling.

Although early papers suggest that perchlorate is not transferred in milk (Zeghal et al., 1992), newer technology with better analytical sensitivity has detected perchlorate in the milk of rats dosed with as little as 0.01 mg/kg-day perchlorate in drinking water at the AFRL/HEST. The perchlorate levels in 5- and 10-day old neonate serum are comparable to those of the mother (Yu et al., 2000), indicating that the pups are in fact exposed to significant levels of perchlorate through the maternal milk. This information highlighted the need for more information regarding the effect of perchlorate exposure on the neonate.

The model structure is shown in Figure 6-37. Table 6-6 provides the physiological parameters used in the lactating and neonatal rat PBPK models. Table 6-7 provides the perchlorate-specific parameters, and Table 6-8 provides the iodide-specific parameters for each.

The model structure was developed to be consistent with the previously discussed structures for the adult male rat, pregnant rat, and fetus. In fact, an important linking to the pregnancy model was required. Since the experimental data used to develop the lactation model were taken from drinking water studies in which the dosing began on GD2, it was necessary to include initial perchlorate concentrations in the tissues at the time of birth (0 hours). In order to
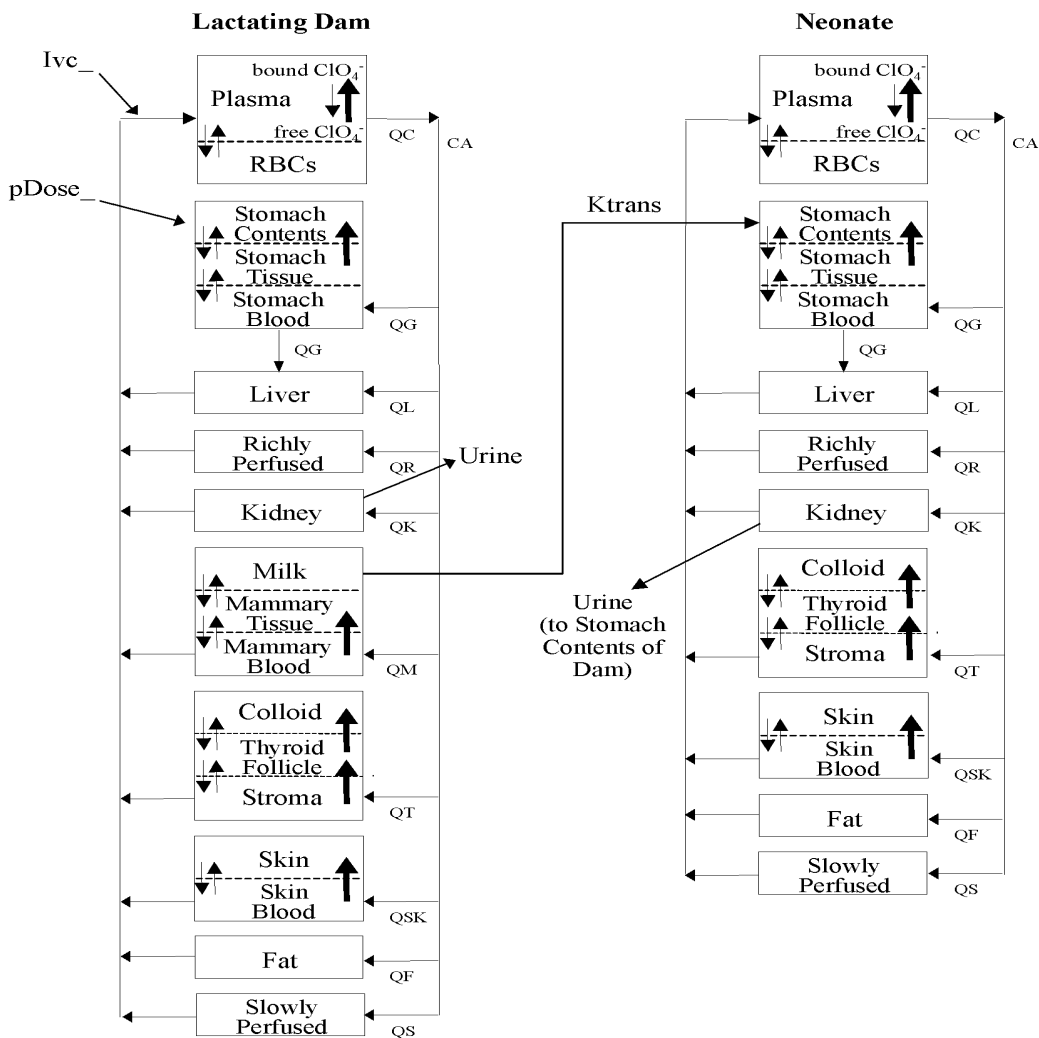
Figure 6-37. Schematic for the lactating dam and neonatal rat PBPK model of perchlorate and iodide distribution (Clewell, 2001b). Boxes represent specific compartments in the model structure. The thyroid consists of the stroma, the follicle, and the colloid, and the stomach consists of the capillary bed, stomach wall, and contents. The skin contains two subcompartments representing the capillary bed and skin tissue. Bold arrows indicate active uptake at NIS sites in the thyroid, skin, mammary gland and GI tract. Plasma binding was also described with Michaelis-Menten terms for the association of perchlorate anion to binding sites with first-order clearance rates for dissociation. Sequestration of the perchlorate (ClO_4^-) and iodide (I^-) anions into milk was also described with Michaelis-Menten kinetics. Permeability area cross products and partition coefficients were used to describe the first-order movement of the perchlorate (ClO_4^-) anion into deeper subcompartments which results from the inherent electrochemical gradient within the tissues. Urinary clearance and transfer of the anions through suckling were represented by first order clearance rates.

TABLE 6-6. PHYSIOLOGICAL PARAMETERS FOR LACTATING DAM AND NEONATE PBPK MODEL (Clewell, 2001b)

Physiological Parameters	Lactation		
Tissue Volumes[a]	Dam	Neonate	Source
Body Weight BW (kg)	0.277 - 0.310	0.0075 - 0.1985	Yu, 2000
Slowly Perfused VSc (%BW)	37.07-40.42	53.92-49.31	Brown et al., 1997
Richly Perfused VRc (%BW)	5.35	5.36	Brown et al., 1997
Fat VFc (%BW)	12.45 - 6.9	0.0 - 4.61	Naismith et al., 1982
Kidney VKc (%BW)	1.7	1.7	Brown et al., 1997
Liver VLc (%BW)	3.4	3.4	Brown et al., 1997
Stomach Tissue VGc (%BW)	0.54	0.54	male rat ClO_4^- kinetics
Gastric Juice $VGJc$ (%BW)	1.68	1.68	Yu, 2000
Stomach Blood $VGBc$ (%VG)	2.9	2.9	Altman & Dittmer, 1971
Skin Tissue $VSkc$ (%BW)	19.0	19.0	Brown et al., 1997
Skin Blood $VSkBc$ (%VSkc)	2.0	2.0	Brown et al., 1997
Thyroid Total $VTtotc$ (%BW)	0.0105	0.0125	Malendowicz & Bednarek, 1986; Florsheim et al., 1966
Thyroid Follicle VTc (%Vttot)	45.89	37.2	Malendowicz & Bednarek, 1986; Conde et al.,1991
Thyroid Colloid $VDTc$ (%VTtot)	45	13.8	Malendowicz & Bednarek, 1986; Conde et al.,1991
Thyroid Blood $VTBc$ (%VTtot)	9.1	49.0	Malendowicz & Bednarek, 1986; Conde et al.,1991
Plasma $VPlasc$ (%BW)	4.7	4.7	Brown et al., 1997, Altman & Dittmer, 1971
Red Blood Cells $VRBCc$ (%BW)	2.74	2.74	Brown et al., 1997, Altman & Dittmer, 1971
Mammary Tissue VMc (%BW)	4.4 - 6.6	—	Knight et al., 1984
Mammary Blood $VMBc$ (%VM)	18.1	—	Assume same % as Thyroid Blood
Milk VMk (L)	0.002	—	Fisher et al., 1990
Blood Flows			
Cardiac Output QCc (L/hr-kg)	14.0 - 21.0	14.0	Hanwell & Linzell, 1973; Brown et al., 1997
Slowly Perfused QSc (%QC)	7.9-1.9	16.9	Brown et al., 1997
Richly Perfused QRc (%QC)	40.8	40.8	Brown et al., 1997
Fat QFc (%QC)	7.0	7.0	Brown et al., 1997
Kidney QKc (%QC)	14.0	14.0	Brown et al., 1997
Liver QLc (%QC)	18.0	18.0	Brown et al., 1997
GI QGc (%QC)	1.61	1.61	Brown et al., 1997
Skin $QSkc$ (%QC)	0.058	0.058	Brown et al., 1997
Thyroid QTc (%QC)	1.6	1.6	Brown et al., 1997
Mammary QMc (%QC)	9.0 - 15.0	—	Hanwell & Linzell, 1973

[a]For calculation of volumes from body weight, a density of 1.0 g/mL was assumed.

TABLE 6-7. PERCHLORATE-SPECIFIC PARAMETERS FOR LACTATING DAM AND NEONATE PBPK MODEL (Clewell, 2001b)[a]

Perchlorate Parameters	Lactation Values		Source
Partition Coefficients (unitless)	**Dam**	**Neonate**	
Slowly Perfused/Plasma PS_	0.31	0.31	Yu et al., 2000
Rapidly Perfused/Plasma PR_	0.56	0.56	Yu et al., 2000
Fat/ Plasma PF_	0.05	0.05	Pena et al., 1976
Kidney/ Plasma PK_	0.99	0.99	Yu et al., 2000
Liver/Plasma PL_	0.56	0.56	Yu et al., 2000
Gastric Tissue/Gastric Blood PG_	1.80	3.21	Yu, 2000; Yu et al., 2000
Gastric Juice/Gastric Tissue PGJ_	2.30	5.64	Yu, 2000; Yu et al., 2000
Skin Tissue/Skin Blood PSk_	1.15	1.15	Yu et al., 2000
Thyroid Tissue/Thyroid Blood PT_	0.13/2.0	0.13/2.0	Chow and Woodbury, 1970; Yu, 2000[b]
Thyroid Lumen/Thyroid Tissue PDT_	7.0	7.0	Chow and Woodbury, 1970; Yu, 2000
Red Blood Cells/Plasma PRBC_	0.73	0.73	Yu et al., 2000
Mammary Tissue/Mammary Blood PM_	0.66	—	Anbar et al., 1959
Milk/Mammary Tissue PMk_	2.39	—	Yu, 2000
Max Capacity, Vmaxc (ng/hr-kg BW)			
Thyroid Follicle Vmaxc_T	1.50E+03	1.50E+03	Fitted[c]
Thyroid Colloid Vmaxc_DT	1.00E+04	1.00E+04	Fitted[c]
Skin Vmaxc_S	8.00E+05	8.00E+05	Fitted
Gut Vmaxc_G	1.00E+06	1.00+06	Fitted
Mammary Tissue Vmaxc_M	2.0E+5/2.0E+4	—	Fitted[b,c]
Milk Vmaxc_Mk	2.00E+04	—	Fitted
Plasma Binding Vmaxc_B	9.00E+03	1.00E+03	Fitted
Affinity Constants, Km (ng/L)			
Thyroid Follicle Km_T	1.00E+05	1.00E+05	Gluzman & Niepomniszcze, 1983; Wolff, 1998
Thyroid Colloid Km_DT	1.0E+09	1.0E+09	Golstein et al., 1992; Wolff, 1998
Skin Km_S	1.00E+05	1.00E+05	Gluzman & Niepomniszcze, 1983; Wolff, 1998
Gut Km_G	1.00E+05	1.00E+05	Gluzman & Niepomniszcze, 1983; Wolff, 1998
Mammary Km_M	1.0E+05	—	Gluzman & Niepomniszcze, 1983; Wolff, 1998
Milk Km_Mk	1.00E+06	—	Fitted
Plasma Binding Km_B	1.00E+04	1.00E+04	Fitted

TABLE 6-7 (cont'd). PERCHLORATE-SPECIFIC PARAMETERS FOR LACTATING DAM AND NEONATE PBPK MODEL (Clewell, 2001b)[a]

	Lactation Values		
Perchlorate Parameters	**Dam**	**Neonate**	**Source**
Permeability Area Cross Products, (L/hr-kg)			
Gastric Blood to Tissue PAGc_	1.00	1.00	Fitted
Gastric Tissue to Juice PAGJc_	1.00	1.00	Fitted
Thyroid Blood to Tissue PATc_	4.0E-05/6.0E-04	4.0E-05/6.0E-05	Fitted[b,c]
Thyroid Tissue to Colloid PADTc_	0.01	0.01	Fitted
Skin Blood to Tissue PASkc_	0.50	1.00	Fitted
Mammary Blood to Tissue PAMc_	0.01	—	Fitted
Mammary Tissue to Milk PAMkc_	0.001/1.0		Fitted
Plasma to Red Blood Cells PRBCc_	1.00	1.00	Fitted
Clearance Values, (L/hr-kg)			
Urinary excretion CLUc_	0.07	0.005	Fitted
Dissociation from Binding Sites Clunbc_	0.034	0.034	Fitted
Transfer from Milk to Pup Ktransc	6.4E-04/1.04E-03	6.4E-04/1.04E-03	Sampson & Jansen, 1984

[a]All parameters listed are notated in the model either by an *i* (for iodide) or *p* (for perchlorate) following an underscore in the parameter name (e.g., PR_*i*, PR_*p*, Vmaxc_T*i*, Vmaxc_T*p*, etc.).
[b]Neonate was given maternal values for Vmax (scaled by body weight) in the absence of data.
[c]Parameters with two values indicate acute and drinking water parameters, respectively.

1   obtain these initial values for tissue loading at birth, the pregnancy model had to include all of

2   the compartments contained in the lactation model (Clewell, 2001a). The pregnancy model was

3   then allowed to run until the day of birth (GD22), and the average tissue concentrations of

4   perchlorate or iodide for the final day of gestation were used as the starting values for the

5   respective tissues in the lactation model (Clewell, 2001b).

6   As discussed, the mammary tissue has been shown to concentrate both perchlorate and

7   iodide during lactation via the NIS symporter. Additionally, hormones produced during lactation

8   such as prolactin which stimulates milk production, have been shown to regulate the mammary

9   NIS. Suckling of the neonatal rats has also been shown to stimulate mammary NIS activity

10   (Tazebay et al., 2000). An additional symporter has been identified in the experiments of

11   Shennan (2001). *In vitro* studies of iodide transport into the mammary gland and the resulting

12   efflux of sulfate from the cells in the absence of sodium cation (Na^+), indicates that another form

13   of transport exists for iodide in the mammary gland in addition to the NIS. Shennan suggests

TABLE 6-8. IODIDE-SPECIFIC PARAMETERS FOR LACTATING DAM AND NEONATE PBPK MODEL (Clewell, 2001b)[a]

Iodide Parameters	Lactation Values		
Partition Coefficients (unitless)	Dam	Neonate	Source
Slowly Perfused/Plasma PS_	0.21	0.21	Halmi et al., 1956
Rapidly Perfused/Plasma PR_	0.40	0.40	Halmi et al., 1956
Fat/Plasma PF_	0.05	0.05	Pena et al., 1976
Kidney/Plasma PK_	1.09	1.09	Perlman et al., 1941
Liver/Plasma PL_	0.44	0.44	Perlman et al., 1941
Gastric Tissue/Gastric Blood PG_	1.00	1.00	Unpublished Lactation Inhibition Study
Gastric Juice/Gastric Tissue PGJ_	1.00	3.50	Unpublished Lactation Inhibition Study
Skin Tissue/Skin Blood PSk_	0.70	0.70	Perlman et al., 1941
Thyroid Tissue/Thyroid Blood PT_	0.15	0.15	Chow and Woodbury, 1970
Thyroid Lumen/Thyroid Tissue PDT_	7.00	7.00	Chow and Woodbury, 1970
Red Blood Cells/Plasma	1.00	1.00	Rall et al., 1950
Mammary Tissue/Mammary Blood PM_	0.66	—	Anbar et al., 1959
Milk/Mammary Tissue PMk_	4.00	—	Yu, 2000
Max Capacity, Vmaxc (ng/hr-kg BW)			
Thyroid Follicle Vmaxc_T	4.00E+04	4.00E+04	Fitted[b]
Thyroid Colloid Vmaxc_DT	6.00E+07	6.00E+07	Fitted[b]
Skin Vmaxc_S	6.00E+04	2.50E+05	Fitted
Gut Vmaxc_G	1.00E+06	2.00E+05	Fitted
Mammary Tissue Vmaxc_M	8.00E+05	—	Fitted
Milk Vmaxc_Mk	5.00E+06	—	Fitted
Affinity Constants, Km (ng/L)			
Thyroid Follicle Km_T	4.00E+06	4.00E+06	Gluzman and Niepomniszcze, 1983
Thyroid Colloid Km_DT	1.00E+09	1.00E+09	Golstein et al., 1992
Skin Km_S	4.00E+06	4.00E+06	Gluzman and Niepomniszcze, 1983
Gut Km_G	4.00E+06	4.00E+06	Gluzman and Niepomniszcze, 1983
Mammary Km_M	4.00E+06	—	Gluzman and Niepomniszcze, 1983
Milk Km_Mk	1.00E+06	—	Fitted
Permeability Area Cross Products, (L/hr-kg)			
Gastric Blood to Gastric Tissue PAGc_	0.80	0.05	Fitted
Gastric Tissue to Gastric Juice PAGJc_	0.60	0.06	Fitted
Thyroid Blood to Thyroid Tissue PATc_	1.00E-04	1.00E-04	Fitted[b]
Thyroid Tissue to Thyroid Colloid PADTc_	1.00E-04	1.00E-04	Fitted[b]
Skin Blood to Skin Tissue PASkc_	0.50	0.02	Fitted
Mammary Blood to Tissue PAMc_	0.02	—	Fitted
Mammary Tissue to Milk PAMkc_	1.00	—	Fitted
Plasma to Red Blood Cells PRBCc_	1.00	1.00	Fitted
Clearance Values, (L/hr-kg)			
Urinary excretion CLUc_	0.03	0.02	Fitted
Transfer from Milk to Pup Ktransc	6.4E-04 - 1.04E-03		Sampson & Jansen, 1984

[a]All parameters listed are notated in the model either by an *i* (for iodide) or *p* (for perchlorate) following an underscore in the parameter name (e.g., PR_*i*, PR_*p*, Vmaxc_T*i*, Vmaxc_T*p*, etc.).
[b]Neonate was given maternal values for Vmax (scaled by body weight) in the absence of data.

1 that this anion transport mechanism is able to transfer perchlorate and iodide into the secretory

2 cells against a concentration gradient. Since the secretory cells are responsible for secreting their

3 contents into the milk, the anion transport mechanism was included in the milk compartment of

4 the Clewell (2001b) model.

5 The structure of the Clewell (2001b) neonatal model is similar to that of the pregnant and

6 fetal rat model, with the exception of the mammary gland compartment as will be described in

7 6.4.2.1.1. In order to simplify the model, all neonates from a single litter were combined in the

8 structure of the model, essentially viewing the entire litter as one entity, or one large neonate.

9 The dose to the neonate is based on the transfer of perchlorate from the maternal milk to the GI

10 contents of the neonate rather than through direct exposure to the drinking water. The 60% of

11 urinary excretion of the neonate is then entered back into the GI contents of the dam in order to

12 account for maternal ingestion of the pup's urine during cleaning, based on the work of Samuel

13 and Caputa (1965).

14 The same challenge posed by the pregnancy model (i.e., to describe perchlorate and iodide

15 distribution in a highly dynamic system) was the objective of the lactating and neonatal rat model

16 (Clewell, 2001b). In addition to total body weight changes in the dam and neonate, maternal

17 mammary tissue and blood flow, cardiac output, fractional body fat and neonatal body weight,

18 and fractional body fat change with respect to time. All tissue volume and blood flow values

19 were adjusted with respect to the changing parameters.

20 Clewell (2001b) assumed the neonate to be nursing at a constant rate, 24 hours a day. This

21 assumption is based on the fact that young nursing rats are unable to go for long periods of time

22 without suckling. The loss through suckling was then described with a first order clearance rate

23 from the mother's milk to the gastric juice of the neonate, based on the experiments of Sampson

24 and Jansen (1984). The milk production rate was assumed to be equal to the amount of milk

25 ingested by the litter.

26

27 **6.4.1 Data and Methods**

28 This section summarizes the data that Clewell (2001b) used for development and validation

29 of the lactating and neonatal rat model structures. Details on experimental methods, including:

30 protocol design, exposure regimen, chemical source and purity, animals (housing, feeding,

1   surgical procedures, etc.), and the analytical methods can be found in the consultative letter and

2   associated reports from AFRL/HEST or papers cited therein.

3

4   **6.4.1.1 AFRL/HEST Experiments in Laboratory Rats**

5   These studies are described in the consultative letter and reports of Clewell (2001b), Yu

6   (2000, 2002), Yu et al. (2000), and Mahle (2000; 2001).

7

8   **6.4.1.1.1 Drinking Water Study**

9   Perchlorate drinking water experiments used in development of the Clewell (2001b) model

10   included this study in which pregnant Sprague-Dawley dams were exposed to drinking water

11   treated with perchlorate from GD 2 through PND5 or PND10 at perchlorate doses of 0.0, 0.01,

12   0.1, 1.0, and 10.0 mg/kg-day. GD0 was determined by the presence of a vaginal plug. Litters

13   were standardized to eight pups (four male and four female, when possible) on PND2. Dams and

14   their litters were euthanized on either PND5 or PND10; maternal and neonatal serum was

15   analyzed for fT4, tT4, T3, and TSH. Maternal serum, thyroid, skin, and gastric contents were

16   analyzed for perchlorate at all doses. Neonatal serum, skin, and GI contents were also analyzed

17   for perchlorate at all doses. Milk was analyzed only on PND10 at all doses. Perchlorate analysis

18   was performed only on maternal gastric tract, mammary tissue, and neonatal gastric tract samples

19   from the PND5 study at the 10.0 mg/kg-day dose. Two hours before euthanization, the dams

20   were given iv doses of 33 mg/kg radiolabeled iodide ($^{125}I^-$) with carrier. Tissue concentrations of

21   iodide were measured in order to determine the inhibition in the various tissues after long-term

22   exposure to perchlorate. This study is described in detail in the consultative letter (Yu, 2000).

23

24   **6.4.1.1.2 Cross-fostering Study**

25   The cross-fostering study involved four groups of rats with varied experimental conditions:

26   true control, control, exposed, and true exposed. True control rats were never dosed with

27   perchlorate. Neonates remained with the dam after birth. In the control group, dams were never

28   exposed to perchlorate in drinking water. However, at the time of birth, the neonates were

29   replaced with pups (less than 24 hours old) that had been exposed to perchlorate throughout

30   gestation (1.0 mg/kg-day to mother through drinking water). In the exposed group, the dams

31   were dosed with 1.0 mg/kg-day perchlorate in drinking water from GD2 to PND10. At the time

1 of birth, the neonates were replaced with pups (less than 24 hours old) that had never been

2 exposed to perchlorate. The true exposed dams were dosed with 1.0 mg/kg-day perchlorate from

3 GD2 to PND10. Neonates remained with their mother after birth. All dams and pups were

4 euthanized on PND10. The skin, GI contents, and serum from the neonates and dam were

5 analyzed for perchlorate. Results indicated that both true control and control (exposed neonates

6 with control dams) showed no perchlorate present on PND10. True exposed and exposed

7 (exposed dams with control litters) showed comparable perchlorate levels on PND10. This study

8 is described in detail in the consultative letters (Mahle, 2000; 2001).

9

10 **6.4.1.1.3 Perchlorate Kinetics Study**

11   In order to evaluate the acute kinetics of perchlorate in the lactating dam and neonate,

12 AFRL/HEST performed a study of the kinetic behavior of perchlorate after the administration of

13 an acute dose. PND10 Sprague-Dawley dams were given 0.1 mg/kg perchlorate by tail-vein

14 injection. The dams were left with their neonates until the time of euthanization at 0.5, 1, 2, 4, 8,

15 or 12 hours post-dosing. Maternal serum, thyroid, stomach contents, skin, and mammary gland

16 were collected and analyzed for perchlorate content at all time points. Neonate serum, stomach

17 contents, and skin were also collected for perchlorate analysis at all time points. Fat, liver,

18 kidney and bladder tissues were also collected from the dam at the eight hour time point.

19 Perchlorate analysis was performed on the serum of the dam and neonates and the maternal

20 thyroid, mammary gland, GI contents, and skin.

21

22 **6.4.1.1.4 Iodide Inhibition Kinetics Study**

23   A study of iodide time course and inhibition kinetics was performed by AFRL/HEST in

24 which Sprague-Dawley timed-pregnant dams were given 1.0 mg/kg body weight perchlorate via

25 tail-vein injection on PND10. The perchlorate dose was followed at two hours post-dosing with

26 a tail-vein injection of carrier free radiolabeled iodide ($^{125}I^-$) at an average dose of 2.10 ng/kg.

27 Dams (n=6) were euthanized after 0.5, 1, 2, 4, 8, and 24 hours. Maternal and neonatal serum,

28 skin, GI contents and tract, as well as the maternal thyroid and mammary gland tissue, were

29 collected and analyzed for total iodide content at each time point. Neonatal serum was pooled by

30 sex in each litter. Neonatal skin and GI contents and tract were analyzed individually.

31

6.4.1.2 Data Published in the Literature

Data available in the literature and used in development and validation of the model are described briefly in this section.

6.4.1.2.1 Sztanyik and Turai, 1988

Five groups of CFY albino rats (BW = 200 to 250 g) were dosed ip with either 370 kBq (0.081 ng) or 740 kBq (1.61 ng) carrier-free radiolabeled iodide (^{131}I$^-$) on PND1 (after 24 hours). Sztanyik and Turai measured the total iodide burden of each litter at 29 hours and on PNDs 2, 5, 7, 9, and 14. Since the litters were not standardized, the number of pups in each litter varied.

6.4.1.2.2 Potter et al., 1959

Four dams of the Long-Evans strain (PND 17-18) were dosed ip with 500 μCi of carrier-free radiolabeled iodide (^{131}I$^-$). Iodide uptake was measured in the milk and plasma of the dam 3, 6, and 24 hours postdosing and in the maternal thyroids 24 hours postdosing.

6.4.2 Lactating and Neonatal Rat Model Development

This section summarizes only the key features that were different than the preceding model structures described in Sections 6.2 and 6.3.

6.4.2.1 Physiological Parameters and Partition Coefficients

Maternal parameters were scaled allometrically based on body weight as previously described for the male rat. Neonatal values were scaled in the same manner as the maternal parameters. However, since the model actually represents several neonates, it was necessary to scale the values for the individual pup first, then to adjust for the total number of pups in the litter as was done in an analogous fashion as for the fetuses in the pregnant rat model (Clewell, 2001a,b).

6.4.2.1.1 Maternal Tissues

During lactation, the mammary gland grows in response to the increased need for milk production by the growing neonates. Knight et al. (1984) measured the mammary gland on several days during lactation. They found the mammary tissue to be 4.4, 5.6, 6.3, and 6.6% of

the maternal body weight on days 2, 7, 14, and 21, respectively. The residual milk was assumed to be 0.002 L based on the model of Fisher et al. (1990). Naismith et al. (1982) examined the change in body fat content of the lactating rat. They reported values for the volume of maternal body fat of 15.2 and 6.9% of the body weight on PND 2 and 16, respectively. The body fat composition of the dam on PND1 was calculated to be 12.4% from the PBPK model for perchlorate and iodide kinetics in the pregnant rat model described in Section 6.3 (Clewell, 2001a).

In order to describe the changes in the physiology of the lactating rat, it was not sufficient to simply scale some of the parameters allometrically. As opposed to the typical growth scenario, some of the tissues in the lactating rat cannot be assumed to increase at the same rate in this dynamic system. Rather, a few tissues, such as the mammary gland and fat, are changing at an accelerated rate in comparison to the other organs. These parameters required additional descriptions for their growth beyond the previously described allometric scaling by body weight. Clewell (2001b) based the approach to modeling these changing parameters on the work of Fisher et al. (1990) with trichloroethylene.

Additionally, the thyroid of the female rat was found by investigators to be significantly larger than that of the male rat (Malendowicz and Bednarek, 1986). Clewell (2001b) assigned values to these parameters based on these data and relevant to the gender and condition (i.e., lactation) of the animal. A value of 1.05% of the maternal body weight was used for the thyroid in the lactation model. The volume fractions of the colloid, follicle, and stroma were given values of 45, 46, and 9% of the thyroid volume. These are significantly different from the values given for the male rat. The volume of the colloid in particular is much greater in the female than the male rat (46 vs. 24% of the thyroid volume). Parameters that were not available specifically for the female were described by adjusting the values for the male rat by body weight.

In the PND10 drinking water study performed by AFRL/HEST (see Attachment #2; Clewell, 2001b), the body weight of the dam showed an average increase of 12% between PND1 and PND10, but did not show a significant difference in weight between dose groups. As a result, Clewell (2001b) calculated the average body weight of the dams for all dose groups for each day of the study and then programmed this changing body weight into the model as a table function.

6.4.2.1.2   Neonatal Tissues

As for the lactating rats, Clewell (2001b) programmed the overall average body weights of the neonates measured on PNDs 3, 5, 7, 9 and 10 into the model is a table function, in order to estimate growth. Naismith et al. (1982) reported the body fat in the pup at PND2 and PND16 to be 0.167 and 3.65% of the neonatal body weight. The amount of body fat in a 41-day old rat was given in Brown et al. (1997) as 4.61% of the body weight.

The volume of the thyroid was studied by Florsheim et al. (1966). The volume of the thyroid was found to increase in a fairly linear relationship with body weight between PND1 and PND22. These investigators reported thyroid volumes of 0.0125, 0.0146, 0.0120, 0.0137, 0.0130, 0.0130, and 0.0131% body weight for neonates on PND1 through 5, 7, and 11. These values were used in a table function in the model to describe the growth of the neonatal thyroid (Clewell, 2001b). The histometry of the thyroid in the neonate was examined by Conde et al. (1991). The authors found a significant difference between the volume fractions of the colloid, follicle and stroma in the neonatal rat versus those in the adult. The reported values of 18.3, 61.4, and 20.3% thyroid volume were used to describe the colloid, follicle, and stroma fractions in the neonatal rat (Clewell, 2001b).

The suckling rate of the neonatal rat has been examined in more than one literature study and has been shown to change over time in response to the growth of the neonatal rats. As the pups grow, they require larger amounts of milk. Sampson and Jansen (1984) measured the amount of milk suckled in rats by removing neonates from the dams for two hours and then allowing the pups to suckle for two hours. This process was repeated throughout the day on several days of lactation. By assuming that the weight gained by the neonates during the suckling period was due to the milk intake and the weight lost while separated from the dam was through excretion, Sampson and Jansen were able to develop an equation that describes the suckling rate of the neonatal rat. Since this equation is dependent on the body weight and growth rate of the neonates, it is able to account for the change over time and the difference between strains and studies. The equation was used in the Clewell (2001b) model which assumed the milk yield of the dam was equal to the suckling rate of the neonate.

1 **6.4.2.1.3 Blood Flows**

2   All maternal and neonatal blood flows that were not directly affected by the changes

3 induced by lactation were scaled by weight from the adult male rat parameters. For those blood

4 flow parameters that change in response to lactation, some additional description was required

5 (Clewell, 2001b). Cardiac output has been shown to increase during lactation (Hanwell and

6 Linzell, 1973). The values given by Hanwell and Linzell (1973) of 14.0, 18.6, 19.0, and

7 21.0 L/hr-kg for days 3, 8, 13, and 23 of lactation were used in the model as a table function to

8 describe the change in cardiac output over time (Clewell, 2001b). Additionally, the blood flow

9 to the mammary tissue was also found to increase during lactation. Reported fractional blood

10 flows to the mammary tissue of 9, 10, 11, 14, 14, and 15% of the cardiac output on PNDs 1, 5,

11 10, 15, 17, and 21, again from Hanwell and Linzell (1973), were used.

12

13 **6.4.2.2 Chemical-Specific Parameters**

14   The various active transport processes, tissue permeabilities and clearance rate (excretion)

15 are described in PBPK models for each species on a chemical-specific basis. This section

16 outlines how the values for perchlorate and iodide used in the lactating and neonatal rat model

17 were derived. The values can be found in Tables 6-7 and 6-8. Details on the derivation can be

18 found in Clewell (2001b).

19

20 **6.4.2.2.1 Affinity Constants and Maximum Velocities for Active Uptake Processes**

21   Whenever possible, chemical specific parameters were kept the same in human and in

22 male, female, neonatal, and fetal rats. However, it was necessary to change a few of the

23 parameters, including the maximum velocity capacity (Vmaxc). The Km values were similar

24 between tissues and between female and male rat and human models. However, the maximum

25 velocity capacity differs between tissues (Wolff and Maurey, 1961). Since values for the tissue

26 maximum velocity capacity for perchlorate (Vmaxc-*p*) were not given in literature, the values

27 were estimated with the model. In order to determine Vmax with the model, the simulation for

28 the tissue of interest was compared to various data sets with several different perchlorate dose

29 levels. The value for Vmaxc within a given compartment was then determined by the best fit of

30 the simulation to the data.

31

6.4.2.2.2 Effective Partitions, Permeability Area Cross Products and Clearance Values

Anbar et al. (1959) measured the mammary gland:blood ratios in the rat four hours after an intra-peritoneal injection of 100 mg radiolabeled perchlorate ($^{36}ClO_4^-$) as potassium perchlorate. They reported an effective partition of 0.66 for the rat mammary gland. Clewell (2001b) used this value in the model. Since the partition for iodide into the mammary gland was not available in the literature, Clewell (2001b) assigned the same effective partition coefficient as used for perchlorate.

When available, iodide partition coefficients were calculated from the tissue:blood ratios measured during the clearance phase of iodide data in the tissue of interest. For example, GI tract and contents were determined from the clearance portion of the data from the iodide kinetic study in the lactating rat.

For tissues in which a clearance was described (urinary clearance and dissociation of perchlorate from the binding sites), a clearance value was determined by fitting the model simulation to the appropriate tissue data. Since perchlorate is quickly excreted in urine and binding has little effect on serum levels at high doses, the simulation for the 10 mg/kg-day dose group was primarily dependent on the urinary clearance value (ClUc_p). The urinary clearance value for perchlorate was therefore based on the fit of the model to the serum data at the high dose. The value obtained in this manner was similar to that determined by fitting the male rat PBPK simulation to urinary perchlorate at several doses (Merrill, 2001a) and to the high dose in the pregnant rat (Clewell, 2001a). The rate of dissociation of perchlorate from the binding sites was fit to the serum data across doses.

6.4.2.3 Lactating Rat and Neonate Model Parameterization and Validation

This section summarizes how Clewell (2001b) used the various data sets to parameterize the model and how the validation exercises were performed.

6.4.2.3.1 Perchlorate Model Parameterization

Clewell (2001b) performed model parameterization for perchlorate using the data obtained for the tissues from the AFRL/HEST drinking water studies on PND5 and PND10. Optimized kinetic parameters (Vmaxc and permeability area) were determined by visually fitting the model simulation to the experimental data. As for the previous model structures (adult male rat, human,

1 pregnant rat and fetus), it was necessary to account for the serum binding of perchlorate in order

2 to adequately describe the serum perchlorate concentrations at the lower doses (0.01 and

3 0.1 mg/kg-day). Figure 6-38 illustrates the importance of binding in the model simulations in the

4 dam on these days.

5

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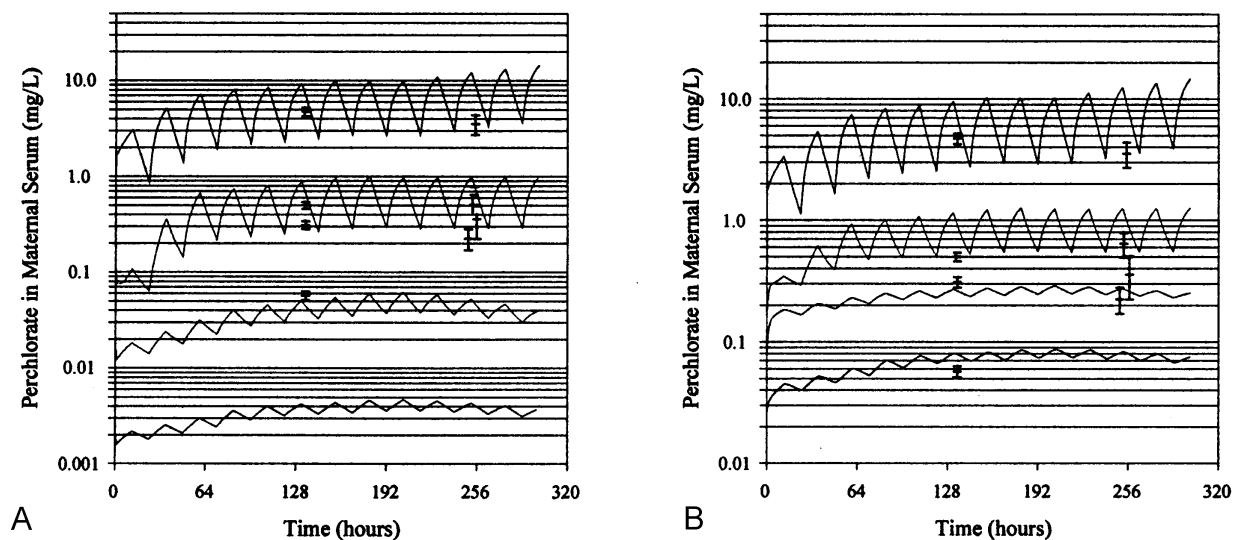
Figure 6-38. Simulations illustrating the necessity of including plasma binding in the lactating dam and neonatal rat PBPK model structure (Clewell, 2001b). Model predictions (lines) versus data time course (mean ± SD) for maternal serum perchlorate concentrations (mg/L) on PND5 and PND10 at doses to the dam of 10.0, 1.0, 0.1, and 0.01 mg/kg-day are shown with (A) and without (B) plasma binding.

1   Figure 6-39 shows the perchlorate tissue concentrations (mg/L) in the lactating dam thyroid

2 (A) and in maternal milk (B) at PND5 and PND10 for the 0.01, 0.1, 1.0 and 10.0 mg/kg-day

3 doses. It was noticed that during the drinking water studies, the daily dose to the dams varied

4 somewhat due to their changing water intake. Therefore, all of the model simulations of the

5 drinking water studies reflect the actual daily dose to the dam, which Clewell (2001b) calculated

6 from the daily water consumption and body weight measurements.

7   Figure 6-40 shows the model simulations of the male and female neonate plasma levels

8 compared to the data obtained in the AFRL/HEST drinking water study. Plasma concentrations
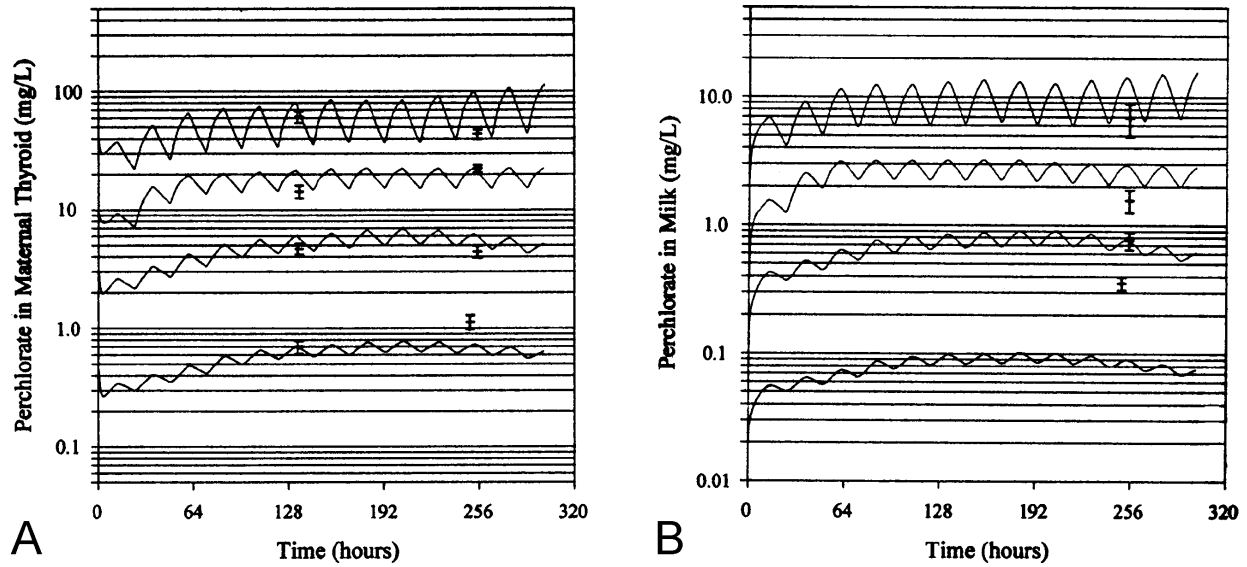
Figure 6-39. Lactating dam and neonatal rat PBPK model predictions (lines) versus data time course (mean ± SD) of perchlorate concentrations (mg/L) in the maternal thyroid (A) and milk (B) on PND5 and PND10 at doses in drinking water to the dam of 10.0, 1.0, 0.1, or 0.01 mg/kg-day perchlorate (Clewell, 2001b).
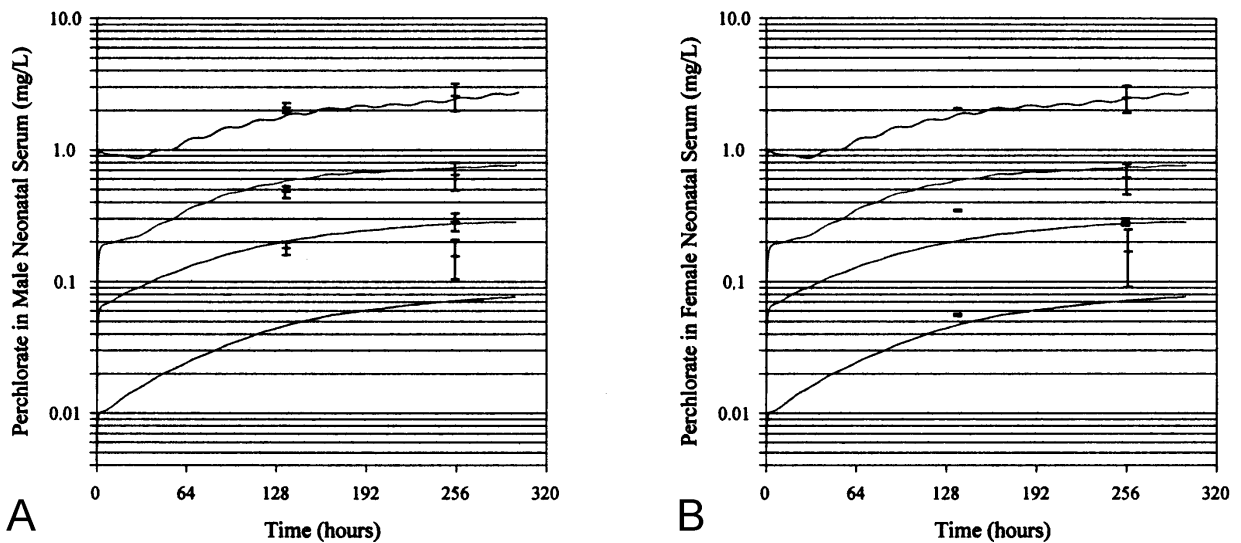
Figure 6-40. Lactating dam and neonatal rat PBPK model predictions (lines) versus data time course (mean ± SD) of perchlorate concentrations (mg/L) in the serum of male (A) and female (B) neonates on PND5 and PND10 at doses in drinking water to the dam of 10.0, 1.0, 0.1, or 0.01 mg/kg-day perchlorate (Clewell, 2001b).

1   varied significantly between the male and female neonates, and Clewell (2001b) noted that the

2   difference appears to be a function of age. At PND5, the male neonatal plasma concentrations

3   were nearly 4 times higher than those of the female neonates in the 0.1 mg/kg-day dose group.

4   By PND10, however, no significant sex difference was found in the plasma perchlorate

5   concentrations at the same dose.

6   Clewell (2001b) fit the male neonatal serum data because the male pups showed higher

7   perchlorate concentrations in the serum than the female pups (Yu, 2000). The neonatal serum

8   was under-predicted by the model in the 0.01 mg/kg-day dose group. Clewell (2001b) strongly

9   asserts that this was due to the fact that the milk concentration was also under-predicted in that

10 same dose group. The three higher doses are well described in the male neonate. The female

11 pups also show acceptable fits at PND10. However, since the PND5 data were much lower in

12 the female than male neonates, the model over-predicts the PND5 time-points in the 0.1 and

13 1.0 mg/kg-day doses. Fits of the model to neonatal skin and GI tract are discussed in Clewell

14 (2001b).

15   As in the maternal model, the clearance value for urinary excretion was determined by the

16 fit of the model to the serum from the 10 mg/kg-day dose, while the lower doses were used to

17 determine the kinetic parameters for the binding in the neonate. Both binding and urinary

18 clearance were considerably lower in the pup than in the dam (Table 6-7).

19

20 **6.4.2.3.2 Iodide Model Parameterization**

21   Clewell (2001b) developed the iodide aspect of the model by visually fitting the model to

22 measured tissue concentrations in the dam and neonate from the control group of the inhibition

23 kinetic study. Only the values for Vmax and permeability area needed to be fit with the model.

24 As shown in Figure 6-41, the model simulations of iodide concentrations (ng/L) after an iv

25 injection of 2.10 ng/kg radioalabeled iodide (^{125}I$^-$) on PND10 versus the experimental data in the

26 lactating dam are shown in the dam serum (A) and thyroid (B) and in male (C) and fetal (D)

27 neonatal serum.

28   The model simulations describe the data well with the exception of the longest time point

29 in the neonates. The clearance value for urinary excretion was determined by fitting the maternal

30 serum prediction to the above data while keeping good fits in the other tissues, such as maternal

31 skin, GI, and mammary gland (Clewell, 2001b). Permeability area values were adjusted to
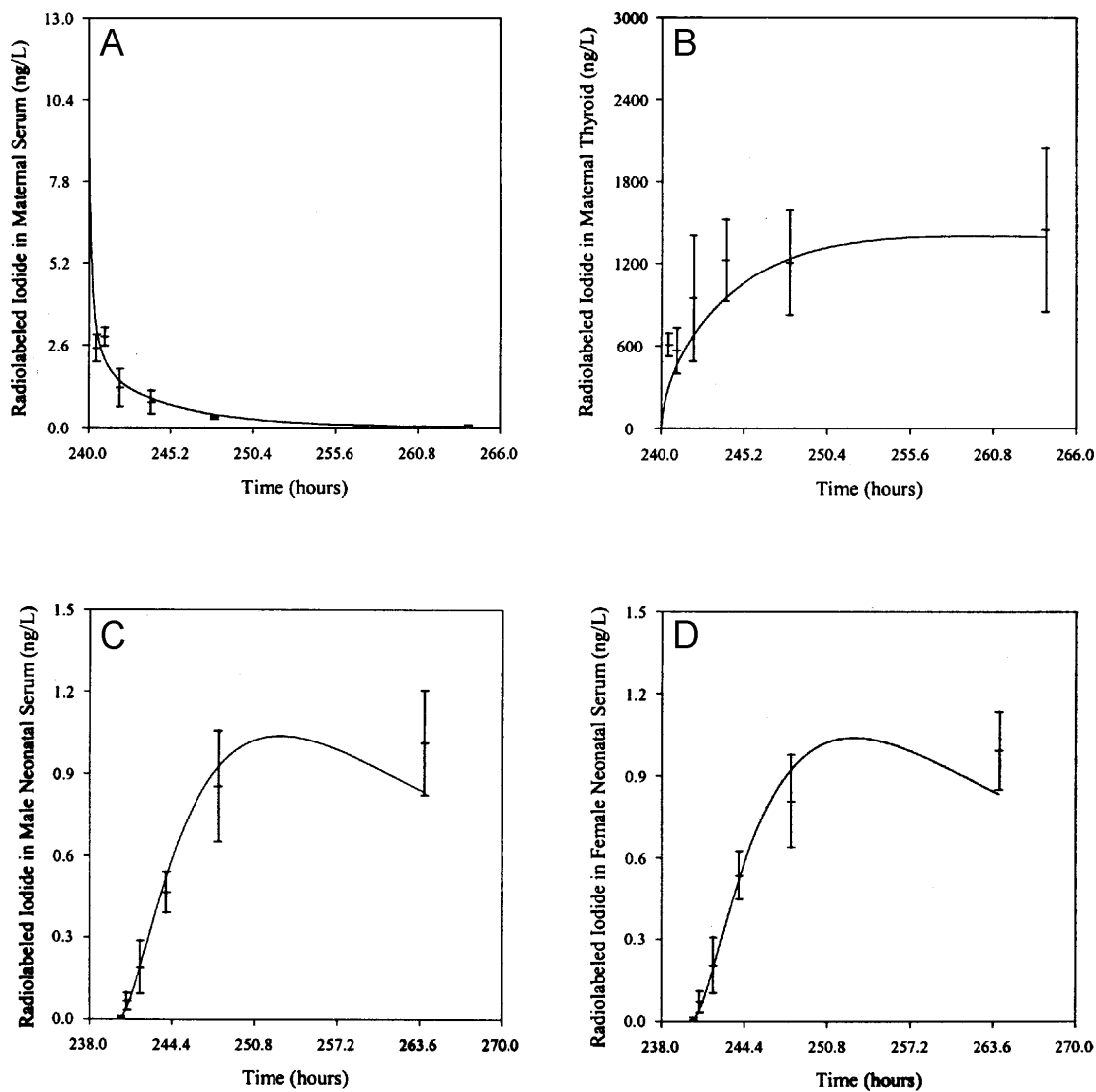
Figure 6-41. Lactating dam and neonatal rat PBPK model predictions (lines) versus data time course (mean ± SD) of iodide concentrations (mg/L) in the maternal serum (A) or thyroid (B) and in male (C) or female (D) neonatal pups on PND10 after an iv dose to the lactating dams of 2.10 ng/kg ^{125}I⁻ (Clewell, 2001b). Data of Yu (2000, 2002).

1 describe the behavior of the iodide data; varying the permeability area values toward 1.0 L/hr-kg

2 generally increased the rate at which uptake and clearance in a particular tissue occurred;

3 decreasing permeability area slowed the uptake and clearance.

1   The behavior of the iodide in the neonatal skin and GI tract and contents appeared to be

2   different from the dam. The iodide tended to stay in the tissue of the neonate longer, requiring a

3   slower clearance in the fetal tissues than was used in the corresponding maternal tissue. As a

4   result, permeability area values used for the GI and skin in the neonate were lower than those

5   used in the dam (Table 6-8). For example, the permeability area value in the skin was

6   determined to be 0.5 L/hr-kg in the dam, but was decreased to 0.02 L/hr-kg in the neonate.

7   However, these values correspond well to the values used for the fetus in the pregnancy model

8   (Clewell, 2001a).

9   The neonatal urinary clearance value was determined to be 0.02 L/hr-kg in the neonate,

10   which is very similar to the maternal value (0.03 L/hr-kg of the dam). This was a surprise,

11   because the neonate was expected to have a much lower rate of excretion than the more mature

12   dam; however, Clewell (2001b) notes that this trend is supported in the literature. Capek and

13   Jelinek (1956) measured the amount excreted by pups at various ages. The neonates required

14   external stimulation by the mother in order to release the urine from their bladders. However,

15   when that stimulation was supplied, the neonates were able to excrete urine at the same rate as an

16   adult rat. Therefore, it is reasonable that the urinary excretion rate is similar between the pup and

17   adult. The amount of iodide lost to urine is then dependent on both the urinary clearance value

18   and the concentration of the ion in the kidney (Clewell, 2001b).

19

20   **6.4.3 Model Validation**

21   The ability of the model to simulate the kinetics of perchlorate in the lactating dam and

22   neonate was tested against the perchlorate time course data collected *in vivo* by AFRL/HEST.

23   Since the study was performed with an acute perchlorate dose, it was necessary to make minor

24   changes in the thyroid perchlorate parameters. The long-term exposure to perchlorate in the

25   drinking water studies that were used to determine the perchlorate parameters is sufficient to

26   induce up-regulation in the thyroid (Yu, 2000). Therefore, the thyroid parameters in the dam at

27   this point would be different from those seen in an acute situation. Clewell (2001b) achieved the

28   model fits to the acute data by altering the partition coefficient (from 2.25 in the drinking water

29   to 0.13 in the acute exposure) and permeability area value (from 6.0E-4 to 4.0E-5) into the

30   thyroid at the basolateral membrane (thyroid follicle). The value for the partitioning into the

31   follicle in a naïve thyroid was calculated as described previously from Chow and Woodbury

1   (1970). The permeability area value in the naïve thyroid follicle was determined by fitting the

2   model prediction to the thyroid data, while keeping good fits in the serum and other tissues.

3   Figure 6-42 shows the model predictions versus the data time course of perchlorate

4   concentrations in maternal serum (A), thyroid (B), or mammary gland (C) and in neonatal serum.

5

6
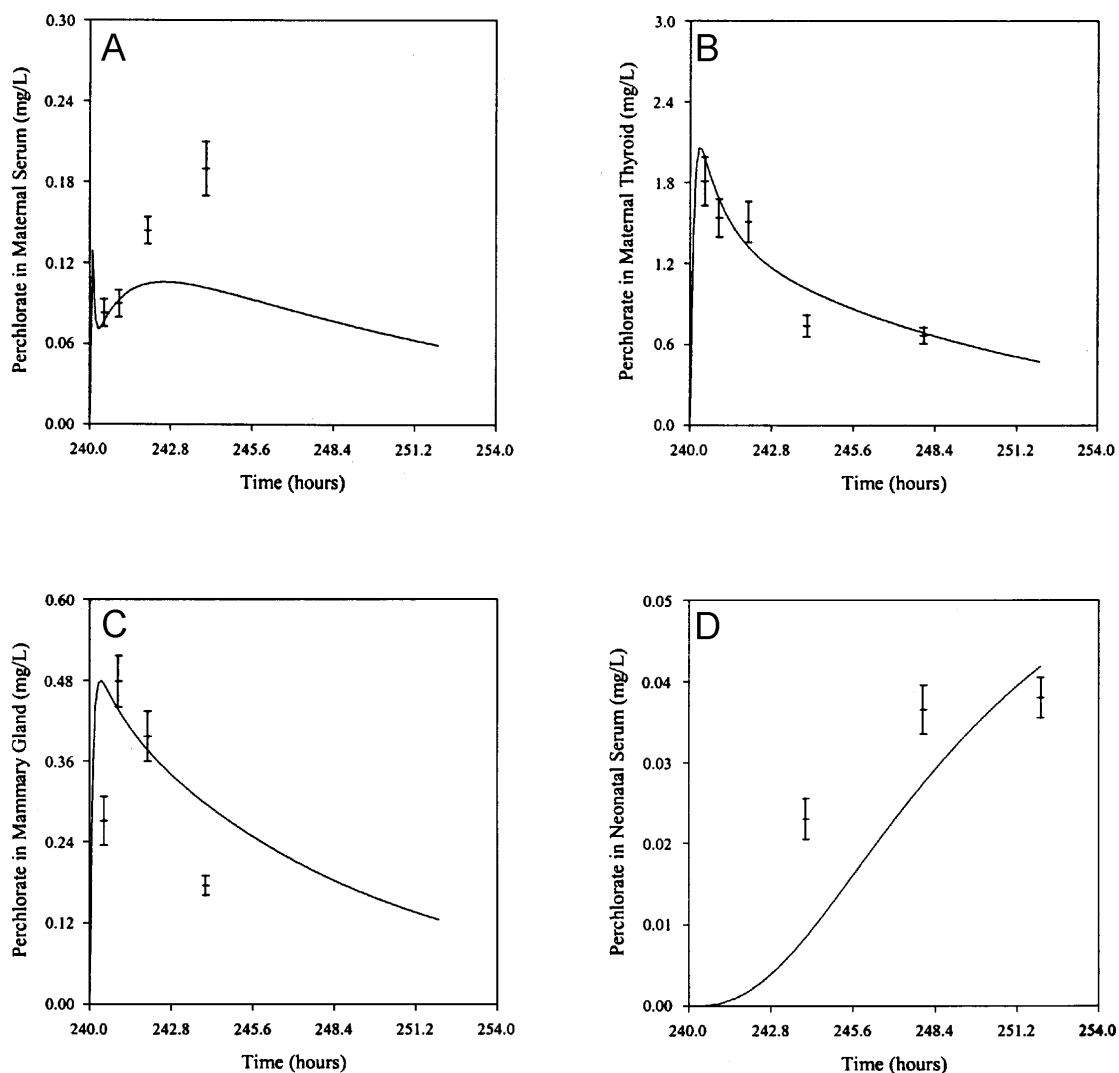
Figure 6-42. Validation for lactating dam and neonatal rat PBPK model (Clewell, 2001b). Model predictions (lines) versus data time course (mean ± SD) of perchlorate in the maternal serum (A), thyroid (B), or mammary gland (C) and in neonatal serum (D) after an iv dose of 1.0×10^6 mg/kg perchlorate on PND10. Data of Yu (2000, 2002) and Yu et al. (2000).

1   The maternal serum is not fit particularly well and the neonatal serum fit could also be

2   improved. Clewell (2001b) notes the difficulties may be due to the use of the iv kinetic data as

3   well as some additional challenges not yet met by the model with respect to the mammary gland.

