

CHEMICAL ANALYSIS, TOXICITY EVALUATION  
AND BIOACCUMULATION EXPOSURE  
OF SEDIMENTS FROM  
HUMBOLDT BAY:

**BASELINE SURVEY II**  
Fiscal Year 1994

**FINAL REPORT**

Prepared for:

U.S. ARMY ENGINEERING DISTRICT  
SAN FRANCISCO CORPS OF ENGINEERS  
San Francisco, California

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Watsonville, California

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THE UNIVERSITY OF CHICAGO

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CHEMICAL ANALYSIS, TOXICITY EVALUATION  
AND BIOACCUMULATION TESTING  
OF SEDIMENTS FROM  
HUMBOLDT BAY

**BASELINE SURVEY II**

**1.0 Introduction**

Under Contract No. DACW07-92-D-002 from San Francisco District, Army Corps of Engineers (SFACOE), ToxScan, Inc. collected and analyzed sediment samples from **Humboldt Bay** for FY 1994, **Baseline Survey II**. Sediments were sampled by Kinnetic Laboratories, Inc., and returned to the ToxScan, Inc. laboratory at Watsonville, CA where they were assigned laboratory number **T-10774** for physical, chemical and bioassay analyses. Bioaccumulation analyses were not performed on tissues exposed to these sediments. Samples collected, composites and analyses are summarized in Table 1.

**2.0 Methods**

**2.1 Sediment Collection**

Sediment sampling was conducted between 31 March and 5 April 1994 from the M/V Celtic and the M/V Sally Kae. Target sampling locations (California state plane coordinates) are listed in Table 1 of the Scope of Services provided by SFACOE (Appendix A, this report). The target locations were placed at or near sampling stations of Baseline Survey I (FY 1993) to establish an historical database of sediment characteristics at Humboldt Harbor. In some cases, sampling stations had to be moved from the target locations in order to sample areas with significant shoaling. Prior to initiating the field program each station's plane coordinates were converted to latitude/longitude coordinates to allow use of a differential Global Positioning System (GPS) for actual field positioning, and target locations were plotted on SFACOE-provided "blue line" pre-dredge survey charts to determine the site and approximate depth of each core. Actual sampling locations are plotted on Figures 1 through 4. Details of each core and grab sample (time collected, depth, location) are summarized in Table 2 and documented in field log sheets (Appendix B).

Horizontal positioning was established with a Trimble series 4000 Differential GPS navigation system with base station set on Corps of Engineers survey markers. Mudline elevations were determined at each core location at the time of sampling with a dual frequency fathometer calibrated by leadline to 0.1 feet at the expected depth range. Mean lower low water (MLLW) mudline elevations were extrapolated using Micronautics, Inc. Tide 1 software, and verified daily with USGS or COE benchmarks.

A preliminary field determination of sediment particle size distribution (PSD) was made at each sampling station, using a Smith-Macintyre grab sampler. Locations that yielded predominantly sand or coarser-grained material (80%  $\Phi \leq 4$ ) were sampled for individual PSD analysis only. Stations that did not meet this criterion and showed greater than one foot of shoaling were sampled for discrete chemistry analysis using the Vibra-core sampler. Composite samples of material from the fine grained sites in each sampling area were made for chemistry, bioassay, and bioaccumulation analysis. Five composite samples were generated: 1) Eureka Upper Channel (EKUP); 2) Samoa Turning Basin (SAMTB); 3) Fields Landing Lower Channel and Turning Basin (FLTBT); and 4) the disposal site reference (REF). Individual samples comprising each composite are indicated in Table 2. The composite samples were homogenized by thorough mixing using Teflon-lined containers and tools.

The vibracore cutting tip and core sample catcher were #306 grade stainless steel; the Vibracore barrel was aluminum. The Smith-Macintyre grab was constructed of galvanized steel. Prior to sampling at each station, the vibracore cutting tip, core catcher and the compositing equipment were all cleaned by the following EPA approved clean-up protocol (the Smith-Macintyre grab was cleaned with Steps 1 and 2 only):

1. Wash with 2% Micro Laboratory Soap
2. Rinse three times with clean water
3. Rinse with 2N nitric acid
4. Final rinse 3x with Milli-Q type I reagent grade DI water
5. Store in cleaned containers until use

**2.1.1 Sample Handling.** Vibracore and Smith Macintyre grab samples were taken during this project. Handling procedures for each sample type are summarized below:

**Vibracore Samples.** Each core sample was measured for total core length. If the core achieved penetration to project dredge depth the desired sample (from dredge depth to sediment surface) was extruded into the compositing container.

**Grab Samples.** Each grab sample was evaluated for grain size, composition, and penetration. Grabs which had "washed out", or which were determined to have insufficient penetration, were rejected.

The individual samples and area composites were placed in appropriate containers in precleaned coolers, on ice, to reduce the temperature to the prescribed 4°C. All samples were transported to ToxScan's chemistry and bioassay facilities in Watsonville under chain of custody at the prescribed temperature. Subsamples of the four composites were subsequently shipped at temperature under chain of custody to Alta Analytical Laboratory Inc., El Dorado Hills, CA for 2,3,7,8-TCDD and 2,3,7,8-TCDF (Dioxins) analysis.

## 2.2 Water Collection

Reference water for bioassay tests was collected at mid depth at the reference site using an EPA protocol-cleaned peristaltic pump and cleaned silicon and teflon hoses. The hose was lowered into the water at one end of the reference site, and the vessel drifted with the current through the reference site while sampling. The water was pumped into five (5) gallon cubitainers which were then stored at 4°C until delivery to the ToxScan laboratory in Watsonville.

## 2.3 Chemical and Physical Sediment Analysis

Sediment samples for chemical and physical analysis were collected in glass containers. Prior to analysis, samples were stored in the laboratory at 4°C. Analyses were conducted according to the following methods:

**Sediment Grain Size** was determined using the methods described in Plumb (1981).

**Interstitial Water Salinity and Total Ammonia** values were determined for centrifuge-extracted sediment pore waters by salinometer-calibrated refractometer (YSI Model 33 Conductivity/Salinity Meter and Atago S-10 or S-28 Hand Held Refractometer), and by pH meter / ammonia probe (Fisher Accumet Model 925 with Orion Ammonia Electrode Model 95-12). One hundred to two hundred grams of sediment were centrifuged at 7,000 to 8,000 rpm until supernatant was clear (15 - 30 minutes).

**Total and Water Soluble Sulfides.** This method was adapted from EPA Method 376.1 (EPA 1983) and Standard Method 4500-S<sup>2</sup>-E (APHA 1992). Sediment samples were mixed with O<sub>2</sub>-free DIW, and treated in a manner similar to aqueous samples. Hydrogen sulfide present in aqueous samples was purged into a zinc acetate trap using nitrogen gas. The sample pH was adjusted to about 4 if total sulfide was to be determined, or left unadjusted for free sulfide determinations. The zinc sulfide precipitate in the trap was oxidized with a known and excess amount of iodine, and the unreacted iodine was back-titrated with thiosulfate.

**Oil and Grease, Total Petroleum Hydrocarbon.** Samples were acidified to a low pH and extracted with fluorocarbon-113 in a separatory funnel. The fluorocarbon layer was separated from each sample, passed over sodium sulfate and collected for analysis of Oil and Grease using an Infrared spectrophotometer scanning the wavelengths from 3200 to 2700 cm<sup>-1</sup>. To determine Total Petroleum Hydrocarbons, this above extract was passed through silica gel which extracted the vegetable oil fractions; the remaining petroleum fraction was then analyzed by Infrared spectrophotometric techniques as described below.

**Total Organic Carbon (TOC).** Analysis for total organic carbon followed the method of Gaudette, et al. (1974). One-to-two grams of sediment were placed in a 500 ml flask to which 10 ml of potassium dichromate (K<sub>2</sub>CR<sub>2</sub>O<sub>7</sub>) had been added. Twenty ml of concentrated sulfuric acid (H<sub>2</sub>SO<sub>4</sub>) was then added while the flask was swirled. After 30 minutes, the sample was diluted to a volume of 200 ml with de-

ionized water (DIW), and 10 ml of phosphoric acid ( $H_3PO_4$ ) and 0.2 g of sodium fluoride (NaF) were added. After more swirling, 15 drops of diphenylamine indicator was added and the sample was titrated with 0.5N ferrous ammonium sulfate.

**Metals.** Analyses for metals employed combinations of the following Varian spectrophotometers: SpectrAA 400P or 400Z with GTA 96 a Graphite Furnace and autosampler; or a SpectrAA 10 with VOA 76 hydride—cold vapor generator and flame autosamplers. Sample preparation prior to analysis by atomic absorption was accomplished by guidelines specified by Chapter 3, Sections 3.2 and 3.3, 7000 series (EPA 1986).

**Organotins.** Organotin species analysis was by the method of Uhler and Durrel (1989). Speciation was done by a n-pentyl derivatization using a Gas Chromatograph with a Flame Photometric Detector. A sediment sample was mixed with 5 ml of hydrobromic acid (HBr), converting cationic butyltins to the bromide complexes, which were then extracted with a toluene-tropolone mixture. Following this extraction a n-pentylmagnesium bromide was used to convert the butyltins to the n-pentyl derivatives. This extract was cleaned by passing it through a Florisil/Silica chromatograph column and then injected into the Gas Chromatograph with a FPD detector where butyltins were quantified.

**Chlorinated Pesticides and PCB's.** Analyses for these constituents were determined by EPA Method 8080 (EPA 1986). Each solid sample was mixed with anhydrous sodium sulfate, placed in an extraction thimble and extracted using acetone and hexane in a Soxhlet extractor. The extract was then dried, concentrated, and, as necessary, underwent a Florisil clean-up. After extraction, a 2 microliter sample was injected into a gas chromatograph and the effluent detected by an electron capture detector.

**Polynuclear Aromatic Hydrocarbons and Phthalates.** Analyses for semivolatile compounds were by GC-MS techniques, following Method 8270 (EPA 1986). Each solid sample was mixed with anhydrous sodium sulfate, placed in an extraction thimble and extracted using acetone and hexane in a Soxhlet extractor. The extract was then dried, concentrated and cleaned up by gel permeation chromatography. After extraction, a 2 microliter sample was injected into a gas chromatograph and the effluent detected by mass spectroscopy.

**TCDD and TCDF (Dioxins).** Sediment samples were analyzed for 2,3,7,8-TCDD and 2,3,7,8-TCDF using EPA Method 8290. These analyses were performed by Alta Analytical Laboratory, Inc., El Dorado Hills, CA.



## **2.4 Bioassay and Bioaccumulation Test Procedures**

### **2.4.1 Suspended Particulate Phase (SPP) Bioassays**

Suspended particulate phase elutriates were prepared by procedures outlined in the "Green Book" (EPA/USACE 1991) using reference site water and test sediments. The test protocol for bivalves was as specified by ASTM (1989). Three concentrations (100%, 50%, 10%) of suspended particulate phase were tested. The lower concentrations were evaluated only if the 100% concentrations produced >50% inhibition of development. Three species were tested in suspended particulate phase bioassays: The larvae of a marine bivalve (the bay mussel, *Mytilus edulis*), a mysid (*Holmesimysis costata*), and a marine teleost fish (the speckled sanddab, *Citharichthys stigmaeus*).

Elutriate sanddab bioassays were performed at the Davenport laboratory, and elutriate bioassays with mysids and bivalve larvae were performed at the Watsonville laboratory. The positioning of test containers and other conditions in the laboratories were designed for uniform exposure to the controlled laboratory environment. Five replicates of test treatments were randomly assigned (complete random design) to the test containers by use of a random numbers generating program.

The sediment samples were placed in cleaned 5-gallon polyethylene buckets with laboratory seawater for elutriate preparation. The sediment to water ratio was 1:4 as specified in the Green Book. The mixtures were agitated by vigorous aeration for 30 minutes. After a one-hour settling period, the elutriates were siphoned off and used as suspended particulate phase media.

#### **2.4.1.1 Bivalve Larvae (*Mytilus edulis*)**

Mussels were induced to spawn by high-temperature stimulation. Eggs and sperm were collected in separate basins filled with aerated seawater at 20°C. Egg density was determined by microscopically counting several 1-ml aliquots taken from the well-mixed egg basin. Fertilization was accomplished by addition of an appropriate amount of sperm suspension, and confirmed by microscopic examination.

The control exposure, performed for quality assurance purposes, used seawater from our laboratory system. Five replicate dishes were used for each test exposure. Temperature, dissolved oxygen, pH and salinity were monitored in each test concentration and in controls at the beginning and end of the test.

Larvae were tested in 250 ml polyethylene beakers containing approximately 200 ml of test solution. After fertilization was confirmed an aliquot containing approximately 6000 fertilized eggs was pipetted into each test beaker. Gentle aeration was provided throughout the 48-hour duration of the test. Five extra beakers were prepared in addition to those required for test and control replicates. These "extra" test containers were not incubated for 48 hours, but rather they were evaluated immediately after

inoculation to provide the "initial recovery" data used to establish the mean number of embryos added to each experimental beaker.

At the end of the 48-hour exposure period the contents of each dish were poured through a 45 $\mu$  nytex screen. Surviving larvae were retained on the screen. The test beaker was rinsed three times with seawater and each successive rinse was poured through the screen to ensure complete transfer of larvae. Larvae were quantitatively transferred from the screen into a graduated cylinder and the volume was adjusted with a seawater-formalin mixture. Contents of the cylinder were mixed by inversion to ensure uniform distribution of larvae, and a 1 ml aliquot was transferred to a Sedgwick-Rafter counting slide for microscopic evaluation. Larvae were scored for evidence of internal tissue inside a complete larval shell. Larvae which had a complete larval shell containing tissue were counted as normal, whereas empty shells and larvae with incomplete shells were scored as abnormal. Data were reported as percent of initial embryos which survived, and percent of survivors which showed normal development, as calculated below.

The raw data resulting from these bioassays included the following:

- Counts of embryos added to five replicate test containers which were not incubated for 48 hours (= initial recovery).
- Counts of normal and abnormal embryos from test containers (five replicates per sample, reference and control) which were incubated for 48 hours.

The results were calculated from these data as follows:

$$\% \text{ Survival} = \frac{\text{No. normal larvae recovered}}{N} \times 100$$

$$\% \text{ Normal} = \frac{\text{No. normal larvae}}{\text{No. normal larvae} + \text{No. abnormal larvae}} \times 100$$

where N = the mean initial number of embryos added (from initial recovery data).

For each test chamber other than controls, % survival data were adjusted to correct for mortality observed in the control exposures by use of **Abbott's correction**:

$$\text{Corrected Sample \% Survival} = 100 - \left( \frac{\text{mean \% control survival} - \% \text{ sample survival}}{\text{mean \% control survival}} \times 100 \right)$$

Percent normal development data were similarly adjusted.

For the bioassay to be considered a valid test, an average of at least 70% of the exposed embryos must survive in the controls; abnormals were counted as mortalities as per the Testing Guidelines contained in SFACOE Public Notice No. 93-2: Response to Comments on Public Notice 92-5.

Following the Scope of Services, the 100% elutriate concentrations were evaluated initially. If Abbott's-corrected survival or normal development values were  $\geq 50\%$ , no further evaluations were performed. If these values were  $\leq 50\%$ , the 10% and 50% elutriate exposures were evaluated and  $EC_{50}$  and/or  $LC_{50}$  calculations were made using the Trimmed Spearman-Kärber method. For  $LC_{50}$  calculations, abnormal larvae and calculated mortalities were added; whereas for  $EC_{50}$  calculations, separate abnormality counts were used, as per Public Notice 93-2 (see above).

A reference toxicant bioassay was also performed for quality assurance purposes, to verify the health and sensitivity of the test organism population. The reference toxicant used was cupric sulfate ( $CuSO_4 \cdot 5H_2O$ ) dissolved in laboratory seawater.

#### 2.4.1.2 Mysid (*Holmesmysis costata*)

Adult mysids (*Holmesmysis costata*) were collected from kelp beds near Monterey, California. The animals were gently aggregated with a dip net, corralled into a submerged bucket without removing them from the water and transported directly to the bioassay lab. In transit, holding tank temperatures were maintained within 2°C of the ambient temperature at sampling. Gentle aeration was supplied from a bottle of compressed oxygen. Throughout testing, the mysids were fed about 50 brine shrimp (*Artemia salina*) nauplii per mysid per day to prevent mortality from starvation and cannibalism.

Mysids were tested in one-liter polycarbonate tanks containing one liter of test solution. To initiate testing, mysids were sorted into groups of 10 in small containers with very small volumes of seawater. Mysids were transferred to the test containers by submerging the containers and slowly tipping the animals into the test medium. During the bioassays, the number of survivors of the original 10 animals per tank were recorded as experimental data at 4, 8, 24, 48, 72, and 96 hours after test initiation. At each of these checkpoints, dead animals (i.e., those nonresponsive to mechanical stimulus) were removed from the test containers.

A reference toxicant bioassay was also performed on the mysids for quality assurance purposes, to verify the health and sensitivity of the test organism population. The reference toxicant used was Sodium Dodecyl Sulfate (SDS) dissolved in laboratory seawater.

#### **2.4.1.3 Teleost Fish (*Citharichthys stigmaeus*)**

Speckled sanddabs were collected by otter trawl from Tomales Bay and kept in holding tanks until transported to the laboratory via overnight delivery. They were allowed to acclimate to laboratory conditions prior to testing. Fish were fed a high protein pellet food during the holding period until 48 hours before test initiation; they were not fed thereafter.

Sanddabs were tested in 10-liter aquaria and were individually transferred from holding tanks to aquaria to start the test. During the bioassays, the number of survivors of the original 10 animals per tank was recorded as experimental data at 4, 8, 24, 48, 72, and 96 hours after test initiation. At each of these checkpoints, dead animals (i.e., those nonresponsive to mechanical stimulus) were removed from the test containers.

A reference toxicant bioassay was also performed on the sanddabs for quality assurance purposes, to verify the health and sensitivity of the test organism population. The reference toxicant used was Sodium Dodecyl Sulfate (SDS) dissolved in laboratory seawater.

#### **2.4.1.4 Initial Mixing Calculations**

In cases where an  $EC_{50}$  or  $LC_{50}$  was obtained, calculations of initial mixing were made using standardized formulae developed by the USACOE and EPA (EPA/ACOE 1977).

#### **2.4.2 Solid Phase (SP) Static Bioassays (Amphipod)**

Solid phase static bioassays were conducted on the harbor sediments simultaneously with control and reference sediments. The amphipod *Rhepoxynius abronius* was tested following procedures outlined in ASTM (1990).

Salinity and total ammonia measurements were made on sediment interstitial water as received; in addition, a final pore-water ammonia measurement was taken from one replicate of each test sediment at test termination. Pore waters were extracted by centrifugation. Interstitial water salinity was measured using a salinometer-calibrated refractometer. Interstitial water ammonia concentrations were measured with an ammonia probe calibrated to three concentration standards (see Sediment Physical and Chemical Analysis- Section 2.3).

In each test, five replicates of each station and reference treatment were randomly assigned to test jars. A 2-cm deep layer of appropriate sediment was added to each jar on the day prior to test initiation, and each test jar was provided with aeration via pasteur pipet. Each test was started on the following day by randomly assigning 20 amphipods to each jar, and continued for 10 days under static conditions with constant illumination and aeration. Daily measurements of environmental test conditions (temperature, salinity, pH, dissolved oxygen) were made in each test container, and the number of animals which had appeared on the sediment surface was noted.

At the end of the ten day exposure period, the contents of each jar were poured through a 0.5 mm sieve and the number of surviving amphipods counted. Survivors from each replicate were transferred into bowls containing control sediment and monitored for their ability to rebury within one hour. Test data for each replicate therefore include number of survivors and number of survivors able to rebury.

Reference toxicant bioassays were performed with each batch of test animals to verify the health and sensitivity of the test organism population. The reference toxicant used was cadmium chloride ( $\text{CdCl}_2$ ) dissolved in laboratory seawater.

#### 2.4.3 Solid Phase (SP) Flow-through Bioassays (Mysid Shrimp and Polychaete Worm)

Solid phase flow-through bioassays with mysids and worms were conducted on the harbor sediments simultaneously with control and reference sediments. Control sediments were collected from Tomales Bay. Testing for both species was performed at the Davenport facility where continuously flowing seawater is available, using testing procedures in EPA/COE (1991).

Mysids (*Holmesimysis costata*) were collected from kelp beds near Monterey, California. The animals were gently aggregated with a dip net, corralled into a submerged bucket without removing them from the water and transported directly to the bioassay lab. In transit, holding tank temperatures were maintained within 2°C of the ambient temperature at sampling. Gentle aeration was supplied from a bottle of compressed oxygen. Throughout testing, the mysids were fed about 50 brine shrimp (*Artemia salina*) nauplii per mysid per day to prevent mortality from starvation and cannibalism.

Polychaete worms (*Nephtys caecoides*) were collected from Tomales Bay and shipped overnight to the bioassay laboratory. They were kept in holding tanks with home sediment and overlying seawater until test initiation.

All sediments were sieved through a 1.0 mm screen to remove indigenous fauna, and a 3.0 cm layer of appropriate sediment was added to each test container. Tanks were then filled with lab seawater, and either twenty polychaete worms (*Nephtys caecoides*) or twenty mysids (*Holmesimysis costata*) were added to each container. Worms were tested in 31 L glass aquaria; mysids were tested in 1.5 L

polycarbonate tanks fitted with small, screened drain holes. The small mysid containers were suspended above the larger worm containers such that when the flow-through seawater system was activated, seawater passed through the mysid tanks, overflowed through the screened drain holes into the worm tanks, then drained to sea.

Solid Phase flow-through bioassays continued for 10 days. At least twice each day, environmental systems were checked for proper functioning. Once each day, the salinity and temperature of the system were measured. Dissolved oxygen and pH values of each tank were measured twice daily.

After the 10-day bioassay period, the contents of each tank were gently washed with seawater through a 0.5-mm nylon screen. The animals were retrieved from the screen and counted. Test data were the number of survivors of each species.

A reference toxicant bioassay was also performed on the mysids for quality assurance purposes, to verify the health and sensitivity of the test organism population. The reference toxicant used was Sodium Dodecyl Sulfate (SDS) dissolved in laboratory seawater.

#### 2.4.4 Bioaccumulation Exposure

Based on evaluations of sediment chemistry, bioaccumulation tissue assessments were not performed on these sediments (see below). However, clams (*Macoma nasuta*) and polychaete worms (*Nephtys caecoides*) were exposed to test and control sediments in an array of 31-liter flow-through glass aquaria, as follows: Five replicates of each harbor composite, reference composite and control sediments were randomly assigned to the test tanks. The control sediment was collected from Tomales Bay, CA. Sediments were screened through a 1.0 mm screen to remove indigenous fauna, and a 3.0 cm layer was added to each tank. Tanks were filled with water and 30 clams and 40 worms were added to each. After a one-hour settling time, the flow-through seawater system was activated and adjusted to a flow rate equivalent to 5 tank/volume changes per 24 hours (6.5 liters/hour).

Bioaccumulation exposures continued for 28 days. At least twice each day, environmental systems were checked for proper function. Each tank was monitored daily for temperature and D.O., and the seawater system was monitored daily for salinity and pH.

After exposure, the contents of each tank were gently washed with seawater through a 0.5-mm nylon screen from which the animals were retrieved. Surviving clams were transferred for two days to filtered flowing seawater for gut evacuation. Surviving worms were transferred to 30-liter flow-through aquaria containing a 3-cm layer of fine, clean sand. Visual inspection of individuals confirmed how much time (typically 24 hours) was necessary for complete gut evacuation in worms. Directly following these treatments, the soft tissues of clams and worms were homogenized, then frozen.

