

Surface Water Ambient Monitoring Program (SWAMP) Synthesis Report on Stream Assessments in the San Diego Region

March 2008



SURFACE WATER AMBIENT MONITORING PROGRAM (SWAMP) SYNTHESIS REPORT ON STREAM ASSESSMENTS IN THE SAN DIEGO REGION

Raphael D. Mazor Ken Schiff

Southern California Coastal Water Research Project 3535 Harbor Blvd., Suite 110 Costa Mesa, CA 92626 www.sccwrp.org

Prepared for the California Regional Water Quality Control Board, San Diego Region (Region 9).

This project was funded by the Surface Water Ambient Monitoring Program.

Technical Report 527_Synthesis

TABLE OF CONTENTS

1
2
3
3
5
6
7
9
9
18
31
33
37
A - 1

LIST OF FIGURES

4
6
3) 9
d 2
3
4
5
6
7
8
9
0
6
7

LIST OF TABLES

Table 1. Watersheds in the San Diego region	4
Table 2. Summary of data sources	5
Table 3. Selected water chemistry constituents	10
Table 4. Mean percent control of each toxic indicator in each watershed	13
Table 5. Physical habitat at sampled sites	14
Table 6. Bioassessment scores by watershed	15
Table 7. Correlations of biological metrics with NMS axes	19
Table 8. Correlations of selected water chemistry constituents with NMS a	axes
and IBI	21
Table 9. Correlations of toxicity endpoints with NMS axes and IBI	23
Table 10. Correlations of physical habitat assessments with NMS axes an	nd IBI
· · ·	24
Table 11. Correlations of landscape metrics with NMS axes and IBI	24

1. ABSTRACT

Watershed managers require regional data to develop biomonitoring tools and contextualize local assessments. However, they often rely on data generated by programs with a more local emphasis, such as studies mandated by pollution discharge permits. These programs typically study only specific sites or stream reaches, The goal of this study was to compile individual data sets from sitespecific programs to see if they could be merged into a regional-scale program. We evaluated if the merged data could be used to a) perform a regional assessment of streams in southern California, and b) identify potential stressors to aquatic life in these streams. Water quality, toxicity, physical habitat, and benthic macroinvertebrate samples were collected from over 100 sites in coastal watersheds in San Diego, Riverside, and Orange counties by six different programs. The data indicated widespread impacts to many water chemistry constituents, with some, like ammonia-N and specific conductivity, exceeded aquatic life thresholds in more than 60% of samples. More than 50% of water and sediment samples were toxic to at least one indicator species (Ceriodaphnia dubia, Hyallela azteca or Selenastrum capricornutum). Of the 708 bioassessment samples included in the study, 80% were in poor condition (i.e., index of biotic integrity < 40). Impacts for all indicators were most severe in urban areas along the coast. Nonmetric multidimensional scaling of benthic macroinvertebrate communities identified two stressor gradients: a strong gradient associated with toxic contaminants in the water (e.g., metals, high specific conductivity, and organics) and a weaker gradient related to eutrophication (e.g., dissolved oxygen and nitrate). The toxic contaminant gradient was strongly associated with development in the watershed, and watersheds with more than 10% developed area were invariably in poor biological health. In conclusion, combined data sets produced potentially biased regional assessments because of their sampling designs, which emphasized evaluations of known impacts. In contrast, such data are more useful for stressor identification, as they effectively capture important gradients in the region. Integration of local programs may prove useful if they are designed in coordination to meet regional as well as local goals.

2. INTRODUCTION

With few exceptions (e.g., EPA 2006), stream monitoring has been applied in largely piecemeal fashion around the country in response to regulatory-based requirements. Southern California serves as a good example. Collectively, more than 12 agencies collect over 300 samples in the 18 major coastal watersheds in just the 6 from San Diego to Ventura. For the most part, these programs employ independent, site-specific monitoring designs that target specific discharge locations (SMC 2007).

Despite the lack of programs that focus beyond specific sites or watersheds, there is a tremendous need for regional scale evaluations of stream health. The first need for regional stream monitoring is to address questions posed by the public that tend to focus on streams as a whole as opposed to just the reaches where there are potential sources of impacts. The second need for regional stream monitoring is to develop assessment tools that watershed managers need for evaluating potential impacts. One such tool is stressorresponse relationships (Van Sickle et al. 2006). Regional scale programs are one way to collect the information necessary for developing stressor-response relationships because they not only capture the full breadth of natural variation, but a wide range of anthropogenically induced impacts. It is this range of stressor impact that is important for anchoring the spectrum of stream responses. The third need for regional stream monitoring is to help set management priorities, which is especially important in these times of limited resources. Regional monitoring programs provide the context of the worst and best streams that allow managers to effectively target the locations of greatest need.

One mechanism to achieve regional scale assessments is to link multiple local- or watershed-scale programs. However, there are many challenges associated with this approach. Individual programs may not measure similar indicators or, if they do, they may not measure the indicators using similar methods. Different programs may also have differing levels of quality assurance. The result is to force all of the data to the lowest level of QA common among them, which may be insufficient for management-level assessments. Finally, even if indicators, methods, and quality assurance were similar, data management can present an enormous hurdle. Undoubtedly, each monitoring program stores its data in different ways, from simple to sophisticated, making the collation of data an unusually burdensome (if not impossible) task.

The goal of this study was to determine if multiple, local scale data sets could be combined to make regional scale assessments. In this case study, we selected data sets collected by the State of California and by numerous dischargers regulated by the National Pollutant Discharge Elimination System (NPDES) permits in the San Diego region to answer two questions of regional importance:

- 1) What is the health of streams in the San Diego region?
- 2) What are the primary stressors responsible for biological responses in the San Diego Region?

The challenge was to combine data collected over 11 different hydrologic units and a time period of 9 years. None of the data was collected in concert, none of the designs was integrated, and none of the data systems were connected in any way. Thus, this study was an evaluation of the ability of such programs to address regional needs

3. METHODS

Setting

The San Diego region includes all coastal watersheds north of the Mexican border and south of the Santa Ana River. Covering portions of Orange, Riverside, and San Diego Counties, the region encompasses nearly 4,000 mi² and ranges from the mountains of the Peninsular Range mountains to the Pacific Coast.

Southern California is characterized by an arid mediterranean climate, with hot dry summers and cool wet winters. Average monthly rainfalls measured at the Lindberg Airport (SDG) in San Diego, California between 1905 and 2006 show that nearly all rain fell between the months of October and April, with hardly any falling between the months of May and September (California Department of Water Resources 2007). The wettest month was January, with an average rainfall of 2.05"). Average annual rainfall at this station was 10.37".

The San Diego Region consists of several coastal rivers and streams that are grouped into 11 hydrologic units (Figure 1, Table 1). The Tijuana River is the largest in the region. Other large rivers include the Santa Margarita, San Luis Rey, San Dieguito, San Diego, Sweetwater, and Otay Rivers (Figure 1). The watersheds extend from the Lagunas, the Cuyamacas, and other mountains of the Peninsular Range. Most of the larger rivers are regulated by large dams. The streams of the San Diego Region have profound effects on coastal ecology and the Southern California Bight (Ackerman and Schiff 2003). Discharging over 300 million m³ annually in typical years, the rivers are an important source of freshwater for San Diego and Mission Bay, as well as several estuaries and coastal wetlands.

Urban development extends along almost the entire coastal strip of the region (23% of the region), although large undeveloped areas remain in coastal northern San Diego County in Camp Pendleton Marine Corps Base. Many smaller coastal watersheds are entirely urbanized. Agricultural land use occurs in 9% of the region, and is most extensive in the San Luis Rey and San Dieguito watersheds. Open space predominates in the interior, as well as in the

aforementioned Camp Pendleton, covering 68% of the region (SANDAG 1998). The extent of undeveloped open space varies among each watershed, from a low of 12% in Pueblo San Diego to a high of 92% in San Juan (Table 1).

Watersheds	Abbreviation	HUC	Area (mi ²)	% Open	% Developed	% Agricultural
San Juan	SJ	901	496	92	7	1
Santa Margarita	SM	902	750	81	13	6
San Luis Rey	SLR	903	560	61	15	24
Carlsbad	CB	904	211	38	50	12
San Dieguito	STO	905	346	18	61	21
Los Peñasquitos	LP	906	162	43	53	4
San Diego	SD	907	440	72	26	2
Pueblo San Diego	PSD	908	56	12	88	0
Sweetwater	SW	909	230	67	29	4
Otay	OT	910	154	70	20	10
Tijuana	TJ	911	463	90	6	4
TOTAL			3868	68	23	9

Table 1. Watersheds in the San Diego region. Land uses are calculated from data provided by SANDAG (1998)



Figure 1. Hydrologic units and land use within the San Diego region. Abbreviations are given in Table 1. Dark green is undeveloped open space. Orange is agricultural land. Gray is developed land. Inset shows location of the San Diego region within California.

Sources of data

This report combines data collected by the State of California's Surface Water Ambient Monitoring Program (SWAMP) with data from California Department of Fish and Game (CDFG), and NPDES monitoring by San Diego and Orange Counties, Camp Pendleton Marine Corps Base, and the Padre Dam Municipal Water District (Table 2). All these assessment programs used a targeted design to select sites of interest for sampling. Most sites were selected in order to assess known disturbances in the watershed, although a few undisturbed sites were targeted to set reference expectations for specific studies. A total of 62 sites were sampled under SWAMP for water chemistry and toxicity. Physical habitat was assessed at all but nine of these sites. Bioassessment samples were collected at 144 sites, of which 35 were located at or within 500 m of sites with water chemistry and toxicity data (Figure 2). All four indicators were measured at 29 sites.

 Table 2. Summary of data sources used in this analysis. Additional data from each of these programs was analyzed in watershed-specific reports.

Program	Years	Watersheds	Indicator	Sites	Samples
SWAMP	2000-2006	11	Bioassessment	71	17
			Water chemistry	62	233
			Toxicity	62	235
			Physical habitat	53	53
California Dept. of Fish and Game	1998-2005	11	Bioassessment	98	408
San Diego County NPDES	2002-2005	9	Bioassessment	45	169
Orange County NPDES	2002-2005	1	Bioassessment	18	87
Camp Pendleton	2004-2006	1	Bioassessment	7	14
Padre Dam MWD	2004-2006	1	Bioassessment	2	10
All programs	1998-2006	11	Bioassessment	144	708
			Water chemistry	62	233
			Toxicity	62	235
			Physical habitat	53	53



Figure 2. Locations of sampling sites. Blue circles are sites sampled under SWAMP for water chemistry, toxicity, and physical habitat. Red triangles are sites sampled for bioassessment.

To aggregate data collected under multiple programs, sites within 500 m of each other were treated as a single site. This distance was based on published measures of spatial correlation of benthic communities in streams (Gebler 2004). Although data used in this assessment cover many years (1998-2006), there was little indication that conditions had changed over the course of this study apart from a few sites in the upper Sweetwater watershed, which were affected by the 2003 Cedar Fires (see reports on specific watersheds for details about specific sites).

Indicators

Multiple indicators were used to assess the sites in the San Diego region. Water chemistry, water and sediment toxicity, benthic macroinvertebrate communities, and physical habitat.

Water chemistry was measured as per the SWAMP Quality Assurance Management Plan (QAMP) (Puckett 2002). Measured indicators included physical measures of water quality (e.g., pH, temperature dissolved oxygen, etc.), inorganics, pesticides, polycyclic aromatic hydrocarbons (PAHs), dissolved metals, pesticides, and polychlorinated biphenyls (PCBs). The Appendix contains a complete list of water chemistry constituents. To evaluate water and sediment toxicity to aquatic life, toxicity assays were conducted on samples from each site as per the SWAMP QAMP (EPA 1993, Puckett 2002). Water toxicity was evaluated with 7-day exposures on the water flea, *Ceriodaphnia dubia*, and 96-hour exposures to the alga *Selenastrum capricornutum*. Both acute and chronic toxicity to *C. dubia* was measured as decreased survival and fecundity (i.e., eggs per female) relative to controls, respectively. Chronic toxicity to *S. capricornutum* was measured as changes in total cell count relative to controls. Sediment toxicity was evaluated with 10-day exposures on the amphipod *Hyallela azteca*. Both acute and chronic toxicity to *H. azteca* was measured as decreased survival and growth (mg per individual) relative to controls, respectively. Chronic toxicity endpoints (i.e., *C. dubia* fecundity, *H. azteca* growth, and *S. capricornutum* total cell count) were used to develop a summary index of toxicity at each site.

Physical habitat was assessed using semi-quantitative observations of 10 components relating to habitat quality, such as embeddedness, bank stability, and width of riparian zone. The assessment protocols are described in The California Stream Bioassessment Procedure (California Department of Fish and Game 2003). Each component was scored on a scale of 0 (highly degraded) to 20 (not degraded). 53 sites were assessed, although data were incomplete at 26 sites. Sites were assessed by the average component score.

To assess the ecological health of the streams in the San Diego region, 708 benthic macroinvertebrate samples were collected at 144 sites. Samples were collected using SWAMP-comparable protocols, as per the SWAMP QAMP (Puckett 2002). Three replicate samples were collected from riffles at each site; at least 300 individuals were sorted and identified from each replicate, creating a total count of over 900 individuals per site. Using a Monte Carlo simulation, all samples were reduced to 500 count for calculation of the Southern California Index of Biotic Integrity (IBI; Ode et al. 2005), a composite of seven metrics summed and scaled from 0 (poor condition) to 100 (good condition).

A GIS analysis was used to calculate simple landscape metrics for each site. Land use data came from the San Diego Association of Governments, and the Tijuana River Watershed GIS Database (SANDAG 1998, CESAR 2000). Both data sources use compatible procedures for identifying and naming land uses. Land use categories were aggregated into three classes: open space, developed land, and agricultural land. Metrics were calculated for the entire contributing watershed, as well as at a local scale (i.e., within 500 m of the sampling site).

Data Analysis

Water quality was assessed by comparing water chemistry constituents to known thresholds, when possible (SDRWQCB 1994, EPA 1997, CCR 2007).

Watersheds were compared by plotting distributions of concentrations of selected constituents. Toxicity was assessed by plotting frequency of samples with endpoints significantly different from controls for each indicator species. Bioassessment samples were assessed by calculating the Southern and Central California Index of Biotic Integrity (IBI, Ode et al. 2005), and comparing samples to a threshold of 40 (i.e., poor or very poor condition versus fair or better condition).

To assess the influence of water chemistry, toxicity, physical habitat, and land use on benthic communities, nonmetric multidimensional scaling (NMS) was used to ordinate bioassessment samples. Because of the lack of synoptic data for many sites, 500-count subsamples were averaged to produce mean abundances for each site. Number of samples per site ranged from 1 to 13. NMS was run with the following parameters: 1000 runs with real data, 100 runs with randomized data, 4 maximum number of axes, 250 maximum number of iterations, 0.2 step length, 0.000001 stability criterion. NMS was run in PC-ORD v 5.12 (McCune and Mefford 2006).

To assess the influence of environmental variables on biotic structure, water chemistry, toxicity, physical habitat, and landscape variables were correlated with NMS axis scores using Spearman's rank correlation (ρ). In addition, these variables were correlated to IBI scores to determine their relationship with biological condition. Correlation strength (based on ρ^2) rather than statistical significance was used to identify strong relationships, as the high number of tests may yield spurious significance and low power.

The SWAMP QAMP guided QA/QC for all data collected under SWAMP (See SWAMP QAMP for detailed descriptions of QA/QC protocols, Puckett 2002). QA/QC officers flagged non-compliant physical habitat, water chemistry, toxicity, and tissue results. No data were excluded as a result of QA/QC violations.

4. RESULTS

Assessment of the watersheds

Many sites showed signs of degraded water chemistry. For example, most sites had elevated nutrients, metals, and other constituents. Several of these constituents occurred in concentrations known to harm aquatic life. For example, more than 60% of samples exceeded applicable aquatic life thresholds of 0.025 mg/L of ammonia-N. Exceedances for specific conductivity, sulfate, selenium, and total phosphorus were nearly as frequent. Some anthropogenic organic constituents, such as diazinon, lack thresholds but were detected at many sites (Figure 3). Table 3 shows selected water chemistry constituents in each watershed. The full list of constituents is included in the appendix.





Concentrations of many constituents were high in most watersheds, although come values were extreme in sites from the Tijuana watershed that receive surface flows from Mexico (e.g., ammonia-N and phosphorus, Figure 4). Although elevated values were found throughout the region, some samples from the Tijuana River were 1 - 2 orders of magnitude more concentrated than other samples. Samples with elevated concentrations of many constituents were also found in the San Juan, Carlsbad, and Los Peñasquitos watersheds (Table 3, Appendix).

					SJ		mann	SM	Jui	<u></u>	SLR			CB		<u>. uppij</u>	STO	1 00	impico.	LP	
Constituent	Symbol	Threshold Units	Source	Mean	SD	n	Mean	SD	n	Mean	SD	n	Mean	SD	n	Mean	SD	n	Mean	SD	n
Physical water quality																					
Alkalinity as CaCO3	Alk	20000 mg/l	EPA 2002	208	109	39	187	49	21	198	84	25	252	43	41	243	129	18	221	88	20
Oxygen, Dissolved	DO	6 mg/L	SDRWQCB 1994			0			0	9.4	3.6	24			0			0			0
рН		6 or 8 pH	SDRWQCB 1994	7.6	1.5	38	7.4	0.8	20	7.7	0.4	24	7.9	1.5	40	8.0	0.4	17	7.9	0.4	19
Salinity	Sal	None ppt		1.05	1.77	32	2.62	6.16	20	0.83	0.52	24			0	2.53	5.47	17			0
Specific conductivity	Cond	1600 µS/cm	CCR 2007	2032	2839	38	4399	9779	20	1516	968	24	3800	4232	40	4316	8781	17	2981	1256	19
Sulfate	SO_4	250* mg/l	SDRWQCB 1994	497	491	39	352	398	21	382	264	25	469	277	41	358	367	18	674	407	20
Nutrients																					
Ammonia as N	NH ₃ -N	0.025 mg/l	SDRWQCB 1994	0.19	0.52	39	0.02	0.05	21	0.05	0.06	25	0.12	0.11	41	0.10	0.12	18	0.09	0.06	20
Nitrate as NO3	NO ₃	None mg/l		1.79	2.20	39	22.48	19.92	20			0	16.00		1	1.96	2.58	17	0		1
Total Phosphorus as P	TP	0.1 mg/l	SDRWQCB 1994	0.22	0.27	39	0.21	0.22	21	0.21	0.24	25	0.14	0.09	41	0.24	0.35	18	0.10	0.16	20
Metals		0																			
Arsenic	As	50 µg/L	SDRWQCB 1994	3.4	2.5	39	2.5	3.9	21	1.3	0.9	25	4.7	2.6	41	2.1	1.7	18	3.4	0.8	20
Cadmium	Cd	5 µg/L	SDRWQCB 1994	0.26	0.34	39	0.04	0.03	21	0.03	0.02	25	0.05	0.04	41	0.03	0.03	18	0.02	0.01	20
Chromium	Cr	50 µg/L	SDRWQCB 1994	0.25	0.22	39	0.23	0.35	21	0.33	0.28	25	1.06	1.05	41	0.18	0.17	18	0.89	0.97	20
Copper	Cu	9 µg/L	EPA 1997	4.05	2.85	39	3.10	2.37	21	4.03	2.60	25	3.55	1.50	41	2.41	1.76	18	4.03	1.58	20
Lead	Pb	2.5 µg/L	EPA 1997	0.02	0.02	39	0.01	0.01	21	0.06	0.05	25	0.05	0.12	41	0.03	0.03	18	0.05	0.08	20
Manganese	Mn	5* µg/L	EPA 2002	148	329	39	92	139	21	133	270	25	127	147	41	135	135	18	141	156	20
Nickel	Ni	52 µg/L	EPA 1997	5.55	6.76	39	0.71	1.22	21	0.97	2.14	25	2.16	1.37	41	0.70	0.79	18	3.38	3.55	20
Selenium	Se	5 µg/L	EPA 2002	7.5	10.4	38	5.9	16.8	20	4.9	4.8	24	10.6	10.1	40	3.7	5.4	17	7.8	3.7	19
Silver	Ag	3.4 µg/L	EPA 1997	0.20	1.22	39	0.09	0.29	21	0.07	0.34	25	0.05	0.28	41	0.00	0.00	18	0.06	0.25	20
Zinc	Zi	120 µg/L	EPA 2002	4.1	3.1	38	2.3	1.5	20	2.7	1.9	24	6.5	6.4	40	2.2	1.7	17	8.4	8.7	19
Organics																					
Benzo(b)fluoranthene		0.0044 ng/L	EPA 2002	3.5	9.6	39	1.0	3.2	21	0	0	25	0	0	41	3.0	5.9	18	0	0	20
PCBs		0.014 ng/L	EPA 2002	2.45	6.68	39	0	0	21	0	0	25	0	0	41	0	0	18	0	0	20
Diazinon		None ng/L		43.97	103.77	38	7.78	18.10	20	0.67	2.43	24	68.39	101.40	40	12.64	14.90	17	50.81	51.65	19
DDE(p,p')		0.00059 ng/L	EPA 2002	0.29	0.66	39	0.90	2.64	21	0.04	0.20	25	1.37	2.24	41	0.17	0.51	18	5.30	12.71	20
DDTs		None ng/L		0.58	1.28	39	1.48	3.89	21	0.04	0.20	25	1.93	2.79	41	0.28	0.96	18	5.65	12.75	20
Dieldrin		0.00014 ng/L	EPA 2002	0.15	0.42	39	0	0	21	0	0	25	0.02	0.16	41	0.11	0.32	18	0	0	20
Disulfoton		None ng/L		3.95	10.28	38	0	0	20	0	0	24	33.33	38.29	40	0	0	17	52.92	64.68	19
Heptachlor epoxide		0.0038 ng/L	EPA 1997	0.21	0.70	39	0.10	0.30	21	0	0	25	0	0	41	0.11	0.32	18	0	0	20
Secbumeton		None ng/L		3.11	13.48	38	1.75	7.83	20	4.50	12.50	24	85.00	153.57	40	0	0	17	131.16	147.98	19

Table 3. Selected water chemistry constituents. SD = Standard deviation. n = number of samples. * = Thresholds may not apply to all samples.

Table 3, continued.

					SD		F	PSD		:	SW			OT			ΤJ	
Constituent	Symbol	Threshold Units	Source	Mean	SD	n	Mean	SD	n	Mean	SD	n	Mean	SD	n	Mean	SD	n
Physical water quality																		
Alkalinity as CaCO3	Alk	20000 mg/l	EPA 2002	242	85	27	191	15	5	191	92	15	233	21	7	400	164	12
Oxygen, Dissolved	DO	6 mg/L	SDRWQCB 1994	9.5	3.0	26	113.3	198.9	4	9.2	2.0	14			0	8.3	4.3	12
рН		6 or 8 pH	SDRWQCB 1994	8.0	0.3	26	8.8	0.8	4	8.0	0.4	14	7.8	0.3	5	7.9	0.7	12
Salinity	Sal	None ppt		1.00	0.45	26	4.19	5.66	4	1.85	1.57	14	1.30	0.85	5	0.76	0.46	12
Specific conductivity	Cond	1600 µS/cm	CCR 2007	1872	830	26	6925	9619	4	3930	3649	14	2478	1539	5	1482	887	12
Sulfate	SO_4	250* mg/l	SDRWQCB 1994	283	159	27	437	394	5	276	190	15	217	40	7	201	133	12
Nutrients																		
Ammonia as N	NH ₃ -N	0.025 mg/l	SDRWQCB 1994	0.05	0.05	27	0.05	0.05	5	0.06	0.05	15	0.08	0.10	7	11.33	16.63	12
Nitrate as NO3	NO ₃	None mg/l				0	0		1			0	9.21	9.14	7			0
Total Phosphorus as P	TP	0.1 mg/l	SDRWQCB 1994	0.12	0.09	27	0.25	0.07	5	0.06	0.04	15	0.01	0.02	7	3.31	3.70	12
Metals		C C																
Arsenic	As	50 µg/L	SDRWQCB 1994	3.2	2.4	27	2.5	1.5	5	11.5	16.8	15	7.7	6.0	7	3.7	2.6	12
Cadmium	Cd	5 µg/L	SDRWQCB 1994	0.03	0.02	27	0.08	0.05	5	0.02	0.02	15	0.02	0.01	7	0.06	0.03	12
Chromium	Cr	50 µg/L	SDRWQCB 1994	0.64	0.74	27	1.22	0.79	5	0.96	1.06	15	0.37	0.30	7	2.67	2.94	12
Copper	Cu	9 µg/L	EPA 1997	4.16	2.06	27	8.23	3.94	5	4.25	3.05	15	2.91	1.16	7	5.14	6.25	12
Lead	Pb	2.5 µg/L	EPA 1997	0.09	0.08	27	0.51	0.28	5	0.07	0.13	15	0.02	0.02	7	0.25	0.27	12
Manganese	Mn	5* µg/L	EPA 2002	60	116	27	61	67	5	54	71	15	41	67	7	238	228	12
Nickel	Ni	52 µg/L	EPA 1997	1.15	1.92	27	3.99	2.81	5	0.78	0.93	15	1.80	3.22	7	9.16	11.13	12
Selenium	Se	5 µg/L	EPA 2002	8.1	6.8	26	77.5	115.8	4	26.6	27.8	14	9.2	7.3	6	7.2	4.6	12
Silver	Ag	3.4 µg/L	EPA 1997	0.00	0.00	27	0.13	0.27	5	0.00	0.00	15	0.39	1.02	7	0.02	0.04	12
Zinc	Zi	120 µg/L	EPA 2002	3.9	2.3	26	13.6	4.1	4	2.9	2.0	14	2.0	0.8	6	4.5	6.6	12
Organics																		
Benzo(b)fluoranthene		0.0044 ng/L	EPA 2002	0.8	4.0	27	5.8	13.0	5	0	0	15	0	0	7	8.8	30.3	12
PCBs		0.014 ng/L	EPA 2002	0	0	27	0	0	5	0	0	15	0	0	7	0	0	12
Diazinon		None ng/L		1.73	5.50	26	26.25	29.17	4	11.36	11.19	14	20.83	26.47	6	16.92	20.93	12
DDE(p,p')		0.00059 ng/L	EPA 2002	0	0	27	0	0	5	0	0	15	1.43	2.51	7	0	0	12
DDTs		None ng/L		0	0	27	0	0	5	0	0	15	2.00	2.83	7	0	0	12
Dieldrin		0.00014 ng/L	EPA 2002	0	0	27	0	0	5	0	0	15	0	0	7	0	0	12
Disulfoton		None ng/L		0	0	26	13.25	26.50	4	4.14	15.50	14	0	0	6	16.17	40.76	12
Heptachlor epoxide		0.0038 ng/L	EPA 1997	0	0	27	0	0	5	0	0	15	0	0	7	0	0	12
Secbumeton		None ng/L		12.92	48.18	26			0			0	0	0	6			0



Figure 4. Concentrations of selected constituents in each sample by watershed. Box and whiskers represent 5th, 25th, 50th, 75th, and 95th percentiles. Dots represent values above the 95th or below the 5th percentile. Numbers below the X-axis represent the number of samples from that watershed for all constituents, except for diazinon (which had one less sample in each watershed).