4   Clewell (2001b) increased the transfer of perchlorate through the milk in the acute studies in

5   order to fit the model derived on drinking water studies to these acute (iv) data. That is, the

6   value for the Vmax into the mammary tissue was increased in order to allow more perchlorate

7   into the mammary compartment, and the permeability area into the milk was decreased in order

8   to minimize the back flow of perchlorate into the mammary from the milk. This essentially

9   forced the perchlorate in the milk to be passed to the neonate rather than return to the mammary

10   tissue of the mother. The Vmaxc for the binding in the neonate was decreased slightly from the

11   value used in the drinking water simulations. This may have been due to increased transfer of

12   iodide in the acute simulations. When the same parameters were used in the mammary

13   compartment that were determined with the drinking water studies, the amount in the mammary

14   tissue was low and the clearance of the mammary was too slow. As a result, acute neonatal

15   serum levels were under-predicted. By adjusting the Vmaxc, the model was able to achieve

16   reasonable fits to the available data in the maternal and neonatal tissues. Clewell (2001b)

17   suggests that different fractions of the dose are transferred through the milk during an acute (iv)

18   exposure versus a drinking water scenario.

19   Figure 6-43 shows the model predictions against the data obtained in the AFRL/HEST

20   cross-fostering study described in Section 6.4.1.1.2. Perchlorate concentrations (mg/L) in the

21   maternal thyroid of dams exposed during gestation (A) or only during lactation (B) show similar

22   results. Perchlorate concentrations (mg/L) in neonatal serum exposed only during gestation (C)

23   or only during lactation (D) also contained similar levels. Because the data were taken on

24   PND10, the sex difference seen at the earlier time points was not present and the simulation is

25   shown for the average of all pups.

26   The model is able to predict the data from the cross-fostering study very well. It is apparent

27   from the data and from the model prediction of the cross-fostering data that the gestational

28   exposure to perchlorate does not affect the perchlorate concentrations of the maternal serum and

29   thyroid or the neonatal serum. This is in agreement with other studies that indicate the rapid

30   clearance of perchlorate in the urine (Yu et al., 2000), but not in agreement with the toxicological

31
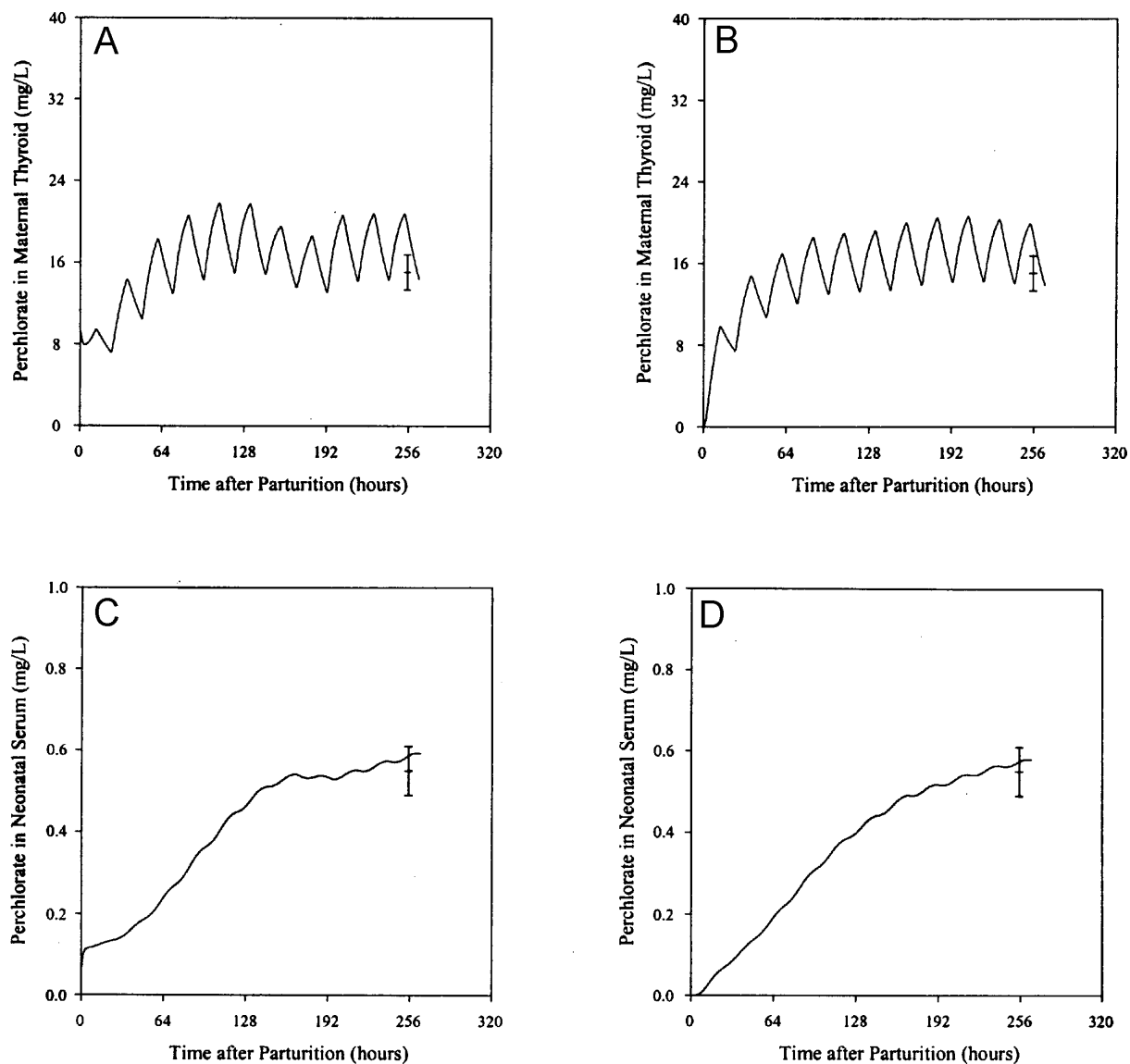
Figure 6-43. Validation for lactating dam and neonatal rat PBPK model (Clewell, 2001b). Model predictions (lines) versus data time course (mean ± SD) of perchlorate in the maternal thyroid during gestation (A) or during lactation only (B) and in the neonatal serum during gestation (C) or during lactation only (D) after an iv dose of 1.0×10^6 mg/kg perchlorate on PND10. Data of Mahle (2001).

1   observations between the 1998 and 2001 developmental neurotoxicological studies performed by

2   Argus Research Laboratories, Inc. (1998; 2001). Differences in the hormone data are discussed

3   in Clewell (2001c) and other differences may be due to strain differences (Fail et al., 1999).

4   From the model, even though the neonatal urinary excretion is much lower than that of the dam

1   (0.005 vs. 0.07 L/hr-kg), the prenatal exposure does not affect the serum levels of the neonate

2   past PND2. This is in accord with the observations made of the BMDL estimates for the post-

3   natal thyroid discussed in Chapter 5.

4   Additional validation exercises were performed by Clewell (2001b), showing reasonably

5   adequate model fits to the data of Potter et al. (1959) and that of Sztanyik and Turai (1988) as

6   shown in Clewell (2001b). Maternal radiolabeled iodide concentrations were overpredicted in

7   the thyroid on PND18. The maternal milk concentrations were also overpredicted for the earlier

8   time point, but were within the range at the later. The model predicted the radiolabeled iodide

9   data obtained in the litters of Sztanyik and Turai (1988) quite well. This indicates that the

10   lactation and neonatal kinetics are characterized accurately.

11   Figure 6-44 shows that the Clewell (2001b) model is able to predict the radiolabeled iodide

12   ($^{125}I^-$) uptake-inhibition data in maternal thyroids on PND10 from the AFRL/HEST "acute" (iv)

13   studies with perchlorate. The inhibition was described well by the model across the range of

14   time points from 0.5 to 24 hours postdosing. The top line indicates the prediction for the control

15   thyroid, and the bottom line shows the effect of perchlorate. The model is able to describe the

16   kinetics of iodide under both conditions.

17   The Clewell (2001b) model is also able to predict the radiolabeled iodide uptake inhibition

18   data from AFRL/HEST obtained after "chronic" drinking water exposures. Figure 6-45 shows

19   the radiolabeled iodide ($^{125}I^-$) concentrations (mg/L) in the maternal thyroids at PND5 after

20   23 days of dosing with perchlorate at 0.0, 0.01, 1.0, and 10.0 mg/kg-day.

21

22   **6.4.4 Summary**

23   Clewell (2001b) highlights some important differences in the lactating dam and neonatal rat

24   model structure that were necessary in order to adequately describe the distribution kinetics of

25   perchlorate and iodide. The loss of iodide and perchlorate in the milk results in much faster

26   clearance rates of the anions from the dam. Studies also suggest that the loss of iodide to the

27   mammary gland and milk decreases the iodide available for the maternal thyroid (Brown-Grant,

28   1961; Yu, 2000; Yu et al., 2000). The thyroidal maximum capacities are lower in the lactating

29   and pregnant dam than in the male rat. Model parameterization in the male rat indicated the need

30   for Vmaxc values for uptake into the follicle of the thyroid of 2.2×10^3 L/hr-kg for perchlorate

31   and 5.5×10^4 L/hr-kg for iodide while the gestation model required values of 1.5×10^3 L/hr-kg
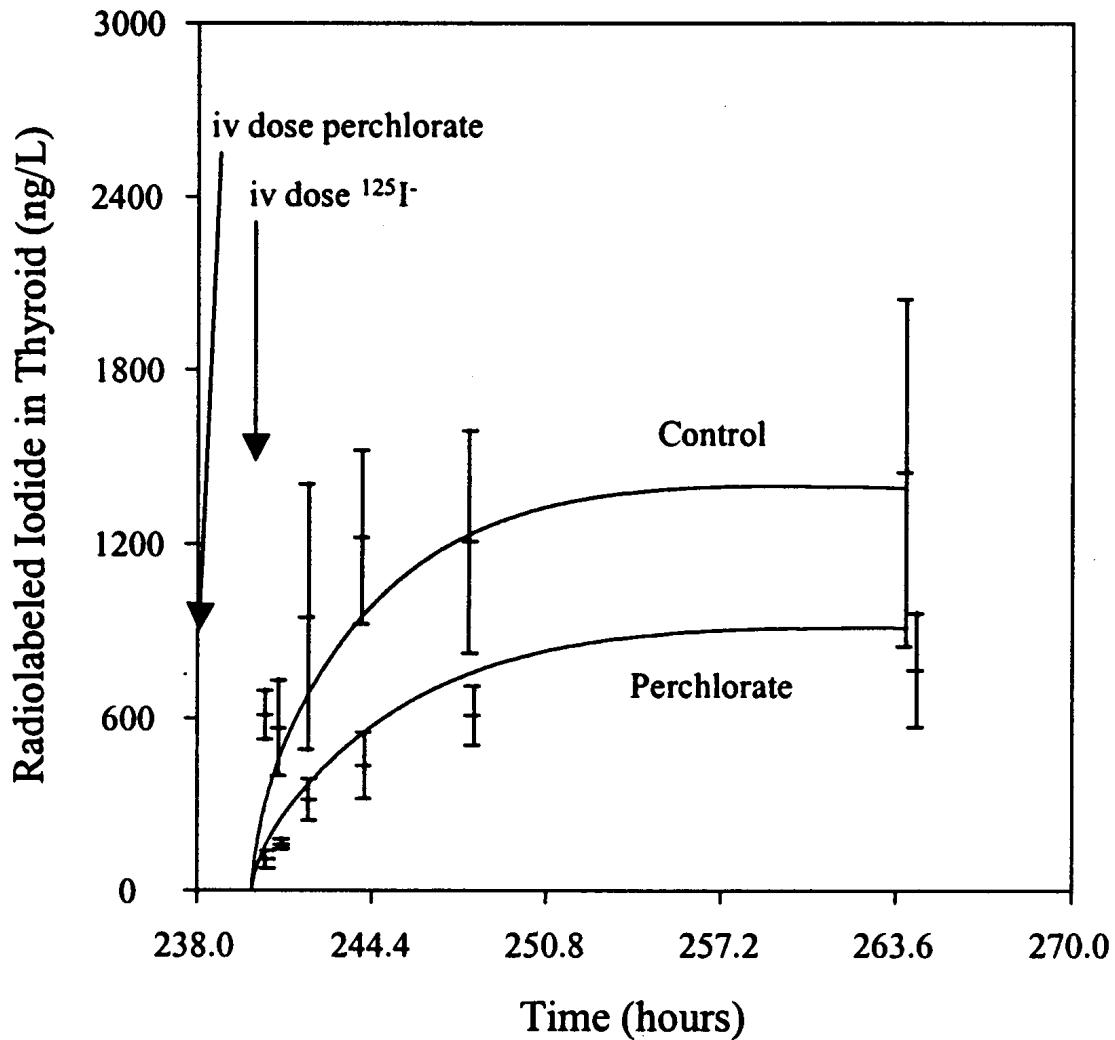
Figure 6-44. Validation for lactating dam and neonatal rat PBPK model (Clewell, 2001b). Model predictions (lines) versus data time course (mean ± SD) of [125]I radiolabeled iodide in the maternal thyroid with and without an iv dose of perchlorate at 1.0 mg/kg perchlorate 2 hours prior to an iv dose of 2.10 ng/kg [125]I to the dam on PND10. The top simulation indicates the control thyroid and the lower indicates the inhibited thyroid. Data of Yu (2000) and Yu et al. (2000).

1   and 4.0 x 10⁴ L/hr-kg for the same parameters. This difference is supported in the literature.

2   Versloot et al. (1997) suggest that the pregnant rat may have a lowered reserve of iodide in the

3   thyroid toward the end of pregnancy, causing increased activity in the thyroid. This may also be

4   true in the lactating rat. The skin of the lactating dam also required a smaller value for Vmaxc
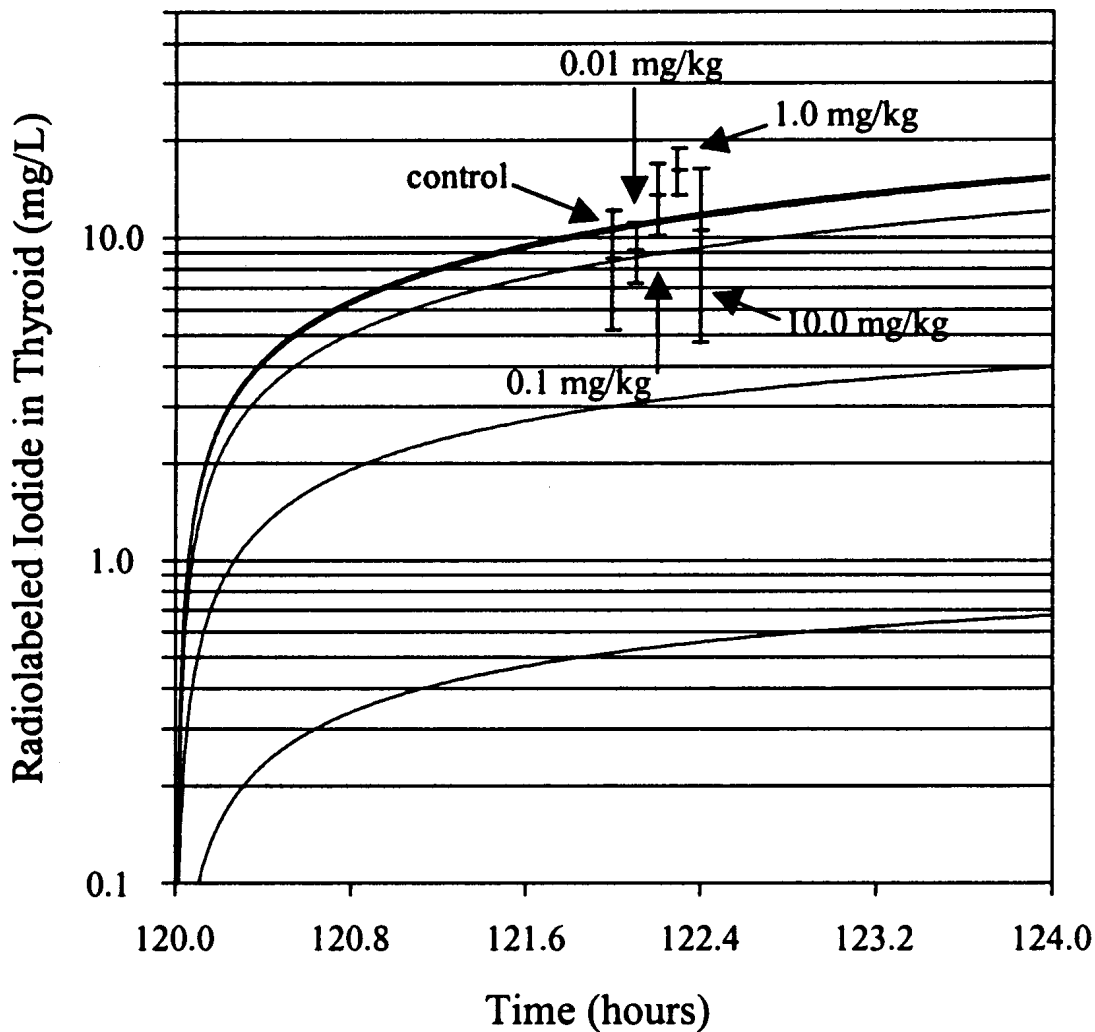
Figure 6-45. Validation for lactating dam and neonatal rat PBPK model (Clewell, 2001b). Model predictions (lines) versus data time course (mean ± SD) of [125]I⁻ radiolabeled iodide in the maternal thyroid on PND5 after 23 days dosing with perchlorate in drinking water at 0.0, 0.1, 1.0, and 10.0 mg/kg-day. All experimental data were taken two hours post-dosing. Data of Yu et al. (2000).

1   than the male rat. This is supported by the work of Brown-Grant and Pethes (1959), who

2   reported higher levels of iodide in the skin of male rats than in female rats. Skin, therefore,

3   appears to be a more important iodide reserve in the male rat than the female.

4   The described PBPK lactation model is able to predict the distribution of perchlorate in the

5   tissues of active uptake and serum of the lactating dam and neonate on PND5 and PND10 after

6   exposure to perchlorate in drinking water. Perchlorate distribution in this dynamic system is

1   described utilizing a pharmacokinetic approach to the modeling and accounting mathematically

2   or physiological changes, such as changing tissue volumes and maternal and neonatal growth.

3   The model predicts the transfer of perchlorate to the neonate and is also able to describe the

4   uptake into tissues of interest in the neonate, such as the GI contents and skin; however, the EPA

5   believes that both the maternal and neonatal serum fits could be improved. This may already be

6   accomplished with the additional data to which Clewell (2001b) alludes or, as noted previously,

7   the radionuclide modeling efforts of the ICRP (2001, 1989) may be informative.

8   The kinetic behavior of iodide is well described with the existing model, in spite of the

9   physiological complexity of the described system. The dam and neonate were accurately

10   simulated at a range of doses that spans four orders of magnitude (2.10 to 33,000 ng/kg) between

11   days 1 and 18 of lactation. The active sequestration of iodide in maternal and neonatal tissues

12   and the transfer of iodide between mother and neonate was described kinetically with the model;

13   data have been simulated at a variety of doses and at various time points up to 14 days after

14   exposure. The fact that the model was able to simulate data from other laboratories under a

15   variety of different conditions attests to the validity of the model structure and its applicability to

16   other studies. This also provides greater confidence in the model structure.

17   The clear differences between the perchlorate data from iv and drinking water studies draw

18   attention to unresolved issues in the transfer kinetics of perchlorate. Although lactational transfer

19   has long been studied, the transport mechanisms of this ion have yet to be elucidated in the

20   literature. A second transporter has been identified in the mammary gland, which actively

21   transports anions against the chemical gradient. However, the relationship of this transporter and

22   the anion concentration resulting from prolonged exposure to the high doses of perchlorate used

23   in these studies is not known. Clewell (2001b) suggests that it is possible that the high anion

24   load resulting from the long-term exposure to perchlorate may have resulted in decreased

25   transport of the ion. It is feasible that the movement of iodide may be regulated in the mammary

26   tissue, because the ion is vital to the development of the newborn. The data obtained between

27   the acute and drinking water studies suggest that a feedback mechanism is in place, because the

28   model over-predicts the milk transfer in the drinking water data when the acute parameters are

29   used. Clewell (2001b) notes in-house experiments that may help resolve these issues are

30   currently underway. Additional data were provided by Yu (2002), but is not clear that all these

31   data have been provided to the Agency or how these will be used to improve the modeling effort.

6.5 APPLICATION OF PBPK MODEL STRUCTURES TO INTERSPECIES EXTRAPOLATION

As discussed in the introduction to this chapter, the purpose of developing the proposed PBPK model structures was to aid interspecies extrapolation. All of the proposed model structures adequately describe both perchlorate and iodide distributions as evidenced by the fit of the model predictions against the experimental data shown in the preceding sections of this chapter. The degree of confidence in the model descriptions differed for the acute (iv) versus chronic (drinking water) data to some degree in the laboratory animals. A rather large degree of intersubject variability was evident among the human subjects, but in general the structures are accepted as quite sound and informative to the task.

The models do not link the perchlorate and iodide kinetics to perturbations in thyroid hormone. The existing data and current structures were not designed to address the complex issues involved with hormone homeostasis of the hypothalamic-pituitary-thyroid feedback axis as described in Chapter 3 or illustrated in the beginning of this chapter. Such a model would need to incorporate the hormone levels in tissues and serum and processes such as hormone production, storage, and secretion in the thyroid; conversion of T4 to T3 in the tissues; deiodination of T4 and T3 to less active forms and a feedback mechanism between the hormone levels, TSH, and the thyroid NIS. Kohn et al. (1996) developed a PBPK model that attempts to describe the effect of dioxin on thyroid hormones. Although perchlorate and dioxin act on the endocrine system through different modes of action, it is likely that a similar approach to that of Kohn et al. (1996) would be required to begin to address the hormone feedback system in the case of perchlorate. Parameterization and validation of such a model system would take a significant number of additional studies.

Nevertheless, the model structures as they exist currently are useful, particularly when employed in the conceptual framework proposed in Section 6.1. Because the models predict perchlorate and iodide kinetics, two relevant dose metrics to the mode of action can be evaluated: (1) the area under the curve (AUC) of perchlorate in the serum and (2) the degree (expressed as a % of baseline) of iodide uptake inhibition in the thyroid.

Because developmental effects are of concern, an argument could be made that peak and not AUC is the appropriate dose metric–the rationale being that any transient dose could be responsible for permanent deficits. However, the AUC values, as opposed to peak

1 concentrations, were used based on the assumption that these dose metrics would represent an

2 averaging of the serum and thyroid perchlorate concentrations and would be better correlated

3 with the inhibition effect on iodide uptake. The correlation was shown to be good between the

4 AUC and the degree of inhibition (see Section 6.5.2). Further, due to the rapid phase of

5 distribution after an iv dose, measurements of concentrations are very difficult to attain

6 experimentally and are more variable. Using simulated peak concentrations after iv injections is

7 potentially problematic due to the inexact modeling of the actual distribution of dose in the tail-

8 vein volume and the exact time of mixing in the whole blood compartment (Merrill, 2001e).

9 It was also observed by EPA that the ratios for peak perchlorate serum values (Merrill, 2001e;

10 Table 6) were in good agreement with those for the perchlorate serum AUC and that the serum

11 AUC were slightly more conservative if really different at all at the lower doses of concern to the

12 risk assessment.

13   The perchlorate AUC concentration in the thyroid was also considered, but the EPA and

14 AFRL/HEST agreed that this was a less satisfactory dose metric based on a number of

15 considerations. These included the following: that the thyroid Vmaxc estimates had to be

16 adjusted to account for upregulation of the NIS, but that this adjustment was more an empirical

17 exercise than a true biological model (since the hormone changes discussed above regulate the

18 NIS); that the thyroid concentrations were not actually measured in the fetus and neonate so that

19 verification of the parameters was not possible; and that the effects of perchlorate are related to

20 its effects on the NIS and secondary impact on thyroid hormone economy rather than to the

21 concentrations in the gland itself. Results of a sensitivity analysis on the adult male rat model

22 structure supported these conclusions (Merrill, 2001e). The results of the sensitivity analysis will

23 be discussed in Section 6.5.1. Thus, the models were exercised to develop human equivalent

24 exposure (HEE) estimates based on internal perchlorate concentration and iodide uptake

25 inhibition, both components of exposure in the proposed EPA model (Merrill, 2001e). The

26 purpose of Section 6.5.2 is to describe the modeling exercises underlying the HEE estimates that

27 are used in Chapter 7.

28

29 **6.5.1 Sensitivity Analysis of Proposed Adult Male Rat Model**

30   A sensitivity analysis was performed on the adult male rat model of Merrill (2001c) in

31 order to determine which parameters had the most significant impact on serum and thyroid AUC

1   perchlorate concentrations. All chemical specific kinetic parameters were increased individually

2   by 1% from the original, optimized values. The model-predicted dosimetrics were recalculated

3   after each change to determine the effect on the AUC estimates. This exercise was performed at

4   the four-hour time point after iv dosing for the 0.1 and 1.0 mg/kg-day doses. The equation

5   describing the calculation of the Sensitivity Coefficient value for each PBPK perchlorate

6   parameter tested is (Merrill, 2001e):

7

8   Sensitivity Coefficient = $\dfrac{(A - B)/B}{(C - D)/D}$,   (6-2)

9

10

11 where:

12

13   A = AUC for either serum or thyroid perchlorate with 1% increased parameter value,

14   B = AUC for either serum or thyroid perchlorate at initial parameter value,

15   C = Parameter value increase 1% over initial parameter value, and

16   D = Original initial starting parameter value.

17

18 Results are presented for the physiological parameters and chemical specific parameters

19 separately. Tables 6-9 and 6-10 provide the results for the 0.1 mg/kg-day dose, and

20 Tables 6-11 and 6-12 provide the results for the 1.0 mg/kg-day dose. The sensitivity coefficients

21 for the AUC estimates in both the thyroid and serum are provided and the changes in predicted

22 AUC estimates for the thyroid and serum are presented in the final two columns (Merrill, 2001e).

23   The sensitivity of serum and thyroid concentrations to model parameters is not linear.

24 At an iv dose level of 1.0 mg/kg, the model prediction of the AUC for serum ClO_4^- concentration

25 is most sensitive to urinary clearance (ClUc_p). A one percent increase in this value, from

26 0.07 to 0.0707 ng/hr-kg, causes a decrease in AUC serum ClO_4^- concentration from 4.69×10^5 to

27 4.63×10^5 ng, with a sensitivity coefficient of -1.271 (Table 6-12). Serum concentration is next

28 most sensitive to the rate ClO_4^- unbinds from plasma proteins (Clunbc_p), with a sensitivity

29 coefficient of -0.869 (Table 6-12).

30

TABLE 6-9. SENSITIVITY ANALYSIS FOR PHYSIOLOGICAL PARAMETERS IN THE ADULT MALE RAT MODEL AT 0.1 mg/kg PERCHLORATE (ClO₄⁻) DOSE (Merrill, 2001e)

Parameter[a]	Original Parameter Value	1% Increase in Parameter Value	AUC Thyroid Sensitivity Coefficient	AUC Serum Sensitivity Coefficient	Increase in AUC Thyroid ClO$_4^-$ (ng)[b]	Increase in AUC Serum ClO$_4^-$ (ng)[c]
BW	3.00E-01	3.03E-01	0.315	0.182	1.88E+06	9.95E+04
Blood Flows (fraction of cardiac output, QCc [L/hr])						
QCc	1.40E+01	1.41E+01	-0.005	-0.006	1.88E+06	9.94E+04
QTc	1.60E-02	1.62E-02	NS[d]	NS	1.88E+06	9.94E+04
QSKc	5.80E-02	5.86E-02	NS	-0.003	1.88E+06	9.94E+04
QGc	1.60E-02	1.62E-02	0.011	0.008	1.88E+06	9.94E+04
QLc	1.70E-01	1.72E-01	NS	NS	1.88E+06	9.94E+04
QKc	1.40E-01	1.41E-01	-0.016	-0.010	1.88E+06	9.93E+04
QFc	6.90E-02	6.97E-02	NS	NS	1.88E+06	9.94E+04
Tissue Volumes (fraction of body weight)						
Vplasc	4.10E-02	4.14E-02	0.155	0.079	1.88E+06	9.94E+04
VRBCc	3.30E-02	3.33E-02	0.192	0.109	1.88E+06	9.95E+04
Vttotc	7.70E-05	7.78E-05	0.187	0.113	1.88E+06	9.95E+04
VDTc	2.44E-01	2.46E-01	0.928	0.114	1.89E+06	9.95E+04
VTBc	1.57E-01	1.58E-01	0.203	0.114	1.88E+06	9.95E+04
VTc	6.00E-01	6.05E-01	0.453	0.114	1.88E+06	9.95E+04
VGc	5.40E-03	5.45E-03	0.197	0.112	1.88E+06	9.95E+04
VGJc	1.68E-02	1.70E-02	0.165	0.091	1.88E+06	9.94E+04
VGBc	4.10E-02	4.14E-02	0.197	0.114	1.88E+06	9.95E+04
VSkc	1.90E-01	1.92E-01	-0.053	-0.023	1.87E+06	9.93E+04
VSkBc	2.00E-02	2.02E-02	0.203	0.117	1.88E+06	9.95E+04
VLc	5.50E-02	5.56E-02	0.197	0.114	1.88E+06	9.95E+04
VKc	1.70E-02	1.72E-02	0.197	0.113	1.88E+06	9.95E+04
VFc	7.40E-02	7.47E-02	0.208	0.118	1.88E+06	9.95E+04

[a]Parameters as defined in Tables 6-1 and 6-2.
[b]AUC Thyroid Concentration using original parameters = 1.88E+06 ng ClO₄⁻.
[c]AUC Serum Concentration using original parameters = 9.94E+04 ng ClO.
[d]NS = sensitivity coefficient less than 0.001.

TABLE 6-10. SENSITIVITY ANALYSIS FOR CHEMICAL SPECIFIC PARAMETERS IN THE ADULT MALE RAT MODEL AT 0.1 mg/kg PERCHLORATE (ClO$_4^-$) DOSE (Merrill, 2001e)

Parameter[a]	Original Parameter Value	1% Increase in Parameter Value	AUC Thyroid Sensitivity Coefficient	AUC Serum Sensitivity Coefficient	Increase in AUC Thyroid ClO$_4^-$ (ng)[b]	Increase in AUC Serum ClO$_4^-$ (ng)[c]
Iodide Tissue/Blood Partition Coefficients						
PS_p	3.10E-01	3.13E-01	0.149	0.085	1.88E+06	9.94E+04
PR_p	5.60E-01	5.66E-01	0.192	0.111	1.88E+06	9.95E+04
PK_P	9.90E-01	1.00E+00	0.192	0.111	1.88E+06	9.95E+04
PL_p	5.60E-01	5.66E-01	0.187	0.108	1.88E+06	9.95E+04
PG_p	1.80E+00	1.82E+00	0.160	0.088	1.88E+06	9.94E+04
PGJ_p	2.30E+00	2.32E+00	0.165	0.090	1.88E+06	9.94E+04
PT_p	1.30E-01	1.31E-01	1.184	0.113	1.90E+06	9.95E+04
PDT_p	7.00E+00	7.07E+00	0.928	0.114	1.89E+06	9.95E+04
PF_p	5.00E-02	5.05E-02	0.197	0.114	1.88E+06	9.95E+04
PSk_p	7.00E-01	7.07E-01	11.154	6.024	2.08E+06	1.05E+05
PRBC_p	8.00E-01	8.08E-01	11.324	6.112	2.09E+06	1.05E+05
PS_p	3.10E-01	3.13E-01	0.149	0.085	1.88E+06	9.94E+04
PR_p	5.60E-01	5.66E-01	0.192	0.111	1.88E+06	9.95E+04
PK_P	9.90E-01	1.00E+00	0.192	0.111	1.88E+06	9.95E+04
PL_p	5.60E-01	5.66E-01	0.187	0.108	1.88E+06	9.95E+04
PG_p	1.80E+00	1.82E+00	0.160	0.088	1.88E+06	9.94E+04
PGJ_p	2.30E+00	2.32E+00	0.165	0.090	1.88E+06	9.94E+04
PT_p	1.30E-01	1.31E-01	1.184	0.113	1.90E+06	9.95E+04
PDT_p	7.00E+00	7.07E+00	0.928	0.114	1.89E+06	9.95E+04
PF_p	5.00E-02	5.05E-02	0.197	0.114	1.88E+06	9.95E+04
PSk_p	7.00E-01	7.07E-01	11.154	6.024	2.08E+06	1.05E+05
PRBC_p	8.00E-01	8.08E-01	11.324	6.112	2.09E+06	1.05E+05
Perchlorate Active Uptake Parameters - Vmaxc (ng/hr-kg BW) Km (ng/L)						
Vmaxc_Tp	2.90E+03	2.93E+03	47.830	6.088	2.77E+06	1.05E+05
Km_Tp	2.50E+05	2.53E+05	45.154	6.090	2.72E+06	1.05E+05
Vmaxc_DTp	1.00E+05	1.01E+05	55.875	6.081	2.92E+06	1.05E+05
Km_DTp	1.00E+08	1.01E+08	55.673	6.081	2.92E+06	1.05E+05
Vmaxc_Gp	1.00E+04	1.01E+04	55.769	6.080	2.92E+06	1.05E+05
Km_Gp	2.00E+05	2.02E+05	55.774	6.081	2.92E+06	1.05E+05
Vmaxc_Sp	6.50E+05	6.57E+05	54.713	5.678	2.90E+06	1.05E+05
Km_Sp	2.00E+05	2.02E+05	55.060	5.811	2.91E+06	1.05E+05

TABLE 6-10 (cont'd). SENSITIVITY ANALYSIS FOR CHEMICAL SPECIFIC PARAMETERS IN THE ADULT MALE RAT MODEL AT 0.1 mg/kg PERCHLORATE (ClO_4^-) DOSE (Merrill, 2001e)

Parameter[a]	Original Parameter Value	1% Increase in Parameter Value	AUC Thyroid Sensitivity Coefficient	AUC Serum Sensitivity Coefficient	Increase in AUC Thyroid ClO_4^- (ng)[b]	Increase in AUC Serum ClO_4^- (ng)[c]
Perchlorate Plasma Binding Parameters						
Vmaxc_B*p*	9.50E+03	9.60E+03	54.857	6.417	2.90E+06	1.06E+05
km_B*p*	1.10E+04	1.11E+04	54.916	5.590	2.91E+06	1.05E+05
Kunbc_*p*	1.00E-01	1.01E-01	54.948	5.096	2.91E+06	1.04E+05
Perchlorate Urinary Clearance and Permeability Area Cross Products (L/hr-kg)						
ClUc_*p*	7.00E-02	7.07E-02	54.047	5.399	2.89E+06	1.05E+05
PAGc_*p*	8.00E-01	8.08E-01	54.905	5.752	2.91E+06	1.05E+05
PAGJc_*p*	8.00E-01	8.08E-01	54.905	5.752	2.91E+06	1.05E+05
PATc_*p*	5.00E-05	5.05E-05	23.273	5.776	2.31E+06	1.05E+05
PADTc_*p*	1.00E-02	1.01E-02	24.398	5.775	2.33E+06	1.05E+05
PASKc_*p*	4.00E-01	4.04E-01	3.759	-4.354	1.95E+06	9.50E+04
PARBCc_*p*	1.00E-01	1.01E-01	3.455	-4.508	1.94E+06	9.49E+04

[a]Parameters as defined in Tables 6-1 and 6-2.
[b]AUC Thyroid concentration using original parameters = 1.88E+06 ng ClO_4^-.
[c]AUC Serum concentration using original parameters = 9.94E+04 ng ClO_4^-.

1   The predicted AUC for total thyroid concentration at a dose level of 1.0 mg/kg-day is most

2   sensitive to changes in the maximum capacity of the thyroid colloid (Vmaxc_DTp). A one

3   percent increase in this value from 1.00×10^5 to 1.0110^5 ng/hr-kg results in an increase in AUC

4   thyroid concentration from 9.84×10^6 to 1.04×10^7 ng (Table 6-12). However, the AUC thyroid

5   concentration is almost equally sensitive to other parameters of saturable processes, including

6   Vmaxc, Km, and the permeability area cross product values of other saturable tissues.

7   With a lower iv dose of 0.1 mg/kg, the blood serum concentration remains sensitive to

8   changes in urinary clearance, but demonstrates increased sensitivity to the parameters of

9   saturable compartments and effective partitioning with skin (PSk_*p*) and red blood cells

10   (PRBC_*p*). Serum concentration is most sensitive to the maximum capacity for plasma binding

11   (Vmaxc_B*p*) at this dose level (Table 6-10).

12   At the lower dose level of 0.1 mg/kg, thyroid concentrations show a similar sensitivity to

13   parameters of saturable processes, including plasma binding, permeability area cross products,

**TABLE 6-11. SENSITIVITY ANALYSIS FOR PHYSIOLOGICAL
PARAMETERS IN THE ADULT MALE RAT MODEL
AT 1.0 mg/kg PERCHLORATE (ClO$_4^-$) DOSE (Merrill, 2001e)**

Parameter[a]	Original Parameter Value	1% Increase in Parameter Value	AUC Thyroid Sensitivity Coefficient	AUC Serum Sensitivity Coefficient	Increase in AUC Thyroid ClO$_4^-$ (ng)[b]	Increase in AUC Serum ClO$_4^-$ (ng)[c]
BW	3.00E-01	3.03E-01	−5.944	−0.534	9.81E+06	4.67E+05
Blood Flows [fraction of cardiac output, QCc (L/hr)]						
QCc	1.40E+01	1.41E+01	−0.192	−0.014	9.84E+06	4.69E+05
QTc	1.60E-02	1.62E-02	0.021	NS[b]	9.84E+06	4.69E+05
QSKc	5.80E-02	5.86E-02	−0.085	0.001	9.84E+06	4.69E+05
QGc	1.60E-02	1.62E-02	0.128	0.005	9.84E+06	4.69E+05
QLc	1.70E-01	1.72E-01	0.021	NS	9.84E+06	4.69E+05
QKc	1.40E-01	1.41E-01	−0.234	−0.021	9.84E+06	4.69E+05
QFc	6.90E-02	6.97E-02	0.021	NS	9.84E+06	4.69E+05
Tissue Volumes (fraction of bodyweight)						
Vplasc	4.10E-02	4.14E-02	−7.734	−0.701	9.80E+06	4.66E+05
VRBCc	3.30E-02	3.33E-02	−7.649	−0.691	9.80E+06	4.66E+05
VTtotc	7.70E-05	7.78E-05	−7.841	−0.683	9.80E+06	4.66E+05
VDTc	2.44E-01	2.46E-01	7.606	−0.683	9.87E+06	4.66E+05
VTBc	1.57E-01	1.58E-01	−7.500	−0.682	9.80E+06	4.66E+05
VTc	6.00E-01	6.05E-01	−2.322	−0.683	9.83E+06	4.66E+05
VGc	5.40E-03	5.45E-03	−7.649	−0.685	9.80E+06	4.66E+05
VGJc	1.68E-02	1.70E-02	−7.883	−0.710	9.80E+06	4.66E+05
VGBc	4.10E-02	4.14E-02	−7.628	−0.682	9.80E+06	4.66E+05
VSkc	1.90E-01	1.92E-01	−8.799	−0.829	9.80E+06	4.65E+05
VSkBc	2.00E-02	2.02E-02	−7.606	−0.680	9.80E+06	4.66E+05
VLc	5.50E-02	5.56E-02	−7.628	−0.683	9.80E+06	4.66E+05
VKc	1.70E-02	1.72E-02	−7.628	−0.685	9.80E+06	4.66E+05
VFc	7.40E-02	7.47E-02	−7.585	−0.676	9.80E+06	4.66E+05

[a]Parameters as defined in Tables 6-1 and 6-2.
[b]Original AUC Thyroid concentration = 9.84E+06 ng ClO$_4^-$.
[c]Original AUC Serum concentration = 4.69E+05 ng ClO$_4^-$.
[d]NS = sensitivity coefficient less than 0.001.

TABLE 6-12. SENSITIVITY ANALYSIS FOR CHEMICAL–SPECIFIC PARAMETERS IN THE MALE RAT MODEL AT 1.0 mg/kg PERCHLORATE (ClO$_4^-$) DOSE (Merrill, 2001e)

Parameter[a]	Original Parameter Value	1% Increase in Parameter Value	AUC Thyroid Sensitivity Coefficient	AUC Serum Sensitivity Coefficient	Increase in AUC Thyroid ClO$_4^-$ (ng)[b]	Increase in AUC Serum ClO$_4^-$ (ng)[c]
Perchlorate Tissue/Blood Partition Coefficients						
PS_p	3.10E-01	3.13E-01	−7.862	−0.728	9.80E+06	4.66E+05
PR_p	5.60E-01	5.66E-01	−7.649	−0.688	9.80E+06	4.66E+05
PK_P	9.90E-01	1.00E+00	−7.649	−0.688	9.80E+06	4.66E+05
PL_p	5.60E-01	5.66E-01	−7.670	−0.692	9.80E+06	4.66E+05
PG_p	1.80E+00	1.82E+00	−7.905	−0.714	9.80E+06	4.66E+05
PGJ_p	2.30E+00	2.32E+00	−7.883	−0.711	9.80E+06	4.66E+05
PT_p	1.30E-01	1.31E-01	12.911	−0.683	9.90E+06	4.66E+05
PDT_p	7.00E+00	7.07E+00	7.606	−0.683	9.87E+06	4.66E+05
PF_p	5.00E-02	5.05E-02	−7.628	−0.684	9.80E+06	4.66E+05
PSk_p	7.00E-01	7.07E-01	−8.885	−0.846	9.80E+06	4.65E+05
PRBC_p	8.00E-01	8.08E-01	−7.649	−0.691	9.80E+06	4.66E+05
Perchlorate Active Uptake Parameters - Vmaxc (ng/hr-kg BW), Km (ng/L)						
Vmaxc_Tp	2.90E+03	2.93E+03	12.890	−0.683	9.90E+06	4.66E+05
Km_Tp	2.50E+05	2.53E+05	−15.745	−0.682	9.76E+06	4.66E+05
Vmaxc_DTp	1.00E+05	1.01E+05	123.554	−0.687	1.04E+07	4.66E+05
Km_DTp	1.00E+08	1.01E+08	120.784	−0.687	1.04E+07	4.66E+05
Vmaxc_Gp	1.00E+04	1.01E+04	122.062	−0.687	1.04E+07	4.66E+05
Km_Gp	2.00E+05	2.02E+05	122.062	−0.687	1.04E+07	4.66E+05
Vmaxc_Sp	6.50E+05	6.57E+05	120.997	−0.806	1.04E+07	4.66E+05
Km_Sp	2.00E+05	2.02E+05	122.914	−0.641	1.04E+07	4.66E+05
Perchlorate Plasma Binding Parameters - Vmaxc (ng/hr-kg BW), Km (ng/L)						
Vmaxc_Bp	9.50E+03	9.60E+03	122.062	−0.500	1.04E+07	4.67E+05
km_Bp	1.10E+04	1.11E+04	122.062	−0.694	1.04E+07	4.66E+05
Kunbc_p	1.00E-01	1.01E-01	122.275	−0.869	1.04E+07	4.65E+05
Perchlorate Urinary Clearance and Permeability Area Cross Products (L/hr-kg)						
ClUc_p	7.00E-02	7.07E-02	115.031	−1.271	1.04E+07	4.63E+05
PAGc_p	8.00E-01	8.08E-01	122.275	−0.685	1.04E+07	4.66E+05
PAGJc_p	8.00E-01	8.08E-01	122.275	−0.686	1.04E+07	4.66E+05
PATc_p	5.00E-05	5.05E-05	100.969	−0.686	1.03E+07	4.66E+05
PADTc_p	1.00E-02	1.01E-02	120.784	−0.687	1.04E+07	4.66E+05
PASKc_p	4.00E-01	4.04E-01	123.341	−0.567	1.04E+07	4.67E+05
PARBCc_p	1.00E-01	1.01E-01	122.062	−0.687	1.04E+07	4.66E+05

[a]Parameters as defined in Tables 6-1 and 6-2.
[b]Original AUC Thyroid concentration = 9.84E+06 ng ClO$_4^-$.
[c]Original AUC Serum concentration = 4.69E+05 ng ClO$_4^-$.

1   and urinary clearance. However, the predicted thyroid concentrations at both dose levels (1.0 and

2   0.1 mg/kg) are most sensitive to a change in Vmax_DT*p*. The Vmax values of the thyroid were

3   established by empirically fitting thyroid radioiodide and perchlorate uptake from several data

4   sets ranging in three orders of magnitude

5

6.5.2 Derivation of Human Equivalent Exposure Estimates

7   As discussed, the following internal dosimetrics were chosen to represent output from each

8   of the PBPK models: area under the curve (AUC) perchlorate concentrations in serum and

9   thyroid; peak serum and thyroid perchlorate concentrations; the total amount of perchlorate

10 excreted in the urine; the AUC for the lactational and placental transfer of perchlorate; and the

11 percent inhibition of iodide uptake into the thyroid. In order to explore the dose-response

12 elationship of these values, the target dosimetrics were evaluated across several doses in both

13 acute and sub-chronic exposure scenarios using previously developed PBPK models at the

14 AFRL/HEST; i.e., the models for the adult male rat (Merrill, 2001c) and human (Merrill, 2001d)

15 described in Section 6.2, the pregnant and fetal rat model (Clewell, 2001a) and the lactating and

16 neonatal rat model (Clewell, 2001b).

17   Acute (iv) pharmacokinetic studies in the adult male rat were used as the basis for this

18 dose-response analysis because iodide uptake inhibition could be correlated to perchlorate levels.

19 Further, as discussed in Section 6.1, the initial inhibition of iodide is viewed in the conceptual

20 model as the important step in the transient phase (Figure 6-2). Transient decrements in T4 can

21 result in permanent neurodevelopmental sequelae. In drinking water studies, upregulation of NIS

22 in the rat is so rapid that it resulted in no measurable thyroid iodide inhibition, so the iv doses

23 were used to estimate this initial insult. The target internal dosimetrics were first calculated in

24 each of the rat models for acute exposure to perchlorate (single *iv* administration) at doses of

25 0.01, 0.1, 1.0, 3.0, 5.0, 10.0, 30.0, and 100.0 mg/kg. In order to correlate perchlorate parameters

26 to data-validated inhibition, the 2 to 4 hr time-frame was used for all acute calculations. The

27 AUC for thyroid and serum were calculated by integrating predicted tissue concentrations from

28 2 to 4 hrs post dosing.

29   These same dosimetrics calculated for acute exposures were also determined for subchronic

30 (drinking water) perchlorate exposures at doses of 0.01, 0.1, 1.0, 3.0, 5.0, 10.0, 30.0, and

31 100.0 mg/kg-day. In order to achieve steady state concentrations, the models were run until the

1   predicted peak and trough heights did not change from one day to the next (Merrill, 2001e).

2   Serum and thyroid perchlorate AUC concentrations were then determined over a 24 hr period

3   (240-264 hrs in male, lactating, and neonatal rats; 480-504 hrs in pregnant and fetal rats).

4   Although the tissues reach steady state perchlorate concentrations within one week, the above

5   time-points were chosen in the lactation and gestation models for their ability to be verified with

6   data (Clewell, 2001a,b). The male rat model was run at the same time as lactation for the sake of

7   consistency with the other models. The total perchlorate AUC in the serum and thyroid were

8   determined from each the models at 240 and 264 hrs (or 480 and 504 hrs). The difference in the

9   two values was then divided by 24 hrs to give the AUC in units of ng/L-hr.

10   The AFRL/HEST experiments (Yu, 2000; Yu et al., 2000) have shown upregulation of the

11   NIS to be both time and dose-dependent. Thus, at lower doses, the rat thyroid was completely

12   upregulated after only a few days of drinking water exposure. Iodide uptake in the thyroid at

13   higher perchlorate doses ($\geq$10 mg/kg-day) was completely restored by the 18th day of exposure,

14   the time of data collection in the pregnant and fetal rats (Clewell, 2001).

15   Drinking water studies in the adult male rats showed elevated perchlorate uptake in the

16   thyroid at drinking water doses of 3.0 mg/kg-day and higher (Yu et al., 2000; Merrill et al.,

17   2001c). Increased perchlorate uptake also results from upregulation of NIS. Since perchlorate is

18   transferred into the thyroid via NIS, the inhibiting anion is "upregulated" along with iodide.

19   In order to simulate increased perchlorate concentrations in thyroids of the 3.0, 10.0, and

20   30.0 mg/kg-day dose groups, the original value for follicular Vmaxc (Vmaxc_Tp) was adjusted

21   to obtain the best fit of the model simulation to experimental data (Table 6-13). Since there were

22   no pharmacokinetic data available for the 5.0 and 100.0 mg/kg-day dose groups, values for

23   Vmaxc_Tp were estimated from a Michaelis-Menten fit to the adjusted Vmaxc's at 3.0, 10.0, and

24   30.0 mg/kg-day doses (Figure 6-46). Target dosimetrics in the male rat were calculated for both

25   originally optimized parameters and these adjusted ("upregulated") parameters.

26   This process of adjusting ("upregulating") the Vmaxc_Tp values was not necessary in the

27   gestation, lactation, or human models, as they were able to successfully describe perchlorate

28   concentrations in serum and thyroid at all measured doses (0.01 – 10.0 mg/kg-day in gestation

29   and lactation; 0.02 – 12 mg/kg-day in human) using one set of model parameters (Clewell,

30   2001a,b; Merrill, 2000). Merrill (2001e) posits that it was not necessary because it is likely that a

31   loss of maternal iodide to the fetus and neonate causes dams to exist in a chronic state of

TABLE 6-13. "UP-REGULATED" VALUES OF VMAXC_Tp^a AFTER PERCHLORATE DRINKING WATER EXPOSURE IN THE ADULT MALE RAT MODEL (Merrill, 2001e)

Drinking Water Dose (mg/kg-day)	Adjusted Vmaxc_Tp (ng/hr-kg)
0.01	2900
0.1	2900
1	2900
3	9000
5	17500[b]
10	32000
30	55000
100	79000[b]

[a]Maximum velocity capacity of active transport in the thyroid follicle.
[b]Data not available for these dose levels.
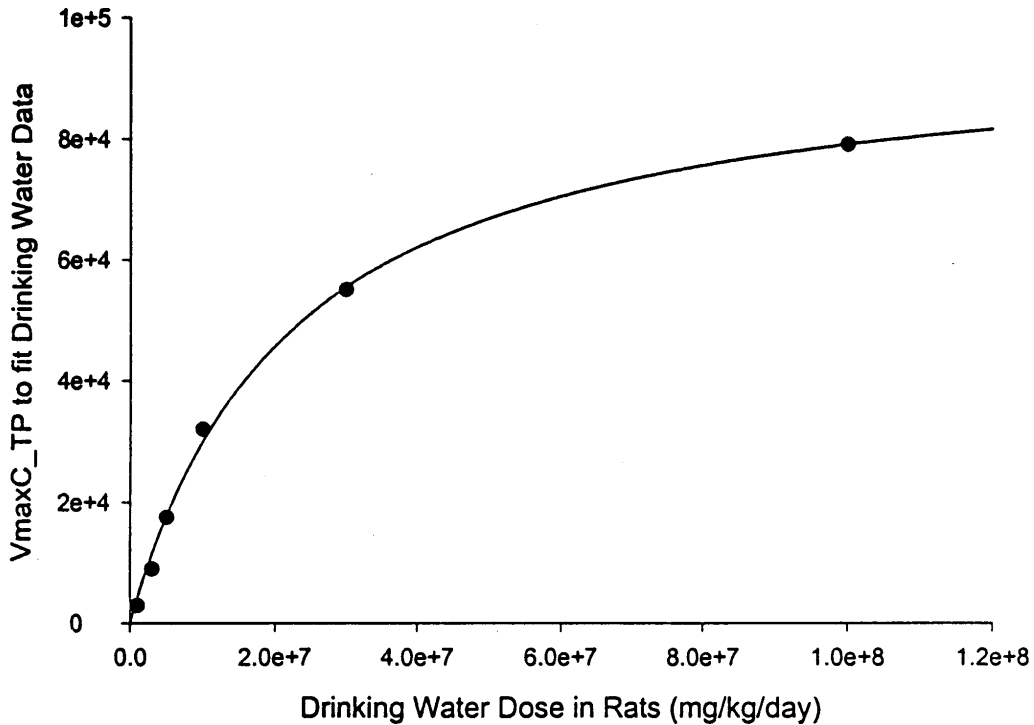
Figure 6-46. Upregulation of maximal capacity (ng/kg-hr) of active transport into the thyroid follicle for perchlorate (Vmaxc_Tp) optimized by fitting to drinking water data in the rat. Upregulation is first noted in the 3.0 mg/kg-day dose group.

1 thyroidal up-regulation. As a result, the effect of perchlorate on the thyroid was less dramatic

2 than in the male rat where a completely naïve system is perturbed by an inhibiting chemical.

3 Thus, the PBPK models for gestation and lactation were able to describe thyroid perchlorate

4 levels at drinking water doses from 0.01 to 10.0 mg/kg-day without adjusting the follicular

5 Vmaxc (Vmaxc_Tp) values with dose.

6   Increased follicular Vmaxc values were not needed to fit the human data likely due to the

7 larger size of the human thyroid colloid versus that of the rat and to the differences in plasma

8 protein binding discussed in Chapter 3.

9   The human PBPK model (Merrill, 2001d) was used to calculate all target dose metrics in

10 both acute and two-week drinking water perchlorate exposures in a 70 kg adult at doses of 0.01,

11 0.1, 1.0, 3.0, 5.0, 10.0, 30.0, and 100.0 mg/kg-day. Acute serum and thyroid perchlorate AUC

12 concentration estimates were calculated with the model over an eight hr time period (from 24 to

13 32 hrs post-exposure) in order to correlate perchlorate parameters to data-validated iodide

14 inhibition. For two-week drinking water exposures, the thyroid and serum perchlorate AUC

15 concentration estimates were calculated over a 24 hr period after serum and thyroid

16 concentrations reached steady state. The 240 to 264 hr time period was chosen for consistency

17 with the male rat model (Merrill, 2001c).

18   The adult human model (Merrill, 2001d) was also used to predict dosimetry in a 15 kg

19 child. The same dosimetrics were run in the model for the child and adult. However, since an

20 average child drinks less water than an adult (approximately 1 L/d as opposed to 2 L/d in the

21 adult), the actual exposures of a child and adult from the same water source would be different.

22 For example, a 15kg child consuming 1 L of contaminated water would receive a daily dose (per

23 kg bodyweight) that was 2.3 times that of a 70 kg adult consuming 2 L of water. Table 2 shows

24 the concentration of the drinking water required to deliver the same dose to a 15 kg child and a

25 70 kg adult. For the purpose of this paper, dosimetric comparisons were calculated using the

26 same dose (mg/kg-day) in the adult and child.

27   Figure 6-47 shows the curve generated from plotting the experimentally-determined percent

28 inhibition versus the corresponding PBPK-derived serum (A) and thyroid (B) perchlorate AUC

29 concentration estimates after acute (iv) exposure in rats. Thyroidal radiolabeled iodide (^{125}I$^-$)

30 uptake measurements were taken two hours after iv administration of perchlorate. The solid line

● - represents experimental data
◆ - represents PBPK model simulation values
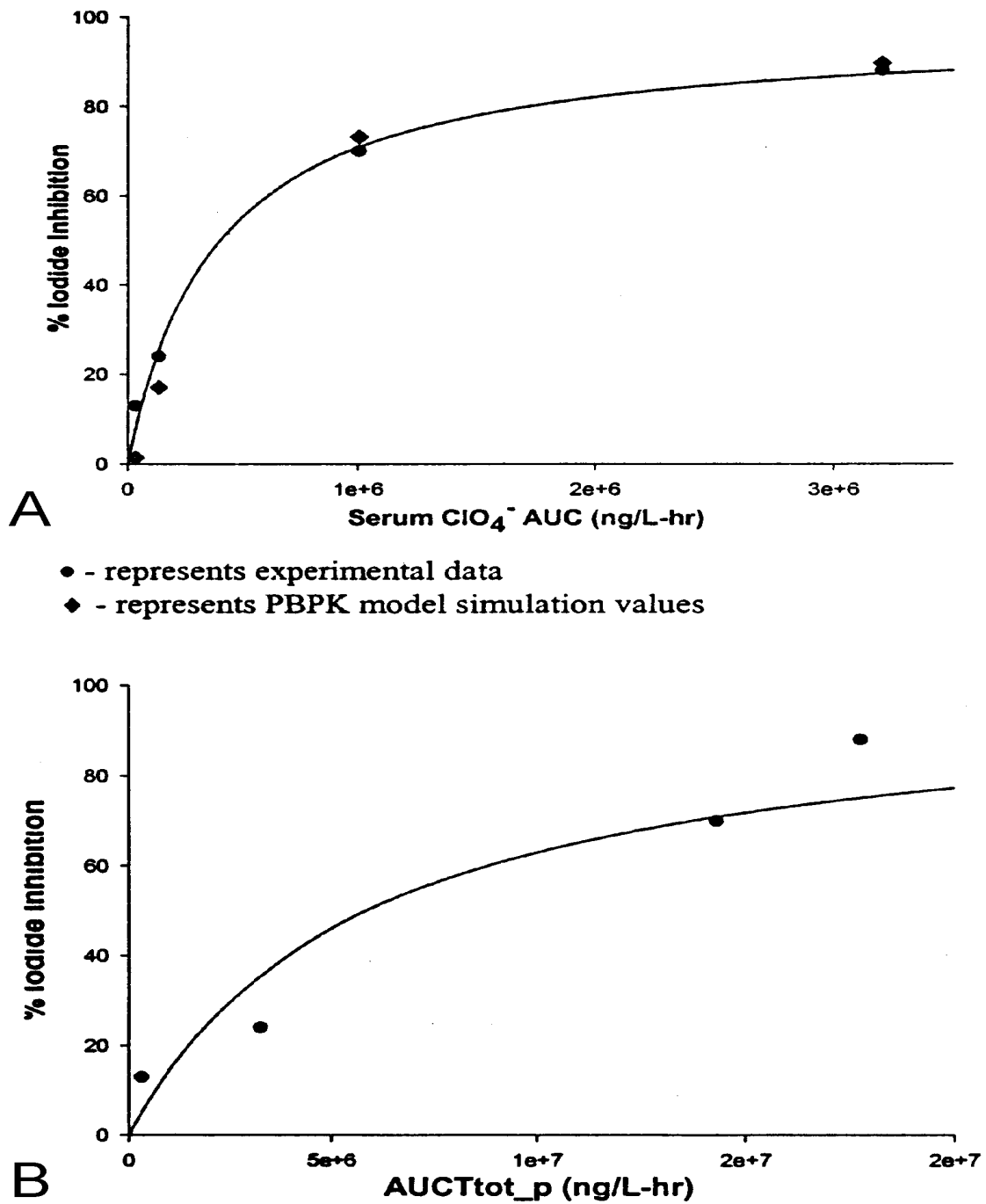
Figure 6-47. Michaelis-Menten fit of the "acute" male rat area under the curve (AUC) for serum (A) and thyroidal perchlorate (AUCTtot_p) in ng/L-hr. Model predictions and actual data shown for percent radiolabeled iodide uptake inhibition after iv injection of perchlorate.