### 3.0 Results

Sediment physical, chemical, and bioassay analyses are summarized in Table 1. Fourteen samples (including one replicate) from North Bay, Entrance and Bar were screened and analyzed only for particle size distribution (PSD). Twenty-seven samples were analyzed for PSD and sediment chemistry: 22 discrete samples plus four composites (EKUP, SAMTB, FLTB, REF) and the Tomales Bay control sediment. Bioassay testing and bioaccumulation exposures were performed on the four composites and on the control sediment; subsamples of these sediments were subcontracted for dioxin (2,3,7,8-TCDD and 2,3,7,8-TCDF) analysis.

#### 3.1 Sediment Physical Analysis

The particle size distributions of the sediment samples and composites are summarized in Table 4 and detailed in Appendix C. Except for NB4, Rep 1, the North Bay, Entrance and Bar samples each contained at least 90% coarse sediments by weight ( $\Phi \leq 4$ ). Coarse sediment composition of the three harbor composites were as follows: Eureka Upper Channel (EKUP) = 61.1%; Samoa Turning Basin (SAMTB) = 81.0%; and Field's Landing Lower Channel and Turning Basin (FLTB) = 42.6%. The disposal site reference (REF) composite contained 59.3% coarse sediments, and the Tomales Bay control sediment contained 97.9% coarse particles.

#### 3.2 Bulk Sediment Chemistry

Results of bulk sediment chemical analyses of the Humboldt Harbor sediment samples and composites are summarized in Table 4. The laboratory reports are presented in Appendix C, and QA/QC reports are presented in Appendix D. Chains of Custody are Presented in Appendix F. The discussion below is generally limited to analyses of the harbor and reference composites; please refer to Appendix C for results of analyses of the individual samples.

**Metals.** The Humboldt Harbor sediment composites were analyzed for ten metals. Except for cadmium, metals concentrations in the Harbor composites were similar to or less than those found in the Reference composite. Within the Harbor composites, Comp FLTB tended to have the highest metals concentrations, and contained cadmium levels twice that of the reference composite. Individual accounts of the ten metals analyzed in these sediments are as follows:

- Arsenic concentrations ranged from 5.7 ppm to 7.0 ppm in the harbor composites. None of the harbor composites exceeded the 7.3 ppm found in the reference composite.
- Cadmium concentrations ranged from 0.1 ppm to 0.2 ppm in the harbor composites. FLTB and EKUP exceeded (by 2.0x) the 0.1 ppm found in the reference composite. It should be noted, however, that the cadmium concentrations found in these sediments were near the detection limit.

- Chromium concentrations ranged from 97 ppm to 100 ppm in the harbor composites. None of the harbor composites exceeded the 110 ppm found in the reference composite.
- Copper concentrations ranged from 12 ppm to 28 ppm in the harbor composites. Only FLTB (1.3x) exceeded the 21 ppm found in the reference composite.
- Lead concentrations ranged from 5.8 ppm to 8.1 ppm in the harbor composites. FLTB (1.09x) and EKUP (1.04x) exceeded the 7.4 ppm found in the reference composite.
- Mercury concentrations ranged from 0.08 ppm to 0.05 ppm in the harbor composites. None of the harbor samples exceeded the 0.20 ppm found in the reference composite.
- Nickel concentrations ranged from 62 ppm to 98 ppm in the harbor composites. Only FLTB (1.01x) exceeded the 97 ppm found in the reference composite.
- Selenium was not detected in any of the harbor composites, nor in the reference composite.
- Silver was not detected in any of the harbor composites, nor in the reference composite.
- Zinc concentrations ranged from 41 ppm to 66 ppm in the harbor composites. Only FLTB (1.06x) exceeded the 62 ppm found in the reference composite.

**Butyltins.** Three organotins (tri-, di-, and mono-butyltin) were measured in the Humboldt Harbor sediment composites. A small amount (1 ppb) of tributyltin was detected in the SAMTB composite. No mono- or tetrabutyltins were detected from the harbor composites, and the reference and control sediments contained no detectable butyltins.

**Semivolatiles.** Phthalate esters and seventeen polynuclear aromatic hydrocarbons (PAHs) were measured in the Humboldt Harbor sediment composites. Phthalate concentrations ranged from 160 to 170 ppb in the harbor composites; none exceeded the 1200 ppb measured in the reference composite. PAH detections were as follows:

LPAHs. Three low molecular weight PAHs were detected in the harbor or reference composites, as follows:

- 2-methylnaphthalene concentrations ranged from <20 ppb to 78 ppb in the harbor composites; FLTB (1.95x) and EKUP (1.03x) exceeded the 40 ppb detected in the reference composite.
- Naphthalene concentrations ranged from 37 ppb to 43 ppb in the harbor composites; all three harbor composites exceeded the <20 ppb detected in the reference composite.
- Phenanthrene concentrations ranged from <20 ppb to 49 ppb in the harbor composites; none of the harbor composites exceeded the 58 ppb detected in the reference composite.



HPAHs. Three high molecular weight PAHs (HPAHs) were detected in the harbor or reference composites, as follows:

- Fluoranthene concentrations ranged from 30 ppb to 42 ppb in the harbor composites; all three harbor composites exceeded the <20 ppb detected in the reference composite.
- Pyrene concentrations ranged from <20 ppb to 47 ppb in the harbor composites; FLTB and EKUP exceeded the <20 ppb detected in the reference composite.
- Benz(a)pyrene concentrations ranged from <20 ppb to 52 ppb in the harbor composites; only SAMTB exceeded the <20 ppb detected in the reference composite.

**Chlorinated Pesticides and PCBs.** The Humboldt Harbor sediment composites were analyzed for the eighteen chlorinated pesticides and four polychlorinated biphenyls (PCBs as Aroclors). None of the harbor composites, reference or Tomales Bay control sediments contained detectable amounts of these substances.

**Dioxins.** The Humboldt Harbor composites were analyzed for 3,7,8-TCDD and 3,7,8-TCDF by Alta Analytical Laboratories, (El Dorado Hills, CA). Only SAMTB (0.51 pg/g 2,3,7,8-TCDF) contained detectable quantities of dioxins.

**Sediment Conventionals.** Total sulfides ranged from 29 ppm to 110 ppm in the harbor sediment composites. The reference composite contained 0.3 ppm total sulfides. Except for a trace amount (0.2 ppm) in the FLTB composite, no water soluble sulfides were found in the harbor composites, nor in the reference composite.

Oil and Grease (31 ppm) was detected only in the FLTB composite; total petroleum hydrocarbons were not detected in the harbor and reference composites.

Percent solids in the harbor composites ranged from 65% to 77% compared to 73% in both the reference composite and the Tomales Bay control sediment; total organic carbon ranged from 0.3% to 0.8% in the harbor composites, compared to 0.4% in the reference composite.

**Sediment Chemistry Summary:** Except for total sulfides, Humboldt Harbor sediments appear to contain no particularly high concentrations of any of the tested substances or compounds when compared to the reference site sediments. Although sulfide concentrations in the harbor composites (29 ppm to 110 ppm) were not particularly high for harbor sediments in general, they exceeded the very low reference site concentration of 0.3 ppm. Concentrations of some PAHs (Naphthalene, Fluoranthene, Pyrene and Benzo(a)pyrene) in some harbor composites exceeded reference site values by 4.2x to 5.2x (calculating non-detects at 0.5 x D.L.), but again, the absolute concentrations (30 ppb to 52 ppb) of these compounds were not high.

### 3.3 Bioassay Test Results

Six sediment toxicity evaluations were conducted on the Humboldt Harbor sediments. Suspended particulate phase (SPP) bioassays and solid-phase (SP) bioassays employed a total of five species, and bioaccumulation exposures were conducted with two species (Table 3). Bioassay test results and statistical evaluations are summarized in Tables 6 through 11. Water quality monitoring data summaries and reference toxicant test results are tabularized in Appendix D. Logs of test animal shipping, receiving, acclimation and holding are contained in Appendix E.

#### 3.3.1 Suspended Particulate Phase (SPP) Bioassays

Suspended Particulate Phase bioassay testing of the Humboldt Harbor Baseline Survey II sediments comprised three species: a bivalve larva (*Mytilus edulis*), a teleost fish (*Citharichthys stigmatosus*) and a mysid shrimp (*Holmesimysis costata*). Results of these bioassays are summarized below, and in Tables 6 through 8.

##### 3.3.1.1 Bivalve Larvae

Adult *Mytilus edulis* were purchased from Carlsbad Aquafarm Inc., Carlsbad, CA. Collection data were not available. The animals were shipped on 13 April 1994 and arrived at ToxScan's Watsonville laboratory on 14 April via Federal Express overnight. The mussels were held in 33.2 - 33.9‰ seawater at 14.3 - 15.2 °C until test initiation on 20 April. Results of the bivalve larvae tests are presented in Table 6.

**Survival.** Mean survival of bivalves in the laboratory seawater control was 101.8%, well above the ASTM (1989) protocol requirements of 70 percent. The reference site 100% elutriate produced 86.4% survival, Abbott's-corrected to 84.9%. Abbott's corrected mean survival in the 100% elutriates of the Humboldt Harbor composites ranged from 76.6% in the FLTB composite to 93.2% in the SAMTB composite. None of the harbor sediment bivalve tests demonstrated enough toxicity to generate an LC<sub>50</sub>.

**Development.** Mean normal development values (adjusted with Abbott's correction) for bivalve larvae exposed to 100% elutriates of the test sediment ranged from 88.3% in the FLTB composite to 99.7% in the EKUP composite. Normal development in the disposal site reference elutriate was 98.2%, Abbott's-corrected to 99.1%. Normal development the laboratory seawater control was 99.0%. None of the Humboldt Harbor sediment bivalve tests demonstrated enough toxicity to generate an EC<sub>50</sub>.

**Reference Toxicant.** The bivalve reference toxicant LC<sub>50</sub> was 5.72 ppb Cu (95% CL: 5.21 - 7.15), and the EC<sub>50</sub> for development was 6.66 ppb (95% CL: 6.21 - 7.15). These values are within ±2 SD of the mean of EC<sub>50</sub>s calculated from previous *Mytilus*: copper reference toxicant tests.

### 3.3.1.2 Mysid Shrimp

Adult *Holmesimysis costata* were collected from kelp beds near Monterey, California by SP Aquatics on 11 May 1994 and transported directly to the bioassay lab. The mysid suspended particulate phase bioassay was initiated that afternoon.

Mean survival of the mysids was 100% in each of the Humboldt Harbor Baseline Survey II sediment elutriates (Table 7). Mean control and reference site composite survival were each 98%. Mysid survival in the harbor composites was not significantly different than reference site survival (Steel's Many-One Rank Test:  $p=0.05$ ,  $k=3$ ).

**Reference Toxicant.** The mysid reference toxicant 96 hour  $LC_{50}$  was 5.46 mg/L SDS (95% CL: 4.45 - 6.71). This value is within  $\pm 2$  SD of the mean of  $LC_{50}$ s calculated from previous *Holmesimysis*: SDS reference toxicant tests.

### 3.3.1.3 Teleost Fish

Speckled sanddabs (*Citharichthys stigmaeus*) were collected from Tomales Bay by John Brezina & Associates. Collection data was not available. The animals arrived at ToxScan's Watsonville laboratory on 6 May 1994 via Federal Express overnight. The sanddabs were held in 33.0 - 33.3‰ seawater at 11.7 - 14.1 °C until test initiation on 10 May.

Mean survival of the sanddabs in the Humboldt Harbor sediment elutriates ranged from 4.0% in the FLTB composite to 96% in the EKUP composite (Table 8). Mean control survival was 100%, and the reference site survival was 98%. Survival in the FLTB composite was significantly diminished from survival in the reference sediment (Steel's Many-One Rank Test:  $p=0.05$ ,  $k=3$ ).

**Reference Toxicant.** The sanddab reference toxicant 96 hour  $LC_{50}$  was 2.87 mg/L SDS (95% CL: 2.66 - 3.10). This value is above  $\pm 2$  SD of the mean of  $LC_{50}$ s calculated from previous *Citharichthys*: SDS reference toxicant tests. This suggests that these fish may be less sensitive indicators of sediment toxicity than those used in prior sediment bioassays in our laboratory.

### 3.3.1.4 Initial Mixing Calculations

Calculations of initial mixing concentrations for disposal depths of 50m, 20m and 10m are detailed in Appendix E (Table E1). Only one composite (FLTB) generated an  $EC_{50}$  value (71.74% SPP) for one test species (*Citharichthys*) and therefore qualified for initial mixing calculations. In this case the factored  $EC_{50}$  (0.71) was higher than the projected concentrations ( $C_{sp}$ ) for each of the three depths. Therefore the Limiting Permissible Concentrations (LPCs) were not exceeded for this sediment.

### 3.3.2 Solid Phase (SP) Static Bioassay (Amphipod)

Test amphipods (*R. abronius*) were collected on 11 April 1994 at West Beach, Whidbey Island, WA by Northwestern Aquatics, Inc.. They arrived via overnight delivery at ToxScan's bioassay facility in Watsonville, CA on 19 April. These amphipods did not require salinity acclimation; they were held at 33‰ to 34.1‰ until testing was initiated on 25 and 26 April with 32.5‰ - 33.0‰ overlying seawater.

Solid phase static bioassay results for the amphipod *Rhepoxynius abronius* are summarized below and in Table 9. Prior to initiation of the bioassay, analysis of interstitial waters found ammonia concentrations in the FLTB composite (57.7 mg/L) exceeded the threshold limit of 30 mg/L for *Rhepoxynius*. Therefore, ammonia-purging procedures were initiated for the FLTB sediments, and a parallel (purged) control was set up using home sediments from Yaquina Bay. After two days of aeration and twice-daily renewals of overlying water, total ammonia levels in the FLTB pore water was sufficiently reduced ( $\leq 30$  mg/L) to begin the test. Twice daily renewals continued for the test duration. The final concentration of total ammonia in the FLTB pore water was 8.6 mg/L (Table 5).

**Survival.** Mean survival of *R. abronius* in the Humboldt Harbor Baseline Survey II sediment composites ranged from 96.9% to 99.0% (versus 93.0% in the home sediment control and 94.0% in the reference site composite. Survival in the harbor composites did not differ significantly from reference site survival (Dunnett's Test:  $p=0.05$ ,  $df=16,3$ ).

**Reference Toxicant.** The amphipod reference toxicant 96 hour  $LC_{50}$  was 0.76  $\mu\text{g/L}$  Cd (95% CL: 0.56 - 1.02). This value is within  $\pm 2$  SD of the mean of  $LC_{50}$ s calculated from previous *Rhepoxynius* : Cd reference toxicant tests.

### 3.3.3 Solid Phase (SP) Flow-Through Bioassays: Mysid Shrimp and Polychaete Worm

Solid phase flow-through bioassay results are summarized below and in Tables 10 (mysid) and 11 (worm).

**Mysid Shrimp Survival.** Adult *Holmesimysis costata* were collected from kelp beds near - Monterey, California by SP Aquatics on 4 May 1994. They were transported directly to the bioassay lab where the mysid SP bioassay was initiated that afternoon.

Mean survival of *H. costata* was 95% to 96% in the Humboldt Harbor composites, in the home sediment control and in the reference composite. Mysid survival in the harbor composites was equal to or greater than survival in the reference site composite.

**Reference Toxicant.** The mysid reference toxicant 96 hour  $LC_{50}$  was 9.28 mg/L SDS (95% CL: 7.61 - 11.3). This value falls above  $\pm 2$  SD of the mean of  $LC_{50}$ s calculated from previous *Holmesimysis*:

SDS reference toxicant tests. This suggests that these mysids may be less sensitive indicators of sediment toxicity than mysids used in other sediment bioassays in our laboratory.

**Polychaete Worm Survival.** Adult *N. caecoides* were collected from Tomales Bay by John Brezina and Associates. Collection data were not available. They were received at the bioassay laboratory on 26 April 1994 via overnight delivery. These animals did not require salinity acclimation; they were held in home sediment with 32.8‰ - 32.9‰ overlying seawater at 10.9 °C - 12.7 °C until testing was initiated on 28 April.

Mean survival of *Nephtys caecoides* in the Humboldt Harbor sediment composites ranged from 93% to 94% (versus 100% in the home sediment control and 89% in the reference composite. Worm survival in each of the harbor composites exceeded survival in the reference site composite.

**Reference Toxicant.** The polychaete reference toxicant 96 hour LC<sub>50</sub> was 107.5 µg/L copper (95%CL: 71.5 - 161.6). Sufficient data were not available to generate a control chart for this species.

### 3.3.4 Bioaccumulation Exposure (Clam and Worm)

Based on EPA and SFACOE review of the sediment chemistry results for the three harbor composites and the reference composite, no assessment of the exposed tissues was deemed warranted for either *Macoma nasuta* or *Nephtys caecoides*. The bulk sediment chemistry data revealed no detectable chlorinated pesticides or PCBs, while total detected organotins (<1 ppb) were close to detection limits. Similarly, analyses for dioxins found trace amounts (0.51 pg/g 2,3,7,8-TCDF; 0.94 pg/g total TCDF) in only one composite (SAMTB). PAHs were detected at low concentrations (98 - 160 ppb) in the composites.

**Bioassay Results Summary.** Except for the sanddab elutriate (SPP) test, the bioassay testing of the Humboldt Bay Baseline Survey II sediments demonstrated little or no toxicity to the test species. In the sanddab test, the high mortality found in the 100% elutriate was not carried through to the 50% and 10% dilutions, and did not produce an initial mixing scenario which exceeded the LPC. High concentrations of initial pore water ammonia (57.7 mg/L) in the FLTB composite may have been the major source of the observed toxicity. The reference toxicant tests for the sanddabs and for the solid phase mysid test produced LC<sub>50</sub>s which were above our control chart limits.

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TABLES





Table 1. Analyses Performed, Humboldt Bay Baseline Survey II (FY 1994). Shaded samples composited; SP = Suspended Phase; SPP = Suspended Particulate Phase.

SAMPLE	Initial Grain Size	Sediment Chemistry	SP + SPP Bioassay	Bioaccumulation <sup>1</sup>
<b>North Bay Channel:</b>				
NB1	YES	NO	NO	NO
NB2	YES	NO	NO	NO
NB3	YES	NO	NO	NO
NB4 Rep 1	YES	YES	NO	NO
NB4 Rep 2	YES	NO	NO	NO
NB5	YES	NO	NO	NO
NB6	YES	NO	NO	NO
NB7	YES	NO	NO	NO
NB8	YES	NO	NO	NO
NB9	YES	NO	NO	NO
NB10	YES	NO	NO	NO
<b>Samoa Turning Basin:</b>				
SAM1	YES	YES	NO	NO
SAM2	YES	YES	NO	NO
SAM3	YES	YES	NO	NO
SAM4	YES	YES	NO	NO
SAM5	YES	YES	NO	NO
<u>Comp. SAMTB:</u>	YES	YES	YES	YES
SAM6-A	YES	YES	NO	NO
SAM6-B	YES	YES	NO	NO
SAM6-C	YES	YES	NO	NO
SAM7	YES	YES	NO	NO
<b>Eureka Upper Channel:</b>				
EK1	YES	YES	NO	NO
<u>Comp. EKUP:</u>	YES	YES	YES	YES
EK2	YES	YES	NO	NO
EK3	YES	YES	NO	NO
EK4	YES	YES	NO	NO
<b>Fields Landing Lower Channel and Turning Basin:</b>				
<u>Comp. FLTB:</u>	YES	YES	YES	YES
FL1	YES	YES	NO	NO
FL2	YES	YES	NO	NO
FL3	YES	YES	NO	NO
FL4	YES	YES	NO	NO
FL5	YES	YES	NO	NO
FL6	YES	YES	NO	NO
FL7	YES	YES	NO	NO
FL8	YES	YES	NO	NO
<b>Entrance Channel, Bar, Reference Site and Control:</b>				
ENT1	YES	NO	NO	NO
ENT2	YES	NO	NO	NO
BAR1	YES	NO	NO	NO
REF	YES	YES	YES	YES
CONTROL	YES	YES	YES	YES

<sup>1</sup> Exposures only; no tissue analyses performed.

Table 2. Sediments Collected, Humboldt Bay Baseline Survey II (FY 1994). Samples collected by vibracore or Smith-Macintyre grab; shaded samples composited.

SAMPLE	DATE	TIME	MUDLINE	Core Penetration (Feet)		California State Plane Coordinates <sup>1</sup>	
				ACHIEVED	SAMPLED	NORTH	EAST
<b>North Bay Channel:</b>							
NB 1	4/1/94	16:47	36.0	GRAB <sup>2</sup>	0.5	525040	1384353
NB 2	4/4/94	14:10	37.5	GRAB	0.5	526977	1380256
NB 3	4/1/94	16:20	34.5	GRAB	0.5	528632	1386625
NB 4 Rep1	4/1/94	15:55	36.5	GRAB	0.5	530690	1387641
NB 4 Rep2	4/1/94	17:23	36.5	GRAB	0.5	530690	1387641
NB 5	4/1/94	15:23	37.0	GRAB	0.8	531747	1389429
NB 6	4/1/94	15:02	35.7	GRAB	0.4	533775	1391711
NB 7	4/1/94	14:31	34.4	GRAB	0.4	535684	1392296
NB 8	4/1/94	14:10	36.5	GRAB	0.5	536655	1392972
NB 9	4/1/94	13:52	35.4	GRAB	0.7	540438	1394733
NB 10	4/1/94	13:33	35.5	GRAB	0.3	552578	1395037
<b>Samoa Turning Basin (SAMTB):</b>							
SAM 1	3/31/94	10:56	34.6	GRAB	0.4	541657	1394834
SAM 2	3/31/94	10:27	36.1	GRAB	0.5	542636	1395296
SAM 3	3/31/94	10:03	33.0	GRAB	0.5	544222	1395985
SAM 4	3/31/94	09:30	35.5	GRAB	0.5	545254	1396359
SAM 5	3/31/94	09:02	34.5	GRAB	0.7	548030	1397957
SAM 6 A	4/1/94	09:54	35.7	GRAB	0.5	547562	1397303
SAM 6 B	3/31/94	16:34	33.9	4.1	3.1	548139	1397482
SAM 6 C	3/31/94	12:10	34.9	3.2	2.1	548418	1397752
SAM 7D	3/31/94	08:36	35.9	GRAB	0.7	548030	1397957
<b>Eureka Upper Channel (EKUP):</b>							
EK 1	4/2/94	13:48	35.8	GRAB	0.5	541580	1395533
EK 2	4/2/94	09:25	24.7	3.5	3.3	543195	1396863
EK 3	4/2/94	14:48	21.9	6.5	6.1	543562	1397576
EK 4	4/2/94	16:46	27.2	GRAB	0.5	543841	1399100
<b>Fields Landing Lower Channel and Turning Basin (FLTBT):</b>							
FL 1	4/4/94	11:15	27.8	GRAB	0.3	513355	1384535
FL 2	4/4/94	12:44	26.5	GRABS	0.5	514065	1384547
FL 3	4/3/94	12:09	27.2	GRABS	0.5	514488	1384092
FL 4	4/3/94	09:23	25.9	3.1	2.1	515355	1384535
FL 5	4/3/94	11:27	26.3	GRAB	0.5	517271	1385416
FL 6	4/3/94	11:05	30.2	GRAB	0.5	519202	1385029
FL 7	4/3/94	10:51	28.4	GRAB	0.5	521204	1383853
FL 8	4/3/94	10:45	25.2	GRAB	0.5	523214	1384976
<b>Entrance Channel, Bar and Reference Site:</b>							
ENT 1	4/5/94	12:00	36	GRAB	0.5	526000	1382500
ENT 2	4/5/94	12:10	48	GRAB	0.5	529100	1380400
BAR 1	4/5/94	11:45	42	GRAB	0.5	530800	1377700
REF 1	4/5/94	08:46	165	GRAB	0.5	524700	1351300

<sup>1</sup> Field measurements of station locations were made in latitude x longitude (see Field Logs, Appendix A), and converted here to California State Plane Coordinates.