Water and sediment toxicity was frequently observed throughout the region, although certain indicator species were more sensitive than others. For example, 59% of all water samples were toxic to the algae *S. capricornutum*. The two arthropod indicators were less sensitive, with the amphipod *H. azteca* suffering increased mortality when exposed to 27% of sediment samples, and the water flea *C. dubia* showing reduced fecundity when exposed to 34% of water samples (Figure 5).



Figure 5. Frequency of toxicity for each endpoint and indicator species are shown in the black bars. Toxicity was determined if sample endpoints were less than 80% of controls, and the difference was significant at the 0.05 level. Weaker (but still significant) results are shown in the gray bars. Empty space above the bars indicate the proportion of samples not indicating toxicity. Numbers above bars indicate number of samples.

Toxicity was observed in every watershed. The frequency of toxicity to chronic endpoints (i.e., *C. dubia* young per female, *H. azteca* growth, and *S. capricornutum* total cell count) ranged from 24% of samples in the Carlsbad watershed to 90% of samples in the Tijuana watershed (Table 4).

				C. d	ubia						Н	. az	teca				S. ca	pricc	ornutu	m	All	
	5	Survi	ival		Υοι	ıng/f	emale	3		Survi	val			Gro	wth		Tota	al cel	l coun	t	Indica	ators
Watershed	Mean	SD	Freq	n	Mean	SD	Freq	n	Mean	SD	Freq	n	Mean	SD	Freq	n	Mean	SD	Freq	n	Freq	n
SJ	89	27	0.11	35	92	26	0.35	34	153	193	0.29	25	80	34	0.12	28	64	35	0.64	39	0.32	161
SM	93	22	0.05	21	95	23	0.20	20	106	34	0.00	13	101	6	0.15	13	96	61	0.52	21	0.20	88
SLR	98	6	0.00	34	80	26	0.32	34	99	19	0.00	16	106	5	0.06	16	82	41	0.50	32	0.21	132
CB	96	34	0.10	39	87	43	0.44	36	121	62	0.45	40	68	34	0.13	40	70	30	0.55	40	0.33	195
STO	91	25	0.06	16	92	25	0.20	15	130	31	0.00	7	101	6	0.00	8	61	38	0.88	17	0.30	63
LP	95	34	0.16	19	104	56	0.33	18	129	79	0.38	16	82	22	0.25	16	56	27	0.84	19	0.40	88
SD	101	4		30	82	26	0.37	30	94	39	0.21	11	92	28	0.36	14	82	44	0.45	29	0.28	114
PSD	42	60	0.50	2	45	64	0.50	2	110	39	0.00	2	109	2	0.00	2	57	35	0.50	2	0.30	10
SW	108	4		2	123	17		2	86	15	0.14	5	102	14	0.20	7	121	100	0.50	4	0.21	20
ОТ	96	9		5	84	28	0.40	5	106	36	0.50	3	81	24	0.33	4	66	26	0.71	7	0.41	24
ТJ	105			1	103			1	125	26	0.40	9	70	48	0.00	10	64	79	0.50	2	0.20	23

Table 4. Mean percent control of each toxic indicator in each watershed. SD = standard deviation. n = number of samples. Freq = frequency of toxicity. -- = No toxicity detected.

Physical habitat ranged from very poor to very good, although the majority of sites showed some signs of degradation. Every watershed contained some sites in good condition, except for watersheds where few sites were assessed. For example, all sites in the Santa Margarita watershed were in very good condition, with mean physical habitat scores greater than 15 (Figure 6, Table 5).

Some components of physical habitat were more often degraded than other components. For example, a large majority of sites had poor scores (< 5) for embeddedness. Degradation of velocity-depth regimes were nearly as bad, with the majority of sites scoring below 10. In contrast, sediment deposition, channel flow, and bank stability were in good condition (score > 15) at the majority of sites (Figure 7, Table 5).

Table 5. Physical habitat at sampled sites. Symbols above the columns indicate the watershed. Numbers indicate number of sites assessed within each watershed. SD = standard deviation.

												-						
		SJ	SN	1	SLR		CB		STO	C	LP		SD)	SW	TO		TJ
		11	4		6		10		5		6		7		1	2		1
Component	Symbol	Mean SD	Mean	SD	Mean S	SD	Mean	Mean	SD	Mean								
Mean score	AvePHAB	10.5 4.7	' 15.3	0.4	13.3 2	2.0	11.4	2.6	14.7	3.1	10.9	4.7	11.1	3.2		8.7	5	
Epifaunal cover	EpiCov	11.0 5.1	16.3	2.2	13.3 3	3.2	12.1	5.6	13.8	6.0	10.7	7.3	10.3	4.6	13	8.5	6	10
Embeddedness	Embed	9.8 6.6	5 3.5	1.9	5.5 6	6.4	3.5	4.1	5.0	5.7	6.7	7.8	5.4	7.0	13	3	1	3
Velocity-Depth Regime	VelDep	8.2 4.4	14.8	2.5	10.0 1	1.1	12.1	4.6	11.2	4.0	10.3	4.5	7.6	4.0	8	5.5	4	9
Sediment Deposition	SedDep	11.0 6.2	13.5	1.7	15.2 1	1.3	13.8	4.9	17.2	0.8	12.7	8.2	15.6	4.8	13	8.5	8	5
Channel Flow	ChanFlo	11.3 6.0	17.3	2.1	15.3 3	3.9	13.9	5.4	16.0	6.2	12.8	5.5	14.7	4.3	13	8	6	18
Channel Alteration	ChanAlt	11.9 7.7	19.3	1.0	14.0 2	2.4	7.8	6.4	17.6	1.9	10.5	7.1	11.3	5.9	18	6.5	9	19
Riffle Frequency	RifFreq	11.2 7.3	17.8	1.7	12.2 4	4.7	10.2	6.0	13.2	7.9	11.0	7.0	7.9	6.1	13	8.5	9	16
Bank Stability	BankStab	11.0 7.5	5 16.5	2.4	14.5 2	2.1	16.3	5.4	18.0	3.4	12.8	5.4	15.3	2.8	18	16.5	4	16
Vegetative Protection	VegPro	11.4 6.8	17.8	1.3	15.3 2	2.9	15.4	5.3	19.2	1.8	12.8	5.3	11.3	3.3	18	14	6	15
Riparian Vegetation	RipVeg	10.5 8.2	16.5	2.4	15.3 3	3.8	8.9	4.7	15.4	6.3	8.2	6.8	11.7	3.4	18	8	8	19



Figure 6. Mean physical habitat scores for all sites within each watershed. Box and whiskers represent 5th, 25th, 50th, 75th, and 95th percentiles. Points represent values above the 95th and below the 5th percentiles. No sites in the Pueblo San Diego watershed were assessed. Numbers indicate number of sites assessed in each watershed.



Figure 7. Scores for each component of physical habitat. Abbreviations are given in Table 5. Box and whiskers represent 5th, 25th, 50th, 75th, and 95th percentiles. Points represent values above the 95th and below the 5th percentiles. Numbers indicate number of sites assessed for each component.

IBI scores covered nearly the entire range of the index, with a low score of 0 and a high score of 93 (Table 6). However, the overwhelming majority (80%) of the 708 samples were below the impairment threshold of 39. Poor conditions were observed in every sample from 61% of sites. Good conditions (IBI > 39) were observed in samples from 39% of sites. Although samples in poor condition were found in every watershed, samples in good condition were absent from smaller coastal watersheds, like Carlsbad, Los Peñasquitos, and Pueblo San Diego. A majority of samples (65%) from the Tijuana watershed were in good condition, as were a near-majority (44%) in the Santa Margarita watershed (Figure 8). Sites with samples in good condition were largely restricted to the interior mountains of the larger watersheds. However, a few samples in good condition were occasionally detected in smaller coastal watersheds in undeveloped portions of southern Orange County and in the Camp Pendleton Marine Corps Base (Figure 9).

Table 6. Bioassessment scores by watershed. SD = Standard deviation. Frequency = frequency of samples in poor condition (i.e., IBI < 40).

				IB		
Watershed	Sites	Samples	Years	Mean	SD	Frequency
SJ	26	132	1998-2005	25	18	0.83
SM	16	113	1998-2006	37	16	0.56
SLR	16	75	1998-2006	28	20	0.73
CB	22	125	1998-2005	13	8	1.00
STO	9	27	2000-2005	24	13	0.89
LP	10	62	1998-2006	16	8	1.00
SD	20	75	1996-2006	17	13	0.96
PSD	2	7	2003-2005	18	6	1.00
SW	10	46	1998-2005	28	20	0.74
ОТ	2	3	2000-2001	25	18	0.67
TJ	16	43	1999-2006	43	18	0.35



Figure 8. Boxplot of IBI scores of bioassessment samples in each watershed. Boxes and whiskers represent 5th, 25th, 50th, 75th, and 95th percentiles; points represent scores above the 95th percentile or below the 5th percentile. The dashed line represents the threshold for impaired conditions (i.e., 40).



Figure 9. Distribution of IBI scores in the San Diego region. A) Mean IBI scores at each site. B) Frequency of samples in poor or very poor condition (i.e., IBI < 40).

Stressor relationships

Ordination of mean taxa abundances yielded a 3-axis solution with moderately low final stress of 17.6. The three axes combined represented 82.6% of the variance in the site-by-taxa matrix, with the third axis representing the largest portion (40%) of this variance, followed by axis 2 (23.9%) and axis 1 (18.7%). No clustering of sites in ordination plots by watershed was evident (Figure 10).



Figure 10. NMS Ordinations of sites in the San Diego region. Each point represents the ordination of the mean abundance of all samples collected at that site. Symbols represent different watersheds. Final stress was 17.6. Numbers in the axis titles is the proportion of variability represented by the axis.

Examination of weighted scores for taxa showed that that several mayflies, stoneflies, and caddisflies were located high on axis 2, while several non-insects were low on axis 2. However, there was considerable diversity within all taxonomic groups; for example, the caddisfly *Hydroptila* had relatively low score of -0.16 on axis 2, and the dipteran *Dasyhelea* had a high score of 0.57. Dipterans, like *Simulium* and Muscidae dominated the low end of axis 1, and non-insects, such as Oligochaeta, Cladocera, Corbicula, and Nematoda were at the high end. No obvious pattern characterized axis 3; the stonefly *Malenka* and the dipteran *Dasyhelea* were at the high end, while several caddisflies *Ochrotrichia*, the mayfly *Baetis*, and the clam *Corbiculum* were at the low end. (Figure 11).



Figure 11. Weighted averages of selected taxa in ordination space. Symbols represent taxonomic groups. Only taxa appearing in 40 or more sites are shown.

Correlation of ordination axes with IBI and metric scores showed that most metrics responded strongly to axis 2 (Figure 12). In general, higher values on axis 2 corresponded to better ecological condition. For example, the IBI as well as the metrics EPT richness and % intolerant individuals had strong positive correlations with axis 2 (ρ^2 of 0.50, 0.50, and 0.54 respectively), and % non-insect taxa had a moderately strong negative relationship ($\rho^2 = 0.35$). Two metrics (i.e., % collectors and % tolerant taxa) showed no strong relationships with any axis, perhaps because of the ubiquity of collector and tolerant taxa at both disturbed and undisturbed sites. No metric showed strong relationships with axis 1 or 3 (Figure 12). However, the IBI showed a unimodal relationship with axis 1 (Table 7, Figure 13).

Table 7. Correlations of biological metrics with NMS axes. n = Number of sites used to calculate correlations. Spearman rank correlations (ρ)
Biological metric
Symbol
NMS1
NMS2
NMS3
n

		opeanin		onoratione	(P)
Biological metric	Symbol	NMS1	NMS2	NMS3	n
Index of biotic integrity	IBI	-0.04	0.70	0.02	44
Frequeny of impaired samples		-0.10	-0.60	-0.07	44
EPT Taxa	EPT tx	0.16	0.58	0.13	44
Coleoptera Taxa	Coleo tx	-0.11	0.71	-0.08	44
Predator Taxa	Pred tx	-0.03	0.53	-0.08	44
% Collectors	% Coll	-0.12	-0.15	0.11	44
% Intolerant	% Intol	0.02	0.74	0.20	44
% Non-Insecta Taxa	% NI tx	0.33	-0.59	-0.04	44
% Tolerant Taxa (8-10)	% Tol tx	0.22	-0.39	0.09	44



Figure 12. Correlations of variables with NMS axes. Length of vectors represent strength and direction of correlation, as measured by Spearman's rank correlation coefficient (ρ).



Correlation analyses revealed that many environmental variables related to water chemistry were correlated with axis 2. For example, many metals, ammonia-n, specific conductivity, sulfate, and several organic constituents (such as diazinon and secbumeton) had strong negative correlations (Spearman's ρ^{2} > 0.2) with this axis (Table 8). In addition, frequency of toxicity for all endpoints were negatively correlated with this axis, further suggesting that axis 2 represents a toxic contamination gradient (Table 9). In contrast, only two variables (NO₃-N and dissolved oxygen) were strongly correlated with axis 1, suggesting that this axis may represent a eutrophication or nutrient enrichment gradient. No water chemistry or toxicity variables were strongly correlated with axis 3 (Figure 12). Many of the variables that had significant correlations with axis 2 also had significant correlations with the IBI (e.g., arsenic, ammonia-N, etc., Table 8-9).

Table 8. Correlations of selected water chemistry constituents with NMS axes and IBI. N = number of sites used to calculate correlations. A) Physical water quality, metals, and nutrients. B) PAHs, PCBs, and pesticides.

		Spearr	nan rar	k correl	ations	(ρ)
A. Water qualityNon-organic constituents	Symbol	NMS1	NMS2	NMS3	IBI	n
Physical water quality and inorganics						
Alkalinity as CaCO3	Alk	0.02	-0.48	0.00	-0.47	35
Sulfate	SO ₄	0.14	-0.51	-0.14	-0.58	35
Oxygen, Dissolved	DO	-0.56	0.05	-0.19	0.10	14
pH		-0.32	-0.15	-0.06	-0.19	34
Salinity	Sal	0.06	-0.49	-0.23	-0.76	24
Specific conductivity	Cond	0.23	-0.61	-0.07	-0.72	34
Temperature		0.10	-0.32	-0.05	-0.55	34
Total Suspended Solids		0.10	-0.11	0.05	-0.13	27
Turbidity		-0.02	-0.19	-0.06	0.02	34
Velocity		0.06	0.02	-0.05	-0.08	35
Metals						
Aluminum		0.16	0.20	0.27	0.10	35
Arsenic	As	0.11	-0.68	-0.08	-0.79	35
Cadmium	Cd	0.15	-0.34	-0.33	-0.31	35
Chromium	Cr	0.03	-0.52	0.02	-0.68	35
Copper	Cu	-0.21	-0.58	-0.26	-0.58	35
Lead	Pb	-0.24	0.06	0.20	-0.04	35
Manganese	Mn	0.23	-0.60	-0.07	-0.46	35
Nickel	Ni	0.31	-0.55	0.10	-0.59	35
Selenium	Se	0.08	-0.61	-0.08	-0.78	34
Silver	Ag	0.22	-0.47	0.08	-0.51	35
Zinc	Zi	-0.11	-0.56	-0.33	-0.62	34
Nutrients						
Ammonia as N	NH ₃ -N	-0.03	-0.57	-0.28	-0.53	35
Nitrate + Nitrite as N		0.01	-0.14	-0.28	-0.31	35
Nitrate as N		0.01	0.03	-0.18	-0.28	27
Nitrate as NO3	NO_3	0.50	0.09	-0.29	-0.20	13
Nitrite as N		-0.07	-0.08	-0.29	-0.24	27
Nitrogen, Total Kjeldahl		-0.15	-0.38	-0.12	-0.38	35
OrthoPhosphate as P		0.20	-0.36	0.03	-0.50	23
Phosphorus as P, Total	TP	0.08	-0.46	-0.16	-0.40	35

Table 0, continueu.	le 8, continue	d.
---------------------	----------------	----

	Spearr	nan ran	k correlations (ρ)
B. Water qualityOrganic constituents	NMS1	NMS2	NMS3 IBI n
PAHs			
Acenaphthene	-0.15	-0.33	0.05 -0.33 35
Benz(a)anthracene	-0.15	-0.33	0.05 -0.33 35
Benzo(b)fluoranthene	-0.07	-0.39	-0.08 -0.22 35
Chrysene	-0.16	-0.34	0.01 -0.30 35
Fluorenes, C2 -	0.36	-0.45	-0.14 -0.41 33
Naphthalenes, C3 -	-0.15	-0.41	-0.10 -0.15 33
Naphthalenes, C4 -	-0.15	-0.34	-0.22 -0.12 33
Phenanthrene	-0.06	-0.39	0.00 -0.32 35
PCBs			
PCBs	0.05	-0.18	-0.20 0.02 35
Pesticides			
DDE(p,p')	0.33	-0.23	-0.25 -0.23 35
DDTs	0.35	-0.23	-0.25 -0.19 35
Demeton-s	0.38	-0.20	-0.09 -0.26 34
Diazinon	0.19	-0.57	-0.12 -0.57 34
Dimethoate	0.15	-0.34	-0.05 -0.21 34
Dioxathion	-0.31	-0.26	0.17 -0.33 34
Disulfoton	0.16	-0.49	-0.02 -0.52 34
Endosulfan sulfate	-0.01	-0.07	-0.35 0.05 35
Endrin Aldehyde	0.35	-0.14	0.05 -0.33 35
HCH, alpha	0.06	-0.42	-0.32 -0.14 34
HCH, delta	0.24	-0.44	-0.17 -0.20 34
Oxadiazon	0.08	-0.67	-0.23 -0.61 34
Oxychlordane	0.32	-0.13	0.05 -0.13 34
Parathion, Methyl	0.09	0.00	-0.32 -0.06 34
Prometon	0.43	-0.15	0.33 -0.46 28
Propazine	0.01	-0.34	-0.23 -0.37 28
Secbumeton	0.51	-0.35	0.12 -0.43 28
Terbuthylazine	0.32	-0.29	0.03 -0.39 28
Thiobencarb	-0.31	-0.15	-0.40 0.08 34

Table 9. Correlations of toxicity endpoints with NMS axes and IBI. N = number of sites used to calculate correlations.

		Spearman rank correlations (p				
Toxicity indicator	Symbol	NMS1	NMS2	NMS3	IBI	n
C. dubia survival (% control)		0.00	0.25	-0.11	0.16	32
C. dubia young/female (% control)		0.01	0.27	-0.20	0.29	32
H. azteca survival (% control)		-0.03	0.45	0.23	0.47	33
H. azteca growth (% control)		0.28	0.02	-0.07	0.13	32
S. capricornutum total cell count (% control)		0.00	0.61	0.08	0.35	34
C. dubia survival frequency	CerSur	0.15	-0.39	0.09	-0.36	32
C. dubia young/female frequency	CerYou	-0.06	-0.46	0.00	-0.40	32
H. azteca survival frequency	HyaSur	-0.03	-0.40	-0.22	-0.42	33
H. azteca growth frequency	HyaGro	-0.04	-0.35	-0.02	-0.36	32
S. capricornutum total cell count frequency	SelTcc	-0.12	-0.47	-0.03	-0.27	34

Physical habitat variables were correlated with both axis 1 and 2, although none with $\rho^2 > 0.2$. The strongest physical habitat variables were riparian vegetation, riffle frequency, channel alteration, and epifaunal cover with ρ^2 with axis 2 of 0.17-0.18; these variables all had significant relationships with the IBI as well. Correlations with axis 1 were weaker, with ρ^2 ranging from 0.10-0.15. Therefore, physical habitat degradation appears to be associated with both water chemistry contamination and eutrophication (Table 10, Figure 12).

	Spearman rank correlations (ρ)					
Physical habitat component	Symbol	NMS1	NMS2	NMS3	IBI	n
Epifaunal cover	EpiCover	-0.26	0.39	0.06	0.45	30
Embeddedness	Embed	-0.31	0.29	0.07	0.53	28
Velocity-depth regime	VelDep	0.00	0.09	-0.12	0.04	28
Sediment deposition	SedDep	0.07	0.08	0.12	-0.13	26
Channel flow	ChanFlo	0.02	0.04	-0.20	-0.08	27
Channel alteration	ChanAlt	-0.39	0.41	0.03	0.64	29
Riffle frequency	RifFreq	-0.34	0.41	0.01	0.64	28
Bank stability	BankStab	0.01	0.22	0.16	-0.02	30
Vegetative protection	VegPro	-0.04	0.28	0.09	0.12	30
Riparian zone	RipZone	-0.28	0.42	0.15	0.59	29
Mean score	AvePHAB	-0.32	0.41	0.02	0.50	28

Table 10. Correlations of physical habitat assessments with NMS axes and IBI. N = number of sites used to calculate correlations.

Analysis of landscape-scale variables suggest that the extent of development in the watershed strongly influences benthic community structure. For example, total developed area in the watershed, as well as percent of developed land in the watershed, both had strong negative relationships with axis 2 (ρ^2 of 0.26 and 0.21, respectively). Landscape metrics reflecting local land use were more weakly correlated with axis 2, with local developed land having a ρ^2 of 0.13. Axis 2 was most strongly correlated with distance from coast ($\rho^2 = 0.53$), perhaps reflecting the higher intensity of development along the coast. No landscape-scale variable correlated strongly with axis 1 or 3 (Table 11, Figure 12).

Table 11. Correlations of landscape metrics with NMS axes and IBI. N = number of sites used to calculate correlations.

Variable	Symbol	Unit	NMS1	NMS2	NMS3	IBI	n
Land use, watershed-wide							
Open space in watershed	Open	log km ²	0.00	0.02	0.05	0.03	44
Agricultural land in watershed		log km ²	0.00	0.03	-0.02	0.02	44
Developed land in watershed	Developed	log km ²	0.13	-0.51	-0.07	-0.70	44
Percent open space in watershed		%	-0.07	0.35	0.17	0.42	44
Percent agricultural land in watershed		%	0.03	0.16	-0.01	0.13	44
Percent developed land in watershed	% Developed	%	0.13	-0.46	-0.12	-0.58	44
Land use, local							
Percent open space within 500 m		%	0.06	0.23	-0.02	0.41	44
Percent agricultural land within 500 m	Ag500	%	-0.14	0.32	0.16	0.23	44
Percent developed land within 500 m		%	0.02	-0.37	-0.10	-0.55	44
Other landscape-scale variables							
Distance from coast	Coast	km	-0.30	0.72	0.00	0.72	44
Watershed area	WSA	log km ²	0.00	-0.12	-0.01	-0.10	44

Inspection of scatterplots revealed that some variables had a strong influence on the IBI. For example, IBI scores were never above 30 where more than 10% of the watershed was developed. Similar relationships were observed with other variables, such as arsenic concentration, frequency of acute toxicity to *C. dubia*, sulfate, and arsenic. In contrast, more linear relationships were observed for other variables, such as distance from coast. Wedge-shaped relationships, were observed for several nutrients, such as ammonia-N and total phosphorus (Figure 14). Several variables showed no discernible relationship, such as dissolved oxygen and frequency of toxicity to *S. capricornutum.*



Figure 14. Relationships between IBI scores and landscape, nutrient, water quality, and toxicity variables.

Ternary plots of sites according to land use in the watershed further suggested a strong role for development in the watershed as limiting biological health (Figure 15). Sites in fair or good condition (mean IBI > 40) were tightly clustered on the right side of plots, where developed land was minimal. Similar patterns were observed for watershed-wide and local scales.



Figure 15. Ternary plots of sites showing land use in A) the contributing watershed, and B) within 500 m. Red dots indicate sites in very poor condition (mean IBI 0 - 20). Pink dots indicate sites in poor condition (mean IBI 20 – 40). Purple dots indicate sites in fair condition (mean IBI 40 – 60). Blue circles indicate sites in good condition (mean IBI 60 – 80).

5. DISCUSSION

Assessment of the region

Impacts to streams in southern California in this study were pervasive, and were associated with a large suite of potential stressors, including multiple water chemistry constituents, toxic waters and sediments, and degraded physical habitat. Impacts to all indicators were observed, and in most cases impacts were widespread. In general, smaller coastal watersheds (e.g., Carlsbad, Los Peñasquitos, Pueblo San Diego, and Otay) were more impacted, suffering from elevated water contaminants, high toxicity, degraded physical habitat, and poor biological condition. However, all watersheds contained sites suffering impacts to multiple indicators.

Despite the prevalence of observed impacts, some watersheds contained sites in good health. Larger watersheds with extensive undeveloped areas (e.g., Santa Margarita, San Diego, and Tijuana) contained sites in moderate to good health, generally clustered in the interior. Bioassessment samples from these sites were frequently in fair or good condition, and contamination of the water column was less severe. The San Juan hydrologic unit was unique in that it contained sites in moderate to good health near the coast. Unlike all other watersheds, the San Juan hydrologic unit contains extensive undeveloped coastal areas, where these sites were located.

Using data from local programs for regional assessments was a qualified success. These programs generated considerable quantities of data within the San Diego region, measuring multiple indicators in all watersheds. However, extrapolating results from the sites in this study to the entire San Diego region should be done cautiously. All sites were targeted for sampling, often because impairment was suspected at many of these sites. Therefore, the assumption that the sites in the study represent the region as a whole is most likely violated, resulting in a regional assessment that may be worse than the true condition.

Sampling was typically driven by municipal stormwater permits or other mandates, resulting in a high concentration of sites in densely populated and highly developed areas along the coast. Because of this focus on urban streams vast areas of the interior, such as the upper San Luis Rey River and the Santa Margarita, contained no sites (Figure 2).

Relationship between stressors, ecological health, and land use

Despite the limits of the data in making regional assessments, they helped determine the relationships between ecological health and potential stressors to

aquatic life. The targeted selection of sites by local programs was adequate to establish gradients of most stressors, such as nutrients or metals in the water column. Poor biological condition was associated with elevated metals, organic constituents, nutrients, specific conductivity, pH, and many other water chemistry constituents. Water and sediment toxicity was more frequent at biologically impacted sites than at sites in good biological condition. Degraded physical habitat was also associated with poor biological condition.

Nonmetric multidimensional scaling of benthic communities suggests ecosystem health was degraded in two different ways: toxic contamination of the water column and eutrophication. These two gradients of degradation correspond to two of the three ordination axes (axes 2 and 1, respectively). Sites located on the low end of axis 2 had elevated concentrations of many contaminants, such as metals and organic constituents, as well as elevated specific conductivity, sulfates, and other water quality constituents. Furthermore, toxicity was frequently evident at sites on the low end of axis 2. In contrast, the eutrophication gradient was strongly related to dissolved oxygen and to concentrations of nitrate; sites on the high end of axis 1 may be in a nutrient-enriched state. Physical habitat scores were related to both axes, suggesting that degradation of physical habitat is associated with both eutrophication and with contamination of the water column. The lack of strong relationships between the third axis and potential stressors suggest that this axis represents either responses to unmeasured stressors, or natural variability in stream communities. Such variability may arise from environmental heterogeneity, as well as biotic processes like dispersal, predation, and competition among stream biota (Power et al. 1998).

The fact that the IBI and its composite metrics were all strongly correlated to axis 2 suggests that the IBI is a good tool to detect impacts from altered water chemistry, as well as degraded physical habitat. However, the weak relationship with axis 1 suggests that it may not be sensitive to impacts caused by eutrophication. A complementary index, either based on macroinvertebrate taxa that respond to this gradient (e.g., *Simulium*, Oligochaeta, Cladocera, etc.) or based on assemblages with higher sensitivity to eutrophication (e.g., periphyton) may improve assessment of impacts related to nutrient enrichment.