1   represents a fit (not a PBPK model simulation) using the Michaelis-Menten type equation given

2   below:

3

4                    $$Y = (A \times AUC_{dose})/(AUC_{dose} + B) \qquad\qquad (6\text{-}3)$$

5

6   Where 'Y' represents the predicted percent inhibition of radioiodide uptake, 'A' represents the

7   maximal percent inhibition of radioiodide uptake, 'B' is related to the affinity of iodide uptake

8   based on serum concentration, and AUC_{dose} represents the AUC at each specific dose of

9   perchlorate. The above equation was also used to derive the dose-response relationship in

10   subsequent figures. The correlation coefficient (r^2) greater than 0.91 in all cases indicated

11   excellent fit for all (see Merrill, 2001e; Table 3).

12   Figure 6-48 shows the PBPK-derived AUC perchlorate concentration estimates for

13   drinking water exposure to the adult male rat versus the calculated percent inhibition of

14   radioiodide in the serum (A) and thyroid (B). The values for AUC of perchlorate concentration

15   in the serum were determined by running the adult male rat model (Merrill, 2001c) across doses.

16   Corresponding percent inhibitions were calculated by putting serum AUC perchlorate

17   concentration values into the equation from Figure 6-47. Human response (thyroid inhibition) to

18   subchronic exposure is similar to that of an acute exposure in the rat. This approach allows the

19   sub-chronic serum levels in the rat be related to iodide uptake in the native thyroid. The values

20   for AUC of thyroid perchlorate concentration (B) were determined by running the male rat model

21   (Merrill, 2001c) at steady state (between 240 and 264 hours of drinking water exposure) across

22   the doses shown. Corresponding percent inhibitions were calculated by putting thyroid AUC

23   values in the equation from Figure 6-47.

24   The actual human iodide inhibition data (Greer et al., 2000) were plotted as a function of

25   the perchlorate AUC concentration estimates for serum and thyroid calculated with the PBPK

26   model in Figure 6-49. The measured percent inhibition of radiolabeled iodide uptake in the

27   serum and thyroid on Day 2 of drinking water exposure to perchlorate is shown versus the

28   PBPK-derived estimates for human volunteers (both male and female). Inhibition data from time

29   points earlier than Day 2 of perchlorate in the human drinking water (Greer et al., 2000) and

30   inhibition data from acute perchlorate dosing in humans were not available. Therefore, the

31   inhibition measurements on Day 2 of perchlorate drinking water exposure were the closest-
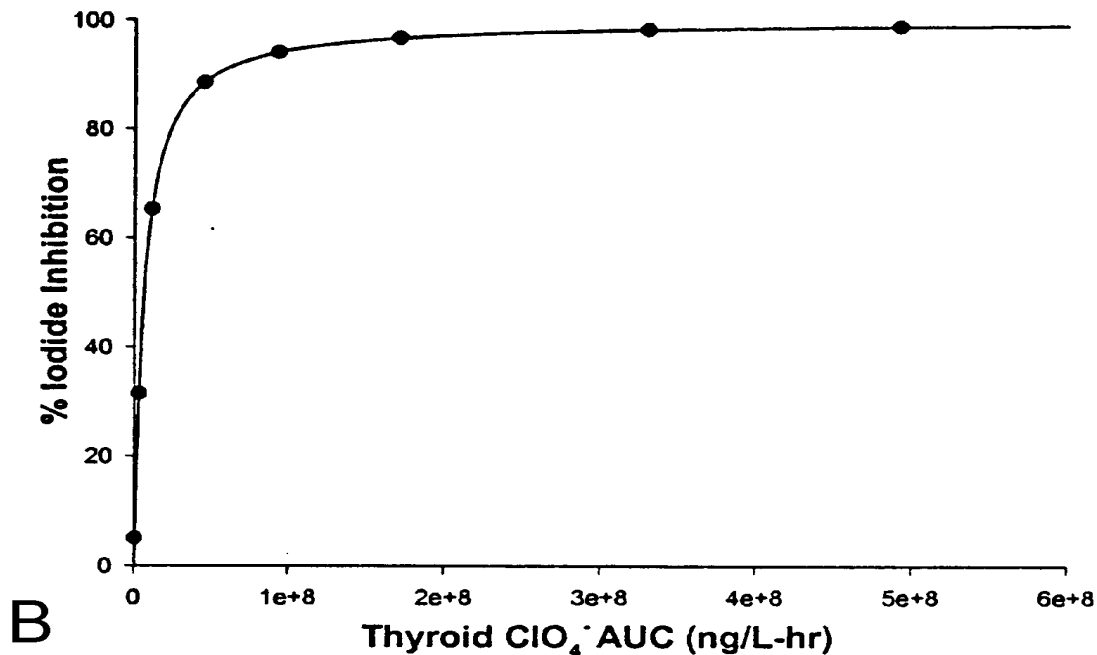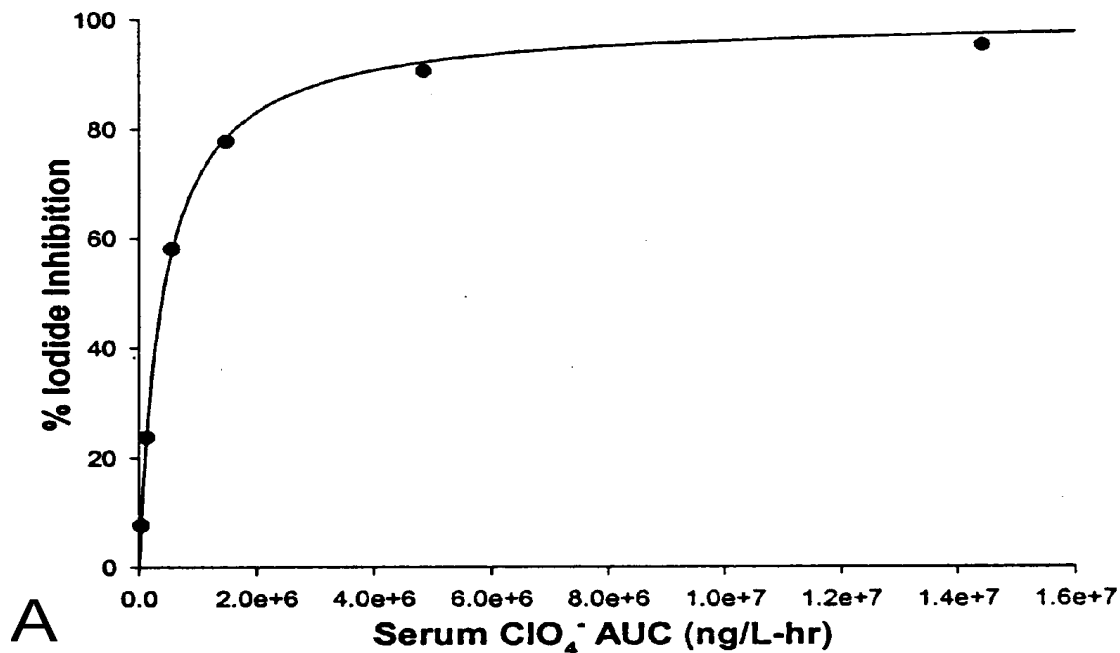
Figure 6-48. Michaelis-Menten fit of the "chronic" male rat area under the curve (AUC) for serum (A) and thyroidal (B) perchlorate (ng/L-hr). Model predictions and actual data shown for percent radiolabeled iodide uptake inhibition after drinking water exposure of perchlorate. Fit for serum calculated percent inhibition of radioiodide uptake calculated from equation used in Figure 6-47 (A) and for thyroid from Figure 6-47(B).
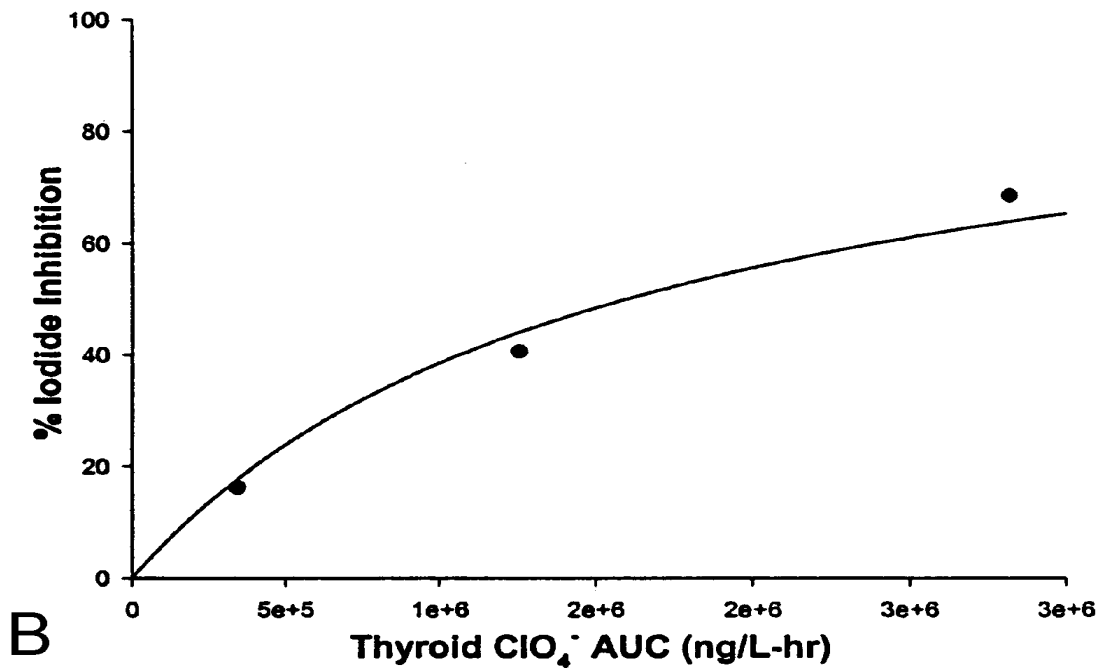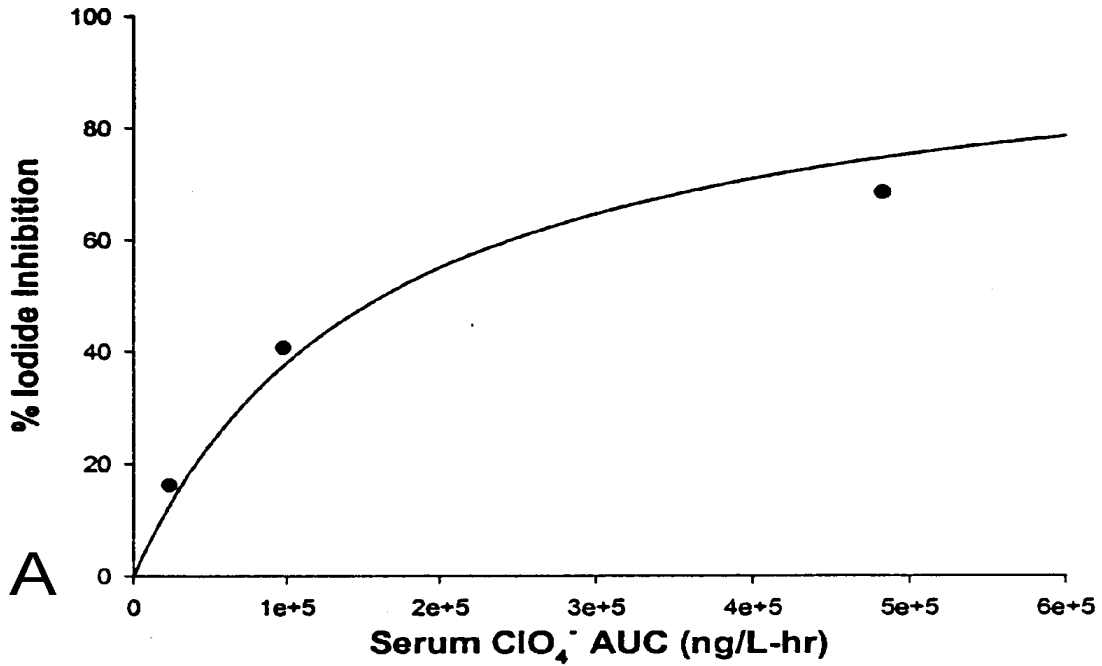
Figure 6-49. Michaelis-Menten fit of the human area under the curve (AUC) for serum (A) and thyroidal (B) perchlorate (ng/L-hr) on exposure Day 2. Model predictions and actual data shown for percent radiolabeled iodide uptake inhibition after drinking water exposure of perchlorate.

available representation of an acute human dose. Measured serum TSH and thyroid hormones indicated that thyroids were in normal homoeostatic state in human volunteers during the entire two week study (Merrill, 2001d).

The HEE estimates were calculated using the models as described in Section 6.1 (Figure 6-4). The HEE that would result in the same perchlorate AUC concentration estimates for serum (A) and thyroid (B) in the human and rat and the corresponding percent inhibition of iodide uptake is presented in Figure 6-50. Values for percent inhibition were determined from the rat serum AUC during drinking water exposures to perchlorate using the Michaelis-Menten equations from Figure 6-47. The correlation coefficient for both the serum and thyroid AUC versus percent iodide uptake inhibition relationship was 0.99.

6.5.3 Summary

The correlation coefficients for the dose-response relationships using the PBPK-model generated HEE estimates between serum and thyroid perchlorate AUC concentration versus iodide inhibition indicated good fits. Tables of the actual estimates and their ratios can be found in Merrill (2001e).

The rat serum ratios (AUC and peak concentrations) change significantly between 0.1 and 3.0 mg/kg-day due to binding of perchlorate by plasma proteins. Plasma binding is saturated at doses greater than 1.0 mg/kg-day. Male rat to human ratios are notably lower than those ratios between rats, as plasma binding of perchlorate occurs to a much lesser extent in humans.

HEE estimates were calculated for both a 15 and 70 kg human. The differences between the 15 and 70 kg human HEE estimates were never greater than 75%, indicating that body weight doesn't significantly affect the target dose metrics. Interestingly, the HEE estimates were greater in the 15 kg child. One might expect the adult and child HEE estimates to be nearly equal, given no parameters were changed in the human model except body weight. However, physiological parameters within the model are linearly scaled by body weight; whereas, chemical-specific parameters are scaled nonlinearly (e.g., as a multiple of body weight to a power of ¾). As indicated later in the sensitivity analysis, the internal dose metrics presented are more sensitive to chemical-specific parameters, especially those describing saturable kinetics. Therefore, the chemical-specific parameter values for the 15 kg child are proportionally greater
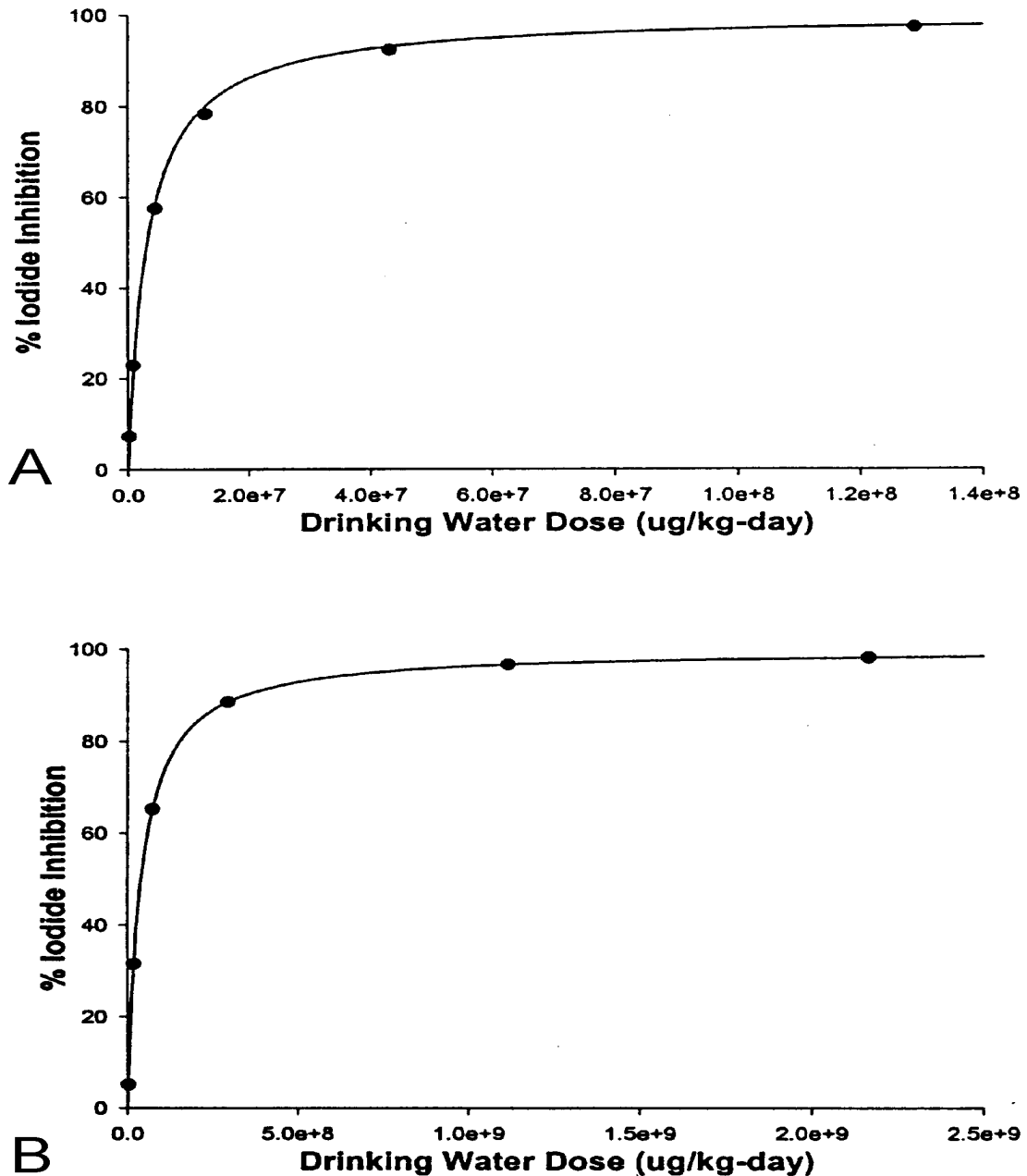
Figure 6-50. Michaelis-Menten fit of the human equivalent exposure (HEE) of perchlorate in drinking water derived from the area under the curve (AUC) for serum (A) or thyroid (B) versus percent predicted inhibition in the rat after an "acute" (iv) dose.

1   (in terms of body weight) than those of the adult. As a result, a slightly higher dose is required to

2   saturate these tissues in a child.

1   When comparing the dose metrics for serum versus thyroid, the HEE estimates calculated

2 from the thyroid were less than the HEE estimates calculated from the serum by a factor of 100 at

3 a 0.01 mg/kg-day dose level. This difference became a factor of 10 starting at the 5.0 mg/kg-day

4 concentration for the 15 kg child and at 10.0 mg/kg-day for the adult.

5   These considerations will be explored in Chapter 7 to develop dosimetric adjustment

6 factors for the observed effect levels.

7

7. DOSE-RESPONSE ASSESSMENTS FOR HUMAN HEALTH

The available database prior to initiation of the perchlorate testing strategy in 1997 (see Chapter 3) on the health effects and toxicology of perchlorate or its salts was very limited. The majority of human data were clinical reports of patients treated with potassium perchlorate for hyperthyroidism resulting from an autoimmune condition known as Graves' disease. Potassium perchlorate still is used diagnostically to test TSH, T3, and T4 production in some clinical settings. The primary effect of perchlorate is to decrease the production of thyroid hormones by competitively inhibiting iodide anion uptake into the thyroid at the *sodium (Na$^+$)-iodide (I$^-$) symporter (NIS)* and by causing a discharge of stored iodide from the thyroid gland.

It was difficult to establish a dose-response for the effects on thyroid function from daily or repeated exposures in healthy humans based on the data in patients with Graves' disease because of a variety of confounding factors, including that the disease itself has effects; that often only a single exposure and not repeated exposures were tested; that only one or two doses were employed; and that often the only effect monitored was iodide release from the thyroid or control of the hyperthyroid state. There were limited data in normal human subjects and laboratory animals that support the effect of perchlorate on thyroid hormones, but the majority of these studies suffer from the same limitations as those with the Graves' disease patients, with respect to the number of doses and exposures. These limitations prevent establishment of a quantitative dose-response estimate for the effects on thyroid hormones after long-term repeated exposures to perchlorate in healthy human subjects.

In addition, on December 14, 2001, after internal peer review of this document, the Agency articulated its interim policy on the use of third-party studies submitted by regulated entities (U.S. Environmental Protection Agency, 2001c). For these purposes, EPA is considering "third party studies" as studies that have not been conducted or funded by a federal agency pursuant to regulations that protect human subjects. Under the interim policy, the Agency will not consider or rely on any such human studies (third-party studies involving deliberate exposure of human subjects when used to identify or quantify toxic endpoints such as those submitted to establish a

1   NOAEL or NOEL for systemic toxicity of pesticides) in its regulatory decision making, whether

2   previously or newly submitted. Some of the clinical studies contained in this database fall in this

3   category of studies not to be considered. However, the scientific and technical strengths and

4   weaknesses of these studies were described before this Agency policy was articulated.

5   Therefore, because of the scientific shortcomings of these studies, they will not be used as

6   "principal studies" in the derivation of an RfD. The ethical issues surrounding the conduct of

7   these studies or their use for regulatory purposes in light of the Agency's interim policy will not

8   be discussed in this document. The Agency is requesting that the National Academy of Sciences

9   conduct an expeditious review of the complex scientific and ethical issues posed by EPA's

10   possible use of third-party studies which intentionally dose human subjects with toxicants to

11   identify or quantify their effects.

12   Thyroid hormone deficiencies, such as those induced by perchlorate, can affect normal

13   metabolism, growth, and development. However, no robust data existed previously with which

14   to evaluate potential target tissues or effects other than those in the thyroid. The data on the

15   thyroid effects were also insufficient for quantitative dose-response assessment. Additionally,

16   there were no data with which to evaluate the effects of perchlorate in potentially susceptible

17   populations, such as developing fetuses; nor were there data on the effects of perchlorate on the

18   reproductive capacity of male or female laboratory animals.

19   Benign tumors had been reported in the thyroids of male Wistar rats and female BALB/c

20   mice treated with repeated, high-dose exposures (2 years at 1,339 mg/kg-day and 46 weeks at

21   2,147 mg/kg-day, respectively) of potassium perchlorate in drinking water, establishing

22   perchlorate as a carcinogen. Benign tumors in the thyroid have been established to be the result

23   of a series of progressive changes that occur in the thyroid in response to interference with

24   thyroid-pituitary homeostasis (i.e., perturbation of the normal stable state of the hormones and

25   functions shared between these two related glands). This progression is similar regardless of the

26   cause of the thyroid hormone interference (Hill et al., 1989; Capen, 1997; Hurley et al., 1998).

27   EPA has adopted the policy that for the dose-response of chemicals that cause disruption in the

28   thyroid but that do not have genotoxic activity (i.e., cause damage to DNA or show other genetic

29   disruption) a threshold for carcinogenicity is to be based on precursor lesions (U.S.

30   Environmental Protection Agency, 1998e).

1   In the case of perchlorate, an overall model based on its mode of action has been developed

2   as shown in Figure 7-1. The model supports iodide inhibition as the key event that precedes the

3   hormone and thyroid changes with subsequent neurodevelopmental and neoplastic sequelae.

4   Focusing on the key event of iodide uptake inhibition allows a harmonized approach to both the

5   "noncancer" and "cancer" toxicity that occurs downstream along the continuum. Thus, one

6   harmonized risk estimate is derived for both sequelae based on their common mode of action.

7

8
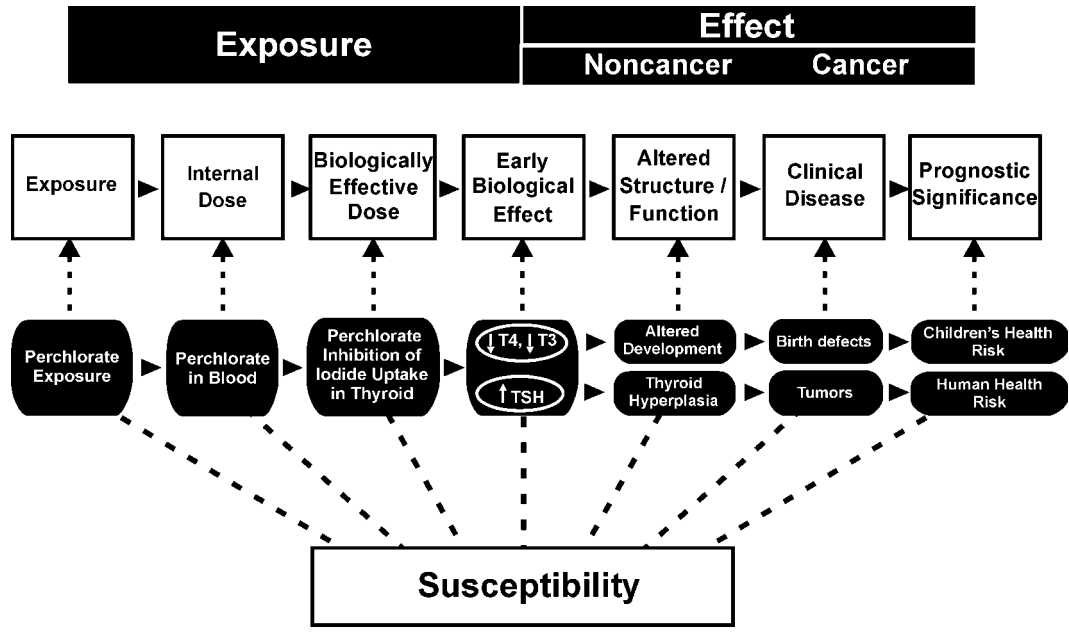
Figure 7-1. Mode-of-action model for perchlorate toxicity proposed by the U.S. EPA (U.S. Environmental Protection Agency, 1998d). Schematic shows the exposure-dose-response continuum considered in the context of biomarkers (classified as measures of exposure, effect, and susceptibility) and level of organization at which toxicity is observed (U. S. Environmental Protection Agency, 1994a; Schulte, 1989). The model maps the toxicity of perchlorate on this basis by establishing casual linkage or prognostic correlations of precursor lesions.

1   This chapter presents the synthesis of the most relevant data for deriving a revised

2   quantitative assessment of human health risk for perchlorate. The new data were consistent with

3   the limited historical characterization and the 1998 EPA assessment in that the anti-thyroid

1 effects remain the focus of concern and the key event of its mode of action remained identified as

2 the inhibition of iodide uptake at the NIS. However, data from the testing strategy allowed a

3 more comprehensive evaluation of the possible sequelae of the iodide uptake inhibition and its

4 thyroid-pituitary axis perturbations with respect to other endpoints, notably effects in dams and

5 their offspring and on nerurodevelopmental, reproductive, and immunotoxicity parameters.

6   The key event is defined as an empirically observable precursor step that is a necessary

7 element of the mode of action or is a marker for such an element. This will be discussed in

8 Section 7.1.1. Section 7.1.2 discusses dosimetric adjustment of effect levels observed in the

9 laboratory animals to human equivalent exposures (HEE). Choice of the point of departure for

10 the assessment based on a quantitative consideration of the key event, observed effects, and

11 weight of the evidence is discussed in Section 7.1.3. Application of factors to account for

12 uncertainty and variability in the extrapolations required to use the data is discussed in Section

13 7.1.4. The overall operational derivation is presented in Section 7.1.5, and the assignment of

14 confidence levels is discussed in Section 7.1.6. Section 7.1.5 also presents a discussion of the

15 cancer assessment in the context of the RfD. Section 7.2 discusses the inhalation reference

16 concentration. Susceptible population considerations are discussed in Section 7.1.5.3. Section

17 7.3 presents a brief summary of the findings.

18

7.1.1 Key Events and Weight of the Evidence

20   Results of the testing strategy have established that the critical target tissue for perchlorate

21 is the thyroid gland, with some remaining concern for adequate characterization of its potential

22 for immunotoxicity, notably contact hypersensitivity. Changes in thyroid weights, three response

23 indices of thyroid histopathology (colloid depletion, hypertrophy and hyperplasia), and thyroid

24 and pituitary hormones were consistently altered across the array of experimental designs

25 represented by the data base. The developmental and reproductive NOAEL and LOAEL values

26 were higher than those associated with thyroid toxicity per se.

27   Figure 7-2 highlights the temporal considerations that have to be superimposed on

28 evaluation of the data from the various studies in laboratory animals and humans in order to

29 characterize the anti-thyroid effects from perchlorate exposure. Conceptually, competitive

30 inhibition of iodide uptake at the NIS by perchlorate is the key event leading to both potential

31 neurodevelopmental and neoplastic sequelae. The decrement in iodide uptake leads to
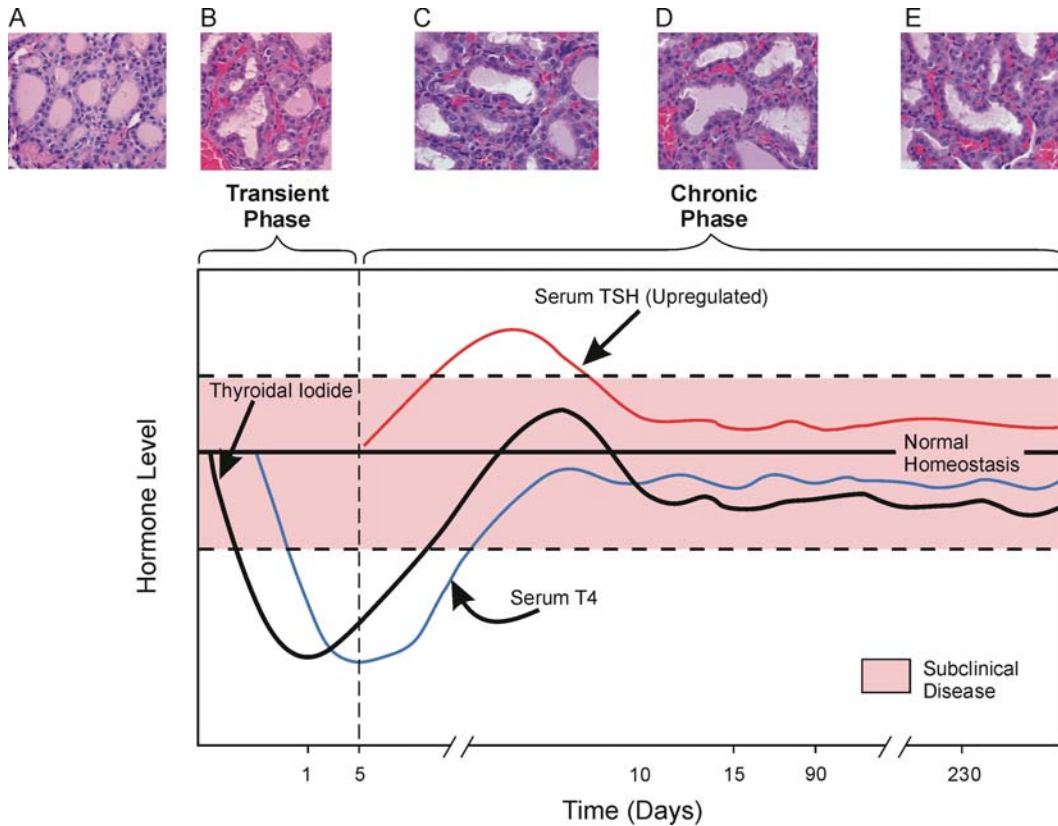
Figure 7-2. Schematic of thyroid and pituitary hormone levels with associated pathology after acute versus chronic dosing with perchlorate. The transient phase is represented by decreases in thyroidal iodide due to the inhibition by perchlorate at the NIS with subsequent drop in T4. The transient drops in T4 can lead to permanent neurodevelopmental sequelae. Once TSH is upregulated via the hypothalamic-pituitary-thyroid feedback, T4 appears to be in normal homeostasis but actually can represent subclinical or undiagnosed disease (hypothyroxinemia). The upregulation of TSH can result in neoplasia. Normal thyroid tissue is represented in Panel A. Panel B shows lace-like colloid depletion which is more pronounced in subsequent panels C, D and E. Panels D and E represent hypertrophy and hyperplasia.

subsequent drops in T4 (and T3) that can lead to permanent neurodevelopmental deficits. Corroborating evidence for this likely outcome given the mode of action of perchlorate comes from the iodide deficiency literature and recent studies showing that maternal hypothyroxinemia (i.e., decrements in T4 with or without concomitant increases in TSH) is linked to poor developmental, neuropsychological and cognitive outcomes (Haddow, et al., 1999; Pop et al.,

1   1999; Morreale de Escobar, et al., 2000). It should be noted that medical concern for

2   hypothyroxinemia remains in the "chronic phase"; i.e., once TSH upregulates to attempt to

3   regulate the hypothalamic-pituitary-thyroid feedback system back to an apparent homeostasis,

4   because this stress on the system essentially represents a "subclinical" disease state. Indeed,

5   adverse outcome in women with hypothyroxinemia per se has been demonstrated because

6   adversity includes the inability of an organism to respond to additional stressors. The system in

7   this case, particularly when considered on a population level, would present a diminished

8   capacity to compensate for other anti-thyroid insults. Since a large percentage of women are

9   believed to already be hypothyroid, the importance of this effect to women in general, pregnant

10   women, and fetuses on a population level can not be discounted. Weiss (2000) has noted that

11   even if the magnitude of effect may be relatively small for most environmental levels, such

12   neurotoxicity is extremely significant for public health.

13   Of notable concern, as previously discussed in Chapter 3, is that the developing fetus is

14   dependent on the mother for its T4 and T3 through parturition, as illustrated in Figure 7-3 for

15   humans with a similar pattern in rats. During the period illustrated in Figure 7-3, a number of

16   critical stages in neural development take place, some of which depend on thyroid hormones.

17   The cell precursors of the brain and spinal cord which compose the central nervous system

18   (CNS) begin to develop early in embryogenesis through the process called neurulation.

19   Beginning early in the second week of gestation in rodents (GD9.5 in rats) and the first month of

20   gestation in humans, specific areas of the CNS begin to form with the neurogenesis and

21   migration of cells in the forebrain, midbrain, and hindbrain. This sequence of developmental

22   processes includes proliferation, migration, differentiation, synaptogenesis, apoptosis, and

23   myelination (Rice and Barone, 2000). As discussed in Chapter 3, thyroid hormones play a role

24   throughout this process, regulating proliferation, migration, and differentiation. Alterations in

25   these processes can result in abnormalities of the brain and developmental delays.

26   The upregulation in TSH in the "chronic phase" (see Figure 6-2) also presents an increased

27   potential for neoplasia because stimulation of the thyroid to produce more T4 and T3 can result

28   in hyperplasia. Both the decrement in T4 and T3 and increase in TSH is shown in Figure 7-1 at

29   the same step along the continuum. Which of these thyroid responses is the most sensitive to

30   hormone changes has not specifically been studied in the perchlorate testing strategy. As noted

31   in the analyses of the studies in Chapter 5, there is a considerable degree of overlap among the
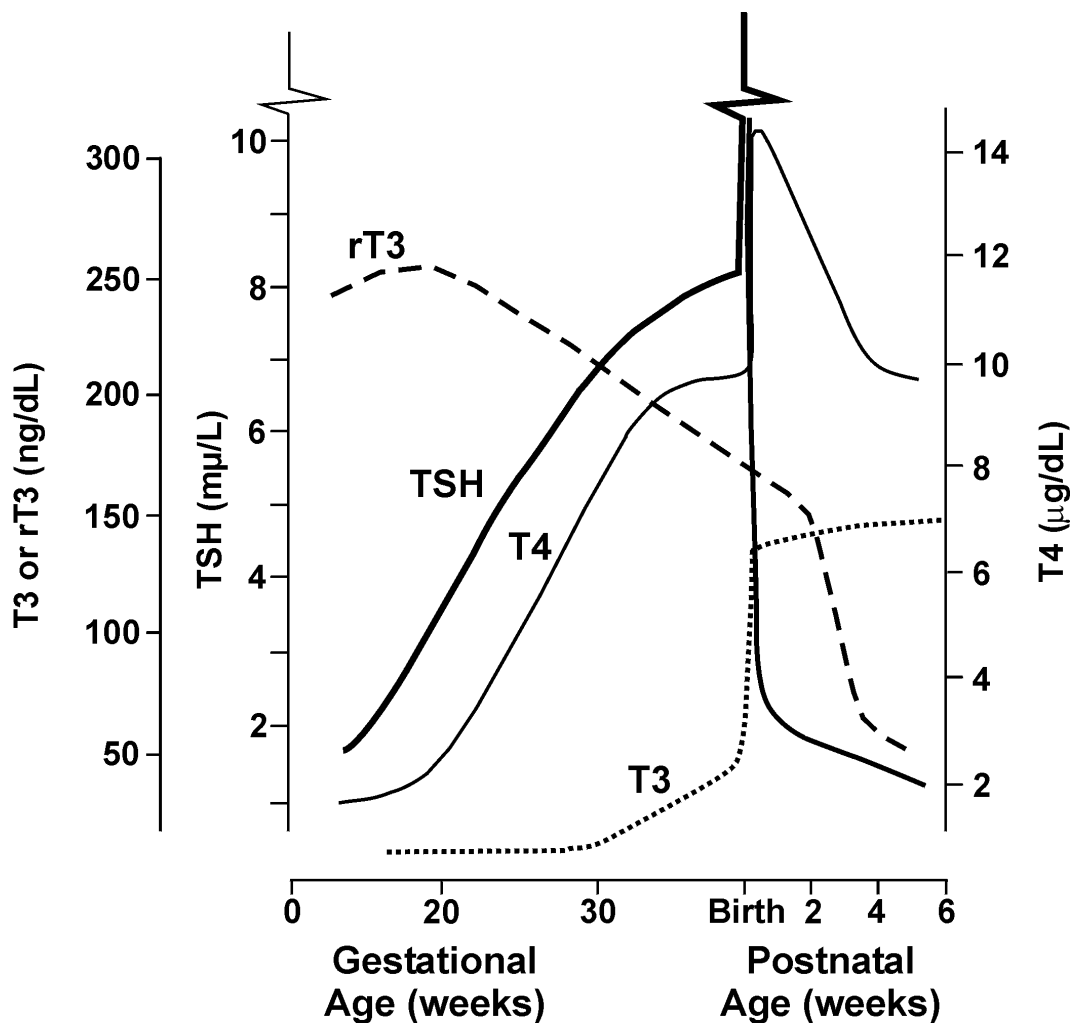
Figure 7-3. Pattern of change in fetal and neonatal thyroid function parameters during pregnancy and extrauterine adaptation in the human (from Fisher, 1996). A similar pattern is thought to exist in the rat (see text for further details).

1   three different diagnoses of thyroid histopathology: colloid depletion, hypertrophy, and

2   hyperplasia.

3   Colloid depletion does appear to be slightly more sensitive across the perchlorate studies.

4   The fact that thyroid follicular colloid depletion is a consistent finding not only across this study,

5   but in rodents in general, would suggest that it is a good indicator of sufficient exposure to inhibit

6   thyroid hormone synthesis. From a physiologic point of view this may be logical and supports

7   the mode-of-action model. If there is any reserve thyroid hormone in the colloid, it is depleted

1   before serum hormones are altered. Once serum levels are altered, TSH is upregulated and

2   hypertrophy and hyperplasia are initiated in an attempt by the gland to restore circulating levels

3   of T4 and T3. The diagnosis of colloid depletion has been reported with a similar compound,

4   sodium chlorate, in the rat (Hooth et al., 2001), with many other chemicals in the rat, and with

5   numerous goitrogens and pharmaceutical agents in the mouse. Colloid depletion in association

6   with hypertrophy and hyperplasia suggests sufficient dose of the compound to inhibit colloid

7   synthesis and decreases of circulating serum thyroid hormone levels sufficient to stimulate TSH.

8   Colloid depletion as the most sensitive indicator is most notable in the pups of the 2001

9   "Effects Study" on GD21 and then immediately post parturition on PND4. Alternatively, as

10 discussed in Chapter 5, it may have been harder to diagnose hypertrophy and hyperplasia in the

11 younger (smaller) and growing glands. The BMDL for colloid depletion increased with post-

12 natal age and by PND21, hyperplasia was also present. In contrast, all three thyroid indices were

13 present in the PND4 pups of the previous Argus Laboratories, Inc. (1998a) study. This may be

14 due to the difference in dosing of the dams. The dams in the 1998 study were only dosed during

15 gestation and, therefore, likely had a greater decrement in thyroid hormones. The dams in the

16 2001 study were dosed for two weeks during cohabitation, sufficient time as evidenced in the

17 data described in Chapter 6, for upregulation of NIS to compensate.

18   Other studies indicate that whichever index is most sensitive could be dependent on dose

19 spacing in the study, age of animals on test, and sacrifice time point. For example, hyperplasia

20 was the most sensitive of the three in the P2-generation adults (19 week F1-generation pups) and

21 these same pups developed thyroid adenomas.

22   The proposed mode of action mapped in Figure 7-1 is supported by correlations between

23 thyroid hormones and TSH and between thyroid hormones or TSH and an objective measure of

24 lumen size from laboratory animals exposed to ammonium perchlorate. There were positive

25 correlations between T3 and T4, and negative correlations between either T3 and T4 and TSH, as

26 expected based on the mode of action model (Appendix 7A). The positive correlation between

27 TSH and decreased follicular lumen size and negative correlation between T4 or T3 and

28 decreased follicular lumen size similarly support the proposed model (Appendix 7A). Some of

29 the correlations used in the 1998 assessment were precluded due to the limited severity scoring

30 system used by the PWG.

1   Additional support for the mode of action comes from data that now allow the linkage of

2   both neurodevelopmental and neoplastic sequelae into the model. These definitive data were not

3   available prior to the 1997 perchlorate testing strategy and especially not before the most recent

4   studies recommended by the 1999 external peer review. The repeat of observed effects on the

5   motor activity and brain morphometry results by new studies allowed definitive determination

6   that perchlorate exposure poses a neurodevelopmental hazard.

7   Repeatability and variability in statistics, sometimes a concern for evaluation of behavioral

8   assays (Cory-Slechta et al., 2001) were addressed by the Bayesian approach employed for the

9   motor activity analysis (Dunson, 2001a) that showed remarkable reproducibility between the two

10   studies despite the deficits previously noted for the Argus Research Laboratories, Inc. (1998a)

11   study. The effects on the size of the corpus callosum measurements were also repeated, and

12   effects on additional brain regions identified. The new data were subject to a more rigorous

13   statistical analysis than in 1998. The profile analysis described in Chapter 5 required that all

14   areas of the brain measured were altered in a dose-dependent fashion and effects were again

15   demonstrated not only in the corpus callosum but other brain regions as well (Geller, 2001d).

16   Likewise the neoplastic potential for perchlorate that had been demonstrated only at high

17   doses in historical studies was confirmed at lower doses by the thyroid adenomas reported by the

18   PWG (Wolf, 2000; 2001) for the F1-generation pups at 19 weeks (P2 parents) from the

19   two-generation reproductive study (Argus Laboratories, Inc., 1999). Consistent with the

20   proposed mode-of-action model, the anti-thyroid effects leading to neoplasia are likely to be via

21   the non-linear mechanism described above. The genotoxicity battery established that perchlorate

22   is not directly damaging to DNA.

23   Thus, the key event for the anti-thyroid effects of perchlorate is its perturbation of the

24   hypothalamic-pituitary-thyroid axis by competitive inhibition of iodide uptake at the NIS. The

25   evidence for this effect is built upon the observation of consistent changes across a range of

26   experimental designs, including various species. These changes demonstrate effects on thyroid

27   and pituitary hormones, increases in thyroid weight, and increases in three different diagnoses of

28   thyroid histopathology (colloid depletion, hypertrophy, and hyperplasia). In addition,

29   corresponding neurodevelopmental (motor activity and brain morphometry) and neoplastic

30   outcomes were observed in special assays; these outcomes are also consistent with the proposed

1   mode of action and provide further evidence to confirm that the perturbation of the thyroid

2   hormone economy should be viewed as adverse.

3   Due to the age and time-dependent nature of the critical effect, no one principal study is

4   being chosen for this derivation. Instead, a weight-of-the-evidence approach will be taken to

5   arrive at a point of departure in Section 7.1.3.

6

7   7.1.2 Dosimetric Adjustment of Exposures Associated with Effect Levels

8   Adjustments for interspecies differences in the internal dose delivered to target tissues

9   should be made before an evaluation of the data array for valid comparisons across endpoints

10   (U.S. Environmental Protection Agency, 1994). Based on the mode of action and the available

11   PBPK model structures, two dose metrics were considered to describe the biologically effective

12   dose for perchlorate: (1) the area under the curve (AUC) for perchlorate in the serum associated

13   with drinking water exposures and (2) the percent of iodide uptake inhibition in the thyroid.

14   These correspond to the different exposure components along the exposure-dose-response

15   continuum in the mode-of-action model (Figure 7-1).

16   As described in Chapter 6, the serum perchlorate AUC was developed as the first dose

17   metric based on data in rats and humans after drinking water exposures. To predict the

18   "transient" phase of initial iodide inhibition in the rat, i.e., before upregulation of the NIS or

19   increases in TSH, the second dose metric was based on RAIU measurements made in adult male

20   rats dosed with perchlorate by iv two hours prior to an iv dose of radiolabled iodide. Table 7-1

21   presents the human equivalent exposures (HEE) estimates calculated using the PBPK models for

22   serum perchlorate AUC as the dose metric. Table 7-2 shows the ratios for this same dose metric

23   that can be applied in the parallelogram approach to arrive at estimates for different life stages

24   used to observe effects in the different experimental endpoints. Fetal rat predictions were based

25   on data developed for GD21. Neonatal rat predictions were based on data for PND10. This

26   approach was taken since PBPK models for human pregnancy and lactation do not exist for

27   perchlorate distribution. The calculation using the ratios approach is described in Chapter 6.

28   The resultant adult HEE values for the different life stages of the rat experiments are shown in

29   Table 7-3.

30   It can be observed in the tables in Merrill (2001e) that the pregnant and lactating rats have

31   significantly higher average serum perchlorate concentrations at the lowest drinking water dose

TABLE 7-1. PBPK-MODEL CALCULATED HUMAN EQUIVALENT EXPOSURES (HEE) TO VARIOUS EXPERIMENTAL DOSES IN THE MALE RAT FOR 15 AND 70 KG HUMAN BASED ON PERCHLORATE AREA UNDER THE CURVE (AUC) IN SERUM OR THYROID AS THE DOSE METRIC (Merrill, 2001e)

Adult Male Rat DW[a] Dose (mg/kg-day)	Human 15 kg HEE (mg/kg-day) based on serum[b] AUC	Human 70 kg HEE (mg/kg-day) based on serum[b] AUC	Human 15 kg HEE (mg/kg-day) based thyroid[b] AUC	Human 70 kg HEE (mg/kg-day) based on thyroid[b] AUC
0.010	0.030	0.021	0.0002	0.0001
0.1	0.145	0.100	0.002	0.001
1.0	0.745	0.505	0.008	0.006
3.0	2.05	1.35	0.052	0.035
5.0	3.35	2.25	0.145	0.098
10.0	6.75	4.45	0.725	0.460
30.0	20.3	13.2	163.0	110.0
100.0	65.0	43.8	490.0	330.0

[a]DW = drinking water.
[b]Calculated from PBPK-derived rat AUC(s) at steady state between 240 and 264 hrs during DW exposure, using upregulated $Vmaxv\_T_P$ values from (Merrill, 2001e: Table 1).

TABLE 7-2. RATIO OF PBPK-DERIVED PERCHLORATE AREA UNDER THE CURVE (AUC) SERUM CONCENTRATIONS IN DRINKING WATER FOR VARIOUS EXPERIMENTAL LIFE STAGES (Merrill, 2001e)

Rat DW[a] Dose (mg/kg-day)	Male Rat: Pregnant Rat	Male Rat: Lactating Rat	Male Rat: Fetal Rat	Male Rat: Neonate Rat	Pregnant Rat: Fetal Rat	Lactating Rat: Neonate Rat
0.01	0.63	0.58	1.44	1.16	2.28	1.99
0.1	0.73	0.54	1.06	0.85	1.46	1.56
1.0	0.90	0.84	1.44	1.01	1.61	1.20
3.0	0.94	0.95	1.67	1.71	1.77	1.80
5.0	0.95	0.98	1.74	2.14	1.82	2.18
10.0	0.96	1.01	1.80	2.70	1.87	2.69
30.0	0.97	1.02	1.84	3.33	1.90	3.26
100.0	0.97	1.03	1.85	3.65	1.92	3.55

[a]DW = drinking water.

TABLE 7-3. PBPK-MODEL CALCULATED HUMAN EQUIVALENT EXPOSURES (HEE) TO VARIOUS EXPERIMENTAL LIFE STAGES IN THE RAT USING SERUM PERCHLORATE AREA UNDER THE CURVE (AUC) AS THE DOSE METRIC

Dose (mg/kg-day)	Human Equivalent Exposure[a] (mg/kg-day)				
	Adult Male Rat	Pregnant Rat	Fetal Rat	Lactating Rat	Neonate Rat
0.01	0.02	0.01	0.03	0.01	0.02
0.1	0.10	0.07	0.10	0.05	0.08
1.0	0.51	0.46	0.73	0.43	0.52
3.0	1.35	1.3	2.3	1.3	2.4
5.0	2.25	2.14	3.92	2.20	4.82
10.0	4.4	4.22	7.9	4.4	11.9
30.0	13.2	12.8	24.3	13.5	43.95
100.0	43.8	42.5	81.0	45.11	160.0

[a]Based on predicting the area under the curve in the blood (AUCB) using the human PBPK model that achieves an equivalent degree to that simulated for the rat experimental regimen associated at different life stages. See Tables 7-1and 7-2 and Chapter 6 for explanation of calculation.

1 (0.01 mg/kg-day). This is likely due to increased binding in the serum (Merrill, 2001e). It has

2 been shown that the estrus cycle affects the concentration of binding proteins within the blood.

3 Thyroxine, which is displaced from plasma proteins by perchlorate, is bound to a greater extent

4 in the pregnant rat (Iino and Greer, 1960). It follows then that perchlorate would also be bound

5 to a greater extent during pregnancy and possibly lactation. Since serum binding affects only the

6 low doses, it is reasonable that the higher doses (1.0 through 100 mg/kg-day) would be similar

7 across the male, pregnant and lactating rats (Merrill, 2001e).

8   Tables 7-4 through 7-7 are a comparable set of tables but are based on using thyroid uptake

9 inhibition as the dose metric. Table 7-5 shows the percent of iodide uptake inhibition predicted

10 at each dose for the various life stages used in the various laboratory rat experiments.

11

12 **7.1.2.1 Choice of Dose Metric**

13   Because developmental effects are of concern, an argument could be made that peak—and

14 not AUC—is the appropriate dose metric with the rationale that any transient dose could be

15 responsible for permanent deficits. However, the AUC values, as opposed to peak

TABLE 7-4. PBPK-MODEL CALCULATED HUMAN EQUIVALENT EXPOSURES (HEE) TO VARIOUS EXPERIMENTAL DOSES IN THE ADULT MALE RAT FOR 15 AND 70 KG HUMAN BASED ON % IODIDE UPTAKE INHIBITION IN THE THYROID

Rat iv Dose (mg/kg)	Adult male rat inhibition at 2-hr post iv dose	Human 15 kg HEE (mg/kg-day)	Human 70 kg HEE (mg/kg-day)
0.01	1.5%	0.006	0.004
0.1	16.3%	0.075	0.048
1.0	74.5%	1.5	0.9
3.0	90.0%	4.8	2.7
5.0	93.5%	8.0	4.9
10.0	96.2%	16.0	9.0
30.0	98.1%	35.0	19.3
100.0	98.7%	50.0	33.0

TABLE 7-5. PBPK-MODEL PREDICTED % INHIBITION OF IODIDE UPTAKE IN THE THYROID[a]

Rat DW [b] Dose (mg/kg-day)	Adult Male Rat	Pregnant Rat	Fetal Rat[c]	Lactating Rat[d]	Neonate Rat[c,d]	70 kg Human
0.01	1.5%	3.2%	-129.1%	0.5%	0.4%	2.8%
0.1	16.3%	30.1%	27.9%	5.3%	1.3%	23.7%
1.0	74.5%	88.7%	81.2%	62.9%	3.0%	80.2%
3.0	90.0%	93.8%	90.3%	92.8%	3.3%	92.3%
5.0	93.5%	97.0%	90.4%	95.8%	3.1%	95.2%
10.0	96.2%	97.9%	97.9%	97.6%	3.8%	97.4%
30.0	98.1%	98.6%	98.9%	98.5%	6.1%	98.9%
100.0	98.7%	98.8%	99.2%	98.8%	13.4%	99.4%

[a]Based on iv administration to rat and drinking water in human.
[b]DW = drinking water
[c]Values for these tissues not validated versus data.
[d]All calculations are for PND10 in lactating and neonatal rat.

TABLE 7-6. RATIOS OF PBPK-DERIVED % IODIDE UPTAKE INHIBITION IN DRINKING WATER FOR VARIOUS EXPERIMENTAL LIFE STAGES[a]

Rat DW[b] Dose (mg/kg-day)	Male Rat: Pregnant Rat	Male Rat: Lactating Rat	Male Rat: Fetal Rat[c]	Male Rat: Neonate Rat[c]	Pregnant Rat: Fetal Rat	Lactating Rat: Neonate Rat[c,d]
0.01	0.48	3.24	−0.01	4.02	−0.02	1.2
0.1	0.54	3.06	0.59	12.75	1.08	4.2
1.0	0.84	1.18	0.92	24.53	1.09	20.7
3.0	0.96	0.97	1.00	27.49	1.04	28.4
5.0	0.96	0.98	1.03	30.45	1.07	31.2
10.0	0.98	0.99	0.98	25.61	1.00	26.0
30.0	0.99	1.00	0.99	16.06	1.00	16.1
100.0	1.00	1.00	1.00	1.37	1.00	7.4

[a]Inhibition in human was PBPK-derived from 2 wks ClO_4^--exposure in drinking water (DW); all rat values simulated from an iv dose.
[b]DW = drinking water
[c]Model predicted in fetal and neonate rats not validated with data.
[d]All calculations are for PND10 in lactating and neonatal rat.

TABLE 7-7. PBPK-MODEL CALCULATED HUMAN EQUIVALENT EXPOSURES (HEE) TO VARIOUS EXPERIMENTAL LIFE STAGES IN THE RAT USING % IODIDE UPTAKE INHIBITION IN THE THYROID AS THE DOSE METRIC

Dose (mg/kg-day)	Human Equivalent Exposure[a] (mg/kg-day)				
	Adult Male Rat	Pregnant Rat	Fetal Rat	Lactating Rat	Neonate Rat
0.01	0.004	0.002	—	0.01	0.02
0.1	0.048	0.026	0.03	0.15	0.61
1.0	0.90	0.756	0.83	1.06	22.05
3.0	2.7	0.259	2.70	2.62	74.2
5.0	4.9	4.70	5.05	4.80	149.2
10.0	9.0	8.82	8.82	8.91	230.5
30.0	19.3	19.1	19.1	19.3	309.96
100.0	33.0	33.0	33.0	33.0	33.0

[a]Based on predicting the % iodide uptake in the thyroid using the human PBPK model that achieves an equivalent degree to that simulated for the rat experimental regimen associated at different life stages. See Tables 7-4 and 7-6 and text for explanation of calculation.

1   concentrations, were used based on the assumption that these dose metrics would represent an

2   averaging of the serum and thyroid perchlorate concentrations and would be better correlated

3   with the inhibition effect on iodide uptake. The correlation was shown to be good between the

4   AUC and the degree of inhibition (Figures 6-47 through 6-50). Further, due to the rapid phase of

5   distribution after an iv dose, measurement of concentrations are very difficult to attain

6   experimentally and are more variable. Using simulated peak concentrations after iv injections is

7   potentially problematic due to the inexact modeling of the actual distribution of dose in the

8   tail-vein volume and the exact time of mixing in the whole blood compartment (Merrill, 2001e).