<sup>2</sup> Grab samples (except Entrance and Bar) were taken only where depth from bottom to project depth was less than 1.5 ft; Entrance and Bar stations were grab sampled due to wind and sea conditions.

Table 3. Biological Assessments, Humboldt Bay Baseline Survey II (FY 1994).

Test Species	SP	SPP	BA
<i>R. abronius</i>	X	-	-
<i>M. edulis</i>	-	X	-
<i>H. costata</i>	X	X	-
<i>C. stigmaeus</i>	-	X	-
<i>N. caecoides</i>	X	-	(x)
<i>M. nasuta</i>	-	-	(x)

X = test performed; (x) = exposure only, no tissue evaluation

SP = Solid Phase; SPP = Suspended Particulate Phase; BA = Bioaccumulation.

Table 4. Sediment Chemistry Summary, Humboldt Bay Baseline Survey II (FY 1994): Composites only; for individual sample results, see Appendix C.

Analyte	Sampling Sections				Detection Limit
	SAMTB	EKUP	FLT B	REF	
<b>METALS</b> (ppm, dry wt)					
Arsenic	5.7	6.3	7.0	7.3	0.1
Cadmium	0.1	0.2	0.2	0.1	0.1
Chromium	100	97	100	110	0.1
Copper	12	20	28	21	0.1
Lead	5.8	7.7	8.1	7.4	0.1
Mercury	0.05	0.06	0.08	0.20	0.02
Nickel	62	81	98	97	0.1
Selenium	ND	ND	ND	ND	0.1
Silver	ND	ND	ND	ND	0.1
Zinc	41	55	66	62	1.0
<b>ORGANOTINS</b> (ppb, dry weight)					
Monobutyltin	ND	ND	ND	ND	1.0
Dibutyltin	ND	ND	ND	ND	1.0
Tributyltin	1	ND	ND	ND	1.0
<b>PAHs</b> (ppb, dry wt)					
2-Methylnaphthalene	ND	41	78	40	20
Naphthalene	43	37	43	ND	20
Acenaphthylene	ND	ND	ND	ND	20
Acenaphthene	ND	ND	ND	ND	20
Fluorene	ND	ND	ND	ND	20
Phenanthrene	49	ND	ND	58	20
Anthracene	ND	ND	ND	ND	20
Fluoranthene	30	39	42	ND	20
Pyrene	36	47	ND	ND	20
Chrysene	ND	ND	ND	ND	20
Benzo(a)anthracene	ND	ND	ND	ND	20
Benzo(b)fluoranthene	ND	ND	ND	ND	20
Benzo(k)fluoranthene	ND	ND	ND	ND	20
Benzo(a)pyrene	52	ND	ND	ND	20
Indeno[1,2,3-CD]pyrene	ND	ND	ND	ND	20
Dibenzo(a,h)anthracene	ND	ND	ND	ND	20
Benzo[ghi]perylene	ND	ND	ND	ND	20
total PAHs	160	160	160	98	20
<b>PHTHALATES</b> (ppb, dry wt)					
total phthalates	160	160	170	1200	10
<b>DIOXINS</b> (pptr, dry wt)					
2,3,7,8-TCDD	ND	ND	ND	ND	0.22-0.33
Total TCDD	ND	ND	ND	ND	"
2,3,7,8-TCDF	ND	0.51	ND	ND	0.13-0.45
Total TCDF	ND	0.94	ND	ND	"

Table 4, continued. Sediment Chemistry Summary, Humboldt Bay Baseline Survey II (FY 1994): Composites only; for individual sample results, see Appendix C.

Analyte	Sampling Sections				Detection Limit
	SAMTB	EKUP	FLT B	REF	
<b>GRAIN SIZE (% dry)</b>					
Coarse Sand/Gravel ( $\Phi < -1$ )	0.8	0.3	0.2	0.0	-
Sand ( $-1 \leq \Phi \leq 4$ )	80.1	60.8	41.4	59.3	-
Silt ( $5 \leq \Phi \leq 8$ )	12.4	26.2	39.4	33.8	-
Clay ( $\Phi \geq 9$ )	6.7	12.7	19.0	6.9	-
<b>MISCELLANEOUS CHEMISTRIES</b>					
Total sulfides (ppm, dry)	29	110	35	0.3	0.1
Water soluble sulfides (ppm, dry)	ND	ND	0.2	ND	0.1
Oil & Grease (ppm, dry)	ND	ND	31	ND	20
Petroleum Hydrocarbons (ppm, dry)	ND	ND	ND	ND	20
% Solids (%)	77	72	65	73	0.1
TOC (%)	0.3	0.7	0.8	0.4	0.1
<b>CHLORINATED PESTICIDES (ppb, dry weight)</b>					
Aldrin	ND	ND	ND	ND	0.5
alpha-BHC	ND	ND	ND	ND	1.0
beta-BHC	ND	ND	ND	ND	1.0
delta-BHC	ND	ND	ND	ND	1.0
gamma-BHC (lindane)	ND	ND	ND	ND	1.0
alpha-Chlordane	ND	ND	ND	ND	5.0
gamma-Chlordane	ND	ND	ND	ND	5.0
4,4'-DDD	ND	ND	ND	ND	1.0
4,4'-DDE	ND	ND	ND	ND	1.0
4,4'-DDT	ND	ND	ND	ND	1.0
Dieldrin	ND	ND	ND	ND	0.5
Endosulfan I	ND	ND	ND	ND	2.0
Endosulfan II	ND	ND	ND	ND	0.5
Endosulfan sulfate	ND	ND	ND	ND	10
Endrin	ND	ND	ND	ND	0.5
Endrin aldehyde	ND	ND	ND	ND	0.5
Heptachlor	ND	ND	ND	ND	0.5
Heptachlor epoxide	ND	ND	ND	ND	10
Toxaphene	ND	ND	ND	ND	30
<b>PCBs (ppb, dry weight)</b>					
PCB 1242	ND	ND	ND	ND	20
PCB 1248	ND	ND	ND	ND	20
PCB 1254	ND	ND	ND	ND	20
PCB 1260	ND	ND	ND	ND	20
total PCBs	ND	ND	ND	ND	20

Table 5. Sediment Pore Water Salinity and Total Ammonia, Humboldt Bay Baseline II (FY 1994).

Composite	Initial (As Received)		Final
	S‰	NH <sub>3</sub>	NH <sub>3</sub>
REF	34.5	8.7	6.2
EKUP	30.5	6.1	4.1
SAMTB	32.0	4.0	2.6
FLTB	33.0	57.7 <sup>1</sup>	8.6

<sup>1</sup> Exceeds threshold value for *R. abronius* bioassay; sample purged as per EPA memorandum of 12/23/93 (see text).

Table 6. Bivalve larvae (*M. edulis*) SPP Bioassays, Humboldt Bay Baseline Survey II (FY1994). See text for explanation of calculations (Mean initial recovery = 4795).

Sample ID	Number		Total Recovered per 1 mL	Resuspended Volume	Total # Normal Larvae Recovered	% Survival	Mean % Survival $\pm$ S.D.	% Normal Development	Mean % Normal Development $\pm$ S.D.	Survival		Normal Development	
	Normal	Abnormal								Abbotts Corrected Value	Mean Corrected Value	Abbotts Corrected Value	Mean Corrected Value
Control	1	126	2	50.5	6363	132.7	98.4	98.4	98.4				
	2	91	3	51	4641	96.8	101.8	96.8	99.0				
	3	84	0	51	4284	89.3	$\pm$	100.0	$\pm$				
	4	104	0	49	5096	106.3	19.24	100.0	1.42				
	5	78	0	51.5	4017	83.8	100.0	100.0					
REF 100%	1	88	2	47	4136	86.3	97.8	97.8	98.2	84.8	98.7	98.7	98.7
	2	70	1	48.5	3395	70.8	86.4	98.6	98.2	69.6	99.5	99.5	99.1
	3	98	1	48	4704	98.1	$\pm$	99.0	$\pm$	96.4	99.9	99.9	$\pm$
	4	85	3	48	4080	85.1	10.16	96.6	1.00	83.6	97.5	97.5	1.01
	5	90	1	49	4410	92.0	98.9	98.9		90.4	99.9	99.9	
EKUP 100%	1	90	0	47.5	4275	89.2	100.0	100.0	98.8	87.6	101.0	101.0	99.7
	2	92	4	49	4508	94.0	93.5	95.8	98.8	92.4	91.8	96.8	$\pm$
	3	78	0	49.7	3877	80.8	$\pm$	100.0	$\pm$	79.4	$\pm$	101.0	$\pm$
	4	94	2	47	4418	92.1	11.08	97.9	1.86	90.5	10.89	98.9	1.88
	5	111	0	48	5328	111.1	100.0	100.0		109.2	101.0	101.0	
SAMTB 100%	1	89	2	47.9	4263	88.9	97.8	97.8	98.0	87.4	98.7	98.7	98.9
	2	101	2	49	4949	103.2	94.8	98.1	98.0	101.4	93.2	99.0	$\pm$
	3	104	3	48.5	5044	105.2	$\pm$	97.2	$\pm$	103.4	$\pm$	98.1	$\pm$
	4	80	1	48	3840	80.1	10.41	98.8	0.56	78.7	10.23	99.7	0.57
	5	98	2	47.3	4635	96.7	98.0	98.0		95.0	98.9	98.9	
FLTB 100%	1	72	12	47.5	3420	71.3	85.7	85.7	87.5	70.1	86.5	86.5	88.3
	2	81	7	47	3807	79.4	78.0	92.0	87.5	78.0	76.6	92.9	$\pm$
	3	77	14	47	3619	75.5	$\pm$	84.6	$\pm$	74.2	$\pm$	85.4	$\pm$
	4	71	11	49.5	3515	73.3	7.61	86.6	2.90	72.0	7.48	87.4	2.93
	5	91	12	47.7	4341	90.5	88.3	88.3		88.9	89.2	89.2	

ENDPOINT ESTIMATES:  
 LC<sub>50</sub> >100%  
 EC<sub>50</sub> >100%  
 EKUP: >100%  
 SAMTB: >100%  
 FLTB: >100%

Table 7. Mysid SPP Bioassays, Humboldt Bay Baseline Survey II (FY 1994).

*Holmesimysis costata*  
Suspended Particulate Phase Bioassay Results  
Humboldt Harbor Sediments

NUMBER OF SURVIVORS (Start n = 10)					
Rep #	Control	REF	EKUP	SAMTB	FLT B
1	10	9	10	10	10
2	10	10	10	10	10
3	9	10	10	10	10
4	10	10	10	10	10
5	10	10	10	10	10
Mean	9.8	9.8	10.0	10.0	10.0
SD	0.45	0.45	0.0	0.0	0.0
Mean % Survival	98	98	100	100	100

1. Data **FAIL** SHAPIRO-WILKS TEST for normality at P=0.01:

W=0.575

D = 1.600

Critical  $W_{(25, 0.01)} = 0.888$

2. Data **FAIL** BARTLETT'S TEST for homogeneity of variance at  $\alpha=0.01$ : At least one group has zero variance.

3. Steel's Many-One Rank test shows **NO significant difference** among sample data and disposal site reference:

Critical value = 17 (0.05, k=3)

	<u>EKUP</u>	<u>SAMTB</u>	<u>FLT B</u>
Rank Sum:	30.0	30.0	30.0



Table 8. Fish (*C. stigmaeus*) suspended particulate phase bioassays, Humboldt Bay Baseline Survey II (FY 1994).

*Citharichthys stigmaeus*  
 Suspended Particulate Phase Bioassay Results  
 Humboldt Harbor Sediments

NUMBER OF SURVIVORS (Start n = 10)					
Rep #	Control	Reference	EKUP	SAMTB	FLTB
1	10	10	10	10	0
2	10	10	10	9	0
3	10	10	10	10	2
4	10	9	9	10	0
5	10	10	9	7	0
Mean	10.0	9.8	9.6	9.2	0.40*
SD	0.0	0.45	0.55	1.3	0.89
Mean % Survival	100	98	96	92	4.0

1. Data **PASS** SHAPIRO-WILKS TEST for normality at P=0.01:

W=0.992

D = 12.00

Critical  $W_{(25, 0.01)} = 0.888$

2. Data **FAIL** BARTLETT'S TEST for homogeneity of variance at  $\alpha=0.01$ : At least one group has zero variance.

3. Steel's Many-One Rank test shows **FLTB** significantly different from the disposal site reference composite.

Critical F value = 17 (0.05, k=4)

	<u>EKUP</u>	<u>SAMTB</u>	<u>FLTB</u>
Rank Sum:	9.6	9.2	0.40*

Table 9. Amphipod (*R. abronius*) solid phase static bioassays, Humboldt Bay Baseline Survey II (FY 1994). Home A and composite FLT B replicates were ammonia-purged (daily renewals) as per EPA/ACOE memo of 21 December 1993.

*Rhepoxynius abronius*  
Solid Phase Static Bioassay Results  
Humboldt Harbor Sediments

NUMBER OF SURVIVORS (Start n = 20)						
Rep #	Home A	Home B	REF	EKUP	SAMTB	FLT B
1	18	19	18	18	19	20
2	19	20	18	17	19	19
3	20	19	18	18	20	17
4	18	18	16	20	19	18
5	20	20	18	20	20	20
Mean	19.0	19.2	17.6	18.6	19.4	18.8
SD	1.00	0.84	0.89	1.34	0.55	1.30
Mean % Survival	95	96	88	93	97	94
Mean % Reburial	100	96.8	98.9	96.9	99.0	97.0

1. Data **PASS** SHAPIRO-WILKS TEST for normality at P=0.01:

W = 0.936                      D = 18.400                      Critical  $W_{(20, 0.01)} = 0.868$

2. Data **PASS** BARTLETT'S TEST for homogeneity of variance at  $\alpha=0.01$ :

Calculated B statistic = 3.14                      Table Chi-square value = 11.34

3. ANOVA test shows **NO significant difference** among sample means and disposal site reference:

Critical F value = 3.24 (0.05, 3, 16)                      Calculated F value = 2.435  
Calculated F > Critical F;  $\therefore$  **Fail to Reject  $H_0$** : all groups equal

4. DUNNETT'S TEST (Mean Comparison Test) shows **NO Humboldt Harbor sample composite with lower survival** than the Humboldt reference composite at P = 0.05:

	<u>EKUP</u>	<u>SAMTB</u>	<u>FLT B</u>
Dunnett's t:	-1.474	-2.654	-1.769
(1-tailed, P=0.05, d.f.=16,3)			

Dunnett table value = 2.23

Table 10. Mysid (*H. costata*) solid phase flow-through bioassays, Humboldt Bay Baseline Survey II (FY 1994).

*Holmesimysis costata*  
 Solid Phase Flow-Through Bioassay Results  
 Humboldt Harbor Sediments

NUMBER OF SURVIVORS (Start n = 20)					
Rep #	Home	REF	EKUP	SAMTB	FLTB
1	19	20	18	19	19
2	19	19	20	19	18
3	20	19	20	19	20
4	18	18	20	20	18
5	19	19	19	19	20
Mean	19.0	19.0	19.4	19.2	19.0
SD	0.71	0.71	0.89	0.45	1.00
Mean % Survival	95	95	97	96	95

1. Data **PASS** SHAPIRO-WILKS TEST for normality at P=0.01:

W = 0.928                      D = 10.000                      Critical  $W_{(20, 0.01)} = 0.868$

2. Data **PASS** BARTLETT'S TEST for homogeneity of variance at  $\alpha=0.01$ :

Calculated B statistic = 2.34                      Table Chi-square value = 11.34

3. ANOVA test shows **NO significant difference** among sample means and disposal site reference:

Critical F value = 3.24 (0.05, 3, 16)                      Calculated F value = 0.293  
 Calculated F > Critical F;  $\therefore$  **Fail to Reject  $H_0$** ; all groups equal

4. DUNNETT'S TEST (Mean Comparison Test) shows **NO Humboldt Harbor sample composite with lower survival** than the Humboldt reference composite at P = 0.05:

	<u>EKUP</u>	<u>SAMTB</u>	<u>FLTB</u>
Dunnett's t:	-0.800	-0.400	0.000
(1-tailed, P=0.05, d.f.=16,3)			

Dunnett table value = 2.23

Table 11. Polychaete worm (*N. caecoides*) solid phase flow-through bioassays, Humboldt Bay Baseline Survey II (FY 1994).

*Nephtys caecoides*  
 Solid Phase Flow-Through Bioassay Results  
 Humboldt Harbor Sediments

NUMBER OF SURVIVORS (Start n = 20)					
Rep #	Home	Reference	EKUP	SAMTB	FLTB
1	20	18	19	18	17
2	20	16	18	20	20
3	20	18	18	18	18
4	20	18	19	18	19
5	20	19	20	20	19
Mean	20.0	17.8	18.8	18.8	18.6
SD	0.0	1.10	0.84	1.10	1.14
Mean % Survival	100	89	94	94	93

1. Data **PASS** SHAPIRO-WILKS TEST for normality at P=0.01:

W = 0.918                      D = 17.600                      Critical  $W_{(20, 0.01)} = 0.868$

2. Data **PASS** BARTLETT'S TEST for homogeneity of variance at  $\alpha=0.01$ :

Calculated B statistic = 0.40                      Table Chi-square value = 11.34

3. ANOVA test shows **NO significant difference** among sample means and disposal site reference:

Critical F value = 3.24 (0.05, 3, 16)                      Calculated F value = 5.652  
 Calculated F > Critical F;  $\therefore$  **Reject  $H_0$** ; all groups equal

4. DUNNETT'S TEST (Mean Comparison Test) shows **NO sample composites produce lower survival** than the Humboldt reference composite at P = 0.05:

	<u>EKUP</u>	<u>SAMTB</u>	<u>FLTB</u>
Dunnett's t:	-1.508	-1.508	-1.206

(1-tailed, P=0.05, d.f.=16,3)

Dunnett table value = 2.23

FIGURES



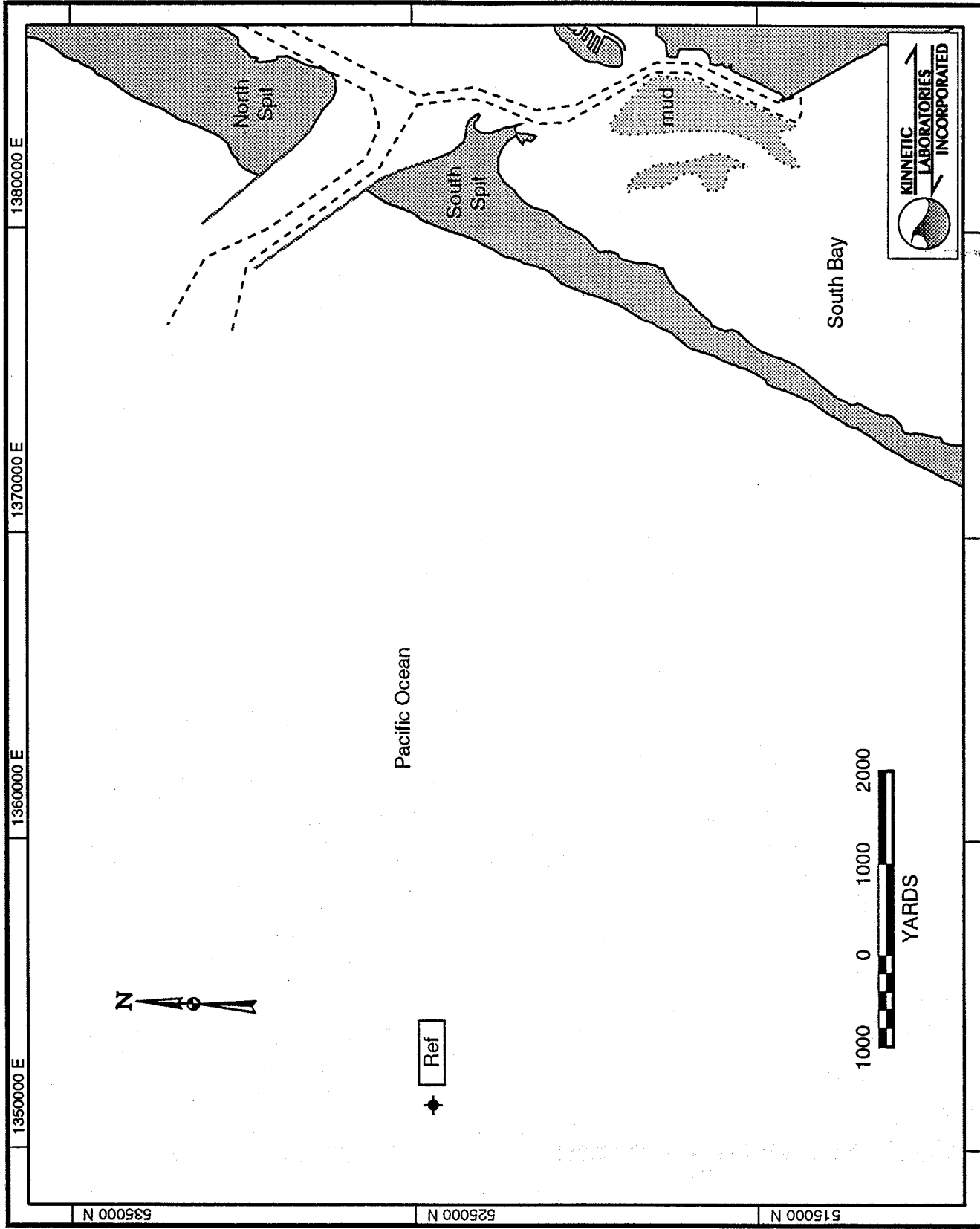


Figure 1. Humboldt Bay FY 1994 sampling locations. Reference station (solid) composite of six grab samples.

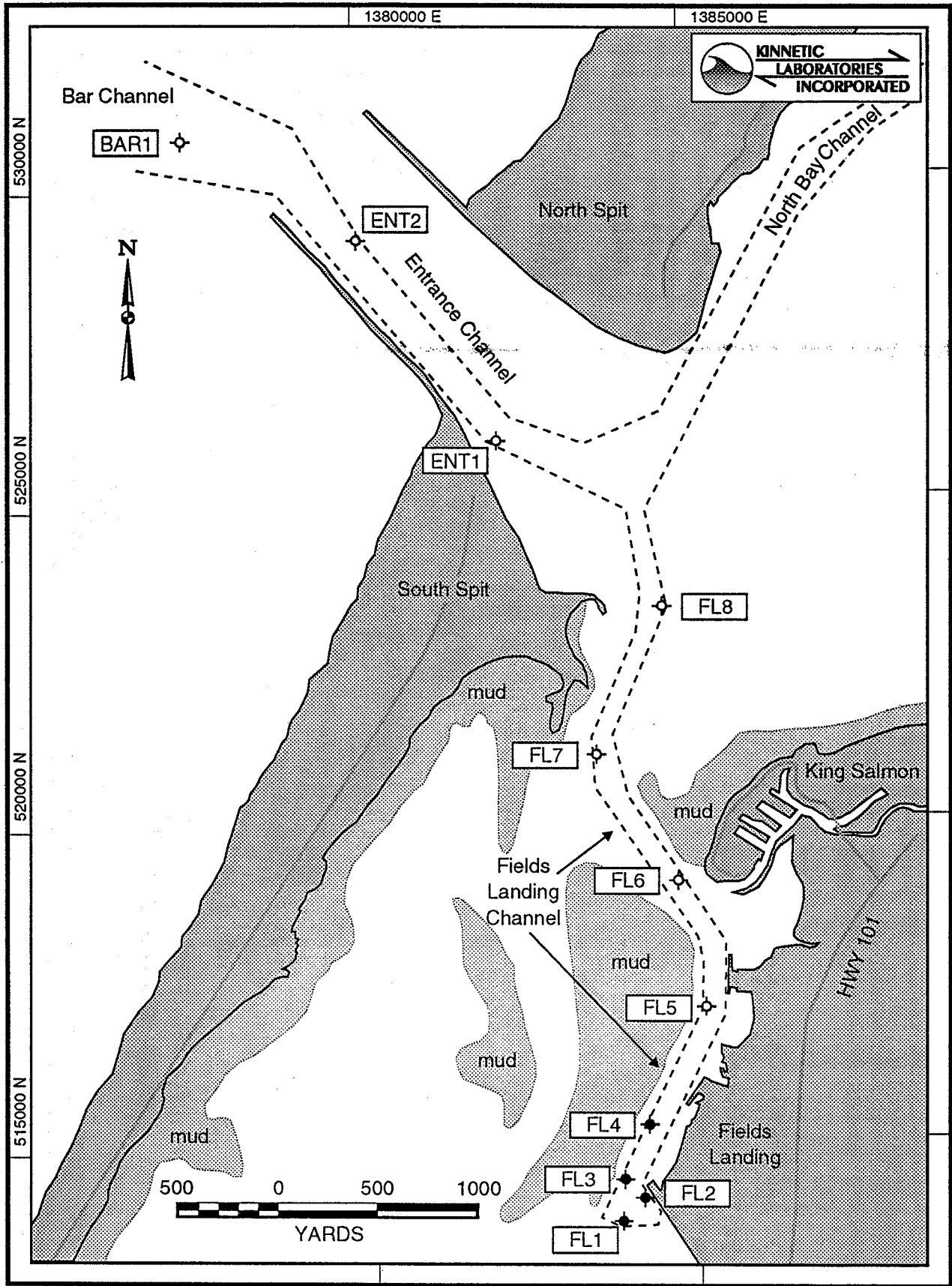


Figure 2. Humboldt Bay FY1994 sampling locations. Stations FL1 through FL8, ENT1, ENT2, and BAR1. Solid stations indicate those used in Fields Landing Lower Channel and Turning Basin (FLTB) composite.