The nature of the data collected for this assessment does not allow identification of stressors that were directly responsible for the observed impacts to ecological health. However, the extent of developed land had one of the strongest associations with poor health observed in this study. It is evident that increased development in watersheds—perhaps as little as 10 percent—could seriously impact stream ecosystems. Similar thresholds have been identified in many other regions of the world (e.g., Hatt et al. 2004, Walsh et al. 2007). Although agricultural land within the watershed was not shown to be associated with impacts to aquatic life, watersheds with extensive agricultural activity were

minimally represented in the data set, Furthermore, agricultural land may include areas that are minimally affected, such as lightly grazed pasture.

Land use may affect stream health at both local as well as watershed scales. Our data showed that both watershed-scale and local-scale land use measured were associated with poor biological integrity. Furthermore, local conditions, as reflected by physical habitat condition, was also associated with biological health. The role near-stream conditions and riparian buffers in biological integrity has long been recognized (Hickey and Doran 2004, Moore and Palmer 2005). However, recent research suggests that watershed condition is more important than local riparian condition (e.g., Walsh et al. 2007). This study supports the finding of other studies that protection of aquatic life may require addressing local habitat, as well as watershed-wide alterations of land use and stream hydrology (Taylor et al. 2004, Walsh et al. 2005).

Conclusions

Most sites within the San Diego region were in poor condition.

Multiple lines of evidence suggested that many sites in the San Diego region were in poor condition. For example, over half of all water samples exceeded applicable aquatic life thresholds for multiple water chemistry constituents, such as ammonia-n, selenium, specific conductivity, or sulfate. Water or sediment toxicity was evident in the majority of samples; toxicity to the alga *Selenastrum capricornutum* was the most widespread affecting 59% of all samples. Impacts to benthic macroinvertebrate communities were particularly prevalent, with 80% of over 700 bioassessment samples in poor or very poor condition. Good bioassessment condition was never observed at 87 of the 144 (60%) sites assessed.

Multiple stressors were associated with poor biological condition

Poor biological condition was associated with many potential stressors, including altered water chemistry, high toxicity and degraded physical habitat. Nonmetric multidimensional scaling showed that benthic communities responded to two different gradients of stressors: toxic contaminants in the water column (e.g., trace metals, organic constituents) and eutrophication (e.g., nutrients, low dissolved oxygen). The toxic contaminant gradient accounted for more of the variability observed in biological communities compared to the eutrophication gradient and was closely related to frequency of toxicity, as well as the index of biotic integrity. Degradation of physical habitat was associated with both toxic contaminant and eutrophication gradients.
Development in the watershed was a strong predictor of biological health.

Sites with extensive development in the contributing watershed were invariably in poor condition. The data suggest that development in as little as 10% of the watershed was enough to degrade biological integrity, although other stressors may affect aquatic life in undeveloped watersheds. Development in the watershed correlated strongly with the toxic contaminant gradient.

Recommendations

SWAMP should integrate its monitoring with other monitoring programs in the region to increase cost-efficiency

SWAMP is an important foundation for monitoring in the San Diego Region. SWAMP provides a high-quality data set that is not constrained to specific waterbodies or pollutant categories. It cannot, however, monitor all waterbodies for all of the important attributes RWQCB staff need for decisionmaking. This study found that hundreds of samples were collected for NPDES monitoring in the San Diego Region, sometimes in the same locations at the same time as SWAMP. SWAMP should look to integrate its monitoring with NPDES monitoring to extend its resources. This was a similar recommendation to what the SPARC had provided to the SWRCB during its most recent external review of the SWAMP program.

SWAMP should redesign its monitoring program to improve effectiveness at addressing important monitoring questions.

One of the primary questions to be addressed by the SWAMP program was "what is the health of streams in the San Diego Region?". Answering this question was hindered due to a potentially biased monitoring design that targeted sites for sampling. Often, these were sites with known sources of pollutants. A probabilistic monitoring design would provide a more accurate assessment of stream health overall while requiring fewer sample sites. The probabilistic design has been used by others including the US EPA's Perennial Stream Assessment (PSA) and the Southern California Stormwater Monitoring Coalition's Regional Watershed Monitoring Program (SMC).

Identify a set of core indicators that can help determine impacts to beneficial uses

Two challenges affected the ability to assess regional stream health and examine stressor-response relationships. First, few bioassessment samples were collected synoptically with water chemistry, toxicity, or physical habitat

assessments. The lack of synoptic data obscures potentially strong stressor relationships, as stresses may wax or wane between sampling events. Second, potentially important indicators were not measured. Impacts such as eutrophication may be detected most effectively using other indicators like periphyton (attached algae). Once again, there is opportunity to integrate and collaborate with local scale monitoring programs such as NPDES and larger scale programs such as PSA and SMC to define a list of core indicators. This also presents another opportunity for cost efficiency, where the hundreds of chemicals monitored during this study need not be measured.

SWAMP should ensure that there is an infrastructure to support its collaborative programs

One problem encountered during this study was the inability to combine data sets from different programs. Differing data structures, QA requirements, plus field and laboratory methods hindered effective progress towards meaningful interpretations of the data. SWAMP should engage in shared information management systems, integrated quality assurance system checks, and common field and laboratory method manuals to ensure that integration among monitoring programs becomes seamless. These activities have already made a start including California's Environmental Data Exchange Network (CEDEN) and the intercalibration exercises being conducted by the SMC.

6. LITERATURE CITED

Ackerman, D. and K. Schiff. 2003. Modeling storm water mass emissions to the Southern California Bight. *Journal of Environmental Engineering* 129: 308-317.

Bêche, L.A., E.P. McElravy and V.H. Resh. 2005. Long-term seasonal variation in the biological traits of benthic-macroinvertebrates in two Mediterranean climate streams in California, USA. *Freshwater Biology* 51:56-75.

Bernstien, B. and Schiff, K. 2002. Stormwater research needs in Southern California. Technical Report 358. Southern California Coastal Water Research Project. Westminster, CA.

California Regional Water Quality Control Board, San Diego Region. 1994. Water quality control plan for the San Diego Region. San Diego, CA. http://www.waterboards.ca.gov/sandiego/programs/basinplan.html

California Department of Fish and Game. 2003. California Stream Bioassessment Procedure: Protocol for Biological and Physical/Habitat Assessment in Wadeable Streams. Available from www.dfg.ca.gov/cabw/cabwhome.html.

California Department of Water Resources. 2007. http://www.water.ca.gov/.

Center for Earth Systems Analysis Research. 2000. Tijuana River Watershed GIS Database. San Diego, CA. Available from <u>http://geography.sdsu.edu/Resources/Data/Clearinghouse/trw.php</u>

Collins, J.N., E.D. Stein, M. Sutula, R. Clark, A.E. Fetscher, L. Grenier, C. Grosso and A. Wiskind. 2007. California Rapid Assessment Method (CRAM) for Wetlands and Riparian Areas. Version 5.0. Available from http://www.cramwetlands.org.

Environmental Protection Agency (EPA). 1993. Methods for measuring acute toxicity of effluents and receiving waters to freshwater and marine organisms, Fourth Edition. EPA 600/4-90/027. US Environmental Protection Agency, Environmental Research Laboratory. Duluth, MN.

Environmental Protection Agency (EPA). 1997. Water quality standards: Establishment of numeric criteria for priority toxic pollutants for the state of California: Proposed Rule. *Federal Register* 62:42159-42208.

Environmental Protection Agency (EPA). 2002. National recommended water quality criteria. EPA-822-R-02-047. Environmental Protection Agency Office of Water. Washington, DC.

Environmental Protection Agency (EPA). 2006. Wadeable Streams Assessment: A collaborative survey of the nation's streams. EPA 841-B-06-002. Environmental Protection Agency. Office of Water. Washington, DC.

Gebler, J.B. 2004. Mesoscale spatial variability of selected aquatic invertebrate community metrics from a minimally impaired stream segment. *Journal of the North American Benthological Society* 23:616-633.

Hatt, B.E., T.D. Fletcher, C.J. Walsh, and S.L. Taylor. 2004. The influence of urban density and drainage infrastructure on the concentrations and loads of pollutants in small streams. *Environmental Management* 34: 112-124.

Hickey, M.B.C. and B. Doran. 2004. A review of the efficiency of buffer strips for the maintenance and enhancement of riparian ecosystems. *Water Quality Research Journal* 39: 311-317.

Moore, A.A. and M.A. Palmer. 2005. Invertebrate biodiversity in agricultural and urban headwater streams: Implications for conservation and management. *Ecological Applications* 15: 1169-1177.

McCune, B. and M. J. Mefford. 2006. PC-ORD. Multivariate Analysis of Ecological Data. Version 5.12 MjM Software, Gleneden Beach, Oregon, U.S.A.

National Academy of Sciences. 1977. Drinking Water and Health. Volume 1. Washington, DC.

National Oceanic and Atmospheric Administration. 2007. National Weather Service data. Available from http://www.wrh.noaa.gov/sgx/obs/rtp/rtpmap.php?wfo=sgx

Ode, P.R., A.C. Rehn and J.T. May. 2005. A quantitative tool for assessing the integrity of southern California coastal streams. *Environmental Management* 35:493-504.

Ode, P.R. 2007. SWAMP Bioassessment Procedures: Standard operating procedures for collecting benthic macroinvertebrate samples and associated physical and chemical data for ambient bioassessment in California. Available from http://mpsl.mlml.calstate.edu/phab_sopr6.pdf

Office of Environmental Health Hazard Assessment (OEHHA). 2006. Draft development of guidance tissue levels and screening values for common contaminants in California Sports Fish: Chlordane, DDTs, Dieldrin, Methylmercury, PCBs, Selenium, and Toxaphene. OEHHA. Sacramento, CA.

Omernik, J. M. 1987. Ecoregions of the conterminous United States. Map (scale 1:7,500,000). Annals of the Association of American Geographers 77:118–125.

Olsen, A.R., J. Sedransk, D. Edwards, C.A. Gotway, W. Liggett, S. Rathburn, K.H. Reckhow and L.J. Young. 1999. Statistical issues for monitoring ecological and natural resources in the United States. *Environmental Management and Assessment* 54:1-45.

Power, M.E., R.J. Stout, C.E. Cushing, P.P. Harper, F.R. Hauer, W.J. Matthews, P.B. Moyle, B. Statzner, and I.R. Wais De Badgen. 1988. Biotic and abiotic controls in river and stream communities. Journal of the North American Benthological Society. 7: 456-479.

Puckett, M. 2002. Quality Assurance Management Plan for the State of California's Surface Water Ambient Monitoring Program: Version 2. California Department of Fish and Game, Monterey, CA. Prepared for the State Water Resources Control Board. Sacramento, CA.

California Regional Water Quality Control Board, San Diego Region. 1994. Water quality control plan for the San Diego Region. San Diego, CA. http://www.waterboards.ca.gov/sandiego/programs/basinplan.html

SANDAG. 1998. Watersheds of the San Diego Region. SANDAG INFO.

Sandin, L. and R.K. Johnson. 2000. The statistical power of selected indicator metrics using macroinvertebrates for assessing acidification and eutrophication of running waters. *Hydrobiologia* 422/423:233-243.

Schiff, K. and P. Ode. In prep. Recommendations for the development and maintenance of a reference condition management program (RCMP) to support biological assessment of California's wadeable streams. Report to the State Water Resources Control Board's Surface Water Ambient Monitoring Program (SWAMP). Southern California Coastal Water Research Project. Costa Mesa, CA.

Stevans, Jr., D.L. and A.R. Olsen. 2004. Spatially balanced sampling of natural resources. *Journal of the American Statistical Association: Theory and Methods* 99:262-278.

Stormwater Monitoring Coalition Bioassessment Workgroup (SMC). 2007. Regional Monitoring of Southern California's Coastal Watersheds. Southern California Coastal Water Research Project. Costa Mesa, CA. Available from: <u>ftp://ftp.sccwrp.org/pub/download/PDFs/539_SMCworkplan.pdf</u>

Taylor, S.L., S.C. Roberts, C.J. Walsh, and B.E. Hatt. 2004. Catchment urbanisation and increased benthic algal biomass in streams: linking mechanisms to management. *Freshwater Biology* 49: 835-851.

Van Sickle, J., J.L. Stoddard, S.G. Paulsen and A.R. Olsen. 2006. Using relative risk of aquatic stressors at a regional scale. *Environmental Management* 38: 1020-1030.

Walsh, C.J., T.D. Fletcher, and A.R. Ladson. 2005. Stream restoration in urban catchments through redesigning stormwater systems: looking to the catchment to save the stream. *Journal of the North American Benthological Society* 690-705.

Walsh, C.J., K.A. Waller, J. Gehling, and R. MacNally. 2007. Riverine invertebrate assemblages are degraded more by catchment urbanisation than by riparian deforestation. *Freshwater Biology* 52: 574-587.

Weigel, B.M. and D.M. Robertson. 2007. Identifying biotic integrity and water chemistry relations in nonwadeable rivers of wisconsin: Toward the development of nutrient criteria. *Environmental Management* 40: 691-708.

Weston Solutions, Inc. 2007. San Diego County Municipal Copermittees 2005-2006 Urban Runoff Monitoring. Final Report. County of San Diego. San Diego, CA. Available at http://www.projectcleanwater.org/html/wg_monitoring_05-06report.html.

7. APPENDICES

APPENDIX

Water quality constituents at each watersh	ed. SD =	standard devi	ation. n =	number of	i saı	mples.	
			Sa	n Juan		Santa	Margarita
Physical water quality and inorganics	Symbol	Units	Mean	SD	n	Mean	SD n
Alkalinity as CaCO3	Alk	mg/l	208	109	39	187	49 21
Chloride		mg/l			0		0
Fine-ASTM		%			0		0
Fine-ASTM, Passing No. 200 Sieve		%	24.9	24.6	25	6.5	7.0 12
Oxygen, Dissolved	DO	mg/L			0		0
Oxygen, Saturation		%	105	33	38	99	25 20
рН		рН	7.6	1.5	38	7.4	0.8 20
Salinity	Sal	ppt	1.05	1.77	32	2.62	6.16 20
Specific conductivity	Cond	µS/cm	2032	2839	38	4399	9779 20
Sulfate	SO_4	mg/l	497	491	39	352	398 21
Suspended Sediment Concentration		%	70.5	117.2	8		0
Temperature		°C	16.8	3.2	38	16.1	4.1 20
Total Organic Carbon		mg/L			0		0
Total Suspended Solids		mg/L	101	241	31	25	48 21
Turbidity		NTU	35.5	76.8	38	33.1	101.0 20
Velocity		ft/s	0.4	0.9	39	0.5	0.9 21
Nutrients							
Ammonia as N	NH ₃ -N	mg/l	0.19	0.52	39	0.02	0.05 21
Nitrate + Nitrite as N		mg/l	0.42	0.52	39	4.86	4.52 21
Nitrate as N		mg/l	0.41	0.51	39	4.84	4.52 21
Nitrate as NO3	NO ₃	mg/l	1.79	2.20	39	22.48	19.92 20
Nitrite as N		ma/l	0.02	0.02	39	0.02	0.01 21
Nitrogen, Total Kieldahl		ma/l	0.80	0.90	39	0.67	0.72 21
OrthoPhosphate as P		mg/l	3.45	20.80	39	2.30	9.79 21
Phosphorus as P.Total	TP	mg/l	0.22	0.27	39	0.21	0.22 21
Metals		U					
Aluminum		µg/L	3.3	8.6	39	4.5	13.5 21
Arsenic	As	µg/L	3.4	2.5	39	2.5	3.9 21
Cadmium	Cd	µg/L	0.26	0.34	39	0.04	0.03 21
Chromium	Cr	µg/L	0.25	0.22	39	0.23	0.35 21
Copper	Cu	µg/L	4.05	2.85	39	3.10	2.37 21
Lead	Pb	µg/L	0.02	0.02	39	0.01	0.01 21
Manganese	Mn	µg/L	148	329	39	92	139 21
Nickel	Ni	µg/L	5.55	6.76	39	0.71	1.22 21
Selenium	Se	µg/L	7.5	10.4	38	5.9	16.8 20
Silver	Ag	µg/L	0.20	1.22	39	0.09	0.29 21
Zinc	Zi	µg/L	4.1	3.1	38	2.3	1.5 20
Bacteria							
Enterococcus		MPN/100 ml	70		1	10	1
Fecal Coliform		MPN/100 ml	170		1	50	1
Total Coliform		MPN/100 ml	190		1	900	1

Appendix, continued.		San	Juan		Santa	Margarita
PAHs	Symbol Units	Mean S	D	n	Mean	SD n
Acenaphthene	ng/L	0	0	39	0	0 21
Acenaphthylene	ng/L	0	0	39	0	0 21
Anthracene	ng/L	0	0	39	0	0 21
Benz(a)anthracene	ng/L	0	0	39	0	0 21
Benzo(a)pyrene	ng/L	1.7	10.8	39	0	0 21
Benzo(b)fluoranthene	ng/L	3.5	9.6	39	1.0	3.2 21
Benzo(e)pyrene	ng/L	1.3	6.3	39	0	0 21
Benzo(g,h,i)perylene	ng/L	4.3	14.4	39	0	0 21
Benzo(k)fluoranthene	ng/L	1.4	8.9	38	0	0 20
Biphenyl	ng/L	0	0	38	0	0 20
Chrysene	ng/L	0.8	3.4	39	0	0 21
Chrysenes, C1 -	ng/L	1.9	7.4	38	0	0 20
Chrysenes, C2 -	ng/L	2.7	9.3	38	0	0 20
Chrysenes, C3 -	ng/L	19.6	111.4	38	0	0 20
Dibenz(a,h)anthracene	ng/L	2.4	15.1	39	0	0 21
Dibenzothiophene	ng/L	1.7	6.4	38	0	0 20
Dibenzothiophenes, C1 -	ng/L	10.4	21.1	38	5.4	8.4 20
Dibenzothiophenes, C2 -	ng/L	18.1	38.9	38	10.5	12.9 20
Dibenzothiophenes, C3 -	ng/L	9.4	25.1	38	2.3	7.1 20
Dimethylnaphthalene, 2,6-	ng/L	0	0	38	0	0 20
Dimethylphenanthrene, 3,6-	ng/L			0		0
Fluoranthene	ng/L	0.82	3.74	39	0	0 21
Fluoranthene/Pyrenes, C1 -	ng/L	0.39	2.43	38	0	0 20
Fluorene	ng/L	0	0	39	0	0 21
Fluorenes, C1 -	ng/L	2.06	6.06	38	0.57	2.55 20
Fluorenes, C2 -	ng/L	0.93	3.26	38	0	0 20
Fluorenes, C3 -	ng/L	5.89	13.27	38	4.27	12.54 20
Indeno(1,2,3-c,d)pyrene	ng/L	3.79	17.96	39	0	0 21
Methyldibenzothiophene, 4-	ng/L			0		0
Methylfluoranthene, 2-	ng/L			0		0
Methylfluorene, 1-	ng/L			0		0
Methylnaphthalene, 1-	ng/L	0	0	38	0	0 20
Methylnaphthalene, 2-	ng/L	0	0	38	0	0 20
Methylphenanthrene, 1-	ng/L	0	0	38	0	0 20
Naphthalene	ng/L	1.34	6.42	39	0	0 21
Naphthalenes, C1 -	ng/L	1.30	6.53	38	0	0 20
Naphthalenes, C2 -	ng/L	1.17	4.07	38	0	0 20
Naphthalenes, C3 -	ng/L	3.65	8.57	38	1.70	5.27 20
Naphthalenes, C4 -	ng/L	7.44	21.33	38	0	0 20
Perylene	ng/L	1.83	8.04	38	0	0 20
Phenanthrene	ng/L	0	0	39	0	0 21
Phenanthrene/Anthracene, C1 -	ng/L	4.99	9.68	38	5.01	8.55 20
Phenanthrene/Anthracene, C2 -	ng/L	2.57	7.07	38	0.50	2.24 20
Phenanthrene/Anthracene, C3 -	ng/L	2.81	8.01	38	0	0 20
Phenanthrene/Anthracene, C4 -	ng/L	0.44	2.74	38	0	0 20
Pyrene	ng/L	2.65	8.10	39	0	0 21
Trimethylnaphthalene, 2,3,5-	ng/L	0	0	38	0	0 20

		San	Juan		Santa Mar	garita
PCBs	Symbol Units	Mean S	D	n	Mean SD	- <u>n</u>
PCB 005	ng/L	0.35	1.71	38	0	0 20
PCB 008	ng/L	0.41	2.50	38	0	0 20
PCB 015	ng/L	0	0	38	0	0 20
PCB 018	ng/L	0	0	38	0	0 20
PCB 027	ng/L	0	0	38	0	0 20
PCB 028	ng/L	0	0	38	0	0 20
PCB 029	ng/L	0	0	38	0	0 20
PCB 031	ng/L	0.11	0.65	38	0	0 20
PCB 033	ng/L	0	0	38	0	0 20
PCB 044	ng/L	0	0	38	0	0 20
PCB 049	ng/L	0	0	38	0	0 20
PCB 052	ng/L	0.32	1.95	38	0	0 20
PCB 056	ng/L	0	0	38	0	0 20
PCB 060	ng/L	0	0	38	0	0 20
PCB 066	ng/L	0	0	38	0	0 20
PCB 070	ng/L	0	0	38	0	0 20
PCB 074	ng/L	0	0	38	0	0 20
PCB 087	ng/L	0.82	2.01	38	0	0 20
PCB 095	ng/L	0	0	38	0	0 20
PCB 097	ng/L	0	0	38	0	0 20
PCB 099	ng/L	0	0	38	0	0 20
PCB 101	ng/L	0	0	38	0	0 20
PCB 105	ng/L	0	0	38	0	0 20
PCB 110	ng/L	0	0	38	0	0 20
PCB 114	ng/L	0	0	38	0	0 20
PCB 118	ng/L	0	0	38	0	0 20
PCB 128	ng/L	0	0	38	0	0 20
PCB 137	ng/L	0	0	38	0	0 20
PCB 138	ng/L	0	0	38	0	0 20
PCB 141	ng/L	0	0	38	0	0 20
PCB 149	ng/L	0	0	38	0	0 20
PCB 151	ng/L	0	0	38	0	0 20
PCB 153	ng/L	0	0	38	0	0 20
PCB 156	ng/L	0	0	38	0	0 20
PCB 157	ng/L	0	0	38	0	0 20
PCB 158	ng/L	0	0	38	0	0 20
PCB 170	ng/L	0	0	38	0	0 20
PCB 174	ng/L	0	0	38	0	0 20
PCB 177	ng/L	0	0	38	0	0 20
PCB 180	ng/L	0	0	38	0	0 20
PCB 183	ng/L	0	0	38	0	0 20
PCB 187	ng/L	0.16	0.72	38	0	0 20
PCB 189	ng/L	0	0	38	0	0 20
PCB 194	ng/L	0.18	1.14	38	0	0 20
PCB 195	ng/L	0.18	1.14	38	0	0 20
PCB 200	ng/L	0	0	38	0	0 20
PCB 201	ng/L	0	0	38	0	0 20
PCB 203	ng/L	0	0	38	0	0 20
PCB 206	ng/L	0	0	38	0	0 20