9   It was also observed by EPA that the ratios for peak perchlorate serum values (Merrill, 2001e:

10   Table 6) were in good agreement with those for the perchlorate serum AUC and that the serum

11   AUC were slightly more conservative if different at all.

12   Merrill (2001e) expressed concern regarding the thyroid values in neonates and fetuses

13   because these values were not validated against experimental data. Fetal and neonatal thyroid

14   were never actually analyzed for perchlorate concentration. In the case of the fetus, kinetic

15   parameters were determined by fitting model simulations of fetal thyroid concentration to

16   available iodide data and assuming that the perchlorate:iodide ratio would be similar to that of

17   the mother. In the case of the neonatal rat, no data were available for thyroid concentrations for

18   either perchlorate or iodide. Thus, model predictions were based on allometrically scaling

19   maternal parameters for thyroid uptake. It was the opinion of the AFRL/HEST authors that while

20   the thyroid parameters in the fetus and neonatal rat were highly informative, they should not be

21   used in the formal risk assessment (Merrill, 2001e). EPA concurs with these considerations and

22   recommendation.

23   In general, the models were believed to provide a good description of perchlorate and

24   iodide disposition in the blood. Using the models to describe dose metrics in the thyroid was

25   viewed as less reliable due to assumptions regarding parameters and the lack of experimental

26   data for validation. The models were able to successfully describe serum perchlorate and iodide

27   concentrations for both acute (based on iv doses) and chronic drinking water in the adult male,

28   pregnant, neonatal and fetal rat, and greater confidence can be afforded these predictions

29   (Merrill, 2001e).

30   Tables 7-3 and 7-7 demonstrate good correspondence in the HEE estimates predicted for

31   both dose metrics at the lower doses for the lactating and neonatal rats, but not for the male adult,

1   pregnant or fetal rats where there is an order of magnitude difference. The iodide inhibition

2   metric predicts a 10-fold lower HEE in both the adult male and pregnant dam when compared to

3   the HEE estimated based on the serum AUC. The fetal rat value for iodide inhibition was

4   viewed as unreliable for the reasons stated above. All of the factors influencing this disparity are

5   not fully appreciated at this time but can reasonably be ascribed to uncertainty in the thyroid

6   descriptions that were not validated with experimental data, and will require additional studies to

7   characterize accurately. For these reasons, the adjustment factor to arrive at an HEE estimate

8   was based on perchlorate serum AUC as the dose metric.

9

10   7.1.3 Point-of-Departure Analysis

11   Various statistical procedures were used for each of the different outcome measures for the

12   various endpoints described in Chapter 5. The weight-of-evidence approach herein relies on the

13   results, and the details on the statistical analyses are provided in Chapter 5 and associated

14   memoranda from EPA and NIEHS scientists. In general, benchmark dose analysis was used for

15   the thyroid histopathology because the EPA advocates the use of quantitative dose-response

16   modeling to diminish the influence of dose-spacing, sample size, and variability on the NOAEL

17   designation (Crump et al., 1995). Likewise, ANOVA was used to evaluate the thyroid and

18   pituitary hormone data (Crofton and Marcus, 2001) although benchmark analyses were also

19   performed as a comparison (Geller, 2001c). The 1998 benchmark analyses for the hormone data

20   from the previous set of studies (Geller, 1998a) is provided in Appendix 7B.

21   Specific Bayesian statistical analyses were employed for the motor activity data and for

22   evaluating the significance of the tumors in the 19-week old F1-generation adult rats (Dunson,

23   2001a,b). Another specific statistical approach, profile analysis, was used to evaluate the brain

24   morphometry effects (Geller, 2001d).

25   Several studies suggest 0.01 mg/kg-day as the exposure dose that is a level of concern for

26   the adverse effects of perchlorate. The first is the profile analysis on brain morphometry effects

27   in PND21 pups in the "Effects Study" (Argus Research Laboratories, Inc., 2001) which

28   demonstrated a dose-dependent and significant effect on the size of the corpus callosum and

29   other brain regions. Statistically significant changes were also demonstrated in the PND9 pups.

30   This effect repeated effects on brain morphometry observed in the previous neurodevelopmental

31   study (Argus Research Laboratories, Inc. 1998a) that were a noted concern to the EPA in the

1   1998 risk assessment. Changes in the corpus callosum at a later time point on PND82 were also

2   observed in that previous study.

3   An increase in the corpus callosum plausibly represents a delay in developing brain

4   structures since this area is known to increase in size and then decrease later during development.

5   Neurodevelopmental toxicity suggestive of delays was also demonstrated by effects on motor

6   activity in both the Argus Research Laboratories, Inc. (1998a) and repeated in the Bekkedal et al.

7   (2000) study. The increases in motor activity represent activity that should have subsided by

8   these test dates. A type of hyperactivity has been noted in monkeys exposed to PCBs (Rice,

9   2000).

10   These effects on brain morphometry and motor activity are of particular concern because

11 the relative sensitivity of laboratory animal assays to adequately characterize the types of deficits

12 related to maternal hypothyroxinemia in large population studies is unknown (Morreale de

13 Escobar, 2000; Haddow et al., 1999; Pop, 1999). Screening neurodevelopmental studies may not

14 have the power to ascertain neurological effects that might result from small changes in the

15 thyroid-pituitary hormone economy. As pointed out by Crofton (1998j), the sensitivity of animal

16 models used to explore the role of thyroid hormones in neural development is currently

17 equivocal. Most of the data collected and published to date were with high doses of thyrotoxic

18 chemicals (e.a., methimazole, propylthiouracil) or with thyroidectomy. It is not known whether

19 the available tests are capable of detecting more subtle changes in nervous system development.

20 An analysis presented by Crofton (1998j) suggested that measurements of nervous system

21 development are less sensitive than measurements of T4. Two reasons for this relationship were

22 presented. First, the brain may be protected from perturbations in circulating concentrations of

23 T4, as demonstrated by upregulation of deiodinases in brain tissue that compensate for very large

24 decreases in circulating T4. The second reason, and one for concern in the context development

25 of this model, is that currently available testing methods, particularly screening methods, may not

26 be sufficiently sensitive. Recent data suggest that the battery is insensitive to alterations in

27 thyroid hormones during development (Goldey, 1995a,b).

28   The 0.01 mg/kg-day dosage as a level for concern was also supported by thyroid

29 histopathology in the database. Changes in colloid depletion observed on PND4 in both the 1998

30 neurodevelopmental study (Argus Research Laboratories, Inc., 1998a) and the newer 2001

31 "Effects Study" (Argus Research Laboratories, Inc. 2001) were demonstrated. The BMDL

1   estimated for those studies on PND4 was 0.33 mg/kg-day, but an estimate of 0.009 mg/kg-day is

2   also obtained with a model demonstrating adequate fit to the data. The BMDL for colloid

3   depletion in pups on GD21 was 0.12 mg/kg-day, but for female pups alone on GD21 was 0.04

4   mg/kg-day. The BMDL estimated for thyroid hypertrophy in weanling pups from the two-

5   generation study (Argus Research Laboratories, Inc., 1999) was 0.06 mg/kg-day. Of notable

6   concern to this consideration was that the BMDL estimates decreased with duration in the 90-day

7   study. The BMDL estimates for colloid depletion were 0.28 and 0.03 mg/kg-day at the 14-day

8   and 90-day time points in the Springborn Laboratories, Inc. (1998) study. The BMDL estimates

9   for hypertrophy were 0.017 and 0.008 mg/kg-day at the 14-day and 90-day time points. This

10   effect of duration was of concern as it was also evident by the observation of tumors in the

11   F1-generation adults at 19 weeks. Both observations suggest concern that duration may

12   recalibrate either the homeostatic interactions of the hypothalamic-pituitary-feedback system or

13   the cellular sensitivity and demand for the thyroid hormones.

14   The thyroid hormone data in a number of studies also designated 0.01 mg/kg-day as a

15   LOAEL. Levels of T4 were significantly decreased and TSH levels statistically increased at this

16   dosage in the dams on GD21 in the same study as the significant brain morphometry

17   measurements in the PND21 pups (Argus Research Laboratories, Inc. 2001), revealing no

18   NOAEL for hypothyroidism in the dams. The pups in that study were also affected at

19   0.01 mg/kg-day. Effects on T3 occurred at GD21, PND5, and PND9 at this dosage. The

20   0.01 mg/kg-day dose was the LOAEL for effects on T4 and TSH at PND21 in the male pups and

21   for TSH in both sexes at PND9 as well. This same dose (0.01 mg/kg-day) was also the LOAEL

22   for decreases in T4 and increases in TSH at the 14-day and 90-day time points in the 90-day

23   study (Springborn Laboratories, Inc., 1998).

24   The ANOVA estimates for hormone data were used to characterize this effect after serious

25   consideration. While in clinical studies a normal range typically is defined by a control healthy

26   population, the ANOVA approach is an equally valid approach in that a statistically significant

27   value represents a shift in the mean for the population. The control group defines the range for

28   the unexposed, presumably healthy population, and statistically significant differences indicate

29   that the mean for an exposed group is outside of that normal range. Circadian fluctuations are

30   addressed because the same fluctuations in the control population occur as in the exposed

31   population. A small shift in the mean of a population can have significant consequences to

1   individuals in the tails of the distributions of those populations. Indeed, such an evaluation

2   underlies the basis for the blood lead level used in the National Ambient Air Quality Standard

3   (Davis and Elias, 1996) and has been noted as an important consideration for neurotoxicity

4   (Weiss, 2000).

5   The notion that continuous data should be considered in the context of the specific dose-

6   response rather than to *a prioro* categories defined outside of the data under analysis is supported

7   in the benchmark dose literature. Murrell et al. (1998) point out that a continuous quantity

8   measurement such as the hormone data should be scaled by the range from background response

9   level to maximum response level (for increasing response functions). The authors go on to note

10   that it is a biological reality that, whatever the mechanism of effect of the toxicant, there is some

11   dose level beyond which no further change in response is seen or is theoretically feasible.

12   In general, there is some type of limitation or saturation phenomenon that occurs at high enough

13   doses (e.g., in the saturation of the symporter capacity, as suggested by the modeling effort in

14   Chapter 6 and the data of Chow and Woodbury [1970] and of Meyer [1998]).

15   An analogy to the case of quantal data for which an effect is defined as a probability metric

16   in which the response reaches a maximum at one, is, that for continuous measures, the extra

17   effect can be defined as the change in effect from background standardized by the total range of

18   response (Murrell et al., 1998). The total response range is not necessarily the response range of

19   the observed responses in a study; rather, it is defined by a determination of the minimum and

20   maximum possible responses according to, for example, a model equation fitted to the data as in

21   the case of benchmark analyses. In all BMD analyses, however, the hormone BMDL estimates

22   were shown to be extremely low (Geller, 1998a; Geller, 2001c). This may not necessarily be

23   surprising given that hormones are operative at low doses by definition, but corresponding

24   changes in thyroid histopathology were more consistent with the ANOVA estimates.

25   Finally, the NOAEL for immunotoxicity suggested by the dermal contact hypersensitivity

26   assay at 0.02 mg/kg-day can be viewed as supportive, especially since deficiencies in this study

27   raise concern for the characterization and because a LOAEL for the effect was demonstrated at

28   0.06 mg/kg-day.

29

30

31

7.1.4 Application of Uncertainty Factors

The types of uncertainty factors (UF) applied for various extrapolations required to arrive at a reference dose were discussed in Chapter 3. Figure 7-4 illustrates schematically that the interspecies and intraspecies UFs embody attributes of both uncertainty and variability. A factor for variability across humans typically is applied to account for potentially susceptible portions of the population. As shown in Figure 7-5 (Jarabek, 1995b), both of these factors typically are broken into components of approximately three each for pharmacokinetics (toxicokinetics) and pharmacodynamic (toxicodynamic) processes. This scheme is consistent with that used by the World Health Organization (WHO) (Jarabek, 1995b).
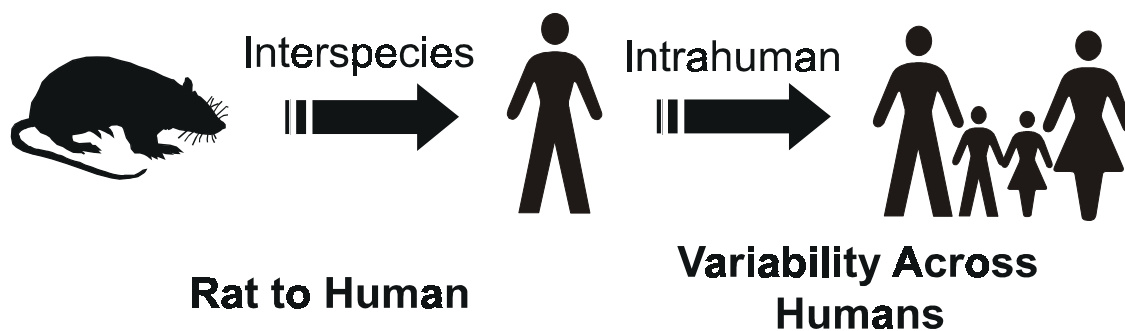
Figure 7-4. Consideration of uncertainty and variability influence interspecies and intrahuman extrapolation.

There were a total of four (4) uncertainty factors applied in this derivation, resulting in a composite factor of 300. The partial factors of 3 represent "halving" of each UF that is believed to be an upper bound on a lognormal distribution; i.e., $10^{0.5}$, so that multiplication of the various partial factors results in a composite of 100 (U.S. Environmental Protection Agency, 1994).

A 3-fold factor for intraspecies variability was retained due to the variability observed in the data and PBPK modeling for the adult humans and because these subjects do not represent kinetic data for the potentially susceptible populations of the hypothyroid or hypothyroxinemic pregnant women and their fetuses. There was also uncertainty in the parallelogram approach to extending the adult structure to address different life stages. These uncertainties might be

```
Lab Animal                    Target                      Tissue
Exposure          ──────→     Dose          ──────→       Response
Concentration                                             
                    ┊                        ┊               ┊
                    ↓                         ↓               ┊
Interspecies      ┌───┐                    ┌───┐              ┊
(10A)             │ 3 │                    │ 3 │              ┊
                  └───┘                    └───┘              ┊
                    ┊                        ┊               ┊
                    ↓                         ↓               ┊
Intrahuman        ┌───┐                    ┌───┐              ┊
(10H)             │ 3 │                    │ 3 │              ┊
                  └───┘                    └───┘              ┊
                    ┊                        ┊               ↓
Human               ↓          Target        ↓          Tissue
Exposure      ←┄┄┄┄┄          Dose     ←┄┄┄┄┄           Response
Concentration                                           
```
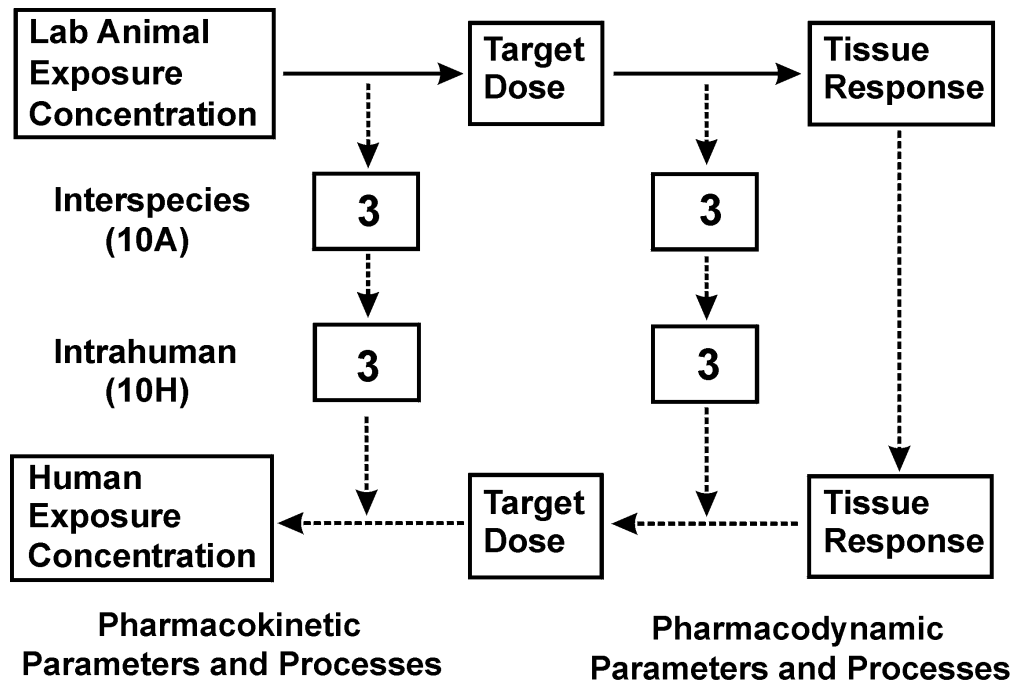
Figure 7-5.   Schematic of uncertainty factor components incorporated into exposure-dose-
response characterization for interspecies and intrahuman extrapolations
(Jarabek, 1995b).

1   mitigated by further development of pregnancy and lactation models or the models might be

2   further validated with radionucleide data using a parallelogram approach between perchlorate

3   and iodide as described in Chapter 6. This reduced factor was a point of considerable debate,

4   especially given the concern over the animal neurodevelopmental assays for adequately

5   characterizing neuropsychological development deficits in susceptible populations. However, it

6   was also discussed that the UF values are not entirely independent; e.g., aspects underlying the

7   duration extrapolation also might underlay the intrahuman UF (Jarabek, 1995b).

8   The interspecies factor was omitted due to general confidence that the extrapolation based

9   on perchlorate distribution (and on iodide inhibition by perchlorate at lower doses) was

10   accurately characterized by the PBPK modeling effort described in Chapter 6. Concern for

11   eliminating this factor was again considered in the context of the lack of independence with other

12   applied UF. The concern that the HEE was not based on iodide inhibition but rather the serum

1   perchlorate AUC was assuaged somewhat by the correlations that demonstrated a close

2   relationship between these two measures.

3   A full 10-fold factor was applied to extrapolate the LOAEL for the brain morphometry,

4   thyroid histopathology, and hormone changes observed at the 0.01 mg/kg-day level. Designating

5   these changes to be adverse is consistent with the proposed mode of action and existing Agency

6   guidance and procedures. The neurotoxicity assessment guidelines (U.S. EPA, 1998a) specify

7   changes in brain structure as adverse. The OPPTS has used thyroid hormone changes to

8   designate effect levels. Finally, the shallow slope of the response curve at these lower levels

9   suggested that a full factor should be applied.

10   A 3-fold factor for duration was applied due to the concern for the biological importance of

11   the statistically significant increase in tumors in the F1-generation pups at 19 weeks (P2, second

12   parental generation). The occurrence of these tumors with a dramatically reduced latency and

13   with a significance in incidence greater than the NTP historical data (Dunson, 2001b) for thyroid

14   tumors in this strain of rat was reason for concern. As discussed earlier, the concerns were that

15   this observation represented the potential for *in utero* programming; and that the decrease in the

16   NOAEL/LOAEL estimates for hormone perturbations and histopathology between the 14-day

17   and 90-day time points represented a recalibration of the regulatory feedback system or changes

18   in cellular sensitivity and demand for thyroid hormones with extended exposures. This factor

19   can also be viewed as part of a data base deficiency because there are no long-term bioassays of

20   perchlorate with contemporary design and data quality. While the original strategy aimed at

21   determining a NOAEL for thyroid histopathology as a precursor lesion to tumors in the 90-day

22   study, this finding in the F1-generation cannot be ignored, especially in light of an emerging

23   appreciation of findings suggesting a phenomenon known as *in utero* imprinting with endocrine

24   disruption (Prins et al., 2001; Phillips et al., 1998; Seckl, 1997). Thus, *in utero* disruption of

25   thyroid hormones in the developing fetus may predispose the developing neonate and adult to

26   future environmental insults to the thyroid gland by making the fetus more sensitive. Weiss

27   (2000) has noted that changes in brain functions occur throughout life and some consequences of

28   early damage may not even emerge until advanced age. This could be exacerbated if

29   environmental insults to the thyroid were to be continued throughout life.

30   The potential for perchlorate to cause immunotoxicity remains a concern so that a 3-fold

31   factor was applied for the database insufficiency. New studies based on recommendations at the

1   1999 external peer review had some deficiencies and reinforced concern about the lack of an

2   accurate characterization of this endpoint.

3

4   **7.1.5 Operational Derivation of the Reference Dose**

5   The HEE for the neonatal rat corresponding to brain morphometry and hormone changes

6   observed in the PND21 pups (also the PND9 pups) at the 0.01 mg/kg-day dosage would be 0.02

7   mg/kg-day (Table 7-3). However, because the dams on GD21 were shown to be hypothyroid

8   (with statistically-significant decreases in T4 and increases in TSH) at this same dosage, and

9   because the temporal windows underlying the neonatal brain morphometry effects are unknown,

10 and because the brain morphometry effects may have occurred *in utero* due to the dams'

11 hormone deficiency, the HEE estimate for dams of 0.01 mg/kg-day was chosen as the operational

12 derivation. It was noted that this choice was not as conservative as using the HEE for iodide

13 inhibition in the dams (0.002 mg/kg-day), but it was viewed as more accurate given the concerns

14 for the reliability of the thyroid estimates.

15   According to Dollarhide (1998), who spoke with Argus laboratory on behalf of the sponsor

16 (PSG), the reported doses were of ammonium perchlorate and not the anion itself. Thus, an

17 adjustment for percent of the molecular weight of the salt from ammonium (15.35%) must also

18 be made. Further, because the analytical methods measure the anion concentration in

19 environmental samples, this is the appropriate expression for the RfD to use while making valid

20 comparisons for risk characterization. Thus, the derivation for an RfD for the perchlorate anion

21 as itself is as follows:

22

23                 $$0.01 \text{ mg/kg-day} \times 0.85 / 300 = 0.00003 \text{ mg/kg-day.}$$ (7-1)

24

25 Note that the appropriate adjustment for any salt of perchlorate (e.g., adjustment by a factor of

26 0.72 for potassium perchlorate) should be made when evaluating toxicity data for similar

27 assessment activities.

28   It is critically important to distinguish the proposed RfD from any guidance value that may

29 result. An RfD would be only one step in the future regulatory process of determining, based on

30 a variety of elements, whether a drinking water standard for perchlorate is appropriate. As with

31 any draft EPA assessment containing a quantitative risk value, that risk estimate is also draft and

1   should be construed at this stage to represent Agency policy. The units for an RfD are mg/kg-

2   day. Conversion of an RfD to a drinking water equivalent level (DWEL) is based on adjusting

3   by body weight (kg) and drinking water consumption (L) to arrive at a level expressed in units of

4   mg/L (ppb). Derivation of a maximum contaminant level goal (MCLG) from a DWEL by the

5   OW typically involves the use of a relative source contribution (RSC) factor to account for non-

6   water sources of exposures such as those discussed in Chapters 8 and 9.

7   Because the effect is viewed to be the result of neurodevelopmental deficits resulting from

8   the hypothyroid or hypothyroxinemic state induced by the mother's exposure, and because

9   developmental neurotoxicity may emerge later in the life or be exacerbated later in life,

10   conversion factors for the adult of 70 kg body weight and 2 L of water per day are considered

11   appropriate. Recent guidance from the OW in its Methodology for Deriving Ambient Water

12   Quality Criteria for the Protection of Human Health (U.S. Environmental Protection Agency,

13   2000) provides a decision flow chart for derivation of the RSC and recommends 80% as a ceiling

14   and 20% as the floor for this factor when data are adequate to estimate sources of exposure.

15   When data are not adequate to estimate other anticipated exposures, OW recommends a default

16   RSC of 20%. (U.S. Environmental Protection Agency, 2000: Chapter 4, Section 4.2.2.4 on

17   apportionment decisions). EPA does not recommend that high-end intakes be assumed for every

18   exposure source since the combination may not be representative of any actually exposed

19   population or individual.

20   A hypothetical adjustment of the 0.00003 mg/kg-day RfD by 70 kg and 2 L would thereby

21   result in a DWEL of 1 ug/L (ppb) and application of an RSC between 0.2 to 0.8 would thereby

22   result in an MCLG in the range of 0.2 to 0.8 ug/L (ppb). These values are in the range of current

23   analytical capabilities. As discussed in Chapter 1, improvements to the analytical methods on the

24   near horizon or expected to be published this spring could result in minimum reporting limits in

25   this range and lower (Yates, 2001).

26   Concern is often expressed in the regulatory arena for the potential added susceptibility of

27   children in developing DWEL estimates based on different conversion factors (15 kg and 1 L).

28   Consequently, the EPA asked for additional PBPK simulations to help inform this dialogue.

29   As shown in Table 7-1, the HEE estimates for a 15 kg human for serum perchlorate AUC can be

30   as great as two-fold higher than those predicted for the 70 kg human due to differences in

31   distribution volumes and excretion. Thus, if the 15 kg and 1 L values are used to convert this 2-

1 fold higher HEE value in an analogous derivation to the adult RfD derivation and DWEL

2 calculation above, an estimate of 1 ppb that is equivalent to the adult conversion results.

3

4 **7.1.5.1 Comparison with Derivation Considering Human Data**

5   It is important to evaluate this derivation in context with the evidence from the available

6 and relevant human data. As described in Chapter 4, the EPA felt that both the observational

7 epidemiological and the human clinical studies have significant scientific and technical

8 limitations that preclude their use as the basis for a quantitative dose-response assessment. The

9 clinical study subject attributes (euthyroid adults) and study design issues (sample size, RAIU

10 time points, etc.) made these data less reliable than the laboratory animal toxicological data to

11 ascertain effect levels for the basis of an RfD derivation. In addition, on December 14, 2001,

12 after internal peer review of this document, the Agency articulated its interim policy on the use of

13 third-party studies submitted by regulated entities (U.S. Environmental Protection Agency,

14 2001c). For these purposes, EPA is considering "third party studies" as studies that have not

15 been conducted or funded by a federal agency pursuant to regulations that protect human

16 subjects. Under the interim policy, the Agency will not consider or rely on any such human

17 studies (third-party studies involving deliberate exposure of human subjects when used to

18 identify or quantify toxic endpoints such as those submitted to establish a NOAEL or NOEL for

19 systemic toxicity of pesticides) in its regulatory decision making, whether previously or newly

20 submitted. Some of the clinical studies contained in this database fall in this category of studies

21 not to be considered. However, the scientific and technical strengths and weaknesses of these

22 studies were described before this Agency policy was articulated. Therefore, because of the

23 scientific shortcomings of these studies, they will not be used as "principal studies" in the

24 derivation of an RfD. The ethical issues surrounding the conduct of these studies or their use for

25 regulatory purposes in light of the Agency's interim policy will not be discussed in this

26 document. The Agency is requesting that the National Academy of Sciences conduct an

27 expeditious review of the complex scientific and ethical issues posed by EPA's possible use of

28 third-party studies which intentionally dose human subjects with toxicants to identify or quantify

29 their effects.

30   These issues not withstanding, a dose of 0.007 mg/kg-day has been suggested by some

31 authors in an abstract (Greer et al., 2000) to be a NOAEL estimate. This was based on an

1   average 6.2 % decrease relative to baseline of RAIU measured on Day 14 of exposure to seven

2   subjects at the 8-hour time point (unpublished data presented in Merrill, 2001a; Attachment #7).

3   The values for RAIU ranged from a 38.6% decrease in a 34-year old female to a 27.9% increase

4   in a 49-year old female at that dosage.

5   Prior to the articulation of the Agency's interim policy, the Agency had conducted a

6   comparison of its reference dose derivation considering the results of the study described above,

7   which falls within the category of a "third-party study" described by the authors as demonstrating

8   a NOAEL in humans. If this study were to be considered in lieu of the laboratory animal data

9   and PBPK modeling, the following would need to be considered. The seven subjects (six

10   females and one male) were euthyroid and ranged in age from 18 to 49. Because this is a limited

11   data set (small sample size), with noted variability and because of relevance to the elderly

12   woman, cardiac risk patient, hypothyroid or hypothyroxinemic pregnant woman, or fetus as the

13   susceptible population is difficult to ascertain, an uncertainty factor of 3-fold for this iodide

14   uptake inhibition level as a minimal LOAEL as well as a 3-fold factor for intrahuman variability

15   would be warranted. This is particularly relevant if this value is viewed in context with the

16   neurodevelopmental effects in laboratory animal data. At a minimum each factor should be

17   3-fold, and discussion with respect to the meaning of these factors with respect to population

18   effects again entertained. None of the human studies of perchlorate reviewed in Chapter 4 have

19   adequately investigated neurodevelopmental outcomes. The concern for duration of exposure

20   was at least a 3-fold factor per the above laboratory animal data discussion and should also be

21   applied, as well as the 3-fold factor for database deficiencies because these considerations and

22   deficiencies are not obviated by the use of human data.

23   Thus, a derivation based on the available human data would estimate the RfD at a

24   maximum of 0.00007 mg/kg-day, an estimate in rather good agreement with that proposed based

25   on the laboratory animal data (0.00003 mg/kg-day). If a larger UF were to be applied to the

26   human data, as could be justified for the intrahuman factor, the resultant estimate would be

27   essentially equivalent to that proposed using the laboratory animal data.

28   The consistency between the estimates based on the laboratory animal versus the human

29   data is likely due, at least in apart, to the use of AFRL/HEST PBPK modeling (Merrill, 2001c,d;

30   Clewell, 2001a,b) to perform the interspecies extrapolation rather than the use of default factors.

31   It should be noted that the original motivation for performing these human studies (as discussed

1   in Chapter 3) in the perchlorate testing strategy was to support such interspecies pharmacokinetic

2   extrapolation and not to derive NOAEL estimates for thyroid effects in the human population. In

3   addition, as noted in Chapter 4, the EPA felt that both the observational epidemiological and the

4   human clinical studies have significant scientific and technical limitations that precluded their

5   use as the basis for a quantitative dose-response assessment. As mentioned previously, under the

6   interim policy articulated on December 14, the Agency will not consider or rely on any such

7   human studies (third-party studies involving deliberate exposure of human subjects when used to

8   identify or quantify toxic endpoints such as those submitted to establish a NOAEL or NOEL for

9   systemic toxicity of pesticides) in its regulatory decision making, whether previously or newly

10   submitted. Nonetheless, the use of both previously published and newly-derived human data by

11   the Air Force in its modeling efforts was important. The AFRL/HEST PBPK model approach

12   allowed EPA to confirm that humans were as sensitive as rats to the iodide uptake inhibition

13   effects of perchlorate at the NIS, the key event for the proposed mode-of action of perchlorate on

14   the thyroid. In addition, the PBPK models increased the accuracy of interspecies extrapolation

15   by allowing the incorporation and integration of ADME data to describe perchlorate and iodide

16   disposition relative to the key event. These two outcomes from the integration of human and

17   animal data in the AFRL/HEST models provide greater confidence than would the laboratory

18   animal data alone that the reference dose that is derived will be protective of human health.

19

20   **7.1.5.2 Comparison with Derivation Based on Tumor Data**

21   To address neoplasia as the other potential adverse endpoint, this section will discuss how

22   an estimate could be derived based on the recently acquired tumor data.

23

24   **7.1.5.2.1 Choice of Dose-Response Procedure**

25   As discussed in Chapter 5, the genotoxicity assays included in the testing strategy

26   determined that perchlorate was not likely to be mutagenic. This was one of the critical

27   determinants in deciding on a dose-response approach for a cancer derivation. The EPA

28   guidance on assessment of thyroid follicular cell tumors (U.S. Environmental Protection Agency,

29   1998a) sets forth data needs to establish the default dose-response procedure that should be used

30   to establish that a chemical has antithyroid activity (i.e., that it is disrupting the thyroid-pituitary

31   hormone status). Table 7-8 lists the default procedures for thyroid carcinogens that would be

TABLE 7-8. DEFAULT DOSE-RESPONSE PROCEDURES FOR THYROID CARCINOGENS (U.S. Environmental Protection Agency, 1998a)

| Example | Array of Effects | | Dose-Response Methodology |
	Mutagenic	Antithyroid	
1	Either or both unknown		Linear
2	Yes	No	Linear
3	No	Yes	Margin of exposure
4	Yes	Yes	Linear and margin of exposure

1   used. The thyroid lesions observed (colloid depletion, hypertrophy, and hyperplasia) are among

2   the required lesions to demonstrate antithyroid activity. Table 7-9 shows the types of data

3   required.

4

5

TABLE 7-9. DATA DEMONSTRATING ANTITHYROID ACTIVITY (U.S. Environmental Protection Agency (1998a)

Required	Desirable
1. Increases in cellular growth	6. Lesion progression
2. Hormone changes	7. Structure-activity relationships
3. Site of action	8. Other studies
4. Dose correlations	
5. Reversibility	

1   What has been proposed in this assessment is the harmonization of the "noncancer" and

2   "cancer" assessment approaches because the target tissue is the thyroid and the mode of action is

3   the same for both the neurodevelopmental and neoplastic sequelae. The proposed RfD based on

4   precursor lesions is analogous to a nonlinear approach and viewed as a protective for thyroid

5   tumors.

6   Perchlorate has clearly demonstrated an effect in both adult, fetal, and neonatal stages in

7   thyroid histopathology, as well as a decrease in lumen size in a dose-dependent fashion. Thyroid

8   and pituitary hormone changes and expected correlations all have been demonstrated for T3, T4,

1 and TSH across an array of studies at different time points. The site of action has been

2 established as competitive inhibition of the iodide symporter although there remains some

3 uncertainty as to whether that is the only locus for the effect (e.g., evidence for intrathyroidal

4 activity) because of the efflux (discharge) phenomenon. Dose-correlations in this case were not

5 with tumors, but rather for precursor lesions (colloid depletion, hypertrophy, hyperplasia, and

6 decreased follicular lumen size). Reversibility has been demonstrated in thyroid weight, colloid

7 depletion, hypertrophy, hyperplasia, and thyroid and pituitary hormones in the 30-day recovery

8 period after the 90-day study in rats and in T4 levels of the various immunotoxicity experiments

9 in mice.

10   Lesion progression was difficult to determine because of dose-spacing and differences in

11 sample size and histological methods among the studies. However, there was a progression

12 within the 90-day study between the 14- and 90-day time points.

13   Analyses of other anions have fairly well established that the mode of action of perchlorate

14 arises from it being an anion that is recognized by the NIS (see Chapter 3).

15   Thus, the appropriate dose-response procedure for perchlorate would be a nonlinear

16 margin-of exposure approach based on demonstration that it is not genotoxic and that its

17 anti-thyroid effects are consistent with a mode of action leading from inhibition of iodide uptake

18 at the NIS through precursor lesions of perturbation of thyroid hormone economy and resultant

19 histopathological changes in the thyroid gland.

20

21 **7.1.5.2.2 Dose-response Assessment for Thyroid Neoplasia**

22   Thyroid adenomas were statistically increased in the high dose (30 mg/kg-day) group of

23 F1-generation animals sacrificed as adults (P2-generation) at 19 weeks in the Argus Research

24 Laboratories, Inc. (1999) two-generation reproductive study. Both the latency and incidence of

25 these tumors were remarkable relative to the entirety of the NTP data base for this type of tumor

26 in this strain of rat (Dunson, 2001b). Colloid depletion, hypertrophy, and hyperplasia were all

27 observed at dosages of 0.3 mg/kg-day and above with BMDL estimates of 0.9, 0.15, and

28 0.0004 mg/kg-day. This last estimate is outside the range of possible dosimetric adjustment so it

29 will not be carried forward, but consideration of the overlap among colloid depletion,

30 hypertrophy, and hyperplasia should be superimposed on the derivation. The HEE values for

31 adult versus neonatal rats are comparable at these dosages. Using the adult male rat dosimetric

1 adjustment factor to correspond to sacrifice date results in HEE estimates of 0.45 and 0.02 for

2 colloid depletion and hypertrophy.

3   Using the nonlinear approach and applying a composite factor of 100 to the HEE estimates

4 to account as above for uncertainty in intrahuman variability, duration, and database deficiencies;

5 and with factor for a minimal LOAEL of 3 to account for the fact that hyperplasia occurred at

6 over an order of magnitude lower than these two thyroid histopathology estimates, results in an

7 RfD derivation in the range of 0.005 to 0.0002 mg/kg-day. Applying a larger uncertainty factor

8 for intrahuman variability would result in a range of 0.002 to 0.00007 mg/kg-day. Thus, the

9 derivation based on tumor outcome data supports the mode-of-action concept and corroborates

10 that the proposed RfD that as derived would be protective of both neurodevelopmental and

11 neoplastic sequelae.

12

13 **7.1.5.3 Possible Susceptibility**

14   Based on the mode-of-action for perchlorate, the competitive inhibition of iodide uptake,

15 and the subsequent perturbation of thyroid hormone homeostasis, a number of factors potentially

16 could cause an increase in susceptibility of a population to perchlorate toxicity. As already

17 indicated by the choice of critical effect, the fetus, and perhaps the developing child, may

18 represent susceptible populations. However, critical data on the steady-state pharmacokinetics

19 and placental dosimetry are lacking to definitively state whether or not there is an inherent

20 pharmacodynamic component to the apparent sensitivity of pups versus dams in the laboratory

21 animal models. Individuals that are iodine deficient may be another susceptible population. The

22 elderly, especially women, and hypothyroid and hypothyroxinemic individuals or those treated

23 with anti-thyroid drugs, may be others more susceptible than the general population to the effects

24 of perchlorate. Patients with cardiac dysfunction or elevated levels of cholesterol may also be at

25 increased risk.

26

27 **7.1.6 Designation of Confidence Levels**

28   Confidence in the principal study is medium. The dose level of 0.01 mg/kg-day was the

29 lowest tested, and it was determined to be a LOAEL (not NOAEL). The small sample size for

30 the critical effect also reduces confidence in the study. Despite the new data, the confidence in

31 the database at this time remains medium because the sensitivity of these animal assays versus

1 evaluation of neuropsychological development in human population studies is not known, and

2 because a concern for potential immunotoxicity remains. Based on confidence in the study and

3 on the database together in setting the overall confidence in the RfD, the confidence in the RfD

4 currently is also medium.

5

6

7.2 INHALATION REFERENCE CONCENTRATION

8 Derivation of an inhalation reference concentration is precluded because there are no

9 inhalation data available with which to characterize dose-response or the portal-of-entry

10 modulation of internal dose. However, the EPA has been questioned as to whether the potential

11 for inhalation exposure of perchlorate from showering with contaminated water poses a health

12 risk. Given the low vapor pressure of perchlorate, it is not likely that it would come out of

13 solution. Further, Giardino et al. (1992) characterized shower particle droplet size as ranging

14 from 200 to 3,000 μm. Thus, there is minimal chance for inhalation or deposition of perchlorate-

15 laden droplets in the respiratory tract.

16

17

7.3 SUMMARY

19 The model based on mode of action for perchlorate served as a useful construct for the

20 integration of a diverse set of data. Results of studies in the testing strategy confirmed that the

21 target tissue for perchlorate is the thyroid and that the key event for its antithyroid effects is the

22 inhibition of iodide uptake at the NIS with corresponding perturbations of thyroid hormone

23 economy. Disturbances in thyroid hormone economy were confirmed to result in thyroid

24 histology as diagnosed by decreases in colloid depletion or follicular lumen size and increases in

25 hypertrophy and hyperplasia. Effects on both neurodevelopmental indices (brain morphometry

26 and motor activity) and neoplasia that could be expected based on the mode of action were also

27 demonstrated. Other developmental and reproductive effects were not observed to be as

28 sensitive as the neurodevelopmental and thyroid histopathological changes. Accurate

29 characterization of the immunotoxicity of perchlorate, notably its potential to cause contact

1   hypersensitivity, either secondarily to these hormone effects or possibly via a direct effect of the

2   anion itself, remains a concern.

APPENDIX 7A

CORRELATION ANALYSES

1
2
3

4   The correlation analyses were of two types. Hormone levels are continuous, ratio-scaled

5 values, so correlations were computed using the conventional Pearson's r statistic. Correlations

6 between ratio-scaled hormone levels and ordinally-scaled standard histology ratings must be

7 computed using nonparametric correlations. To compare variables from the different scales, it is

8 simplest to recode the data by converting the variable values into rank scores. Spearman's rank

9 order (r_s) was used to compute the correlation between the rankings of two variables. When there

10 were ties in the ranks, as there were in this data set, each value was assigned the mean of the

11 ranks that they would otherwise occupy. A correlation coefficient was then computed for the

12 rankings of the variables of interest.

13   An alternative statistic used for comparing the data sets was Kendall's tau, best thought of

14 as a measure of agreement or concordance between two sets of ranked data. It searches for the

15 number of inversions in two sets of ranked data (i.e., observations are ranked according to the

16 first variable, then reranked according to the second, and the number of interchanges that occur is

17 used to compute the statistic). The Spearman and Kendall statistics produced nearly identical

18 results. Statistics were computed using SAS® software (PROC RANK and PROC CORR,

19 SAS Institute, Cary, NC). All statistics corresponding to Figures 7A-1 through 7A-7 can be

20 found in Tables 7A-1 through 7A-6.

21
22

TABLE 7A-1. PEARSON'S r CORRELATIONS (n = 96) BETWEEN THYROID HORMONES AND TSH IN RATS OF THE CALDWELL et al. (1995) 14-DAY STUDY

	T3	T4	TSH
T3	1.00 p = 0.00	0.81 p = 0.0001	−0.65 p = 0.0001
T4		1.00 p = 0.00	−0.67 p = 0.0001
TSH			1.00 p = 0.00

TABLE 7A-2. SPEARMAN'S r_S CORRELATIONS (n = 95) BETWEEN THE RANK ORDER OF HORMONE LEVELS AND HISTOLOGICAL SEVERITY RATING DECREASE IN FOLLICULAR LUMEN SIZE (LS) IN RATS OF THE CALDWELL et al. (1995) 14-DAY STUDY

	LS
T3	-0.74 $p = 0.0001$
T4	-0.70 $p = 0.0001$
TSH	0.79 $p = 0.0001$
FH	0.75 $p = 0.0001$

TABLE 7A-3. PEARSON'S r CORRELATIONS (n = 223) BETWEEN THYROID HORMONES AND TSH IN RATS FOR THE COMBINED 14- AND 90-DAY DATA OF THE SPRINGBORN LABORATORIES, INC. (1998) SUBCHRONIC RAT STUDY

	T3	T4	TSH
T3	1.00 $p = 0.00$	0.42 $p = 0.0001$	-0.18 $p = 0.007$
T4		1.00 $p = 0.00$	-0.20 $p = 0.0027$
TSH			1.00 $p = 0.00$

TABLE 7A-4. PEARSON'S r CORRELATIONS (n = 104) BETWEEN THYROID HORMONES AND TSH FOR THE 14-DAY DATA OF THE SPRINGBORN LABORATORIES, INC. (1998) SUBCHRONIC RAT STUDY

	T3	T4	TSH
T3	1.00 $p = 0.00$	0.36 $p = 0.0001$	-0.11 $p = 0.27$
T4		1.00 $p = 0.00$	0.20 $p = 0.04$
TSH			1.00 $p = 0.00$

TABLE 7A-5. PEARSON'S r CORRELATIONS (n = 119) BETWEEN THYROID HORMONES AND TSH OF THE 90-DAY DATA OF THE SPRINGBORN LABORATORIES, INC. (1998) SUBCHRONIC RAT STUDY

	T3	T4	TSH
T3	1.00 p = 0.00	0.66 p = 0.0001	−0.40 p = 0.0001
T4		1.00 p = 0.00	−0.38 p = 0.0001
TSH			1.00 p = 0.00

**TABLE 7A-6. PEARSON'S r CORRELATIONS (n = 22 to 27) BETWEEN THYROID HORMONES AND TSH FOR THE F1 RAT PUPS ON PND5 IN THE DEVELOPMENTAL NEUROTOXICITY STUDY
(Argus Research Laboratories, Inc., 1998a)**

	T3	T4	TSH
T3	1.00 p = 0.00	0.87 p = 0.0001	−0.43 p = 0.03
T4		1.00 p = 0.00	−0.57 p = 0.0046
TSH			1.00 p = 0.00

1   In general, positive correlations were expected between T3 and T4 and between TSH and

2   the histopathology rating. Negative correlations were expected between T4 and TSH and

3   between T4 and histopathology.

4   Figure 7A-1 shows the correlations between T3 and T4 and between T4 and TSH levels

5   from the 14-day Caldwell et al. (1995) study in rats. Robust relationships are illustrated:

6   a positive correlation is shown between T3 and T4; whereas, the T4 and TSH varied inversely.

7   Hormone levels also correlated highly with decrease in follicular lumen size. Figure 7A-2 shows

8   the rank of T4 level and TSH level versus the severity rating for follicular lumen size to be highly

9   correlated inversely. Figure 7A-3 shows the correlations for the combined 14-day and 90-day

10   time points (male and female) from the subchronic study performed in rats (Springborn
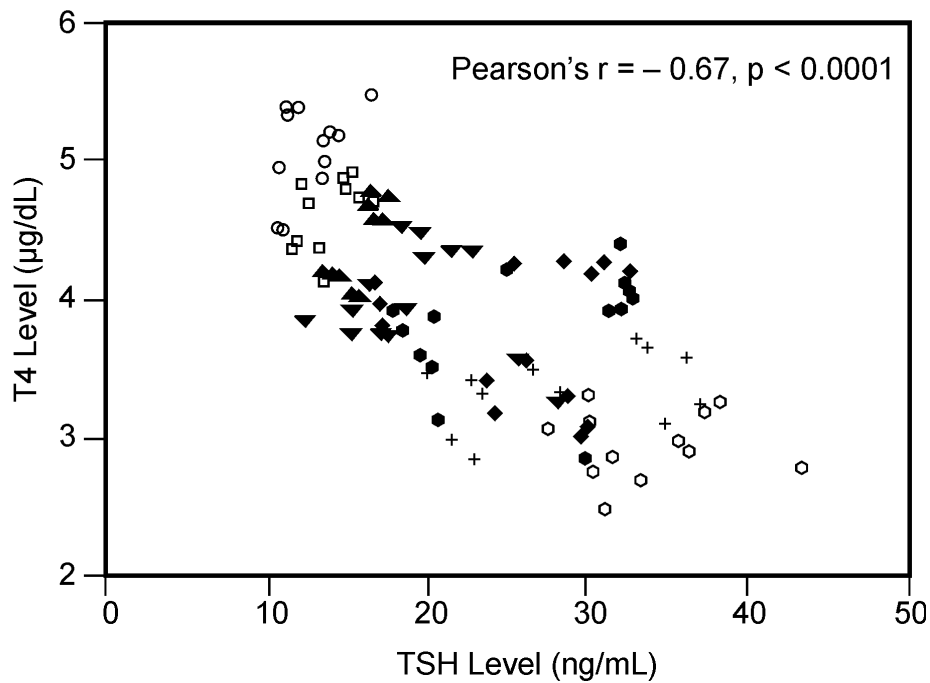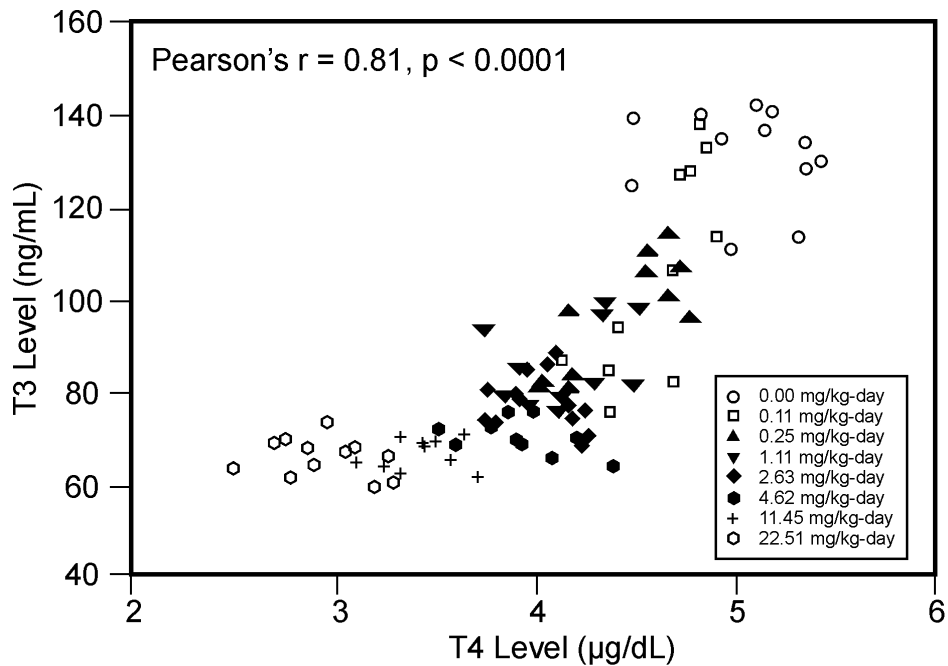
Figure 7A-1. Correlations between T3 versus T4 (top panel) and T4 versus TSH (bottom panel) in rats of the 14-day Caldwell et al. (1995) study (Geller, 1998a). Data of Channel (1998a) and Crofton (1998a).
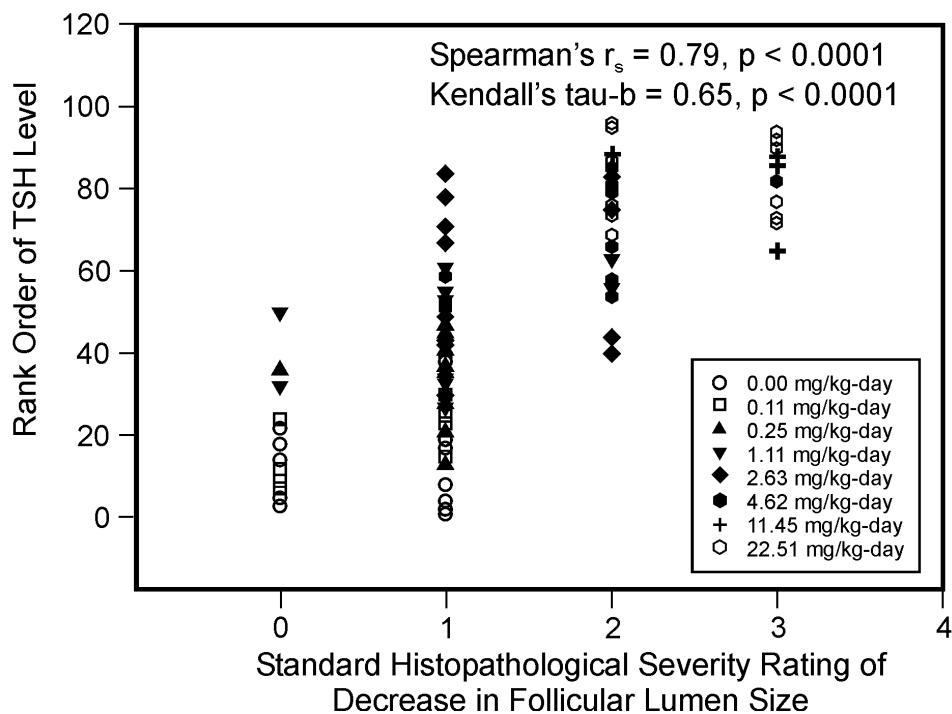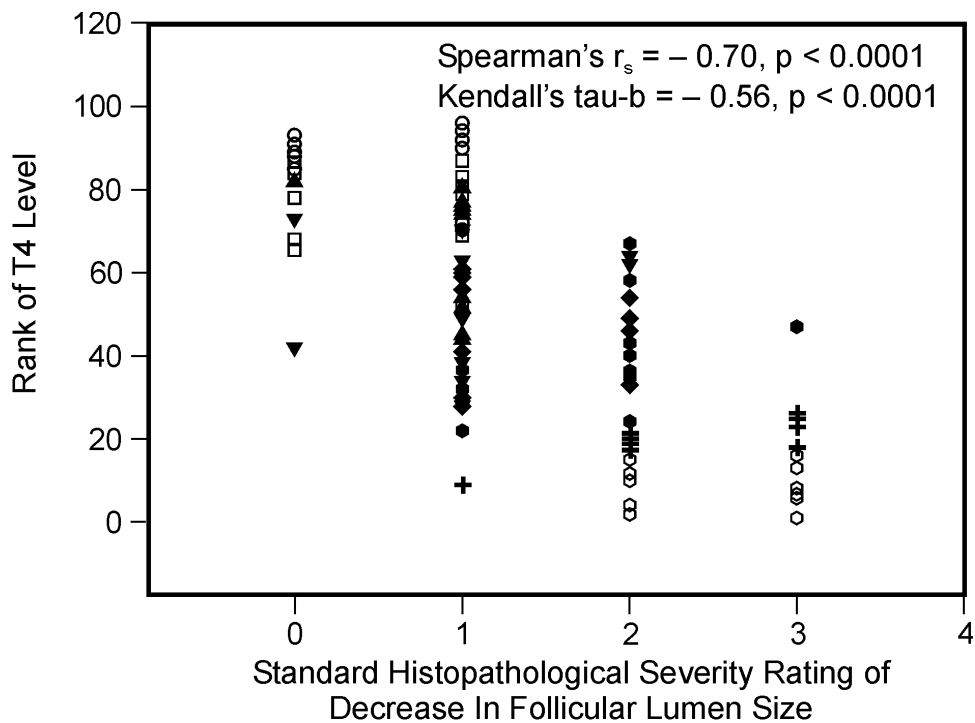
Figure 7A-2. Correlations between the rank order of T4 (top panel) and TSH (bottom panel) versus decrease in follicular lumen size in rats of the 14-day Caldwell et al. (1995) study (Geller, 1998a). Data of Channel (1998a) and Crofton (1998a).

1   Laboratories, Inc., 1998). As shown in Figure 7A-3 (top panel), T3 and T4 were highly

2   significantly correlated, with low levels of T3 and T4 associated with high doses. Both T4 and

3   TSH were significantly negatively correlated (bottom panel). After 14-days of dosing

4   (Figure 7A-4), T3 and T4 are highly associated (top panel), but there is an unexpected positive

5   relation between T4 and TSH (bottom panel). At the 90-day time point, there are the expected

6   strong correlations between T3 and T4 (Figure 7A-5, top panel) and between T4 and TSH

7   (bottom panel).

8   Correlations also were performed on the data from the neurodevelopmental study for the

9   PND5 pups (Argus Research Laboratories, Inc., 1998a). T3 and T4 were strongly positively

10   correlated, and T4 and TSH were negatively correlated (Figure 7A-6). Figure 7A-7 (top panel)

11   shows that T4 is negatively associated with a significant decrease in lumen area. Figure 7A-7

12   (bottom panel) also shows that TSH is positively correlated with a decrease in lumen size.

13   In total, these correlations lent strong support to the mapping model proposed. Strong

14   correlations were observed between T3 and T4 levels, T3 or T4, and TSH levels, and hormone

15   levels and a decrease in thyroid lumen size. These relationships were most definitive in the

16   Caldwell et al. (1995) study, in which strong correlations existed between the elements of the

17   thyroid hormone homeostasis feedback loop and between hormone levels and severity ratings for

18   lumen size decrease as a measure of thyroid histopathology. In the subchronic (Springborn

19   Laboratories, Inc., 1998) study, correlations were established between hormone levels across

20   both the 14- and 90-day dosing points and for each time point individually. At 14 days of dosing,

21   the expected inverse relationship between T4 and TSH was not found. At the 90-day dosing

22   point, the inverse relationships between T3 or T4 and TSH were found.

23   Similar relationships were observed in pups on PND5 of the developmental neurotoxicity

24   study (Argus Research Laboratories, Inc., 1998a; York, 1998c). The T4 and TSH were

25   significantly correlated negatively, as expected. The T3, T4, and TSH were all significantly

26   correlated with decrease in lumen size. The correlations in the rat studies support the model that

27   manipulations resulting in decreased levels of circulating thyroid hormone are linked to thyroid

28   histopathological changes that are thought to result directly from elevation of TSH.