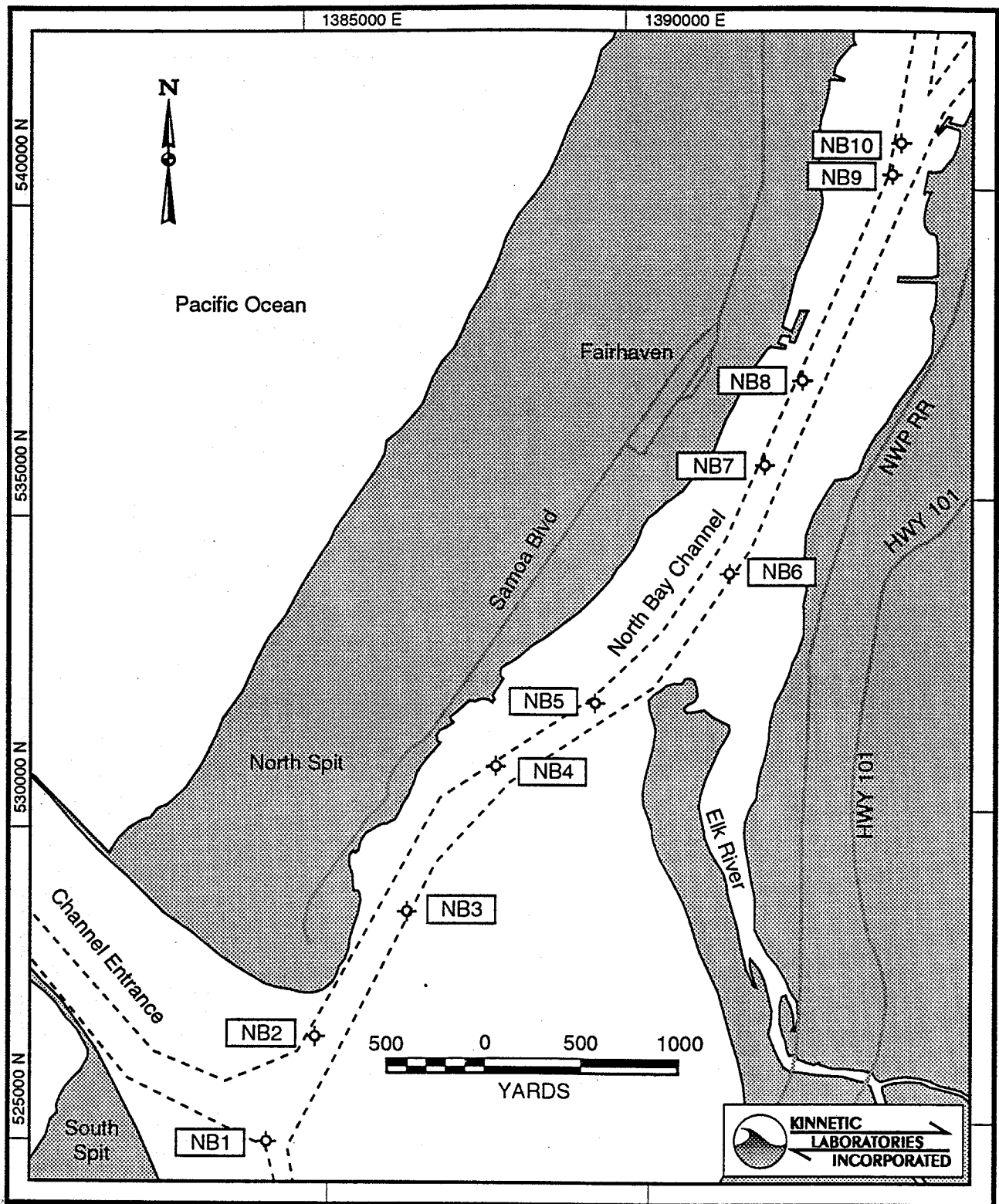


Figure 3. Humboldt Bay FY1994 sampling locations. Stations NB1 through NB10.

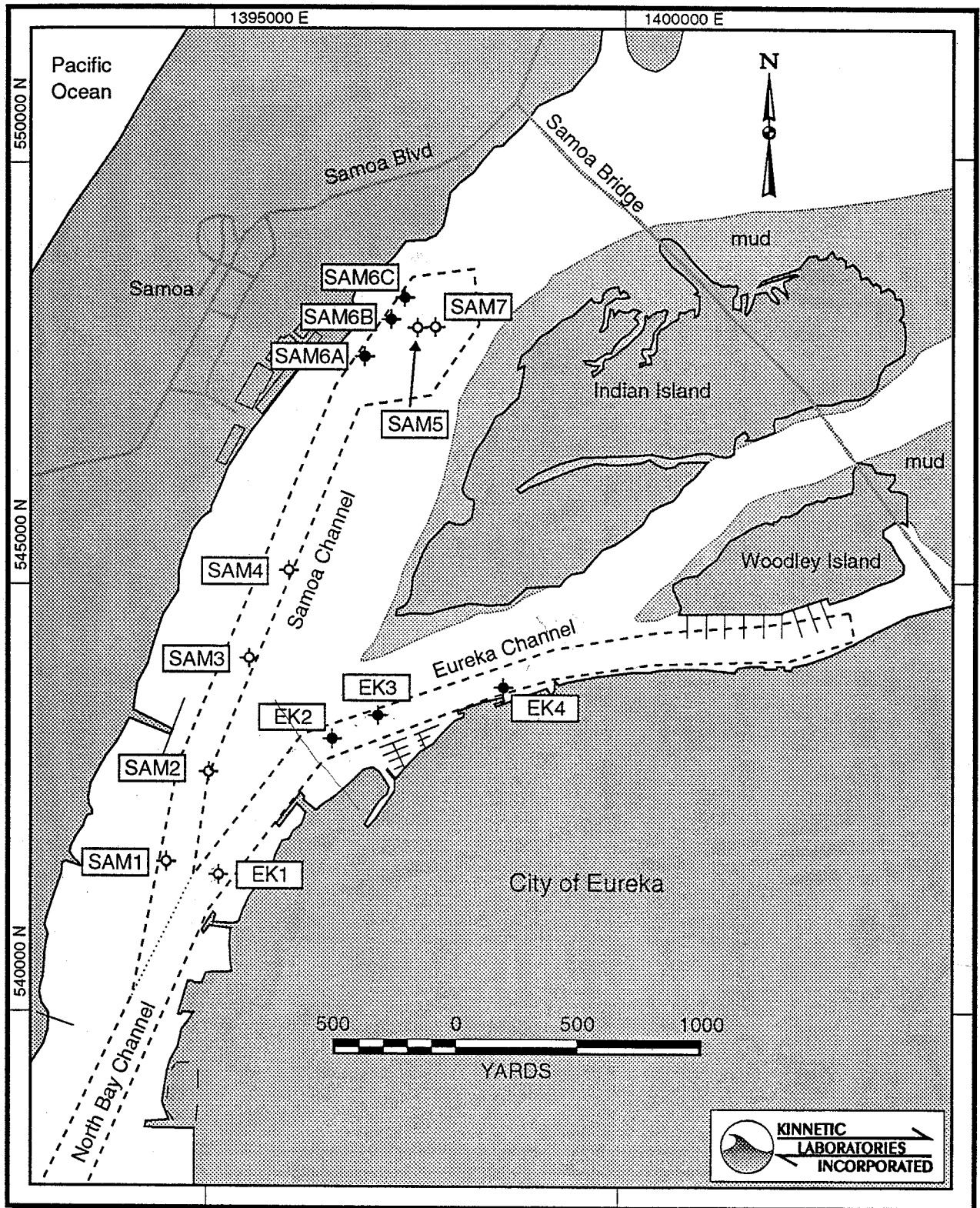


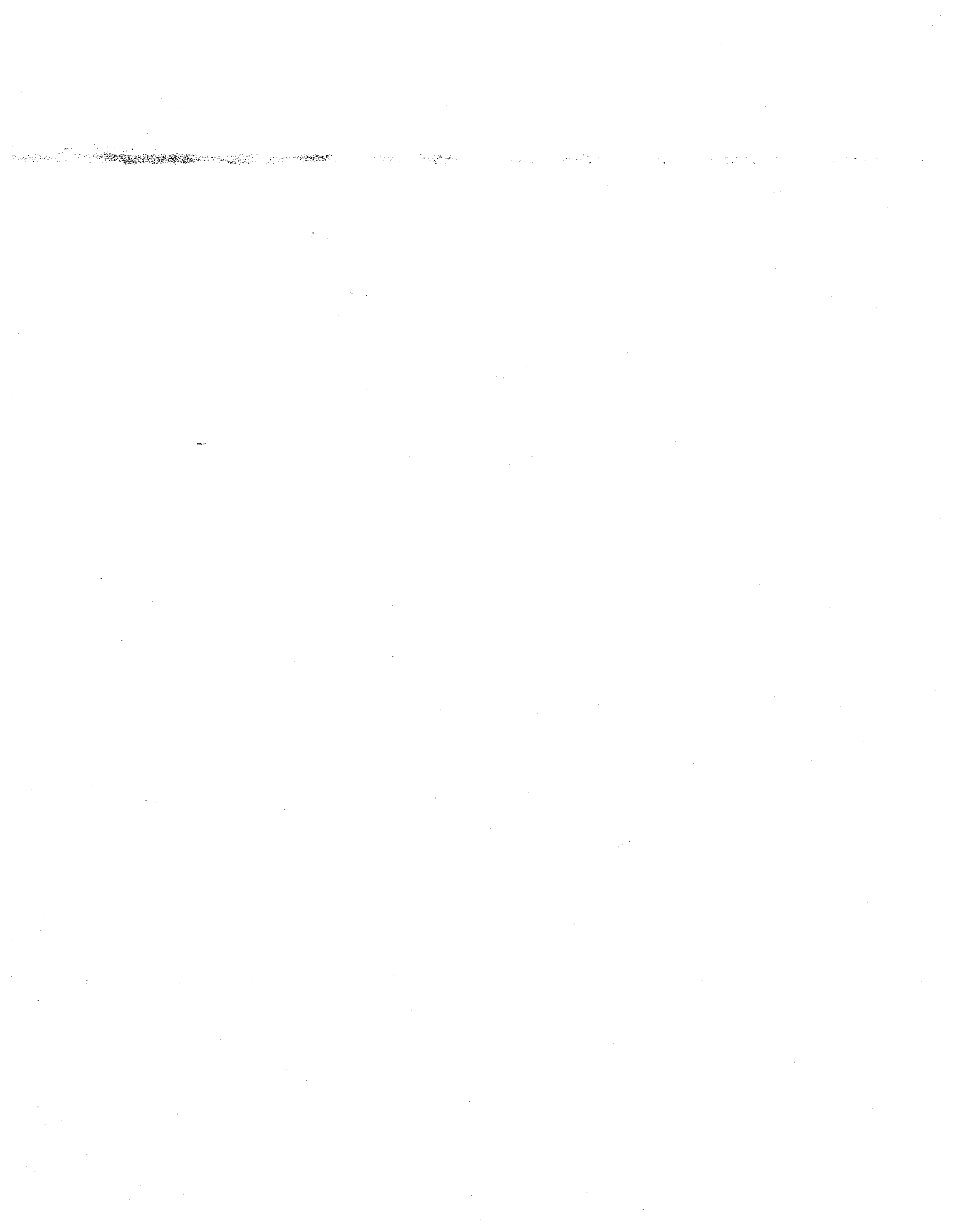
Figure 4. Humboldt Bay FY1994 sampling locations. Stations EK1 through EK4 and SAM1 through SAM7. Solid stations indicate those used in Eureka Upper Channel (EKUP) and Samoa Turning Basin (SAMTB) composites.

# APPENDICES



# Appendix A

## Scope of Services



September 1, 1994  
Revised Scope of Services  
Sediment Chemical, Bioassay and Bioaccumulation Testing  
Humboldt Harbor FY 94 Maintenance Dredging

Note: This Revised Scope of Service contains the following three specific modifications to the Revised Scope of Services dated April 8, 1994, which the contractor should address in his revised proposal:

Change 1 (section 5a). The following has been added (in bold) to section 5a: This task has been completed since the award of the initial contract and the number of samples requiring chemistry, including composited samples, is 27.

Change 2 (section 7d2). The following has been added (in bold) to section 7d(2): This task has been completed since the award of the contract and was determined necessary for Composite FL.

Change 3 (section 8c(1)). The Corps and EPA have decided to not conduct tissue chemistry on tissues archived by the contractor for bioaccumulation evaluation. This decision was made by the two agencies after review of sediment chemistry, which was provided by the contractor as part of this contract. Therefore, this section has been deleted from this revised scope of service and no longer required.

1. PURPOSE. The purpose of this contract is to perform bulk sediment analyses, suspended particulate bioassays, solid phase bioassays, and bioaccumulation testing of sediments collected from Humboldt Harbor and Bay. The testing will assist in determining whether the material from Humboldt Harbor and Bay is suitable for aquatic disposal in compliance with Section 103 of the Marine Protection Research and Sanctuaries Act.

2. THE CONTRACTOR'S RESPONSIBILITY. The Contractor shall furnish all necessary labor, facilities, equipment, and materials to perform the work described under this contract. The Contractor's representative shall be available to meet with Government personnel as requested by the USACE San Francisco District. The Contractor shall perform the services in accordance with this statement of work and the general provisions. Any modifications in equipment and/or methodology from those outlined in this Scope of Services must be approved by the San Francisco District (SFD). In order to adhere to the project schedule, all requests for modification or variations in equipment or procedures shall be forwarded to the SFD at the earliest date/time to ensure a timely review. The Contractor shall comply with all pertinent provisions of the U.S. Army Corps of Engineers Safety and Health Requirements Manual EM-385-1-1, date October 1984. The Contractor shall provide transportation and access from shore to the sampling vessel to a representative of the U.S. Army Corps of Engineers who may be present during sampling.

SEDIMENT SAMPLING LOCATIONS

a. Samoa, Eureka, Fields Landing, North Bay, Bar and Entrance Channels. Sediment samples shall be taken at those sites

listed in Table 1 (shown in Figure 1). A total of three composites shall be made according to the compositing scheme in Table 2.

b. A sufficient amount of sediment shall be collected from each location specified in Table 1, so that a representative amount of sediment is included from each sampling location in each composite, and that there is sufficient composited sediment to run the initial suite of sediment chemistry, bioassays and bioaccumulation and also one additional sediment chemistry, suspended particulate phase, and solid phase toxicity bioassay on each composite should a re-test be necessary. Sufficient individual sediment from each sediment location within a composite area shall be taken to conduct individual sediment chemistry analyses.

c. All of the samples shall have their containers physically marked as to area, sample location, and purpose of sampling. The Contractor shall furnish SFD an inventory of all samples taken and delivered, and their respective labels.

d. Sediment samples shall be placed in appropriate containers and stored following methodologies described in the manual. Care shall be taken to ensure that the containers are completely filled by the samples and that air bubbles are not trapped in the containers. All samples shall be stored immediately at 4°C and not frozen or dried. The Contractor shall provide the ice and ice chests or chest freezers to be used in the field to maintain samples at 4 C. These samples shall be stored at 4 C until testing initiated.

e. That portion of each individual sediment sample remaining after analyses shall be archived at 4°C. for possible additional chemical analyses until completion of the work and acceptance of the final report. Disposal of all sediments remaining at the end of testing shall be the Contractor's responsibility.

f. The Contractor shall provide the mudline elevations at each sample gathering location in reference to mean lower low water.

g. The Contractor shall maintain a daily field activity log listing the beginning and ending time for every and all phases of operation.

h. Formal chain-of-custody procedures shall be followed and documented.

#### 4. SEDIMENT SAMPLING EQUIPMENT

a. Sediments in the Samoa, Eureka, Fields Landing, and North Bay channels shall be sampled with vibracore equipment. Each of the sampling locations within Humboldt Bay and Harbor sampled by vibracore shall be sampled from mudline to project depths (MLLW) listed on Table 1. Material below the required depths listed on Table 1 shall not be



used for testing. Where there is less than a foot of sediment at the sampling location or attempts to sample with the Vibracore equipment has failed, sediment samples at that location shall be obtained with either a Van Veen Grab sampler or a pipe dredge sampler. Samples from the Bar and Entrance channels, reference site, and control site shall be sampled using either a Van Veen Grab Sampler or equivalent, or a pipe dredge.

b. A fathometer shall be used to ensure vertical control of sampling. Horizontal positioning equipment with an accuracy of ten (10) feet is required to locate sampling points within the harbor. An accuracy of fifty (50) feet is required to locate the sampling site of the reference area.

c. Each individual sediment core sample taken in the Humboldt channels shall be taken within an area bounded by a 50-foot radius having its center located at the coordinates provided above or as approved by the government representative. In the event that there is insufficient sediment to sample between mudline elevation and the sampling depth listed above, with either the vibracore or grab sampler, the contractor shall locate as close as possible to the original sampling site, a new sampling location (inside the channel lines) which will provide sufficient sediment for sampling.

d. Care shall be taken during sampling to avoid contamination of sediment. All coring devices, if possible, shall be composed of or lined with a noncontaminating material such as cellulose buterate or lexan. If this is not possible, the Contractor must document what steps will be taken to prevent contamination of sediments during sampling as well as during storage prior to initiation of testing. Any samples indicating external contamination due to handling shall require resampling at no additional cost to the SFD.

## 5. SEDIMENT CHEMICAL, PHYSICAL AND GEOLOGICAL CHARACTERIZATION.

a. Grain size analyses shall be completed for all individual sediment samples taken in each of the Humboldt Harbor and Bay channels. Individual sediment samples taken in the Bar, Entrance, and North Bay channels, which are found to not be predominantly sand (if <80% retained on #200 sieve), and are not included in a compositing area, shall be analyzed for the parameters specified in Table 3. All composited sediments from Humboldt Harbor channels, the reference site, and the control, and all individual sediments sampled within the Fields landing, Samoa, and Eureka channels, shall be analyzed for the parameters specified in Table 3. In addition, for each composited sediment, Dioxin/Furan analyses shall be conducted. The required detection limits are also given in Table 3. The results shall be reported in dry weight. This task has been completed since the award of the initial contract and the number of samples requiring chemistry, including composited samples, is 27.

b. All analyses must be conducted using EPA approved methodologies that are suitable for marine sediments and which yield the required detection limits with good precision and accuracy. Appropriate clean-up procedures shall be employed that remove as much of the interfering material as possible from the sample without compromising the integrity of the sample or increasing the detection limits.

c. The presence of major "unknown" analytes on gas chromatograms or reconstructed ion chromatography (GC/MS) should be noted.

d. Grain size analysis and hydrometer readings shall be performed in accordance with the grain size procedure found in "Procedures for Handling and Chemical Analysis of Sediment and Water Samples, U.S. Army Corps of Engineers Technical Committee on Criteria for Dredged and Fill Material (Plumb 1981)".