PCBs Symbol Units Mean SD n Mean SD n PCB-1016 ng/L 0 0 38 0 20 PCB-1016 ng/L 0 1 0 1 0 1 PCB-1221 ng/L 0 1 0 1 0 1 PCB-1224 ng/L 0 1 0 1 0 1 PCB-1254 ng/L 0 1 0 1 0 1 PCB-1254 ng/L 0 1 0 1 0 1 PCBs ng/L 0 0 38 0 0 20 Aldrin ng/L 0 0 38 0 0 20 Atraton ng/L 0 0 38 0 0 20 Atraton ng/L 0 0 38 0 20 20 Atraton	· · · · · · · · · · · · · · · · · · ·		San Juan			Santa Margarita		
PCB 209 ng/L 0 0 38 0 0 20 PCB-1016 ng/L 0 1 0 1 PCB-1221 ng/L 0 1 0 1 PCB-1232 ng/L 0 1 0 1 PCB-1242 ng/L 0 1 0 1 PCB-1248 ng/L 0 1 0 1 PCB-1260 ng/L 0 0 38 0 0 20 Attaron ng/L 0 0 38 0 0 20 Attaron ng/L 0 0 38 0 0 20 Attaron ng/L 0 0 38 0 0 20 At	PCBs	Symbol Units	Mean S	SD	n	Mean	SD n	
PCB-1016 ng/L 0 1 0 1 PCB-1221 ng/L 0 1 0 1 PCB-1232 ng/L 0 1 0 1 PCB-1242 ng/L 0 1 0 1 PCB-1248 ng/L 0 1 0 1 PCB-1254 ng/L 0 1 0 1 PCB-1254 ng/L 0 1 0 1 PCB-1254 ng/L 0 3 0 0 2 Aldrin ng/L 0 0 38 0 0 20 Atrazine ng/L 0 0 38 0 0 20 Azinphos enthyl ng/L 0 0 38 0 0 20 Azinphos methyl ng/L 0 0 38 0 0 0 DelataBHC ng/L 0 0 38 0	PCB 209	ng/L	0	0	38	0	0 20	
PCB-1221 ng/L 0 1 0 1 PCB-1232 ng/L 0 1 0 1 PCB-1242 ng/L 0 1 0 1 PCB-1248 ng/L 0 1 0 1 PCB-1254 ng/L 0 1 0 1 PCB-1260 ng/L 0 3 0 0 2 Pesticides ng/L 0 38 0 0 2 Aldrin ng/L 0 0 38 0 2 0 Aspon ng/L 0 0 38 0 2 0 Atraton ng/L 0 0 38 0 2 0 Azinphos methyl ng/L 0 0 38 0 2 0 Azinphos methyl ng/L 0 1 0 1 0 1 Bolstar ng/L 0.38	PCB-1016	ng/L	0		1	0	1	
PCB-1232 ng/L 0 1 0 1 PCB-1242 ng/L 0 1 0 1 PCB-1248 ng/L 0 1 0 1 PCB-1254 ng/L 0 1 0 1 PCB-1260 ng/L 0 1 0 1 PCB-1260 ng/L 0 1 0 1 Pesticides ng/L 0 0 38 0 0 21 alpha-BHC ng/L 0 0 38 0 0 20 Atraton ng/L 0 0 38 0 0 20 Azinphos ethyl ng/L 0 0 38 0 0 20 DatashCane, tersh- ng/L 0 0 38 0 0 1 Chiordene, tersh- ng/L 0 1 0 1 0 1 Chiordene, agamma- ng/L <td>PCB-1221</td> <td>ng/L</td> <td>0</td> <td></td> <td>1</td> <td>0</td> <td>1</td>	PCB-1221	ng/L	0		1	0	1	
PCB-1242 ng/L 0 1 0 1 PCB-1248 ng/L 0 1 0 1 PCB-1260 ng/L 0 1 0 1 PCB-1260 ng/L 0 1 0 1 Pesticides ng/L 0 1 0 1 Aldrin ng/L 0 0 38 0 0 21 Aldrin ng/L 0 0 38 0 20 Atraton ng/L 0 0 38 0 20 Atraton ng/L 0 0 38 0 20 Atrazine ng/L 0 0 38 0 20 Atraton ng/L 0 1 0 1 1 1 1 Bolstar ng/L 0 1 0 1 0 1 1 1 1 1 1 1 1 1 <td< td=""><td>PCB-1232</td><td>ng/L</td><td>0</td><td></td><td>1</td><td>0</td><td>1</td></td<>	PCB-1232	ng/L	0		1	0	1	
PCB-1248 ng/L 0 1 0 1 PCB-1260 ng/L 0 1 0 1 PCB-1260 ng/L 2.45 6.68 39 0 0 21 Pesticides ng/L 0 0 39 0 0 21 Aldrin ng/L 0 0 38 0 20 Aspon ng/L 0 0 38 0 20 Atrazine ng/L 0 0 38 0 20 Azinphos ethyl ng/L 0 0 38 0 20 Azinphos methyl ng/L 0 0 38 0 20 Chiordane (tech) ng/L 0 1 0 1 0 1 Chiordene, trans ng/L 0 1 0 1 0 1 Chiordene, trans ng/L 0.03 0.66 8 0 20	PCB-1242	ng/L	0		1	0	1	
PCB-1254 ng/L 0 1 0 1 PCB-1260 ng/L 2.45 6.68 39 0 0 2 Pesticides 1 0 1 0 1 Aldrin ng/L 0 1 0 1 0 1 Ametryn ng/L 0 0 38 0 20 Aspon ng/L 0 0 38 0 20 Atrazine ng/L 0 0 38 0 20 Azinphos ethyl ng/L 0 0 38 0 20 Azinphos methyl ng/L 0 0 38 0 20 Chordane, (tsc. ng/L 0 1 0 1 0 1 Bolstar ng/L 0 38 0 20 Chordene, alpha- ng/L 0 38 0 20 Chlordene, alpha- ng/L<	PCB-1248	ng/L	0		1	0	1	
PCB-1260 ng/L 0 1 0 1 Pectsicides ng/L 2.45 6.68 39 0 0.21 Aldrin ng/L 0 0 39 0 0.21 Aldrin ng/L 0 0 38 0 0.20 Ametryn ng/L 0 0 38 0 0.20 Asspon ng/L 0 0 38 0 0.20 Atration ng/L 0 0 38 0 0.20 Atrazine ng/L 0 0 38 0 0.20 Azinphos methyl ng/L 0 0 38 0 0.20 Azinphos methyl ng/L 0 0 38 0 0.20 Catophenothion ng/L 0 1 0 1 0 1 Chlordane, (cis- ng/L 0.03 0.16 38 0 20 Chlordane, gamma- ng/L 0.03 0.16 38 0 20 <th< td=""><td>PCB-1254</td><td>ng/L</td><td>0</td><td></td><td>1</td><td>0</td><td>1</td></th<>	PCB-1254	ng/L	0		1	0	1	
PCBs ng/L 2.45 6.68 39 0 0 21 Pesticides ng/L 0 0 39 0 0 21 alpha-BHC ng/L 0 1 0 1 0 1 Ametryn ng/L 0 0 38 0 20 Arazine ng/L 0 0 38 0 20 Azinphos ethyl ng/L 0 0 38 0 20 Azinphos methyl ng/L 0 0 38 0 20 Azinphos methyl ng/L 0 0 38 0 20 Carbophenothion ng/L 0 0 38 0 20 Chlordane, (is- ng/L 0.33 0.16 38 0 20 Chlordane, trans- ng/L 0.33 0.16 38 0 20 Chlordane, trans- ng/L 0 38 0	PCB-1260	ng/L	0		1	0	1	
Pesticides Aldrin ng/L 0 0 39 0 0 21 alpha-BHC ng/L 0 0 38 0 0 20 Arston ng/L 0 0 38 0 0 20 Atraton ng/L 0 0 38 0 0 20 Atrazine ng/L 0 0 38 0 0 20 Azinphos ethyl ng/L 0 0 38 0 0 20 Azinphos methyl ng/L 0 0 38 0 0 20 Azinphos methyl ng/L 0 1	PCBs	ng/L	2.45	6.68	39	0	0 21	
Aldrinng/L00390001alpha-BHCng/L010101Ametrynng/L00380020Asponng/L00380020Atratonng/L00380020Atratonng/L00380020Azinphos ethylng/L00380020Azinphos methylng/L0038020Deta-BHCng/L01011Bolstarng/L0038020Chlordane, (tech)ng/L01011Chlordane, trans-ng/L0.030.1638020Chlordene, alpha-ng/L0038020Chlordene, alpha-ng/L0038020Chlordene, alpha-ng/L0038020Chlordene, alpha-ng/L0038020Chlordene, alpha-ng/L0038020Chlordene, alpha-ng/L0038020Chlordene, alpha-ng/L0038020Chlordene, alpha-ng/L0038020DDD(o,p')ng/L0 <td>Pesticides</td> <td></td> <td>_</td> <td>_</td> <td></td> <td></td> <td></td>	Pesticides		_	_				
alpha-BHC ng/L 0101Ametryn ng/L 00380020Araton ng/L 00380020Atrazine ng/L 00380020Azinphos ethyl ng/L 00380020Azinphos methyl ng/L 00380020Azinphos methyl ng/L 00380020Carbophenothion ng/L 00380020Carbophenothion ng/L 00380020Chlordane, cis- ng/L 00380020Chlordane, trans- ng/L 0.030.1638020Chlordene, aghpa- ng/L 0.030.1638020Chlordene, aghpa- ng/L 0038020Chlordene, aghpa- ng/L 0038020Chlordene, aghma- ng/L 0038020Chlorpyrifos ng/L 0038020Chlorpyrifos ng/L 0038020Chlorpyrifos methyl ng/L 0038020DDD(o,p') ng/L 0038020DDD(o,p') ng/L 0.030.1639 <t< td=""><td>Aldrin</td><td>ng/L</td><td>0</td><td>0</td><td>39</td><td>0</td><td>0 21</td></t<>	Aldrin	ng/L	0	0	39	0	0 21	
Ametryn ng/L 0038000Aspon ng/L 00380020Atraton ng/L 00380020Atrazine ng/L 00380020Azinphos ethyl ng/L 00380020Azinphos methyl ng/L 00380020beta-BHC ng/L 010101Bolstar ng/L 00380020Chordane (tech) ng/L 010101Chlordane, trans- ng/L 0.291.4938020Chlordane, trans- ng/L 0.3802020Chlordene, talpha- ng/L 0.341.1638020Chlordene, tans- ng/L 038020Chlordene, tans ng/L 038 </td <td>alpha-BHC</td> <td>ng/L</td> <td>0</td> <td>_</td> <td>1</td> <td>0</td> <td>1</td>	alpha-BHC	ng/L	0	_	1	0	1	
Aspon ng/L 0 0 38 0 0 20 Atraton ng/L 0 0 38 0 0 20 Atraton ng/L 0 0 38 0 0 20 Azinphos ethyl ng/L 0 0 38 0 0 20 Azinphos methyl ng/L 0 0 38 0 0 20 beta-BHC ng/L 0 0 38 0 0 20 Carbophenothion ng/L 0.29 1.49 38 0 0 20 Chlordane, trans- ng/L 0.29 1.49 38 0 20 20 Chlordane, trans- ng/L 0.03 0.16 38 0 20	Ametryn	ng/L	0	0	38	0	0 20	
Arraton ng/L 0 0 38 0 0 20 Atrazine ng/L 0 0 38 0 0 20 Azinphos ethyl ng/L 0 0 38 0 0 20 Azinphos methyl ng/L 0 0 38 0 0 20 beta-BHC ng/L 0 0 38 0 0 20 Carbophenothion ng/L 0 0 38 0 0 20 Chlordane (tech) ng/L 0 1 0 1 1 1 1 1 1 1 1 1 1 1 1 1 1 0 20 Chlordene, gamma- ng/L 0.03 0 20 1 1 1 1 1 1 1 1 0 20 20 20 20 20 20 20 20 20 20 <td< td=""><td>Aspon</td><td>ng/L</td><td>0</td><td>0</td><td>38</td><td>0</td><td>0 20</td></td<>	Aspon	ng/L	0	0	38	0	0 20	
Arrazine ng/L 0 0 38 0 0 20 Azinphos ethyl ng/L 0 0 38 0 0 20 Azinphos methyl ng/L 0 0 38 0 0 20 beta-BHC ng/L 0 1 0 1 0 1 Bolstar ng/L 0 0 38 0 0 20 Carbophenothion ng/L 0 1 0 1 0 1 Chlordane, (tech) ng/L 0.29 1.49 38 0 0 20 Chlordene, alpha- ng/L 0.33 0.16 38 0 0 20 Chlordene, gamma- ng/L 0.34 1.16 38 0 0 20 Chlorpyrifos ng/L 0 0 38 0 0 20 Colorphyrifos methyl ng/L 0 0 38 0	Atraton	ng/L	0	0	38	0	0 20	
Azinphos ethyl ng/L 0 0 38 0 0 20 Azinphos methyl ng/L 0 0 38 0 0 20 beta-BHC ng/L 0 0 38 0 0 20 Carbophenothion ng/L 0 0 38 0 0 20 Chlordane (tech) ng/L 0.29 1.49 38 0 0 20 Chlordane, trans- ng/L 0.33 0.16 38 0 0 20 Chlordene, agmma- ng/L 0.34 1.16 38 0 0 20 Chlordene, gamma- ng/L 0 0 38 0 0 20 Chlordene, gamma- ng/L 0 0 38 0 0 20 Chlordene, gamma- ng/L 0 0 38 0 0 20 Chlordene, gamma- ng/L 0 0 38 0 0 20 Chlordene, gamma- ng/L 0 0	Atrazine	ng/L	0	0	38	0	0 20	
Azinphos methyl ng/L 0 0 38 0 0 20 beta-BHC ng/L 0 1 0 1 0 1 Bolstar ng/L 0 0 38 0 0 20 Carbophenothion ng/L 0 0 38 0 0 20 Chlordane (tech) ng/L 0.29 1.49 38 0 0 20 Chlordane, trans- ng/L 0.03 0.16 38 0 0 20 Chlordene, agamma- ng/L 0.34 1.16 38 0 0 20 Chlordene, gamma- ng/L 0 0 38 0 0 20 Chlordpyrifos ng/L 0 0 38 0 0 20 Chlordpyrifos methyl ng/L 0 0 38 0 0 20 Dod(o,p') ng/L 0.24 0.71 38 <t< td=""><td>Azinphos ethyl</td><td>ng/L</td><td>0</td><td>0</td><td>38</td><td>0</td><td>0 20</td></t<>	Azinphos ethyl	ng/L	0	0	38	0	0 20	
beta-BHC ng/L 0101Bolstar ng/L 0038020Carbophenothion ng/L 01011Chlordane, (tech) ng/L 0.291.4938020Chlordane, trans- ng/L 0.030.1638020Chlordene, alpha- ng/L 0.330.1638020Chlordene, agamma- ng/L 0.341.1638020Chlordene, agamma- ng/L 038020Chlordene, agamma- ng/L 0038020Chlordene, agamma- ng/L 0038020Chlordprifos ng/L 0038020Chlordprifos methyl ng/L 0038020Coumaphos ng/L 0038020DDD(o,p') ng/L 0.030.16390.2020DDDL(o,p') ng/L 0.030.1638020DDDL(o,p') ng/L 0.030.1638020DDDL(o,p') ng/L 0.030.16390.050.22DDDL(o,p') ng/L 0.0380020DDL(o,p') ng/L 0.0380020DDT(o,p') ng/L 0.230.83390.431.5721DDTs <t< td=""><td>Azinphos methyl</td><td>ng/L</td><td>0</td><td>0</td><td>38</td><td>0</td><td>0 20</td></t<>	Azinphos methyl	ng/L	0	0	38	0	0 20	
Boistar ng/L 0 0 38 0 0 20 Carbophenothin ng/L 0 0 38 0 0 20 Chlordane (tech) ng/L 0.29 1.49 38 0 20 Chlordane, trans- ng/L 0.33 0.16 38 0 20 Chlordene, alpha- ng/L 0.34 1.16 38 0 20 Chlordene, gamma- ng/L 0.38 0 20 Chlordene, gamma- ng/L 0 0 38 0 20 Chlorpyrifos ng/L 0 0 38 0 20 Chlorpyrifos 0 20	beta-BHC	ng/L	0	-	1	0	1	
Carbophenothionng/L000380020Chlordane (tech)ng/L01011Chlordane, cis-ng/L0.291.49380020Chlordane, trans-ng/L00380020Chlordene, alpha-ng/L00380020Chlordene, gamma-ng/L0.341.1638020Chlordene, gamma-ng/L0038020Chlorpyrifosng/L0038020Chlorpyrifosng/L0038020Coumaphosng/L0038020DDD(o,p')ng/L0038020DDD(o,p')ng/L0038020DDD(o,p')ng/L0.030.1638020DDD(o,p')ng/L0.030.1638020DDD(o,p')ng/L0.030.1638020DDD(o,p')ng/L0.030.1638020DDD(p,p')ng/L0.030.1638020DDT(o,p')ng/L0.0338020DDT(o,p')ng/L038020DDT(p,p')ng/L038020DDT(p,p')ng/L038020	Bolstar	ng/L	0	0	38	0	0 20	
Chlordane (tech)ng/L0101Chlordane, cis-ng/L0.291.49380020Chlordane, trans-ng/L0.030.1638020Chlordene, alpha-ng/L0.341.1638020Chlordene, gamma-ng/L0.341.1638020Chlordene, gamma-ng/L0038020Chlordene, spanna-ng/L0038020Chlordene, spanna-ng/L0038020Chlordene, spanna-ng/L0038020Chlordene, spanna-ng/L0038020Chlordene, spanna-ng/L0038020Chlordene, spanna-ng/L0038020Chlordene, spanna-ng/L0038020Chlordene, spanna-ng/L0038020Chlordene, spanna-ng/L0038020Dob(o,p')ng/L0.240.7138020DDD(o,p')ng/L0.030.16390.902.64DDE(o,p')ng/L0.030.16390.431.57DDT(o,p')ng/L0380020DDT(o,p')ng/L0380020DDT(o,p') <td>Carbophenothion</td> <td>ng/L</td> <td>0</td> <td>0</td> <td>38</td> <td>0</td> <td>0 20</td>	Carbophenothion	ng/L	0	0	38	0	0 20	
Chlordane, cis- ng/L 0.29 1.49 38 0 0 20 Chlordane, trans- ng/L 0.03 0.16 38 0 0 20 Chlordene, alpha- ng/L 0.38 0 0 20 Chlordene, gamma- ng/L 0.34 1.16 38 0 20 Chlordpryifos ng/L 0 0 38 0 20 Chlordpyrifos methyl ng/L 0 0 38 0 20 Coumaphos ng/L 0 0 38 0 20 DDD(o,p') ng/L 0.24 0.71 38 0 20 DDD(o,p') ng/L 0.38 0 20<	Chlordane (tech)	ng/L	0		1	0	1	
Chlordane, trans- ng/L 0.03 0.16 38 0 0 20 Chlordene, alpha- ng/L 0.34 1.16 38 0 0 20 Chlordene, gamma- ng/L 0.34 1.16 38 0 0 20 Chlordenvinphos ng/L 0 0 38 0 0 20 Chlordenyrifos ng/L 0 0 38 0 0 20 Chlordenyrifos ng/L 0 0 38 0 0 20 Chlorpyrifos ng/L 0 0 38 0 0 20 Couraphos ng/L 0.24 0.71 38 0 0 20 DDD(o,p') ng/L 0.03 0.16 39 0.05 0.22 21 DDE(o,p') ng/L 0.03 0.16 39 0.05 0.22 21 DDE(o,p') ng/L 0.03 0.16 <td< td=""><td>Chlordane, cis-</td><td>ng/L</td><td>0.29</td><td>1.49</td><td>38</td><td>0</td><td>0 20</td></td<>	Chlordane, cis-	ng/L	0.29	1.49	38	0	0 20	
Chlordene, alpha- ng/L 0 0 38 0 0 20 Chlordene, gamma- ng/L 0.34 1.16 38 0 0 20 Chlordene, gamma- ng/L 0 0 38 0 0 20 Chlorpyrifos ng/L 0 0 38 0 0 20 Chlorpyrifos methyl ng/L 0 0 38 0 0 20 Coumaphos ng/L 0 0 38 0 0 20 Dacthal ng/L 0.24 0.71 38 0 0 20 DDD(o,p') ng/L 0.03 0.16 39 0.05 0.22 21 DDE(o,p') ng/L 0.03 0.16 39 0.02 20 20 DDE(o,p') ng/L 0.03 0.16 39 0.02 20 DDT(o,p') ng/L 0.23 0.83 39 0.43	Chlordane, trans-	ng/L	0.03	0.16	38	0	0 20	
Chlordene, gamma- ng/L 0.34 1.16 38 0 0 20 Chlorfenvinphos ng/L 0 0 38 0 0 20 Chlorpyrifos ng/L 0 0 38 0 0 20 Chlorpyrifos methyl ng/L 0 0 38 0 0 20 Ciodrin ng/L 0 0 38 0 0 20 Coumaphos ng/L 0 0 38 0 0 20 Dacthal ng/L 0.24 0.71 38 0 0 20 DDD(o,p') ng/L 0.03 0.16 39 0.05 0.22 21 DDE(o,p') ng/L 0.03 0.16 38 0 0 20 DDE(p,p') ng/L 0.03 0.16 38 0 0 20 DDE(p,p') ng/L 0.23 0.83 39 0.43 1.57 21 DDTs ng/L 0.58 1.28 39 <t< td=""><td>Chlordene, alpha-</td><td>ng/L</td><td>0</td><td>0</td><td>38</td><td>0</td><td>0 20</td></t<>	Chlordene, alpha-	ng/L	0	0	38	0	0 20	
Chlorfenvinphos ng/L 0 0 38 0 0 20 Chlorpyrifos ng/L 0 0 38 0 0 20 Chlorpyrifos methyl ng/L 0 0 38 0 0 20 Ciodrin ng/L 0 0 38 0 0 20 Coumaphos ng/L 0 0 38 0 0 20 Dacthal ng/L 0.24 0.71 38 0 0 20 DDD(o,p') ng/L 0.03 0.16 39 0.05 0.22 21 DDE(o,p') ng/L 0.03 0.16 38 0 0 20 DDE(o,p') ng/L 0.03 0.16 38 0 0 20 DDE(o,p') ng/L 0.23 0.66 39 0.90 2.64 21 DDMU(p,p') ng/L 0.23 0.83 0 0	Chlordene, gamma-	ng/L	0.34	1.16	38	0	0 20	
Chlorpyrifos ng/L 0 0 38 0 0 20 Chlorpyrifos methyl ng/L 0 0 38 0 0 20 Ciodrin ng/L 0 0 38 0 0 20 Coumaphos ng/L 0 0 38 0 0 20 Dacthal ng/L 0.24 0.71 38 0 0 20 DDD(o,p') ng/L 0.33 0.16 39 0.05 0.22 21 DDE(o,p') ng/L 0.03 0.16 39 0.05 0.22 21 DDE(o,p') ng/L 0.03 0.16 39 0.90 2.64 21 DDMU(p,p') ng/L 0.29 0.66 39 0.90 2.64 21 DDT(o,p') ng/L 0.23 0.83 0 0 20 DDT(p,p') ng/L 0.23 0.83 0 0 20	Chiortenvinphos	ng/L	0	0	38	0	0 20	
Chiorpyritos metnyi ng/L 0 0 38 0 0 20 Ciodrin ng/L 0 0 38 0 0 20 Coumaphos ng/L 0 0 38 0 0 20 Dacthal ng/L 0.24 0.71 38 0 0 20 DDD(o,p') ng/L 0.03 0.16 39 0.05 0.22 21 DDE(o,p') ng/L 0.03 0.16 38 0 0 20 DDE(p,p') ng/L 0.03 0.16 38 0 0 20 DDE(p,p') ng/L 0.29 0.66 39 0.90 2.64 21 DDMU(p,p') ng/L 0 0 38 0 0 20 DDT(o,p') ng/L 0 0 38 0.10 0.45 20 DDT(p,p') ng/L 0.58 1.28 39 1.48	Chiorpyritos	ng/L	0	0	38	0	0 20	
Cloarin ng/L 0 0 38 0 0 20 Coumaphos ng/L 0 0 38 0 0 20 Dacthal ng/L 0.24 0.71 38 0 0 20 DDD(o,p') ng/L 0.03 0.16 39 0.05 0.22 21 DDE(o,p') ng/L 0.03 0.16 38 0 0 20 DDE(o,p') ng/L 0.03 0.16 38 0 0 20 DDE(o,p') ng/L 0.03 0.16 38 0 0 20 DDE(p,p') ng/L 0.29 0.66 39 0.90 2.64 21 DDMU(p,p') ng/L 0 0 38 0 0 20 DDT(o,p') ng/L 0.23 0.83 39 0.43 1.57 21 DDTs ng/L 0.58 1.28 39 1.48	Chiorpyritos metnyi	ng/L	0	0	38	0	0 20	
Coumapnos ng/L 0 0 38 0 0 20 Dacthal ng/L 0.24 0.71 38 0 0 20 DDD(o,p') ng/L 0.03 0.16 39 0.05 0.22 21 DDE(o,p') ng/L 0.03 0.16 39 0.05 0.22 21 DDE(o,p') ng/L 0.03 0.16 38 0 0 20 DDE(p,p') ng/L 0.03 0.16 38 0 0 20 DDT(o,p') ng/L 0.29 0.66 39 0.90 2.64 21 DDMU(p,p') ng/L 0 0 38 0 0 20 DDT(o,p') ng/L 0 0 38 0 0 20 DDTs ng/L 0.58 1.28 39 1.48 3.89 21 delta-BHC ng/L 0 0 38 0 <	Ciodrin	ng/L	0	0	38	0	0 20	
Dactrial Ig/L 0.24 0.71 36 0 0 20 DDD(o,p') ng/L 0 0 38 0 0 20 DDD(p,p') ng/L 0.03 0.16 39 0.05 0.22 21 DDE(o,p') ng/L 0.03 0.16 38 0 0 20 DDE(o,p') ng/L 0.03 0.16 38 0 0 20 DDE(p,p') ng/L 0.29 0.66 39 0.90 2.64 21 DDMU(p,p') ng/L 0 0 38 0 0 20 DDT(o,p') ng/L 0 0 38 0.10 0.45 20 DDTs ng/L 0.23 0.83 39 0.43 1.57 21 DDTs ng/L 0.58 1.28 39 1.48 3.89 21 delta-BHC ng/L 0 0 38 0	Coumaphos	ng/L	0 24	0 71	38	0	0 20	
DDD(0,p) ng/L 0 0 38 0 0 20 DDD(p,p') ng/L 0.03 0.16 39 0.05 0.22 21 DDE(0,p') ng/L 0.03 0.16 38 0 0 20 DDE(p,p') ng/L 0.29 0.66 39 0.90 2.64 21 DDMU(p,p') ng/L 0 0 38 0 0 20 DDT(o,p') ng/L 0 0 38 0.10 0.45 20 DDT(p,p') ng/L 0.23 0.83 39 0.43 1.57 21 DDTs ng/L 0.58 1.28 39 1.48 3.89 21 delta-BHC ng/L 0 1 0 1		ng/L	0.24	0.71	30 20	0	0 20	
DDD(p,p') ng/L 0.03 0.16 39 0.03 0.22 21 DDE(o,p') ng/L 0.03 0.16 38 0 0 20 DDE(p,p') ng/L 0.29 0.66 39 0.90 2.64 21 DDMU(p,p') ng/L 0 0 38 0 0 20 DDT(o,p') ng/L 0 0 38 0.10 0.45 20 DDT(p,p') ng/L 0.23 0.83 39 0.43 1.57 21 DDTs ng/L 0.58 1.28 39 1.48 3.89 21 delta-BHC ng/L 0 1 0 1 1 Demeton-s ng/L 0 0 38 0 0 20 Dichlofenthion ng/L 0 0 38 0 0 20 Dichlorvos ng/L 0 0 38 0 0 20 Dichlorvos ng/L 0 0 38 0 0	DDD(0,p)	ng/L	0 02	0 16	20	0.05	0 20	
DDE(0,p') ng/L 0.03 0.10 38 0 0 20 DDMU(p,p') ng/L 0 0 38 0 0 20 DDT(o,p') ng/L 0 0 38 0.10 0.45 20 DDT(o,p') ng/L 0 0 38 0.10 0.45 20 DDT(p,p') ng/L 0.23 0.83 39 0.43 1.57 21 DDTs ng/L 0.58 1.28 39 1.48 3.89 21 delta-BHC ng/L 0 1 0 1 1 1 Demeton-s ng/L 0 0 38 0 0 20 Dichlofenthion ng/L 0 0 38 0 0 20 Dichlorvos ng/L 0 0 38 0 0 20 Dichlorvos ng/L 0 0 38 0 0 20 Dichlorvos ng/L 0 0 38 0 0	DDD(p,p)	ng/L	0.03	0.10	20	0.05	0.22 21	
DDE(p,p') ng/L 0.29 0.00 39 2.04 21 DDMU(p,p') ng/L 0 0 38 0 0 20 DDT(o,p') ng/L 0.23 0.83 39 0.43 1.57 21 DDTs ng/L 0.23 0.83 39 0.43 1.57 21 DDTs ng/L 0.58 1.28 39 1.48 3.89 21 delta-BHC ng/L 0 1 0 1 0 1 Demeton-s ng/L 0 38 0 0 20 Diazinon ng/L 43.97 103.77 38 7.78 18.10 20 Dichlofenthion ng/L 0 0 38 0 0 20 Dichlorvos ng/L 0 0 38 0 0 20 Dicrotophos ng/L 0 0 38 0 0 20 Diredtrin ng/L 0.15 0.42 39 0 21 <t< td=""><td>DDE(0,p)</td><td>ng/L</td><td>0.03</td><td>0.10</td><td>20</td><td>0 00</td><td>264 21</td></t<>	DDE(0,p)	ng/L	0.03	0.10	20	0 00	264 21	
DDMO(p,p) ng/L 0 0 38 0 0 20 DDT(o,p') ng/L 0.23 0.83 39 0.43 1.57 21 DDTs ng/L 0.58 1.28 39 1.48 3.89 21 delta-BHC ng/L 0 1 0 1 0 1 Demeton-s ng/L 0 0 38 0 0 20 Diazinon ng/L 43.97 103.77 38 7.78 18.10 20 Dichlofenthion ng/L 0 0 38 0 0 20 Dichlorvos ng/L 0 0 38 0 0 20 Dicrotophos ng/L 0 0 38 0 0 20 Dieldrin ng/L 0 0 38 0 0 20 Dicrotophos ng/L 0 0 38 0 0 20 Dimethoate ng/L 0.15 0.42 39 0 21 <td>DDE(p,p)</td> <td>ng/L</td> <td>0.29</td> <td>0.00</td> <td>20</td> <td>0.90</td> <td>2.04 21</td>	DDE(p,p)	ng/L	0.29	0.00	20	0.90	2.04 21	
DDT(p,p') ng/L 0 0 38 0.10 0.43 20 DDT(p,p') ng/L 0.23 0.83 39 0.43 1.57 21 DDTs ng/L 0.58 1.28 39 1.48 3.89 21 delta-BHC ng/L 0 1 0 1 0 1 Demeton-s ng/L 0 0.38 0 0.20 0 0.43 0.20 Diazinon ng/L 0 1 0 1 0 1 Dichlofenthion ng/L 43.97 103.77 38 7.78 18.10 20 Dichlorvos ng/L 0 0 38 0 0 20 Dicrotophos ng/L 0 0 38 0 0 20 Dieldrin ng/L 0.15 0.42 39 0 0 21 Dimethoate ng/L 1.05 6.49 38 0 0 20 Dioxathion ng/L 0 0 38 </td <td>DDT(a p')</td> <td>ng/L</td> <td>0</td> <td>0</td> <td>20</td> <td>0 10</td> <td>0 20</td>	DDT(a p')	ng/L	0	0	20	0 10	0 20	
DDT(p,p) ng/L 0.23 0.83 39 0.43 1.37 21 DDTs ng/L 0.58 1.28 39 1.48 3.89 21 delta-BHC ng/L 0 1 0 1 0 1 Demeton-s ng/L 0 0 38 0 0 20 Diazinon ng/L 43.97 103.77 38 7.78 18.10 20 Dichlofenthion ng/L 0 0 38 0 0 20 Dichlorvos ng/L 0 0 38 0 0 20 Dicrotophos ng/L 0 0 38 0 0 20 Dieldrin ng/L 0.15 0.42 39 0 0 20 Dimethoate ng/L 0.05 6.49 38 0 0 20 Dioxathion ng/L 0 0 38 0 0 20	DDT(0,p)	ng/L	0 23	0 83	30	0.10	0.45 20	
bb1s ng/L 0.38 1.28 39 1.48 3.69 21 delta-BHC ng/L 0 1 0 1 0 1 Demeton-s ng/L 0 0 38 0 0 20 Diazinon ng/L 43.97 103.77 38 7.78 18.10 20 Dichlofenthion ng/L 0 0 38 0 0 20 Dichlorvos ng/L 0 0 38 0 0 20 Dicrotophos ng/L 0 0 38 0 0 20 Dimethoate ng/L 0.15 0.42 39 0 0 21 Dinxathion ng/L 0 0 38 0 0 20	DDT_{c}	ng/L	0.23	1.00	20	1 / 9	3 20 21	
Ing/L 0 1 0 1 Demeton-s ng/L 0 0 38 0 0 20 Diazinon ng/L 43.97 103.77 38 7.78 18.10 20 Dichlofenthion ng/L 0 0 38 0 0 20 Dichlorvos ng/L 0 0 38 0 0 20 Dicrotophos ng/L 0 0 38 0 0 20 Dimethoate ng/L 0.15 0.42 39 0 0 21 Divertion ng/L 0.05 6.49 38 0 0 20	dolta PHC	ng/L	0.56	1.20	39	1.40	3.09 21	
Definition ng/L 0 <	Domoton c	ng/L	0	0	20 1	0	0.20	
Diazition ng/L 43.97 103.77 38 7.78 18.10 20 Dichlofenthion ng/L 0 0 38 0 0 20 Dichlorvos ng/L 0 0 38 0 0 20 Dicrotophos ng/L 0 0 38 0 0 20 Dieldrin ng/L 0.15 0.42 39 0 0 21 Dimethoate ng/L 1.05 6.49 38 0 0 20	Diazinon	ng/L	43.07	103 77	20	7 79	19 10 20	
Dichlorent non ng/L 0 0 38 0 0 20 Dichlorvos ng/L 0 0 38 0 0 20 Dicrotophos ng/L 0 0 38 0 0 20 Dieldrin ng/L 0.15 0.42 39 0 0 21 Dimethoate ng/L 1.05 6.49 38 0 0 20	Diazinon	ng/L	43.97	103.77	20	1.10	10.10 20	
Dicrotophos ng/L 0 1 0 0 1 0 0 1 0 0 1 0 0 1 0 0 1 0 0 1 0 0 1 0 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1	Dichloryos	ng/L	0	0	20	0	0 20	
Dieldrin ng/L 0 10 0 0 10 0 0 10 0 0 10 0 0 10 0 0 10 0 10 <t< td=""><td>Dicrotophos</td><td>ng/L</td><td>0</td><td>0</td><td>20</td><td>0</td><td>0 20</td></t<>	Dicrotophos	ng/L	0	0	20	0	0 20	
Dimethoate ng/L 0.15 0.42 0	Dieldrin	ng/L	0 15	0 42	30	0	0 20	
Dintellioale 19/∟ 1.00 0.49 30 0 0 20 Dioxathion no/I 0 0.38 0 0.20	Dimethoate	ng/L	1 05	6 10	20	0	0 21	
	Dioxathion	ng/⊑	1.05	0. 4 9 N	38	0	0 20	