29
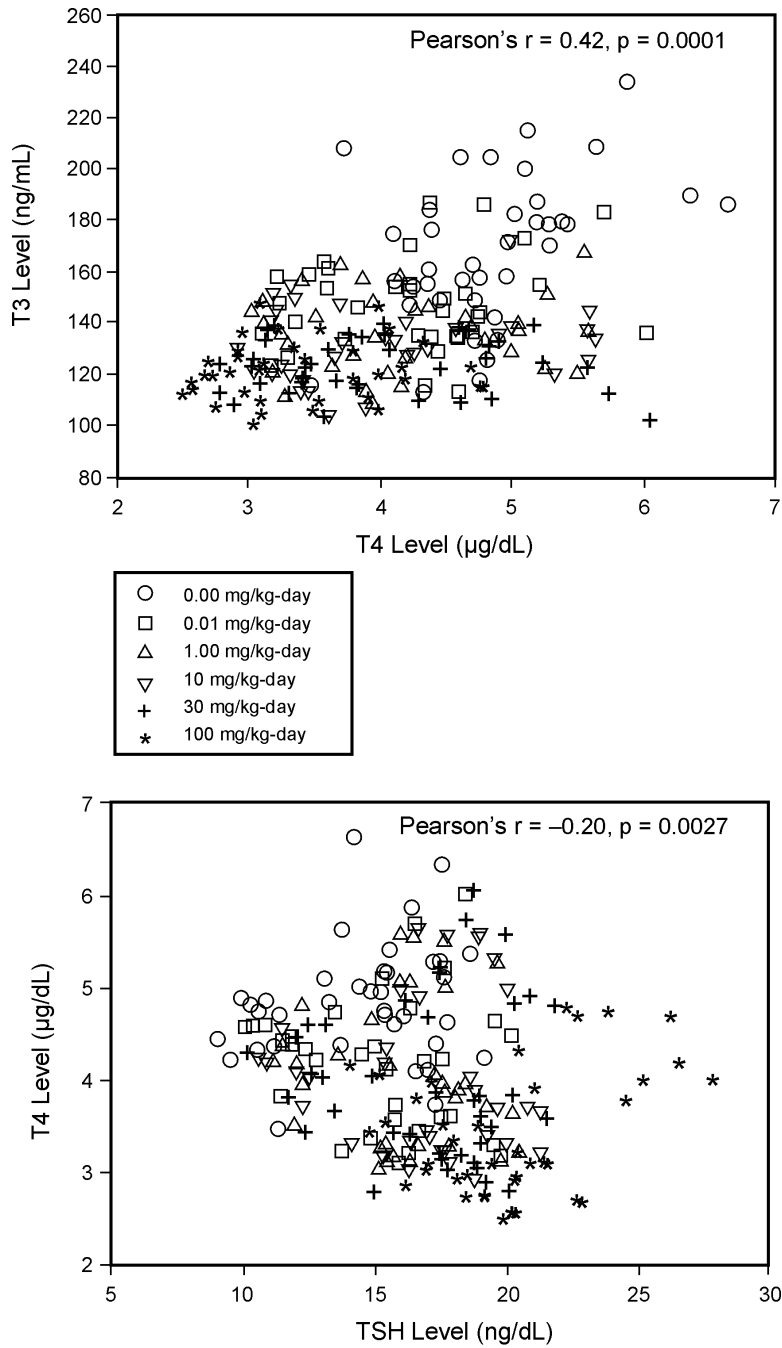
Figure 7A-3. Correlations between T3 versus T4 (top panel) and T4 versus TSH (bottom panel) for the combined male and female data of the 14-day and 90-day time points from the Springborn Laboratories Inc. (1998) subchronic study (Geller, 1998a).
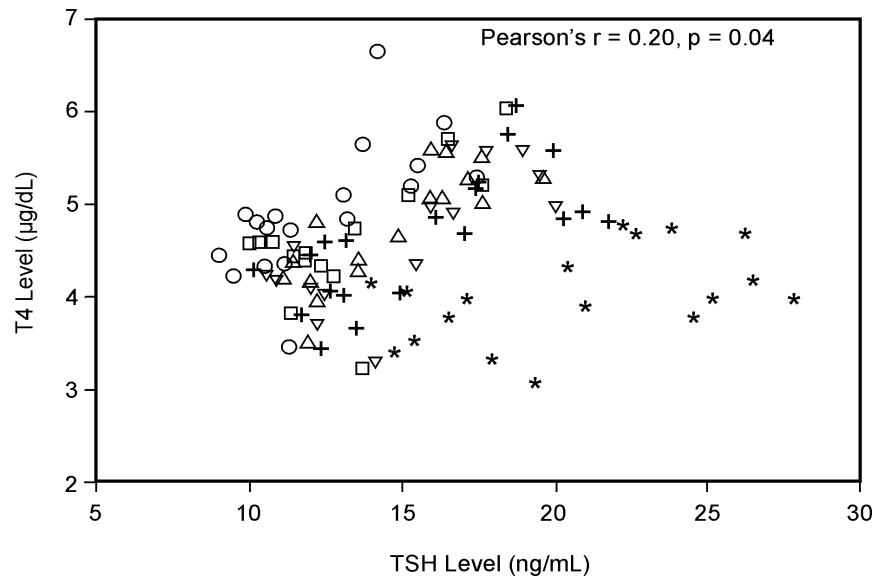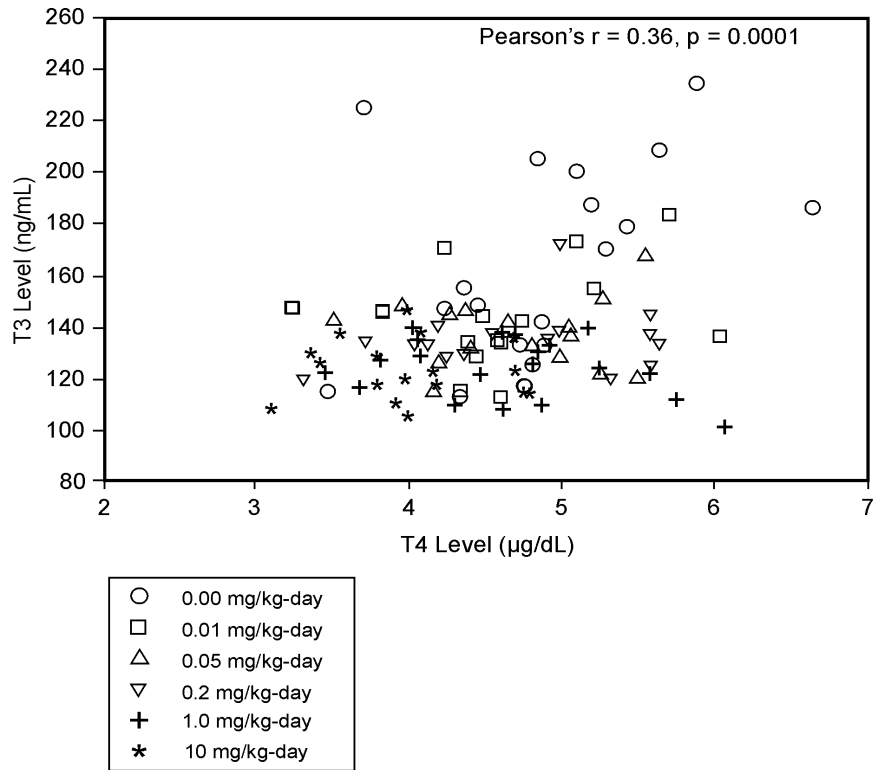
Figure 7A-4. **Correlations between T3 versus T4 (top panel) and T4 versus TSH (bottom panel) for the combined male and female data of the 14-day time point from the Springborn Laboratories Inc. (1998) subchronic study in rats (Geller, 1998b).**
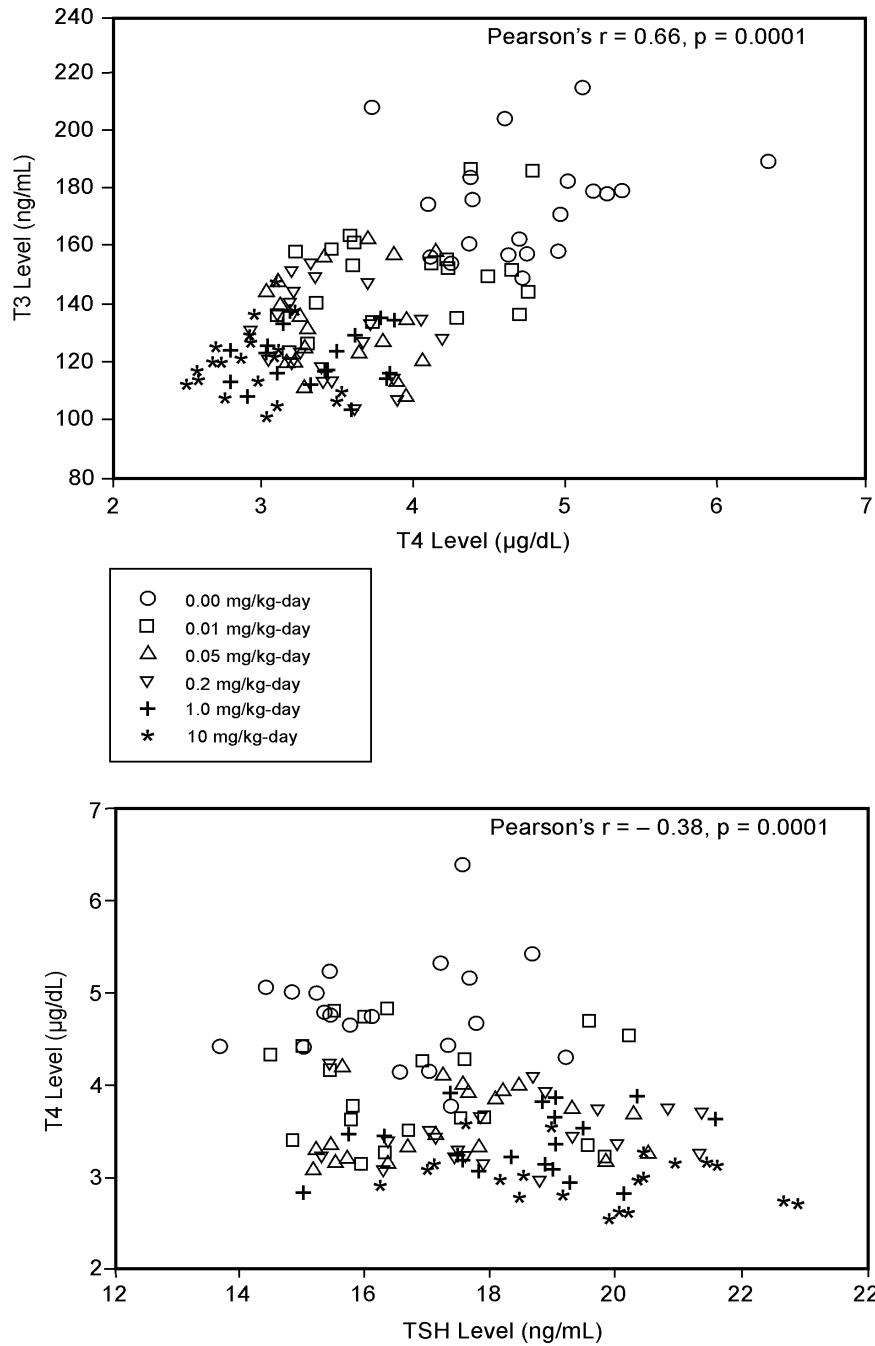
Figure 7A-5. Correlations between T3 versus T4 (top panel) and T4 versus TSH (bottom panel) for the combined male and female data of the 90-day time point from the Springborn Laboratories Inc. (1998) subchronic study in rats (Geller, 1998b).
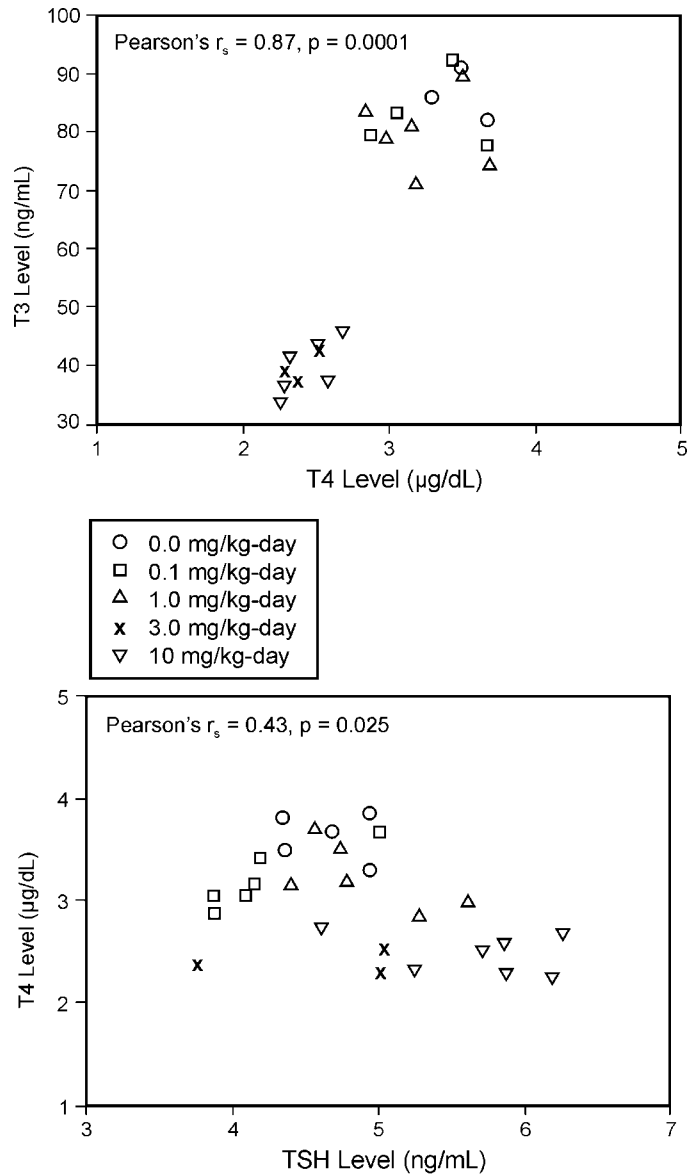
Figure 7A-6. **Correlations between T3 versus T4 (top panel) and T4 versus TSH (bottom panel) for the F1-generation rat pups on PND5 in the developmental neurotoxicity study (Geller, 1998b). Data of Argus Research Laboratories, Inc. (1998a), York (1998c), Channel (1998c), and Crofton (1998f).**
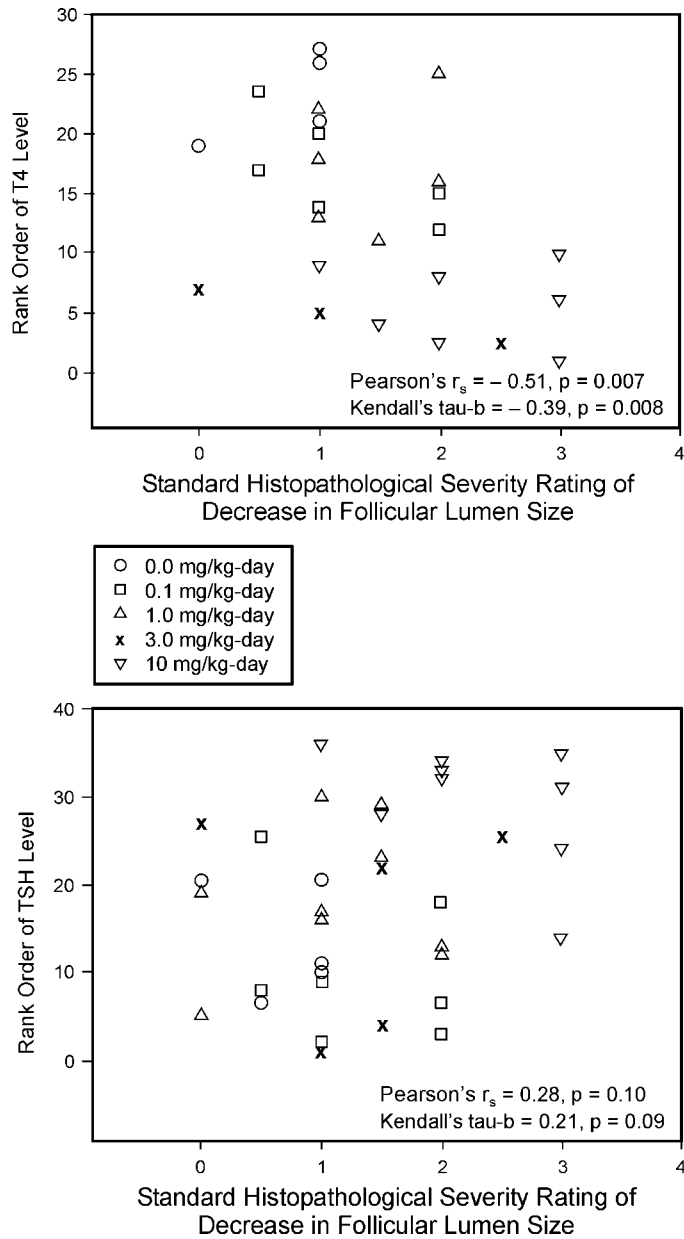
Figure 7A-7. Correlations between the rank order of T4 (top panel) and TSH (bottom panel) versus histopathology severity rating of the decrease in follicular lumen size for the postnatal day 5 (PND5) pups in the 1998 neurodevelopmental study (Geller, 1998b). Data of Argus Research Laboratories, Inc. (1998b), Channel (1998c), and Crofton (1998e, f).

Appendix 7B

Benchmark Dose Statistics for Hormone Analyses

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5   As mentioned in Chapter 5, benchmark dose analyses were performed in addition to the

6   ANOVA for all hormone data. Benchmark analysis of the 2001 "Effects Study" is presented in

7   Geller (2001c). This appendix presents analyses performed on the other data sets provided in the

8   1998 assessment.

9   For the continuous hormone data, the BMD and BMDL estimates were calculated using a

10   variety of benchmark response (BMR) values. Generally, the BMR was equal to a response 10%

11   less than the control mean (i.e., 10% of the actual control response was subtracted from the

12   estimate of the control value generated by the fit to the data). This is a less rigorous standard

13   than the (control minus 5% of control) BMR that provided a close match to NOAELs in the

14   evaluation of BMD for developmental toxicity by Kavlock et al. (1995) although this may be

15   warranted because other endpoints (thyroid hormone and histopathology) are being evaluated.

16   For the natural log (ln) transformed data, this means subtracting the constant 0.1053 from the

17   control value, which is equivalent to multiplying the control value by 0.90. The BMD and

18   BMDLs at 20 and 30% less than control and control standard deviations also are provided as a

19   yardstick for evaluating how other clinical criteria may affect the estimates. Hormone data were

20   fit with polynomial (linear or quadratic) or power functions (Table 7B-1).

21

22

TABLE 7B-1. CONTINUOUS FUNCTIONS USED IN BENCHMARK DOSE (BMD) MODELING

Power function	$f(dose) = control + slope * dose^{power}$
Polynomial function	$f(dose) = \beta0 + \beta1 * dose + \beta2 * dose^2 + ...$
(includes linear and quadratic)	

1   Adequacy of fit for continuous data was evaluated by the statistical goodness-of-fit

2   ($-2 \times$ log likelihood ratio) test provided by the EPA BMD program output, visual comparison,

1	and whether the fit was biologically plausible. The latter criterion in most cases,
2	non-monotonicities in the function fit to the data, precluded a fit from consideration. In general,
3	the second order quadratic fits suffered from minima or maxima between the data points from the
4	two highest data points in a given experiment. This consideration also precluded the use of
5	polynomials of higher than second order because these higher order polynomials generally had a
6	local maxima or minima between data points (dose levels) and did not model the data plausibly.
7	It should be noted that the interpretation of the test for constant variance included in the output of
8	the version of the BMD software (version 0.96) was not reliable.
9	

7B.1 Benchmark Dose Estimates Submitted to U.S. Environmental Protection Agency

12	Two sets of BMD calculations were derived from the Caldwell et al. (1995) 14-day study
13	and submitted to the EPA (Dollarhide and Dourson, 1997). One set was calculated for TSH and
14	T4 levels for males and females separately using the THC (polynomial fit) module of the Crump
15	software, and the model coefficients were restricted to be nonnegative to prevent
16	non-monotonicity. This resulted in linear fits to curvilinear data, and the fits were judged to be
17	poor by both visual inspection and statistical goodness-of-fit criteria (Geller, 1998a).
18	An alternative approach to calculating BMD estimates based on additional risk also was
19	derived using the Kodell-West algorithm (Kodell-West, 1993). This model generates a quadratic
20	fit to the dose-response data using a maximum likelihood estimator, defines an adverse effect
21	level based on the variability present in the data, and then calculates additional risk. The EPA
22	recalculated these fits using Kodell's SAS® program (Geller, 1998a). The EPA estimates
23	correspond to those previously reported, as shown in Table 7B-2 of Appendix 7B. The
24	coefficients of the fits are provided in Table 7B-3. None of the fits to the data reached statistical
25	significance, and all contain minima (T3 and T4) or maxima (TSH) within the dose range tested.
26	Again, the lack of fit raises difficulties with interpretation and suggests that these estimates
27	should not be used as the basis for risk assessment. The EPA also calculated BMD estimates on
28	ln-transformed data because the Kodell-West algorithm assumes constant variance, and the
29	transformed data is more likely to fit this assumption. The BMD estimates calculated with the ln
30	transform, however, were virtually identical to those of the previous estimates.
31	

**TABLE 7B-2. BENCHMARK DOSE (BMD) ESTIMATES FOR MALE HORMONE
DATA OF CALDWELL et al. (1995) 14-DAY RAT STUDY, USING
KODELL-WEST ALGORITHM**

Responders	BMD Associated with 1% Additional Risk (mg/kg-day)		BMD Associated with 10% Additional Risk (mg/kg-day)		BMD:N(L)OAEL 1%; 10%
TSH	EPA[a]	D&D, 1997[b]	EPA[a]	D&D, 1997[b]	1.11
k = 3	0.832	0.823	2.078	2.074	0.75; 1.87
k = 2	0.176	0.172	0.972	0.970	0.16; 0.88
ln TSH					1.11
k = 3		0.845		2.115	0.76; 1.91
k = 2		0.181		0.987	0.16; 0.89
T3	EPA[a]	D&D, 1997[b]	EPA[a]	D&D, 1997[b]	0.11[c,d]
k = 3	0.980	0.983	2.485	2.495	8.1; 22.59
k = 2	0.209	0.207	1.146	1.151	1.9; 10.42
lnT3					0.11[c,d]
k = 3		0.891		2.244	8.1; 20.4
k = 2		0.190		1.042	1.73; 9.47
T4	EPA[a]	D&D, 1997[b]	EPA[a]	D&D, 1997[b]	0.11[c,d]
k = 3	0.797	0.658	1.969	1.639	7.25; 17.9
k = 2	0.172	0.136	0.927	0.774	1.56; 8.43
ln (T4)					0.11[c,d]
k = 3		1.002		2.490	9.11; 22.64
k = 2		0.215		1.169	1.95; 10.63

[a]EPA refers to BMD estimates calculated using SAS® software received from Dr. Ralph Kodell for Kodell-West
 calculations (Geller, 1998a).
[b]D&D refers to BMDs included in Dollarhide and Dourson (1997).
[c]LOAEL; otherwise, value indicates NOAEL.
[d]LOAEL from combined male and female.

TABLE 7B-3. COEFFICIENTS AND GOODNESS-OF-FIT STATISTICS OF KODELL-WEST (QUADRATIC POLYNOMIAL) MODEL FITS TO MALE HORMONE DATA OF CALDWELL et al. (1995) 14-DAY RAT STUDY[a]

Responders	B0	B1	B2	Dose (mg/kg-day) of Global Max/Min	p of Fit2[b]
TSH	17.182	2.895	-0.0914	max: 15.84	<0.00001
ln TSH	2.825	0.1269	-0.004202	max: 15.11	<0.00001
T3	112.871	-8.987	0.3169	min: 14.18	<0.00001
lnT3	4.7114	-0.09702	0.0034	min: 14.27	<0.00001
T4	4.7712	-0.1791	0.00445	min: 20.11	<0.00001
ln (T4)	1.563	-0.0414	0.0009	min: 23.00	0.00012

[a]Coefficients generated by using SAS software received from Dr. Ralph Kodell (Geller, 1998a). Identical coefficients were generated by using EPA BMD software.
[b]$p > 0.05$ denotes significant fit. Goodness-of-fit derived using -2 log (likelihood ratio) test from EPA BMD software (see Geller, 1998a).

7B.2 U.S. Environmental Protection Agency Benchmark Dose Estimates for Thyroid and Pituitary Hormones

The hormone data from the Caldwell et al. (1995) subchronic (Springborn Laboratories, Inc., 1998) and rabbit developmental studies (Argus Research Laboratories, Inc., 1998c) were best fit by unrestricted power functions. The hormone data from the developmental neurotoxicity study (Argus Research Laboratories, Inc., 1998a; York, a,b,c,d,e) and mouse immunotoxicity study (Keil et al., 1998) were fit by either unrestricted power or polynomial functions. It is noted that the unrestricted power function fits generally have an extremely high slope as dose approaches zero. Tables 7B-4 through 7B-14 provide the statistics for each study.

Many of the BMDL estimates derived from these studies were lower than the NOAEL or LOAEL values derived by ANOVA, particularly those derived from power function fits. Murrell et al. (1998) suggested that this occurs when sampling statistics (i.e., small group sample sizes and few dose groups) play a large role in inflating NOAELs while depressing BMDL estimates. This may be the case for some of the data examined herein. Murrell et al. (1995) suggested that under such conditions using the BMD point estimate, rather than the lower confidence limit, would be a more accurate representation of the dose-response behavior.

TABLE 7B-4. BENCHMARK DOSE (BMD) ESTIMATES USING POWER FUNCTION FIT TO COMBINED MALE AND FEMALE HORMONE DATA OF CALDWELL et al. (1995) 14-DAY RAT STUDY (Benchmark response based on 10% change from control value.)

Endpoint	p of Fit	BMD	BMDL	NOAEL/ LOAEL	BMD: N(L)OAEL	BMDL: N(L)OAEL	BMR: 10% control SD
TSH[a]	0.272	0.014	0.0002	0.44	0.032	4.55e-4	1.29 1.88
ln TSH[a]	0.099	0.017	0.002	0.44	0.039	4.55e-3	−0.1053
Female TSH[b]	0.077	0.19	0.032	0.1	1.90	0.32	1.125 0.48
Female ln(TSH)[a]	0.50	0.078	0.035	0.1	0.78	0.35	−0.1053
Male TSH	No significant fits to male TSH or male ln(TSH) data						
T3[a]	0.107	0.00035	0.00	0.1[c]	0.0035	NA	13.07 10.21
lnT3[a]	0.091	0.0004	2e-6	0.1[c]	0.004	2.00e-5	−0.1053
T4[a]	0.303	0.243	0.096	0.1[c]	2.43	0.96[c]	0.506 0.321
ln (T4)[d]	0.172	0.340	0.0997	0.1[c]	3.40	1.00[c]	−0.1053

[a]Unrestricted quadratic: fit nonmonotonic, not significant. Restricted polynomial (linear): fit not significant.
[b]Unrestricted quadratic: fit monotonic but not significant. Restricted polynomial (linear): fit not significant.
[c]LOAEL; otherwise, value is NOAEL.
[d]Unrestricted quadratic: fit not significant, global minimum at approximate high dose. Restricted polynomial (linear): fit not significant.

1   The BMD estimates calculated with a benchmark response of 10% less than control on the

2 TSH hormone dose-response data are spread over 2.5 orders of magnitude, a similar range to that

3 seen in the distribution of NOAELs calculated for TSH. The BMDL estimates are distributed

4 more widely, over 5 orders of magnitude. These reflect the steepness of the confidence limits on

5 the slope at low doses.

6   The T3 BMD estimates are spread over approximately two orders of magnitude, similar to

7 the variability seen across studies in the LOAEL and NOAEL estimates. The T3 BMD estimates

8 are 100-fold lower than the NOAEL/LOAEL estimates, however. A BMDL could be calculated

**TABLE 7B-5. BENCHMARK DOSE (BMD) AND BMD 95% LOWER LIMIT (BMDL)
ESTIMATES USING POWER FUNCTION FIT TO COMBINED MALE AND FEMALE
HORMONE DATA OF CALDWELL et al. (1995) 14-DAY RAT STUDY
(Benchmark response based on 10, 20, and 40% changes from control value.)**

Endpoint	p of Fit	BMD BMDL (10%)	BMD BMDL (20%)	BMD BMDL (40%)	Mean	NOAEL
TSH	0.272	0.014 0.0002	0.083 0.0038	0.507 0.0604	12.861	0.44
ln(TSH)[a]	0.099	0.002	0.043	1.11		0.44
T3	0.0108	0.00035 0.00	0.0338 0.000036	3.27 0.042[c]	130.69	0.10[b]
ln(T3)[a]	0.091	0.000002	0.000642	0.478		0.10[b]
T4	0.303	0.243 0.096	2.28 1.299	21.44 16.78	5.06	0.10[b]
ln(T4)[a]	0.172	0.100	1.213	16.89		0.10[b]

[a]For ln-transformed data, only BMDL estimates are displayed.
[b]LOAEL, not NOAEL.
[c]BMDL calculation failed at some values. This means BMDL value may not be accurate.

1 for only one of the data sets, and this value was approximately 10,000 times lower than the

2 LOAEL. The BMD estimates comprising the 25th to 75th percentiles for T4 cover the same

3 2.5 orders of magnitude as those covered by the NOAEL and LOAEL estimates for T4. The

4 BMDL estimates for this same percentile range are distributed a little more widely, but do

5 include the range of T4 NOAEL and LOAEL estimates.

6

7B.3   Summary of U.S. Environmental Protection Agency Benchmark Dose Analyses

9   The BMD analyses of previously reported estimates for the hormone data of Caldwell et al.,

10 (1995)14-day study in rats (Dollarhide and Dourson, 1997) were shown to be based on

11 inadequate model fits. The EPA was able to successfully model the hormone data. However,

12 these estimates raised a number of issues with respect to approaches for these types of data.

13 An alternative may be to pursue a model form of the Hill equation which recently has been used

14 for endocrine disruption data (Barton et al., 1998).

TABLE 7B-6. BENCHMARK DOSE (BMD) AND BMD 95% LOWER LIMIT (BMDL) ESTIMATES FOR COMBINED MALE AND FEMALE HORMONE DATA OF 14-DAY TIME POINT IN THE SPRINGBORN LABORATORIES, INC. (1998) SUBCHRONIC STUDY (Benchmark response based on 10% change from control value.)

Endpoint	Model	p of Fit	BMD	BMDL	NOAEL/ LOAEL	BMDL: N(L)OAEL	BMD: N(L)OAEL	BMR: 10% control SD
TSH	Power	0.45	0.037	0.000075	0.01	0.0075	3.7	1.26
	Quadratic	0.069	Fit significant, but not monotonic		0.01			2.52
ln TSH	Power	0.43	0.043	Could not calculate	0.01	NA	4.3	−0.1053
	Quadratic	Fit not significant, nonmonotonic			0.01			
T3	Power	0.41	0.000033	Lower limit includes 0	0.01[a]	NA	0.0033	16.65 38.51
	Quadratic	Fit not significant, nonmonotonic			0.01[a]			
lnT3	Power	0.35	0.000168	Lower limit includes 0	0.01[a]	NA	0.0168	−0.1053
	Quadratic	Fit not significant, nonmonotonic			0.01[a]			
T4	Power	0.203	1.16	0.0035	1.0	0.0035	1.16	0.506
	Quadratic[b]	0.12	3.27	1.09	1.0	1.09	3.27	0.603
ln (T4)	Power	0.22	1.64	0.04	1.0	0.04	1.64	−0.1053
	Quadratic[b]	0.16	3.25	1.06	1.0	1.06	3.25	

[a]LOAEL; otherwise, value is NOAEL.
[b]Global minimum of quadratic function is at dose ≈9.50 mg/kg-day.

TABLE 7B-7. BENCHMARK DOSE (BMD) AND BMD 95% LOWER LIMIT (BMDL) ESTIMATES FOR COMBINED MALE AND FEMALE HORMONE DATA OF 14-DAY TIME POINT IN THE SPRINGBORN LABORATORIES, INC. (1998) SUBCHRONIC STUDY
(Benchmark response based on 10, 20, and 40% changes from control value.)

Endpoint	Model	p of Fit	BMD BMDL (10%)	BMD BMDL (20%)	BMD BMDL (40%)	Mean	NOAEL
T4	Power	0.203	1.16 0.0035	12.73 1.21	138.94 38.33	5.066	1.0
ln(T4)	Power	0.22	0.037	3.899	36.48		1.0
T3	Power	0.41	0.000033 —	0.207 —	129.39 0.129[a]	166.5	0.01[b]
ln(T3)	Power	0.35	Lower limit includes 0	0.000054[a]	43.16[a]		0.01[b]
TSH	Power	0.45	0.037 0.000076	0.326 0.005	2.89 0.36	12.616	0.01
ln(TSH)	Power	0.43	0.0015	0.098	6.587		0.01

[a]BMDL calculation failed at a number of values. This means BMDL value may not be accurate.
[b]LOAEL, not NOAEL.

TABLE 7B-8. BENCHMARK DOSE (BMD) ESTIMATES FOR COMBINED MALE AND FEMALE HORMONE DATA OF 90-DAY TIME POINT IN THE SPRINGBORN LABORATORIES, INC. (1998) SUBCHRONIC STUDY
(Benchmark response based on 10% change from control value.)

Endpoint	p of Fit	BMD	BMDL	NOAEL/ LOAEL	BMD: N(L)OAEL	BMDL: N(L)OAEL	BMR: 10% Control SD
TSH[a]	0.42	0.269	0.018	0.05	5.38	0.36	1.633 1.464
ln TSH[a]	0.40	0.492	0.0796	0.05	9.84	1.6	−0.1053
T3[a]	0.01	No fit	No fit	0.01[b]	NA	NA	17.50 18.924
lnT3[a]	0.01	No fit	No fit	0.01[b]	NA	NA	NA
T4[a]	0.14	6e-6	Lower limit includes 0	0.01[b]	6e-4	NA	0.475 0.576
ln (T4)[a]	0.17	1.10e-5	0.00	0.01[b]	1.1e-3	∞	−0.1053

[a]Unrestricted quadratic: fit nonmonotonic, not significant. Restricted polynomial (linear): fit not significant.
[b]LOAEL; otherwise, value is NOAEL.

**TABLE 7B-9. BENCHMARK DOSE (BMD) AND BMD 95% LOWER
LIMIT (BMDL) ESTIMATES FOR COMBINED MALE AND FEMALE
HORMONE DATA OF 90-DAY TIME POINT IN THE SPRINGBORN
LABORATORIES, INC. (1998) SUBCHRONIC STUDY
(Benchmark response based on 10, 20, and 40% changes from control value.)**

	Model	p of Fit	BMD BMDL (10%)	BMD BMDL (20%)	BMD BMDL (40%)	Mean	NOAEL
T4	Power	0.14	0.000006 —	0.01 0.000001	15.09 0.52[a]	4.75	0.01[b]
ln(T4)	Power	0.165	0.00	0.004	4.87		0.01[b]
T3	Power	0.01		No significant fit		174.96	0.01[b]
ln(T3)	Power	0.01		No significant fit			0.01[b]
TSH	Power	0.43	0.272 0.019	8.808 2.404	285.52 73.80	16.33	0.05
ln(TSH)	Power	0.40	0.082	7.94	405.14		0.05

[a]BMDL calculation failed at a number of values. This means BMDL value may not be accurate.
[b]LOAEL not NOAEL.

TABLE 7B-10. BENCHMARK DOSE (BMD) AND BMD 95% LOWER LIMIT (BMDL) ESTIMATES FOR HORMONE AND THYROID MORPHOMETRY DATA OF F1-GENERATION PUPS AT PND5 IN THE DEVELOPMENTAL NEUROTOXICITY STUDY
(Argus Research Laboratories, Inc., 1998a, and Channel, 1998c)[a]
(Benchmark response based on 10% change from control value.)

Endpoint	Model	p of Fit	BMD	BMDL	NOAEL or LOAEL	BMD: N(L)OAEL	BMDL: N(L)OAEL	BMR: 10% Control SD
TSH	Linear	0.50	4.64	3.77	3.0	1.55	1.26	0.45 0.465
	Power	0.31	4.48	1.43	3.0	1.49	0.48	
ln TSH	Linear	0.48	5.51	4.43	3.0	1.84	0.54	−0.1054
	Power	0.30	5.03	2.11	3.0	1.68	0.70	
T3	Neither linear, quadratic, or power FCNS fit data	<0.00001 for all	No fit	No fit	0.1	NA	NA	
lnT3	Neither linear, quadratic, or power FCNS fit data	<0.00001 for all	No fit	No fit	0.1	NA	NA	
T4	Nonmonotonic quadratic significant fit	0.50 min = 7.45 mg/kg	*1.26*	*0.98*	*1.0*	*1.26*	*0.98*	0.341 0.370
ln (T4)	Nonmonotonic quadratic significant fit	0.50 min = 7.14 mg/kg	*1.18*	*0.92*	*1.0*	*1.18*	*0.92*	
Morphometry	Control-10% Control (=31.78); SD = 0.37 Nonmonotonic quadratic significant fit Power FCN BMDL interval includes 0.00	0.19 global min = 6.81 mg/kg	*1.053*	*0.644*	*1.00*	*1.053*	*0.644*	
ln (morph)	Control-10% Control (= 0.341); SD = 0.37 Nonmonotonic quadratic significant fit Power FCN BMDL computational failures	0.19 global min = 7.01 mg/kg	*0.822*	*0.538*	*1.00*	*0.822*	*0.538*	

[a]Italics denote estimates derived from nonmonotonic fits to data. FCN = function and SD = standard deviation.

TABLE 7B-11. BENCHMARK DOSE (BMD) AND BMD 95% LOWER LIMIT (BMDL) ESTIMATES FOR HORMONE DATA OF F1-GENERATION PUPS AT PND5 IN THE DEVELOPMENTAL NEUROTOXICITY STUDY (Argus Research Laboratories, Inc., 1998a, and Channel, 1998c) (Benchmark response based on 10, 20, and 40% changes from control value.)

	p of Fit	BMD BMDL (10%)	BMD BMDL (20%)	BMD BMDL (40%)	Mean	NOAEL
T4	0.50[a]	1.26[a] 0.973[a]	2.89[a] 2.16[a]	BMD set to[a] 1,000[a]	3.41	1.0
ln(T4)	0.50[a]	0.92[a]	NC[a]	NC[a]		1.0
T3	<0.00001	NC	NC	NC	87.97	0.1
ln(T3)	<0.00001		NC	NC		0.1
TSH	0.50	4.64 3.77	9.30 7.55	18.61 15.10	4.51	3.0
ln(TSH)	0.48	NC	NC	NC		3.0

[a]Underlined values from nonmonotonic fits to data. (NC = not computed.) The BMDL calculation failed at a number of values. This means BMDL value may not be accurate.

TABLE 7B-12. BENCHMARK DOSE (BMD) AND BMD 95% LOWER LIMIT (BMDL) ESTIMATES USING THE LINEAR MODEL FIT TO THE MOTOR ACTIVITY DATA OF F1-GENERATION PUPS AT PND14 IN THE DEVELOPMENTAL NEUROTOXICITY STUDY (Argus Research Laboratories, Inc., 1998a) (Benchmark response based on 10% change from control value.)

Endpoint	p of Fit	BMD	BMDL	NOAEL/ LOAEL	BMD: N(L)OAEL	BMDL: N(L)OAEL	BMR: 10% control SD
Movement[a]	0.72	1.94	1.04	None	NA	NA	24.45 162.75
Time[b]	0.69	1.33	0.66	None	NA	NA	18.60 184.78

[a]Number of movements.
[b]Time spent in activity.

TABLE 7B-13. BENCHMARK DOSE (BMD) AND BMD 95% LOWER LIMIT (BMDL) ESTIMATES USING THE POWER MODEL FIT TO THE HORMONE DATA OF FEMALE RABBITS ON GESTATION DAY 29 IN THE DEVELOPMENTAL STUDY (Argus Research Laboratories, Inc., 1998c) (Benchmark response based on 10% change from control value.)

Endpoint	p of Fit	BMD	BMDL	NOAEL/ LOAEL	BMD: N(L)OAEL	BMDL: N(L)OAEL	BMR
TSH, ln TSH						NA	No effect of dose
T3, ln T3						NA	No effect of dose
T4	0.06	0.54	Lower limit includes 0	0.1	5.4	NA	0.187
ln (T4)	0.0503	1.69	0.002	0.1	16.9	0.02	0.1053

TABLE 7B-14. BENCHMARK DOSE (BMD) AND BMD 95% LOWER LIMIT (BMDL) ESTIMATES USING THE POWER MODEL FIT TO THE HORMONE DATA OF FEMALE RABBITS ON GESTATION DAY 29 IN THE DEVELOPMENTAL STUDY (Argus Research Laboratories, Inc., 1998c) (Benchmark response based on 10, 20, and 40% changes from control value.)

	p of Fit	(10%)	(20%)	(40%)	Mean	NOAEL
T4	0.06	0.54 —	7.05 —	91.76 0.63	1.874	0.1
ln(T4)	0.05	1.69 0.0018	10.97 0.033	86.19 7.278		0.1
T3						No effect
ln(T3)						No effect
TSH						No effect
ln(TSH)						No effect

8. SCREENING ECOLOGICAL RISK ASSESSMENT FOR PERCHLORATE

8.1 INTRODUCTION

As discussed in Section 1.1, perchlorate salts including ammonium, potassium, sodium, and magnesium perchlorate, are manufactured as oxidizer components for propellants and explosives. The manufacture or use of perchlorate salts has been reported in most of the states of the continental United States (Figure 1-3). In some areas involved with the manufacture, use, or disposal of perchlorate salts, perchlorate, as the anion dissociated from these salts, has contaminated soils or ground or surface waters (Figure 1-4). These releases of perchlorate into the environment have been confirmed to have occurred in 20 states, clustered primarily in the southwestern United States where most sampling has occurred (Figures 1-3 and 1-4). Currently, there is a research need to determine whether perchlorate ion is causing any potential effects on ecosystems or ecosystem components. This chapter presents a screening-level ecological risk assessment of environmental contamination by perchlorate. In organization, it follows the Guidelines for Ecological Risk Assessment (U.S. Environmental Protection Agency, 1998c).

8.1.1 Management Goals and Decisions

The discovery that perchlorate release in some sites has contaminated ground and surface waters in certain locations has raised public and regulatory agency concerns. Much concern has focused on potential public exposures through drinking water and on the possible needs to improve analytical and treatment methods and to develop drinking water regulations (Section 1.4). Consequently, an extensive scientific assessment effort is underway to address those concerns (Section 1.5). A balanced approach requires assessing ecological effects as well. The goal of this screening-level ecological risk assessment is, therefore, to indicate the likelihood that adverse ecological effects (i.e., toxicity to specific organisms or effects on aquatic or terrestrial ecosystems) will result from observed levels of environmental contamination by perchlorate. The results of this assessment may be used to address the following questions:

1 • Are ecological risks best characterized as *de minimis* (exposures clearly are below levels of

2   concern), *de manifestis* (risks are clearly significant and require management action to reduce

3   exposures); or somewhere in between and requiring further characterization?

4 • Are analytical detection methods for determining levels of perchlorate in the environment

5   sufficient, or is it likely that adverse ecological effects occur at levels below current detection

6   limits?

7 • Is the available ecotoxicological information on perchlorate sufficient, or are additional studies

8   needed?

9

8.1.2 Scope, Complexity, and Focus

11   In the previous ERD version of this document (U.S. EPA, 1998d), the available

12 information for this ecological risk assessment was characterized as "very limited" and the

13 assessment was characterized as "screening-level." Information about the environmental levels

14 of perchlorate to which organisms were exposed and about its effects on diverse taxonomic

15 groups was practically nonexistent. Since then, additional information has become available that

16 improves the database in some respects. Most significantly, additional data are available on

17 effect levels in aquatic animals, an aquatic plant, a terrestrial plant, and a soil invertebrate; some

18 of these data are for chronic exposures. Effect levels in rodents have been reevaluated as part of

19 the human health risk assessment for perchlorate, and the ecological implications of those

20 changes are reflected herein. In addition, surveys have been conducted at several sites of known

21 or suspected perchlorate contamination, and environmental and biological materials have been

22 analyzed for perchlorate. Nonetheless, the level of knowledge on this issue must still be

23 characterized as limited because the number of species tested is still quite minimal, and the site

24 surveys focused only on the range of exposures at those sites. This ecological risk assessment is

25 therefore still a screening-level, rather than definitive, assessment. The materials used in the

26 1998 ERD and those that are new to this present draft, are described in this section.

27   **Interagency Perchlorate Steering Committee Report.** Perchlorate Ecological Risk

28 Studies is a report of the IPSC's Ecological Risk/Transport and Transformation Subcommittee,

29 dated November 13, 1998 (Interagency Perchlorate Steering Committee, 1998). This report

30 presents a literature review on perchlorate toxicity to nonmammalian organisms, recognizing that

31 few published studies exist, and a rationale for the selection of a battery of ecotoxicology tests

1   conducted for the USAF Armstrong Laboratory by EA Engineering, Science and Technology,

2   Inc. It then summarizes those test results, discusses the findings in the context of observed

3   exposures, discusses uncertainties, and makes recommendations for further study. The present

4   report constitutes a reevaluation of much of the same information from EPA's perspective,

5   except that EPA did not examine the open literature studies reviewed by the IPSC subcommittee.

6   **Test Battery Reports.** The EA Engineering, Science and Technology, Inc. (1998) final

7   report, *Results of Acute and Chronic Toxicity Testing with Sodium Perchlorate*, dated November

8   1998, details the test methods and results of the ecotoxicology battery. A follow-up report (EA

9   Engineering, Science and Technology, Inc., 2000) details the test methods and results from

10   additional chronic toxicity testing with the freshwater amphipod *Hyalella azteca* and the fathead

11   minnow *Pimephales promelas*.

12   **Block Environmental Services, Inc., Report.** The report, LC_{50} *Aquatic Toxicity Test*

13   *Results for Ammonium Perchlorate—A Two-Species Chronic Definitive Bioassay* (Block

14   Environmental Services, Inc., 1998) presents additional bioassay results that were not included in

15   the IPSC report.

16   **Algal Toxicity Testing.** The EA Engineering, Science and Technology, Inc. (1999) final

17   report, *Results of Algal Toxicity Testing with Sodium Perchlorate*, dated September 1999, details

18   the test methods and results of the ecotoxicological testing with the algae, *Selenastrum*

19   *capricornutum*.

20   **Frog Embryo Teratogenesis Assay: *Xenopus* (FETAX) Study.** The report, FETAX

21   Analysis of Ammonium Perchlorate (Dumont and Bantle, 1998), prepared by the Department of

22   Zoology, Oklahoma State University, and dated May 22, 1998, presents results of the Frog

23   Embryo Teratogenesis Assay: *Xenopus* (FETAX) conducted with ammonium perchlorate.

24   Recent data received by the EPA that the Agency has not yet fully reviewed indicate effects on

25   thyroid function, metamorphosis and sex ratio in developing *Xenopus laevis* (Goleman et al.,

26   2002). These data are made available with this document to the external peers for their review.

27   **Phytotransformation Study.** Two sets of studies report on the accumulation and potential

28   degradation of perchlorate by plants. The study, *Laboratory Characterization of*

29   *Phyto-transformation Products of Perchloroethylene (PCE), Trichloroethylene (TCE) and*

30   *Perchlorate* (Nzengung, n.d.; Nzengung et al., 1999), examined perchlorate distribution and

31   degradation in experimental systems containing sand, aqueous perchlorate solution, and rooted

1   cuttings of woody plant species. This study also examined systems containing chopped leaves or

2   microbial mats and aqueous perchlorate solution. A second study, *Potential Species for*

3   *Phytoremediation of Perchlorate* (Susarla et al., 1999a; Susarla et al., 2000a), reported

4   perchlorate depletion from test media over a ten day period by 13 vascular plant species and their

5   potential for phytoremediation of perchlorate contaminated sites.

6   **Biotransport Investigation Studies.** These studies assess the potential for

7   bioaccumulation of perchlorate in food webs by answering the question of whether perchlorate is

8   present in biological receptors. The report *Scientific and Technical Report for Perchlorate*

9   *Biotransport Investigation: A Study of Perchlorate Occurrence in Selected Ecosystems* (Parsons,

10   2001) examined perchlorate concentrations in site media and in various ecological receptors at

11   six sites with known or suspected perchlorate contamination: (1) sites associated with withdraw

12   of irrigation water from the Colorado River in the vicinity of Yuma, Arizona; (2) Las Vegas

13   Wash and Lake Mead near Las Vegas, Nevada; (3) Allegany Ballistics Laboratory, Rocket

14   Center, West Virginia; (4) Holloman Air Force Base in Otero County, New Mexico; (5) Naval

15   Surface Warfare Center, Indian Head, Maryland; and (6) Longhorn Army Ammunition Plant,

16   Karnack, Texas. Additional data are available for one of these sites, Longhorn Army

17   Ammunition Plant (LHAAP), Texas, in a paper published by Smith et al. (2001). In both studies,

18   ion chromatography with an AS-16 analytical column was used to measure for perchlorate

19   concentrations. Analyses with this analytical column have been shown to be superior than other

20   columns for detecting and quantifying perchlorate (Ellington and Evans, 2000; Susarla et al.,

21   2000b).

22   All these sites, except for those in the vicinity of Yuma, are associated with localized

23   contamination related to the manufacture, handling, or use of perchlorate in solid propellants.

24   The Yuma sites are approximately 250 miles downstream along the Colorado River from the Las

25   Vegas Wash and Lake Mead sites; the report suggests that there is no localized source of the

26   perchlorate; therefore, the most likely potential source of any perchlorate contamination in these

27   soils is believed to be Colorado River irrigation water. However, portions of the Yuma Proving

28   Grounds are drained by washes that pass near some of the agricultural locations sampled, and the

29   information provided in the report was not sufficient for ruling out the possibility of

30   contamination from the Yuma Proving Grounds.

31

8.2 PROBLEM FORMULATION

The characteristics of perchlorate and its sources are described earlier in this document (Chapters 1 and 2). Because this assessment is site independent, this problem formulation focuses on the selection of assessment endpoints, derivation of the conceptual model, and the analysis plan.

8.2.1 Assessment Endpoints

In ecological risk assessment, assessment endpoints are operational definitions of the environmental values to be protected. They are chosen based on policy goals and societal values, their ecological relevance, and their susceptibility to the stressor and are defined in terms of an entity and a property of that entity. The assessment endpoints for this ecological risk assessment are described in the following five subsections.

8.2.1.1 Fish Community Richness and Productivity

Fish communities are valued societally and are ecologically important. The productivity of these communities is important in terms of the support of fisheries. Species richness is important in terms of maintaining biodiversity. This importance is reflected by the use of species sensitivity distributions in the derivation of national ambient water quality criteria and the use of fish species richness as an important component of bioassessment procedures for enforcement of the Clean Water Act.

8.2.1.2 Aquatic Invertebrate Community Richness and Productivity

Aquatic invertebrate communities have little direct societal value but are important to energy and nutrient dynamics in aquatic ecosystems. The productivity of these communities is important in terms of trophic support of fisheries, of other groups of aquatic species, and of some terrestrial insectivores. Species richness is important in terms of maintaining biodiversity. This importance is reflected by the use of species sensitivity distributions in the derivation of national ambient water quality criteria and the use of invertebrate species richness as an important component of bioassessment procedures for enforcement of the Clean Water Act.

8.2.1.3 Aquatic Plant Richness and Productivity

Algae and other aquatic plants have little direct societal value but are important to energy and nutrient dynamics in aquatic ecosystems. Species richness is important in terms of maintaining biodiversity. Because of their importance to the trophic support of fisheries and other aquatic consumers, productivity is an important endpoint for this assemblage.

8.2.1.4 Soil Invertebrate Community Richness and Productivity

Soil invertebrate communities have little direct societal value, but, in nearly all terrestrial ecosystems, they are important to energy and nutrient dynamics and to maintenance of soil structure. The productivity of these communities is also important in terms of trophic support of some terrestrial insectivores. Species richness is important in terms of the policy of maintaining biodiversity.

8.2.1.5 Terrestrial Plant Richness and Productivity

Terrestrial plants are valued highly by society for production of food, fiber, and timber, as well as their aesthetic value. The primary valued property of terrestrial plants is their productivity. As autotrophs, plants are the basis of energy and nutrient dynamics in most terrestrial or aquatic food webs. Moreover, species richness is important in terms of the policy of maintaining biodiversity.

8.2.1.6 Population Productivity of Herbivorous Wildlife

Herbivorous wildlife are included as an endpoint entity because of the apparent bioconcentration of perchlorate in plant foliage. The meadow vole (*Microtus pennsylvanicus*) is used as a representative species for this group. Population productivity is used as the endpoint property because growth and reproduction are generally sensitive properties and because herbivores are valued for their production of food for human and nonhuman carnivores.

8.2.2 Conceptual Models

The conceptual model describes the relationships between sources of perchlorate and the endpoint receptors (Figure 8-1). Sources include spills during the flushing of rockets; the combustion of rocket fuel; the improper disposal of rocket fuel, open burn or open detonation
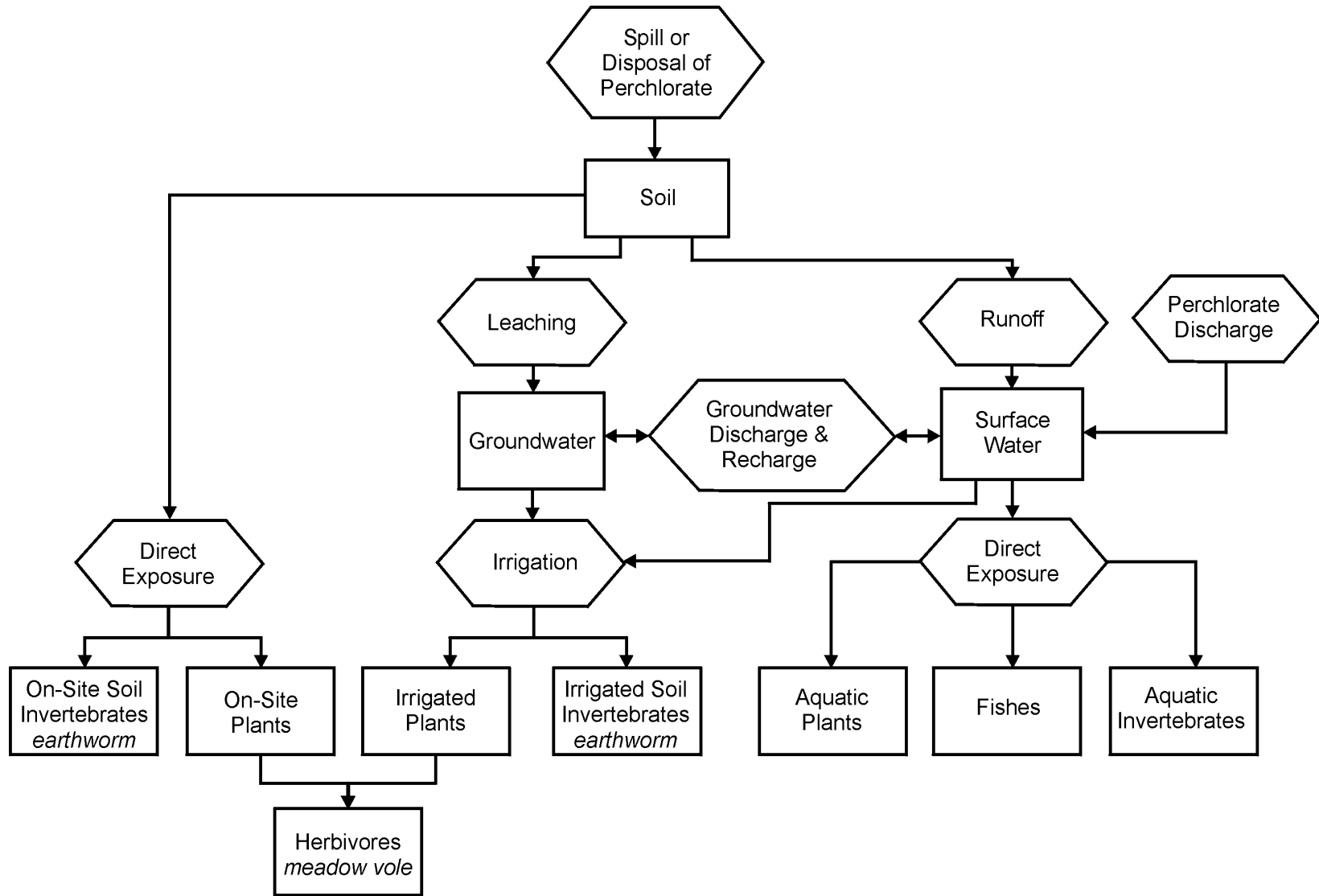
Figure 8-1. A conceptual model of exposure of ecological endpoint receptors to perchlorate. Specific endpoint taxa are identified in italics; all other endpoints are defined at the community level. Processes are designated by hexagonal boxes, compartments by rectangular boxes.

1   operations, explosives, or manufacturing wastes; and the aqueous discharge of waste water from

2   manufacturing of perchlorate. The most recent information on perchlorate content in fertilizers

3   demonstrates that fertilizer use is unlikely to constitute an environmentally significant source of

4   perchlorate contamination, and ecological risks from this source are not considered further (see

5   Chapter 9). Spills contaminate the soil at the site and, through leaching and run-off, contaminate

6   the surface water and groundwater. The discharge of groundwater to surface water may result in

7   locally high levels of perchlorate in surface waters. Aquatic communities are exposed directly to

8   contaminated surface water; soil invertebrate and plant communities are exposed to perchlorate

9   in soil at the spill site and through irrigation with either surface or groundwater; and herbivorous

10   terrestrial wildlife are exposed through their consumption of plants that have bioconcentrated

11   perchlorate. However, the potential for transfer of perchlorate further up the terrestrial food web

12   is currently unknown.

13   This conceptual model is relatively simple because it excludes some potential routes and

14   receptors. Dietary exposures are excluded from aquatic systems because, as of this writing,

15   available data have not shown perchlorate to bioconcentrate to any significant extent.

16   Information newly received form the U.S. Army Corps of Engineers (Condike, 2001) report on

17   the analysis of environmental samples from perchlorate-contaminated water bodies near

18   McGregor Naval Weapons Industrial Reserve Plant (NWIRP), TX, and purports to show fish

19   tissue concentrations that exceed comparable water concentrations. These data suggest that

20   perchlorate not only accumulates but is bioconcentrated. This information, which has not yet

21   been fully reviewed by the U.S. EPA, is herewith made available with this document to external

22   peers for their review.