**TABLE 1. Humboldt Sampling Locations**

CHANNEL	SAMPLE	NORTHING	EASTING	Estimated depth to mudline (MLLW)	Sample to maximum Depth of (MLLW)
<b>North Bay</b>					
	NB1	525,070	1,384,200	GRAB	37
	NB2	525,920	1,383,850	GRAB	37
	NB3	528,610	1,386,270	GRAB	37
	NB4	530,600	1,387,800	GRAB	37
	NB5	531,750	1,389,435	GRAB	37
	NB6	533,710	1,391,365	GRAB	37
	NB7	535,691	1,392,300	36	37
	NB8	537,165	1,392,987	36	37
	NB9	538,680	1,393,630	35	37
	NB10	540,530	1,394,465	36	37
<b>SAMOA</b>					
	SAM1	541,698	1,394,581	36.5	37
	SAM2	542,620	1,34,962	35.5	37
	SAM3	544,057	1,395,362	35-36	37
	SAM4	545,480	1,396,110	35-36	37
	SAM5	547,270	1,397,500	34-35	37
	SAM6-A	547,592	1,396,836	36	37
	SAM6-B	547,998	1,397,080	36	37
	SAM6-C	548,368	1,397,403	36	37
	SAM7	548,480	1,398,061	36	37
<b>EUREKA</b>					
	EK1	541,498	1,395,132	36.5	37
	EK2	543,115	1,396,720	26.5	28
	EK3	543,600	1,397,863	27	28
	EK4	543,792	1,398,985	26.5	28
<b>FIELDS LANDING</b>					
	FL1	513,800	1,383,820	27.5-28	28

CHANNEL	SAMPLE	NORTHING	EASTING	Estimated depth to mudline (MLLW)	Sample to maximum Depth of (MLLW)
	FL2	514,070	1,384,130	27.5-28	28
	FL3	514,250	1,383,790	28-30	28
	FL4	515,660	1,384,580	28	28
	FL5	517,305	1,385,100	27	28
	FL6	519,220	1,384,600	27	28
	FL7	521,140	1,383,510	25	28
	FL8	523,300	1,384,500	27	28
ENTRANCE	ENT1	526,110	1,382,040	Grab	45
	ENT2	529,240	1,379,860	Grab	45
BAR	BAR1	531,010	1,377,490	GRAB	45
Reference site	RF	40°49'41"	124°18'34"	GRAB OR PIPE DREDGE	165-165' or 26.5-27.0 fathoms
Control Site	Tomaes Bay	38°13'50"	172°57'40"		

**Table 2. Compositing Plan**

<b>Composite</b>	<b>SAMPLE</b>
<b>EKUP</b>	<b>EK1</b>
<b>EKUP</b>	<b>EK2</b>
<b>EKUP</b>	<b>EK3</b>
<b>EKUP</b>	<b>EK4</b>
<b>SAMTB</b>	<b>SAM6-A</b>
<b>SAMTB</b>	<b>SAM6-B</b>
<b>SAMTB</b>	<b>SAM6-C</b>
<b>FLTB</b>	<b>FL1</b>
<b>FLTB</b>	<b>FL2</b>
<b>FLTB</b>	<b>FL3</b>
<b>FLTB</b>	<b>FL4</b>
<b>Ref</b>	<b>Reference</b>

\* Only placed in composite if >80% passes through #200 sieve

Table 3 Designation of Parameters for Analysis

Detection Limit (mg/kg dry wt) <sup>(a)</sup>

Parameters

Sediment  
Conventionals

TOC	0.1%
Oil and Grease	20
TPH	20
Grain Size	NA
Total Solids	0.1%
Total and Water Soluble Sulfides	0.1

Metals

Ag	0.1
As	0.1
Cd	0.1
Cr	0.1
Cu	0.1
Hg	0.02
Ni	0.1
Pb	0.1
Se	0.1
Zn	1.0

Organic  
Compounds

Phthalate esters	0.01
Butyltins <sup>(b)</sup>	0.001
PCBs <sup>(c)</sup>	0.02

Pesticides- <sup>(e)</sup>

Aldrin	0.002
Alpha-BHC	0.002
Beta-BHC	0.002
Delta-BHC	0.002
Gamma-BHC	0.002
Alpha-Chlordane	0.002
Gamma-Chlordane	0.002
4,4'-DDD	0.002
4,4'-DDE	0.002
4,4'-DDT	0.002
Dieldrin	0.002
Endosulfan I	0.002
Endosulfan II	0.002
Endosulfan Sulfate	0.01
Endrine	0.002
Heptochlor	0.002
Heptochlor Epoxide	0.01
Toxaphene	0.03

TCDD/TCDF <sup>(f)</sup>

1 (part per trillion)

PAHs <sup>(d)</sup>

NAPHTHALENE	0.02
ACENAPHTHYLENE	0.02
ACENAPHTHENE	0.02
FLUORENE	0.02
PHENANTHRENE	0.02
ANTHRACENE	0.02
FLUORANTHENE	0.02
PYRENE	0.02
CHRYSENE	0.02
BENZO (A) ANTHRACENE	0.02
BENZO (B) FLUORANTHENE	0.02

BENZO (K) FLUORANTHENE	0.02
BENZO (A) PYRENE	0.02
INDENO (1,2,3-CD) PYRENE	0.02
DIBENZ (A,H) ANTHRACENE	0.02
BENZO (GHI) PERYLENE	0.02

- (a) Report as mg/kg dry wt., unless otherwise noted.
- (b) Mono-, Di-, and Tributyltin.
- (c) Reported as Aroclor equivalents 1242, 1248, 1254, and 1260 and total PCB.
- (d) All compounds on EPA Method 610 list.
- (e) All compounds on EPA Method 608 list.
- (f) Only on composited sediments

SAMTB (Samoa Turning Basin)

SAM-6-C  
SAM-6-B  
SAM-6-A

SAM-7

SAM-5

SAM-4

EKUP (Eureka)

SAM-3

EK-2-3

EK-3

EK-4

SAM-2

EK-1

NB-10

NB-9

NB-7

NB-6

NB-5

NB-4

NB-3

NB-2

ENT-1

NB-1

FL-6

FL-7

FL-8

FL-5

FL-4

FL-3

FL-2

FL-1

FLTB (Fields Landing Turning Basin)



HUMBOLDT HARBOR  
FIGURE 1 - Samples and Composites





Note: Throughout the following discussions on bioassays the term Manual refers to the Evaluation of Dredged Material for Ocean Disposal, Testing Manual (EPA-503/8-91/001, February 1991) developed by the EPA Office of Marine and Estuarine Protection and U. S. Army Corps of Engineers, available through the Corps of Engineers' Waterways Experiment Station, Telephone (601)634-2571.

6. SUSPENDED PARTICULATE PHASE BIOASSAYS.

a. Sediment and Water Collection. The Contractor shall collect and preserve all sediment samples as described in sections 3 and 4 above and in the Manual. Water shall be clean, uncontaminated seawater of appropriate salinity, pH and temperature. Sufficient water shall be collected to perform the required tests. Seawater from any suitable location may be used provided it does not exceed applicable EPA quality criteria for marine waters and is of constant quality. Contractors shall be able to provide evidence that water meets these criteria, if necessary. Testing shall be conducted on the composited samples as specified in sections 3 and 4 above.

b. Preparation of the Bioassay Phase.

(1) Suspended Particulate Phase Bioassay. Phase preparation shall follow the procedure in the Manual for the suspended phase.

(2) Water Samples. Preparation of water samples shall follow the Manual.

(3) Sediment Sample. Composited sediment samples from Humboldt Bay and Harbor shall be prepared according to the manual. In addition to the treatment composites, there shall be the control water, reference water, and reference sediments. The control and reference water may be the same if the animals are being held before testing in the same water to be used for the bioassays.

c. Collection and Maintenance of Test Species.

(1) Species Selection. Three species shall be used: (1) Larvae of (pacific oyster) *Crassostreaa gigas* or (bay mussel) *Mytilus edulis* (% normal development to D stage) (2) (mysid shrimp) *Holmesimysis sculpta*, and (3) (juvenile sanddab) *Citharichthys stigmatæus*.

(2) Organism Handling and Holding. Organisms shall be held no longer than two weeks. The SFD must approve additional holding time. Experiments shall be designed and performed so that organisms are handled as minimally as possible. Procedures for handling are found in the Manual. The physiological and biological needs of the test organisms must be met at all times.

d. Bioassay Testing of the Suspended Phase. Five replications of each treatment (including control) shall be performed. If greater than 10% of the control dies during any test, that test must be repeated at no additional expense to the SFD. However, control mortalities of 30% are acceptable in zooplankton

bioassays. Conditions and procedures shall follow those found in the Manual, unless otherwise noted.

e. Deviations From the Manual. If there is an odor of hydrogen sulfide, the water shall be aerated until the odor of hydrogen sulfide is no longer detected. The Contractor shall measure NH<sub>3</sub> in the test containers. If the NH<sub>3</sub> concentration is elevated, the water shall be aerated until the concentration is adequately reduced before introducing the test organisms.

f. Experimental Design. The design is a completely randomized design with three dilutions per dredging area per species, three reference sediments, and a control.

<u>Suspended Particulate Treatments</u>	<u>As a Reference</u>	<u>As a Control</u>
For each dredge area:	(1) 100% marine water	(1) 100% culture water
(1) 100% Suspended Particulate Phase	(The following use reference sediment)	Note: May be the same as reference water
(2) 50% suspended particulate phase	(2) 100% suspended particulate phase	
(3) 10% suspended particulate phase	(3) 50% suspended particulate phase	
	(4) 10% suspended particulate phase	

The test organisms and treatment shall be randomly assigned to test containers. The variable measured shall be percent survival except for the bivalve larvae test for which both survival and percent normal development are measured. The EC50 and LC50 shall both be calculated according to ASTM E724-89. Each species shall be considered a separate test. The 100% suspended particulate phase may be run first. If mortalities (or abnormal development) of 50% or less occur by 48-96 hours, the 50% and 10% dilutions need not be run. If greater than 50% mortality (or abnormal development) occurs by 48-96 hours, the test must be rerun at the Contractor's expense using the full series of dilutions (100%, 50%, 10% and control).

g. Data Analysis for Suspended Particulate Bioassays.

(1) If total survival or percent normal development in the test medium is equal to or higher than survival in the reference or control, visual inspection of the data is adequate and no statistical analyses are needed.

(2) A table or tables shall be provided for each species tested, giving the number of organisms tested, the total number of

surviving organisms for each time period and each treatment, the mean, and the standard deviation.

(3) If mean percent survival or normal development in the control is greater than any of the other treatments, for the bioassays, than additional statistical analyses shall be performed. The statistical analyses shall be as described in the Manual. Any deviations from the Manual must be approved by the Government. The results of all statistical analyses shall be presented in tabular form.

(4) If 50 percent or greater mortality or abnormal development occurs in the highest concentration of test medium, than a LC50 or EC50 must be calculated as described in the Manual.

## 7. SOLID PHASE BIOASSAY

a. Sediment and Water Collection. The Contractor shall collect and preserve all sediment and water samples as described in sections 3 and 4 above and in the Manual. Compositing sediment samples shall be prepared and handled according to the Manual. For control sediment, the Contractor shall procure unpolluted sediment that is compatible with the test organisms and preferably from where they were collected. The control sediment must meet the needs of the organisms. The bioassays shall be conducted with a flow-through seawater system except for the test using the amphipod. Seawater of approximately 15°C, 30-32 ppt salinity should be passed through a sand filter and flow into each aquarium at a rate that will replace the aquarium volume at least once every 12 hours. The flow should be directed to achieve good mixing without disturbing the layer of sediment on the aquarium bottom. Water for all bioassays will be clean, uncontaminated seawater of appropriate salinity, pH and temperature. Seawater from any suitable location may be used provided it does not exceed applicable EPA quality criteria for marine waters and is of constant quality.

### b. Collection and Maintenance of Test Species.

(1) Species Collection. It is recommended that collection of species should include at least 20% more than the minimum requirement.

(2) Species Selection. Three species shall be used:  
(1) (Amphipod) *Rhepoxynius abronius*; (2) (burrowing polychaete) *Nephtys caecoides*, and (3) (mysid shrimp) *Holmesimysis costata*.

(3) Organism Handling and Holding. Organisms shall be held no longer than two weeks. The SFD must approve additional holding time. Experiments shall be designed and performed so that organisms are handled as minimally as possible. Procedures for handling are found in the Manual. The physiological and biological needs of the organisms must be met at all times.

c. Solid Phase Preparation and Experimental Design. The test treatments shall consist of the dredged material samples, a reference, and a control. Five replications of each treatment shall be

performed. Each replicate shall consist of at least 20 organisms of each of these species. The dredged material treatments, references, and control shall be prepared as described in the Manual. However, only whole sediments shall be used in the solid phase tests. Layering of test sediments or control sediments over reference sediments is no longer acceptable. The purpose of the control is to verify the health of test organisms and the acceptability of test conditions. It also provides for quality assurance. If the mean survival in the control is less than 90 percent, the test must be repeated at no additional cost to the SFD. The variable measured shall be percent survival. Each species shall be considered a separate test.

d. Solid Phase Testing. Conditions and procedures for the 10-day solid phase bioassay shall follow ASTM (E 1367-90), 1991 and the Manual. In the event of a discrepancy between the ASTM and the Manual the Contractor shall contact the Corp's Contracting Officer for clarification.

(1) The following table<sup>1</sup> contains test condition acceptability ranges for organisms used to evaluate dredged material.

**MARINE AND ESTUARINE AMPHIPOD TOXICITY TEST APPLICATION CONDITIONS\***

Parameter	<u>Rhepoxynius</u>	<u>Ampelisca</u>	<u>Eohaustorius</u>	<u>Leptocheirus</u>
Temperature (°C)	15	20	15	25
Overlying Salinity (ppt)	>25	>20	2-34	2-32
Grain Size (% silt/clay)	<90	>10	full range	full range
Ammonia (total mg/L, ph 7.7)*	<30	<30	<60	<60
Ammonia (UI** mg/L, ph 7.7)*	<0.4	<0.4	<0.8	<0.8
Sulfides	***	**	**	***

A framework for deciding whether observed sediment (or elutriate) toxicity may be due to ammonia is presented in EPA/USACE (1993: Appendix F). This document should be consulted if ammonia is suspected to be a contaminant of concern.

\*\* Unionized

\*\*\* Hydrogen Sulfide is not likely to be a problem in these tests if adequate oxygen levels are maintained in the overlying water.

(2) The contractor shall conduct measurements of interstitial ammonia, ph, and salinity on the sediments prior to the beginning and end of each bioassay test. Whenever chemical evidence of ammonia is present at toxicologically important levels prior to initiation of the test, ammonia in the interstitial water shall be reduced to below 20 mg/l before adding the benthic test organism. Ammonia levels in the interstitial water can be reduced by sufficiently aerating the sample at saturation and replacing two

volumes of water per day. During the test, the contractor shall ensure that ammonia concentrations remain within the acceptable range by conducting the toxicity test with continuous flow or volume replacement not to exceed two volumes per day. This task has been completed since the award of the contract and was determined necessary for Composite FL.

e. Data Analysis For Solid Phase Bioassay.

(1) If total survival in the test medium is equal to or higher than in the reference, visual inspection of the data is adequate and no statistical analyses are needed for that test.

(2) A table or tables shall be provided for each species tested, giving the number of organisms tested, the total number of surviving organisms for each treatment, the means, and the standard deviation.

(3) If mean percent survival in the reference is greater than any of the other treatments, for the bioassays, then additional statistical analyses shall be performed. The statistical analyses shall be as described in the Manual except that multiple t-test shall not be used. Alternative statistical methods must be approved by the SFD. The results of all statistical analyses shall be presented in tabular form.

8. BIOACCUMULATION.

a. Sediment and Water Collection. The Contractor shall collect and preserve all sediment and water samples as described in sections 3 and 4 above and in the Manual. Compositing sediment samples shall be prepared and handled according to the Manual. For control sediment, the Contractor shall procure unpolluted sediment that is compatible with the test organisms and preferably from where they were collected. The control sediment must meet the needs of the organisms.

The bioassays shall be conducted with a flow-through seawater system except for the test using the amphipod. Seawater of approximately 15°C, 30-32 ppt salinity should be passed through a sand filter and flow into each aquarium at a rate that will replace the aquarium volume at least once every 12 hours. The flow should be directed to achieve good mixing without disturbing the layer of sediment on the aquarium bottom. Water for all bioassays will be clean, uncontaminated seawater of appropriate salinity, pH and temperature. Seawater from any suitable location may be used provided it does not exceed applicable EPA quality criteria for marine waters and is of constant quality.

b. Collection and Maintenance of Test Species.

(1) Species Collection. It is recommended that collection of species should include at least 20% more than the minimum requirement.

1. Table taken from Memorandum dated December 21, 1993, "Technical Panel Recommendations Concerning Use of Acute Amphipod Tests in Evaluation of Dredged Material."

(2) Species Selection. Two species shall be used: (1) Macuma nasuta and (2) Nephtys caecoides

(3) Organism Handling and Holding. Organisms shall be held no longer than two weeks. The SFD must approve additional holding time. Experiments shall be designed and performed so that organisms are handled as minimally as possible. Procedures for handling are found in the Manual. The physiological and biological needs of the organisms must be met at all times.

c. Solid Phase Preparation and Experimental Design. The test treatments shall consist of the dredged material samples, a reference, and a control. Five replications of each treatment shall be performed. Each replicate shall consist of at least 20 organisms of each of these species. The dredged material treatments, references, and control shall be prepared as described in the Manual. However, only whole sediments shall be used in the solid phase tests. Layering of test sediments or control sediments over reference sediments is no longer acceptable. The purpose of the control is to verify the health of test organisms and the acceptability of test conditions. It also provides for quality assurance. If the mean survival in the control is less than 90 percent, the test must be repeated at no additional cost to the SFD. This data must be reported to the SFD. The variable measured shall be percent survival. Each species shall be considered a separate test.

(1) Tissue Analyses. After Corps and EPA review, it was determined that, based on the low levels of contaminants measured in the sediment chemistry, tissue analyses is not necessary and therefore not required in this revised scope of service.

(2) Number of Samples. Five replicates from each of the treatments shall be tested for the parameters listed in Table 3. Survivors within each replicate shall be pooled as necessary to provide sufficient tissue for testing. The treatments shall consist of the dredged material samples, the references, and the control.

a. The results shall be reported in dry weight. Percent moisture shall also be reported.

b. Procedure. Suggested procedures for specific constituents are given in the Manual. The method selected must yield the required detection limits with good precision and accuracy.

c. Solid Phase Testing. Conditions and procedures shall follow those found in the Manual for the 28-day solid phase bioassay. Observations and water quality measurements (temperature, pH salinity, dissolve oxygen shall be made daily.

(1) If the test sediment has an odor of hydrogen sulfide or has elevated ammonia levels, prior to introducing the organisms let

1. Table taken from Memorandum dated December 21, 1993, "Technical Panel Recommendations Concerning Use of Acute Amphipod Tests in Evaluation of Dredged Material."

the sediment settle in tank and then aerate until the ammonia concentration is sufficiently reduced and there is sufficient oxygen (approximately 4ppm) at the sediment-water interface being careful not to oxidize the sediment. One hour after the addition of the organism, the water in the tank shall be analyzed for hydrogen sulfide, ammonia, and dissolved oxygen. This information shall be included in the final report.

d. Data Analysis and Presentation.

NA

9. QUALITY ASSURANCE AND QUALITY CONTROL.

a. The Contractor and subcontractors shall have an established quality control plan which is based on Environmental Protection Agency's quality control program as outlined in Handbook for Analytical Quality Control in Water and Wastewater Laboratories, USEPA 600/4-79-019, March 1979, EPA Office of Research and Development, Cincinnati, Ohio (Handbook). This plan shall also comply with the manual.

b. Quality control charts will be used for precision and accuracy (see section 6.1-6.3 of the Handbook). Percent recovery will be the control chart statistic for controlling accuracy. The industrial statistic "I" will be the control chart statistic for controlling precision. When it is discovered that any analysis is out of control from the standpoint of either precision or accuracy, all analyses since the last in control point will be repeated.

c. Upon completion of the analyses, the laboratory shall prepare a quality control report which includes the precision and accuracy of data generated on the analyzed samples.

d. As an absolute minimum, the following quality control measures shall be taken with each group of samples analyzed:

(1) A reagent blank per batch of samples shall be analyzed.

(2) One duplicate analyses per 10-20 samples shall be made, and precision data shall be reported in the quality control report.

(3) At least one audit or reference sample (EPA, NBS or other EPA- acceptable sources) for each constituent (if available) shall be analyzed (per batch or one per 10-20 samples whichever is less) and reported in the quality control report. This audit sample (marine or

1. Table taken from Memorandum dated December 21, 1993, "Technical Panel Recommendations Concerning Use of Acute Amphipod Tests in Evaluation of Dredged Material."

estuarine sediment and tissue) shall be within the same concentration range as the samples that are being analyzed.

(4) Spiked samples shall be analyzed in order to address analytical accuracy. At least one per 10-20 samples must be spiked with an appropriate standard in order to address accuracy. The concentration of the spike shall be within 200% of the detection limit.

(5) Printouts from all AA and GC analyses shall be kept on file in the event that any concerns arise with the data.

e. All laboratory analyses shall be completed within the recommended holding time for each analytical method.

f. In addition to following quality control procedures described in the Handbook, quality control procedures described for specific analytical methods shall also be followed.

g. All GC analyses require confirmation using a second column which is different from the one used in the initial GC analysis.

h. Standard reference toxicant tests shall be conducted on all species. The results shall be reported in the report.

#### 10. RELEASE OF DATA.

All data, reports, and materials obtained as a result of this contract shall become the property of the U.S. Government and shall be turned over to the SFD upon completion of this work. No data shall be released by the Contractor to any other party other than the SFD without expressed written permission from the SFD.

#### 11. RESPONSIBILITY FOR FIELD WORK.

The Contractor shall be responsible for all damages to persons and property that occur as a result of actions by the Contractor's employees in connection with execution of the work.

#### 12. REPORT PREPARATION.

a. The contractor shall prepare a project report according to the following format.

(1) Introduction. This section shall include a discussion of the purpose and a description of the project.

(2) Materials and Methods. This section shall include:

a. Narrative description of the material, methods and equipment used to perform the project

1. Table taken from Memorandum dated December 21, 1993, "Technical Panel Recommendations Concerning Use of Acute Amphipod Tests in Evaluation of Dredged Material."



tasks.

b. Daily field activity log which includes tidal stage and weather conditions.

c. Inventory of all samples taken and explanation of how used in the tests.

d. Diagrams and figures as appropriate including location map of the sampling areas and sample locations within each area.

(3) Results. The Contractor shall include a narrative of the chemical characterization test results as well as the tables and graphs as described earlier. Any unusual laboratory or field observations shall also be described.

(4) References.

(5) Include appendixes

Appendix A -Scope of Work

Appendix B- Field Sampling Log Sheets/Field Notes

Appendix C- Grain Size data/graphs

Appendix D- QA/QC Data Plan and Report

(6) Text material shall be typed on good quality 8 1/2 by 11 inch bond paper with a 1 1/2-inch margin on the right, and 1-inch at the top and bottom.

(7) Drawings or plates shall be no larger than 20 inches by 11 inches with sufficient margin for binding on the left side and shall include a geographical scale.

(8) Each draft report shall be reviewed by the Corps of Engineers and comments returned to the Contractor. The Contractor shall address comments, correct typographical errors, and otherwise revise the document in accordance to the Contracting Officer's or his Authorized Representative's comments and questions.

#### **Period of Service**

##### **Check Point One:**

Pre-sampling Conference

Within 2 days of receiving the notice to proceed the contractor shall contact the Corps contract representative and provide the proposed dates for sampling.

##### **Check Point Two:**

1. Table taken from Memorandum dated December 21, 1993, "Technical Panel Recommendations Concerning Use of Acute Amphipod Tests in Evaluation of Dredged Material."

Within 15 workdays of receiving the notice to proceed the contractor shall complete the sampling.

**Check Point Three:**

Within 60 workdays following the sampling the contractor shall submit 3 copies of the draft report.

**Check Point Four:**

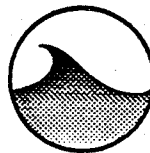
Within 10 workdays of receiving the Corps comments on the draft report, the contractor shall submit 10 copies of the final report.

## Appendix B

### Field Sampling Log Sheets



# HUMBOLDT



**KINNETIC  
LABORATORIES  
INCORPORATED**

## CORE SAMPLING DATA SHEET

Date: 3/31/94 Vessel: Celtic

Captain: Phil Glen Crew: Kranschmidt, Filios, m. Kacher.

GENERAL OBSERVATIONS: N.E. corner of turning basin.  
Medium to fine grain sand - very little silt  
Shell hash & large wood debris - No odor - Grey/Brown  
Take PSD only (Clear & calm)

Location I.D.: SAM-70 Time: 0836

Coordinates: 40° 49' 01.39  
124 10 35.83

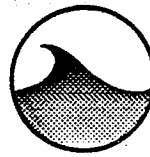
Grab # 1 Grab Core Length Obtained: 8" Core Length Sampled Take entire depth

Sample: Composite/Discrete Discrete Core subsampled: Yes/No No Sub Sample Interval: NA

Water Depth (+/-) Tide	M.L.L.W. Sampling Depth =	<u>37</u>
<u>35.25</u>		
<u>32.8</u> (+/-) <u>-.59</u>	=	M.L.L.W. Mudline Depth = <u><del>34.55</del> 35.95</u>
<u>- (-.7)</u>		
	Core Length =	<u><del>2.45</del> 4.05</u>

COMMENTS: Took a grab to characterize grain size first,  
Based on grain size will not core

HUMBOLDT



KINNETIC  
LABORATORIES  
INCORPORATED

CORE SAMPLING DATA SHEET

Date: 3/31/94 Vessel: Celtic

Captain: Phil Glen Crew: K.K., W.F., M.E.