Pesticides Symbol Units Mean SD n Mean SD n Disuffon ng/L 3.95 10.28 3.95 0.02 22 Endosulfan I ng/L 0.14 0.38 39 0.05 0.22 21 Endosulfan II ng/L 0.04 0.243 39 0.05 0.22 21 Endrin Aldehyde ng/L 0.05 0.22 39 0 0 21 Endrin Aldehyde ng/L 0 0.38 0 0.22 12 Endrin Aldehyde ng/L 0 0.38 0 0.20 Ethoprop ng/L 0 0.38 0 0.20 Fenchlorphos ng/L 0 0.38 0 0.20 Fenthion ng/L 0 0.38 0 0.20 Gamma-BHC (Lindane) ng/L 0.04 0.24 38 0 0.20 Heptachlor ng/L 0.13 0.34			San	Juan		Santa	Margarit	а
Disulfoton ng/L 3.95 10.28 38 0 0 0 0 Endosulfan I ng/L 0.14 0.38 39 0.05 0.22 21 Endosulfan II ng/L 0.10 0.31 39 0.05 0.22 21 Endroin didebude ng/L 0.05 0.22 39 0 0.22 11 Endrin Aldebude ng/L 0.05 0.22 39 0 0.22 12 Endrin Aldebude ng/L 0 0.38 0 0.22 12 Endrin Ketone ng/L 0 0.38 0 0.22 12 Ethion ng/L 0 0.38 0 0.20 13 0.02 14 0 0 38 0 0.20 Fenchlorphos ng/L 0 0.38 0 0.20 1 0 0 38 0 0.20 Fenchlorphos ng/L 0 0.38	Pesticides	Symbol Units	Mean S	SD	n	Mean	SD	n
Endosulfan I ng/L 0.14 0.38 39 0.05 0.22 21 Endosulfan sulfate ng/L 0.04 0.24 39 0 0 21 Endosulfan sulfate ng/L 0.05 0.22 29 0 0 21 Endrin ng/L 0 0.38 0 0 20 Endrin Aldehyde ng/L 0 0.38 0 0 20 Ethoprop ng/L 0 0.38 0 0 20 Fenchlorphos ng/L 0 0.38 0 0 20 Fenthrothion ng/L 0 0.38 0 0 20 Fenthrothion ng/L 0 0.38 0 0 20 Gamma-BHC (Lindane) ng/L 0 0.38 0 0 20 HCH, alpha ng/L 0.04 3.38 0 0 20 HCH, delta ng/L 0.13	Disulfoton	ng/L	3.95	10.28	38	0	0	20
Endosulfan II ng/L 0.04 0.24 9 0 0.21 Endosulfan sulfate ng/L 0.10 0.31 39 0.05 0.22 21 Endrin ng/L 0.05 0.22 39 0 0 21 Endrin Aldehyde ng/L 0 0.38 0 0.20 Ethion ng/L 0 0.38 0 0.20 Ethion ng/L 0 0.38 0 0.20 Fenchlorphos ng/L 0 0.38 0 0.20 Fenchlorphos ng/L 0 0.38 0 0.20 Fensitothion ng/L 0 0.38 0 0.20 Fensitothion ng/L 0 0.38 0 0.20 Gamma-BHC (Lindane) ng/L 0.04 0.24 38 0 0.20 HCH, alpha ng/L 0.03 38 0 0.20 21 Heptachlor ng/L	Endosulfan I	ng/L	0.14	0.38	39	0.05	0.22	21
Endosulfan sulfate ng/L 0.10 0.31 39 0.05 0.22 39 0 0 21 Endrin Aldehyde ng/L 0 0.38 0 0 0 21 Endrin Aldehyde ng/L 0 0.38 0 0 20 Ethion ng/L 0 0.38 0 0 20 Fenchlorphos ng/L 0 0.38 0 0 20 Fenchlorphos ng/L 0 0.38 0 0 20 Fenchlorphos ng/L 0 0.38 0 0 20 Fentitothion ng/L 0 0.38 0 0 20 Gamma-BHC (Lindane) ng/L 0 1 0 1 0 1 0 1 0 20 HCH, beta ng/L 0 0.38 0 20 20 20 20 20 20 20 20 20	Endosulfan II	ng/L	0.04	0.24	39	0	0	21
Endrin ng/L 0.05 0.22 39 0 0 21 Endrin Aldehyde ng/L 0 0 38 0 0 21 Endrin Ketone ng/L 0 0 38 0 0 20 Ethoprop ng/L 0 0 38 0 0 20 Famphur ng/L 0 0 38 0 0 20 Fenitrothion ng/L 0 0 38 0 0 20 Fensulfothion ng/L 0 0 38 0 0 20 Fondros ng/L 0 0 38 0 0 20 Fondros ng/L 0 0 38 0 0 20 Gamma-BHC (Lindane) ng/L 0.04 0 38 0 0 20 HCH, gamma ng/L 0.13 0.34 38 0 0 20 <td>Endosulfan sulfate</td> <td>ng/L</td> <td>0.10</td> <td>0.31</td> <td>39</td> <td>0.05</td> <td>0.22</td> <td>21</td>	Endosulfan sulfate	ng/L	0.10	0.31	39	0.05	0.22	21
Endrin Aldehyde ng/L 0 0 38 0 0 21 Endrin Ketone ng/L 0 0 38 0 0 20 Ethion ng/L 0 0 38 0 0 20 Famphur ng/L 0 0 38 0 0 20 Fenchlorphos ng/L 0 0 38 0 0 20 Fensulfohion ng/L 0 0 38 0 0 20 Fenchlorphos ng/L 0 0 38 0 0 20 Fensulfohion ng/L 0 0 38 0 0 20 Gamma-BHC (Lindane) ng/L 0 0 38 0 0 20 HCH, detta ng/L 0 0 38 0 0 20 HCH, detta ng/L 0 0 38 0 0 20	Endrin	ng/L	0.05	0.22	39	0	0	21
Endrin Ketone ng/L 0 0 38 0 0 20 Ethion ng/L 0 0 38 0 0 20 Ethoprop ng/L 0 0 38 0 0 20 Fenchlorphos ng/L 0 0 38 0 0 20 Fenchlorphos ng/L 0 0 38 0 0 20 Fencentorthion ng/L 0 0 38 0 0 20 Foncfos ng/L 0 0 38 0 0 20 Generators ng/L 0 0 38 0 0 20 0 <td>Endrin Aldehyde</td> <td>ng/L</td> <td>0</td> <td>0</td> <td>39</td> <td>0</td> <td>0</td> <td>21</td>	Endrin Aldehyde	ng/L	0	0	39	0	0	21
Ethion ng/L 0 0 38 0 0 20 Ethoprop ng/L 0 0 38 0 0 20 Famphur ng/L 0 0 38 0 0 20 Fenchlorphos ng/L 0 0 38 0 0 20 Fensulfothion ng/L 0 0 38 0 0 20 Fonofos ng/L 0 0 38 0 0 20 gamma-BHC (Lindane) ng/L 0 1 0 1 0 20 HCH, beta ng/L 0 13 0.34 38 0 0 20 HCH, delta ng/L 0.13 0.34 38 0 0 20 HCH, detta ng/L 0.13 0.34 38 0 20 20 Heptachlor ng/L 0.21 0.70 39 0.10 0	Endrin Ketone	ng/L	0	0	38	0	0	20
Ethoprop ng/L 0 0 38 0 0 20 Famphur ng/L 0 0 38 0 0 20 Fenchlorphos ng/L 0 0 38 0 0 20 Fensulfothion ng/L 0 0 38 0 0 20 Gensulfothion ng/L 0 0 38 0 0 20 Gensulfothion ng/L 0 0 38 0 0 20 gamma-BHC (Lindane) ng/L 0 1 0 1 0 1 0 20 HCH, Japha ng/L 0.13 0.34 8 0 0 20 HCH, detta ng/L 0 0 38 0 0 20 Heptachlor ng/L 0 0 38 0 0 20 Heptachlor ng/L 0 0 38 0	Ethion	ng/L	0	0	38	0	0	20
Famphur ng/L 0 0 38 0 0 20 Fenchlorphos ng/L 0 0 38 0 0 20 Fensulfothion ng/L 0 0 38 0 0 20 Fensulfothion ng/L 0 0 38 0 0 20 Genthion ng/L 0 0 38 0 0 20 gamma-BHC (Lindane) ng/L 0 1 0 1	Ethoprop	ng/L	0	0	38	0	0	20
Fenchlorphos ng/L 0 0 38 0 0 20 Fensufforbion ng/L 0 0 38 0 0 20 Fensufforbion ng/L 0 0 38 0 0 20 Fenthion ng/L 0 0 38 0 0 20 garma-BHC (Lindane) ng/L 0.04 0.24 38 0 0 20 HCH, Jeta ng/L 0.0 38 0 0 20 HCH, detta ng/L 0 0 38 0 0 20 HCH, detta ng/L 0 0 38 0 0 20 Heptachlor ng/L 0.10 0.30 21 Hexachlorobenzene ng/L 0.10 0.30 20 Heptachlor ng/L 0 0 38 0 0 20 Mathibion ng/L 0 0 38	Famphur	ng/L	0	0	38	0	0	20
Fenitrothion ng/L 0 0 38 0 0 20 Fensulfothion ng/L 0 0 38 0 0 20 Fenthion ng/L 0 0 38 0 0 20 gamma-BHC (Lindane) ng/L 0 1 0 1	Fenchlorphos	ng/L	0	0	38	0	0	20
Fensulfothion ng/L 0 0 38 0 0 20 Fenthion ng/L 0 0 38 0 0 20 Fonofos ng/L 0 0 38 0 0 20 gamma-BHC (Lindane) ng/L 0 0 38 0 0 20 HCH, apha ng/L 0 0 38 0 0 20 HCH, detta ng/L 0 0 38 0 0 20 HCH, detta ng/L 0 0 38 0 0 20 Heptachlor ng/L 0 0 38 0 0 20 Heptachlor poxide ng/L 0.18 0.68 38 0 20 20 Matathion ng/L 0 0 38 0 20 20 20 20 20 20 20 20 20 20 20	Fenitrothion	ng/L	0	0	38	0	0	20
Fenthion ng/L 0 0 38 0 0 20 Fonofos ng/L 0 0 38 0 0 20 gamma-BHC (Lindane) ng/L 0.04 0.24 38 0 0 20 HCH, alpha ng/L 0.04 0.24 38 0 0 20 HCH, beta ng/L 0.13 0.34 38 0 0 20 HCH, delta ng/L 0 0 38 0 0 20 Heptachlor ng/L 0 0 38 0 0 20 Heptachlor epoxide ng/L 0.18 0.68 38 0 0 20 Leptophos ng/L 0 0 38 0 20 0 0 20 Methoxychlor 0 20 0 20 0 23 0 21 0 20 20 0 20 20	Fensulfothion	ng/L	0	0	38	0	0	20
Fonofos ng/L 0 0 38 0 0 20 gamma-BHC (Lindane) ng/L 0 1 0 1 0 1 HCH, alpha ng/L 0.04 0.24 38 0 0 20 HCH, beta ng/L 0 0 38 0 0 20 HCH, detta ng/L 0 0 38 0 0 20 Heptachlor ng/L 0 0 38 0 0 20 Heptachlor epoxide ng/L 0.21 0.70 39 0.10 0.30 21 Hexachlorobenzene ng/L 0.18 0.68 38 0 20 Mathition ng/L 0 0 38 0 20 20 Methidathion ng/L 0 0 38 0 20 20 Methidathion ng/L 0 0 38 0 20 <t< td=""><td>Fenthion</td><td>ng/L</td><td>0</td><td>0</td><td>38</td><td>0</td><td>0</td><td>20</td></t<>	Fenthion	ng/L	0	0	38	0	0	20
gamma-BHC (Lindane) ng/L 0 1 0 1 HCH, alpha ng/L 0.04 0.24 38 0 0 20 HCH, beta ng/L 0 38 0.10 0.45 20 HCH, delta ng/L 0.13 0.34 38 0 0 20 HCH, gamma ng/L 0 0 38 0 0 20 Heptachlor ng/L 0.18 0.68 38 0 0 20 Leptophos ng/L 0.18 0.68 38 0 0 20 Mathtion ng/L 0 0 38 0 0 20 Methidathion ng/L 0 0 38 0 0 20 Methidathion ng/L 0 0 38 0 0 20 Methidathion ng/L 0 0 38 0 0 20 Methox	Fonofos	ng/L	0	0	38	0	0	20
HCH, alpha ng/L 0.04 0.24 38 0 0 20 HCH, beta ng/L 0 0 38 0.10 0.45 20 HCH, delta ng/L 0.13 0.34 38 0 0 20 HcH, gamma ng/L 0 0 38 0 0 20 Heptachlor ng/L 0 0 39 0 0 21 Heptachlor epoxide ng/L 0.21 0.70 39 0.10 0.30 21 Hexachlorobenzene ng/L 0 0 38 0 0 20 Mathion ng/L 0 0 38 0 0 20 Methoxychlor ng/L 0 0 38 0 0 20 Mirex ng/L 0 0 38 0 0 20 Mirex ng/L 0 0 38 0 0	gamma-BHC (Lindane)	ng/L	0		1	0		1
HCH, beta ng/L 0 0 38 0.10 0.45 20 HCH, detta ng/L 0.13 0.34 38 0 0 20 HCH, gamma ng/L 0 0 38 0 0 20 Heptachlor ng/L 0.21 0.70 39 0.10 0.30 21 Heptachlor epoxide ng/L 0.18 0.68 38 0 0 20 Leptophos ng/L 0 0 38 0 0 20 Mathinon ng/L 0 0 38 0 0 20 Methidathion ng/L 0 0 38 0 0 20 Methoxychlor ng/L 0 0 38 0 0 20 Mitex ng/L 0 0 38 0 0 20 Methoxychlor ng/L 0 0 38 0 20	HCH, alpha	ng/L	0.04	0.24	38	0	0	20
HCH, delta ng/L 0.13 0.34 38 0 0 20 HCH, gamma ng/L 0 0 38 0 0 20 Heptachlor ng/L 0.21 0.70 39 0.10 0.30 21 Heptachlor epoxide ng/L 0.21 0.70 39 0.10 0.30 21 Hexachlorobenzene ng/L 0.18 0.68 38 0 0 20 Leptophos ng/L 0 0 38 0 0 20 Mathion ng/L 0 0 38 0 0 20 Methidathion ng/L 0 0 38 0 0 20 Methoxychlor ng/L 0 0 38 0 0 20 Mirex ng/L 0 0 38 0 0 20 Nated ng/L 0 0 38 0 0 <td>HCH, beta</td> <td>ng/L</td> <td>0</td> <td>0</td> <td>38</td> <td>0.10</td> <td>0.45</td> <td>20</td>	HCH, beta	ng/L	0	0	38	0.10	0.45	20
HCH, gamma ng/L 0 0 38 0 0 20 Heptachlor ng/L 0.21 0.70 39 0.10 0.32 21 Heptachlor epoxide ng/L 0.18 0.68 38 0 0 20 Leptophos ng/L 0 0 38 0 0 20 Malathion ng/L 0 0 38 0 0 20 Methidathion ng/L 0 0 38 0 0 20 Methidathion ng/L 0 0 38 0 0 20 Methidathion ng/L 0 0 38 0 20 Methidathion ng/L 0 0 38 0 20 Methidathion ng/L 0 0 38 0 20 Mirex ng/L 0 0 38 0 20 Naled ng/L	HCH, delta	ng/L	0.13	0.34	38	0	0	20
Heptachlor ng/L 0 0 39 0 0 21 Heptachlor epoxide ng/L 0.21 0.70 39 0.10 0.30 21 Hexachlorobenzene ng/L 0.18 0.68 38 0 0 20 Leptophos ng/L 0 0 38 0 0 20 Malathion ng/L 0 0 38 0 0 20 Methoxychlor ng/L 0 0 38 0 0 20 Metinate ng/L 0 0 38 0 0 20 Metinate ng/L 0 0 38 0 0 20 Mirex ng/L 0 0 38 0 20 20 Nonachlor, cis- ng/L 0.05 0.23 38 0.05 0.22 20 Narathlor, Ethyl ng/L 0.01 0 38 0	HCH. gamma	na/L	0	0	38	0	0	20
Heptachlor epoxide ng/L 0.21 0.70 39 0.10 0.30 21 Hexachlorobenzene ng/L 0.18 0.68 38 0 0 20 Leptophos ng/L 0 0 38 0 0 20 Malathion ng/L 0 0 38 0 0 20 Methos ng/L 0 0 38 0 0 20 Methoxychlor ng/L 0 0 38 0 0 20 Mirex ng/L 0 0 38 0 0 20 Mirex ng/L 0 0 38 0 0 20 Nonachlor, cis- ng/L 0 0 38 0 0 20 Nonachlor, trans- ng/L 0.03 0.16 38 0.05 0.22 20 Oxadiazon ng/L 0.01 0.38 0 20 <t< td=""><td>Heptachlor</td><td>na/L</td><td>0</td><td>0</td><td>39</td><td>0</td><td>0</td><td>21</td></t<>	Heptachlor	na/L	0	0	39	0	0	21
Hexachlorobenzene ng/L 0.18 0.68 38 0 0.20 Leptophos ng/L 0 0 38 0 0.20 Malathion ng/L 0 0 38 0 0.20 Merphos ng/L 0 0 38 0 0.20 Methidathion ng/L 0 0 38 0 0.20 Mirex ng/L 0 0 38 0 20 Malath ng/L 0 0 38 0 20 Malath ng/L 0 0 38 0 20 Malath ng/L 0.03 0.16 38 0.05 0.22 20 <t< td=""><td>Heptachlor epoxide</td><td>na/L</td><td>0.21</td><td>0.70</td><td>39</td><td>0.10</td><td>0.30</td><td>21</td></t<>	Heptachlor epoxide	na/L	0.21	0.70	39	0.10	0.30	21
Leptophos ng/L 0 0 38 0 0 20 Malathion ng/L 0 0 38 0 0 20 Merphos ng/L 0 0 38 0 0 20 Methoxychlor ng/L 0 0 38 0 0 20 Methoxychlor ng/L 0 0 38 0 0 20 Methoxychlor ng/L 0 0 38 0 0 20 Mirex ng/L 0 0 38 0 0 20 Malathion ng/L 0 0 38 0 0 20 Minate ng/L 0 0 38 0 0 20 Nonachlor, trans- ng/L 0.05 0.23 38 0.05 0.22 20 Oxadiazon ng/L 0 0 38 0 20 20	Hexachlorobenzene	ng/L	0.18	0.68	38	0	0	20
Malathion ng/L 0 0 38 0 20 Merphos ng/L 0 0 38 0 20 Methidathion ng/L 0 0 38 0 20 Methoxychlor ng/L 0 0 38 0 20 Mevinphos ng/L 0 0 38 0 20 Mirex ng/L 0 0 38 0 20 Mirex ng/L 0 0 38 0 20 Maled ng/L 0 0 38 0 20 Nonachlor, cis- ng/L 0.05 0.23 38 0.05 0.22 20 Nonachlor, trans- ng/L 0.05 0.23 38 0.05 0.22 20 Oxychlordane ng/L 0.11 0.31 38 0.5 2.22 20 Parathion, Ethyl ng/L 0 0 38	Leptophos	na/L	0	0	38	0	0	20
Marphos ng/L 0 0 38 0 0 20 Methidathion ng/L 0 0 38 0 0 20 Methoxychlor ng/L 0 0 38 0 0 21 Mevinphos ng/L 0 0 38 0 0 20 Mirex ng/L 0 0 38 0 0 20 Molinate ng/L 0 0 38 0 0 20 Naled ng/L 0.03 0.16 38 0 0 20 Nonachlor, cis- ng/L 0.03 0.16 38 0.05 0.22 20 Natation ng/L 0.05 0.23 38 0.05 0.22 20 Nonachlor, trans- ng/L 0.05 0.23 38 0.05 0.22 20 Oxychlordane ng/L 0.01 0.38 0.02 20	Malathion	ng/L	0	0	38	0	0	20
Methidathion ng/L 0 0 38 0 0 20 Methoxychlor ng/L 0 0 39 0 0 21 Mevinphos ng/L 0 0 38 0 0 20 Mirex ng/L 0 0 38 0 0 20 Molinate ng/L 0 0 38 0 0 20 Naled ng/L 0.03 0.16 38 0.05 0.22 20 Nonachlor, cis- ng/L 0.05 0.23 38 0.05 0.22 20 Oxadiazon ng/L 0.11 0.31 38 0.05 0.22 20 Oxadiazon ng/L 0.11 0.31 38 0.05 0.22 20 Parathion, Ethyl ng/L 0 0 38 0 20 20 Phosphamidon ng/L 0 0 38 0	Merphos	ng/L	0	0	38	0	0	20
Instruction Ingle Ingle	Methidathion	ng/l	0	0	38	0	0	20
Metriciphos ng/L 0 0 38 0 0 20 Mevinphos ng/L 0 0 38 0 0 20 Mirex ng/L 0 0 38 0 0 20 Molinate ng/L 0 0 38 0 0 20 Naled ng/L 0.03 0.16 38 0.05 0.22 20 Nonachlor, cis- ng/L 0.05 0.23 38 0.05 0.22 20 Nonachlor, trans- ng/L 0.05 0.23 38 0.05 0.22 20 Oxadiazon ng/L 0.05 0.23 38 0.05 0.22 20 Oxadiazon ng/L 0.05 0.23 38 0.05 0.22 20 Parathion, Ethyl ng/L 0.011 0.31 38 0.05 0.22 20 Phorate ng/L 0 0 38 0 0 20 Phosphamidon ng/L 0 0	Methoxychlor	ng/l	0	0	39	0	0	21
Mirex ng/L 0 0 38 0 0 20 Molinate ng/L 0 0 38 0 0 20 Naled ng/L 0 0 38 0 0 20 Nonachlor, cis- ng/L 0.03 0.16 38 0.05 0.22 20 Nonachlor, trans- ng/L 0.05 0.23 38 0.05 0.22 20 Oxadiazon ng/L 46.21 164.92 38 22.15 68.41 20 Oxychlordane ng/L 0.11 0.31 38 0.05 0.22 20 Parathion, Ethyl ng/L 0 0 38 0 0 20 Phorate ng/L 0 0 38 0 0 20 Phosphamidon ng/L 0 0 38 0 0 20 Prometon ng/L 0 0 38 0	Mevinphos	ng/l	0	0	38	0	0	20
Instruct IngrL 0 0 38 0 0 20 Naled ng/L 0 0 38 0 0 20 Naled ng/L 0.03 0.16 38 0 0 20 Nonachlor, cis- ng/L 0.05 0.23 38 0.05 0.22 20 Oxadiazon ng/L 46.21 164.92 38 22.15 68.41 20 Oxychlordane ng/L 0.11 0.31 38 0.05 0.22 20 Parathion, Ethyl ng/L 0 0 38 0 20 20 Phorate ng/L 0 0 38 0 0 20 Phosmet ng/L 0 0 38 0 0 20 Prometryn ng/L 0 0 38 0 0 20 Propazine ng/L 0 0 38 0 0 <td>Mirex</td> <td>ng/l</td> <td>0</td> <td>0</td> <td>38</td> <td>0</td> <td>0</td> <td>20</td>	Mirex	ng/l	0	0	38	0	0	20
Instruction Ing/L 0	Molinate	ng/L	0 0	Ő	38	Ő	0	20
Nonachlor, cis- ng/L 0.03 0.16 38 0.05 0.22 20 Nonachlor, trans- ng/L 0.05 0.23 38 0.05 0.22 20 Oxadiazon ng/L 46.21 164.92 38 22.15 68.41 20 Oxychlordane ng/L 0.11 0.31 38 0.05 0.22 20 Parathion, Ethyl ng/L 0.11 0.31 38 0.05 0.22 20 Parathion, Methyl ng/L 0 0 38 0 0 20 Phorate ng/L 0 0 38 0 0 20 Phosmet ng/L 0 0 38 0 0 20 Prometon ng/L 0 0 38 0 0 20 Prometon ng/L 0 0 38 0 0 20 Propazine ng/L 0 0 38 0 0 20 Simazine ng/L 0.89 5.52	Naled	ng/L	0	Ő	38	Ő	0	20
Nonachlor, trans- ng/L 0.05 0.10 0.05 0.23 38 0.05 0.22 20 Oxadiazon ng/L 46.21 164.92 38 22.15 68.41 20 Oxychlordane ng/L 0.11 0.31 38 0.05 0.22 20 Parathion, Ethyl ng/L 0.11 0.31 38 0.05 0.22 20 Parathion, Methyl ng/L 0 0 38 0 0 20 Phorate ng/L 0 0 38 0 0 20 Phorate ng/L 0 0 38 0 0 20 Phosemet ng/L 0 0 38 0 0 20 Prometon ng/L 0 0 38 0 0 20 Prometon ng/L 0 0 38 0 0 20 Prometryn ng/L 0 0 38 0 0 20 Propazine ng/L 0	Nonachlor cis-	ng/L	0.03	0 16	38	0.05	0.22	20
Instruction, runing Ing/L 0.00 0.120 0.00 0.121 0.00 Oxadiazon ng/L 46.21 164.92 38 22.15 68.41 20 Oxychlordane ng/L 0.11 0.31 38 0.05 0.22 20 Parathion, Ethyl ng/L 0 0 38 0 0 20 Parathion, Methyl ng/L 0 0 38 0 0 20 Phorate ng/L 0 0 38 0 0 20 Phorate ng/L 0 0 38 0 0 20 Phosemet ng/L 0 0 38 0 0 20 Prometon ng/L 0 0 38 0 0 20 Prometryn ng/L 0 0 38 0 0 20 Propazine ng/L 0 0 38 0 0 20 Simazine ng/L 0.89 5.52 38 35.90	Nonachlor trans-	ng/L	0.05	0.23	38	0.05	0.22	20
Oxychlordane ng/L 0.11 0.31 38 0.05 0.22 20 Parathion, Ethyl ng/L 0 0 38 0 0 20 Parathion, Methyl ng/L 0 0 38 0 0 20 Phorate ng/L 0 0 38 0 0 20 Phorate ng/L 0 0 38 0 0 20 Phosmet ng/L 0 0 38 0 0 20 Phosphamidon ng/L 0 0 38 0 0 20 Prometon ng/L 0 0 38 0 0 20 Prometryn ng/L 0 0 38 0 0 20 Propazine ng/L 0 0 38 0 0 20 Simazine ng/L 0.89 5.52 38 35.90 100.34 20 Simetryn ng/L 0 0 38 0 0 <	Oxadiazon	ng/L	46 21	164.92	38	22 15	68 41	20
Parathion, Ethyl ng/L 0 0 38 0 0 20 Parathion, Methyl ng/L 0 0 38 0 0 20 Phorate ng/L 0 0 38 0 0 20 Phorate ng/L 0 0 38 0 0 20 Phosmet ng/L 0 0 38 0 0 20 Phosphamidon ng/L 0 0 38 0 0 20 Prometon ng/L 0 0 38 0 0 20 Prometryn ng/L 0 0 38 0 0 20 Propazine ng/L 0 0 38 0 0 20 Simazine ng/L 3.11 13.48 38 1.75 7.83 20 Simetryn ng/L 0 0 38 0 0 20 Simetryn ng/L 0 0 38 0 0 20 <td>Oxychlordane</td> <td>ng/L</td> <td>0.11</td> <td>0.31</td> <td>38</td> <td>0.05</td> <td>0.22</td> <td>20</td>	Oxychlordane	ng/L	0.11	0.31	38	0.05	0.22	20
Parathion, Methyl ng/L 0 0 38 0 0 20 Phorate ng/L 0 0 38 0 0 20 Phorate ng/L 0 0 38 0 0 20 Phosmet ng/L 0 0 38 0 0 20 Phosphamidon ng/L 0 0 38 0 0 20 Prometon ng/L 0 0 38 0 0 20 Prometryn ng/L 0 0 38 0 0 20 Propazine ng/L 0 0 38 0 0 20 Secbumeton ng/L 3.11 13.48 38 1.75 7.83 20 Simazine ng/L 0 0 38 0 0 20 Sulfotep ng/L 0 0 38 0 0 20 Sulfotep ng/L 0 0 38 0 0 20	Parathion Ethyl	ng/L	0.11	0.01	38	0.00	0.22	20
Phorate ng/L 0 0 38 0 0 20 Phosmet ng/L 0 0 38 0 0 20 Phosmet ng/L 0 0 38 0 0 20 Phosphamidon ng/L 0 0 38 0 0 20 Prometon ng/L 0 0 38 0 0 20 Prometon ng/L 0 0 38 0 0 20 Prometon ng/L 0 0 38 0 0 20 Prometryn ng/L 0 0 38 0 0 20 Propazine ng/L 0 0 38 0 0 20 Secbumeton ng/L 3.11 13.48 38 1.75 7.83 20 Simazine ng/L 0 0 38 0 0 20 Sulfotep ng/L 0 0 38 0 0 20 20	Parathion Methyl	ng/L	0 0	Ő	38	Ő	0	20
Instate ng/L 0	Phorate	ng/L	0	Ő	38	Ő	0	20
Phosphamidon ng/L 0 0 38 0 0 20 Prometon ng/L 0 0 38 0 0 20 Prometon ng/L 0 0 38 0 0 20 Prometryn ng/L 0 0 38 0 0 20 Propazine ng/L 0 0 38 0 0 20 Secbumeton ng/L 3.11 13.48 38 1.75 7.83 20 Simazine ng/L 0.89 5.52 38 35.90 100.34 20 Simetryn ng/L 0 0 38 0 0 20 Sulfotep ng/L 0 0 38 0 0 20 Tedion ng/L 0 0 38 0 0 20	Phosmet	ng/L	0	0	38	0	0	20
Prometon ng/L 0 0 38 0 0 20 Prometryn ng/L 0 0 38 0 0 20 Prometryn ng/L 0 0 38 0 0 20 Prometryn ng/L 0 0 38 0 0 20 Propazine ng/L 0 0 38 0 0 20 Secbumeton ng/L 3.11 13.48 38 1.75 7.83 20 Simazine ng/L 0.89 5.52 38 35.90 100.34 20 Simetryn ng/L 0 0 38 0 0 20 Sulfotep ng/L 0 0 38 0 0 20 Tedion ng/L 0.05 0.23 38 0 0 20	Phosphamidon	ng/L	0	Ő	38	Õ	0	20
Prometryn ng/L 0 0 38 0 0 20 Propazine ng/L 0 0 38 0 0 20 Secbumeton ng/L 3.11 13.48 38 1.75 7.83 20 Simazine ng/L 0.89 5.52 38 35.90 100.34 20 Simetryn ng/L 0 0 38 0 0 20 Sulfotep ng/L 0 0 38 0 0 20 Tedion ng/L 0 0 38 0 0 20	Prometon	ng/L	0	0	38	0	0	20
Propazine ng/L 0 0 38 0 0 20 Secbumeton ng/L 3.11 13.48 38 1.75 7.83 20 Simazine ng/L 0.89 5.52 38 35.90 100.34 20 Simetryn ng/L 0 0 38 0 0 20 Sulfotep ng/L 0 0 38 0 0 20 Tedion ng/L 0 0 38 0 0 20	Prometryn	ng/L	0	0	38	0	0	20
Inspiration ng/L 3.11 13.48 38 1.75 7.83 20 Simazine ng/L 0.89 5.52 38 35.90 100.34 20 Simetryn ng/L 0 0 38 0 0 20 Sulfotep ng/L 0 0 38 0 0 20 Tedion ng/L 0 0 38 0 0 20	Pronazine	ng/L	0	0	38	0	0	20
Simazine ng/L 0.89 5.52 38 35.90 100.34 20 Simetryn ng/L 0 0 38 0 0 20 Sulfotep ng/L 0 0 38 0 0 20 Tedion ng/L 0.05 0.23 38 0 0 20	Sectumeton	ng/L	3 1 1	13.48	38	1 75	7 83	20
Simetryn ng/L 0.03 0.02 00.04 20 Sulfotep ng/L 0 0.38 0 0.20 Tedion ng/L 0 0.38 0 0.20	Simazine	ng/L	0.11	5 52	38	35 90	100 34	20
Sulfotep ng/L 0 <th< td=""><td>Simetryn</td><td>ng/L</td><td>0.09</td><td>0.02</td><td>38</td><td>00.00</td><td>100.04</td><td>20</td></th<>	Simetryn	ng/L	0.09	0.02	38	00.00	1 00.04	20
Tedion ng/l 0.05 0.23 38 0 0.20	Sulfoten	ng/L	0	0	38	0	0	20
	Tedion	ng/L	0.05	0 23	38	0	0	20