23   Wildlife are assumed to have negligible exposure from air or from direct exposure to soil.

24   Exposures of wetlands to groundwater or surface water are not included explicitly because their

25   exposures and effects are assumed to be equivalent to irrigation exposures. That is, plants and

26   invertebrates are assumed to be exposed to pore-water concentrations equal to surface or

27   groundwater concentrations. Exposures to contaminated sediments also are not included

28   explicitly because they are believed to be equivalent to surface water exposures. Perchlorate salts

29   are highly water soluble and the anion is unlikely to adsorb to anionic particles, such as soils or

30   humic substances, to a significant extent. Therefore, sediment exposures are expected to be

31   dominated by exposure to pore water, which is assumed to be equal to surface water.

1   **8.2.3 Analysis Plan**

2   This screening assessment uses existing information to determine whether the existing

3   environmental contamination by perchlorate poses a clearly significant risk, insignificant risk, or

4   an ambiguous risk. The analysis of effects will consist of the derivation of screening benchmarks

5   through the application of conservative extrapolation models. The analysis of exposure for

6   ecological endpoints consists of measured concentrations reported in Chapter 1 or derived from

7   Parsons (2001) or Smith et al. (2001). Soil exposure estimates are based on exposure to

8   perchlorate in irrigation water.

9

10

11   **8.3 ANALYSIS**

12   **8.3.1 Characterization of Exposure**

13   **8.3.1.1 Water Concentrations**

14   As previously described, fishes, aquatic invertebrates, and aquatic plants may be exposed

15   directly to concentrations of perchlorate in surface waters. These concentrations may result from

16   surface run-off from perchlorate-contaminated soil, from leaching of perchlorate from

17   contaminated soil via shallow groundwater, or from direct discharge of aqueous wastes. Surface

18   or groundwater may be used for irrigation, resulting in direct exposure of soil invertebrates or

19   plants (Figure 8-1).

20   Perchlorate salts are dissolved readily given the conditions under which the contamination

21   has occurred, releasing the perchlorate anion and the associated cation. Sorption is not expected

22   to attenuate perchlorate because it absorbs weakly to most soil minerals, and natural chemical

23   reduction in the environment is not expected to be significant. Consequently, perchlorate is both

24   very mobile in aqueous systems and persistent for many decades under typical ground and

25   surface water conditions (Section 1.1).

26   Limited information is available on perchlorate concentrations in surface waters.

27   Perchlorate from an ammonium perchlorate manufacturing area has been detected at 4 to 16 μg/L

28   downstream in Lake Mead and the Colorado River (Section 1.2). Information on the frequency

29   or central tendency (mean or median) of perchlorate detection in those water bodies was not

30   available for this review, but it is assumed that some aquatic organisms are exposed chronically

1   to concentrations as high as 16 μg/L. On the other hand, perchlorate concentrations have been

2   measured as high as 0.37% (37×10^6 μg/L) in groundwater-monitoring wells at facilities that

3   manufacture or test rocket motors and at 280 μg/L in public water supply wells (Section 1.2)

4   Smaller surface water bodies, including some that are supplied primarily by groundwater, are

5   likely to exist near sites of soil contamination and to have perchlorate concentrations much

6   higher than those reported for Lake Mead and the Colorado River. A spring associated with the

7   Las Vegas Wash site had concentrations of 1.0 to 1.3×10^5 μg/L in surface water (Parsons,

8   2001). Perchlorate concentrations in a pond (INF Pond) that receives water from the pump and

9   treat system at the Longhorn Army Ammunition Plant near Karnack, TX ranged from 30,776 to

10   31,438 μg/L in November 1999 (Smith et al., 2001) and ranged from 3500 to 3800 μg/L in

11   September 2000 (Parsons, 2001). It is also possible that, within large water bodies, there are

12   locally elevated concentrations at sites of groundwater discharge. In the vicinity of a sediment

13   delta created by the Las Vegas Wash in Las Vegas Bay of Lake Mead, Parsons (2001) documents

14   a maximum perchlorate concentration of 68 μg/L in surface water. At the Allegany Ballistics

15   Laboratory in Rocket Center, WV, discharge water from a Comprehensive Environmental

16   Response Compensation, and Liability Act (CERCLA) groundwater pump and treat facility to

17   the North Branch Potomac River contained 250 to 280 μg/L perchlorate (Parsons 2001). Surface

18   water concentrations in Town Gut Marsh adjacent to the Naval Surface Warfare Center at Indian

19   Head, MD ranged from not detected (reporting limit = 4.0 μg/L) to 25 μg/L. It should be noted

20   that the groundwater pump and treat facilities either at Longhorn Army Ammunition Plant or

21   Allegany Ballistics Laboratory were not equipped with facilities to treat perchlorate in water.

22   Surface water concentrations in Harrison Bayou below the discharge point for the INF pond

23   at LHAAP also ranged from undetectable (reporting limit = 4.0 μg/L) to 4.0 μg/L (Parsons,

24   2001; Smith et al., 2001). However, Smith et al. (2001) point out that water from the pond is

25   discharged to Harrison Bayou only during periods when Harrison Bayou is flowing, and neither

26   study apparently sampled Harrison Bayou when water was being discharged from the pond.

27   Therefore, higher concentrations of perchlorate in surface water of Harrison Bayou are likely to

28   be measured at other times.

29   It is assumed that irrigation waters pumped from Lake Mead or the Colorado River are in

30   the range of downstream concentrations given above (i.e., 4 - 16 μg/L). Groundwater irrigation

1   may be contaminated at levels similar to those observed in public water supplies ($\leq$280 μg/L),

2   unless the well is appreciably nearer a perchlorate-contaminated site.

3

4   **8.3.1.2 Aquatic Bioaccumulation**

5   As discussed above, little information has been previously available on the potential for the

6   perchlorate ion to accumulate in animal tissues. The studies outlined in the Parsons (2001)

7   report sought to answer the question whether perchlorate is present in ecological receptors.

8   In these studies, concentrations of perchlorate in aquatic vegetation, fish, amphibians, aquatic

9   invertebrates, and birds were compared to surface-water, pore-water, and sediment

10   concentrations from the same water body. This information is supplemented by the additional

11   studies conducted at LHAAP by Smith et al. (2001).

12   When perchlorate concentrations in physical media (i.e., surface water or sediment) were

13   greater than the reporting limits for biological media ($\geq$300 ppb [μg/L or μg/kg] in Parsons

14   [2001]), concentrations in aquatic vegetation were similar to or greater than the concentrations in

15   surface water or pore water; but concentrations in fish, amphibians, or invertebrates were less.

16   In Smith et al. (2001) reported the detection of high concentrations of perchlorate in the INF

17   Pond and lower concentrations in aquatic vegetation and in animals than in surface water or

18   sediments.

19   In Parsons (2001), when perchlorate concentrations in the physical media were lower,

20   concentrations in aquatic vegetation or amphibians were in a few cases greater than the

21   concentrations in surface water or sediment; but in most cases, perchlorate was not detected in

22   aquatic receptors. However, our understanding of bioaccumulation of perchlorate in this lower

23   concentration range is limited because the reporting limits in the Parsons (2001) studies for

24   perchlorate in animal tissues (i.e., 300-400 μg/kg) were greater then the reporting limits for

25   surface water or pore water (i.e., $\approx$ 4 μg/L) or for sediments (i.e., $\approx$ 80 μg/L).

26   Although Smith et al. (2001) do not identify their reporting limits, their reporting limits for

27   biological tissues appear to be less (i.e., $\approx$70 μg/kg based on their lowest detected concentration)

28   than those of Parsons (2001). In the Smith et al. (2001) study of LHAAP, detected perchlorate

29   concentrations were similar in surface water (44-85 μg/L), sediments (78 μg/kg), and fish tissues

30   (83-131 μg/kg) at Goose Prairie Creek. In Harrison Bayou, the single detected concentration in

31   surface water (4 μg/L) was less than detected concentrations in animal tissues (86-356 μg/kg).

1   However, as the authors discuss, the measured concentration in surface water in Harrison Bayou

2   is likely less than when water is being discharged from the INF Pond (Smith et al., 2001). In

3   addition, the study did not collect sufficient samples from any one site and medium or species for

4   any significant statistical comparisons to be made.

5   Information newly received form the U.S. Army Corps of Engineers (Condike, 2001) report

6   on the analysis of environmental samples from perchlorate-contaminated water bodies near

7   McGregor Naval Weapons Industrial Reserve plant (NWIRP), TX, and purports to show fish

8   tissue concentrations that exceed comparable water concentrations. These data suggest that

9   perchlorate not only accumulates but is bioconcentrated. This information, which has not yet

10   been fully reviewed by the U.S. EPA, is herewith made available with this document to external

11   peers for their review.

12   The above information indicates that perchlorate may bioaccumulate in aquatic organisms

13   living in contaminated waters, but it does not resolve the question of whether perchlorate may

14   bioconcentrate in the tissues of aquatic organisms to levels exceeding the surface water

15   concentrations. The existing data are also insufficient to determine whether there is further

16   trophic transfer of perchlorate within aquatic food webs.

17

18   **8.3.1.3 Soil Levels**

19   On-site soils may be contaminated by direct spills of perchlorate solutions from flushing

20   rockets, combustion of rocket fuel, improper disposal of rocket fuel, open burn/open detonation

21   operations, explosives, or manufacturing wastes. Perchlorate concentration measurements at

22   disposal sites range from less than 1 to 1470 mg/kg (Parsons, 2001). Off-site soils may be

23   contaminated via irrigation (Figure 8-1). Because of the high water solubility of perchlorate

24   salts, perchlorate is unlikely to accumulate via adsorption to irrigated soils, and aqueous

25   perchlorate was not found to adsorb to sand in laboratory reactors (Nzengung, n.d.). By gross

26   approximation, then, soil concentrations (expressed as milligrams per kilogram) would be

27   unlikely to exceed the concentrations (expressed as milligrams per liter) in irrigation water.

28   Similarly, concentrations of perchlorate in soil pore water may be assumed to be equal to the

29   concentration in irrigation water, both in the field and in soil toxicity tests. However, the

30   concentration of perchlorate salts in irrigated soils with high evaporation rates cannot be ruled

31   out. At the Yuma site, soils are irrigated with water from the Colorado River, and concentrations

1   of perchlorate in surface-water samples collected near the irrigation intake locations ranged from

2   0.003 to 0.006 mg/L. In surface soil, the single detection (0.090 mg/kg) was well above the

3   concurrently-measured water concentrations, as were the perchlorate detection limits in soil

4   (0.079 to 0.080 mg/kg). The relatively higher detection limits in soil, the limited nature of the

5   sampling in soil and water, and the lack of information about potential sources other than

6   irrigation water (see Section 8.1.2) complicate the interpretation of the presence and fate of

7   perchlorate in irrigated soils.

8

9   **8.3.1.4 Uptake by Vegetation**

10   Several laboratory experiments have examined plant uptake of perchlorate from solution

11 culture. Experiments with candidate plants for use in the phytoremediation of perchlorate-

12 contaminated sites showed that perchlorate may concentrate in vegetation (Nzengung, n.d.;

13 Susarla et al., 2000a). Nzengung (n.d.) used rooted cuttings of woody plants, willow (*Salix* spp.),

14 Eastern Cottonwood (further identified only as "poplar"), and eucalyptus (*Eucalyptus cineria*)

15 planted in sand with nutrient solution containing perchlorate at 20 or 100 mg/L for 24 to 42 days.

16 In each case, perchlorate was taken up and concentrated in aerial plant parts, especially leaves.

17 Concentration factors, expressed as the ratio of leaf concentration (mg/kg wet weight) to initial

18 solution concentration (mg/L), ranged from 7.5 to 25.

19   Susarla et al. (2000a) used seedlings or rooted cuttings of 13 vascular plant species, planted

20 in sand with nutrients, and exposed for ten days to 0.2, 2.0 or 20 mg/L perchlorate. These

21 researchers also reported depletion of perchlorate from test media. Qualitative analyses

22 suggested accumulation of perchlorate in the aerial tissues of most of the species analyzed.

23 Using their data and the approach reported by Nzengung above, we calculated concentration

24 factors ranging from 0 to 330.

25   Nzengung (n.d.) and Susarla et al. (2000a) reported that perchlorate accumulated primarily

26 in the leaves, followed by stems, then roots. Predicted perchlorate breakdown products, chlorate,

27 chlorite, and chloride were detected in plant tissues in both studies, but quantitative evidence was

28 not presented. In addition to this lack of quantitative data, there are other concerns related to the

29 potential for plants to degrade perchlorate. First, information concerning accumulation and

30 potential transformation is limited to a few studies by these two laboratories. Second, the

31 method used for perchlorate analysis yielded estimates of perchlorate in fertilizer that were

1 subsequently found to be overestimated by 30 to 150% (Susarla et al., 2000b). Third, no

2 physiological explanation has been suggested for why plants should accumulate this salt far in

3 excess of concentrations in water or groundwater, though it appears this may be simply a

4 function of water uptake rates to meet transpirational losses. Fourth, these two studies were

5 short-term, material depletion studies, a type of study we believe will overestimate long-term

6 accumulation rates because some of the "response" is likely the result of factors not related to the

7 chemical in question. There is ample evidence from salt accumulation studies of plants to

8 suggest that the initial increases in perchlorate accumulation by plants may be due to a salt effect;

9 that is, nutrient salts are initially indistinguishable from perchlorate salts in that they all represent

10 an ionic imbalance across the cell wall. One of the confounding issues that can only be

11 determined with longer-term studies is the effect of increased cell sap salinity on additional

12 perchlorate uptake. As sap salinity increases, there should be an increase in H_2O uptake, further

13 increasing the perchlorate concentrations. This will continue only until a certain concentration of

14 salts, including perchlorate, is reached, at which time the plant will close its stomata and shunt

15 sap salts to vacuoles.

16 　　　In addition to the above stated concerns, there is no reason to expect that these are steady-

17 state concentration factors. These experiments were designed to quantify phytotransformation of

18 an initially introduced perchlorate quantity, rather than bioconcentration, with respect to an

19 ambient perchlorate concentration. As the perchlorate-amended nutrient solution was transpired,

20 and some perchlorate was taken up, it was replenished by solution, without added perchlorate;

21 thus, perchlorate in the test chamber diminished throughout the experiment. Concentration

22 factors that would be observed at steady state, such as may result from continual irrigation with

23 perchlorate-contaminated water, cannot be estimated from this study. Because of the

24 uncertainties associated with both perchlorate accumulation and degradation by plants, a simple,

25 conservative, screening-level assumption that concentrations in leaves can exceed water

26 concentrations by a factor of 100 was made.

27 　　　If irrigation is from surface water sources similar to the Colorado River or Lake Mead, with

28 concentrations as high as 16 μg/L, then plant concentrations are assumed to be as high as

29 1.6 mg/kg. If irrigation is from groundwater sources similar to potable water supplies, with

30 concentrations as high as 280 μg/L, then plant concentrations will be assumed to be as high as

31 28 mg/kg.

1   Concentrations in plant tissues and soils also have been measured in the field. Ellington

2   et al. (2001) measured perchlorate concentrations in leaves of tobacco, *Nicotiana tabacum* var.

3   K326, field-grown in soil amended with Chilean saltpeter, which is naturally high in perchlorate.

4   Perchlorate concentrations (± SD) in leaf lamina from the 1999 crop were 96.0 ± 0.6 mg/kg dry

5   weight and 14.6 ± 0.1 mg/kg wet weight; concentration in a composite soil sample collected in

6   December 1999 was 0.34 ± <0.01 mg/kg dry weight. The concentration factor in this study was

7   approximately 43, on the basis of wet weight in leaf lamina and dry weight in soil.

8   The field studies by Parsons (2001) found that, for various sites, wet-weight perchlorate

9   concentrations in terrestrial vegetation samples were 1.5 to 80 times the wet-weight

10   concentrations in soil samples. The data from one site (i.e., Building 25C) at LHAAP (Smith

11   et al., 2001) seem to indicate greater concentration factors, but the soil and plant samples were

12   taken at different times of the year (i.e., January and October, respectively) and only one sample

13   each of three plant species was analyzed.

14   Soil-to-vegetation concentration factors derived from the above field studies were similar

15   in magnitude, but when using them for risk assessment care should be taken to note the different

16   bases; exposure concentration was variously reported as mg/kg wet weight in soil or mg/kg dry

17   weight in soil. The maximum measured concentration in vegetation at irrigated sites in the

18   vicinity of Yuma, Arizona was 1.0 mg/kg wet weight. At sites with soil contamination related to

19   the manufacture, handling, or use of perchlorate in solid propellants, maximum plant

20   concentrations were 428 mg/kg wet weight at a spring; 99.2 mg/kg wet weight at a site upstream

21   from Lake Las Vegas in the Las Vegas Wash area of the Lake Mead Recreational Area, Nevada;

22   and 300 mg/kg wet weight at the Burn Area of Allegany Ballistics Laboratory, West Virginia.

23   In most cases, detection limits were ~0.4 mg/kg wet weight.

24

25   **8.3.1.5 Herbivore Exposure**

26   The representative herbivore selected for this assessment, *M. pennsylvanicus*, has a diet

27   consisting mainly of monocot and dicot shoots, has an estimated food consumption rate of

28   0.005 kg/day wet weight, and a body weight of 0.044 kg (Sample and Suter, 1994). Using the

29   assumptions stated above, daily exposures resulting from surface water and groundwater

30   irrigation may be as high as 0.18 mg/kg-day and 3.2 mg/kg-day, respectively. Daily exposures

31   resulting from maximum measured concentrations in plants range from 0.11 mg/kg-day at the

1   irrigated sites in the vicinity of Yuma to 49 mg/kg-day for the sites with direct soil

2   contamination.

3   In the Parsons (2001) studies, except when concentrations in surface soils were high (i.e.,

4   ≥9000 μg/kg), perchlorate was not detected in terrestrial birds, mammals, or insects with

5   reporting limits of 300 to 400 μg/kg. The vertebrates collected varied substantially between

6   sites, but the birds collected include the mourning dove (*Zenaida macroura*), tree swallow

7   (*Tachycineta bicolor*), roughwing swallow (*Stelgidopteryx serripennis*), lesser nighthawk

8   (*Chordeiles acutipennis*), nighthawk (*C. minor*); Gambel's quail (Callipepla gambelii), starling

9   (*Sturnus vulgaris*); American crow (*Corvus brachyrhynchus*), eastern bluebird (*Sialia sialis*),

10   eastern phoebe (*Sayornis phoebe*), and blue grosbeak (*Guiraca caerulea*). The mammals

11   collected include the cactus mouse (*Peromyscus eremicus*), rock pocket mouse (*Chaetodipus*

12   *intermeius*), Audubon's cottontail (*Sylvilagus audubonii*), deer mouse (*P. maniculatus*), long-

13   tailed pocket mouse (*Perognathus formosus*), western pipestrelle (*Pipistrellus hesperus*), house

14   mouse (*Mus musculus*), white-footed mouse (*Peromyscus leucopus*) meadow vole (*Microtus*

15   *pennsylvanicus*), Merriam's kangaroo rat (*Dipodomys merriami*), desert pocket mouse

16   (*C. penicillatus*), hispid cotton rat (*Sigmodon hispidus*), western harvest mouse (*Reithrodontomys*

17   *megalotis*), marsh rice rat (*Oryzomys palustris*); northern short-tailed shrew (*Blarina*

18   *brevicauda*), racoon (*Procyon lotor*), eastern harvest mouse (*R. fulvescens*), little brown bat

19   (*Myotis lucifugus*), eastern cottontail (*S. floridanus*). At those sites where perchlorate

20   concentrations in surface soils were high, perchlorate concentrations in potential herbivore

21   tissues were generally an order of magnitude or more less than that in vegetation. At one site,

22   Yuma, with lower perchlorate concentrations in soil (i.e., mean of all results = 81 μg/kg),

23   perchlorate was detected in a single terrestrial reptile sample (brush lizard, *Urosaurus graciosus*),

24   but this detection was lower than the mean perchlorate concentration in vegetation. Although

25   detected soil concentrations were lower (i.e., 50 to 322 μg/kg) in Smith et al. (2001), the

26   concentrations of perchlorate in two composite samples of livers from harvest mice

27   (*Reithrodontomys fulvescens*) were several orders of magnitude less than the detected

28   concentrations in their potential food, plant leaves or seeds.

29

30

8.3.2 Characterization of Effects

8.3.2.1 Aquatic Organisms

Effects on the richness and productivity of fish, aquatic invertebrate, and aquatic plant communities are jointly characterized using the procedures for deriving Tier II water quality values (U.S. Environmental Protection Agency, 1993; Suter and Tsao, 1996). Tier II values are derived where data are not sufficient for deriving ambient water quality criteria (AWQC). The Tier II value derivation procedures account for missing information with approximately 80% confidence.

Test results potentially useful for deriving Tier II values were available for five aquatic species (Table 8-1). In acute tests (48 and 96 h) with sodium perchlorate, using the water flea *Daphnia magna*, the amphipod *Hyalella azteca*, and the fathead minnow *Pimephales promelas*, the endpoints lethality and inhibition were studied. In seven-day tests with a different water flea (*Ceriodaphnia dubia*) and with *P. promelas*, acute lethality was studied in addition to more sensitive endpoints, including the number of offspring per female (*C. dubia*) and growth rate (i.e., body weight; *P. promelas*). A seven-day test with *C. dubia* generally is here used in place of a chronic (i.e., twenty-one day) test because test organisms produce three broods during the test; a seven-day test with *P. promelas* is arguably subchronic because of the test's short duration relative to the organism's lifespan (Suter, 1990; Norberg-King, 1990). A 35-day, early-life-stage (ELS) test with *Pimephales*, here used in place of a chronic test, showed no significant effects on any standard endpoint (survival, growth or biomass) at the highest concentration tested. However, all larvae exposed to perchlorate concentrations, including the lowest concentration of 28 mg/L, exhibited redness and swelling that was not observed in the larvae exposed to the control water. This finding suggests the presence of subtle effects that could be ecologically significant and raises doubt about whether a chronic No-Observed-Effect-Concentration (NOEC) has been adequately determined for this species.

Steps followed in the derivation of the Tier II value for sodium perchlorate are presented in Table 8-2. The secondary acute value (SAV), 5 mg/L (as ClO_4^-), is derived to be protective of 95% of species during short-term exposures with 80% confidence. The secondary chronic value (SCV), 0.6 mg/L (as ClO_4^-), likewise is derived to be protective of 95% of species during long-term exposures with 80% confidence. A sodium chloride control test showed that some of the toxicity to *P. promelas* was potentially attributable to the sodium cation. These tests suggest

TABLE 8-1. RESULTS OF PERCHLORATE TOXICITY TESTS IN AQUATIC AND TERRESTRIAL SPECIES

| Test Species | Test Description | | Endpoints (as mg/L perchlorate in water or mg/kg in soil or sand)[a] | | | | |
	Age	Duration	Acute LC$_{50}$ (95% CL)	NOEC	LOEC	ChV	IC$_{25}$ (95% CL)
Sodium perchlorate (NaClO$_4$)[b] tests (EA Engineering, Science and Technology, Inc., 1998)							
Daphnia magna	<24 hr	Acute (48-hr)	490 (406 - 591)	—	—	—	—
Pimephales promelas	12 - 13 days	Acute (96-hr)	1,655 (1,507 - 1,817)	—	—	—	—
Ceriodaphnia dubia	<24 hr	Chronic (7-day)	66 (40-144) [48-h]	10	33	18.2	17 (8.1 - 20.5)
Pimephales promelas	<24 hr	Subchronic (7-day)	614 (540 - 714) [96-h]	155	280[c]	208[c]	212[c] (175 - 231)[c]
Lactuca sativa	<24 hr	Subchronic (7-day)	614 (540 - 714) [96-h]	155	280[c]	208[c]	212[c] (175 - 231)[c]
Lactuca sativa		Chronic definitive (28-d, sand)		<80	80	<80	41
Lactuca sativa		Chronic definitive (28-d, soil)		40	40	56.6	30
Lactuca sativa		Chronic definitive (28-d, sand)		20	40	28.3	34.3
Eisenia foetida		Acute definitive (7 day/14 day, soil)	4,450/4,450	—	—	—	—
Sodium perchlorate (NaClO$_4$)[b] tests (EA Engineering, Science and Technology, Inc., 2000)							
Pimephales promelas	Embryo	Chronic (35-day, early life stage)	> 490 [96-hr]	> 490[d] <28[e]	> 490[d] 28[e]	> 490[d]	> 490[d] <28[e]
Hyalella azteca	7 - 14 days	Chronic definitive (28-day)		> 1000	> 1000	> 1000	> 1000

TABLE 8-1 (cont'd). RESULTS OF PERCHLORATE TOXICITY TESTS IN AQUATIC AND TERRESTRIAL SPECIES

Test Species	Test Description		Endpoints (as mg/L perchlorate in water or mg/kg in soil or sand)[a]				
	Age	Duration	Acute LC$_{50}$ (95% CL)	NOEC	LOEC	ChV	IC$_{25}$ (95% CL)
Sodium perchlorate (NaClO$_4$)[b] tests (EA Engineering, Science, and Technology, 1999)							
Selenastrum capricornutum	7 days	Acute (96-hr)	—	500	1,200	775	615 (149-1,126)
Ammonium perchlorate (NH$_4$ClO$_4$)[f] tests (Block Environmental Services, Inc., 1998)							
Ceriodaphnia dubia	<24 hr[g]	Chronic (6-day)	77.8 [6-days]	9.6	24	15	24
Pimephales promelas	<24 hr[g]	Subchronic (7-day)	270 [7-days]	9.6	96	30	114
Ammonium perchlorate (NH$_4$ClO$_4$)[f] tests (Dumont and Bantle, 1998)							
Xenopus	Embryo	96-hr	420[h]	—	—	—	—
Xenopus	Embryo	96-hr	336[h]	—	—	—	—

[a]Notation: LC$_{50}$ = Concentration lethal to 50% of individuals; NOEC = No-observed-effect concentration; LOEC = Lowest-observed-effect concentration; ChV = Chronic value; IC$_{25}$ = Concentration inhibiting a process (e.g., growth, reproduction) by 25%; CL = confidence limits.

[b]Sodium chloride control showed no adverse effects of sodium ion except as indicated. Reported values are based on nominal concentrations.

[c]Sodium chloride control showed significant adverse effects attributable to sodium cation at highest test concentration. Effects observed at this perchlorate concentration may have been caused in part by sodium ion toxicity.

[d]Standard endpoints: survival, growth, biomass

[e]Although there were no effects on standard endpoints at any tested concentration, the investigators reported that all larvae exposed to perchlorate concentrations, including the lowest concentration of 28 mg/L, exhibited redness and swelling, which was not observed in the larvae exposed to the control water.

[f]Ammonium control was not used; adverse effects of ammonium ion cannot be ruled out at all effect concentrations. *C. dubia* and *P. promelas* results are based on measured concentrations. *Xenopus* results are based on nominal concentrations. Confidence limits are not reported.

[g]Not reported; assumed based on standard protocols.

[h]IC$_{50}$ for malformations.

TABLE 8-2. PROCEDURE FOR DERIVING TIER II WATER QUALITY VALUES FOR SODIUM PERCHLORATE

Step	Value (mg/L ClO_4^-)	Rationale
Identify the lowest genus mean acute value (GMAV)	66	Lowest GMAV is for genus *Ceriodaphnia* (based on *C. dubia*)
Determine the final acute value factor (FAVF), a factor that compensates for lack of data on a sufficient number of taxonomic groups	13.2 (unitless)	The FAVF varies according to the number of specified taxonomic groups for which GMAVs were available. In this case, two specified values were available (a nonsalmonid fish and a planktonic crustacean), of which one is a daphnid; the value selected from the FAVF table (U.S. Environmental Protection Agency, 1993; Suter and Tsao, 1996) is 13.2.
Calculate the secondary acute value (SAV)	5.0	SAV=GMAV ÷ FAVF = 66 ÷ 13.2
Identify three or more acute-chronic ratios (ACRs), which are ratios of acute value (AV) to chronic value (CV) for a given species (but ratios must be geometrically averaged within any single genus)	3.6, 8.0 (range, <3.4 - >59), 17.9	ACRs can be derived for two species in different genera. For *C. dubia*: ACR=AV ÷ CV = 66 ÷ 18.2 = 3.6 For *P. promelas*, two AVs are available. The lower (614) is thrown out because the larval stage is not standard for acute tests; the higher (1,655) is used. Three CVs are available: >490 for standard endpoints, and <28 for redness and swelling, in the 35-d ELS test; and 208 for survival in the 7-d test. There is uncertainty as to the interpretation of the ELS test results; the 7-d result is used and the two results from the ELS are used to determine a range, shown in parentheses: ACR = 1,655 ÷ 208 (range, >490 - <28) = 8.0 (range, <3.4 - >59) No ratio is possible for *H. azteca* because we are unable to calculate CV due to no acute toxicity. Because a third value is not available, a default value of 17.9 (which provides 80% confidence based on other toxicants) is substituted, according to the Tier II method.
Derive the secondary acute-chronic ratio (SACR)	8.0 (range, <6.0 - >15.6)	The SACR is the geometric mean of the ACRs. (The uncertainty range associated with the *P. promelas* value is carried through and shown in parentheses.)
Derive the secondary chronic value (SCV)	0.60 (range, <0.32 - >0.83)	SCV=SAV ÷ SACR, 5.0 ÷ 8.0 (The uncertainty range associated with the *P. promelas* value is carried through and shown in parentheses.)

1   the possibility that if perchlorate were associated with a less toxic cation, the SCV may be lower

2   than is necessary to protect against perchlorate ion toxicity. Further tests with perchlorate may be

3   needed to assess potentially less toxic cations.

4   Similar chronic (or subchronic) tests were conducted with ammonium perchlorate

5   (Table 8-1). Results, expressed as ClO_4^-, were very similar for *C. dubia*, but *P. promelas* was

6   more sensitive to ammonium perchlorate than to sodium perchlorate. However, Tier II values for

7   ammonium perchlorate are not presented for several reasons, including the lack of ammonium

8   controls which makes it difficult to determine whether the observed effects were caused by the

9   perchlorate anion; the lack of acute values for *C. dubia* and *P. Pimephales*; and the fact that the

10   FETAX (*Xenopus*) test is designed to detect teratogenic potential, and the embryo is not a

11   particularly sensitive life stage for toxicity. When perchlorate is administered as the ammonium

12   salt, the ammonium ion concentration expressed on an ammonia-nitrogen (in milligrams of

13   NH_3-N/L) basis is 14% of the respective perchlorate ion concentration. A Lowest-Observed-

14   Effect-Concentration (LOEC) for *C. dubia* of 24 mg/L perchlorate (Table 8-1) thus corresponds

15   to 3.4 mg NH_3-N/L. Based on a species mean chronic value (SMCV) of 13 mg NH_3-N/L for

16   *C. dubia* exposed to ammonia alone (U.S. Environmental Protection Agency, 1998h), the former

17   value is probably too low to be responsible for the observed effects[1]. On the other hand, the

18   LOEC observed with *P. promelas* at 96 mg/L (Table 8-1) corresponds to 14 mg NH_3-N/L, which

19   exceeds a SMCV of 3.09 mg NH_3-N/L (U.S. Environmental Protection Agency, 1998h).

20   Therefore, ammonium exposure alone could have been responsible for the effects of ammonium

21   perchlorate observed in *P. promelas*.

22   The SAV and SCV derived above based on sodium perchlorate are probably protective

23   even if ammonium perchlorate is the form released, however. Calculated NH_3-N concentrations

24   corresponding to those values are below the acute and chronic ambient water quality criteria for

25   ammonia, regardless of pH (U.S. Environmental Protection Agency, 1998h). While SAV and

26   SCV are not calculated for plants, it appears that there is little perchlorate or ammonium toxicity

27   to the alga *Selenastrum* in toxicity studies (Table 8-1).

28

---

[1]Ammonia/ammonium toxicity increases as test-water pH increases (U.S. Environmental Protection Agency, 1998e). The value of 13 mg NH_3-N/L corresponds to a pH of 8.0; however, unless the test water pH had exceeded 8.8, it is doubtful that 3.4 mg NH_3-N/L was responsible for the observed effects.

1 **8.3.2.2   Terrestrial Organisms**

2   **Plants**. The only available phytotoxicity information comes from 28-day seedling growth

3 tests of lettuce (*Lactuca sativa*) performed in soil and sand cultures with sodium perchlorate (EA

4 Engineering, Science and Technology, Inc., 1998). Although the exposure was to sodium

5 perchlorate solution added to the solid media, the results may be expressed as milligrams per

6 kilogram soil or sand, wet weight, or as milligrams per liter of irrigation solution. Growth was a

7 more sensitive response than germination or survival. The quartile inhibitory wet-weight

8 concentrations (IC_{25}s) for growth in soil and sand were 78 mg/kg (293 mg/L) and 41mg/kg

9 (160 mg/L), respectively. Survival was reduced 26% at 420 mg/kg (2,520 mg/L) in soil and 39%

10 at 180 mg/kg (840 mg/L) in sand. To account for interspecies variance, a factor of 10 is applied

11 to the lowest IC_{25} to obtain a screening benchmark of 4 mg/kg as a wet-weight concentration in

12 soil (or 16 mg/L as a concentration in irrigation solution).

13   **Soil Invertebrates**. The only available toxicity data for soil invertebrates is a 14-day acute

14 lethality test of the earthworm (*Eisenia foetida*) performed in artificial soil irrigated with sodium

15 perchlorate. The LC_{50} at both 7 and 14 days was 4,450 mg/kg as a wet-weight concentration in

16 soil. No factors or other models are available to extrapolate from that LC_{50} to chronic effects on

17 survival, growth, or fecundity or to extrapolate from this species to the soil invertebrate

18 community as a whole. Therefore, the factors applied for aquatic communities in cases where

19 there is only one LC_{50} (see Section 8.3.2.1) to obtain a conservative estimate of a soil screening

20 benchmark for soil community effects, are as follows:

21   Threshold   = LC_{50} ÷ (factor for interspecies variance × acute-chronic ratio)

22   =   4,450 mg/kg ÷ (242 × 18)

23   =   1 mg/kg as a wet-weight concentration in soil.

24 The equivalent aqueous phase benchmark is 2.8 mg/L. This approach requires the assumptions

25 that the variance among soil species is approximately the same as among aquatic species, and

26 that the distribution of acute-chronic ratios across chemicals is approximately the same for both

27 communities. The interspecies variance factor is the one for a test species that has not been

28 demonstrated to be highly sensitive.

29   **Herbivores.** The human health risk assessment for perchlorate uses 0.01 mg/kg-day as the

30 LOAEL from which the RfD is derived (Chapter 7). That value is based on perturbations in

31 thyroid and pituitary hormones, thyroid histopathology, and changes in brain morphometry in P0

1   dams on GD21 and F1-generation rat pups on PND5, PND10, and PND22. Because the

2   representative species for the herbivore endpoint (meadow vole) is a rodent, that value is used

3   here as well. The population-level implications of this effect are unknown; however, it seems

4   likely that such effects on the thyroid, pituitary, and brain could diminish survivorship and

5   fecundity and diminish population production. To account for interspecies variance and LOAEL

6   to NOAEL extrapolation, an uncertainty factor of 10 is applied to obtain a dietary screening

7   benchmark for herbivores of 0.001 mg/kg body weight-day, or ~0.01 mg/kg as a wet-weight

8   concentration in plant tissue (see exposure assumptions in Section 8.3.1.5).

9

CHAPTER 9. EVALUATION OF EVIDENCE FOR INDIRECT EXPOSURES

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The primary purpose of this document is to derive human and ecological risk estimates for perchlorate. As indicated in Chapter 1, pollution of drinking water supplies is the major concern. Most perchlorate salts are used as solid oxidants or energy boosters in rockets or ordnance; therefore, much of the perchlorate-tainted waterways in the U.S. can be traced to military operations, defense contracting, or associated manufacturing facilities. Figure 1-5 shows that the perchlorate anion could potentially be found in many natural waterways that are used for irrigation or consumed by livestock or wildlife. Thus, it is logical to question whether there are means through which humans might consume perchlorate other than drinking water. This question is compounded by the chemical nature of perchlorate, which grants it long life under typical environmental conditions (Urbansky, 1998; Urbansky and Schock, 1999; Espenson, 2000).

As discussed in Section 7.1.5, once a reference dose for perchlorate is established, any burden posed by exposure routes other than potable water necessarily requires that the contaminant's concentration in a water supply be lowered by an equivalent amount if it is determined to calculate a maximum contaminant level goal (MCLG). A relative source contribution (RSC) of between 20% to 80% is used to adjust the RfD according to the decision framework presented in the EPA's methodology for deriving ambient water quality criteria (U.S. Environmental Protection Agency, 2000).

Because polluted waters are used for irrigation, there are also questions concerning absorption, elimination, and retention in food plants. However, this issue becomes considerably less important if it can be demonstrated that the irrigation water is perchlorate-free. Likewise, there are concerns that animals raised for food would consume plants that had received perchlorate-tainted water. As described in Chapter 8, studies are being conducted to assess the occurrence of perchlorate in biological fluids and tissues of animals and plants in affected regions in recognition of the inter-connectedness of the food chain/food web continuum.

1   While much of the perchlorate problem can be traced to specific sites, a few reports have

2   suggested that fertilizers could represent another source of perchlorate in the environment (TRC

3   Environmental Corporation, 1998). These will be addressed in further detail in Section 9.1.1.

4   Sporadic detection of perchlorate in fertilizers was initially alarming because of the widespread

5   use of fertilizers in production farming. In addition to the ecological impact, this raised the issue

6   of assigning responsibility for clean-up costs. Because of the dependence of U.S. agriculture on

7   chemical commodity fertilizers, it was clear that assessment of any possible role of fertilizers

8   would require investigation.

9   This chapter summarizes the available data on the potential for exposure through runoff,

10   erosion, fertilizer, and groundwater movement. Evidence concerning the potential of perchlorate

11   to contaminate soil, sediment, vegetation, livestock and wildlife is also evaluated.

12

13

9.1 FERTILIZERS AS SOURCES OF PERCHLORATE SALTS

9.1.1 The Potential Role of Fertilizers

16   Recently, attention has been drawn to the possible roles of fertilizers as a source of

17   perchlorate contamination for two reasons. First, perchlorate-tainted agricultural runoff could

18   lead to pollution of natural waterways used as drinking water sources. Second, there is a

19   potential for food plants to take up and retain any soluble compounds—including perchlorate

20   salts—and thus provide an alternate route of exposure. It has long been known that Chile

21   possesses caliche ores rich in sodium nitrate ($NaNO_3$) that coincidentally are also a natural source

22   of perchlorate (Schilt, 1979; Ericksen, 1983). The origin of the sodium perchlorate ($NaClO_4$) in

23   the caliche deposits remains an area of debate, but perchlorate is present and can be incorporated

24   into any products made from the caliche.

25   An examination of two manufacturing lots found perchlorate concentrations below 2 mg/g,

26   (i.e., < 0.2% w/w) with some lot-to-lot variability (Urbansky et al., 2001). Presently, the caliche-

27   derived products are sold in the U.S. only by Sociedad Quimíca y Minera (SQM), but other

28   companies have mineral rights to some Chilean deposits and mines (U.S. Environmental

29   Protection Agency 2001b) and are potential sources of caliche-derived products. SQM has now

30   modified its refining process to produce a fertilizer that contains less than 0.1 mg/g (<100 µg/g)

1 of perchlorate, further reducing any environmental release (Lauterbach, 2001). Because nitrate

2 salts (saltpeters) find use as fertilizers, these natural resources have been mined and refined to

3 produce commercial fertilizers for domestic use or for export. Chilean nitrates make up about

4 0.1% of the U.S. market. Most U.S. fertilizers are derived from other raw materials other than

5 sodium nitrate and ammonium nitrate (NH_4NO_3), which is often used for purposes similar to

6 $NaNO_3$, is manufactured from methane, nitrogen, and oxygen. There is no evidence that any

7 ammonium nitrate is derived from Chilean caliche. On account of its low usage, perchlorate

8 from Chilean nitrates cannot represent a continuing, significant anthropogenic source of

9 perchlorate nationwide, especially with its lowered perchlorate content.

10

9.1.2 Raw Material Use

12 As with many commodity chemicals, large scale purchases are dictated by cost of raw

13 materials, which are in turn influenced by transportation costs. Consequently, proximate (rather

14 than distant) sources of agricultural chemicals are likely to play the greatest roles in production

15 farming. Additionally, processing aids (e.g., clays) are likely to be derived from the nearest

16 sources.

17 Commodity chemicals used as agricultural fertilizers contain fairly high concentrations of

18 one, or sometimes two, of the primary plant nutrients, expressed as nitrogen (N), phosphorous (as

19 the oxide P_2O_5), or potassium (as the oxide K_2O). Trace metals (e.g., copper) can be applied

20 separately or along with these primary nutrients on a farm site. The primary phosphorus sources

21 are ammonium phosphates and triplesuperphosphate (a hydrous calcium phosphate). The

22 primary potassium source is potassium chloride. A mixture of synthetic and natural components

23 are used in fertilizer manufacture, described in detail elsewhere (U.S. Environmental Protection

24 Agency, 2001b).

25 Fertilizer application in production farming is highly dependent on the crop and the native

26 soil. Agriculture is influenced by climate, weather, topography, soil type, and other factors that

27 are generally similar within a geographical region; therefore, crops and fertilizer use are also

28 similar within such a region. For example, the Corn Belt relies heavily on urea and anhydrous

29 ammonia as nitrogen sources. Ammonium nitrate finds greater use in tobacco farming, and

30 potassium magnesium sulfate finds more use in milk-producing states. Because all plants require

31 the same primary nutrients, there is some fertilizer usage to provide these regardless of crop.

1   Local soil conditions also dictate what nutrients should be augmented, causing there to be large

2   regional variations.

3   Consumer fertilizer (specialty) products can be distributed over large geographical regions

4   because of the nature of the market. For example, major manufacturers have a limited number of

5   sites dedicated to blending multiple-nutrient formulations. These products are often sold as

6   bagged fertilizers through home-improvement centers, nurseries, florists, horticulturists, and

7   department (or other retail) stores. Unlike agricultural fertilizers, consumer products are usually

8   multi-nutrient formulations. In addition, trace metals are sometimes incorporated directly into

9   them. Because the average user will apply only a very small amount of trace metals (or even

10   primary nutrients) relative to a production farm, it is more economical, more practical, and more

11   convenient to use multiple-nutrient formulations. Moreover, the average consumer does not have

12   the wherewithal to disperse careful doses of several single-component fertilizers at the

13   appropriate times of the growing season.

14   Because fertilizer application on production farms is geographically delimited, there is

15   considerable interest in knowing which commodity chemicals might contain perchlorate—at

16   least in terms of dosing. Such information might suggest regions which should be investigated

17   for perchlorate contamination.   Moreover, it will be important to know what crops might be

18   affected—if any.

19

20   ## 9.1.3 Fertilizer Analysis Studies

21   Aside from the analyses of Chilean caliche, there were no studies to suggest that any other

22   processed fertilizer or raw material might contain perchlorate prior to 1998. That year, the

23   Ecosystems Research Division of the EPA's National Exposure Research Laboratory (NERL-

24   ERD) found perchlorate in several samples that were not derived from Chile saltpeter (Susarla,

25   1999a). This finding was later duplicated by other investigators from the North Carolina State

26   University College of Agriculture. However, the presence of perchlorate could only be

27   confirmed in consumer products, not in agricultural fertilizers. Moreover, subsequent analyses

28   of bags of the same materials acquired at a later date (likely from different manufacturing lots)

29   did not show perchlorate (Susarla et al., 2000). The choice of fertilizers did not account for the

30   possibility that the same raw materials must have been used in a variety of products at a point in

31   time. Additionally, a few major companies are responsible for making a large number of

1   products under several brand names. Furthermore, some companies rely on toll manufacturing

2   so that the products are actually made by another company to meet a specific formulation.

3   Accordingly, an error or contamination associated with one raw material could affect a variety of

4   products without regard to company or application.

5   Perchlorate was found in six of eight lawn and garden fertilizers tested, according to a

6   report provided to the EPA by the U.S. Air Force Materiel Command (TRC, 1998). However,

7   the report's authors were careful to point out that the results were obtained from a single

8   sampling event and that raw material usage was variable; therefore, no general conclusions could

9   be drawn. These qualifiers are consistent with the limitations enumerated above, but they do

10   point towards a temporal contamination of some products.

11   This study helped bring to light a number of important issues for trace analysis of

12   fertilizers. First, most of the research on determining perchlorate to that time had been focused

13   on either finished potable water or raw source water (Urbansky, 2000). Second, fertilizers are

14   considerably more complicated matrixes than dilute water solutions. Third, a solid fertilizer is

15   not a homogeneous substance. In particular, multi-component formulations used as lawn and

16   garden fertilizers are macroscopically heterogeneous and it is possible to sort out the particles

17   visually. Thus, representative subsampling becomes a key issue. Fourth, the effectiveness of

18   leaching out any perchlorate ion into an aqueous phase was unknown. Fifth, the products chosen

19   did not reflect the chemical fertilizers used for production farming, but rather the ingredients

20   used for lawn and garden fertilizers during a specific time period.

21   Around the same time, the U.S. Air Force Research Laboratories (AFRL) performed a

22   study to assess interlaboratory corroboration; that is, the ability of different labs to analyze the

23   same sample and get the same result (AFRL, 1999; Eldridge, 2000). A variety of techniques

24   performed by multiple laboratories showed acceptable agreement on the concentrations of

25   perchlorate in solutions prepared from the purchased products. Several limitations (such as

26   product choice and sampling difficulties with heterogeneous solid products) made it impossible

27   to gain an understanding of agricultural fertilizer use, and the AFRL intentionally restricted its

28   use of the data to evaluating interlaboratory agreement. However, data from the AFRL study was

29   sufficient to confirm independently that some lawn and garden fertilizer products did contain

30   perchlorate during a certain period of time.

1   Subsequently, the Water Supply and Water Resources Division of the EPA's National Risk

2   Management Research Laboratory (NRMRL) conducted its own survey of fertilizers in a

3   collaboration with the Oak Ridge National Laboratory (Urbansky et al., 2000a; Urbansky et al.,

4   2000b). In addition to a variety of products purchased from retailers, products were purchased

5   from farming supply stores (e.g., 50-lb bags of urea or ammonium nitrate) in Indiana, Ohio,

6   Kentucky, Pennsylvania, and Tennessee. In addition, commodity chemical samples were

7   collected from local distributors in Ohio and Indiana. These included urea, potassium chloride,

8   ammonium monohydrogen phosphate, and granular triplesuperphosphate, among others.

9   Samples were leached or dissolved and subjected to complexation electrospray ionization mass

10   spectrometry (cESI-MS) or ion chromatography (IC). Of 45 tested products, the only ones that

11   were found to contain any perchlorate were those based on Chile saltpeter. While this study was

12   the first to include the same products used on agricultural production farms, it did not address the

13   issues of sampling, product inhomogeneity, or geographical source variation.

14   In an effort to better address sampling, raw material usage, and other issues, the EPA

15   undertook an additional study of fertilizers. The project was divided into two phases, the first

16   part of which evaluated the testing laboratories for their ability to identify and quantitate

17   perchlorate in a fertilizer matrix. In the second phase, samples gathered under the supervision of

18   state agricultural agents were homogenized and sent to the laboratories for analysis using a

19   method established by the EPA (U.S. Environmental Protection Agency, 2001a). This

20   investigation was the most thorough in terms of including agriculturally relevant materials used

21   to manufacture a wide variety of specialty products or sold directly to farmers. It also spanned all

22   major national suppliers of these products. Although it reflected only a temporal snapshot, as

23   had all of the other studies, the survey of fertilizers incoporated the greatest number of unique

24   samples, quality control tests, and standardized practices, as well as other design improvements.

25   Four laboratories analyzed all of the materials, and some samples were analyzed by additional

26   laboratories. No other materials were found to contain perchlorate at measurable concentrations,

27   and the EPA concluded that the only clearly identifiable fertilizer source of perchlorate was

28   caliche. The data collected in this endeavor were additionally used to evaluate laboratory

29   performance and further validate the method (Urbansky and Collette, 2001). A set of archived

30   samples of all the Phase 2 materials was analyzed while evaluating an alternate ion

1   chromatographic column and independently verified all of the results reported in U.S.

2   Environmental Protection Agency (2001a) (DeBorba and Urbansky, 2001).

3   The findings reported in U.S. Environmental Protection Agency (2001a) are the most

4   comprehensive in terms of the types of materials tested, the manufacturers, the number of

5   laboratories analyzing each field sample of material, and the quality control checks. In these

6   regards, it represents our best understanding of fertilizers in terms of perchlorate content. While

7   the presence of perchlorate in the materials gathered in late 1998 through early 1999 remains

8   enigmatic, there is no evidence to support the concern that there is ongoing or routine perchlorate

9   contamination in the U.S. fertilizer supply. Reports in 1999 may have reflected the temporal

10   contamination of one or more raw materials or merely an error in manufacture. Based on the

11   studies reported to date (Collette and Williams, 2000; Gu et al., 2000; Urbansky et al., 2000a;

12   Urbansky et al., 2000b; Robarge et al., 2000; EPA, 2001b; Williams et al., 2001; DeBorba and

13   Urbansky, 2001), there is a consensus among researchers from the EPA, the fertilizer industry,

14   and other federal and state laboratories that currently used fertilizers are negligible contributors

15   to environmental perchlorate contamination. Even imported Chile saltpeter or products derived

16   from it contribute minimally due to their low use and low perchlorate content. Consequently, the

17   EPA has concluded that further investigation is unwarranted (U.S. Environmental Protection

18   Agency, 2001b).

19   IMC-Agrico, a major North American fertilizer manufacturer, has instituted its own

20   monitoring program for its raw materials and products as a result of continuing interest among

21   the scientific, industrial, and regulatory communities. These products include various potassium

22   ores (langbeinite, sylvinite), potash-based products (potassium chloride, potassium sulfate and

23   potassium magnesium sulfate), and phosphate products (ammonium monohydrogen phosphate,

24   ammonium dihydrogen phosphate and granular triplesuperphosphate). After more than 100

25   analyses using the latest method (EPA, 2001a), IMC reported to the EPA that no perchlorate was

26   detected in any of the materials it tested during a period spanning nearly three years. In addition,

27   IMC states that it has analyzed Magruder check samples for perchlorate. The Magruder check

28   sample program is jointly administered by the Association of American Plant Food Control

29   Officials and The Fertilizer Institute; it bears the name of a chemist from the F. S. Royster Guano

30   Company named E. W. Magruder, who initiated the program in 1922. The program selects,

31   prepares, and distributes samples of various materials and finished products to subscribing

1   laboratories and then collects and analyzes the data. Magruder samples reflect monthly

2   snapshots taken from the entire fertilizer industry. Perchlorate has not been detected in any IMC

3   product or any of 16 Magruder samples, according to IMC (personal communication from

4   William L. Hall).

5

6   **9.1.4 Complicating Factors**

7   It is worth pointing out at the U.S. Geological Survey (USGS) and Air Force Research

8   Laboratories have found perchlorate in isolated samples of sylvite taken from New Mexico

9   (Harvey et al., 1999). The USGS is engaged in additional sampling of North American mining

10   sites in order to assess whether there are natural mineral deposits of potassium perchlorate in

11   sylvite or sylvinite. Because little is known about the mechanisms of perchlorate formation in

12   the natural environment (which are assumed to be meteorological in nature), it is not clear

13   whether these findings represent a low-level background to be expected in evaporite mineral

14   deposits or not. Nonetheless, perchlorate has not been detected in any samples of agricultural

15   grade potassium chloride (0-0-62 or 0-0-60) taken under the direction of the EPA or by IMC-

16   Agrico. Accordingly, it appears that this mineral commodity does not suffer from inclusions of

17   perchlorate salts to any environmentally relevant extent.

18   Decades ago, ammonium nitrate was prepared from Chilean sodium nitrate by ion

19   exchange rather than by gaseous reactants. It appears that cost began to prohibit this practice for

20   fertilizer-grade ammonium nitrate. Nonetheless, some facilities appear to have continued the

21   practice for explosives-grade ammonium nitrate that was used for blasting in mining operations

22   throughout the American Southwest. It is unlikely that reliable data can be obtained from more

23   than the past 10 years or so. Prior to the establishment of nitric acid and ammonia factories,

24   natural saltpeters played significant roles in American agriculture. Thus, there may be

25   contamination of groundwater in regions where these materials were used historically. The lack

26   of information concerning natural attenuation, as well as a limited knowledge of hydrogeology,

27   makes it difficult to determine where and how such problem sites might be found. For this

28   reason, monitoring for perchlorate under the EPA's Unregulated Contaminant Monitoring Rule

29   can be expected to provide some of the most useful information.

30   Even though perchlorate was identified in some fertilizer products and was presumably

31   introduced through a contaminated raw material, this incident appears to have been entirely

1 isolated. Furthermore, awareness within the fertilizer industry and the environmental community

2 is now heightened to the point that it appears unlikely to happen again.

3

4

5 **9.2 MONITORING FATE AND TRANSPORT IN LIVING PLANTS**

6 Due to the reported occurrence of perchlorate in certain water resources and in certain

7 fertilizer products, several groups have begun to address the extent and significance of

8 perchlorate uptake by plants. For example, if produce is grown using irrigation water tainted

9 with perchlorate, or if agricultural soil is amended with perchlorate-tainted fertilizer, this might

10 constitute a route of human exposure if perchlorate is taken up and retained in the edible parts of

11 produce plants. The possibility of a relevant exposure route would be increased if perchlorate

12 was found to bioaccumulate and if it was shown to survive the various processes that edible

13 plants undergo before being consumed. Unfortunately, experimental results that definitively

14 gauge the extent of risk from this route of exposure have not yet been published. However, some

15 progress toward this goal has been made.

16

17 **9.2.1 Difficulties in Analyzing Plant Tissues and Other Environmental**
18   **Samples for Perchlorate**

19 One problem that has delayed accurate and definitive studies of perchlorate uptake by

20 edible plants is the difficulty of analyzing for perchlorate in plant materials. Ion chromatography

21 is currently the recommended method for routine analysis of inorganic ions such as perchlorate.

22 It is a sensitive, reliable, and easily-implemented technique when perchlorate occurs in a matrix

23 that has a relatively low level of total dissolved solids (TDS). Unfortunately, extracts of plant

24 materials contain high concentrations of TDS, inorganic ions, amino acids, sugars, fatty acids,

25 and nucleotides—all of which contribute to the ionic strength of the sample (Ellington and

26 Evans, 2000). In such matrices with high TDS/ionic strength, other ions can overwhelm the

27 conductivity detector and effectively mask the signal from perchlorate. Ion chromatography is

28 not unique in this regard. Other techniques and methods suitable for reasonably dilute drinking

29 water matrices (Urbansky et al., 2000c; Magnuson et al., 2000a, b; Urbansky et al., 1999;

30 Urbansky and Magunson, 2000) cannot be readily applied to fertilizers or botanical and

31 physiological fluids. The problems of trace ionic analysis have led to the development of other

1   methods that rely on expensive instrumentation, but are not generally available, such as

2   asymmetric waveform ion mobility mass spectrometry (Handy et al., 2000; Ells et al., 2000) or

3   tandem mass-spec (MS-MS) systems (Koester et al., 2000).

4   Recently Ellington and Evans (2000) have reported an IC-based method using an enhanced

5   clean-up procedure for the quantitation of perchlorate in plant materials that greatly reduces

6   interferences from dissolved matter. The minimum reporting level (MRL) of perchlorate in

7   lettuce and tomato was found to be approximately 250 μg/g on a wet mass basis. Lettuce and

8   tomato were chosen as representative plants because they are considered high priority candidates

9   for screening foodstuffs (Ellington and Evans, 2000). Perchlorate was spiked into the extraction

10   water for one half of the duplicate freeze-dried samples, while one half were extracted with pure

11   water. In the absence of other ions, some perchlorate is lost to the alumina used for the clean-up;

12   however, this should not impact application of the method to plant material because most

13   extracts have sufficient ionic strength. Note that perchlorate was not detected in any produce,

14   nor was the method applied to any edible plants that were grown with intentional exposure to

15   perchlorate.

16

17   ## 9.2.2 Ecological Transport

18   In the laboratory setting, some plant species will absorb perchlorate when exposed to

19   contaminated irrigation water. Uptake by plants has been explored for possible use in

20   phytoremediation (Nzengung, 1999; 2000). Some investigators have speculated that bacteria are

21   responsible for this phenomenon in plants. Perchlorate-reducing monera have been identified by

22   several laboratories, and cultured from a variety of sources (including Las Vegas Wash

23   sediments, food processing sludge, soils, and sewage sludge); (Logan, 1998; Coates et al., 1999;

24   Coates et al., 2000; Kim and Logan, 2001; Wu et al., 2001; Logan, 2001). Recent work showing

25   perchlorate reduction in saline solution suggests that attenuation may be possible even in briny

26   locations (e.g., the Las Vegas Wash) or in fertilizer-laden farm runoff (Logan et al., 2001; Okeke

27   et al., 2001). This suggests that perchlorate-reducing bacteria are present at significant levels in

28   the environment. On the other hand, the bacteria isolated thus far prefer oxygen over nitrate over

29   perchlorate. In order to for perchlorate reduction to occur, the water must be anoxic and all of

30   the nitrate must have been consumed. Moreover, these bacterial cultures require a suitably moist

31   environment; arid soils or regions with low rainfall may not sustain their growth. Natural

1   attenuation probably varies around the nation, depending on local factors. Accordingly, it is not

2   possible to draw any meaningful conclusions about the ecological impact of fertilizers that

3   contain perchlorate, for they may or may not be applied in areas where this type of bacterial

4   degradation can occur.

5   Another factor that has prevented the early materialization of definitive data on risk from

6   perchlorate in edible plants is that many researchers who have addressed plant uptake of

7   perchlorate are primarily interested in other aspects of the problem. For example, Ellington et al.

8   (2001) have applied the optimized IC-based method described above first to the analysis of

9   perchlorate in tobacco plants and tobacco products. Tobacco was chosen because it is grown in

10   some locations in soils amended with Chile saltpeter.