GENERAL OBSERVATIONS: Calm + Clear  
S.E. side of turning basin - Just west (north) of channel side  
Medium to fine grain sand w/ shell hash - Very little silt  
no odor - Grey/brown - 1 large shell  
Very little current - @ peak low

Location I.D.: SAM-5D Time: 0902

Coordinates: 40° 49' 01.39

129 10 35.83

Grab 1 8" penetration

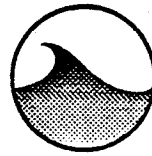
Core#: \_\_\_\_\_ Core Length Obtained: NA Core Length Sampled NA

Sample: Composite/Discrete Core subsampled: Yes/No Sub Sample Interval: NA

Water Depth (+/-) Tide	=	M.L.L.W. Sampling Depth	=	<u>37</u>
<u>33.75</u>				
<u>35.25</u> (+/-) <u>(-.79)</u>	=	M.L.L.W. Mudline Depth	=	<del>32.91</del> <u>34.54</u>
		Core Length	=	<u>NA</u> <u>2.46</u>

COMMENTS: Took Grab First to characteriz PSD  
will not do core based on grain size.  
Took PSD only  
last position from last years Job was on side of channel

HUMBOLDT



KINNETIC  
LABORATORIES  
INCORPORATED

CORE SAMPLING DATA SHEET

Date: 3/31/94 Vessel: Celtic

Captain: P. Glen Crew: KK, W.F., M.K

GENERAL OBSERVATIONS: West edge of channel near marker  
Calm & Clear -  
Med. to fine grain sand w/ small shell hash - very little silt.  
Brownish green in color - Found Fine clay like silt & sand  
below 5 cm - No odor

Location I.D.: SAM-40 Time: 0930

Coordinates: 40° 48' 33.58"  
124° 10' 55.71"

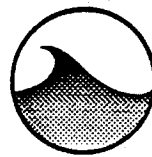
Core #: Grab Core Length Obtained: 6" penetration Core Length Sampled: NA

Sample: Composite/Discrete Discrete Core subsampled: Yes/No NA Sub Sample Interval: NA  
will not keep PSD only

Water Depth (+/-) Tide		M.L.L.W. Sampling Depth =	<u>37</u>
<u>34.75</u> (+/-) <u>(-.70)</u>	=	M.L.L.W. Mudline Depth =	<u><del>34.05</del> 35.45</u>
		Core Length =	<u><del>3.05</del> 1.56</u>

COMMENTS: Took Grab First to characterize PSD. Decided  
to include this site w/ the composite will  
come back to core

HUMBOLDT



KINNETIC  
LABORATORIES  
INCORPORATED

CORE SAMPLING DATA SHEET

Date: 3/31/94 Vessel: Celtic

Captain: B. Glen Crew: KK, WIF, M.K

GENERAL OBSERVATIONS: Slight breeze (clear)  
Grabbed on the Eastern side of channel. Went land of  
site from last years plot on the chart - LAT LARVAS Did not  
match -  
Med. to fine grain sand w/ shell hash - very little silt. no odor  
grey/brown - no organic debris

Location I.D.: SAM-3 Time: 0955 (#1 attempt)  
1003 (#2 attempt)

Coordinates: 40° 48' 23.29 } #2  
124° 11' 00.23 }

Grab #2  
 Core#: \_\_\_\_\_ Core Length Obtained: NA Core Length Sampled NA

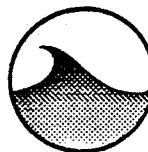
Sample: Composite/Discrete Discrete Core subsampled: Yes/No Sub Sample Interval: NA

Water Depth (+/-) Tide M.L.L.W. Sampling Depth = 32  
32.75 (+/-) (-.32) = M.L.L.W. Mudline Depth = ~~32.43~~ 33.07  
 Core Length = 3.23

COMMENTS: no Take Grab first to characterize PSD  
Made two attempts - First attempt we drifted out of  
the channel.  
Will use for PSD characterization only



HUMBOLDT



KINNETIC  
LABORATORIES  
INCORPORATED

CORE SAMPLING DATA SHEET

Date: 3/31/94 Vessel: Celtic

Captain: P. Glen Crew: KK, W.F, M.K

GENERAL OBSERVATIONS: Clear + slight breeze  
Near center of channel between marker 2 & the Loading Dock  
to the S.W of marker #2  
Med. grain sand w/ shell hash - very little silt. - NO odor or no  
organic debris

Location I.D.: SAM-2 Time: 1027

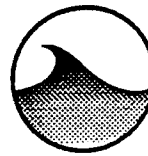
Coordinates: 40° 48' 07.45" N  
124° 11' 08.68" W

~~Grab~~ Grab #1  
 Core #: \_\_\_\_\_ Core Length Obtained: 6" penetration Core Length Sampled NA  
 Sample: Composite (Discrete) Core subsampled: Yes/No Sub Sample Interval: NA.

Water Depth (+/-) Tide M.L.L.W. Sampling Depth = 37  
36.05 (+/-) (+.05) = M.L.L.W. Mudline Depth = 36.1  
 Core Length = \_\_\_\_\_

COMMENTS: Took Grab sample first to characterize PSD  
Will not use for compositing. PSD only

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CORE SAMPLING DATA SHEET

Date: 3/31/94 Vessel: Celtic

Captain: P. Glen Crew: Kronshardt, M. K, W.F

GENERAL OBSERVATIONS: wind 10 knts (clear) - off wood chip piles  
very southern extent of some channel  
Medium to coarse sand w/ shell hash. very little silt. - no smell  
Brown in color  
Went to last times coordinates

Location I.D.: SAM-10 Time: 1056

Coordinates: 40° 47' 57.67"  
124° 11' 14.37"

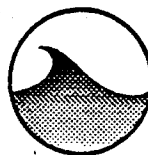
Grab #1 5" penetration  
 Core#: \_\_\_\_\_ Core Length Obtained: NA Core Length Sampled NA

Sample: Composite Discrete Core subsampled: Yes/No Sub Sample Interval: NA

Water Depth (+/-) Tide = M.L.L.W. Sampling Depth = 37  
35.25 (+/-) (+.64) = M.L.L.W. Mudline Depth = ~~35.89~~ 34.61  
 Core Length = 2.39

COMMENTS: Took 2 Grab first to characterize PSD  
PSD only

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CORE SAMPLING DATA SHEET

Date: 3/31/94 Vessel: Celtic  
 Captain: P. Glen Crew: KL, M.R., W.F.

GENERAL OBSERVATIONS: Wind 10-15 kts (Clear) - High current speed (Flood)  
 Dark Fine sand w/ silt - little shell hash + little organic debris - no oden  
 hard clay/sand below 2'

(Core #1 - Pen. 3.2' kept 2.1' - discarded bot. 1.1') (Core #4 - Penetrated 4' - recovered 2.0' - kept 1.8')  
 (Core #2 - recovered 2.3' - kept 2.0') (Core #5 - Pen. 4.5' - recovered 3.3' - kept 2.1')  
 (Core #3 - Penetrated 3.5' - recovered 2.0') (Core #6 - Pen. 4.0' - recovered 2.8' - kept 2.1')  
 (Core #7 - Pen 4.5' - recovered 3.9' - kept 2.1')  
 (Core #8 - Pen 4.5' - " 3.0 - kept 2.1')

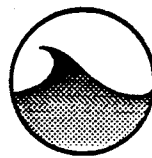
Location I.D.: SAM-6C Time: #1 (1210) #2 (1305) #3 (1323) #4 (1347)  
#5 (1404) #6 (1423) #7 (1443) #8 (1509)  
 Coordinates: 40° 49' 05.18"  
124° 10' 38.62"

Core #: \_\_\_\_\_ Core Length Obtained: 3.2, 2.3, 3.5, 2.0, 3.3, 2.6, 3.9, 3.0  
2.1, 2.0, 2.0, 1.8 Core Length Sampled 2.1, 2.0, 2.0, 1.8, 2.1  
2.1, 2.1, 2.1  
 Sample: Composite Discrete Core subsampled: Yes  No  Sub Sample Interval: 1/4

Water Depth (+/-) Tide M.L.L.W. Sampling Depth = 37  
36.95 (+/-) - (+ 2.07) = M.L.L.W. Mudline Depth = 34.88  
 ↑  
~ 1200hrs Core Length = 2.12

COMMENTS: dropped buoy on last times coordinates.  
Took 8 cores ~~2.1~~' each + composited together - to fill 8 gal's  
made 1 chem sample for the 8 cores + the rest went  
towards the SAM composite  
generated sample Humsed0007  
1 spoon went towards chemistry composite for each core

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CORE SAMPLING DATA SHEET

Date: 3/31/94 Vessel: Celtic
Captain: P. Glen Crew: K.K., W.F., M.K.

GENERAL OBSERVATIONS: ~15 kn winds - clear - current beginning to ebb
Very fine sand & silty clay throughout the 5' - no horizons - slight H2S smell - dark grey
not much in the way of large organic matter - no shell hash
(Core #1 pen - 4.5' 4.1' recovery - kept 3.1') (Core #2 4.2' pen - 3.66' recovery - kept 3.1')
(Core #3 pen - 4.5' 3.75' recov. kept 3.1') (#4 4.1' pen - 3.75' recov - kept 3.1')
(#5 - 4.5' pen - 3.66' recov. - kept 3.1') (#6 - 4.5' pen - 3.5' recov - kept 3.1')
Very strong current by the end of sampling.

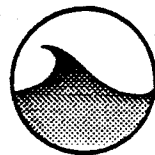
Location I.D.: SAM-6B Time: #1(1634) #2(1655) #3(1714) #4(1745)
Coordinates: 40° 49' 02.35" N
124° 10' 42.04" W

Core#: Core Length Obtained: 4.1, 3.66, 3.75, 3.75 Core Length Sampled 3.1, 3.1, 3.1, 3.1, 3.1
Sample: Composite/Discrete Core subsampled: Yes/No Sub Sample Interval: NA

Water Depth (+/-) Tide M.L.L.W. Sampling Depth = 37
39.75 (+/-) 5.86 = M.L.L.W. Mudline Depth = 33.89
-(+5.86) Core Length = 3.1'
6 cores

COMMENTS: changed position slightly from last time to slightly shallower.
Water (~100' to the N of old position - same distance offshore ~200' off Pacific pier). 1.5 spans towards chemistry composite for each core
Generated sample HUNAYSEDO008 From the 6 cores - rest of mud went to SAM compos.

## HUMBOLDT


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## CORE SAMPLING DATA SHEET

Date: 4/1/94 Vessel: CelticCaptain: P. Glen Crew: KK, M.K, W.F.GENERAL OBSERVATIONS: Fast current (Ebb) - Cloudy - CalmLocation I.D.: SAM-6A Time: 0855Coordinates: 40° 48' 57.29"124° 10' 46.13"

Core#: \_\_\_\_\_ Core Length Obtained: \_\_\_\_\_ Core Length Sampled \_\_\_\_\_

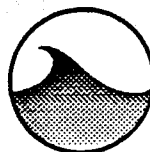
Sample: Composite/Discrete Core subsampled: Yes/No Sub Sample Interval: \_\_\_\_\_

Water Depth (+/-) Tide

36.75 (+/-) - (-.05)M.L.L.W. Sampling Depth = 37'0M.L.L.W. Mudline Depth = 36.8Core Length = .2

COMMENTS: Station abandoned due to lack of dredgable material - will move to find alternate site. The site sampled last time also is too deep. Suspect prop washed out area.

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CORE SAMPLING DATA SHEET

Date: 4/1/94 Vessel: Celtic

Captain: R. Glen Crew: KK, W.F, M. K

GENERAL OBSERVATIONS: high speed ebb current. searched around for shallower sediments & came up w/ this location (66m from previous location) Fine sand on top w/ silt turning ~~more~~ silt/clay like below 5cm Brown/grey - darker below - no odor. some organics. some shell hash  
~~The tide~~

Location ID.: Stm-6A (alt) Time: Grab: 0942 <sup>not used</sup> Grab #1: 0954

Grab coordinates: START	$\left\{ \begin{array}{l} 40^{\circ} 48' 56.61'' \\ 124^{\circ} 10' 44.18'' \end{array} \right.$	Finish	$\left\{ \begin{array}{l} 40^{\circ} 48' 56.82'' \\ 124^{\circ} 10' 44.05'' \end{array} \right.$	#1: <u>0954</u>
				#2: <u>1006</u>
				#3: <u>1009</u>
				#4: <u>1039</u>
				#5: <u>1049</u>

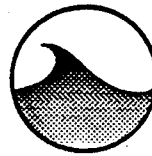
Grab penetration: #1(5") #2(5") #3(6") #4(5") #5(6")  
 Core#: \_\_\_\_\_ Core Length Obtained: \_\_\_\_\_ Core Length Sampled: NA

Sample: Composite/Discrete Core subsampled: Yes/No Sub Sample Interval: NA

Water Depth (+/-) Tide	M.L.L.W. Sampling Depth =	<u>37.0</u>
<u>35.25</u> (+/-) <u>-(-.43)</u>	M.L.L.W. Mudline Depth =	<u>35.68</u>
	Core Length =	<u>1.32</u>

COMMENTS: Took Grab first to characterize sediment (PSD)  
 Took 6 grabs for composite sam - Generated sample Huma4sed0009 for discrete sediment chemistry  
 - Took sediment only from center & away from edges of grab.  
 - The tide changed direction in the middle of sampling moving us slightly off position

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CORE SAMPLING DATA SHEET

Date: 4-1-94 Vessel: Celtic

Captain: P. Glen Crew: KK, M.K, W.F.

GENERAL OBSERVATIONS: Overcast + CALM - strong Flood current

Core #1 - 40' pen, 2.33' recovery, kept 2.2'  
 Medium to coarse grain sand & shell hash. There was a thin layer of silty clay a few inches down from the surface. Overall very little silt  
No odor

Location I.D.: SAM-4(D) Time: #1(1217)

Coordinates: 44° 48' 33.58"  
124° 10' 55.71"

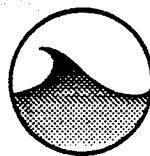
Core #: \_\_\_\_\_ Core Length Obtained: 2.33' Core Length Sampled 2.2'

Sample: Composite/Discrete Discrete Core subsampled: Yes/No Sub Sample Interval: \_\_\_\_\_

Water Depth (+/-) Tide	M.L.L.W. Sampling Depth =	<u>37.0</u>
<u>35.25</u>		
<u>33.5</u> (+/-) - <u>(.91)</u>	M.L.L.W. Mudline Depth =	<u>34.34</u>
	Core Length =	<u>2.66</u>

COMMENTS: Decided to scrap this site as part of the composite. We were fooled by the thin layer of silty clay that was found in yesterdays grab.

## HUMBOLDT


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## CORE SAMPLING DATA SHEET

Date: 4/1/94 Vessel: CelticCaptain: P. Glen Crew: KK, W.F.M.K

GENERAL OBSERVATIONS: Overcast + calm (strong flood current)  
Sand (med to coarse), gravel + large shell hash, very little silt - no odor

Location ID.: NB-10 Time: 1333
 Ordinate: 40° 49' 45.6" N  
124° 11' 15.28" W

 Grab  
 Core #: 1 Core Length Obtained: 3" penetration Core Length Sampled: NA

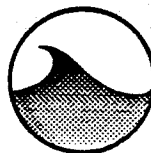
 Sample: Composite/Discrete Discrete Core subsampled: Yes/No No Sub Sample Interval: NA

Water Depth (+/-) Tide

36.6 (+/-) -(+2.86) =M.L.L.W. Sampling Depth = 37M.L.L.W. Mudline Depth = 35.49Core Length = 1.51COMMENTS: Took preliminary grab to characterize PSD
Generated sample HUM94SE0012 to be analyzed for PSD only



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CORE SAMPLING DATA SHEET

Date: 4/1/94 Vessel: Celtic

Captain: P. Glen Crew: KK, M.E, W.F.

GENERAL OBSERVATIONS: Overcast calm flood current.  
Med to coarse grain sand w/ small shell hash - very little silt.  
NO odor - A few large shells - no organic debris. Brownish  
color

Location I.D.: NB-9 Time: 1352

Coordinates: 40° 47' 45.60"  
124° 18' 15.28"

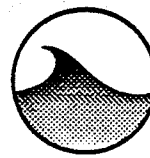
Grab Core#: Grab Core Length Obtained: 8" penetration Core Length Sampled NA

Sample: Composite Discrete Core subsampled: Yes No Sub Sample Interval: NA

Water Depth (+/-) Tide	M.L.L.W. Sampling Depth =	<u>32</u>
<del>37.0</del> (+/-) - <u>(+3.37)</u>	M.L.L.W. Mudline Depth =	<u>35.38</u>
<u>38.75</u>	Core Length =	<u>1.62</u>

COMMENTS: Took preliminary grab to characterize PSD  
Generated sample HUM945200014 for PSD only

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CORE SAMPLING DATA SHEET

Date: 4/1/94 Vessel: Celtic

Captain: P. Glen Crew: KK, M.K., LWF

GENERAL OBSERVATIONS: partly cloudy - calm - strong flood current.  
starfish in sample  
medium to coarse sand - very little silt - some large & small shell hash  
No odor - no organic debris - some small gravel

Location I.D.: B-8 Time: 1410

Coordinates: 40° 47' 07.79"  
124° 11' 36.94"

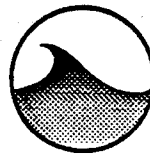
Grab Core#: Grab Core Length Obtained: Grab penetrated 6" Core Length Sampled: NA

Sample: Composite/Discrete Discrete Core subsampled: Yes/No No Sub Sample Interval: NA

Water Depth (+/-) Tide		M.L.L.W. Sampling Depth =	<u>37</u>
<u>40.25</u> (+/-) - <u>(+3.7)</u>	=	M.L.L.W. Mudline Depth =	<u>36.55</u>
		Core Length =	<u>.45</u>

COMMENTS: Took preliminary Grab for PSD characterization  
Moved site from last years position because the  
MLLW depth was 49.05'  
Generated sample HUM94SED0014 For PSD only

## HUMBOLDT


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## CORE SAMPLING DATA SHEET

Date: 4/1/94 Vessel: CelticCaptain: P. Glen Crew: KK, W.F., M.K

GENERAL OBSERVATIONS: partly cloudy & calm  
Went to last times position - very edge of channel - might have  
been slightly to the west of the channel  
Fine to medium sand w/ small gravel & small shell hash - very little silt.  
Gray sand - looks clean

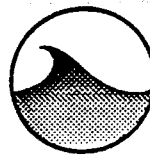
Location I.D.: NB-7 Time: 1431Coordinates: 40° 46' 58.03"124° 11' 45.41"

Grab  
 Core#: \_\_\_\_\_ Core Length Obtained: NA Core Length Sampled NA

Sample: Composite/Discrete Core subsampled: Yes/No Sub Sample Interval: \_\_\_\_\_Water Depth (+/-) Tide M.L.L.W. Sampling Depth = 3738.6 (+/-) - (4.16) = M.L.L.W. Mudline Depth = 34.4Core Length = 2.6

COMMENTS: Took preliminary grab for PSD only characterization  
Generated sample HUM95ed0015 for PSD only

## HUMBOLDT



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## CORE SAMPLING DATA SHEET

Date: 4-1-94 Vessel: Celtic  
 Captain: P. Glen Crew: KK, MK, W.F

GENERAL OBSERVATIONS: mostly cloudy & calm - strong flow current.  
went to site preselected by COE. position given by R. Mattison  
Mouth of Elk River.  
Medium grain sand - large shell hash - brown algae - no odor - gray in color  
small pebbles

Location I.D.: NB-6 Time: 1502

Coordinates: 40° 46' 39.03"  
124° 11' 52.39"

Grab Core#: \_\_\_\_\_ Core Length Obtained: Grab penetration ~ 5" Core Length Sampled: NA

Sample: Composite/Discrete Discrete Core subsampled: Yes  No  Sub Sample Interval: NA

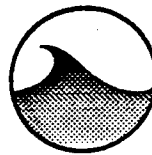
Water Depth (+/-) Tide M.L.L.W. Sampling Depth = 37

40.4 (+/-) - (+4.73) = M.L.L.W. Mudline Depth = 35.72

Core Length = 1.25

COMMENTS: Took preliminary grab to characterize PSD  
Generated sample Hums45E00016 for PSD only

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CORE SAMPLING DATA SHEET

Date: 4/1/94 Vessel: Celtic

Captain: P. Glen Crew: KL, W, F, M, K

GENERAL OBSERVATIONS: mostly cloudy - slight breeze - incoming tide.  
Medium grain sand w/ very little shell has - small globs of fine  
mud - NO organics -

1' swell

Location I.D.: NB-5 Time: 1523

Coordinates: 40° 46' 18.43"  
124° 12' 21.38"

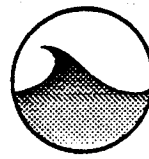
Grab Core#:                      Grab penetration: 10" penetration  
 Core Length Obtained:                      Core Length Sampled NA

Sample: Composite  Discrete  Core subsampled: Yes  No  Sub Sample Interval: NA

Water Depth (+/-) Tide		M.L.L.W. Sampling Depth =	<u>37</u>
<u>42.1</u>	(+/-) - <u>(+5.05)</u>	=	M.L.L.W. Mudline Depth = <u>37.05</u>
		Core Length	= <u>-05</u>

COMMENTS: At dredge depth  
Took Grab to characterize PSD  
Generated sample HUM95E0017 for PSD only

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CORE SAMPLING DATA SHEET

Date: 4/1/94 Vessel: Celtic  
 Captain: P. Glen Crew: KK, W.F, M.K

GENERAL OBSERVATIONS: mostly cloudy - 5 knot wind - 1-2' swell  
Fine sand & silty mud. grabed in a very small shallower area  
some shell - Fine enough for chem sampling. - grey in color.  
No odor

Location I.D.: NB-4 Time: ~1555  
 Coordinates: 40° 46' 07.54"  
124° 12' 44.27

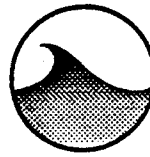
Grab Grab penetration: 7"  
 Core#: \_\_\_\_\_ Core Length Obtained: NA Core Length Sampled NA

Sample: Composite/Discrete Discrete Core subsampled: Yes/No No Sub Sample Interval: M

Water Depth (+/-) Tide	M.L.L.W. Sampling Depth =	<u>37.0</u>
<u>41.8</u>		
<del>40.1</del> (+/-) - <u>(+5.22)</u>	M.L.L.W. Mudline Depth =	<u>36.58</u>
	Core Length =	<u>.42</u>

COMMENTS: moved slightly away from last times position into shallower water  
took grab to characterize PSD  
Generated sample # HUM94SE00018 for PSD ← NO GOOD  
may have to redo for chemistry redo  
discard

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CORE SAMPLING DATA SHEET

Date: 4-1-94 Vessel: Celtic

Captain: Phil Glen Crew: KK, M.K, W.I

GENERAL OBSERVATIONS: *Mostly sunny w/ slight breeze. w/ swell  
- Makes fathometer readings difficult - went to position  
given by R. Mattison from Kerry Guy*

*Medium to coarse sand w/ shell hash - gray in color - very little silt.  
no organics - no smell*

Location I.D.: NB-3 Time: 1620

Coordinates: 40° 45' 46.95"  
124° 12' 56.80"

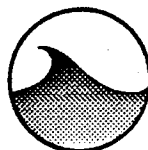
Grab Grab Pen: ~6"  
Core#: \_\_\_\_\_ Core Length Obtained: NA Core Length Sampled NA

Sample: Composite/Discrete Discrete Core subsampled: Yes/No No Sub Sample Interval: NA

Water Depth (+/-) Tide	M.L.L.W. Sampling Depth =	<u>37.0</u>
<u>40.05</u> (+/-) - <u>(+5.61)</u>	M.L.L.W. Mudline Depth =	<u>34.44</u>
	Core Length =	<u>2.56</u>

COMMENTS: *Grabbed to characterize grain size  
Generated sample Hum945ED0019 for PSD only*

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## CORE SAMPLING DATA SHEET

 Date: 4/1/94 Vessel: Celtic

 Captain: Phil Glen Crew: KL, MR, W.F.

 GENERAL OBSERVATIONS: Mostly clear slight breeze - 2-3' swells -  
very large surf in entrance  
clean fine to medium sand w/ small shell hash - no odor -  
no pieces of wood in sample

 Location I.D.: KL NB-~~2~~1 Time: 1647

 Coordinates: 40° 45' 10.9"  
124° 13' 25.13"

 Core#: Grab Core Length Obtained: NA Core Length Sampled: NA

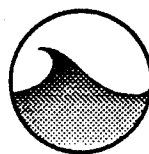
 Sample: Composite/Discrete Core subsampled: Yes/No Sub Sample Interval: NA

Water Depth (+/-) Tide	M.L.L.W. Sampling Depth =	<u>37.0</u>
<u>41.7</u> (+/-) - <u>(+5.7)</u> =	M.L.L.W. Mudline Depth =	<u>36.0</u>
	Core Length =	<u>1.0</u>

 COMMENTS: Took Grab to characterize grain size
Generated sample HUM94SED0020 for PSD only



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CORE SAMPLING DATA SHEET

Date: 4/1/94 Vessel: Celtic

Captain: Phil Glen Crew: KK, M.K, W.F.