		San	Santa Margarita			
Pesticides	Symbol Units	Mean S	SD i	n M	lean SD	n
Terbufos	ng/L	0	0	38	0	0 20
Terbuthylazine	ng/L	0	0	38	0	0 20
Terbutryn	ng/L	0	0	38	0	0 20
Tetrachlorvinphos	ng/L	0	0	38	0	0 20
Thiobencarb	ng/L	3.95	24.33	38	0	0 20
Thionazin	ng/L	0	0	38	0	0 20
Tokuthion	ng/L	0	0	38	0	0 20
Toxaphene	ng/L	0		1	0	1
Trichlorfon	ng/L	0	0	38	0	0 20
Trichloronate	ng/L	0	0	38	0	0 20

			San	Luis Rey		Ca	rlsbad	
Physical water quality and inorganics	Symbol	Units	Mean	SD	n	Mean	SD	n
Alkalinity as CaCO3	Alk	mg/l	198	84	25	252	43	41
Chloride		mg/l	95	129	3			0
Fine-ASTM		%			0	24.4	27.1	30
Fine-ASTM, Passing No. 200 Sieve		%			0	24.9	20.0	10
Oxygen, Dissolved	DO	mg/L	9.4	3.6	24			0
Oxygen, Saturation		%	98	39	24	102	32	40
рН		рН	7.7	0.4	24	7.9	1.5	40
Salinity	Sal	ppt	0.83	0.52	24			0
Specific conductivity	Cond	µS/cm	1516	968	24	3800	4232	40
Sulfate	SO_4	mg/l	382	264	25	469	277	41
Suspended Sediment Concentration		%			0			0
Temperature		°C	17.1	4.5	24	18.5	3.0	40
Total Organic Carbon		mg/L			0	2.24		1
Total Suspended Solids		mg/L	45	79	25	15		1
Turbidity		NTU	12.0	19.0	24	3.5	3.9	40
Velocity		ft/s	0.5	0.9	25	0.7	1.4	41
Nutrients								
Ammonia as N	NH ₃ -N	mg/l	0.05	0.06	25	0.12	0.11	41
Nitrate + Nitrite as N		mg/l	5.13	5.50	25	6.35	11.47	41
Nitrate as N		mg/l	5.10	5.49	25	3.70		1
Nitrate as NO3	NO_3	mg/l			0	16.00		1
Nitrite as N		mg/l	0.03	0.04	25	0		1
Nitrogen, Total Kieldahl		mg/l	0.89	1.18	25	0.49	0.20	41
OrthoPhosphate as P		mg/l	31.00		1	0.12	0.07	41
Phosphorus as P,Total	TP	mg/l	0.21	0.24	25	0.14	0.09	41
Metals		•						
Aluminum		µg/L	6.0	6.4	25	14.7	71.5	41
Arsenic	As	µg/L	1.3	0.9	25	4.7	2.6	41
Cadmium	Cd	µg/L	0.03	0.02	25	0.05	0.04	41
Chromium	Cr	µg/L	0.33	0.28	25	1.06	1.05	41
Copper	Cu	µg/L	4.03	2.60	25	3.55	1.50	41
Lead	Pb	µg/L	0.06	0.05	25	0.05	0.12	41
Manganese	Mn	µg/L	133	270	25	127	147	41
Nickel	Ni	µg/L	0.97	2.14	25	2.16	1.37	41
Selenium	Se	µg/L	4.9	4.8	24	10.6	10.1	40
Silver	Ag	µg/L	0.07	0.34	25	0.05	0.28	41
Zinc	Zi	µg/L	2.7	1.9	24	6.5	6.4	40
Bacteria								
Enterococcus		MPN/100 ml	93		1	490		1
Fecal Coliform		MPN/100 ml	170		1	900		1
Total Coliform		MPN/100 ml	1600		1	1600		1

Appendix, continued.		San Luis	Rev		Car	sbad	
PAHs	Symbol Units	Mean SD		n	Mean S	SD	n
Acenaphthene	ng/L	0	0	25	1	5	41
Acenaphthylene	ng/L	0	0	25	0	0	41
Anthracene	ng/L	0	0	25	4	14	41
Benz(a)anthracene	ng/L	0	0	25	0	0	41
Benzo(a)pyrene	ng/L	0	0	25	0	0	41
Benzo(b)fluoranthene	ng/L	0	0	25	0	0	41
Benzo(e)pyrene	ng/L	0	0	25	0	0	41
Benzo(g,h,i)perylene	ng/L	0.3	1.3	25	0	0	41
Benzo(k)fluoranthene	ng/L	0	0	24	0	0	40
Biphenyl	ng/L	0	0	24	0	0	40
Chrysene	ng/L	0	0	25	0	0	41
Chrysenes, C1 -	ng/L	0	0	24	0	0	10
Chrysenes, C2 -	ng/L	0	0	24	0	0	10
Chrysenes, C3 -	ng/L	0	0	24	0	0	10
Dibenz(a,h)anthracene	ng/L	0	0	25	0	0	41
Dibenzothiophene	ng/L	0.3	1.7	24	0	0	10
Dibenzothiophenes, C1 -	ng/L	0	0	24	0	0	10
Dibenzothiophenes, C2 -	ng/L	0	0	24	0	0	10
Dibenzothiophenes, C3 -	ng/L	0	0	24	0	0	10
Dimethylnaphthalene, 2,6-	ng/L	0	1	24	0	0	40
Dimethylphenanthrene, 3,6-	ng/L	0	0	18			0
Fluoranthene	ng/L	0	0	25	0	0	41
Fluoranthene/Pyrenes, C1 -	ng/L	0	0	24	0	0	10
Fluorene	ng/L	0	0	25	1	4	41
Fluorenes, C1 -	ng/L	0	0	24	0	0	10
Fluorenes, C2 -	ng/L	0	0	24	28.82	5.41	10
Fluorenes, C3 -	ng/L	0	0	24	0	0	10
Indeno(1,2,3-c,d)pyrene	ng/L	0	0	25	0	0	41
Methyldibenzothiophene, 4-	ng/L	0	0	18			0
Methylfluoranthene, 2-	ng/L	0	0	18			0
Methylfluorene, 1-	ng/L	0	0	18			0
Methylnaphthalene, 1-	ng/L	0	0	24	0	0	40
Methylnaphthalene, 2-	ng/L	0	0	24	0	0	40
Methylphenanthrene, 1-	ng/L	0	0	24	0	0	40
Naphthalene	ng/L	0.20	1.01	25	4.27	11.60	41
Naphthalenes, C1 -	ng/L	0.46	2.25	24	0	0	10
Naphthalenes, C2 -	ng/L	1.34	3.81	24	0	0	10
Naphthalenes, C3 -	ng/L	0	0	24	0	0	10
Naphthalenes, C4 -	ng/L	0	0	24	0	0	10
Perylene	ng/L	0	0	24	0	0	40
Phenanthrene	ng/L	0	0	25	4	14	41
Phenanthrene/Anthracene, C1 -	ng/L	0	0	24	0	0	10
Phenanthrene/Anthracene, C2 -	ng/L	0	0	24	0	0	10
Phenanthrene/Anthracene, C3 -	ng/L	0	0	24	0	0	10
Phenanthrene/Anthracene, C4 -	ng/L	0	0	24	0	0	10
Pyrene	ng/L	0	0	25	0	0	41
Trimethylnaphthalene, 2,3,5-	ng/L	0	0	24	0	0	40

		San Luis F	Rey	Carlsbad		
PCBs	Symbol Units	Mean SD	- <u>n</u>	Mean SD		n
PCB 005	ng/L	0	0 24	0	0	40
PCB 008	ng/L	0	0 24	0	0	40
PCB 015	ng/L	0	0 24	0	0	40
PCB 018	ng/L	0	0 24	0	0	40
PCB 027	ng/L	0	0 24	0	0	40
PCB 028	ng/L	0	0 24	0	0	40
PCB 029	ng/L	0	0 24	0	0	40
PCB 031	ng/L	0	0 24	0	0	40
PCB 033	ng/L	0	0 24	0	0	40
PCB 044	ng/L	0	0 24	0	0	40
PCB 049	ng/L	0	0 24	0	0	40
PCB 052	ng/L	0	0 24	0	0	40
PCB 056	ng/L	0	0 24	0	0	40
PCB 060	ng/L	0	0 24	0	0	40
PCB 066	ng/L	0	0 24	0	0	40
PCB 070	ng/L	0	0 24	0	0	40
PCB 074	ng/L	0	0 24	0	0	40
PCB 087	ng/L	0	0 24	0	0	40
PCB 095	ng/L	0	0 24	0	0	40
PCB 097	ng/L	0	0 24	0	0	40
PCB 099	ng/L	0	0 24	0	0	40
PCB 101	ng/L	0	0 24	0	0	40
PCB 105	ng/L	0	0 24	0	0	40
PCB 110	ng/L	0	0 24	0	0	40
PCB 114	ng/L	0	0 24	0	0	40
PCB 118	ng/L	0	0 24	0	0	40
PCB 128	ng/L	0	0 24	0	0	40
PCB 137	ng/L	0	0 24	0	0	40
PCB 138	ng/L	0	0 24	0	0	40
PCB 141	ng/L	0	0 24	0	0	40
PCB 149	ng/L	0	0 24	0	0	40
PCB 151	ng/L	0	0 24	0	0	40
PCB 153	ng/L	0	0 24	0	0	40
PCB 156	ng/L	0	0 24	0	0	40
PCB 157	ng/L	0	0 24	0	0	40
PCB 158	ng/L	0	0 24	• 0	0	40
PCB 170	ng/L	0	0 24	0	0	40
PCB 174	ng/L	0	0 24	0	0	40
PCB 177	ng/L	0	0 24	0	0	40
PCB 180	ng/L	0	0 24	0	0	40
PCB 183	ng/L	0	0 24	0	0	40
PCB 187	ng/L	0	0 24	0	0	40
PCB 189	ng/L	0	0 24	0	0	40
PCB 194	ng/L	0	0 24	0	0	40
PCB 195	ng/L	0	0 24	0	0	40
PCB 200	ng/L	0	0 24	0	0	40
PCB 201	ng/L	0	0 24	0	0	40
PCB 203	ng/L	0	0 24	0	0	40
PCB 206	ng/L	0	0 24	0	0	40

Appendix, continued.		San Lu	is Rev		Ca	rlsbad
PCBs	Symbol Units	Mean S	D	n	Mean	SD n
PCB 209	ng/L	0	0	24	0	0 40
PCB-1016	ng/L	0		1	0	1
PCB-1221	ng/L	0		1	0	1
PCB-1232	ng/L	0		1	0	1
PCB-1242	ng/L	0		1	0	1
PCB-1248	ng/L	0		1	0	1
PCB-1254	ng/L	0		1	0	1
PCB-1260	ng/L	0		1	0	1
PCBs	ng/L	0	0	25	0	0 41
Pesticides	5					
Aldrin	na/L	0	0	25	0.05	0.22 41
alpha-BHC	ng/L	0		1	0	1
Ametryn	ng/L	0	0	24	0	0 40
Aspon	ng/l	0	0	24	0	0 40
Atraton	ng/l	0	0	24	0	0 40
Atrazine	ng/L	8 13	39 80	24	11 88	27 66 40
Azinphos ethyl	ng/L	0.10	00.00	24	0	0 40
Azinphos methyl	ng/L	0	Ő	24	1 00	6.32 40
beta-BHC	ng/L	0	Ŭ	1	0	1
Bolstar	ng/L	0	0	24	0 0	0 40
Carbonhenothion	ng/L	0	Ő	24	4 00	12 15 40
Chlordane (tech)	ng/L	0	Ū	1	0	11
Chlordane cis-	ng/L	0.04	0.20	24	0.03	0 16 40
Chlordane trans-	ng/L	0.01	0.20	24	0.00	0.10 10
Chlordene alpha-	ng/L	0	0	24	0	0 40
Chlordene, damma-	ng/L	0	0	24	0 35	1 44 40
Chlorfenvinnhos	ng/L	0	0	24	0.00	0 40
Chlorovrifos	ng/L	0	0	24	0	0 40
Chlorpyritos methyl	ng/L	0	0	24	0	0 40
Ciodrin	ng/L	0	0	24	0	0 40
Coumanhos	ng/L	0	0	24	0	0 40
Dacthal	ng/L	0	0	24	0 23	0 49 40
	ng/L	0	0	24	0.20	0.45 40
DDD(0,p)	ng/L	0	0	25	0.00	0.33 40
DDE(p,p)	ng/L	0	0	24	0.00	0.22 41
DDE(0,p)	ng/L	0 04	0 20	25	1 37	2 24 41
DDE(p,p)	ng/L	0.04	0.20	20	1.07	2.24 41
DDT(o, p')	ng/L	0	0	24	0	0 40
DDT(0,p)	ng/L	0	0	24	0 44	
DDT _e	ng/L	0 04	0 20	25	1 03	2 70 /1
delta BHC	ng/L	0.04	0.20	20	1.55	2.75 41
Demeton s	ng/L	0	0	2/	5.00	13 /0 /0
Diazinon	ng/L	0.67	243	24	68 30	101 40 40
Dichlofenthion	ng/L	0.07	2.40	24	00.00	0 40
Dichloryos	ng/L	0	0	24	0	0 40
Dicrotophos	ng/L	0	0	24	1 00	632 40
Dieldrin	ng/⊑	0	0	25	0.02	0.52 40
Direction	ng/⊑	0	0	20	0.0Z	14 46 40
Diovathion	ng/⊑	0	0	24 24	0.00 4 00	12 15 40
DIOXALIIIUII	IIY/L	U	U	4 4	+.00	12.10 40

		San Luis Rev			Carlsbad		
Pesticides	Symbol Units	Mean SE	<u>)</u> ı	n	Mean	SD n	
Disulfoton	ng/L	0	0	24	33.33	38.29 40	
Endosulfan I	ng/L	0	0	25	0.05	0.22 41	
Endosulfan II	ng/L	0	0	25	0.43	1.31 41	
Endosulfan sulfate	ng/L	0	0	25	0.02	0.16 41	
Endrin	ng/L	0	0	25	0.08	0.37 41	
Endrin Aldehyde	ng/L	0	0	25	0.51	1.73 41	
Endrin Ketone	ng/L	0	0	24	0	0 40	
Ethion	ng/L	0	0	24	0	0 40	
Ethoprop	ng/L	0	0	24	0	0 40	
Famphur	ng/L	0	0	24	0	0 40	
Fenchlorphos	ng/L	0	0	24	0	0 40	
Fenitrothion	ng/L	0	0	24	0	0 40	
Fensulfothion	ng/L	0	0	24	0	0 40	
Fenthion	ng/L	0	0	24	0	0 40	
Fonofos	ng/L	0	0	24	0	0 40	
gamma-BHC (Lindane)	ng/L	0		1	0	1	
HCH, alpha	ng/L	0	0	24	0.33	0.88 40	
HCH, beta	ng/L	0	0	24	0.46	2.54 40	
HCH, delta	ng/L	0	0	24	0.15	0.43 40	
HCH, gamma	ng/L	0	0	24	0.08	0.35 40	
Heptachlor	ng/L	0	0	25	0	0 41	
Heptachlor epoxide	ng/L	0	0	25	0	0 41	
Hexachlorobenzene	ng/L	0	0	24	0.12	0.27 40	
Leptophos	ng/L	0	0	24	0	0 40	
Malathion	ng/L	0	0	24	0.93	5.85 40	
Merphos	ng/L	0	0	24	0	0 40	
Methidathion	ng/L	0	0	24	1.00	6.32 40	
Methoxychlor	ng/L	0	0	25	0.02	0.16 41	
Mevinphos	ng/L	0	0	24	5.00	13.40 40	
Mirex	ng/L	0	0	24	0	0 40	
Molinate	ng/L	0	0	24	0	0 40	
Naled	ng/L	0	0	24	5.00	13.40 40	
Nonachlor, cis-	ng/L	0	0	24	0	0 40	
Nonachlor, trans-	ng/L	0	0	24	0.05	0.22 40	
Oxadiazon	ng/L	5.08	8.87	24	64.74	292.81 40	
Oxychlordane	ng/L	0	0	24	0	0 40	
Parathion, Ethyl	ng/L	0	0	24	0	0 40	
Parathion, Methyl	ng/L	0	0	24	0.75	4.74 40	
Phorate	ng/L	0	0	24	0	0 40	
Phosmet	ng/L	0	0	24	0	0 40	
Phosphamidon	ng/L	0	0	24	0	0 40	
Prometon	ng/L	0	0	24	7.18	20.81 40	
Prometryn	ng/L	0	0	24	0	0 40	
Propazine	ng/L	0	0	24	7.00	14.18 40	
Secbumeton	ng/L	4.50	12.50	24	85.00	153.57 40	
Simazine	ng/L	19.17	60.61	24	0	0 40	
Simetryn	ng/L	0	0	24	0	0 40	
Sulfotep	ng/L	0	0	24	0	0 40	
Tedion	ng/L	0	0	24	0	0 40	

repondix, continuoui							
		San Luis Rey			ey Carlsbad		
Pesticides	Symbol Units	Mean	SD	n	Mean	SD n	
Terbufos	ng/L	0	0	24	0	0 40	
Terbuthylazine	ng/L	7.29	15.43	24	242.40	488.92 40	
Terbutryn	ng/L	0	0	24	0	0 40	
Tetrachlorvinphos	ng/L	0	0	24	1.00	6.32 40	
Thiobencarb	ng/L	0	0	24	0	0 40	
Thionazin	ng/L	0	0	24	0	0 40	
Tokuthion	ng/L	0	0	24	0	0 40	
Toxaphene	ng/L	0		1	0	1	
Trichlorfon	ng/L	0	0	24	0	0 40	
Trichloronate	ng/L	0	0	24	0	0 40	