11   Ellington and Evans (2000) obtained green (uncured) tobacco leaves from the Coastal Plain

12   Experiment Station (CPES) in Tifton, GA in late July 1999. The plants grew in soil that had

13   been amended with two fertilizer products, one of which was Chile saltpeter. The perchlorate

14   level in the Chile saltpeter was 1.5 mg/g, consistent with contemporaneous reports (Urbansky

15   et al., 2001; personal communication from W.P. Robarge). Perchlorate was also found in a

16   6-6-18 plant food that had been applied to the same soil. While 3% of the nitrogen was from

17   nitrate, the perchlorate concentration was only 36 μg/g; whereas, based on the typical perchlorate

18   content in Chile saltpeter, it should have been about eight times larger if all of the nitrate were

19   from Chile saltpeter. This suggests that synthetic nitrates were also part of the fertilizer's

20   constitution. Perchlorate concentrations in the dried tissue varied from 12.5 to 165 μg/g,

21   depending on the portion of the leaf examined and the curing process employed. Soil samples

22   leached with deionized water contained 0.3 μg/g on a dry weight basis. EPA researchers also

23   analyzed several off-the-shelf cigarettes (2 brands), cigars (1 brand), and chewing tobacco

24   (7 brands) and found perchlorate concentrations ranging from 0.4 to 21.5 μg/g (undried), and

25   only one product that contained none (Wolfe et al., 1999; Ellington et al., 2001). They confirmed

26   the IC results by chlorine NMR spectrometry and capillary electrophoresis. Collectively, these

27   observations argue that tobacco plants can take up perchlorate from perchlorate-contaminated

28   fertilizers via the soil. Furthermore, they indicate the importance of investigating whether crop

29   plants can accumulate perchlorate in their edible portions and whether any contamination can

30   persist through the processing that precedes consumption.

1   Several groups have looked at the accumulation of perchlorate in various inedible plants as

2   a potential means of fate and remediation. Perchlorate-tainted water from the Las Vegas Wash

3   enters Lake Mead and the Colorado River and therefore has the potential to affect the potable

4   water of many people as well as the irrigation water used for much of the lettuce produced in the

5   U.S. Salt cedar (*Tamarix ramosissima*) is an invasive woody plant that grows prolifically in and

6   around the Las Vegas Wash. Salt cedar consumes and transpires an enormous amount of water

7   when it is actively growing. Furthermore, it accumulates and secretes salt. For these reasons,

8   Urbansky et al., (2000d) have analyzed samples of salt cedar that were taken from the Las Vegas

9   Wash. They found perchlorate at 5-6 μg/g in dry twigs extending above the water and 300 μg/g

10   in stalks immersed in the water from a plant growing in a contaminated stream, suggesting that

11   salt cedar plays a role in the ecological distribution of perchlorate.

12   Still others have investigated plant uptake with the specific goal of identifying remediation

13   strategies for perchlorate. The biodegradation of perchlorate in woody plants has been

14   investigated as a means of phytoremediation (Nzengung et al., 1999; Nzengung and Wang,

15   2000). Nzengung et al. (1999) and Nzengung and Wang (2000) found that willow trees (genus

16   *Salix*) were able to decontaminate aqueous solutions containing 10–100 mg/L of perchlorate to

17   below the method detection limit of 2 μg/L and suggest that two distinct phytoprocesses were at

18   work in their studies. Specifically, they observe evidence for rhizodegradation from the exudates

19   released from the plant, and—more importantly from the standpoint of relevance for food safety

20   issues—they see accumulation in branches and leaves. Only about 11% of the perchlorate spiked

21   into the water in which the trees were grown was found to phytodegrade in 26 days. The

22   majority of perchlorate that was removed from solution after 26 days was found in the leafs.

23   Longer term experiments suggest that the perchlorate did not accumulate in the leaves, but was

24   very slowly transformed there as well. Generally, the perchlorate level in the leaves increased to

25   a maximum before decreasing to undetectable levels after perchlorate was completely removed

26   from solution. Nzengung et al. assumed that the phytodegradation pathway of perchlorate leads

27   to chloride. Moreover, Nzengung et al. explored the role of other anions in the removal of

28   perchlorate in solution. They found that the perchlorate removal rate was decreased as the NO_3^-

29   level was increased. This was attributed to competing reactions in which both anions were

30   utilized as electron acceptors. Clearly this has relevance for the food safety issue and should be

31   investigated further. For example, the type of fertilizer used in food crop production may have

1   an effect on the degree to which perchlorate is taken up, depending on the major components of

2   the fertilizer.

3   Susarla and coworkers have published results of their investigations on transformation of

4   perchlorate by a wide range of plant types. For example, Susarla et al. (1999b,c) have performed

5   screening studies to determine what species might show potential for further investigations of

6   perchlorate phytoremediation. Thirteen vascular plant species were selected for evaluation in

7   these preliminary experiments. This included four tree species, four herbaceous wetland species,

8   four aquatic species, and one herbaceous upland species. Laboratory-scale experiments were

9   conducted in order to, among other things, evaluate the ability of these plants to remove

10 perchlorate from solution, evaluate the role of nutrients on perchlorate removal, and determine

11 the fate of perchlorate removed form solution (e.g., plant tissue distribution, accumulation versus

12 breakdown). Each of these topics is indirectly relevant to the issue of uptake by edible plants.

13   For all of these experiments, perchlorate concentrations of 0.2, 2.0 and 20 mg/L were tested

14 in aqueous and sand treatments for ten-day periods. Perchlorate was found to be depleted from

15 solution in the presence of all but two species. Susarla et al. (1999a,c) used a system of five

16 categories to classify the performance of the species based on the degree to which they depleted

17 the solution. None of the trees tested were included in the highest category of performance, but

18 some of the wetland and aquatic plants were. Plant tissue (e.g., roots, stems, leaves) were

19 analyzed from samples that demonstrated the maximum drop in perchlorate concentration.

20 Susarla et al. (1999a,c) report perchlorate, or some transformation metabolite (chlorate, chlorite,

21 chloride), in all tissues analyzed. Results of these studies suggested significant influences on

22 depletion of perchlorate from, among other things, growth substrate (sand versus aqueous

23 treatment), the level of nutrients, stage of plant maturity, and the presence of other ions. All of

24 these influences should prove to be valuable insights when considering the uptake of perchlorate

25 by edible plants. Based on screening studies, additional studies focused on the

26 phytotransformation of perchlorate by the aquatic plant parrot-feather (*Myriophyllum*

27 *aquaticum*); (Susarla et al., 1999b; Susarla et al., 1999c).

28   Tobacco is one crop for which the use of Chilean nitrate salts can be documented in some

29 locations. In northern Kentucky, these products are primarily used for seedling beddings rather

30 than fertilizing fields; for various reasons, ammonium nitrate is preferred by many farmers in

31 Kentucky. Such preferences vary throughout tobacco-producing states and regions, however.

1   Data on application of Chile saltpeter is sparse, and it is not possible to estimate the ecological

2   impact in any meaningful way. There can be no question that at least some vascular plants

3   absorb perchlorate from their local environments. Furthermore, perchlorate has been found in a

4   number of plants and animals living in contaminated environs (Smith et al., 2001). An obvious

5   concern raised by finding measurable perchlorate concentrations in plant tissues is whether this

6   ion can affect food crops and what factors might influence its uptake and accumulations. These

7   issues shall be considered next.

8

9.2.3 Extrapolating to Food Plants

10   Because so much U.S. produce is fertilized with perchlorate-free chemical commodities,

11   the risk from exposures via fertilizers is small. Some crops (e.g., corn, wheat, and rice) are

12   fertilized with materials that are unquestionably perchlorate-free. Additionally, there is no reason

13   to suspect any perchlorate associated with growing grains. However, the risk of exposure

14   resulting from irrigation with perchlorate-tainted water in the American Southwest is unknown.

15   At present, there are no efforts to test fruits and vegetables for perchlorate. Many of the studies

16   on uptake by plants have been based on concentrations higher than those encountered in

17   irrigation water. Furthermore, some products derived from Chile saltpeter are known to be

18   among those used on California citrus crops.

19   One of the few studies of perchlorate uptake by edible plants is the ongoing work of

20   Hutchinson and coworkers with lettuce grown in a greenhouse with perchlorate-tainted irrigation

21   water. Lettuce is of particular importance for assessing the risk of perchlorate to the food supply

22   since much of the lettuce produced in the U.S. is irrigated by water that is fed by the Las Vegas

23   Wash, which is contaminated with perchlorate. Also, lettuce has a high water content and

24   virtually the entire above-ground plant is consumed without cooking or processing. These

25   characteristics would present a potential risk if lettuce efficiently accumulates perchlorate.

26   Hutchinson and coworkers are irrigating lettuce plants with five different concentrations of

27   perchlorate (0.1, 0.5, 1.0, 5.0, and 10.0 μg/L) for a period of 90 days following planting.

28   At various intervals of time they divide the plants into green tissue and root samples and analyze

29   each sample for perchlorate using an analytical method adapted from Ellington and Evans

30   (2000). Their results show an accumulation of perchlorate into the green tissue. The level of

31   perchlorate built up steadily over the first 50–60 days of the experiments, then generally leveled

1   off. At about 50 days into the experiment, the lettuce irrigated with 10.0 ppm perchlorate

2   exhibits a perchlorate content of about 3 mg/g on a lettuce dry matter basis. Since lettuce is

3   about 90% water, this would amount to about 0.300 mg/g on a wet weight basis. The amount of

4   perchlorate detected in the leaves is generally linear with dosing level for a given day.

5   Experiments are underway to determine whether lettuce has the capability to degrade perchlorate

6   if the supply of the contaminant is stopped. However, this determination is frustrated by the fact

7   that lettuce continues to grow. Therefore, a decline in concentration (e.g., expressed in mg/g)

8   does not adequately reflect the situation. The preliminary results from these studies (Hutchinson

9   et al., 2000) suggest that, when complete, they will constitute considerable progress on the issue

10   of exposure to perchlorate from edible plants.

11   Even if many food plants can be shown to absorb and retain perchlorate, the primary source

12   of the contaminant is irrigation water polluted from defense-related activities. Because these

13   activities are reasonably localized geographically, most of the country's agricultural products

14   should be perchlorate-free, e.g., corn, wheat, rice, milk. On the other hand, some types of

15   produce are supplied almost entirely by regions dependent on contaminated irrigation water.

16   Therefore, these sites represent possible exposure routes for most of the nation via foods such as

17   lettuce.

18   Historically, much of the emphasis on fertilizer pollution from agricultural runoff has been

19   on fertilizers applied to the soil. However, potassium nitrate is usually applied to the leaves of

20   citrus trees when a potassium deficiency is found by analyzing leaf tissue. Such foliar

21   application would not necessarily contribute significantly to runoff type pollution of waterways,

22   but could lead to the absorption of contaminants through the leaves and wood. There are no

23   reliable data on the sources of potassium nitrate used for citrus crops. While it is known that

24   absorption of anions similar to perchlorate (e.g., pertechnetate) are affected by the ionic strength

25   and compostion of the surrounding solution, little is known about the factors that influence

26   perchlorate influx via roots or leaves. In addition, the fate of absorbed perchlorate in the plants is

27   also unknown. It may be that xylem-supplied tissues, such as leaves, are the final repository

28   rather than phloem-supplied tissues, such as fruits.

29   These issues and more have begun to be examined by the EPA, but there are many

30   unknowns (U.S. Environmental Protection Agency, 2001b). Until such time as quantitative

31   studies are performed on various species to determine what factors influence the absorption,

1 accumulation, and distribution of perchlorate in plants, it is not possible to estimate whether

2 foods can serve as meaningful contribution to the body burden or to the risk posed to humans

3 from perchlorate contamination. Even if they do, there is considerable peace of mind in knowing

4 that fertilizers and water supplies are generally not providing any perchlorate to the plants in the

5 first place. Consequently, only a small number of foods are worth considering for further study.

6 On the other hand, it is not known to what extent other countries rely on natural saltpeters to

7 fertilize food crops. Moreover, it is not known whether fruits and vegetables absorb and retain

8 the perchlorate ion. Therefore, it is not possible to say whether fruits and vegetables grown

9 outside the U.S. serve as a possible exposure routes at this time. Depending on the season,

10 imported oranges, apples, and grapes and their juices are consumed throughout the U.S.

11 Because there are no data on perchlorate in imported produce, no data on perchlorate in

12 U.S. produce, and no data from controlled laboratory experiments on uptake in fruit crops, it is

13 impossible to assess whether these foods can contribute to perchlorate consumption in humans or

14 whether drinking water constitutes the entire body burden. However, the available information

15 on fertilizers and irrigation water suggests that foods do not contribute to the body burden. At

16 the present time, the available data point towards drinking water as the principal exposure

17 pathway for humans.

18

19

20 **9.3 SUMMARY**

21 Despite some initial findings implicating fertilizers as a source of perchlorate, more

22 thorough and better designed studies that were conducted subsequently have not found this to be

23 the case. Current fertilizer manufacturing practices and raw material sources make it unlikely

24 that perchlorate contamination could occur widely and without discovery. While some plants

25 may absorb or even accumulate perchlorate in specific tissues, there are many unknowns with

26 regard to the edible portions of nutritionally and agriculturally important crops. Many factors

27 influence transport of ions, and current understandings of plant physiology and botany suggest

28 perchlorate uptake would be reduced as a result of such factors. Even if perchlorate uptake

29 occurred in some food crops, perchlorate contamination is localized geographically outside of

30 major agricultural regions, minimizing possibility of uptake in edible produce. While

31 perchlorate-tainted irrigation water may be a source available for uptake of perchlorate by plants,

1	this is again localized, and has not been proven to occur at the concentrations of perchlorate that

2	are observed environmentally. Difficulties in analyzing many plant or animal tissues originally

3	were obstacles to executing appropriate studies, but these problems have generally been solved.

4	Ideally, more data would be available on food plants, but current evidence suggests that drinking

5	water is the primary exposure pathway to perchlorate for humans.

6

10. MAJOR RISK CHARACTERIZATION CONCLUSIONS

10.1 HUMAN HEALTH

This section summarizes major findings regarding human health presented in Chapters 1, 2, 3, 4, 5, and 7.

10.1.1 Hazard Potential

Perchlorate is an anion that originates as a contaminant in ground and surface waters from the dissolution of ammonium, potassium, magnesium, or sodium salts. Ammonium perchlorate is the oxidizer and primary ingredient in solid propellant for rocket motors. Perchlorate salts also are used on a large scale as a component of air bag inflators and in the manufacture of pyrotechnics and explosives. Solid rocket inventories are growing at a significant rate as systems reach the end of their service life: the solid rocket disposal inventory is expected to be over 164 million lb by the year 2005. Because the accepted method for removal and recovery of solid rocket propellant is high-pressure water washout, a large amount of aqueous solution containing ammonium perchlorate is generated. A number of locations where perchlorate has been detected in groundwater or surface waters are in areas associated with the development, testing, or manufacturing of aerospace materials. Perchlorate contamination also occurs when explosives are used extensively, e.g., open burn/open detonation operations and some mining activities.

Perchlorate is rapidly absorbed from the gastrointestinal tract, whereas dermal and inhalation exposures are not expected to be significant exposure routes for the general public. The known mode of action for perchlorate is that it acts as a competitive inhibitor of active iodide uptake by the *sodium (Na⁺)-iodide (I⁻) symporter (NIS)* in most mammals, including humans, laboratory test species, and wildlife. This decrease in intrathyroidal iodide results in a decreased production of T3 and T4 thyroid hormones. Decrements in thyroid hormones can cause permanent neurodevelopmental deficits and impair adult organisms as well. A decrease in thyroid hormones can also potentially perturb the hypothalamic-pituitary-thyroid axis to increase the pituitary's production of TSH and, consequently, stimulate the thyroid to increase production

1   of thyroid hormone in an attempt to compensate. Prolonged stimulation of the thyroid by TSH

2   may result in thyroid neoplasia, particularly in rodents known to be sensitive. Tumors have

3   occurred in rats dosed with high levels of perchlorate for long periods and at much lower doses in

4   relatively young adult animals (19 weeks) dosed *in utero* and during development. These

5   findings have raised concerns about the *in utero* imprinting of the regulatory system responsible

6   for controlling thyroid hormone economy.

7   The target tissue for systemic effects of perchlorate has been identified as the thyroid. The

8   key event of its mode of action is iodide uptake inhibition at the NIS. Changes in the thyroid

9   hormone homeostasis result in histopathological changes in the thyroid, including: colloid

10 depletion, follicular hypertrophy, follicular hyperplasia, and decrease in follicular lumen size.

11 If perchlorate exposure is stopped, the thyroid histopathological effects have been shown to be

12 reversible after exposures as long as 90-days in rats, but incomplete recovery of thyroid

13 hormones occurs in this same time period. There are also some case studies in humans treated

14 therapeutically with perchlorate that indicate reversibility of thyroid hormone changes after years

15 of exposure.

16   Other potentially adverse and permanent effects from decreased thyroid hormone include

17 effects during development *in utero* and early growth, particularly effects on the nervous system

18 if the pregnant mother was hypothyroxinemic or hypothyroid. Laboratory animal assays

19 performed in response to recommendations made at the peer review in 1999 and as part of the

20 perchlorate testing strategy confirmed neurodevelopmental effects observed in previous studies.

21 Changes in brain morphometry and motor activity were observed. The potential for major

22 disturbances in thyroid hormone homeostasis to disturb reproductive capacity or to induce

23 immune effects also exists. The ability of perchlorate to cause contact hypersensitivity is

24 suggested but remains not well characterized. Finally, a remarkable conservation of the thyroid

25 hormone regulatory system has been demonstrated across species. Inhibition of iodide uptake by

26 the NIS has been shown in pharmacokinetic studies to be very similar across species, including

27 humans.

28

29 **10.1.2 Dose Response**

30   The revised RfD is based on an assessment that reviewed a set of studies that were

31 developed to explicitly evaluate these potential toxicities. The quantitative estimate of risk is

1   based on laboratory animal data because there are no good observational epidemiological data

2   concerning human subjects representative of the critical sensitive populations (hypothyroxinemic

3   pregnant women or children) or that have evaluated neurodevelopmental outcomes; nor have

4   adequate clinical studies been performed. A harmonized approach was proposed based on the

5   key event of iodide inhibition and its relationship to disturbances in the hypothalamic-pituitary-

6   thyroid axis as evidenced by effects on thyroid and pituitary hormones, thyroid histopathology,

7   and brain morphometry. Using these precursor lesions as the basis for the point-of-departure is

8   considered to be protective for cancer development as well as for neurodevelopmental sequelae.

9   The database supported a point-of-departure for the RfD deviation at 0.01 mg/kg-day based

10   on changes in maternal thyroid and pituitary hormones and on changes in the brain morphometry

11   and thyroid and pituitary hormones of fetal and neonatal pups. A composite uncertainty factor of

12   300 was applied in the derivation. An adjustment also was made for administration of

13   perchlorate as ammonium perchlorate. The RfD is for perchlorate as the anion because that is

14   what is sampled and analyzed in environmental media and because the salts of perchlorate

15   readily dissolve. Uncertainty factors were applied for intrahuman extrapolation, the use of a

16   LOAEL, concern regarding the lack of studies of longer duration and database deficiencies.

17   Confidence in the study, the database, and the RfD is rated as medium. A major uncertainty is

18   the sensitivity that the screening neurodevelopmental studies provide to protect against

19   neuropsychological deficits of exposures that might occur within critical developmental windows

20   or in susceptible human populations.

21   The daily perchlorate exposure to the human population that is likely to be without

22   appreciable risk of either cancer or noncancer toxicity during a lifetime is 0.00003 mg

23   perchlorate/kg-day. It again is noted that this RfD is specific for the anion because that is what is

24   detected in most environmental samples and because most salts of perchlorate readily dissolve.

25   Because of the application of uncertainty factors, this dose is approximately 1/300 of the dose

26   that resulted in brain morphometry and thyroid changes in pups and hypothyroid status

27   (decreased T4 and increased TSH) in rat mothers (Argus Research Laboratories Inc, 2001) and in

28   their pups both during gestation (GD21) and in the post-natal period (PND4 through PND21).

29

30

1   ## 10.1.3 Risk Characterization

2   Comprehensive risk characterization for the perchlorate contamination issue, as discussed

3   in Chapter 1 (see Figure 1-5), requires accurate information on exposure levels determined by a

4   validated analytical method. Dose-response estimates such as the value derived herein can then

5   be used to gauge the potential toxicity of those exposures. Exposure can be either direct, most

6   likely by ingestion, or indirect, such as by consumption of contaminated food. When using the

7   dose-response assessment derived herein to compare with exposure estimates, one should remain

8   keenly aware that many of these exposure aspects have not yet been characterized accurately for

9   perchlorate. Fate and transport information do not exist to track the spatial and temporal

10   distribution of perchlorate; the potential for evaporative concentration in soils has not been

11   characterized, nor has its uptake in plants or herbivores. In addition, there are uncertainties

12   remaining in the dose-response estimate itself. These concerns also should be considered

13   whenever attempting to characterize the risk to a specific human population exposed to a

14   particular scenario.

15

16   ### 10.1.3.1 Direct Exposures

17   Typically the RfD is used as a comparison for oral ingestion, such as by drinking water.

18   The RfD is compared with an exposure estimate of the drinking water concentration to

19   characterize potential toxicity. When making this comparison, the assumptions underlying

20   derivation of the RfD must be kept in mind. The RfD is intended to be protective of susceptible

21   populations exposed daily. The frequency and magnitude of exposure is a key attribute of

22   accurate dose-response characterization (Jarabek, 1995c) and an equally important component of

23   risk characterization. Transient decreases in T4 can cause permanent neurodevelopmental

24   deficits. Thus, the degree to which the particular suspected population at risk fits with the

25   underlying assumptions of the RfD derivation should be kept in mind. Finally, the degree of

26   imprecision in the derivation of an RfD should be taken into account. The RfD estimates are not

27   intended to serve as "bright line" estimates. By definition, there is an order of magnitude

28   uncertainty around the estimate. This generally translates into a range of approximately

29   three-fold below to three-fold above the RfD, but also depends on the nature of the effects used

30   as the basis.

31

1   **10.1.3.2   Indirect Exposures**

2   Where crops are irrigated with perchlorate-contaminated water, indirect human exposures

3   may result. A number of factors need to be considered in estimating human exposure through

4   crops.

5   Concentration in plant parts as a result of root uptake normally is calculated using a soil-to-

6   plant transfer factor that is expressed as the ratio of plant to soil concentration. If perchlorate is

7   subject to evaporative concentration in irrigated soils, then soil concentration, and therefore

8   uptake, may be higher than that expected simply based on concentration in irrigation water. If a

9   leaf crop such as lettuce is spray-irrigated, perchlorate could be concentrated evaporatively on

10   external leaf surfaces. Because perchlorate salts have high water solubility, this contamination

11   probably would be removed largely by washing. On the other hand, if perchlorate is

12   phytodegraded, as one study has suggested (Nzengung, n.d.), soil or plant concentrations may be

13   lower than otherwise expected. Studies are needed to determine the behavior and fate of

14   perchlorate in plant-soil-water systems, including studies that simulate leaf crop irrigation and

15   that account for full life cycles of crops.

16   Besides estimates of perchlorate concentrations in crops, the calculation of human daily

17   intake depends on the number of crop types that are contaminated, the extent to which a

18   particular individual obtains the crops from a contaminated source, and the individual's daily

19   consumption of the crops. These factors may vary widely in the exposed population, and

20   methods for accounting for the combined variability should be used in characterizing these

21   exposures.

22   Methods for estimating human exposures resulting from crop uptake of soil-deposited

23   contaminants are presented in Chapters 6 (Determining Exposure Through the Terrestrial Food

24   Chain) and 10 (Risk Assessment) of the EPA document, "Methodology for Assessing Health

25   Risks Associated with Multiple Pathways of Exposure to Combustor Emissions (EPA 600/

26   R-98/137)." That document currently is undergoing revision and is scheduled for final release in

27   January 2002. If the needed information can be obtained on perchlorate behavior and fate, the

28   methods described therein can be used to develop estimates of human exposure and risk.

29

30

10.1.4 Major Uncertainties and Research Needs

Reliable exposure estimates are required to accurately and comprehensively characterize the risk of perchlorate contamination. This section will briefly summarize research needs associated with aspects of uncertainty about the human health risk dose-response estimate that were highlighted in Chapter 7.

The greatest need for continued improvement in the dose-response assessment is a more accurate characterization of the linkage between the key event of the mode of action (i.e., inhibition of iodide uptake in the thyroid gland), subsequent changes in thyroid hormones, and the correlation to outcome measures in hypothyroxinemic pregnant animals and their pups. Because this need must be addressed in the fetal compartment as well, accurate characterization of toxicokinetics during pregnancy and lactation also are required. More definitive studies of the degree of change in perturbation of the hypothalamic-pituitary-thyroid axis (i.e., change in hormone levels) that is associated with thyroid histology, and with neurobehavioral deficits especially, would improve the confidence in the accuracy of the exposure-dose-response continuum. The current studies may need to be repeated with larger sample sizes and lower doses, and new studies may be needed to evaluate effects on fetal hormone levels and neurodevelopmental measures both in the laboratory and in a survey of the human population. Research on potential factors influencing sensitivity is also critically requisite. Animal models of thyroid impairment such as iodide deficiency and "womb to tomb" exposure designs should be explored. Finally, mechanistic determinants of these toxicokinetic and toxicodynamic parameters and processes should be further characterized.

10.2 ECOTOXICOLOGY

10.2.1 Aquatic Life

Procedures for deriving Tier II water quality values were used in Section 8.3.2.1 to jointly characterize the potential effects of the perchlorate ion on the richness and productivity of fish, aquatic invertebrate, and plant communities. Tier II values are derived when data are not sufficient for deriving ambient water quality criteria. The Tier II value derivation procedures account for missing information with approximately 80% confidence. In this case, the Tier II

1   values derived, termed secondary acute and chronic values, were 5 and 0.6 mg/L (i.e., 5,000 and

2   600 μg/L), respectively; difficulties associated with the interpretation of one test result in an

3   uncertainty range for the secondary chronic value of < 0.32 to > 0.83 mg/L (< 320 to

4   > 830 μg/L). Perchlorate levels reported for large surface waters (as high as 16 μg/L) and ground

5   waters (as high as 280 μg/L in public supply wells) are well below the secondary acute and

6   chronic values. Thus, at these exposure levels, the likelihood of effects on the richness and

7   productivity of fish, aquatic invertebrate, and plant communities appears to be low. However,

8   because much higher perchlorate concentrations have been reported in monitoring wells at rocket

9   motor manufacturing or testing sites (37×10^6 μg/L) and in groundwater-dominated surface

10   water systems close to sites of contamination (3500 to 1.3×10^5 μg/L), sites clearly exist that

11   have perchlorate concentrations high enough to cause toxicity to aquatic life. These sites include

12   springs, such as that sampled along Las Vegas Wash in Nevada (Parsons, 2001) and the INF

13   Pond at Longhorn Army Ammunition Plant in Texas (Parsons, 2001; Smith et al., 2001). On the

14   other hand, concentrations below the Tier II values were detected in larger water bodies

15   immediately adjacent to sites of contaminations, such as in Lake Mead immediately adjacent to

16   the mouth of the Las Vegas Wash (less than 4 to 68 μg/L). Water discharged from a CERCLA

17   groundwater pump-and-treat facility that was not equipped to treat perchlorate at Allegany

18   Ballistics Laboratory to the North Branch Potomac River contained 250 to 280 μg/L perchlorate

19   (Parsons, 2001).

20   Where high levels of contamination exist, sensitive aquatic organisms such as daphnids

21   may be the most likely to experience effects; in the reported tests, effects were seen on both

22   survival and reproduction (neonates per organism). A teratogenicity assay, FETAX, showed

23   malformations in frog embryos occurring at only slightly lower concentrations than lethality,

24   indicating that perchlorate is probably not a potent developmental toxicant. Tier II values are not

25   estimated for plants, but results from algal toxicity tests suggest that even at the higher

26   perchlorate concentrations associated with rocket motor manufacturing, risk of toxicity to aquatic

27   plants is low.

28   The perchlorate anion can be associated with various cations including sodium,

29   ammonium, and potassium. When sodium perchlorate was tested, the sodium cation was not

30   toxic to daphnids in sodium chloride control tests but did show toxicity to minnows.

31   Ammonium controls were not used in tests with ammonium perchlorate, but ammonium ion is a

1   known toxicant with toxicity that varies according to water temperature and pH. In any aquatic

2   system where perchlorate is present, attention should be given to determining the concentrations

3   of potentially toxic cations that may contribute to ecological effects.

4   Based on a secondary chronic value of 600 μg/L (uncertainty range, < 320 to > 830 μg/L)

5   for perchlorate, the analytical detection methods for perchlorate in water are sufficient. The

6   detection limit achieved for perchlorate in water was 4 μg/L (Parsons, 2001; Smith et al., 2001),

7   which is much less than the secondary chronic value. Thus, the likelihood that adverse

8   ecological effects will occur below detection limits is low.

9

10.2.2 Risks to Consumers of Aquatic Life

11   Information from Parsons (2001) and Smith et al. (2001) indicate that perchlorate may

12   bioaccumulate in aquatic invertebrates and fish in contaminated waters, but perchlorate is not

13   expected to bioconcentrate in these organisms to levels exceeding the surface water

14   concentrations. Therefore, there currently is no indication that consumers of aquatic

15   invertebrates or fish are at increased risk of effects from bioconcentration in areas where

16   perchlorate concentrations in surface water occur. However, there is some uncertainty about the

17   potential for bioaccumulation of perchlorate at low concentrations (i.e., 4 to 300 μg/L in water)

18   because of the higher detection limits for perchlorate in animal tissues, which were 300 to 400

19   μg/kg in Parsons (2001) and about 70 μg/kg in Smith et al. (2001). Furthermore, perchlorate

20   may bioconcentrate (i.e., to levels exceeding those in water) in aquatic plants; therefore,

21   consumers of aquatic plants may be at greater risk than consumers of aquatic invertebrates or

22   fish, but information is not available concerning effect levels in aquatic herbivores.

23

10.2.3 Terrestrial Life

10.2.3.1   Plants

26   Terrestrial plants may be exposed to perchlorate in soil at disposal sites and at sites

27   irrigated with contaminated surface water or groundwater. Perchlorate concentrations in soil at

28   disposal sites range from less than 1 to 1470 mg/kg (Parsons, 2001) and can be higher than the

29   screening benchmark of 4 mg/kg and even higher than the lethal concentrations ($\geq$180 mg/kg;

30   EA Engineering, Science and Technology, Inc., 1998).

1   In the absence of reliable information concerning the accumulation of perchlorate in

2   irrigated soils, it may be assumed that soil concentrations equal irrigation-water concentrations

3   (Section 8.3.1.3). Reported surface-water concentrations in the Colorado River, 4 to 16 μg/L,

4   would translate to 0.004 to 0.016 mg/kg. At the Yuma site, there was a single detection in

5   surface soil of 0.090 mg/kg; all other measurements were below the detection limits of 0.079 to

6   0.080 mg/kg (Parsons, 2001). Even the single detected concentration is a factor of 44 lower than

7   the benchmark value. The reported groundwater concentration in public wells of 280 μg/L

8   would translate to 0.28 mg/kg, which is a factor of 14 lower than the benchmark value. Hence,

9   perchlorate does not appear to constitute a hazard to plants irrigated with surface water.

10 However, given the large uncertainties concerning exposure, a hazard from groundwater

11 irrigation cannot be precluded.

12   Based on this screening benchmark of 4 mg/kg for perchlorate, the analytical detection

13 methods for perchlorate in soil are sufficient for determining whether soils will cause toxicity to

14 plants, and there is little likelihood of adverse ecological effects occurring at levels below

15 detection limits. The detection limit achieved for perchlorate in soils was generally 75-80 μg/kg

16 (Parsons, 2001), but there was at least one soil sample where the reporting limit was 803 μg/kg.

17 However, all of these limits are less than the screening benchmark.

18

19 **10.2.3.2   Soil Invertebrates**

20   Soil invertebrates may be exposed to perchlorate in soil at disposal sites and at sites

21 irrigated with contaminated surface water or groundwater. Perchlorate concentration

22 measurements at disposal sites range from less than 1 to 1470 mg/kg (Parsons, 2001) and,

23 therefore, can exceed the soil screening benchmark of 1 mg/kg. In the absence of reliable

24 information concerning the accumulation of perchlorate in irrigated soils, it may be assumed that

25 soil concentrations equal irrigation water concentrations (Section 8.3.1.3). Reported surface

26 water concentrations in the Colorado River, 4 to 16 μg/L, would translate to 0.004 to

27 0.016 mg/kg in soils. At the Yuma site, the single detection in surface soil was 0.090 mg/kg with

28 detection limits of 0.079 to 0.080 mg/kg. This detected concentration is a factor of 11 lower than

29 the soil screening benchmark value (1 mg/kg). The reported groundwater concentration in public

30 wells of 280 μg/L would translate to 0.28 mg/kg, which is a factor of 4 lower than the

31 benchmark value. Hence, perchlorate does not appear to constitute a hazard to soil invertebrates

1   in soil irrigated with surface water. However, given the large uncertainties concerning exposure,

2   a hazard from groundwater irrigation cannot be precluded.

3   Based on this screening benchmark of 1 mg/kg for perchlorate, the analytical detection

4   methods for perchlorate in soil are sufficient, and there is little likelihood of adverse ecological

5   effects occurring at levels below detection limits. The detection limit achieved for perchlorate in

6   soils was generally 75-80 μg/kg (Parsons, 2001), but there was at least one soil sample where the

7   reporting limit was 803 μg/kg. However, all of these limits are less than this screening

8   benchmark.

9

10   **10.2.3.3   Herbivores**

11   Exposures of voles to perchlorate based on measured plant concentrations at rocket motor

12   manufacturing or testing sites (0.11 mg/kg day to a maximum of 49 mg/kg day) exceed both the

13   LOAEL of 0.01 mg/kg/day and the screening benchmark of 0.001 mg/kg day**.** Estimated

14   exposures of voles consuming plants on sites irrigated with surface water (0.18 mg/kg day) and

15   groundwater (3.2 mg/kg day) also exceed the LOAEL and the screening benchmark. Hence,

16   there is a potential hazard to all herbivorous wildlife living in areas that may be irrigated with

17   contaminated water. At disposal sites, wildlife would be at risk from the effects of loss of food

18   and habitat from toxic effects on plants, as well as the potential for direct toxic effects via

19   consumption of perchlorate-tainted food or water.

20   Assuming a water ingestion rate of 0.21 g/g-day (U.S. EPA, 1993a,b), the screening

21   benchmark for herbivores is equivalent to a water concentration of 4.8 μg/L. Perchlorate levels

22   reported for large surface waters (as high as 16 μg/L) are greater than this concentration. Much

23   higher perchlorate concentrations have been reported in monitoring wells at rocket motor

24   manufacturing or testing sites (37×10^6 μg/L) and in groundwater-dominated surface water

25   systems close to sites of contamination (3500 to 1.3×10^5 μg/L), and rodent exposures via

26   drinking water at these sites would exceed the rodent NOAEL.

27   Based on screening level benchmarks for herbivores, the analytical detection methods for

28   perchlorate in plant tissues may not be sufficient for the detection of concentrations potentially

29   toxic to herbivores even though the analytical detection methods for perchlorate in water are

30   sufficient. The detection limits achieved for perchlorate in water and in plant tissues were 4 μg/L

31   and 0.4 mg/kg, respectively (Parsons, 2001; Smith et al., 2001).

10.2.3.4 Carnivores

Available evidence indicates that concentrations in terrestrial invertebrates are less than the concentrations in plants and similar to that in soils. As a result, there currently is no indication that terrestrial carnivores are at additional risk from perchlorate. Risks of direct toxic effects are therefore lower for carnivores than herbivores. In locations where perchlorate levels are sufficient to significantly affect herbivores, carnivores are more likely to be affected by loss of prey than by perclorate toxicity. Therefore toxic effects are not quantified.

10.2.4 Uncertainties

This discussion of uncertainties is limited to qualitative uncertainties associated with major gaps in the data available for ecological risk assessment of perchlorate. This is because, as with other screening assessments, quantitative uncertainties are treated through the use of conservative assumptions. It is also because the data gaps are the major sources of uncertainty, not imprecision or inaccuracy of the available data.

10.2.4.1 Uncertainties Concerning Aquatic Risks

Aquatic Exposures. The primary uncertainty associated with this assessment of aquatic risks is the paucity of data on perchlorate occurrence in surface waters. For lack of systematic sampling and analysis, the spatial and temporal distribution of perchlorate in water is unknown. It is not certain whether the reported concentrations in water represent the highest existing levels. This is not a large source of uncertainty for this screening assessment if it is assumed that sampling has been biased to areas of highest likely contamination. However, it would be a major source of uncertainty in any subsequent definitive assessment.

Aquatic Effects. While the effects of perchlorate on some species of algae are known, the effects on aquatic macrophytes are unknown. As a result, risks to aquatic primary producers are estimated using only the chronic toxicity test results for the alga *Selenastrum*. Because of physiological differences between algae and vascular plants, effects on aquatic primary producers are not adequately assessed. In addition, it us unknown how or if physiological variations among various species of algae or plants may affect their susceptibility to perchlorate.

Algae, aquatic macrophytes, and terrestrial leaf litter are the bases of food chains in many aquatic ecosystems. Because perchlorate has been shown to concentrate in leaves of terrestrial

1   plants and aquatic plants, the potential for direct impacts to primary consumers (i.e., planktonic

2   and benthic invertebrate communities) is a concern that could not be addressed in this

3   assessment.

4   A 35-day, early-life stage (ELS) test with *Pimephales*, generally regarded as a chronic test

5   but short of a full-life-cycle test, showed no significant effects on any standard endpoint

6   (survival, growth or biomass) at the highest concentration tested (490 mg/L). However, all

7   larvae exposed to perchlorate concentrations, including the lowest concentration of 28 mg/L,

8   exhibited redness and swelling, which was not observed in the larvae exposed to the control

9   water. This finding suggests the presence of subtle effects that could be ecologically significant

10   and raises doubt about whether a chronic NOEC has been adequately determined for this species.

11   This uncertainty is displayed as a range surrounding the secondary chronic value (i.e., < 0.32 to

12   > 0.83 mg/L). Because of the inequality signs, even the width of the range is uncertain. For this

13   reason, and because of the potential for chronic effects caused by thyroid dysfunction, chronic

14   effects should be investigated in a full life cycle test.

15   The uncertainty factors in the secondary chronic value are high because of the lack of test

16   results for aquatic organisms other than fathead minnows, amphipods, and daphnids.

17

18   **10.2.4.2   Uncertainties Concerning Terrestrial Risks**

19   **Terrestrial Exposure**. The available data concerning aqueous perchlorate levels is sparse

20   and has not been collected systematically. As a result, the spatial and temporal distribution of

21   perchlorate in irrigation water is unknown. It is not clear that the reported concentrations in

22   water represent the highest existing levels. This is not a major source of uncertainty for this

23   screening assessment if it is assumed that sampling has been biased to areas of highest likely

24   contamination. However, it would be a major source of uncertainty in any subsequent definitive

25   assessment.

26   The fate of perchlorate in soil, including its tendency for evaporative concentration, is not

27   well characterized. As a result, soil concentrations were assumed to be equal to irrigation water

28   concentrations. This assumption could be low by multiple orders of magnitude if evaporative

29   concentration occurs with perchlorate, as it does with metals. The limited data for irrigated soils

30   near Yuma (Parsons, 2001) do not support the occurrence of such a high degree of evaporative

1   concentration, but neither are they sufficient to rule out concentration by up to a factor of 10 or

2   so. More information on the fate of perchlorate in irrigated soils is needed.

3   The bioconcentration of perchlorate by plants suggests that perchlorate may be elevated in

4   leaves and leaf litter to levels that may affect invertebrate herbivores and soil invertebrate

5   communities. For lack of data concerning dietary toxicity, risks to invertebrates by this route

6   were not assessed.

7   Available toxicity data for rodents suggest that vertebrate herbivores may be sensitive to

8   low levels of perchlorate in plant tissues; concentrations potentially causing toxicity are

9   calculated to be lower than those currently detectable by chemical analyses of plants. In Parsons

10 (2001), detection limits for plants were generally about 0.4 mg/kg wet weight; similar detection

11 limits were achieved by Ellington and Ellis (2000) and Ellington et al. (2001), as compared to an

12 exposure benchmark of 0.01 mg/kg in plant tissue for a representative herbivore (see Section

13 8.3.2.2). Therefore, lower detection limits for perchlorate in plant tissues may be needed to

14 completely assess the risks to vertebrate herbivores.

15   **Terrestrial Effects**. The toxicity of perchlorate to nonmammalian vertebrate wildlife is

16 unknown. As a result, risks to birds, reptiles, and amphibians could not be assessed.

17   The toxicity of perchlorate to terrestrial invertebrates, other than acute lethality to

18 earthworms, is unknown. As a result, risks to other terrestrial invertebrates were inadequately

19 assessed.

20

21 ## 10.2.5 Research Needs

22   Three questions were asked of the screening ecological risk assessment for perchlorate:

23 • Are ecological risks best characterized as *de minimis* (exposures clearly are below levels of

24   concern), *de manifestis* (risks are clearly significant and require management action to reduce

25   exposures); or somewhere in between and requiring further characterization?

26 • Are analytical detection methods for determining levels of perchlorate in the environment

27   sufficient, or is there a likelihood of adverse ecological effects occurring at levels below current

28   detection limits?

29 • Is the available ecotoxicological information on perchlorate sufficient, or are additional studies

30   needed?

1   In the immediate vicinity of facilities that were involved in the manufacture, use, or

2   disposal of perchlorate salts, particularly facilities involved in handling of solid rocket

3   propellents, ecological exposure can exceed levels of concern and management actions may be

4   needed to reduce these exposures. Site-specific risk assessments would be needed to guide

5   remediation of such locally contaminated sites. Farther from such facilities, ecological exposures

6   appear to be below levels of concern.

7   The analytical detection methods for perchlorate are generally sufficient, and there appears

8   to be no indication of adverse ecological effects occurring at levels below detection limits, except

9   that detection limits in plant tissues are not low enough to ensure that risks to herbivores are

10   detected. Additionally, there is some uncertainty about the potential for bioaccumulation at low

11   concentrations of perchlorate in surface water, because of differences in the analytical detection

12   limits between water and animal tissues.

13   The available ecotoxicological information on perchlorate is sufficient for this screening-

14   level ecological risk assessment. However, additional ecotoxicological studies could reduce the

15   uncertainties about the toxicity of perchlorate to other potential ecological receptors.

16   While the available information may yield an adequate screening level ecological risk

17   assessment, the following research needs for exposure and effects analysis deserve mention.

18

19   **10.2.5.1   Exposure**

20   Concerning exposure, at least three important issues remain unresolved:

21   • Because the available data on accumulation in terrestrial and aquatic vascular plants are from

22   studies that were not designed to quantify accumulation factors, the accumulation of

23   perchlorate in terrestrial and aquatic plants should be further investigated.

24   • Because of the potential for evaporative concentration, the fate of perchlorate in irrigated soils

25   should be investigated.

26   • Because the concentrations that have potential for dietary toxicity to vertebrate herbivores are

27   less than the limits of detection currently achievable by chemical analysis of plants, analytical

28   methods for plant tissues that could lower the limits of detection should be investigated.

29

30

10.2.5.2 Effects

Also requiring further attention are issues related to the effects of potential perchlorate exposure:

• The effects of exposure of aquatic plants should be determined.

• The effects of exposure of noncrustacean invertebrates should be determined.

• The effects of dietary exposure to perchlorate should be determined in birds and in herbivorous or litter-feeding invertebrates.

• The effects of dietary and cutaneous exposure to perchlorate should be determined for adult amphibians and aquatic reptiles.

• If perchlorate occurs at significant levels in estuarine systems, its toxicity in saline waters should be determined.

10.2.5.3 Site-Specific Investigations

Some of the research needs that were listed in the previous ERD of this document have been met by the research conducted by the US Air Force IERA (Parsons, 2001) in which perchlorate concentrations in environmental media (i.e., surface soils, surface water, sediments, and pore water) and biological tissues (i.e., terrestrial plants, invertebrates, reptiles, birds, and mammals and aquatic vegetation, invertebrates, fish, amphibians, reptiles, and birds) were surveyed at six sites with known perchlorate contamination. These data are supplemented by additional sampling at one of the sites, Longhorn Army Ammunition Plant in Texas, by Smith et al. (2001). These studies do address some questions about exposure that were expressed in the previous ERD of this document (U.S. EPA, 1998d), i.e:

   • Because concentrations of perchlorate in water are poorly known, and concentrations in soil and biota are unknown, a survey of perchlorate contamination should be conducted.

   • Because, contrary to expectations, perchlorate accumulates to high concentrations in terrestrial vascular plants, the accumulation of perchlorate in aquatic plants and in animals should be investigated.

However, these studies were screening-level surveys that took small numbers of samples during limited periods of time. In addition, the studies were not designed to address questions about the effects of exposure. In some locations, concentrations in environmental media were high enough

1   that toxicity to ecological receptors was highly likely (i.e., the risks were *de manifestis*), and in

2   other locations toxicity could not be ruled out (i.e., the risks could not be termed *de minimus*).

3   Therefore, systematic sampling is needed in these locations to more definitively quantify

4   exposures and effects, so that the likelihood, nature and extent of ecological risks may be

5   quantified, appropriate remedial alternatives may be designed, and effectiveness of site cleanup

6   may be judged. In addition, site surveys may be required in other locations where perchlorate

7   contamination is suspected.

8

9

10.3 CHARACTERIZATION PROGRESS SUMMARY

11   Despite the fact that the appreciation of widespread perchlorate contamination emerged

12   only five years ago, considerable progress has been made in hazard identification and

13   quantitative dose-response characterization for both the human health and ecotoxicological risks

14   of potential perchlorate exposures. The thyroid has been confirmed as the target tissue in

15   humans, laboratory animals, and wildlife. The key event of the mode of action for perchlorate is

16   iodide uptake inhibition at the NIS with the potential for both subsequent neurodevelopmental

17   and neoplastic sequelae. A harmonized human health reference dose has been proposed to be

18   protective for both sequelae based on a mode of action model. Data insufficiencies for various

19   ecotoxicological receptors and for accurate exposure estimates precludes other than a screening-

20   level assessment at this time. Additional research is needed to determine the contribution of

21   exposure sources other than drinking water. This requires more progress in the area of analytical

22   methods to extend current approaches to other media.

23   As with any risk assessment, additional insights and new research will continue to change

24   our understanding as the knowledge base is informed with new data and as the scientific and

25   technical areas relevant to the particular risk characterization mature and evolve. Work

26   dedicated to the areas defined in this chapter should allow continued improvement of the risk

27   characterizations for perchlorate in the future.

28

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APPENDIX A

Schematics of Study Designs for Neurodevelopmental, Two-Generation Reproductive and Developmental Studies

Dosage Period
DG 0 to DG 31 or DL 10

Acclimation 14 Days	Cohabitation Maximum 7 Days	

F0 Female Rats
Arrive at
Testing Facility

DG 0
Dosage
Begins

DG 21:
First
Possible
Delivery

DG 25:
Last
Possible
Delivery

DG 31 or DL 10:
End of Dosage

DL 10:
Scheduled
Sacrifice
(thyroid
evaluations)

DG 31:
Scheduled
Sacrifice
(rats that did
not deliver)

DL 22:
Scheduled
Sacrifice
(weaning)

PARTURITION

DP 5: Litters Culled to 8 Pups and
Blood Collection

DP 10: 5 Dams and Litters Selected
for Blood Collection

F1 GENERATION
(daily clinical observations and sexual
maturation, weekly body weights, feed
consumption, and blind observations)

SUBSET 1
100 Male and Female Pups
(one male/female
pup/litter/dosage group)

SUBSET 2
100 Male and Female Pups
(one male/female
pup/litter/dosage group)

SUBSET 3
100 Male and 100 Female Pups
(one male/female
pup/litter/dosage group)

SUBSET 4
100 Male and 100 Female Pups
(one male/female pup/litter/dosage
group)

DP 12: SCHEDULED SACRIFICE,
BRAIN WEIGHTS, AND
NEUROHISTOLOGICAL
EXAMINATION
6 Male and 6 Female Pups/Dosage Level
(total of 30 male and 30 female pups)

DPs 23-25 AND 30-32: PASSIVE
AVOIDANCE TESTING

DPs 14, 18, 22, AND 59: MOTOR
ACTIVITY

DPs 80-86: SCHEDULED
SACRIFICE,
NEUROHISTOLOGICAL
EXAMINATION,
REGIONAL BRAIN WEIGHTS
6 Male and 6 Female/Dosage Group
(30 male and 30 female total)

DPs 59-63 AND 66-70:
WATER MAZE TESTING

DPs 23 AND 60: AUDITORY
STARTLE HABITUATION

DPs 67-69: SCHEDULED
SACRIFICE

DPs 90-92: SCHEDULED SACRIFICE
AND BLOOD COLLECTION

Abbreviations:

DG = Day of (Presumed) Gestation
DP = Day Postpartum
DL = Day of Lactation

Figure A-1. **Schematic of the neurobehavioral developmental study of ammonium perchlorate administered orally in drinking water to SD rats (Argus Research Laboratories, Inc., 1998a).**

First Day
of Exposure

P1 Male Rats	Premating	Cohabitation	a
	10 weeks	3 weeks	

Last Day
of Exposure [b]

Natural
Delivery

P1 Female Rats	Presumed Gestation	Lactation
10 weeks	3 weeks	3 weeks

F1 Generation

Preweaning
3 weeks

Day 21
Postpartum
Weaning [c]

Last Day
of Exposure [F1] [c]

F1 Generation Rats	Postweaning	Cohabitation	a	Lactation
	10 weeks	3 weeks		3 weeks

F2 Generation

Postpartum
3 weeks

Day 21 [c]

■■■■ = Dosage Period

a = Male rats sacrificed after determination of sufficient number
    of pregnancies
b = P1 generation female rats sacrificed
c = F1 generation dams and F2 generation litters sacrificed

Figure A-2. Schematic of the oral (drinking water), two-generation (one litter per generation) reproduction study of ammonium perchlorate in SD rats (Argus Research Laboratories, Inc., 1998b).

a = Blood samples taken from does for thyroid and pituitary hormone (T3, T4, TSH) analyses.
b = Fetal evaluations (external examinations and soft tissue and skeletal examinations).

Figure A-3. Schematic of the oral (drinking water) developmental toxicity study of ammonium perchlorate in New Zealand rabbits (Argus Research Laboratories, Inc., 1998c).

List of Acronyms and Abbreviations

Acronym	Definition
ΔA°_{rxn}	Helmholtz free energy of reaction
ΔG°_{f}	Gibbs free energy of formation
ΔG°_{rxn}	Gibbs free energy of reaction
ΔS°_{univ}	net entropy of the universe
a-p	anterior-posterior
Ab	antibody
ACSL	advanced continuous simulation language
ADHD	attention deficit hyperactivity disorder
ADME	absorption, distribution, metabolism, and elimination
AFB	air force base
AFRL	U.S. Air Force Research Laboratories
AFRL/HEST	Air Force Research Laboratory/Human Effectiveness Directorate
AIDS	acquired immunodeficiency syndrome
AITD	autoimmune thyroid disease
ANCOVA	analysis of covariance
AP	ammonium perchlorate
ATP	adenosine triphosphate
AUC	area-under-the-curve
AV	acute value
AWQC	ambient water quality criteria
BF_4^-	tetrafluoroborate
BMD	benchmark dose
BMDL	benchmark dose lower confidence limit
BMR	benchmark response

Acronym	Definition
BW	body weight
C'	complement
CA DHS	California Department of Health Services
cAMP	cyclic adenosine monophosphate
CBC	complete blood count
CCL	Contaminant Candidate List
CD4/CD8	cluster of differentiation — cellular markers 4 and 8
CDC	Centers for Disease Control and Prevention
CERCLA	Comprehensive Environmental Response Compensation Liability Act
cESI-MS	complexation electrospray ionization mass spectrometry
CFU	colony-forming units
CHS	contact hypersensitivity
ChV	chronic value
Cl_2	chlorine
CI	confidence interval
ClO^-	hypochlorite
ClO_4^-	perchlorate
ClUC-p	perchlorate urinary clearance
CNS	central nervous system
CP	cyclophosphamide
CPES	Coastal Plain Experiment Station
CPM	counts per minute
Cs^+	cesium
CsCl	cesium chloride
CTL	cytotoxic T-lymphocyte
CV	coefficient of variation
DAF	dosimetric adjustment factor

Acronym	Definition
DEQ	Department of Environmental Quality
DIT	diiodotyrosine
DNA	deoxyribonucleic acid
DNCB	dinitrochlorobenzene
DoD	Department of Defense
DoE	Department of Energy
DTH	delayed-type hypersensitivity
DWEL	drinking water equivalent level
E:T	effector to target cell
EAR	estimated average requirement
EGF	epidermal growth factor
ELISA	enzyme linked immunosorbant assay
ELS	early-life stage
EPA	U.S. Environmental Protection Agency
EPL	Experimental Pathology Laboratories, Inc.
ER	endoplasmic reticulum
$E°$	standard electric potential
F	Faraday constant
F1	first generation
F2	second generation
FAVF	Final acute value factor
FCN	function
FETAX	Frog Embryo Teratogenesis Assay: *Xenopus*
FGF	fibroblast growth factor
FH	follicular epithelial cell hypertrophy or hyperplasia
FIFRA	Federal Insecticide, Fungicide, and Rodenticide Act
fT4	free thyroxine

Acronym	Definition
GA	Golgi apparatus
GD	gestation day
GGTP	g-glutamyl transpeptidase
GI	gastrointestinal
GMAV	Genus mean acute value
gsp	GTP-binding protein mutation
Gy	Gray (equal to 100 rads)
H^+	hydrogen
H_2O_2	hydrogen peroxide
hCG	human chorionic gonadotropin
$HClO_4$	perchloric acid
HEE	human equivalent exposure
HOCl	hypochlorous
I^-	iodide
IC	ion chromatography
IC_{25}	quartile inhibitory concentration
ICD-9	International Classification of Diseases, 9th Revision
ID	iodine deficiency
IFN	interferon
IGF-1	insulin-like growth factor
IgG	immunoglobulin G
IgM	immunoglobulin M
ip	intraperitoneally
IPSC	Interagency Perchlorate Steering Committee
IRIS	Integrated Risk Information System
IU	international unit
IUDR	uridine

Acronym	Definition
iv	intravenously
K^+	potassium
K_2O	potassium oxide
Km	Michaelis-Menten affinity constant
KNO_3	potassium nitrate
LC_{50}	concentration lethal to 50% of population
LD	lactation day
LHAAP	Longhorn Army Ammunition Plant
Li^+	lithium
LLNA	local lymph node assay
ln	natural log
LOAEL	lowest-observed-adverse-effect level
LOEC	lowest-observed-effect concentration
LOEL	lowest-observed effect level
LP	lymphoproliferation
LS	Lumen size
LY	lysosomes
M-W RST	Mann-Whitney Rank Sum Test
MCA	3-methyl cholanthrene
MCL	maximum contaminant level
MDL	minimum detection limit
MF	modifying factor
$Mg(ClO_4)_2$	magnesium perchlorate
MIT	monoiodotyrosine
MMIA	1-methyl-2-mercaptoimidazole
MANOVA	multiple analysis of variance
MCLG	maximum contaminant level goal

Acronym	Definition
MRL	minimum reporting limit
mRNA	messenger ribonucleic acid
MS-MS	mass spec — mass spec
MTD	maximum tolerated dose
n	number of electrons or number of moles
n.d.	no date
N-P-K ratio	nitrogen-phosphorous-potassium ratio
Na^+	sodium
$NaClO_4$	sodium perchlorate
$NaNO_3$	sodium nitrate
NAS	National Academy of Sciences
NASA	National Aeronautics and Space Administration
NCE	Normochromatic erythrocyte
NCEA	National Center for Environmental Assessment
NDEP	Nevada Division of Environmental Protection
NERL-ERD	Natural Exposure Research Laboratory's Ecosystems Research Division
NH_4^+	ammonium
NH_4ClO_4	ammonium perchlorate
NH_4NO_3	ammonium nitrate
NHEERL	National Health and Environmental Effects Research Laboratory
NIEHS	National Institute for Environmental Health Sciences
NIS	sodium iodide symporter
NK	natural killer
NMR	nuclear magnetic resinance
NO_3^-	nitrate
NOAEL	No-Observed-Adverse-Effect Level
NOEC	No-Observed-Effect Concentration

Acronym	Definition
NPDWR	National Primary Drinking Water Regulations
NRMRL	National Risk Management Research Laboratory
NTP	National Toxicology Program
O_2	oxygen
OEHHA	Office of Environmental Health Hazard Assessment
OEPP	Office of Emergency Response and Remediation
OPPTS	Office of Prevention, Pesticides, and Toxic Substances
ORD	Office of Research and Development
OSWER	Office of Solid Waste and Emergency Response
OW	Office of Water
p	probability
P	pressure
P1	parental generation
P_2O_5	disphosphorus pentoxide
p53	*p53* tumor suppressor gene
PA	prealbumin
PAS	periodic acid shift
PBI	protein-bound iodide
PBPK	physiologically based pharmacokinetic
PCE	polychromatic erythrocyte
PCB	polychlorinated biphenyl
PFC	plaque-forming cell
PHG	public health goal
PII	plasma inorganic iodide
PND	post-natal day
PP	post partum
PP-TH	plasma protein-thyroid hormone

Acronym	Definition
ppb	parts per billion
ppm	parts per million
PQL	practical quantitation limit
PSG	Perchlorate Study Group — consortium of defense contractors
PT-p	thyroid follicle:stroma partition coefficient
PTU	propylthiouracil
PWG	Pathology Work Group
QA/QC	quality assurance/quality control
R	ideal gas constant
RAIU	radioactive iodine uptake
ras	*ras* protooncogene
Rb^+	rubidium
RDA	recommended dietary allowance
RfC	inhalation reference concentration
RfD	oral reference dose
RIA	radioimmunoassay
RL	reproducibility limits
RO	reverse osmosis
r_s	Spearman's rank order
RSC	relative source contribution
rT_3	reverse triiodothyronine
SACR	secondary acute-chronic ratio
SAV	secondary acute value
sc	subcutaneously
SCN	thiocyanate
SCV	secondary chronic value
SD	standard deviation

Acronym	Definition
SD rats	Spraque-Dawley strain
SDWA	Safe Drinking Water Act
SE	standard error of the mean
SGOT	serum glutamyl oxacetic transsaminase
SGPT	serum glutamyl pyruvic transaminase
SLA	soluble *Listeria* antigen
SMCV	species mean chronic value
SNK	Student Newman Keuls
SRBC	sheep red blood cell
SRLB	Sanitation and Radiation Laboratory Branch
T	temperature
T2	diiodothyronine
T3	triiodothyronine
T4	thyroxine or tetraiodothyronine
T4 GLUC	T4-glucuronide conjugate
TBG	thyroid-binding globulin
TCE	trichloroethylene
TDS	total dissolved solids
Tg	thyroglobulin
TH	thyroid hormone
TPO	thyroid peroxidase
TRH	thyrotropin-releasing hormone
TSCA	Toxic Substances Control Act
TSH	thyroid-stimulating hormone
tT4	total thyroxine
UCMR	Unregulated Contaminant Monitoring Rule
UDPGTs	uridine diphosphyl glucuronosyl transferases

Acronym	Definition
UF	uncertainty factor
USAF	United States Air Force
USGS	United States Geological Survey
USN	United States Navy
V	volume
Vmaxc	Michaelis-Menten maximum velocity capacity
W_{exp}	expansion work
WHO	World Health Organization
WPAFB	Wright Patterson Air Force Base
WSWRD	Water Supply and Water Resources Division

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Perchlorate: Not Only Rocket Science

Footprints	**Moral Limits**	**Upsetting Offsets**
Five Steps to Lower Toxics	*State Versus Feds On Clean Air Policy*	*Just a Way for Rich To Avoid Change?*

It's Not Only Rocket Science

The perchlorate story follows the arc of other major controversial chemical management challenges, such as dioxin, where the initial focus on end-of-pipe controls missed key sources in the environment

By Andrew Rak

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The ongoing debate over the potential health effects from exposure to low levels of perchlorate is overshadowing an equally important dialogue on the sources of contamination. Perchlorate first became a headline concern when it was discovered that it entered the environment from rocket fuel and munitions at Department of Defense sites — seemingly just another legacy of the Cold War — but it can also be found in fertilizer, bleach, fireworks, road flares, and blasting compounds. It can be present as an ingredient or impurity in lubricating oils, matches, and automotive airbag deployment initiators. It is associated with aluminum, rubber, dye, and paint manufacturing, leather tanning, and pulp processing. Solutions used in water and wastewater treatment plants for disinfection have also been identified as a potential problem.