GENERAL OBSERVATIONS: Going back to site for 2<sup>nd</sup> time to get a grab for a chemistry sample.  
 Fine sand - very little silt - very confusing - we were within a few meters of the last grab but this time there was no fine mud - there was a crab in the grab. no odor  
 Cannot use NB-4 rep 1 for chemistry because the sample was contaminated (handled improperly)

Location I.D.: NB-4 rep 2 Time: 1723

Coordinates: 40° 46' 07.54"  
124° 12' 44.27"

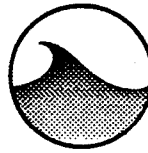
Grabbed Grab penetration: ~ 2"  
 Core#: \_\_\_\_\_ Core Length Obtained: NA Core Length Sampled NA

Sample: Composite/Discrete Discrete Core subsampled: Yes/No Sub Sample Interval: NA

Water Depth (+/-) Tide M.L.L.W. Sampling Depth = 37  
42.25 (+/-) -(+5.7) = M.L.L.W. Mudline Depth = 36.55  
 Core Length = ~.45

COMMENTS: Took Grab for Sediment Chemistry. However, due to grain size it will only be a PSD sample  
 \*see NB-4 rep 1  
 Generated sample HUMB94SEDOO21 for PSD only

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CORE SAMPLING DATA SHEET

Date: 4/2/94 Vessel: Celtic  
Captain: Phil Glenk Crew: Kk

GENERAL OBSERVATIONS: Not far from last years position. First it didn't look right - moved it more towards center of channel - medium to large grain sand with little silt. Large shell has some pebbles & wood chips - 1 stone - dark grey - no odor.

Location I.D.: EK-1 0835  
Coordinates: 40° 47' 55.79"  
124° 11' 13.99"

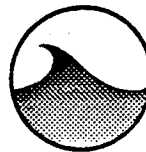
Core#: \_\_\_\_\_ Penetration: \_\_\_\_\_  
Core Length Obtained: NA Core Length Sampled: NA  
Sample: Composite/Discrete Core subsampled: Yes/No Sub Sample Interval: NA

Water Depth (+/-) Tide = M.L.L.W. Sampling Depth = 37  
37.8 (+/-) -(+2.1) = M.L.L.W. Mudline Depth = 35.7  
Core Length = 1.3

COMMENTS: Took Grab to characterize grain size.  
Generated Sample Hum 94sed0022 for PSD only.

*NOT USED - WRONG COORDINATES*

HUMBOLDT



KINNETIC  
LABORATORIES  
INCORPORATED

CORE SAMPLING DATA SHEET

Date: 4/2/94 Vessel: Celtic

Captain: P. Glen Crew: KK, MK, W.F

GENERAL OBSERVATIONS: Overcast - slight breeze. Strong Ebb current. Fine sand, silt & clay - very little shell hash - slight H<sub>2</sub>S smell  
coarser sand on top 6"

(Core #1 - 4.5' pen, 3.5' recovery, kept 3.3) (Core #4 - 4.5' pen, 3.9' recov., kept 3.3)  
(Core #2 - 4.0' pen, 2.6' recov., kept all) (Core #5 - 5' pen, 2.2' recovery, kept all)  
(Core #3 - 4.5' pen, 3.5' recovery, kept 3.3)

Location I.D.: EK-2 Time: #1(0925) #2(0958) #3(1031) #4(1105)

Coordinates: 40° 48' 13.36" NO GOOD - #5(136) #5(11:55) #5(1209)  
124° 10' 48.49" NO GOOD

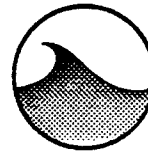
Core #: \_\_\_\_\_ Core Length Obtained: 3.5, 2.6, 3.9, 3.9 Core Length Sampled 3.9, 2.6, 3.3, 3.3

Sample: Composite/Discrete Core subsampled: Yes No Sub Sample Interval: NA

Water Depth (+/-) Tide M.L.L.W. Sampling Depth = 28.0  
25.65 (+/-) - (+.96) = M.L.L.W. Mudline Depth = 24.7  
Core Length = 3.3

COMMENTS: Did prelim grab for grain size then went into coring  
Generated sample HUM94SE00023 for EK-2 discrete chemistry analysis  
Had problems at this site w/ tubes breaking + sediment washing out.

HUMBOLDT



KINNETIC  
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CORE SAMPLING DATA SHEET

Date: 4/2/94 Vessel: Celtic

Captain: R Glen Crew: LK, M.R, W.C

GENERAL OBSERVATIONS:

*Fine to medium sand w/ small shell hash, brownish to grey, no odor - Small amount of silt. Estimated that fine fraction was less than 80%*

Location I.D.: EK1 (redo) Time: 1348

Coordinates: 40° 47' 57.08" N  
124° 11' 05.26" W

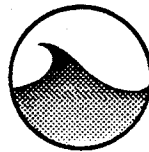
Core#: GRAB Core Length Obtained: NA Core Length Sampled: NA

Sample: Composite/ Discrete Core subsampled: Yes/No Sub Sample Interval: \_\_\_\_\_

Water Depth (+/-) Tide	M.L.L.W. Sampling Depth =	<u>37</u>
<u>37.75</u> (+/-) <u>-(+1.94)</u>	M.L.L.W. Mudline Depth =	<u>35.81</u>
	Core Length =	<u>1.19</u>

COMMENTS: *Re grabed for PSD characterization. Changed location per memo from COE*

HUMBOLDT



KINNETIC  
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CORE SAMPLING DATA SHEET

Date: 4/2/94 Vessel: Celtic

Captain: Phil Glen Crew: KK, M.K., W.F.

GENERAL OBSERVATIONS: Partly foggy & breezy  
Soft clay + silt w/ fine sand throughout - very soft on top 6" - dark grey - slight H<sub>2</sub>S odor  
occasional small shell hash.

(Core #1 - penetrated 6.5' where we met rejection, recovered 4.75' kept it all)  
 (Core #2 - penetrated 7.0 ft - recovered 5.66' - kept it all)  
 (Core #3 - penetrated 7.5 ft - recovered 6.5' kept 6.1')  
 (Core #4 - penetrated 7.5 ft - recovered 6.0' kept all but bottom 2")  
 (Core #5 - penetrated 7.5 ft - recovered 7.0' kept 6.1')

Location I.D.: EK-3 Time: #1(1448) #2(1506) #3(1522) #4(1538)  
#5(1550)

Coordinates: 40° 48' 17.16"  
124° 10' 39.34"

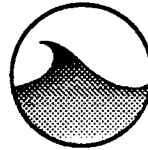
Core #: \_\_\_\_\_ Core Length Obtained: 4.75, 5.66, 6.5, 6.0, Core Length Sampled 4.75, 5.66, 6.1, 6.0  
7.0 6.1 5.8

Sample: Composite/Discrete Core subsampled: Yes/No Sub Sample Interval: NA

Water Depth (+/-) Tide M.L.L.W. Sampling Depth = 28  
24.95 (+/-) - 43.1 = M.L.L.W. Mudline Depth = 21.85  
 Core Length = 6.15

COMMENTS: went to last times position, appeared to be too close to  
channel edge so we moved 200m closer  
Did total of 5 cores - Generated sample # Hums4SE00024 for digester chemistry

HUMBOLDT



KINNETIC  
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CORE SAMPLING DATA SHEET

Date: 4/2/94 Vessel: Celtic  
 Captain: Phil Glen Crew: VK, MK, WF

GENERAL OBSERVATIONS: mostly cloudy & breezy - near slack tide  
soft silty clay - very little sand. No Organics - very very little  
shell hash. Slight H<sub>2</sub>S smell

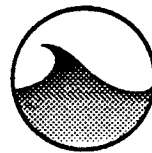
Location I.D.: EK-4 Time: #1 (1646) #2 (1656) #3 (1703) #4 (1711)  
#5 (1718)  
 Coordinates: 40° 48' 20.29" N  
124° 10' 19.61" W

Core#: Crabs Core Length Obtained: NA Core Length Sampled: NA  
 Sample: Composite/Discrete Core subsampled: Yes/No No Sub Sample Interval: NA

Water Depth (+/-)	Tide	M.L.L.W. Sampling Depth	=	<u>28.0</u>
<u>32.85</u>	(+/-) <u>-5.04</u>	M.L.L.W. Mudline Depth	=	<u>27.17</u>
		Core Length	=	<u>.83</u>

COMMENTS: Not enough muds for coring so we took 39 grabs to composite  
w/ the EK sediments - removed ~ 1/2 gal from each grab to allow  
equal weighing  
Generated sample # Humber sed0025 as a discrete - rest of mud  
went towards composite

HUMBOLDT



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INCORPORATED

CORE SAMPLING DATA SHEET

Date: 4-3-94 Vessel: Celtic

Captain: Phil Glen Crew: KK, W.F.M.K

GENERAL OBSERVATIONS: Clear - slight breeze - slack tide - starting to ebb  
Soft mud w/ fine sand turning to hard pack sand below 2' - grey - no shells or  
Core #1 - 4' pen. - 2.0' recovery - kept all) (Core #5 - 4' pen 2.5' recovery kept 2.1) <sup>no organics</sup>  
(Core #2 - 4' pen. 3.1 recovery - kept 2.1) (Core #6 - 4' pen 2.68' recovery kept 2.1) <sup>no 300</sup>  
(Core #3 4' pen 3.0 recovery - kept 2.1) (Core #7 - 4' pen 2.75' recovery kept 2.1) <sup>clam</sup>  
(Core #4 4' pen 2.2' recovery - kept 2.1) (Core #8 - 4' pen 2.0' recovery kept all)

Location I.D.: FL-4 Time: #1(0923) #2(0945) #3(0957) #4(1013)

Coordinates: 40° 43' 35.27" N #5(1022) #6(1032) #7(1040) #8(1052)  
124° 13' 19.57" W

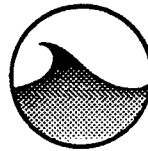
Core #: \_\_\_\_\_ Core Length Obtained: NA Core Length Sampled: NA

Sample: Composite/Discrete Core subsampled: Yes/No Sub Sample Interval: NA

Water Depth (+/-) Tide M.L.L.W. Sampling Depth = 28  
31.5 (+/-) -t 5.6 = M.L.L.W. Mudline Depth = 25.9  
Core Length = 2.1

COMMENTS: Took 8 cores for sediment Composite  
The vibrocore stopped vibrating after the 3rd core. However,  
we were able to penetrate by dropping it  
Generated sample # HUM 94 SED0033 for discrete chemistry

HUMBOLDT



KINNETIC  
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CORE SAMPLING DATA SHEET

Date: 4/3/94 Vessel: Celtic  
 Captain: Phil Glen Crew: VK, M.K, W.F

GENERAL OBSERVATIONS: Mostly clear - westerly winds 20-25 knots - cool  
FL-8: Medium coarse sand - no shell hash - clean looking - very little silt  
no odor  
FL-7: Same as FL-8 except more shell hash + little more silt.

Location I.D.: FL-8 + FL7 Time: FL8: 1045 FL7: 1051  
 Coordinates: 40° 52' 44" 53.02 N 40° 44' 32.88 N  
FL-8 { 124° 13' 16.43 W FL-7 { 124° 13' 30.36 W

Grab  
 Core#: \_\_\_\_\_ Core Length Obtained: N Core Length Sampled: NA  
 Sample: Composite Discrete Core subsampled: Yes/No Sub Sample Interval: NA

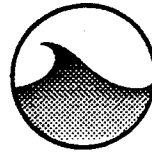
FL-7 times are given in the ( )

Water Depth (+/-) Tide		M.L.L.W. Sampling Depth =	<u>28</u>
<u>28.25</u> (+/-) - <u>+3.1</u>	=	M.L.L.W. Mudline Depth =	<u>25.15</u>
<u>(31.3)</u> - <u>(+2.9)</u>			<u>(28.4)</u>
		Core Length =	<u>2.85</u>
			<u>(-.4)</u>

COMMENTS: Took initial grab to characterize PSD at both stations  
Very hard to stay on position - went coordinates given by <sup>last time</sup> COE for  
Generated Sample # HUMB0027 for PSD only from FL8  
Generated Sample # HUMB0028 for PSD only from FL-7



HUMBOLDT



KINNETIC  
LABORATORIES  
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CORE SAMPLING DATA SHEET

Date: 4/3/94 Vessel: Celtic

Captain: Phil Glen Crew: KK, A.K, W.F.

GENERAL OBSERVATIONS: Mostly Clear w/ skt wind sat of West  
Some silt (Silt fraction less than 80%)

- FL-6: Medium to coarse sand w/ some shell hash - There was a thin layer of mud
- FL-5: Medium to fine grain sand w/ very little shell hash & wood debris. Brown to grey in color - estimate less than 80% fines - no odor

Location I.D.: FL-6 & FL-5 Time: FL-6: ~~1105~~ 1105 FL-5: 1127

Coordinates: FL-6 { 40° 44' 13.4" 124° 13' 14.42" } FL-5 { 40° 43' 59.42" 124° 13' 09.77" }

Grab Core Length Obtained: NA Core Length Sampled: NA

Sample: Composite/Discrete Core subsampled: Yes/No Sub Sample Interval: NA

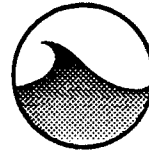
FL-5 times in ( )

Water Depth (+/-)	Tide	M.L.L.W. Sampling Depth =	<u>28</u>
<u>32.7</u>	<u>(+/-) - (+2.52)</u>	M.L.L.W. Mudline Depth =	<u>30.2</u>
<u>(28.4)</u>	<u>(-2.14)</u>	Core Length =	<u>-1.8</u>
			<u>(1.7)</u>

COMMENTS: Took initial grab to characterize PSD at both sites  
Very hard to hold position due to wind & current  
We were in too deep of water @ FL-6 - however the shallower spots appeared to be out of channel

Generated sample number 0039 for site FL-6 (PSD only)  
Generated sample number 0030 for site FL-5 (PSD only)

HUMBOLDT



CORE SAMPLING DATA SHEET

Date: 4/3/94 Vessel: Celtic

Captain: Phil Glen Crew: KK, MK, WJ

GENERAL OBSERVATIONS: Winds out of West @ 25+ knts - strong ebb current  
Soft mud w/ fine sand. Grey in color

Location I.D.: FL-4 Time: Grab: 1141

Coordinates: 40° 43' 35.70"  
124° 13' 20.01"

↑ not sure if in channel

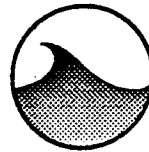
Grab  
Core#: \_\_\_\_\_ Core Length Obtained: NA Core Length Sampled: NA

Sample: Composite/Discrete Core subsampled: Yes/No Sub Sample Interval: \_\_\_\_\_

Water Depth (+/-) Tide M.L.L.W. Sampling Depth = 28.0  
28.2 (+/-) 1.78 = M.L.L.W. Mudline Depth = 26.4  
Core Length = 1.6

COMMENTS: Did an initial grab for PSD characterization.

HUMBOLDT



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LABORATORIES  
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CORE SAMPLING DATA SHEET

Date: 4/3/94 Vessel: Celtic

Captain: Phil Glen Crew: L.K., M.K., W.F.

GENERAL OBSERVATIONS: Very strong winds (clear)  
Soft mud/clay w/ organic debris & eel grass. no odor  
Presence of Rango Clams & polychaetes - little sand but present  
Light Brown softer material on top 4" turning greyer & sandier  
below 4"

Location I.D.: FL-3 Time: Grab #1(1209) #2(1224) #3(1229) #3(1234) <sup>mistake</sup>

Coordinates: 40° 43' <sup>26.59</sup> ~~54.42~~" N  
124° 13' <sup>25.04</sup> ~~07.77~~" W  
#4(1250) #5(1257) #6(1306) #7(1310)  
#8(1330)

Grab Core #: \_\_\_\_\_ Core Length Obtained: NA Core Length Sampled: NA

Sample: Composite/Discrete Core subsampled: Yes/No Sub Sample Interval: NA

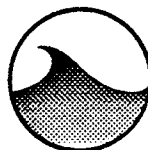
Water Depth (+/-) Tide M.L.L.W. Sampling Depth = 28

28.4  
26.6 (+/-) -(1.2) = M.L.L.W. Mudline Depth = 27.2

Core Length = .8

COMMENTS: Took 8 grabs for sediment + chemistry  
Very hard to anchor up. AT very edge of channel. Water too  
deep toward center of channel  
Generated sample # HUM94SED0031 for discrete sediment chemistry  
left over mud composited w/ ~~EX1, EX2, + EX4~~ → FL1, FL2, + FL4 (RR K.K.)

HUMBOLDT



KINNETIC  
LABORATORIES  
INCORPORATED

CORE SAMPLING DATA SHEET

Date: 4/4/94 Vessel: Celtic  
 Captain: Phil Glen Crew: KK, WF

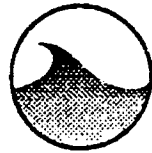
GENERAL OBSERVATIONS: Clear wind out of N.W @ 10kts.  
Very soft loose mud - very slightly clay like - slight H<sub>2</sub>S -  
no shells has. Some green algae, worm tubes, razor clams  
Very little if any sand. light brown on top 2" turning  
dark grey

Location I.D.: FL-1 Time: #1(1115) #2(1125) #3(1132) #4(1141)  
#5(1147) #6(1156) #7(1207) #8(1214)  
 Coordinates: 40° 43' 20.10"  
124° 13' 24.65"

Core#: Grab Core Length Obtained: NA Core Length Sampled: NA  
 Sample: Composite Discrete Core subsampled: Yes No Sub Sample Interval: NA

Water Depth (+/-) Tide M.L.L.W. Sampling Depth = 28  
31.75 (+/-) - (+3.96) = M.L.L.W. Mudline Depth = 27.79  
 Core Length = .21

COMMENTS: on edge of channel to find enough material  
Did 8 grabs - used the top 3" only  
Generated sample # HumaUSE00034 for discrete chemistry



**HUMBOLDT**

**CORE SAMPLING DATA SHEET**

Date: 4/4/94 Vessel: Celtic

Captain: Phil Glen Crew: EK, W.F.

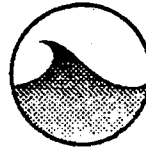
GENERAL OBSERVATIONS: Wind: 10-15 kts out of N.W - Clear  
Very loose brown mud - no shell hash - no sand - Grey + more  
clay like below 411 - some green algae - polychaetes

Location I.D.: FL-2 Time: #1(1244) #2(1257) #3(1252) #4(1302)  
#5(1310) #6(1317) #7(1330) #8(1341)  
 Coordinates: 40° 43' 22.53"  
124° 13' 18.99"

Core#: \_\_\_\_\_ Core Length Obtained: NA Core Length Sampled NA  
 Sample: Composite/Discrete Core subsampled: Yes/No Sub Sample Interval: NA

Water Depth (+/-) Tide	M.L.L.W. Sampling Depth =	<u>28.0</u>
<u>28.8</u> (+/-) <u>(+2.3)</u>	= M.L.L.W. Mudline Depth =	<u><del>27.5</del> 26.5</u>
	Core Length =	<u>1.5</u>

COMMENTS: TOOK 8- grabs for FL sediment composite  
went to last times coordinates - would have core @ this location.  
However the vibracore stopped working  
Generated sample # Hum94SE00035 for discrete chemistry


**HUMBOLDT**
**CORE SAMPLING DATA SHEET**

 Date: 4/4/94 Vessel: Celtic

 Captain: Phil Glen Crew: KK, W.F.

 GENERAL OBSERVATIONS: Winds out of N.W. @ 20knts - 2-3' swell
Medium to coarse grain sand w/ some small shell hash  
No silt.
Grey in color.  
No odor

 Location I.D.: NB-2 Time: 1410

 Coordinates: 40° 45' 29"
124° 14' 19"

Grab

 Core#: \_\_\_\_\_ Core Length Obtained: NA Core Length Sampled: NA

 Sample: Composite/Discrete \_\_\_\_\_ Core subsampled: Yes/No \_\_\_\_\_ Sub Sample Interval: NA

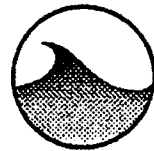
Water Depth (+/-) Tide	M.L.L.W. Sampling Depth =	<u>37</u>
<u>30.5</u> (+/-) <u>+1.72</u>	=	M.L.L.W. Mudline Depth = <u>37.48</u>
<u>38.2</u>	Core Length	= <u>-1.48</u>

COMMENTS: Took Grab to characterize PSD.

Went to edge of channel to grab

Generated Sample # Humused0037 for PSD only

HUMBOLDT



KINNETIC  
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CORE SAMPLING DATA SHEET

Date: 4/5/94 Vessel: Sally Kae  
Captain: Mike Crew: K.K. W.F.

GENERAL OBSERVATIONS: overcast - slight breeze - 5' swell  
Fine sand + silt - Thin layer (w/lem) on surface of  
most grabs. Some mollusks + polychaetes.