Appendix, continued.			San Die	otito		Los	Peñasqui	tos
Physical water quality and inorganics	Symbol	Units	Mean SE))	n	Mean	SD	n
Alkalinity as CaCO3	Alk	ma/l	243	129	18	221	88	20
Chloride		ma/l			0			0
Fine-ASTM		%			0	33.3	31.8	13
Fine-ASTM, Passing No. 200 Sieve		%	18.1	7.1	7	32.2	53.0	3
Oxygen, Dissolved	DO	mg/L			0			0
Oxygen, Saturation		%	86	19	17	107	34	19
pH		pН	8.0	0.4	17	7.9	0.4	19
Salinity	Sal	ppt	2.53	5.47	17			0
Specific conductivity	Cond	µS/cm	4316	8781	17	2981	1256	19
Sulfate	SO4	mg/l	358	367	18	674	407	20
Suspended Sediment Concentration		%			0			0
Temperature		°C	15.1	6.0	17	19.4	4.4	19
Total Organic Carbon		mg/L			0			0
Total Suspended Solids		mg/L	18	23	18	0		1
Turbidity		NŤU	5.9	5.3	17	14.5	26.2	19
Velocity		ft/s	0.3	0.5	18	1.1	1.5	20
Nutrients								
Ammonia as N	NH ₃ -N	mg/l	0.10	0.12	18	0.09	0.06	20
Nitrate + Nitrite as N		mg/l	0.44	0.60	18	0.52	0.70	20
Nitrate as N		mg/l	0.42	0.58	18	0		1
Nitrate as NO3	NO_3	mg/l	1.96	2.58	17	0		1
Nitrite as N		mg/l	0.02	0.03	18	0		1
Nitrogen, Total Kjeldahl		mg/l	1.08	0.81	18	0.58	0.55	20
OrthoPhosphate as P		mg/l	0.16	0.15	18	0.04	0.04	20
Phosphorus as P,Total	TP	mg/l	0.24	0.35	18	0.10	0.16	20
Metals		-						
Aluminum		µg/L	3.9	5.4	18	5.7	7.5	20
Arsenic	As	µg/L	2.1	1.7	18	3.4	0.8	20
Cadmium	Cd	µg/L	0.03	0.03	18	0.02	0.01	20
Chromium	Cr	µg/L	0.18	0.17	18	0.89	0.97	20
Copper	Cu	µg/L	2.41	1.76	18	4.03	1.58	20
Lead	Pb	µg/L	0.03	0.03	18	0.05	0.08	20
Manganese	Mn	µg/L	135	135	18	141	156	20
Nickel	Ni	µg/L	0.70	0.79	18	3.38	3.55	20
Selenium	Se	µg/L	3.7	5.4	17	7.8	3.7	19
Silver	Ag	µg/L	0.00	0.00	18	0.06	0.25	20
Zinc	Zi	µg/L	2.2	1.7	17	8.4	8.7	19
Bacteria								
Enterococcus		MPN/100 ml	2400		1	11		1
Fecal Coliform		MPN/100 ml	240		1	500		1
Total Coliform		MPN/100 ml	1600		1	1600		1

Appendix, continued.		San [Dieguito		Los P	eñasquit	tos
PAHs	Symbol Units	Mean S	SD	n	Mean S	D	n
Acenaphthene	ng/L	0	0	18	0	0	20
Acenaphthylene	ng/L	0	0	18	0	0	20
Anthracene	ng/L	0	0	18	0	0	20
Benz(a)anthracene	ng/L	0	0	18	0	0	20
Benzo(a)pyrene	ng/L	1.0	4.4	18	0	0	20
Benzo(b)fluoranthene	ng/L	3.0	5.9	18	0	0	20
Benzo(e)pyrene	ng/L	0.9	4.0	18	0	0	20
Benzo(g,h,i)perylene	ng/L	10.1	42.9	18	0	0	20
Benzo(k)fluoranthene	ng/L	0	0	17	0	0	19
Biphenyl	ng/L	0	0	17	0	0	19
Chrysene	ng/L	0	0	18	0	0	20
Chrysenes, C1 -	ng/L	0	0	17	0	0	4
Chrysenes, C2 -	ng/L	3.9	16.2	17	0	0	4
Chrysenes, C3 -	ng/L	0.7	3.0	17	0	0	4
Dibenz(a,h)anthracene	ng/L	0	0	18	0	0	20
Dibenzothiophene	ng/L	0	0	17	0	0	4
Dibenzothiophenes, C1 -	ng/L	6.6	8.4	17	0	0	4
Dibenzothiophenes, C2 -	ng/L	13.6	13.0	17	0	0	4
Dibenzothiophenes, C3 -	ng/L	4.5	9.9	17	0	0	4
Dimethylnaphthalene, 2,6-	ng/L	2	8	17	0	0	19
Dimethylphenanthrene, 3,6-	ng/L			0			0
Fluoranthene	ng/L	6.99	24.36	18	0	0	20
Fluoranthene/Pyrenes, C1 -	ng/L	0.63	2.60	17	0	0	4
Fluorene	ng/L	0	0	18	0	0	20
Fluorenes, C1 -	ng/L	1.29	3.65	17	0	0	4
Fluorenes, C2 -	ng/L	0	0	17	27.93	2.53	4
Fluorenes, C3 -	ng/L	2.93	5.49	17	8.28	16.55	4
Indeno(1,2,3-c,d)pyrene	ng/L	2.93	12.42	18	0	0	20
Methyldibenzothiophene, 4-	ng/L			0			0
Methylfluoranthene, 2-	ng/L			0			0
Methylfluorene, 1-	ng/L			0			0
Methylnaphthalene, 1-	ng/L	0	0	17	0	0	19
Methylnaphthalene, 2-	ng/L	0	0	17	0	0	19
Methylphenanthrene, 1-	ng/L	1	6	17	0	0	19
Naphthalene	ng/L	0	0	18	1.75	7.83	20
Naphthalenes, C1 -	ng/L	0	0	17	0	0	4
Naphthalenes, C2 -	ng/L	2.81	11.57	17	0	0	4
Naphthalenes, C3 -	ng/L	2.06	5.82	17	5.45	10.90	4
Naphthalenes, C4 -	ng/L	1.25	3.52	17	0	0	4
Perylene	ng/L	0	0	17	0	0	19
Phenanthrene	ng/L	2	11	18	0	0	20
Phenanthrene/Anthracene, C1 -	ng/L	7.96	15.35	17	0	0	4
Phenanthrene/Anthracene, C2 -	ng/L	3.74	7.48	17	0	0	4
Phenanthrene/Anthracene, C3 -	ng/L	3.28	10.12	17	0	0	4
Phenanthrene/Anthracene, C4 -	ng/L	0.64	2.64	17	0	0	4
Pyrene	ng/L	24.61	82.69	18	0	0	20
Trimethylnaphthalene, 2,3,5-	ng/L	0	0	17	0	0	19

		San Diegu	ito	Los Peña	squitos
PCBs	Symbol Units	Mean SD	n	Mean SD	n
PCB 005	ng/L	0	0 17	0	0 19
PCB 008	ng/L	0	0 17	0	0 19
PCB 015	ng/L	0	0 17	0	0 19
PCB 018	ng/L	0	0 17	0	0 19
PCB 027	ng/L	0	0 17	0	0 19
PCB 028	ng/L	0	0 17	0	0 19
PCB 029	ng/L	0	0 17	0	0 19
PCB 031	ng/L	0	0 17	0	0 19
PCB 033	ng/L	0	0 17	0	0 19
PCB 044	ng/L	0	0 17	0	0 19
PCB 049	ng/L	0	0 17	0	0 19
PCB 052	ng/L	0	0 17	0	0 19
PCB 056	ng/L	0	0 17	0	0 19
PCB 060	ng/L	0	0 17	0	0 19
PCB 066	ng/L	0	0 17	0	0 19
PCB 070	ng/L	0	0 17	0	0 19
PCB 074	ng/L	0	0 17	0	0 19
PCB 087	ng/L	0	0 17	0	0 19
PCB 095	ng/L	0	0 17	0	0 19
PCB 097	ng/L	0	0 17	0	0 19
PCB 099	ng/L	0	0 17	0	0 19
PCB 101	ng/L	0	0 17	0	0 19
PCB 105	ng/L	0	0 17	0	0 19
PCB 110	ng/L	0	0 17	0	0 19
PCB 114	ng/L	0	0 17	0	0 19
PCB 118	ng/L	0	0 17	0	0 19
PCB 128	ng/L	0	0 17	0	0 19
PCB 137	ng/L	0	0 17	0	0 19
PCB 138	ng/L	0	0 17	0	0 19
PCB 141	ng/L	0	0 17	0	0 19
PCB 149	ng/L	0	0 17	0	0 19
PCB 151	ng/L	0	0 17	0	0 19
PCB 153	ng/L	0	0 17	0	0 19
PCB 156	ng/L	0	0 17	0	0 19
PCB 157	ng/L	0	0 17	0	0 19
PCB 158	ng/L	0	0 17	0	0 19
PCB 170	ng/L	0	0 17	0	0 19
PCB 174	ng/L	0	0 17	0	0 19
PCB 177	ng/L	0	0 17	0	0 19
PCB 180	ng/L	0	0 17	0	0 19
PCB 183	ng/L	0	0 17	0	0 19
PCB 187	ng/L	0	0 17	0	0 19
PCB 189	ng/L	0	0 17	0	0 19
PCB 194	ng/L	0	0 17	0	0 19
PCB 195	ng/L	0	0 17	0	0 19
PCB 200	ng/L	0	0 17	0	0 19
PCB 201	ng/L	0	0 17	0	0 19
PCB 203	ng/L	0	0 17	0	0 19
PCB 206	na/l	0	0 17	0	0 19

		San D	ieguito	Los	Peñasquitos
PCBs	Symbol Units	Mean S	SD n	Mean	SD n
PCB 209	ng/L	0	0 17	0	0 19
PCB-1016	ng/L	0	1	0	1
PCB-1221	ng/L	0	1	0	1
PCB-1232	ng/L	0	1	0	1
PCB-1242	ng/L	0	1	0	1
PCB-1248	ng/L	0	1	0	1
PCB-1254	ng/L	0	1	0	1
PCB-1260	ng/L	0	1	0	1
PCBs	ng/L	0	0 18	s 0	0 20
Pesticides	-				
Aldrin	ng/L	0	0 18	0.15	0.67 20
alpha-BHC	ng/L	0	1	0	1
Ametryn	ng/L	0	0 17	, O	0 19
Aspon	ng/L	0	0 17	' Û	0 19
Atraton	ng/L	0	0 17	6.84	29.82 19
Atrazine	ng/L	0	0 17	28.68	35.70 19
Azinphos ethvl	ng/L	0	0 17	′ 0	0 19
Azinphos methyl	ng/L	0	0 17	4.21	12.61 19
beta-BHC	ng/l	0	1	0	1
Bolstar	ng/l	0	0 17	· 0	0 19
Carbophenothion	ng/l	0	0 17	9.37	19.00 19
Chlordane (tech)	ng/l	0	1	0	1
Chlordane cis-	ng/L	0.06	0 24 17	, o	0 19
Chlordane trans-	ng/L	0.00	0.21 17	, 0	0 19
Chlordene alpha-	ng/L	0 0	0 17	0.63	2 75 19
Chlordene, damma-	ng/L	0	0 17	0.00 0.37	1 16 19
Chlorfenvinnhos	ng/L	0	0 17	' 0.07	0 19
Chlorovrifos	ng/L	0	0 17	, 0	0 19
Chlorpyrifos methyl	ng/L	0	0 17	, 0	0 19
Ciodrin	ng/L	0	0 17	, 0	0 19
Coumanhos	ng/L	0	0 17	263	11 47 10
Dacthal	ng/L	0	0 17	2.00 0.16	0 37 19
	ng/L	0	0 17	· 0.10	0.07 10
DDD(0,p)	ng/L	0	0 12	0.05	0 22 20
DDE(p,p)	ng/L	0	0 17	/ 0.00 / 0	0.22 20
DDE(0,p)	ng/L	0 17	0.51.18	530	12 71 20
DDE(p,p)	ng/L	0.17	0.01 10	, 0.00 , 0	0 10
DDMO(p,p')	ng/L	0	0 17	, 0	0 19
DDT(0,p)	ng/L	0 11	0 47 19	0 30	0 02 20
	ng/L	0.11	0.47 10	0.30 5 5 65	12 75 20
dolta PHC	ng/L	0.20	0.30 10	, J.05 0	12.75 20
	ng/L	0	0 17	, U	0 10
Diazinon	ng/L	12.64	14 00 17	7 50 91	51 65 10
Diablefonthion	ng/L	12.04	14.90 17	, 00.01	0 10
Dichloruco	ng/L	0	0 17	, U	0 19
Diction vos	ng/L	0		U דריד י	17 00 10
Diciolophos	ng/L	0 11		1.31	17.90 19
Directhooto	ng/L	0.11	0.32 18	0 U	0 20
Dimethoate	ng/L	0	0 17	24.08	30.77 19
Dioxathion	ng/L	U	0 17	10.53	10.10 19

		San D	Dieguito		Los	Peñasquit	os
Pesticides	Symbol Units	Mean S	SD	n	Mean	SD	n
Disulfoton	ng/L	0	0	17	52.92	64.68	19
Endosulfan I	ng/L	0.06	0.24	18	0.05	0.22	20
Endosulfan II	ng/L	0	0	18	0.54	1.54	20
Endosulfan sulfate	ng/L	0.06	0.24	18	0.05	0.22	20
Endrin	ng/L	0	0	18	0.05	0.22	20
Endrin Aldehyde	ng/L	0	0	18	0.15	0.67	20
Endrin Ketone	ng/L	0	0	17	0	0	19
Ethion	ng/L	0	0	17	0	0	19
Ethoprop	ng/L	0	0	17	2.11	9.18	19
Famphur	ng/L	0	0	17	0	0	19
Fenchlorphos	ng/L	0	0	17	0	0	19
Fenitrothion	ng/L	0	0	17	0	0	19
Fensulfothion	ng/L	0	0	17	0	0	19
Fenthion	ng/L	0	0	17	4.21	12.61	19
Fonofos	ng/l	1.57	6.48	17	0	0	19
gamma-BHC (Lindane)	ng/l	0	00	1	0	•	1
HCH alpha	ng/l	0	0	17	2 05	8 71	19
HCH beta	ng/l	0	Ő	17	0.42	1 39	19
HCH delta	ng/l	0.06	0 24	17	0.05	0.23	19
HCH gamma	ng/l	0.06	0.24	17	0.05	0.23	19
Hentachlor	ng/L	0.00	0.21	18	0.00	0.20	20
Heptachlor enoxide	ng/L	0 11	0 32	18	0	0	20
Hexachlorobenzene	ng/L	0.11	0.02	17	0 15	0.29	10
Lentonhos	ng/L	0	0	17	0.10	0.20	10
Malathion	ng/L	0	0	17	18 95	82 59	10
Mernhos	ng/L	0	0	17	10.33	02.00	10
Methidathion	ng/L	0	0	17	0	0	10
Methoxychlor	ng/L	0	0	10	0.05	0 22	20
Methoxychiol	ng/L	0	0	17	8.07	17.07	10
Mirey	ng/L	0 06	0.24	17	0.97	17.97	10
Molipato	ng/L	0.00	0.24	17	15 70	27.46	10
Naled	ng/L	0	0	17	10.79	18 10	10
Nonachlor cis	ng/L	0	0	17	10.55	10.10	10
Nonachior, cis-	ng/L	0	0	17	0	0	10
Avadiazan	ng/L	0 8 6 5	15.04	17	47.01	44.26	10
Oxadiazon	ng/L	0.00	15.04	17	47.01	44.20	10
Derothion Ethyl	ng/L	0	0	17	2 1 1	0 10	19
Parathion, Eury	ng/L	0	0	17	2.11	9.10	19
Paratinon, Mennyi	ng/L	0	0	17	0.20	23.47	19
Phorate	ng/L	0	0	17	0	0	19
Phosmet	ng/L	0	0	17	0	0	19
Phosphamidon	ng/L	0	0	17	0	0	19
Prometon	ng/L	0	0	17	0	0	19
Prometryn	ng/L	0	0	17	0	0	19
Propazine	ng/L	0	0	17	17.11	35.25	19
Second	ng/L	0	0	17	131.16	147.98	19
Simazine	ng/L	0	0	17	0	0	19
Simetryn	ng/L	0	0	17	0	0	19
Sulfotep	ng/L	0	0	17	0	0	19
ledion	ng/L	0.12	0.33	17	0	0	19

		San Dieg	uito	Los Peñasquitos			
Pesticides	Symbol Units	Mean SD	n	Mean	SD n		
Terbufos	ng/L	0	0 17	0	0 19		
Terbuthylazine	ng/L	0	0 17	380.37	373.84 19		
Terbutryn	ng/L	0	0 17	0	0 19		
Tetrachlorvinphos	ng/L	0	0 17	0	0 19		
Thiobencarb	ng/L	0	0 17	69.74	218.99 19		
Thionazin	ng/L	0	0 17	0	0 19		
Tokuthion	ng/L	0	0 17	2.11	9.18 19		
Toxaphene	ng/L	0	1	0	1		
Trichlorfon	ng/L	0	0 17	0	0 19		
Trichloronate	ng/L	0	0 17	2.11	9.18 19		

Appendix, continued.			Sar	n Diego		Pueblo	San Diego
Physical water quality and inorganics	Symbol	Units	Mean	SD	n	Mean	SD n
Alkalinity as CaCO3	Alk	ma/l	242	85	27	191	15 5
Chloride		mg/l	330		1		0
Fine-ASTM		%			0		0
Fine-ASTM, Passing No. 200 Sieve		%			0		0
Oxygen, Dissolved	DO	mg/L	9.5	3.0	26	113.3	198.9 4
Oxygen, Saturation		%	102	32	26	175	98 4
pH		pН	8.0	0.3	26	8.8	0.8 4
Salinity	Sal	ppt	1.00	0.45	26	4.19	5.66 4
Specific conductivity	Cond	µS/cm	1872	830	26	6925	9619 4
Sulfate	SO ₄	mg/l	283	159	27	437	394 5
Suspended Sediment Concentration		%			0		0
Temperature		°C	19.3	4.1	26	25.7	5.4 4
Total Organic Carbon		mg/L			0		0
Total Suspended Solids		mg/L	34	104	27	38	42 5
Turbidity		NŤU	7.2	8.9	26	6.2	7.4 4
Velocity		ft/s	0.8	1.1	27	4.0	8.9 5
Nutrients							
Ammonia as N	NH ₃ -N	mg/l	0.05	0.05	27	0.05	0.05 5
Nitrate + Nitrite as N		mg/l	15.64	57.07	27	0.41	0.47 5
Nitrate as N		mg/l	15.62	57.08	27	0.39	0.45 5
Nitrate as NO3	NO ₃	mg/l			0	0	1
Nitrite as N		ma/l	0.03	0.04	27	0.02	0.02 5
Nitrogen, Total Kieldahl		mg/l	0.79	0.36	26	1.81	0.45 5
OrthoPhosphate as P		mg/l	0.10		1	32.00	1
Phosphorus as P,Total	TP	mg/l	0.12	0.09	27	0.25	0.07 5
Metals		•					
Aluminum		µg/L	7.9	9.0	27	7.4	9.4 5
Arsenic	As	µg/L	3.2	2.4	27	2.5	1.5 5
Cadmium	Cd	µg/L	0.03	0.02	27	0.08	0.05 5
Chromium	Cr	µg/L	0.64	0.74	27	1.22	0.79 5
Copper	Cu	µg/L	4.16	2.06	27	8.23	3.94 5
Lead	Pb	µg/L	0.09	0.08	27	0.51	0.28 5
Manganese	Mn	µg/L	60	116	27	61	67 5
Nickel	Ni	µg/L	1.15	1.92	27	3.99	2.81 5
Selenium	Se	µg/L	8.1	6.8	26	77.5	115.8 4
Silver	Ag	µg/L	0.00	0.00	27	0.13	0.27 5
Zinc	Zi	µg/L	3.9	2.3	26	13.6	4.1 4
Bacteria							
Enterococcus		MPN/100 ml	520		1	11	1
Fecal Coliform		MPN/100 ml	900		1	900	1
Total Coliform		MPN/100 ml	1600		1	1600	1

		San	Diego		Pueblo	San Diego
PAHs	Symbol Units	Mean S	SD	n	Mean	SD n
Acenaphthene	ng/L	1	3	27	0	05
Acenaphthylene	ng/L	0	0	27	0	05
Anthracene	ng/L	0	0	27	0	05
Benz(a)anthracene	ng/L	0	2	27	3	75
Benzo(a)pyrene	ng/L	0.6	3.0	27	4.6	10.3 5
Benzo(b)fluoranthene	ng/L	0.8	4.0	27	5.8	13.0 5
Benzo(e)pyrene	ng/L	0.4	2.3	27	0	05
Benzo(g,h,i)perylene	ng/L	0.8	4.3	27	5.9	13.2 5
Benzo(k)fluoranthene	ng/L	0	0	26	7.3	14.5 4
Biphenyl	ng/L	0	0	26	0	04
Chrysene	ng/L	0.3	1.5	27	4.3	9.6 5
Chrysenes, C1 -	ng/L	0	0	26	0	04
Chrysenes, C2 -	ng/L	0	0	26	0	04
Chrysenes. C3 -	na/L	0	0	26	0	04
Dibenz(a,h)anthracene	ng/L	0	0	27	6.4	14.4 5
Dibenzothiophene	na/L	0	0	26	3.7	7.5 4
Dibenzothiophenes. C1 -	na/L	0	0	26	18.4	32.9 4
Dibenzothiophenes, C2 -	na/L	0	0	26	35.6	65.5 4
Dibenzothiophenes, C3 -	ng/l	0	0	26	1.4	2.8.4
Dimethylnaphthalene, 2.6-	na/L	0	0	26	4	74
Dimethylphenanthrene, 3.6-	ng/l	0	0	19	0	04
Fluoranthene	ng/l	0.68	3.54	27	4.68	6.69 5
Fluoranthene/Pyrenes, C1 -	ng/l	0.60	3.04	26	0	0 4
Fluorene	ng/l	0	0	27	0	0.5
Fluorenes, C1 -	ng/L	0	0	26	0	04
Fluorenes C2 -	ng/l	0	0	26	0	0 4
Fluorenes C3 -	ng/L	0	0	26	17 38	34 75 4
Indeno(1.2.3-c.d)pyrene	ng/L	0.83	4 33	27	7 14	15 97 5
Methyldibenzothionhene 4-	ng/L	0.00	0	19	2.38	4 76 4
Methylfluoranthene 2-	ng/L	0	0	19	2.00	0.4
Methylfluorene 1-	ng/L	0	0	19	0	04
Methylnaphthalene 1-	ng/L	0	0	26	0	04
Methylnaphthalene, 7	ng/L	0	0	26	0	04
Methylphenanthrene 1-	ng/L	0	0	26	0	04
Nanhthalene	ng/L	0 19	0.97	27	1 23	2765
Naphthalenes C1 -	ng/L	0.10	0.07	26	0	0.4
Nanhthalenes C2 -	ng/L	0.51	1 80	26	4 93	9854
Naphthalenes, C2	ng/L	1 15	3 31	26	7.43	8 64 4
Nanhthalenes C4 -	ng/L	1.10	0.01	26	2 00	5 80 4
Dervlene	ng/L	0	0	20	2.30	0.00
Phenanthrana	ng/∟	0	1	20	5	75
Phenanthrene/Anthracene C1	ng/L	0.77	272	26	6 00	8 20 1
Phenanthrene/Anthracene, C1 -	ng/∟	0.77	2.12	20	11 00	14 07 4
Dhenanthrane/Anthracana C2	ny/L	0.03	2.23	20	2 55	5 10 4
Phononthrono/Anthrocono C4	ny/L	0	0	20	2.05	0.10 4
Pyrene	ng/L	0 77	2 0 S 0	20	1 /5	3 22 5
Trimethylnanhthalana 235	ny/L	0.77	J.90 A	26	0 0	J.2J J 0 4
	ny/∟	0	0	20	0	04

		San Dieg	0	Pueblo Sa	n Diego
PCBs	Symbol Units	Mean SD	n	Mean SE) n
PCB 005	ng/L	0	0 26	6 0	04
PCB 008	ng/L	0	0 26	6 0	04
PCB 015	ng/L	0	0 26	6 0	04
PCB 018	ng/L	0	0 26	6 0	04
PCB 027	ng/L	0	0 26	6 0	04
PCB 028	ng/L	0	0 26	6 0	04
PCB 029	ng/L	0	0 26	6 0	04
PCB 031	ng/L	0	0 26	6 0	04
PCB 033	ng/L	0	0 26	6 0	04
PCB 044	ng/L	0	0 26	6 0	04
PCB 049	ng/l	0	0 26	S 0	04
PCB 052	ng/l	0	0.26	S 0	04
PCB 056	ng/L	0 0	0.26	, 0 , 0	04
PCB 060	ng/L	ů 0	0.26	, e	04
PCB 066	ng/L	0	0 20	, 0 , 0	04
PCB 070	ng/L	0	0 20	, 0 , 0	04
PCB 074	ng/L	0	0 20	, 0 8 0	0 4
	ng/L	0	0 20		04
	ng/L	0	0 20		04
	ng/L	0	0 20		04
	ng/L	0	0 20		04
	ng/L	0	0 20		04
	ng/L	0	0 20		04
PCB 105	ng/L	0	0 20		04
PCB 110	ng/L	0	0.26	5 U	04
PCB 114	ng/L	0	0 20		04
PCB 118	ng/L	0	0.26	5 U	04
PCB 128	ng/L	0	0 26	5 U	04
PCB 137	ng/L	0	0 26	6 0	04
PCB 138	ng/L	0	0 26	6 0	04
PCB 141	ng/L	0	0 26	6 0	04
PCB 149	ng/L	0	0 26	6 0	04
PCB 151	ng/L	0	0 26	6 0	04
PCB 153	ng/L	0	0 26	6 0	04
PCB 156	ng/L	0	0 26	6 0	04
PCB 157	ng/L	0	0 26	6 0	04
PCB 158	ng/L	0	0 26	6 0	04
PCB 170	ng/L	0	026	6 0	04
PCB 174	ng/L	0	026	6 0	04
PCB 177	ng/L	0	026	6 0	04
PCB 180	ng/L	0	026	6 0	04
PCB 183	ng/L	0	0 26	6 0	04
PCB 187	ng/L	0	0 26	6 0	04
PCB 189	ng/L	0	0 26	6 0	04
PCB 194	ng/L	0	0 26	6 0	04
PCB 195	ng/L	0	0 26	6 0	04
PCB 200	ng/L	0	0 26	6 0	04
PCB 201	na/L	0	0 26	6 0	04
PCB 203	na/L	0	0 26	6 Ū	04
PCB 206	ng/l	0	0 26	s 0	04