While some research has addressed the relative importance of multiple sources, the perception that military activities are the overwhelming contributor to the environment may be hindering appropriate and effective regulation. While perfect knowledge about the sources of perchlorate is unobtainable, continuing to focus solely on a few point sources will not adequately spur effective public health regulation.

Extensive toxicological studies have been undertaken related to the potential health risks from exposure to perchlorate, including a National Research Council review of the Environmental Protection Agency's toxicity assessment. The chemical may pose a health risk if taken up by the thyroid, where it disrupts the production of hormones and affects metabolism and neurodevelopment. But toxicity is only part of a risk-based approach to public health. As the NRC famously declared in 1983 in the Red Book — more properly "Risk Assessment in the Federal Government: Managing the Process" — only by combining toxicity data with information about the sources of exposure to a contaminant can a thorough risk-based approach be constructed. In other words, the same scientific rigor and expertise applied to the investigation of a chemical's toxicity should be applied to the identification of sources that lead to significant exposures.

A thorough examination of the sources of exposure to perchlorate has not been completed, and this lack of knowledge hinders attempts by regulatory agencies to provide an effective risk-based approach for protecting public health. Through an

understanding of sources in the environment and releases to the environment, regulatory agencies can take appropriate steps to directly reduce exposure and control public health risk.

While defense activities have often been assumed to be the dominant fraction of perchlorate's environmental presence, military contributions alone cannot be the explanation for the geographically dispersed detection of the compound. Nor can military sources be responsible for the perchlorate detected in a wide variety of food stuffs, including dairy items and produce. Sampling of dairy products from Maryland and organic lettuce from Wisconsin have found trace amounts of perchlorate; however, any direct connection to military point sources at these locations is lacking, thereby suggesting the importance of other contributors.

Early impressions about perchlorate sources helped federal and state regulators develop preliminary plans for addressing contamination. However, more complete evaluations may increase the focus on newly identified sources. In a 2005 letter to the federal Environmental Protection Agency, state regulatory officials wrote that "none of the nine water supplies that have tested positive for perchlorate in Massachusetts appear to have any connection to military bases or activities." The use of perchlorate-containing explosives in construction activities, and even in bleach, were among the new sources identified by the state. Earlier this year, Maine Drilling and Blasting agreed to contribute $1 million to the city of Westford, Massachusetts, for remediating perchlorate associated with construction explosives. Grappling with the risk from other sources is just a matter of time.

The advent of new scientific fingerprinting methods that distinguish natural from man-made sources will allow regulatory managers to tackle this multi-faceted source challenge with targeted risk reduction measures. As new sources are identified and quantified, lingering misperceptions about the

To holistically address the risks a more comprehensive assessment of the sources contributing to exposures is necessary

role of point and non-point sources of perchlorate should be displaced.

The perchlorate story follows the arc of other major controversial chemical management challenges, such as dioxin, where the initial focus on end-of-pipe controls from incinerators and chlorine, and pesticide production proved to miss key sources in the environment. During the multi-decade interagency and public debate over the appropriate health risk estimates for dioxin, EPA issued a pivotal emissions report. In 2000, the emissions inventory analysis for dioxin found that backyard burning of plastics and other waste in rural areas was a major remaining non-point source of deposition to soil. This surprising finding shifted the regulatory debate and highlighted areas where the traditional focus on point sources and end-of-pipe enforcement approaches could not be applied. Any new reduction strategy for dioxin emissions would likely have to include multiple non-point sources in order to be effective. The same dynamic appears to be developing for perchlorate.

While early data helped regulatory agencies focus on perchlorate as an emerging problem, to holistically address its risks a more comprehensive assessment of the sources contributing to environmental exposures is necessary. With improved sampling technology and the additional data that are being generated relating to dietary and other exposure routes, food safety regulators, environmental agencies, and local governments will be able to more effectively regulate the use, disposal, and cleanup of perchlorate, and reduce public exposure. Information on unusual suspects will increase the government's grasp of the problem and enhance risk mitigation as our understanding advances.

At the federal level, EPA has elected to address

exposure to perchlorate in groundwater and drinking water using authorities under Superfund and the Safe Drinking Water Act. Although the agency has yet to make a final regulatory determination for perchlorate in drinking water, some state regulatory agencies have already promulgated regulations, with California and Massachusetts setting enforceable limits of 6 parts per billion and 2 parts per billion, respectively. However, it is not clear if these regulations will result in a meaningful public health benefit because we do not know what role drinking water plays in overall exposure to perchlorate. Addressing the need for appropriate regulation requires a more complete understanding of the sources (and relative amounts) of synthetic and natural perchlorate in the environment.

While EPA has elected to address perchlorate in groundwater and drinking water, other regulatory agencies and researchers have paid attention to potentially more widespread exposures in the diet. Ongoing food monitoring surveys have found perchlorate in mothers' milk and cows' milk, as well as in infant-formula. Perchlorate has also been found in lettuce, spinach, cantaloupe, cabbages, and beets. The Food and Drug Administration has completed studies of the occurrence of perchlorate in food items, but has yet to issue any health advisories. Likewise the Department of Agriculture conducts monitoring through its National Residue Program. But an analysis of the relative sources of perchlorate and how these sources may influence entry into the food chain has not been conducted. In the future, the NRP may generate key opportunities for risk reductions should trends related to perchlorate in food warrant mitigation.

A continued focus on military sources will not provide a complete analysis of the sources of perchlorate. The defense establishment uses perchlorate in numerous items as an oxidizer. While the military continues to purchase large amounts of perchlorate for solid rocket motors and other items (6.2–8.1 million pounds per year from 2004–2006), solid perchlorate inside a rocket motor casing is unavailable for release to the environment. Industrial recycling of rocket motors, which previously released wash water into the environment, is now a closed loop system. Annual military training uses of perchlorate in detonators, initiators, and ground burst simulators, which are confined to certain ranges, is approximately 1.6 million pounds, and substitutes are now being employed for some of these uses. As a result of remediation activities, substitution efforts, best management practices, and compliance with Clean Water Act permits, defense-related releases of perchlorate should be diminishing.

In California, one of the states thought to be most contaminated with perchlorate, the military collaborated with state officials to investigate reports of widespread contamination. In 2003, the Defense Department and state regulators established a partnership to investigate the presence of perchlorate throughout California to locate previously unidentified threats to public water supplies. More than 900 military sites were screened using a consensus-based protocol; 870 of these sites were deemed to be not of concern. State regulators agreed that military installations and formerly used defense sites did not appear to be significantly impacting California public drinking water wells. As a result of the collaboration, California also adopted many of the department's best management practices for perchlorate.

The department began an extensive perchlorate monitoring program and, by 2008, had collected more than 47,500 samples at 309 locations nationwide to define the scope of its perchlorate contamination problem. The results of this monitoring program — posted and annually updated on the Defense Environmental Network Information Exchange website — show that the vast majority of perchlorate samples from military locations were below 4 parts per billion, and large groundwater plumes of perchlorate exist at only a limited number or military sites.

While the military appears to be defining and controlling many of its potential sources of perchlorate, it is unclear what controls are being put in place elsewhere. Recent research announced in the Fourth National Report on Human Exposure to Environmental Chemicals by the Centers for Disease Control and Prevention together with a number of academic findings suggest that exposure to perchlorate is widespread, indicating there may be many undocumented sources. These sources should be thoroughly evaluated in order to better understand total human exposure.

The contribution of perchlorate from the use of sodium hypochlorite for various household, drinking water, and commercial disinfection purposes may be an important non-point source contributor to total perchlorate exposure. Sodium hypochlorite, a bleach, generates perchlorate in storage and when it is exposed to sunlight. The compound is used in a variety of household applications (household bleach is a 3–6 percent solution) and industrial applications (water and waste water disinfection solutions are 12–15 percent solutions) related to disin-

fection. Nearly $3 billion of sodium hypochlorite bleach is sold globally each year. In a 2008 study, perchlorate contamination was found to occur in more than 90 percent of sodium hypochlorite samples.

The most recent part of the perchlorate source picture to come to light is the volume of the compound entering the United States in fireworks. Recent data from the Department of Commerce indicate a large increase in firework imports over a five-year span, from 174.7 million pounds in 2002 to 271.2 million pounds in 2006. Potassium perchlorate constitutes up to 70 percent of the chemical fraction in fireworks. Based on these data, the American Pyrotechnics Association estimates that 14.2 million pounds of perchlorate entered the country in fireworks in 2002, and 21.8 million pounds in 2006. Imports represent about 90 percent of fireworks used in the United States. For 2006, the amount of perchlorate imported in fireworks represents nearly three times the amount purchased by the military. While importation does not necessarily equate with amounts released into the environment, these data highlight an important potential non-point source.

Recent field studies also support focusing on perchlorate discharges from fireworks. EPA analyzed water in an Oklahoma lake before and after fireworks displays. Testing performed 14 hours after an event showed that perchlorate levels rose 24 to 1,028 times above the pre-display baseline. The agency found that concentrations of perchlorate peak about 24 hours after a display and then decrease to the baseline within 20 to 80 days. Thus, Fourth of July activities and sporting events may be important in future efforts to define sources and routes of exposure. Efforts to restrict and manage releases at the local level may be able to control exposure.

The millions of emergency road

Perchlorate Will Accumulate

In 2008 NASA's Phoenix Lander found perchlorate in three different Martian soil samples. That discovery prompted a re-analysis of the soil samples we had brought back from an expedition a year earlier to Antarctica's Dry Valleys, a site used as a Mars analogue for training because of the subzero temperatures and extreme aridity. For both Earth and Mars, the evidence implies that given the right conditions and a source of chlorine, perchlorate will accumulate.

Free of anthropogenic influences, the Dry Valleys provide an ideal location for such studies. The Antarctic soil samples had been collected from pits dug in three types of Dry Valley microclimate zones: coastal (wet), inland (arid), and highland (hyperarid). Samples were collected from every identifiable soil horizon (boundaries between different kinds of soil) down to the ice-cemented soil. All the samples were reanalyzed for perchlorate.

Samuel Kounaves

To our amazement, perchlorate was found in all the highland soils, in all the horizons from the surface to the ice-cemented soil. Its concentration ranged up to 630 parts per billion and in a continuous vertical profile. In contrast, for the inland valley soil horizons, it was found to vary with a more heterogeneous distribution, while in the coastal valleys it appeared very randomly distributed, approaching 1,100 ppb in one isolated soil horizon, and totally absent in others, with no regular or discernible pattern.

How the perchlorate is deposited in these valley soils is clearly evident from its correlation to nitrate and chloride, both of which have been shown to be atmospherically formed and deposited. Thus, the best explanation for its changing distribution in the soil when moving from the upper to lower valleys is the difference in the amount of liquid water present and its effect, leading to depletion or concentration of the perchlorate. In the highlands the atmospheric deposition of perchlorate is left undisturbed, while in the lower wet valleys its distribution becomes chaotic.

Combining our findings with those for the Arctic, North America, and other regions provides an emerging picture for the global presence of natural perchlorate. In addition, our results from Antarctica clearly point to the conclusion that even though natural perchlorate is atmospherically and homogeneously deposited, over time it will accumulate at high levels at some locations while it will be nonexistent at others.

Our results support the hypothesis that perchlorate must also have a variety of long-term, widely, and irregularly distributed sinks. Since aqueous perchlorate is chemically stable in the natural environment, its lack of accumulation in the ocean or aquifers may also be attributable to microbial utilization in anaerobic or low nitrate media.

To help us understand the full impact of perchlorate, we need to more accurately determine its global distribution and accumulation patterns, its interactions with terrestrial ecology, and its atmospheric formation mechanisms.

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flares sold annually contain significant concentrations of perchlorate, which is often released into the environment. A study in Santa Clara, California, found that 88,184 pounds of road flares are used in the county annually with a corresponding perchlorate content of 5,732 pounds. Field testing has shown maximum concentrations of perchlorate in runoff leaving highways of 314,000 parts per billion after road flare use. Considering the volume nationwide, discarded or partially used emergency flares may be another important non-point source of exposure. Communities could take steps to consider the use of alternative safety-lighting systems such as those adopted by Seattle and other local police organizations to reduce the contribution from flares.

Natural sources of perchlorate from geological materials, such as potash ore, playa crust, and hanksite, also contribute to environmental perchlorate. Natural deposits have been found in the Texas panhandle and north-central New Mexico. Other research suggests that perchlorate may be produced via atmospheric processes, then enters the food chain through rainfall. Atmospheric formation may explain the discovery of perchlorate in soil and ice from the Antarctic [See ANOTHER VIEW, page 41] where concentrations reach up to 1100 micrograms per kilogram. Finally, there is a possible connection between tropospheric ozone and the formation of perchlorate in plants, a factor that could in part explain the presence of perchlorate in agricultural products.

One source of perchlorate in the environment that is receiving some additional attention is the over 100 million pounds of Chilean nitrate fertilizer — which contains perchlorate at levels of 100,000 parts per billion —that have been applied in the United States. There are over 400,000 pounds still being applied annually to commercial agricultural land and homeowner gardens. In 2006, Texas Tech University scientists reported in the journal *Environmental Science & Technology* that perchlorate-containing Chilean nitrate fertilizer likely accounts for more low-level contamination in the United States than all military and industrial sources put together.

Perhaps the most important development driving the understanding of sources of perchlorate in the environment is new forensic fingerprinting technology that can help differentiate man-made and natural types of perchlorate. Isotopic methods are powerful tools when applied to the intractable problems of source attribution for groundwater contaminants. Elements in compounds can have widely different isotopic ratios based on their mode of formation. Stable ratios have a fingerprint, allowing scientists to distinguish sources from one another. The isotopic method for perchlorate is available for use through some university and Department of Energy laboratories. Scientific validation of the new method is underway.

The new clarity made available through forensic inquiries into sources may provide a much better understanding about the proportion of contributions of perchlorate in the environment from commercial, consumer, agricultural, and military sources. Forensic methods distinguished synthetic from natural sources of perchlorate at contaminated sites in southern California and elsewhere, and proved helpful in disentangling possible sources. A 2009 study by the U.S. Geological Survey used forensic isotopic methods to identify historic use of fertilizer as the most likely cause of groundwater contamination in areas of Long Island and concludes that these findings may have national implications.

The growing concern over the possible health effects of exposure to low levels of perchlorate should not overshadow the investigation of perchlorate sources. There appears to be a growing body of evidence that perchlorate sources should be reappraised, particularly in light of findings that perchlorate in fireworks, bleach, safety flares, and Chilean fertilizer may outweigh military-industrial uses. The new data on sources in the environment and releases to the environment should be used to better inform regulatory decisions on controlling exposure. More importantly, the application of fingerprinting methods that distinguish natural from man-made sources will empower regulatory managers to tackle this multi-faceted source challenge with tailored risk reduction measures. As consumer, natural, military, and agricultural sources continue to be identified and quantified, misperceptions about the role of point and non-point sources should be displaced.

While early data allowed regulatory agencies to focus initially on perchlorate as an emerging problem, to holistically address the risks from exposure a more comprehensive assessment of sources is necessary. More information about the unusual suspects among perchlorate sources will enhance risk mitigation and protect public health. With improved sampling technology, food safety regulators, environmental agencies, and local governments will be able to control the use, disposal, and cleanup of perchlorate to reduce public exposure. •

Particulate Oxidative Burden Associated with Firework Activity

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Firework events are capable of inducing particulate matter (PM) episodes that lead to exceedances of regulatory limit values. As short-term peaks in ambient PM concentration have been associated with negative impacts on respiratory and cardiovascular health, we performed a detailed study of the consequences of firework events in London on ambient air quality and PM composition. These changes were further related to the oxidative activity of daily PM samples by assessing their capacity to drive the oxidation of physiologically important lung antioxidants including ascorbate, glutathione and urate (oxidative potential, OP). Twenty-four hour ambient PM samples were collected at the Marylebone Road sampling site in Central London over a three week period, including two major festivals celebrated with pyrotechnic events: Guy Fawkes Night and Diwali. Pyrotechnic combustion events were characterized by increased gas phase pollutants levels (NO_x and SO_2), elevated PM mass concentrations, and trace metal concentrations (specifically Sr, Mg, K, Ba, and Pb). Relationships between NO_x, benzene, and PM_{10} were used to apportion firework and traffic source fractions. A positive significant relationship was found between PM oxidative burden and individual trace metals associated with each of these apportioned source fractions. The level of exposure to each source fraction was significantly associated with the total OP. The firework contribution to PM total OP, on a unit mass basis, was greater than that associated with traffic sources: a 1 μg elevation in firework and traffic PM fraction concentration was associated with a 6.5 ± 1.5 OP^T μg^{-1} and 5.2 ± 1.4 OP^T μg^{-1} increase, respectively. In the case of glutathione depletion, firework particulate OP (3.5 ± 0.8 OP^{GSH} μg^{-1}) considerably exceeded that due to traffic particles (2.2 ± 0.8 OP^{GSH} μg^{-1}). Therefore, in light of the elevated PM concentrations caused by firework activity and the increased oxidative activity of this PM source, there is value in examining if firework derived PM is related to acute respiratory outcomes.

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Introduction

Anthropogenic emissions from firework displays are responsible for particulate matter (PM) episodes with the greatest concentrations measured in the United Kingdom (*1*). Nightly celebrations occur annually over a two to three week period to celebrate Diwali and Guy Fawkes Night during October and November. Small size pyrotechnic events (including hand-held sparklers, cherry bombs, roman candles representing British category 2 and US class 1.4G) are set off by individuals in parks and back gardens. Larger professional displays (British category 4 and US class 1.3G) also occur where pyrotechnics are projected to attain explosion heights of up to 200 m. Regardless of the type of firework ignited, the combustion products are similar: high SO_2 and NO gaseous releases (*3, 2*), dense metal-rich PM plumes with a large organic carbon component (*4*).

The firework physicochemical signature has been extensively characterized internationally: Washington State, US (*5*); California, US (*6*); Texas, US (*7*); Montreal, Canada (*8*), Hisar, Hyderabad, and Thumba, India (*10, 9, 2*); Beijing, China (*11*); Saalbach, Austria (*12*); United Kingdom (*1*); Mainz, Germany (*4*); Milan, Italy (*13*); and Valencia, Spain (*3*). These previous reports unanimously agree firework activity contributes to elevated ambient particulate metal concentrations in respirable size fraction (i.e., PM with an aerodynamic diameter of <10 μm, PM_{10}). Fireworks and related explosives account for substantial airshed emissions in the UK: 3 tonnes Cu, 10 tonnes Sr, 65 tonnes Ba, 73 tonnes Mg, and 100 tonnes K (*14*). These ambient emissions for Cu and Mg represented 6 and 8% respectively of annual atmospheric releases in the UK during 2000 (*14*). Each of these metals acts as a coloring agent and are paired with oxidizers, including nitrates, sulfates, and perchlorates. This is known as black powder and approximately 74% consists of KNO_3, 10% sulfur, and 16% carbon in the form of charcoal (*15*). Pyrotechnic displays on celebratory dates often cause PM_{10} mass concentrations to exceed 24 h limit values (50 μg m^{-3}) as set by the European Directive 2008/50/EC. Even though this concentration is not to be exceeded on more than 35 days annually, the UK has declared in its National Air Quality Strategy that it is inappropriate to control air quality infringements resulting from social and cultural activities, including bonfires and pyrotechnic displays (*16*).

To date only a handful of small scale studies have addressed the potential adverse health effects stemming from exposure to firework emissions. Pulmonary function monitored in nine patients (seven healthy and two with chronic respiratory disease) exposed to a six times increase in PM_{10} (110 μg m^{-3} five hour experiment average which included a fifteen minute peak attaining concentrations in excess of 3.8 mg m^{-3}) from firework events was noted to cause a significant maximal midexpiratory flow rate decrease in the susceptible individuals (*17*). A potential impact on lung function was also supported by observations from a 1972 study where a 113% increase in emergency room visits by individuals with chronic respiratory disease was reported following a firework episode in Honolulu (*18*). More recently, Becker et al. (*19*) reported a near fatal and a fatal asthma exacerbation of two severely asthmatic children following exposure to elevated PM concentration from hand-held fireworks.

Despite the extensive use of fireworks to commemorate special events internationally, no literature has yet described the toxicity associated with firework PM on the respiratory system. We therefore investigated toxicologically relevant features of ambient PM collected over a three week period

in central London during a period of intense firework activity including Diwali and Guy Fawkes Night. An oxidative potential (OP) metric was formulated for each daily PM sample based on their capacity to deplete antioxidants from a validated synthetic respiratory track lining fluid (RTLF) (20). The biological pertinence of this acellular model relates to the contention that inhaled PM mediates adverse health effects through the induction of oxidative stress at the air-lung interface (21). Generation of reactive oxygen species has been demonstrated following PM challenge in various extracellular and cellular compartments (21), with evidence that different components (biological material, polycyclic aromatic hydrocarbons/quinones, trace metals) can cause oxidative stress through different pathways. Thus the OP metric used in this study integrates the contribution of a range of PM components into a single biologically informative expression.

Materials and Methods

Sampling Site Description. Sampling was conducted at the Marylebone Road kerbside site in central London, UK. Details of sampling site have been previously reported (22). Given the proximity of the sampling site to traffic emissions, considerable PM and gas species contribution from traffic sources was found. Fireworks and bonfires were ignited across the city during the sampling period by individuals and at large-scale organized public displays. Large-scale organized events commenced after dusk and extended for approximately 30 min on November second (8 displays), third (17 displays), fourth (4 displays), fifth (6 displays), ninth (2 displays), and 10th (2 displays), 2007. The number of firework incidences reported to the London Fire Brigade was used as a surrogate measure of firework displays ignited by individuals. A total of 2427 incidences were logged in the inner London Boroughs over the first ten days of November 2007, significantly greater than the outer Boroughs (1720) (23). Specifically in the London Borough of Westminster, an inner city borough in which the Marylebone Road sampling site is located, 99 firework related incidences occurred with 26 of these events reported in the immediate Marylebone area (23). An indication of the number of bonfires lit by individuals was also estimated using London Fire Brigade statistics: 262 incidences of runaway bonfires, recorded as secondary fires (nonproperty location fires free of injury), were logged on November 5, 2007, representing the highest number of calls during the year (23).

Instrumentation. PM with an aerodynamic diameter between 1 and 10 μm (PM$_{1-10}$) was collected on polyurethane foams (PUFs) using the Airborne Sample Analysis Platform (Thermo ASAP) located on the roof of the monitoring station at the Marylebone Road site. Foams were exposed for 24 h periods at 200 L min^{-1}. PUFs were changed daily at 9:30GMT for samples collected between October 24, 2007 and November 6, 2007 and at 15:00GMT November 7, 2007 to November 13, 2007. PM collected on these foams was used for trace metal analysis and assessment of PM oxidative potential.

Ambient PM$_{10}$ mass concentrations were measured by the UK Automatic Urban and Rural Network at the Marylebone Road sampling site using a Tapered Element Oscillating Microbalance (TEOM) with 15 min resolution. In addition, gaseous pollutant concentrations (SO$_2$, NO, NO$_2$) and meteorological parameters (temperature, wind direction, and wind speed) were measured on an equivalent time basis. Ambient benzene concentrations were measured at an hourly resolution by Defra's Automatic Hydrocarbon Monitoring Network using a Perkin-Elmer Online system. This consisted of an ATD400, sampling onto a carbon impregnated trap at

-30 °C for 40 min every hour and an Autosystem XL gas chromatograph with a PLOT column and flame ionization detector.

Chemicals and Chelex Water Preparation. All water, unless otherwise stated, was deionized and ultrafiltered using an Elga-stat filtration system. All chemicals used were obtained from Sigma Chemical Company Ltd. and were of analytical grade. Ultrapure Chelex100 resin treated water was utilized to eliminate background metal contamination when assessing oxidative potential (20).

Sample Preparation. PM$_{1-10}$ samples collected on PUFs were extracted in a 5% high pressure liquid chromatography (HPLC)-grade methanol prepared in Chelex resin treated ultrapure water, pH 7.0 to achieve a final PM concentration of 150 μg mL^{-1}. To ensure PM suspensions were homogeneous, resuspension was performed using a probe sonicator (MSE Soniprep150, 23 kHz, generator with a titanium probe) operated at an amplitude of 15 μm for 30 s. Further details are provided in the Supporting Information regarding the extraction efficiency of the PUFs and comparison to hourly PM$_{1-10}$ mass concentrations measurements. Interexperimental positive (residual oil fly ash) and negative (M120, carbon black) PM control samples were also resuspended using the same procedure. Compositional details of the residual oil fly ash (24, 25) and M120 (27, 26) control particles have been published previously.

Assessment of Oxidative Potential. The oxidative potential of PM was assessed *in vitro* by measuring the depletion of antioxidants by standardized concentrations of resuspended particulate in a synthetic RTLF. This chemical model contained physiologically relevant concentrations (200 μM) of urate (UA), ascorbate (AA), and glutathione (GSH) adjusted to pH 7.0 and incubated at 37 °C with equal mass concentrations of PM samples (50 μg mL^{-1}) for four hours. Briefly, AA and UA concentrations were determined using reverse phase HPLC with electrochemical detection (Jones Chromatography, Hengoed, Wales). Total glutathione (GSx) and glutathione disulfide (GSSG) concentrations were determined with the GSSG-reductase-5,5′-dithio-bis(2-nitrobenzoic acid) (DNTB) recycling assay. Subtracting two times the measured GSSG from the GSx concentration yielded GSH. Further information regarding antioxidant measurements have been described elsewhere (28). The Supporting Information contains details describing the calculation method of the AA and GSH OP metrics.

Total PM Metal Analysis. An aqua regia solution was prepared in Chelex-100 resin treated water. This digestion media was added to the resuspended PM solution such that final concentrations of 2.8% HNO$_3$, 5.1% HCl, and 7.5 μg mL^{-1} PM were achieved. Samples were digested in a microwave system (Mars 240/50) for 30 min at 180 °C on full power (1600W). Following digestion, samples were further diluted with Chelex-100 resin treated water to attain a final acid and PM concentration of ca. 1.9% HNO$_3$, 3.4% HCl, and 5 μg mL^{-1} PM. Concentrations of K, Ca, Mg, Fe, Cu, Zn, Sr, V, Ba, and Pb in solution were determined using inductively coupled plasma mass spectrometry (ELAN DRC ICP-MS, MSF008). Dilutions of a certified multielement standard solution (VI CertiPUR Merck, Lot. No.OC529648) were used for calibration. Six NIST 1648a Urban PM control samples and water blanks were run in addition to samples; 3σ of the water blank elemental concentrations were used to establish minimum detection limits.

Results and Discussion

Air Mass Origin. The dominating parameters influencing the concentration of PM species measured at the Marylebone Road sampling site were local and regional emissions. Pyrotechnic and biomass burning (in the form of Guy Fawkes Night celebratory bonfires) were known to occur locally but
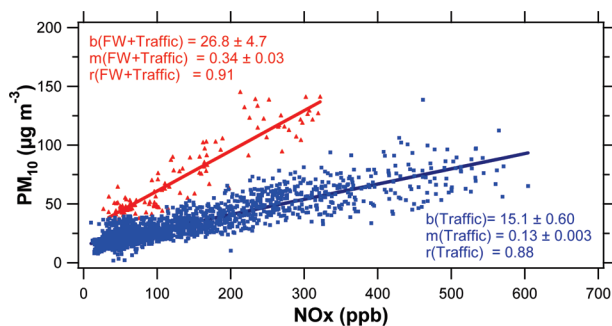
FIGURE 1. Relationship between 15 min mean NO$_x$ and PM$_{10}$ mass concentrations for the entire campaign period. The fraction of PM$_{10}$ derived from vehicle sources is colored in blue squares. Red triangular markers indicate the PM attributed to firework and bonfire events as estimated by NO$_x$ concentrations.

also took place across the United Kingdom, likely contributing to an elevated regional PM background level. The relevance of each combustion event, local or regional, to the sampling site was influenced by varying wind patterns. However, proximity of the known firework events to the sampling site and PM$_{10}$ episode timings implicated local London combustion events with the observed PM episodes. A single large-scale firework event (the Lord Mayor Show) occurred at 18:30GMT on November 10, 2007 along the River Thames (3 km southeast of the sampling site) but did not result in a PM$_{10}$ episode at Marylebone Road. Meteorological data obtained from Heathrow Airport indicated winds were from the northwest inhibiting the firework plume from reaching the sampling site. In contrast, firework activity on November 3 and 9, 2007 (at locations north and northwest of Marylebone Road) enhanced the magnitude of measured PM$_{10}$ concentration given the sampled air mass on these days originated from the north and northwest, respectively. The highest PM$_{10}$ concentrations measured during the campaign, however, occurred on the early morning of November 5, 2007 and were associated with an air mass which traveled through London from the northeast. Unlike all other discussed events, the latter period was dominated by high pressure conditions with near stagnant wind speeds (1.24 ± 0.44 m s^{-1}) which limited PM dispersion.

Nitrogen Oxides (NO$_x$) and PM$_{10}$ Relationship. A relationship between NO$_x$ and PM$_{10}$ mass concentration was used to determine the fraction of PM$_{10}$ attributable to differing combustion contributions (Figure 1) (*29, 30*). Complete regression source apportionment details are described in the Supporting Information. Briefly, this regression analysis segregated the data set into two groups to represent pyrotechnics/bonfires and traffic (*31*) and traffic alone (including vehicular tail pipe emissions, road dust resuspension, and tire and brake pad wear) sources such that each group was characterized by distinct PM$_{10}$-NO$_x$ slope. The former mass concentration grouping corresponded to (i) periods of organized firework-bonfire displays, November 3, 2007; (ii) Guy Fawkes Night, November 4 and 5, 2007; (iii) Diwali, November 9, 2007. Parallel NO$_x$ increases with firework activity have been identified previously (*2, 32*) and were suggested to originate from the black powder KNO$_3$ component (*4*). Allan et al. (*33*) evaluated the contribution of biomass combustion sources to regional NO$_x$ concentrations during the REPARTEE 2 campaign (October 16, 2007-November 9, 2007) which overlapped with the current study: a high resolution time-of-flight Aerosol Mass Spectrometer (AMS) was operated at the Regent's Park urban background site (further site details are provided in the Supporting Information). The AMS measured PM$_{2.5}$ total organics concentrations were deconvolved using Positive Matrix Factorization into four factors profiling different source

attributions: oxygenated organic aerosol, cooking organic aerosol, solid fuel organic aerosol (SFOA), and hydrocarbon-like organic aerosol (HOA). A univariate linear regression between HOA and NO$_x$ was performed in additional to a bilinear fit including both HOA and SFOA factors with NO$_x$. This latter approach did not yield improved Pearson r-values compared to univariate regression results. The authors concluded that despite the NO$_x$ emissions from solid fuel combustion, on the regional scale during REPARTEE 2, traffic dominated NO$_x$ emissions relative to solid fuel combustion in central London as more local sources were present. Thus, the enhanced rate of PM$_{10}$ emission relative to NO$_x$ identified via the regression analysis in the current study is likely isolated to pyrotechnic combustion sources and not from bonfire or other solid fuel combustion emissions.

The validity of assigning the lower PM$_{10}$-NO$_x$ slope to traffic-related emissions was evaluated by considering two week periods void of firework and bonfire activity before and after the campaign (October 9-24, 2007 and November 15-28, 2007). A similar traffic PM$_{10}$-NO$_x$ relationship was found for these periods (slope 0.14 μg m^{-3} ppb^{-1}; y-intercept 19 μg m^{-3}; $r = 0.81$; $p < 0.001$; $N = 2695$) as during the campaign (slope 0.13 μg m^{-3} ppb^{-1}; y-intercept 15 μg m^{-3}; $r = 0.87$; $p < 0.001$; $N = 1817$).

The uncertainty of traffic and firework/bonfire apportioned PM$_{10}$ fractions were determined as the combined uncertainties of each input variable derived in accordance with the ISO Guide to the expression of uncertainty in measurement (GUM) (*34*). Calculation details are provided in the Supporting Information. The average uncertainty for 15 min resolution firework apportioned PM$_{10}$ mass concentrations was 3.7 μg m^{-3}. For the 15 min averaged traffic PM$_{10}$ fraction the uncertainty on mass concentrations was estimated as 1.5 μg m^{-3}.

When interpreting temporal traffic PM$_{10}$ source fraction fluctuations (Figure 2), it is useful to note that the Marylebone Road site is located on the south side of the carriageway in a street canyon such that a vortex is created at sufficiently high wind speeds coming perpendicular to the street (*22, 35, 36*). Consequently, strong southerly winds and those flowing parallel to the street yield increased PM concentrations. Two scenarios may result from northerly wind depending on wind speed: high velocities will cause air recirculation leading to the lowest contribution of local traffic emissions to the site, while low wind speeds enable local traffic emissions to be measured directly and thus the highest PM concentrations. During the campaign, northerly winds were associated with near-stagnant wind conditions (<1 m s^{-1}). Consequentially the maximum traffic component PM$_{10}$ concentration measured was 16.3 ± 14.2 μg m^{-3} (33 ± 26% of the total PM$_{10}$ mass concentration) on average during these periods. In contrast, the lowest concentrations of this source fraction (5.1 ± 7.8 μg m^{-3} or 15 ± 14% of the total PM$_{10}$ mass concentration) were measured when higher wind speeds induced the microscale recirculation effect within this street canyon.

The firework PM component was isolated to episodes which were rapidly dispersed with only acute air quality degradations. These celebratory combustion events included (i) November 3, 2007 19:45GMT - November 4, 2007 3:45GMT (15.7 ± 5.1 μg m^{-3} PM$_{10}$ average), (ii) November 4, 2007 18:15GMT - November 5, 2007 5:00GMT (44.3 ± 17.6 μg m^{-3}), and (iii) November 9, 2007 20:45−23:45GMT (49.9 ± 10.0 μg m^{-3}). This source fraction represented 33 ± 9%, 41 ± 13%, and 60 ± 5% of these total PM$_{10}$ mass concentrations for each of the noted periods.

To further elucidate differences between these two combustion processes and other possible contributing anthropogenic emissions, PM$_{1-10}$ chemical characteristics, PM number concentration, and gaseous SO$_2$ fluctuations were
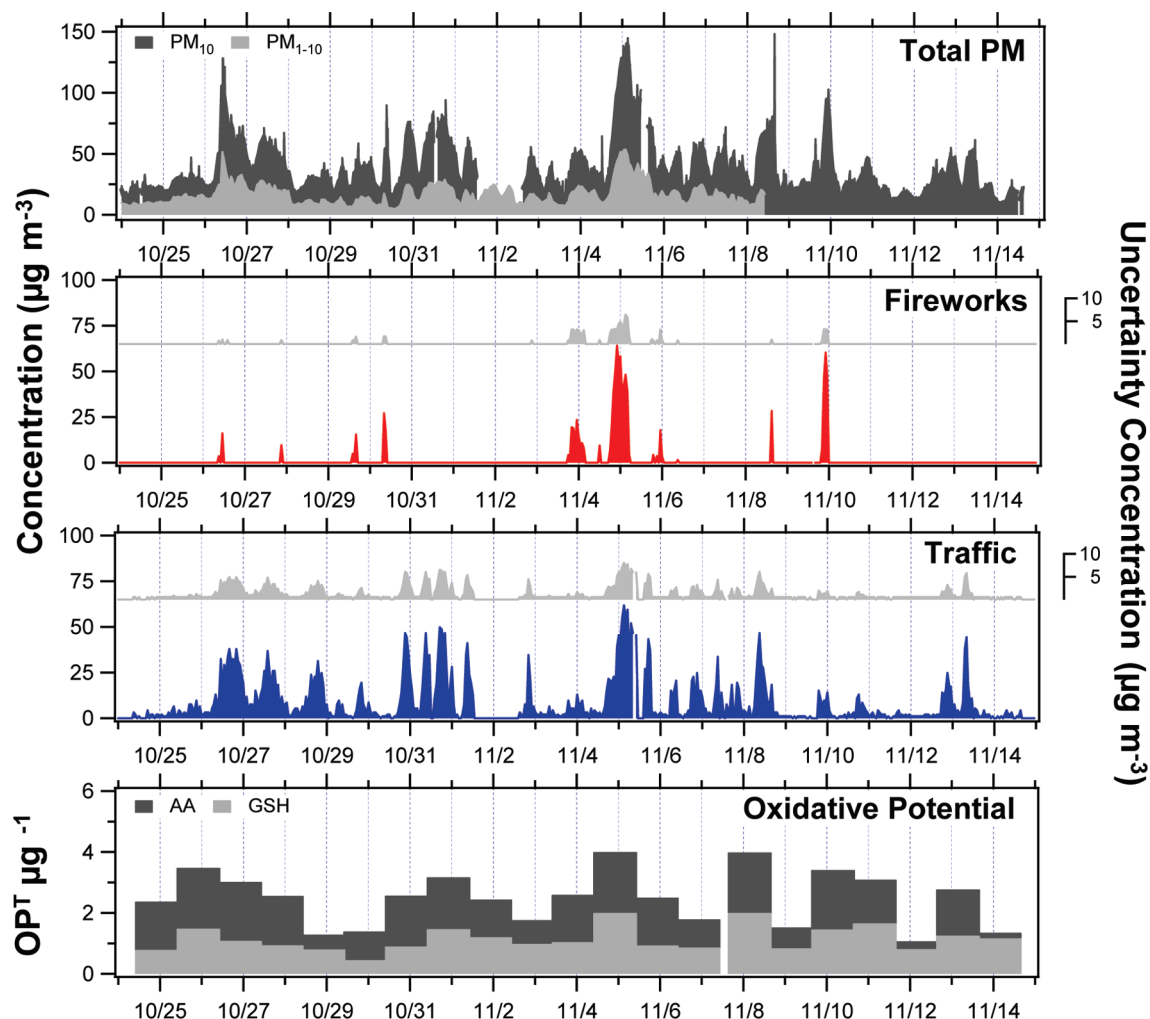
FIGURE 2. Time series of (i) PM$_{10}$ mass concentrations measured by the TEOM (1 h average) and PM$_{1-10}$ mass concentrations estimated by merged SMPS-APS measurements (1 h average). One hour resolution source apportioned PM$_{10}$ firework-bonfire (ii) and traffic (iii) fractions (left axis) with the associated uncertainty for each fraction colored in light gray (right axis). Twenty-four hour averaged PM$_{1-10}$ ascorbate (AA, light gray) and glutathione (GSH, dark gray) oxidative potential per unit mass.

assessed; the latter two are described in the Supporting Information.

Trace Metals. The temporal fluctuations of PM$_{1-10}$ Sr, K, Mg, Ba, and Pb concentrations were found to be similar; the maximum concentration measured for all these metals was for the 24-h sample collected over November 4 to 5, 2007, representing a period of intense pyrotechnic activity (Figure 3-A). An additional rise was observed for the 24-h November 9 to 10, 2007 p.m. sample, also corresponding with a firework combustion period. No significant concentration changes during these celebratory periods were found for metals predominately associated with vehicular nontail pipe emissions (Figure 3-B). Despite the elevated Cu firework emission factors estimated by Passant et al. (*14*), no Cu concentrations increases were found during periods of known firework activity. Ba concentrations reflected firework and traffic emission fluctuations, where the latter was the result of brake pad abrasion (*37, 38*). Following PM size distribution results presented in the Supporting Information, the authors acknowledge that the sampled size fraction (PM$_{1-10}$) does not represent the total metal concentration associated with firework activity. As instrumentation available for daily PM filter sampling was limited to the 1 and 10 μm cut points, the reported firework PM parameters measured were therefore underestimated given that much of the PM derived from pyrotechnic combustion events was found below 1 μm.

Trace metal enrichment factors were determined for 24-h PM$_{1-10}$ samples collected on November 4–5, 2007 and October 28–29, 2007 relative to the November 13–14, 2007 sample (Figure 4). Pyrotechnic combustion related emissions were sampled during the former as predicted from source apportionment results and the occasion of city-wide organized firework displays. In contrast, the latter two samples were primarily subject to continuous traffic emissions given the lack of apportioned firework PM component in parallel with no organized firework events. The sample collected over Guy Fawkes Night celebrations was associated with large increases of firework combustion derived trace metals. The enrichment factor calculated for the nonfirework period maintained approximately unit values for trace metals without a vehicular source. Ba, Sr, and Pb sustained the largest increases (28-, 25-, and 19-fold, respectively) on the pyrotechnic date considered. However, Ba, unlike Sr and Pb, was also associated with an increase factor of 1.4 on the nonfirework days reflective of a traffic emission contribution.

All of the measured trace metals had the potential of originating from either pyrotechnic or urban sources. To determine the elemental composition associated specifically with firework derived PM, bivariate correlations were performed with Sr and the other measured trace metals. The strongest correlation ($p < 0.0001$, $N = 21$) existed between K ($r = 0.98$), Pb ($r = 0.96$), Ba ($r = 0.92$), and Mg ($r = 0.73$)
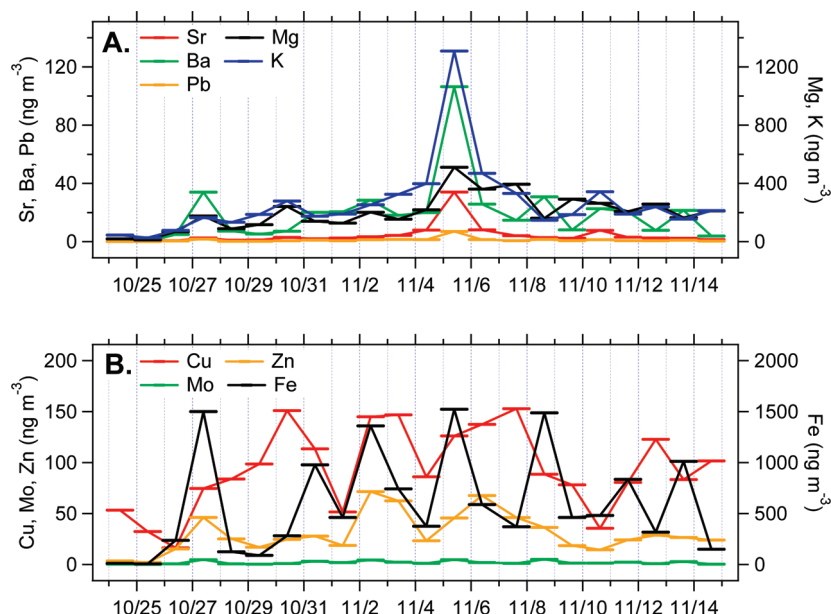
FIGURE 3. Twenty-four hour averaged PM_{1-10} concentrations for metals attributable (A; Sr, Ba, Pb, Mg, K) and not attributable (B; Cu, Mo, Zn, Fe) to firework combustion emissions.
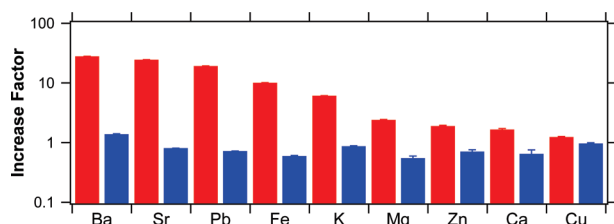

FIGURE 4. Ratio of trace metal concentrations as measured for PM samples run on (1) October 28, 2007 9:30 GMT - October 29, 2007 9:30 GMT (no fireworks events, blue bars) and (2) November 4, 2007 9:30 GMT - November 5, 2007 9:30 GMT (firework events, red bars) to PM samples collected between November 13, 2007 15:00 GMT - November 14, 2007 15:00 GMT.

confirming the elemental firework composition described by previous studies with the exception of Cu (3, 5). A weaker significant association was identified with Fe concentrations ($r = 0.43$, $p = 0.05$). No correlation was established between Sr with Ca, Cu, and Zn despite reported use of these species in fireworks as orange ($CaCO_3$, $CaCl_2$, $CaSO_4 \cdot xH_2O$), blue ($CuCO_3$, $CuSO_4 \cdot 5H_2O$, $CuCl$), and white (ZnO) coloring, respectively (39).

Oxidative Potential. Bivariate correlations were conducted with the bulk PM mass and trace metals for the ascorbate and glutathione dependent OP components on a unit mass basis. Individual metals, representative of the source apportioned firework and traffic PM fractions, were all found to be positively associated the OP parameters. A combination of the combustion derived metals were correlated with $OP^{GSH} \mu g^{-1}$ related to both traffic (Ba, $r = 0.62$, $p = 0.003$; Mo, $r = 0.57$, $p = 0.007$; Fe $r = 0.57$, $p = 0.01$; Cu) and firework (Pb, $r = 0.54$, $p = 0.01$; Sr, $r = 0.48$, $p = 0.03$) emissions. However only traffic emissions related metals were significantly associated with $OP^{AA} \mu g^{-1}$: Mo ($r = 0.48$, $p = 0.03$), Fe ($r = 0.47$, $p = 0.03$) and Ba ($r = 0.46$, $p = 0.04$). A subset of the measured trace metals (Zn, Ca, Cu) lacked an association with antioxidant depletion. The oxidative loss of antioxidants from the synthetic RTLF model is sensitive only to intrinsic redox active PM constituents: redox active metals and quinones. With the exception of Fe and Pb, the contribution of the trace metals noted in the bivariate correlation analysis to reactive oxygen species production, and thus AA and GSH depletion, has not been reported in

the literature. These species probably represent surrogates of constituents with intrinsic redox active properties which were not quantified.

A heterogeneous panel of metals, representing different source fractions, were found to be predictor/surrogate measures of $OP^{AA} \mu g^{-1}$ and $OP^{GSH} \mu g^{-1}$; the former was related to traffic and the latter was associated with both traffic and firework related combustion products. Consideration of the individual trace metals in a multivariate regression method was not feasible as the colinearity of the metal concentrations would confound results. Multivariate regressions were instead performed between the ascorbate, glutathione, and total OP $(\mu g\ PM_{1-10})^{-1}$ metrics and apportioned PM_{10} source fractions such that OP dose effects were estimated for an increase of each PM source fraction in the ambient airshed. The PM_{10}(traffic), PM_{10}(firework), and PM_{10}(other) fractions were averaged to a 24-h periods to correspond with the ASAP PUF PM_{1-10} sampling intervals. As only transient firework concentrations were measured during the campaign, the validity of averaged daily firework PM_{10} concentrations was assessed based on the number of 15 min measurements included in the 24 h average. For averaging periods comprised of only a single firework concentration measurement ($N = 5$), the timing of the episode was evaluated to avoid inclusion of misclassified firework data points; this possibility is acknowledged given the limitations in the methodology employed to segregated traffic from traffic+firework measurements (Figure 1). For these firework events isolated to a 15 min averaged period, the authors dismiss the likelihood that measurements which occurred during the late morning hours (October 26, 2007 9:00 and November 6, 2007 9:30GMT) were in fact derived from a firework source and were excluded from the regression analysis. A further restraint on the regression analysis was caused by the significant negative correlation ($r = 0.63$, $p = 0.002$) between the PM_{10}(traffic) and PM_{10}(other) fractions, arising as an artifact of the definition of PM_{10}(other) (Equation S10). Consequently, the PM_{10}(other) fraction was excluded from the regression analysis, and the predicted mean contribution of this fraction to each OP metric was calculated from the traffic and firework regression coefficients.

Significance was sustained at the 95% confidence level for all OP metrics when a multivariate model was constructed to explain the measured variance (Table 1). The plot of

TABLE 1. Multivariate Regression Analysis of OP μg^{-1} (Ascorbate, Glutathione, and Total) with PM_{10} Traffic and Firework Fractions[a]

metric	PM source fraction (μg (μg PM_{10})$^{-1}$)	explanatory variable			model	
		B	SE	p-value	r	p-value
OP^{AA} ug^{-1}	traffic	3.1	1.2	0.05	0.79	0.05
	firework	3.1	1.3	0.05		
	constant	0.4	0.4	0.30		
OP^{GSH} ug^{-1}	traffic	2.2	0.8	0.03	0.89	0.008
	firework	3.5	0.8	0.005		
	constant	0.2	0.2	0.36		
OP^T ug^{-1}	traffic	5.2	1.4	0.01	0.90	0.006
	firework	6.5	1.5	0.005		
	constant	0.6	0.4	0.18		

[a] B-values indicate the estimated change in OP μg^{-1} for an increase in 1 μg of each apportioned source fraction. The associated standard errors (SE) for regression coefficients and each model are also presented.

regression residuals against standardized predicted values exhibited random scatter around the zero line for all three OP metric models. The fireworks PM factor was implicated with the greatest increase in OP^{GSH} μg^{-1} and OP^T μg^{-1} for each 1 μg dose (GSH: 3.5 ± 0.8 μg^{-1}, $p = 0.005$; Total: 6.5 ± 1.5 μg^{-1}, $p = 0.005$) compared to the traffic fraction (GSH: 2.2 ± 0.8 μg^{-1}, $p = 0.03$; Total: 5.2 ± 1.4 μg^{-1}, $p = 0.01$). Therefore, the firework PM contributed more substantially per unit mass to the PM OP in this acellular model than the traffic related PM_{10} component. Moreover, the predicted oxidative burden associated with the PM_{10}(other) fraction was 0.51 μg^{-1} and 0.96 μg^{-1} for the glutathione and total OP models, respectively. This suggested the bulk of the oxidative activity of the particles was related to the traffic and pyrotechnic sources. In contrast to the GSH and total OP metrics, the oxidative burden associated with the traffic and firework PM_{10} fractions was equivalent (3.1 ± 1.2 μg^{-1}, $p = 0.05$) in the OP^{AA} μg^{-1} regression model. These results further highlight that depletion of each antioxidant is sensitive to a heterogeneous panel of metals. The OP predicted to be derived from the PM_{10}(other) source fraction for the case of ascorbate related oxidative activity was low (0.44 μg^{-1}) compared to the traffic and firework sources, similar to the glutathione and total OP results. It is also important to note when interpreting these results that the reported oxidative potential, in particular for the firework component, was underpredicted: the lower cut point (1 μm) of the sampled size fraction prevented collection of the entire firework PM size distribution as described in the Supporting Information.

In conclusion, this study highlighted distinctions between ambient PM and gas phase species resulting from firework or traffic activity. Furthermore, ambient PM_{10} concentrations were deconvolved into source apportioned fractions which were coupled with measurements of particulate oxidative burden. The *in vitro* PM total OP per unit mass associated with pyrotechnic activity was shown to be significantly greater than that associated with traffic derived PM emissions. From the current study it was not clear if the measured pyrotechnic PM trace metals were the drivers for the measured OP or simply represent surrogates for some unmeasured components. Thus, the authors recommend further examination of a possible relationship between firework exposure and acute respiratory outcomes.

Acknowledgments

Appendix A

AA	ascorbate
ASAP	airborne sample analysis platform
GSH	reduced glutathione
HPLC	high pressure liquid chromatography
ICP-MS	inductively coupled plasma mass spectrometry
NO_x	nitrogen oxides
OP	oxidative potential
$OP^T m^{-3}$	total particulate oxidative burden per cubic meter of sampled air
$OP^T \mu g^{-1}$	total particulate oxidative burden per unit mass PM
$OP^{AA} \mu g^{-1}$	particulate OP with respect to AA depletion per unit mass PM
$OP^{GSH} \mu g^{-1}$	particulate OP with respect to GSH depletion per unit mass PM
PM	particulate matter
PM_{10}	particulate matter with an aerodynamic diameter of 10 μm
PM_{1-10}	particulate matter with an aerodynamic diameter between 1 and 10 μm
PUF	polyurethane foams
RTLF	respiratory tract lining fluid
TEOM	tapered element oscillating microbalance

Supporting Information Available

The supplemental methodology details the instrumentation employed for continuous size distribution measurements, sampling location, campaign period, and source apportionment uncertainty calculations. Additional results are included discussing temporal variation of PM physical (size distributed PM number and mass concentrations) and gaseous pollutant (SO_2) characteristics. This material is available free of charge via the Internet at http://pubs.acs.org.

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Weighing fireworks' effects

By *Trent Spiner*
Created *07/03/2010 - 00:00*
Northwood lake
Research suggests contamination

When Jim Vaillancourt looks out over Northwood Lake from his porch tomorrow night, he won't be surprised to see two neighbors locked in their annual contest for the biggest, longest and most awesome fireworks show. After all, last year's battle went for about an hour and a half.

But as revelers plan to light fireworks across the area tomorrow, one state official said preliminary research shows fireworks can harm local water bodies because their residue can cause cancer, contaminate water supplies and poison wildlife.

State limnologist Jody Connor said he plans to continue researching the effect of fireworks from this weekend after finding that the heavy metals used by some manufacturers are already affecting the state's water bodies.

"I think a lot of people think fireworks are beautiful and have no toxins in them, but we are trying to educate people there are some toxins; even the cardboard can cause pollution," he said.

While Connor's research is preliminary, it has helped residents from at least two local municipalities ask for a fireworks ban near water bodies.

In Northfield, Vaillancourt asked selectmen last year to consider a ban while they were discussing a permitting process for lighting fireworks.

"I can't imagine that over the weekend people who are just pounding hours' worth of fireworks out over the lake can't have some kind of detrimental effect over the long term," he said.

Vaillancourt, president of the Northwood Lake Watershed Association, said he is trying to educate residents within a mile of the lake about how their actions can pollute the water. Since speaking with selectmen, banning fireworks has dropped down his list of issues after the lake required an emergency treatment last month to fight an invasive species of milfoil.

When he discusses it with neighbors, he said many argue that the safety concerns of using fireworks over land outweigh environmental concerns.

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NORTHWOOD LAKE

Weighing fireworks' effects - page 2

Research suggests contamination

In Franklin, resident Martin Russo told councilors that he sees more trash from bob houses than fireworks. Rep. David Palfrey said septic systems and runoff from other sources do more damage than fireworks, according to minutes of the meeting.

Councilors questioned how the ordinance would work and who would enforce it.

"I think they felt it was another level of regulation and they didn't want it," Smith said.

Connor said some companies are researching fireworks that contain fewer heavy metals.

"All these metals eventually fall to the earth, and they eventually get into the lakes," Connor said. "I'm not sure there's enough research out there, but eventually we'll have additional research that tells us how dangerous they may be to the atmosphere and to the water."

Trent Spiner can be reached at 369-3306 or tspiner@cmonitor.com.

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What does gas, oil and boat motor exhaust add to the water?
By Robert Frypp - 07/03/2010 - 8:10 am

Not to mention road salt that eventually makes it's way into the water as well when the snow melts? Because I'd worry a lot more about those effects than the minuscule, if any, effect a couple errant firework remnants might/could add to the water once a year.

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