Appendix, continued.		San Di	eao	Pueblo	San Diego
PCBs	Symbol Units	Mean SD	n n	Mean	SD n
PCB 209	ng/L	0	0 26	<u> </u>	04
PCB-1016	ng/L	0		0	1
PCB-1221	ng/L	0		0	1
PCB-1232	ng/L	0		0	1
PCB-1242	ng/L	0		0	1
PCB-1248	ng/L	0		0	1
PCB-1254	ng/L	0		0	1
PCB-1260	ng/L	0		0	1
PCBs	ng/L	0	0 27	' 0	05
Pesticides	-				
Aldrin	ng/L	0	0 27	' 0	05
alpha-BHC	ng/L	0		0	1
Ametryn	ng/L	0	0 26	6	0
Aspon	ng/L	0	0 26	6 0	04
Atraton	ng/L	0	0 26	6	0
Atrazine	ng/L	3.27	16.67 26	6	0
Azinphos ethyl	ng/L	0	0 26	6 0	04
Azinphos methyl	ng/L	0	0 26	6 0	04
beta-BHC	ng/L	0		0	1
Bolstar	ng/L	0	0 26	6 0	04
Carbophenothion	ng/L	0	0 26	6 0	04
Chlordane (tech)	ng/L	0		0	1
Chlordane, cis-	ng/L	0	0 26	6 0	04
Chlordane, trans-	ng/L	0	0 26	6 0	04
Chlordene, alpha-	ng/L	0	0 26	6 0	04
Chlordene, gamma-	ng/L	0	0 26	6 0	04
Chlorfenvinghos	ng/L	0	0 26	6 6	04
Chlorpyrifos	ng/L	0	0 26	6 6	04
Chlorpyrifos methyl	ng/L	0	0 26	6 6	04
Ciodrin	ng/L	0	0 26	6 0	04
Coumaphos	ng/L	0	0 26	6 6	04
Dacthal	ng/L	0	0 26	6 6	04
DDD(o,p')	ng/L	0	0 26	6 0	04
DDD(p,p')	ng/L	0	0 27	7 0	05
DDE(o,p')	ng/L	0	0 26	6 6	04
DDE(p,p')	ng/L	0	0 27	7 0	05
DDMU(p.p')	ng/L	0	0 26	6 6	04
DDT(o.p')	ng/L	0	0 26	6 6	04
DDT(p,p')	ng/L	0	0 27	7 0	05
DDTs	ng/L	0	0 27	7 0	05
delta-BHC	ng/L	0		0	1
Demeton-s	ng/L	0	0 26	6 0	04
Diazinon	ng/L	1.73	5.50 26	6 26.25	29.17 4
Dichlofenthion	ng/L	0	0 26	6 6	04
Dichlorvos	ng/L	0	0 26	6 0	04
Dicrotophos	ng/L	0	0 26	s 0	04
Dieldrin	ng/L	0	0 27	7 O	05
Dimethoate	ng/L	0	0 26	3 O	04
Dioxathion	ng/L	0	0 26	5 73.50	94.18 4

		San I	Diego	Pueblo	San Diego
Pesticides	Symbol Units	Mean S	Dn	Mean	SD n
Disulfoton	ng/L	0	0 26	13.25	26.50 4
Endosulfan I	ng/L	0	0 27	0	05
Endosulfan II	ng/L	0	0 27	0	05
Endosulfan sulfate	ng/L	0	0 27	0	05
Endrin	ng/L	0	0 27	0	05
Endrin Aldehyde	ng/L	0	0 27	0	05
Endrin Ketone	ng/L	0	0 26	0	04
Ethion	ng/L	0	0 26	0	04
Ethoprop	ng/L	0	0 26	0	04
Famphur	ng/L	0	0 26	0	04
Fenchlorphos	ng/L	0	0 26	0	04
Fenitrothion	ng/L	0	0 26	0	04
Fensulfothion	ng/L	0	0 26	0	04
Fenthion	ng/L	0	0 26	0	04
Fonofos	ng/L	0	0 26	0	04
gamma-BHC (Lindane)	ng/L	0	1	0	1
HCH. alpha	na/L	0	0 26	0	04
HCH, beta	na/L	0	0 26	0	04
HCH, delta	na/L	0	0 26	0	04
HCH, gamma	ng/L	0	0 26	0	04
Heptachlor	na/L	0	0 27	0	05
Heptachlor epoxide	na/L	0	0 27	0	05
Hexachlorobenzene	na/L	0	0 26	0	04
Leptophos	na/L	0	0 26	0	04
Malathion	na/L	1.27	6.47 26	0	04
Merphos	na/L	0	0 26	0	04
Methidathion	na/L	0	0 26	0	04
Methoxychlor	na/L	0	0 27	0	0 5
Mevinphos	na/L	0	0 26	0	04
Mirex	ng/L	0	0 26	0	04
Molinate	na/L	0	0 26	0	04
Naled	na/L	0	0 26	0	04
Nonachlor, cis-	na/L	0	0 26	0	04
Nonachlor, trans-	na/L	0	0 26	0	04
Oxadiazon	na/L	7.23	9.52 26	19.00	15.53 4
Oxvchlordane	ng/L	0	0 26	0	04
Parathion. Ethyl	na/L	0	0 26	0	04
Parathion, Methyl	na/L	0	0 26	0	04
Phorate	na/L	0	0 26	0	04
Phosmet	ng/L	0	0 26	0	04
Phosphamidon	na/L	0	0 26	0	04
Prometon	na/L	0	0 26		0
Prometryn	na/L	0	0 26		0
Propazine	na/L	8.23	26.98 26		0
Secbumeton	na/L	12.92	48.18 26		0
Simazine	na/L	10.46	25.68 26		0
Simetryn	na/L	0	0 26		0
Sulfotep	na/l	0	0 26	0	04
Tedion	ng/L	0	0 26	0	04

		San Diego			Pueblo San Diego		
Pesticides	Symbol Units	Mean	SD	n	Mean SD	n	
Terbufos	ng/L	0	0	26	0	04	
Terbuthylazine	ng/L	16.42	64.61	26		0	
Terbutryn	ng/L	0	0	26		0	
Tetrachlorvinphos	ng/L	0	0	26	0	04	
Thiobencarb	ng/L	4.46	22.75	26	0	04	
Thionazin	ng/L	0	0	26	0	04	
Tokuthion	ng/L	0	0	26	0	04	
Toxaphene	ng/L	0		1	0	1	
Trichlorfon	ng/L	0	0	26	0	04	
Trichloronate	ng/L	0	0	26	0	04	

- 			Swe	etwate	r	C	Dtav		Ti	juana	
Physical water quality and inorganics	Symbol	Units	Mean	SD	n	Mean	SD	n N	<i>lean</i>	SD	n
Alkalinity as CaCO3	Alk	mg/l	191	92	15	233	21	7	400	164	12
Chloride		mg/l	1740	622	2			0			0
Fine-ASTM		%			0			0			0
Fine-ASTM, Passing No. 200 Sieve		%			0	2.3		1			0
Oxygen, Dissolved	DO	mg/L	9.2	2.0	14			0	8.3	4.3	12
Oxygen, Saturation		%	99	25	14	123	31	5	90	51	12
рН		pН	8.0	0.4	14	7.8	0.3	5	7.9	0.7	12
Salinity	Sal	ppt	1.85	1.57	14	1.30	0.85	5	0.76	0.46	12
Specific conductivity	Cond	µS/cm	3930	3649	14	2478	1539	5	1482	887	12
Sulfate	SO_4	mg/l	276	190	15	217	40	7	201	133	12
Suspended Sediment Concentration		%			0			0			0
Temperature		°C	17.9	6.7	14	15.5	2.1	5	19.4	7.0	12
Total Organic Carbon		mg/L			0			0			0
Total Suspended Solids		mg/L	7	6	14	22	37	7	36	51	12
Turbidity		NŤU	4.1	5.0	14	2.5	1.4	5	12.3	13.3	12
Velocity		ft/s	1.1	1.2	15	1.2	1.8	6	0.8	1.2	12
Nutrients											
Ammonia as N	NH ₃ -N	mg/l	0.06	0.05	15	0.08	0.10	7	11.33	16.63	12
Nitrate + Nitrite as N		mg/l	5.21	8.04	15	2.11	2.11	7	0.23	0.32	12
Nitrate as N		mg/l	5.19	8.01	15	2.08	2.07	7	0.18	0.33	12
Nitrate as NO3	NO ₃	mg/l			0	9.21	9.14	7			0
Nitrite as N	Ū	ma/l	0.02	0.03	15	0.03	0.04	7	0.05	0.11	12
Nitrogen, Total Kieldahl		ma/l	0.72	0.35	15	0.73	0.37	7	13.00	15.53	12
OrthoPhosphate as P		ma/l	0.02		1	2.01	5.29	7			0
Phosphorus as P, Total	TP	mg/l	0.06	0.04	15	0.01	0.02	7	3.31	3.70	12
Metals		0									
Aluminum		µg/L	5.5	8.2	15	1.3	2.0	7	5.7	5.1	12
Arsenic	As	µg/L	11.5	16.8	15	7.7	6.0	7	3.7	2.6	12
Cadmium	Cd	µg/L	0.02	0.02	15	0.02	0.01	7	0.06	0.03	12
Chromium	Cr	µg/L	0.96	1.06	15	0.37	0.30	7	2.67	2.94	12
Copper	Cu	µg/L	4.25	3.05	15	2.91	1.16	7	5.14	6.25	12
Lead	Pb	µg/L	0.07	0.13	15	0.02	0.02	7	0.25	0.27	12
Manganese	Mn	µg/L	54	71	15	41	67	7	238	228	12
Nickel	Ni	µg/L	0.78	0.93	15	1.80	3.22	7	9.16	11.13	12
Selenium	Se	µg/L	26.6	27.8	14	9.2	7.3	6	7.2	4.6	12
Silver	Ag	µg/L	0.00	0.00	15	0.39	1.02	7	0.02	0.04	12
Zinc	Zi	µg/L	2.9	2.0	14	2.0	0.8	6	4.5	6.6	12
Bacteria											
Enterococcus		MPN/100 ml	2400		1	210		1			0
Fecal Coliform		MPN/100 ml	900		1	500		1			0
Total Coliform		MPN/100 ml	1600		1	1600		1			0

		Swe	Sweetwater		Otay		Т		Tijuana	
PAHs	Symbol Units	Mean	SD	n	Mean S	SD	n	Mean	SD	n
Acenaphthene	ng/L	0	0	15	0	0	7	1	4	12
Acenaphthylene	ng/L	0	0	15	0	0	7	1	4	12
Anthracene	ng/L	0	0	15	0	0	7	1	3	12
Benz(a)anthracene	ng/L	0	0	15	0	0	7	3	12	12
Benzo(a)pyrene	ng/L	0	0	15	0	0	7	6.6	23.0	12
Benzo(b)fluoranthene	ng/L	0	0	15	0	0	7	8.8	30.3	12
Benzo(e)pyrene	ng/L	0	0	15	0	0	7	12.2	42.1	12
Benzo(g,h,i)perylene	ng/L	0	0	15	0	0	7	13.8	47.6	12
Benzo(k)fluoranthene	ng/L	0	0	14	0	0	6	2.8	9.6	12
Biphenyl	ng/L	0	0	14	0	0	6	7	16	12
Chrysene	ng/L	0	0	15	0	0	7	9.7	33.5	12
Chrysenes, C1 -	ng/L	0	0	14	0	0	6	18.9	65.5	12
Chrysenes, C2 -	ng/L	0	0	14	0	0	6	24.3	78.5	12
Chrysenes, C3 -	ng/L	0	0	14	0	0	6	23.3	80.8	12
Dibenz(a,h)anthracene	ng/L	0	0	15	0	0	7	1.7	5.9	12
Dibenzothiophene	ng/L	0	0	14	0	0	6	3.5	9.8	12
Dibenzothiophenes, C1 -	ng/L	0	0	14	3.6	5.6	6	35.1	82.8	12
Dibenzothiophenes, C2 -	ng/L	0	0	14	7.4	8.6	6	95.5	271.1	12
Dibenzothiophenes, C3 -	ng/L	0	0	14	0	0	6	89.0	264.3	12
DimethyInaphthalene, 2,6-	ng/L	0	0	14	0	0	6	31	80	12
Dimethylphenanthrene, 3,6-	ng/L	0	0	14			0	14.02	43.42	12
Fluoranthene	ng/L	0.53	2.06	15	0	0	7	26.98	71.43	12
Fluoranthene/Pyrenes, C1 -	ng/L	0	0	14	0	0	6	58.74	177.90	12
Fluorene	ng/L	0	0	15	0	0	7	7	22	12
Fluorenes, C1 -	ng/L	0	0	14	0	0	6	43.72	121.07	12
Fluorenes, C2 -	ng/L	0	0	14	0	0	6	0	0	12
Fluorenes, C3 -	ng/L	0	0	14	0	0	6	0	0	12
Indeno(1,2,3-c,d)pyrene	ng/L	0	0	15	0	0	7	4.53	15.70	12
Methyldibenzothiophene, 4-	ng/L	0	0	14			0	12.58	31.73	12
Methylfluoranthene, 2-	ng/L	0	0	14			0	5.89	20.41	12
Methylfluorene, 1-	ng/L	0	0	14			0	16.57	44.06	12
Methylnaphthalene, 1-	ng/L	0	0	14	0	0	6	10	32	12
Methylnaphthalene, 2-	ng/L	0	0	14	0	0	6	15	51	12
Methylphenanthrene, 1-	ng/L	0	0	14	0	0	6	11	29	12
Naphthalene	ng/L	0.47	1.82	15	0	0	7	9.83	34.06	12
Naphthalenes, C1 -	ng/L	0	0	14	0	0	6	26.53	84.37	12
Naphthalenes, C2 -	ng/L	0	0	14	0	0	6	83.41	240.10	12
Naphthalenes, C3 -	ng/L	0	0	14	0	0	6	155.11	427.76	12
Naphthalenes, C4 -	ng/L	0	0	14	0	0	6	48.74	120.60	12
Perylene	ng/L	0	0	14	0	0	6	2.58	8.95	12
Phenanthrene	ng/L	0	0	15	0	0	7	26	57	12
Phenanthrene/Anthracene, C1 -	ng/L	0.36	1.34	14	0	0	6	69.21	157.09	12
Phenanthrene/Anthracene, C2 -	ng/L	0.41	1.53	14	0	0	6	157.18	438.58	12
Phenanthrene/Anthracene, C3 -	ng/L	0	0	14	0	0	6	109.16	347.01	12
Phenanthrene/Anthracene, C4 -	ng/L	0	0	14	0	0	6	27.80	90.90	12
Pyrene	ng/L	0.44	1.70	15	0	0	7	26.23	70.99	12
Trimethylnaphthalene, 2,3,5-	na/l	0	0	14	0	0	6	19	40	12

Ap	pendix.	continued.
· • P	ponany	oomaaaa

		Sweetw	Sweetwater		weetwater Otay		Tijuan		а	
PCBs	Symbol Units	Mean SD	n	Mean SD	n Mea	n SD		n		
PCB 005	ng/L	0	0 14	0	06	0	0	12		
PCB 008	ng/L	0	0 14	0	06	0	0	12		
PCB 015	ng/L	0	0 14	0	06	0	0	12		
PCB 018	ng/L	0	0 14	0	06	0	0	12		
PCB 027	ng/L	0	0 14	0	06	0	0	12		
PCB 028	ng/L	0	0 14	0	06	0	0	12		
PCB 029	ng/L	0	0 14	0	06	0	0	12		
PCB 031	ng/L	0	0 14	0	06	0	0	12		
PCB 033	ng/L	0	0 14	0	0 6	0	0	12		
PCB 044	ng/L	0	0 14	0	0 6	0	0	12		
PCB 049	ng/L	0	0 14	0	0 6	0	0	12		
PCB 052	ng/l	0	0 14	0	0.6	0	0	12		
PCB 056	ng/l	0 0	0 14	Ő	0.6	0	Õ	12		
PCB 060	ng/l	0 0	0 14	Ő	0.6	0	Õ	12		
PCB 066	ng/L	0 0	0 14	õ	0 6	Õ	õ	12		
PCB 070	ng/L	0	0 14	Õ	0.6	Õ	ñ	12		
PCB 074	ng/L	0	0 14	0	0 6	0	ñ	12		
PCB 087	ng/L	0	0 14	0	0 0	0	0	12		
	ng/L	0	0 14	0	0 0	0	0	12		
	ng/L	0	0 14	0	0 0	0	0	12		
	ng/L	0	0 14	0		0	0	12		
PCD 099	ng/L	0	0 14	0		0	0	12		
	ng/L	0	0 14	0	0 0	0	0	12		
	ng/L	0	0 14	0	0 0	0	0	12		
	ng/L	0	0 14	0	0 0	0	0	12		
PCB 114	ng/L	0	0 14	0	06	0	0	12		
PCB 118	ng/L	0	0 14	0	06	0	0	12		
PCB 128	ng/L	0	0 14	0	06	0	0	12		
PCB 137	ng/L	0	0 14	0	06	0	0	12		
PCB 138	ng/L	0	0 14	0	06	0	0	12		
PCB 141	ng/L	0	0 14	0	06	0	0	12		
PCB 149	ng/L	0	0 14	0	06	0	0	12		
PCB 151	ng/L	0	0 14	0	06	0	0	12		
PCB 153	ng/L	0	0 14	0	06	0	0	12		
PCB 156	ng/L	0	0 14	0	06	0	0	12		
PCB 157	ng/L	0	0 14	0	06	0	0	12		
PCB 158	ng/L	0	0 14	0	06	0	0	12		
PCB 170	ng/L	0	0 14	0	06	0	0	12		
PCB 174	ng/L	0	0 14	0	06	0	0	12		
PCB 177	ng/L	0	0 14	0	06	0	0	12		
PCB 180	ng/L	0	0 14	0	06	0	0	12		
PCB 183	ng/L	0	0 14	0	06	0	0	12		
PCB 187	ng/L	0	0 14	0	06	0	0	12		
PCB 189	ng/L	0	0 14	0	06	0	0	12		
PCB 194	ng/L	0	0 14	0	06	0	0	12		
PCB 195	ng/L	0	0 14	0	06	0	0	12		
PCB 200	ng/L	0	0 14	0	06	0	0	12		
PCB 201	na/L	0	0 14	0	06	0	0	12		
PCB 203	na/L	0	0 14	0	0 6	0	0	12		
PCB 206	na/L	0	0 14	0	0 6	0	0	12		

Annendix	continued
Appendix,	continueu.

		Swe	Sweetwater		Otay		Т	ijuana	
PCBs	Symbol Units	Mean	SD	n	Mean	SD	n Mean	SD	n
PCB 209	ng/L	0	() 14	0	0	60	0	12
PCB-1016	ng/L	0		1	0		1		0
PCB-1221	ng/L	0		1	0		1		0
PCB-1232	ng/L	0		1	0		1		0
PCB-1242	ng/L	0		1	0		1		0
PCB-1248	ng/L	0		1	0		1		0
PCB-1254	ng/L	0		1	0		1		0
PCB-1260	ng/L	0		1	0		1		0
PCBs	ng/L	0	() 15	0	0	7 0	0	12
Pesticides	Ū								
Aldrin	ng/L	0	() 15	0	0	7 0	0	12
alpha-BHC	na/L	0		1	0		1		0
Ametryn	na/L			0	0	0	6		0
Aspon	na/L	0	() 14	0	0	6 0	0) 12
Atraton	na/L			0	0	0	6		0
Atrazine	na/L			0	6.80	16.66	6		0
Azinphos ethyl	ng/l	0	() 14	0	0	6 0	0	12
Azinphos methyl	ng/l	0	() 14	Ő	0	6 0	0	12
beta-BHC	ng/l	0		1	Ő	· ·	1		0
Bolstar	ng/l	0	() 14	Ő	0	6 0	0	1 12
Carbonhenothion	ng/L	Ő	í) 14	Ő	0	6 0	0	12
Chlordane (tech)	ng/L	Ő		1	Ő	Ũ	1	0	0
Chlordane cis-	ng/L	Ő	(14	Ő	0	6 0	0	1 12
Chlordane trans-	ng/L	Ő	Ì) 14	Ő	0	6 0	0	12
Chlordene alpha-	ng/L	Ő	Č	14	0 0	0	6 0	0	1 12
Chlordene, damma-	ng/L	0	Ì	14	0	0	6 0	0	1 12
Chlorfenvinnhos	ng/L	0	Č	די ק 14 ה	0	0	6 0	0	12
Chlorpyrifos	ng/L	0	Ì	14	0	0	6 200	6 93	12
Chlorovrifos methyl	ng/L	0	, i	די כ 1/1 ר	0	0	6 <u>2.00</u>	0.00	12
Ciodrin	ng/L	0	, i	די כ 1/1 ר	0	0	6 0	0	12
Coumanhos	ng/L	0	, i	די כ 1/1 ר	0	0	6 0	0	12
Dacthal	ng/L	0	, i	די נ 14 ר	0	0	6 0	0	12
DDD(o p')	ng/L	0		די נ 1/1 ר	0	0	6 0	0	12
DDD(0,p')	ng/L	0		די ל 15 (0	0	0 0 7 0	0	12
DDE(p,p)	ng/L	0	, i	טי 11 ר	0	0	, 0 6 0	0	12
DDE(0,p)	ng/L	0	, i	די ל 15 (1/3	2 5 1	7 0	0	12
DDE(p,p)	ng/L	0	, i	טי 14 ר	1.45	2.51	7 0 6 0	0	12
DDT(a p')	ng/L	0		יי 14 ר	0	0	0 0 6 0	0	12
DDT(0,p)	ng/L	0) 1 4) 15	0.57	0 08	0 0 7 0	0	12
DDT _e	ng/L	0		15	2 00	2 83	7 0	0	12
delta RHC	ng/L	0	,	1	2.00	2.05	1 0	0	0
Domoton o	ng/L	0		ו 1/1 ר	0	0	ו ה ח	0	12
Diazinon	ng/L	11.26	11 10	יי 14 ב	20.02	26.47	0 0 6 1600	20.02	12
Diablefonthion	ng/L	11.30	11.13	יי 14 כ	20.03	20.47	6 10.92	20.93	12
Dichloride	ng/L	0		J 14	0	0		0	
DicritorVos	ng/L	0	(14 כ איז ב	0	U	0 U	0	
Diciolophos	ng/L	0	(14 ב אר	0	0	0 U 7 0	0	
Dieluilli	ng/L	0	(15 כ 14 כ	0	0	<i>i</i> 0	0	12
Dimethoate	ng/L	0		14 כ יייי	0	U		0	
Dioxathion	ng/L	34.43	52.78	314	0	U	o 96.67	231.08	- 12

		Swe	Sweetwater		Otay		Ti	ijuana		
Pesticides	Symbol Units	Mean	SD	n	Mean	SD	n	Mean	SD	n
Disulfoton	ng/L	4.14	15.50	14	0	() 6	16.17	40.76	12
Endosulfan I	ng/L	0	0	15	0	() 7	0	0	12
Endosulfan II	ng/L	0	0	15	0	() 7	0	0	12
Endosulfan sulfate	ng/L	0	0	15	0	() 7	0	0	12
Endrin	ng/L	0	0	15	0	() 7	0	0	12
Endrin Aldehyde	ng/L	0	0	15	0	() 7	0	0	12
Endrin Ketone	ng/L	0	0	14	0	() 6	0	0	12
Ethion	ng/L	0	0	14	0	() 6	0	0	12
Ethoprop	ng/L	0	0	14	0	() 6	0	0	12
Famphur	ng/L	0	0	14	0	() 6	0	0	12
Fenchlorphos	ng/L	0	0	14	0	() 6	0	0	12
Fenitrothion	na/L	0	0	14	0	() 6	0	0	12
Fensulfothion	na/L	0	0	14	0	Ć) 6	0	0	12
Fenthion	ng/L	0	0	14	0	() 6	0	0	12
Fonofos	ng/L	0	0	14	0	() 6	4.50	15.59	12
gamma-BHC (Lindane)	ng/l	0		1	0		1			0
HCH alpha	ng/l	0	0	14	Ő	() 6	0	0	12
HCH beta	ng/l	0	Ő	14	Ő	(6	0	0	12
HCH, delta	ng/l	0	Ő	14	Ő	() 6	0	0	12
HCH gamma	ng/L	0	0	14	Ő	(6 (0	0	12
Hentachlor	ng/L	0	0	15	0	() 7	Ő	0	12
Heptachlor enoxide	ng/L	0	0	15	0	() 7	Ő	0	12
Heyachlorobenzene	ng/L	0	0	14	0	(, ,)	0	0	12
Lentonhos	ng/L	0	0	14	0	(0	0	0	12
Malathion	ng/L	0	0	1/	0		0	0	0	12
Membos	ng/L	0	0	1/	0		0	0	0	12
Methidathion	ng/L	0	0	1/	0		0	0	0	12
Methoxychlor	ng/L	0	0	15	0		, U	0	0	12
Methoxychiol	ng/L	0	0	1/	0	(, , , ,	0	0	12
Miroy	ng/L	0	0	14	0		0	0	0	12
Molinato	ng/L	0	0	14	0		0	0	0	12
Neled	ng/L	0	0	14	0		0	0	0	12
Noncohlor ein	ng/L	0	0	14	0		0	0	0	12
Nonachior, trans	ng/L	0	0	14	0		0	0	0	12
Avadiazon	ng/L	11 14	11 50	14	20 22	20 12	0 6	0	0	12
Oxadiazon	ng/L	11.14	14.50	14	20.33	30.13	0 0	0	0	12
Derethion Ethyl	ng/L	0	0	14	0			0	0	12
Parathion, Ethyl	ng/L	0	0	14	0			0	0	12
Paratinon, Mennyi	ng/L	0	0	14	0		0	0	0	12
Phonate	ng/L	0	0	14	0			0	0	12
Phosphamidan	ng/L	0	0	14	0			0	0	12
Phosphamidon	ng/L	0	0	14	0	(0	0	0	12
Prometon	ng/L			0	0		0			0
Prometryn	ng/L			0	0	(0			0
Propazine	ng/∟			0	0	(0			0
Second	ng/L			0	0	(16			0
Simazine	ng/L			0	0	(16			0
Simetryn	ng/L	~	~	0	0	(16	~	~	0
Sulfotep	ng/L	0	0	14	0	(16	0	0	12
Iedion	ng/L	0	0	14	0	() 6	0	0	12
SWAMP Synthesis Report on Stream Assessments in the San Diego Region

· · · ·		Sweetwater		Otay	Tijuana		a	
Pesticides	Symbol Units	Mean SD	n	Mean SD	n	Mean SD		n
Terbufos	ng/L	0	0 14	0	06	0	0	12
Terbuthylazine	ng/L		0	0	06			0
Terbutryn	ng/L		0	0	06			0
Tetrachlorvinphos	ng/L	0	0 14	0	06	0	0	12
Thiobencarb	ng/L	0	0 14	0	06	0	0	12
Thionazin	ng/L	0	0 14	0	06	0	0	12
Tokuthion	ng/L	0	0 14	0	06	0	0	12
Toxaphene	ng/L	0	1	0	1			0
Trichlorfon	ng/L	0	0 14	0	06	0	0	12
Trichloronate	ng/L	0	0 14	0	06	0	0	12

Appendix, continued.