Location I.D.: Ref. Time: START: 0846 Finish: 1058  
Coordinates: 40° 44' 59"  
124° 20' 34"

Grabs.  
Core#: \_\_\_\_\_ Core Length Obtained: NA Core Length Sampled: NA  
Sample: Composite Discrete Core subsampled: Yes/No Sub Sample Interval: NA

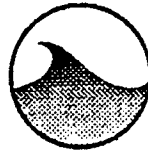
Water Depth (+/-) Tide M.L.L.W. Sampling Depth = \_\_\_\_\_  
~165 (+/-) \_\_\_\_\_ = M.L.L.W. Mudline Depth = \_\_\_\_\_  
Core Length = \_\_\_\_\_

COMMENTS: Took grabs around above coordinates using Loranc.  
TOOK 25 grabs  
Generated composite sample # HUM94 SED0043  
Also filled 3 5gal cubitainers w/ water

SFCOE MAINTENANCE DREDGING FY94

# HUMBOLDT

## CORE SAMPLING DATA SHEET



**KINNETIC  
LABORATORIES  
INCORPORATED**

Date: 4/5/94 Vessel: Sally Kay

Captain: Nike Crew: KK, wif

GENERAL OBSERVATIONS: overcast drizzle (calm) - 5' + swells  
coarse sand & shell hash - no silt - wood debris

Location I.D.: Bar Time: 1145

Coordinates: 40° 46' 06"

1240 14' 54"

Core#: NA Core Length Obtained: NA Core Length Sampled NA

Sample: Composite/Discrete Discrete Core subsampled: Yes/No Sub Sample Interval: NA

Water Depth (+/-) Tide M.L.L.W. Sampling Depth = \_\_\_\_\_

42' (+/-) \_\_\_\_\_ = M.L.L.W. Mudline Depth = \_\_\_\_\_

Core Length = \_\_\_\_\_

COMMENTS: Took grab for PSD characterization only  
went to coordinates used last time

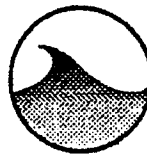
Logone for NA11



SFCOE MAINTENANCE DREDGING FY94

HUMBOLDT

CORE SAMPLING DATA SHEET



KINNETIC  
LABORATORIES  
INCORPORATED

Date: 4/5/94 Vessel: Sally Kay

Captain: \_\_\_\_\_ Crew: KIC, etc

GENERAL OBSERVATIONS: 3' swell - cloudy & drizzle

Medium grain sand w/ a verry small amount of small shell hash - brown to grey in color

Location I.D.: ENT 1 Time: 1200 hrs

Coordinates: 40° 46' 06" 45' 20"  
124° 19' 54" 13' 50"

Core#: NA Core Length Obtained: NA Core Length Sampled: NA

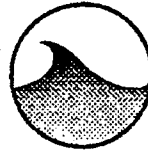
Sample: Composite/Discrete Discrete Core subsampled: Yes  No  Sub Sample Interval: NA

Water Depth (+/-) Tide \_\_\_\_\_ M.L.L.W. Sampling Depth = 37  
\_\_\_\_\_ (+/-) \_\_\_\_\_ = M.L.L.W. Mudline Depth = \_\_\_\_\_  
Core Length = \_\_\_\_\_

6 faths

COMMENTS: Took Grab to characterize PSD only  
Went to positions used last time  
Generated sample # HUM94SEDD0045  
Loran C for NAV

HUMBOLDT



KINNETIC  
LABORATORIES  
INCORPORATED

CORE SAMPLING DATA SHEET

Date: 4/5/94 Vessel: Sally Kae

Captain: Mike Crew: IK, W.F

GENERAL OBSERVATIONS: Cloudy w/ drizzle - wind 10 knots  
Medium grain sand w/ small shell trash - very little silt

Location I.D.: ENT2 Time: 1210  
hrs

Coordinates: 40° 45' 50.68"  
1240 ~~14~~ 14' 18"

Core#: \_\_\_\_\_ Core Length Obtained: NA Core Length Sampled NA

Sample: Composite/Discrete Core subsampled: Yes/No Sub Sample Interval: NA

Water Depth (+/-) Tide M.L.L.W. Sampling Depth = \_\_\_\_\_  
8 fathoms (+/-) \_\_\_\_\_ = M.L.L.W. Mudline Depth = \_\_\_\_\_  
8 fathoms Core Length = \_\_\_\_\_

COMMENTS: Took grab to characterize PSD only  
Generated sample # Hum94SED0042  
Loan C for NAV

## Appendix C

### Chemistry Results



**ToxScan, Inc.**  
Environmental Chemistry and Bioassay Laboratories  
Tel: 408/724-4522 FAX: 408/724-3188

San Francisco Army Corps of Engineers  
Humboldt Maintenance Dredge Sediments  
T-10774  
C-1

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**Percent Solids  
(%)**

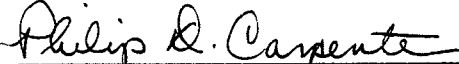
---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 19, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as percent:

---

<u>Sample Identification</u>	<u>% Solids</u>
SAM-7D	80
SAM-5D	80
SAM-3D	75
SAM-4D	78
SAM-2D	82
SAM-1D	81
SAM-6C	80
SAM-6B	75
SAM-6A	75
SAM-4(D)	78
SAM-6 Composite	77
EK-1	78
EK-2	76
EK-3	74
EK-4	63
EK Composite	72

Detection Limit = 20 ppm

  
Laboratory Director

**Percent Solids  
(%)**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 19 & May 9, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as percent:

---

<u>Sample Identification</u>	<u>% Solids</u>
NB-4 Rep 1	74
FL-8	77
FL-7	76
FL-6	69
FL-5	74
FL-3	70
FL-4	64
FL-1	64
FL-2	56
FL-Composite	65
Reference Composite	73
Home Sediment	81

Detection Limit = 20 ppm

ND = None Detected -

  
Laboratory Director

**Sulfides**  
**mg/Kg (ppm)**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 26-27, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) as indicated:

---

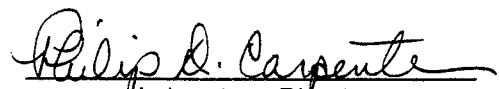
<u>Sample Identification</u>	<u>Total Sulfides as received</u>	<u>Total Sulfides dry weight</u>	<u>Water Soluble Sulfides as received</u>	<u>Water Soluble Sulfides dry weight</u>
SAM-7D	ND	ND	ND	ND
SAM-5D	ND	ND	ND	ND
SAM-3D	5.2	6.9	ND	ND
SAM-4D	ND	ND	ND	ND
SAM-2D	ND	ND	ND	ND
SAM-1D	ND	ND	ND	ND
SAM-6C	14	18	ND	ND
SAM-6B	45	60	ND	ND
SAM-6A	0.3	0.4	ND	ND
SAM-4(D)	0.6	0.8	ND	ND
SAM-6 Composite	22	29	ND	ND
EK-1	ND	ND	ND	ND
EK-2	150	200	0.3	0.4
EK-3	48	65	ND	ND
EK-4	47	75	ND	ND
EK Composite	76	110	ND	ND

Detection Limit

0.1

0.1

ND = None Detected

  
Laboratory Director



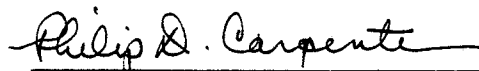
**Sulfides  
 mg/Kg (ppm)**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**DATE COMPLETED:** April 26-27 & May 9, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) as indicated:

<u>Sample Identification</u>	<u>Total Sulfides as received</u>	<u>Total Sulfides dry weight</u>	<u>Water Soluble Sulfides as received</u>	<u>Water Soluble Sulfides dry weight</u>
NB-4 Rep 1	15	20	ND	ND
FL-8	3.2	4.2	ND	ND
FL-7	5.5	7.2	ND	ND
FL-6	69	100	0.5	0.7
FL-5	2.8	3.8	ND	ND
FL-3	34	49	ND	ND
FL-4	29	45	ND	ND
FL-1	11	17	ND	ND
FL-2	26	46	ND	ND
FL-Composite	23	35	0.1	0.2
Reference Composite	0.2	0.3	ND	ND
Home Sediment	0.3	0.4	ND	ND

Detection Limit 0.1 0.1

ND = None Detected

  
 Laboratory Director

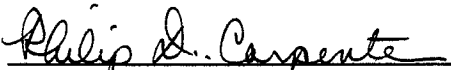
**Total Organic Carbon (TOC)  
(%)**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: May 9, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as percent:

---

<u>Sample Identification</u>	<u>TOC as received</u>	<u>TOC dry weight</u>
SAM-7D	ND	ND
SAM-5D	ND	ND
SAM-3D	0.4	0.5
SAM-4D	ND	ND
SAM-2D	ND	ND
SAM-1D	ND	ND
SAM-6C	0.2	0.3
SAM-6B	0.3	0.4
SAM-6A	0.3	0.4
SAM-4(D)	ND	ND
SAM-6 Composite	0.3	0.3
EK-1	ND	ND
EK-2	0.3	0.4
EK-3	0.4	0.5
EK-4	0.7	1.0
EK Composite	0.5	0.7
Detection Limit		0.1

  
Laboratory Director

**Total Organic Carbon (TOC)**  
(%)


---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: May 9, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as percent:

---

<u>Sample Identification</u>	<u>TOC as received</u>	<u>TOC dry weight</u>
NB-4 Rep 1	0.3	0.5
FL-8	0.08	0.1
FL-7	0.3	0.4
FL-6	0.5	0.7
FL-5	0.3	0.4
FL-3	0.5	0.7
FL-4	0.6	0.9
FL-1	0.6	0.9
FL-2	0.7	1.3
FL-Composite	0.5	0.8
Reference Composite	0.3	0.4
Home Sediment	ND	ND

Detection Limit 0.1

  
Laboratory Director

**Metals**  
**mg/Kg (ppm)**  
**As Received**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 28 - May 23, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) as received:

---

<u>Element</u>	<u>SAM-7D</u>	<u>SAM-5D</u>	<u>SAM-3D</u>	<u>SAM-4D</u>	<u>SAM-2D</u>	<u>SAM-1D</u>
Arsenic	4.7	5.1	6.2	4.3	3.9	4.4
Cadmium	ND	ND	0.1	ND	ND	ND
Chromium	62	89	85	50	59	53
Copper	4.6	4.8	26	4.1	4.6	4.6
Lead	4.9	4.6	7.8	4.0	3.4	3.7
Mercury	0.02	0.02	0.06	0.02	0.02	0.02
Nickel	33	33	76	33	33	32
Selenium	ND	ND	0.1	ND	ND	ND
Silver	ND	ND	ND	ND	ND	ND
Zinc	25	26	53	23	25	25

Mercury Detection Limit = 0.02

All other Detection Limits = 0.1

ND = None Detected

  
Laboratory Director

**Metals**  
**mg/Kg (ppm)**  
**As Received**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 28 - May 23, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) as received:

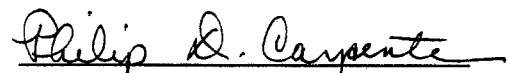
---

<u>Element</u>	<u>SAM-6C</u>	<u>SAM-6B</u>	<u>SAM-6A</u>	<u>SAM-4(D)</u>	<u>SAM-6 Composite</u>
Arsenic	4.3	4.3	5.4	5.0	4.4
Cadmium	ND	0.1	0.1	ND	0.1
Chromium	78	72	86	49	79
Copper	7.8	9.1	12	7.2	10
Lead	4.3	4.5	24.9	4.3	4.5
Mercury	0.04	0.04	0.06	0.02	0.04
Nickel	42	46	54	37	48
Selenium	ND	ND	ND	ND	ND
Silver	ND	ND	ND	ND	ND
Zinc	30	32	36	27	31

Mercury Detection Limit = 0.02

All other Detection Limits = 0.1

ND = None Detected

  
Laboratory Director

**Metals**  
**mg/Kg (ppm)**  
**As Received**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 DATE COMPLETED: April 28 - May 23, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) as received:

<u>Element</u>	<u>EK-1</u>	<u>EK-2</u>	<u>EK-3</u>	<u>EK-4</u>	<u>EK Composite</u>
Arsenic	3.7	4.2	4.7	4.5	4.5
Cadmium	ND	ND	ND	0.1	0.1
Chromium	54	69	69	75	70
Copper	4.7	11	11	18	14
Lead	3.5	5.1	5.9	6.3	5.5
Mercury	0.02	0.04	0.05	0.05	0.04
Nickel	37	52	54	66	59
Selenium	ND	ND	ND	0.1	ND
Silver	ND	ND	ND	ND	ND
Zinc	24	36	37	47	40

Mercury Detection Limit = 0.02

All other Detection Limits = 0.1

ND = None Detected

*Philip D. Carpenter*  
 Laboratory Director

**Metals**  
**mg/Kg (ppm)**  
**As Received**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 28 - May 23, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) as received:

---

<u>Element</u>	<u>NB-4 Rep 1</u>	<u>FL-8</u>	<u>FL-7</u>	<u>FL-6</u>	<u>FL-5</u>	<u>FL-3</u>
Arsenic	5.0	3.8	4.3	2.5	5.0	4.9
Cadmium	0.1	ND	0.1	0.1	ND	0.1
Chromium	76	68	91	60	65	76
Copper	18	7.1	8.6	8.5	8.5	15
Lead	6.2	4.2	4.3	3.5	3.8	5.5
Mercury	0.05	0.04	0.04	0.05	0.04	0.05
Nickel	67	50	50	43	50	64
Selenium	ND	ND	ND	ND	ND	ND
Silver	ND	ND	ND	ND	ND	ND
Zinc	43	31	31	28	31	42

Mercury Detection Limit = 0.02

All other Detection Limits = 0.1

ND = None Detected

  
Laboratory Director

**Metals**  
**mg/Kg (ppm)**  
**As Received**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 28 - May 23, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) as received:

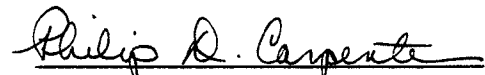
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<u>Element</u>	<u>FL-4</u>	<u>FL-1</u>	<u>FL-2</u>	<u>FL Composite</u>	<u>Reference Composite</u>	<u>Home Sediment</u>
Arsenic	5.1	5.2	5.0	4.5	5.3	3.3
Cadmium	0.1	0.1	0.2	0.1	0.1	0.1
Chromium	79	75	68	68	77	23
Copper	18	21	21	18	15	4.5
Lead	5.6	6.1	6.6	5.3	5.4	2.3
Mercury	0.05	0.08	0.06	0.05	0.14	0.02
Nickel	67	71	64	63	71	15
Selenium	ND	ND	ND	ND	ND	ND
Silver	ND	ND	0.06	ND	ND	ND
Zinc	45	47	46	43	45	15

Mercury Detection Limit = 0.02

All other Detection Limits = 0.1

ND = None Detected

  
Laboratory Director



**Metals**  
**mg/Kg (ppm)**  
**Dry Weight**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 28 - May 23, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) on a dry weight basis:

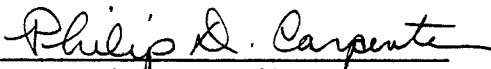
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<u>Element</u>	<u>SAM-7D</u>	<u>SAM-5D</u>	<u>SAM-3D</u>	<u>SAM-4D</u>	<u>SAM-2D</u>	<u>SAM-1D</u>
Arsenic	5.9	6.3	8.2	5.5	4.7	5.4
Cadmium	ND	ND	0.2	ND	ND	ND
Chromium	77	110	110	64	72	66
Copper	5.7	6.0	34	5.2	5.6	5.7
Lead	6.1	5.7	10	5.2	4.2	4.6
Mercury	0.02	0.02	0.08	0.02	0.02	0.02
Nickel	41	41	100	43	40	40
Selenium	ND	ND	0.2	ND	ND	ND
Silver	ND	ND	ND	ND	ND	ND
Zinc	31	32	71	30	30	31

Mercury Detection Limit = 0.02

All other Detection Limits = 0.1

ND = None Detected

  
Laboratory Director

**Metals**  
**mg/Kg (ppm)**  
**Dry Weight**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 28 - May 23, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) on a dry weight basis:

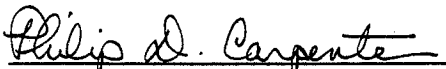
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<u>Element</u>	<u>SAM-6C</u>	<u>SAM-6B</u>	<u>SAM-6A</u>	<u>SAM-4(D)</u>	<u>SAM-6 Composite</u>
Arsenic	5.4	5.7	7.2	6.4	5.7
Cadmium	ND	0.1	0.1	ND	0.1
Chromium	98	97	120	63	100
Copper	10	12	16	9.2	12
Lead	5.4	6.0	6.6	5.5	5.8
Mercury	0.05	0.05	0.08	ND	0.05
Nickel	53	62	72	47	62
Selenium	ND	ND	ND	ND	ND
Silver	ND	ND	ND	ND	ND
Zinc	38	43	48	35	41

Mercury Detection Limit = 0.02

All other Detection Limits = 0.1

ND = None Detected

  
Laboratory Director

**Metals  
mg/Kg (ppm)  
Dry Weight**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 28 - May 23, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) on a dry weight basis:

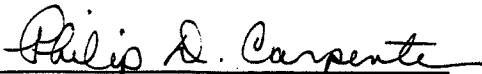
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<u>Element</u>	<u>EK-1</u>	<u>EK-2</u>	<u>EK-3</u>	<u>EK-4</u>	<u>EK Composite</u>
Arsenic	4.7	5.6	6.4	7.2	6.3
Cadmium	ND	ND	ND	0.1	0.2
Chromium	69	91	93	120	97
Copper	6.0	14	15	29	20
Lead	4.5	6.7	7.9	10	7.7
Mercury	0.03	0.05	0.06	0.08	0.06
Nickel	48	69	74	100	81
Selenium	ND	ND	ND	0.2	ND
Silver	ND	ND	ND	ND	ND
Zinc	30	47	50	75	55

Mercury Detection Limit = 0.02

All other Detection Limits = 0.1

ND = None Detected

  
Laboratory Director

**Metals**  
**mg/Kg (ppm)**  
**Dry Weight**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 28 - May 23, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) on a dry weight basis:

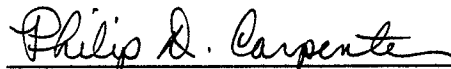
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<u>Element</u>	<u>NB-4 Rep 1</u>	<u>FL-8</u>	<u>FL-7</u>	<u>FL-6</u>	<u>FL-5</u>	<u>FL-3</u>
Arsenic	6.7	4.9	5.7	3.7	6.8	6.9
Cadmium	0.1	ND	0.1	0.2	ND	0.1
Chromium	100	88	120	87	88	110
Copper	24	9.3	11	12	11	21
Lead	8.4	5.4	5.7	5.1	5.1	7.9
Mercury	0.06	0.05	ND	0.07	0.05	0.07
Nickel	90	64	65	62	68	92
Selenium	ND	ND	ND	ND	ND	ND
Silver	ND	ND	ND	ND	ND	ND
Zinc	58	41	40	40	42	60

Mercury Detection Limit = 0.02

All other Detection Limits = 0.1

ND = None Detected

  
Laboratory Director

**Metals**  
**mg/Kg (ppm)**  
**Dry Weight**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 28 - May 23, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) on a dry weight basis:


---

<u>Element</u>	<u>FL-4</u>	<u>FL-1</u>	<u>FL-2</u>	<u>FL Composite</u>	<u>Reference Composite</u>	<u>Home Sediment</u>
Arsenic	8.0	8.1	8.9	7.0	7.3	4.1
Cadmium	0.2	0.2	0.1	0.2	0.1	0.1
Chromium	120	120	120	100	110	28
Copper	29	32	38	28	21	5.6
Lead	8.7	9.5	12	8.1	7.4	2.9
Mercury	0.08	0.13	0.10	0.08	0.20	0.03
Nickel	110	110	110	98	97	19
Selenium	ND	ND	ND	ND	ND	ND
Silver	ND	ND	0.1	ND	ND	ND
Zinc	70	74	82	66	62	19

Mercury Detection Limit = 0.02

All other Detection Limits = 0.1

ND = None Detected

  
Laboratory Director


**Oil & Grease**  
**Standard Method 5520C**  
**mg/Kg (ppm)**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 25 and May 3, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) as indicated:

---

<u>Sample Identification</u>	<u>Oil &amp; Grease as received</u>	<u>Oil &amp; Grease dry weight</u>
SAM-7D	ND	ND
SAM-5D	ND	ND
SAM-3D	ND	ND
SAM-4D	ND	ND
SAM-2D	ND	ND
SAM-1D	ND	ND
SAM-6C	ND	ND
SAM-6B	ND	ND
SAM-6A	ND	ND
SAM-4(D)	ND	ND
SAM-6 Composite	ND	ND
EK-1	ND	ND
EK-2	ND	ND
EK-3	ND	ND
EK-4	ND	ND
EK Composite	ND	ND
Detection Limit	10	20

  
Laboratory Director



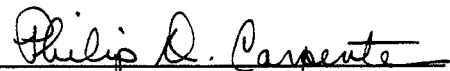
**Total Petroleum Hydrocarbons  
Standard Method 5520F  
mg/Kg (ppm)**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 25 and May 3, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) as indicated:

---

<u>Sample Identification</u>	<u>Total Petroleum Hydrocarbons as received</u>	<u>Total Petroleum Hydrocarbons dry weight</u>
SAM-7D	ND	ND
SAM-5D	ND	ND
SAM-3D	ND	ND
SAM-4D	ND	ND
SAM-2D	ND	ND
SAM-1D	ND	ND
SAM-6C	ND	ND
SAM-6B	ND	ND
SAM-6A	ND	ND
SAM-4(D)	ND	ND
SAM-6 Composite	ND	ND
EK-1	ND	ND
EK-2	ND	ND
EK-3	ND	ND
EK-4	ND	ND
EK Composite	ND	ND
Detection Limit	10	20

  
Laboratory Director



**ToxScan, Inc.**  
Environmental Chemistry and Bioassay Laboratories  
Tel: 408/724-4522 FAX: 408/724-3188

San Francisco Army Corps of Engineers  
Humboldt Maintenance Dredge Sediments  
T-10774  
C-21

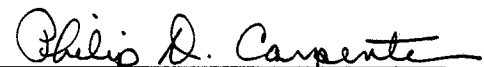
**Total Petroleum Hydrocarbons  
Standard Method 5520F  
mg/Kg (ppm)**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 25, May 3 & 12, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as milligrams per kilogram (parts per million) as indicated:

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<u>Sample Identification</u>	<u>Total Petroleum Hydrocarbons as received</u>	<u>Total Petroleum Hydrocarbons dry weight</u>
NB-4 Rep 1	ND	ND
FL-8	ND	ND
FL-7	ND	ND
FL-6	ND	ND
FL-5	ND	ND
FL-4	ND	ND
FL-1	ND	ND
FL-2	ND	ND
FL-Composite	ND	ND
Reference Composite	ND	ND
Home Sediment	30	37
Detection Limit	10	20

  
Laboratory Director

Organotin Speciation  
 $\mu\text{g/Kg}$  (ppb)  
As Received

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 20-21, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion) as received:

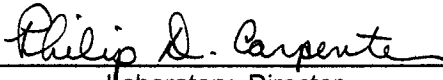
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<u>Sample ID</u>	<u>Monobutyltin</u>	<u>Dibutyltin</u>	<u>Tributyltin</u>	<u>Tetrabutyltin</u>	<u>% TPT SUR</u>
SAM-7D	ND	ND	ND	ND	51
SAM-5D	ND	ND	ND	ND	62
SAM-3D	ND	ND	ND	ND	60
SAM-4D	ND	ND	ND	ND	59
SAM-2D	ND	ND	ND	ND	49
SAM-1D	15	ND	ND	ND	51
SAM-6C	ND	ND	ND	ND	48
SAM-6B	ND	ND	2	ND	46
SAM-6A	ND	ND	ND	ND	51
SAM-4(D)	ND	ND	ND	ND	61
SAM-6 Composite	ND	ND	1	ND	62
EK-1	ND	ND	ND	ND	67
EK-2	ND	ND	ND	ND	60
EK-3	ND	ND	1	ND	60
EK-4	1	ND	2	ND	67
EK Composite	ND	ND	ND	ND	57

TPT Sur = Tripropyltin surrogate recovery

ND = None Detected

Detection Limit = 1 ppb

  
Laboratory Director

**Organotin Speciation**  
**µg/Kg (ppb)**  
**As Received**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 20-21 & May 9, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion) as received:


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<u>Sample ID</u>	<u>Monobutyltin</u>	<u>Dibutyltin</u>	<u>Tributyltin</u>	<u>Tetrabutyltin</u>	<u>% TPT SUR</u>
NB-4 Rep 1	ND	ND	ND	ND	57
FL-8	ND	ND	ND	ND	56
FL-7	ND	ND	ND	ND	64
FL-6	ND	ND	ND	ND	60
FL-5	ND	ND	ND	ND	67
FL-3	ND	ND	ND	ND	61
FL-4	ND	ND	1	ND	69
FL-1	ND	ND	ND	ND	67
FL-2	ND	1	1	ND	51
FL-Composite	ND	ND	ND	ND	63
Reference Composite	ND	ND	ND	ND	57
Home Sediment	ND	ND	ND	ND	54

TPT Sur = Tripropyltin surrogate recovery

ND = None Detected

Detection Limit = 1 ppb

  
Laboratory Director

**Organotin Speciation**  
**µg/Kg (ppb)**  
**Dry Weight**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 20-21, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion) on a dry weight basis:

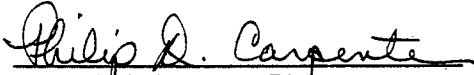
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<u>Sample ID</u>	<u>Monobutyltin</u>	<u>Dibutyltin</u>	<u>Tributyltin</u>	<u>Tetrabutyltin</u>	<u>% TPT SUR</u>
SAM-7D	ND	ND	ND	ND	51
SAM-5D	ND	ND	ND	ND	62
SAM-3D	ND	ND	ND	ND	60
SAM-4D	ND	ND	ND	ND	59
SAM-2D	ND	ND	ND	ND	49
SAM-1D	19	ND	ND	ND	51
SAM-6C	ND	ND	ND	ND	48
SAM-6B	ND	ND	3	ND	46
SAM-6A	ND	ND	ND	ND	51
SAM-4(D)	ND	ND	ND	ND	61
SAM-6 Composite	ND	ND	1	ND	62
EK-1	ND	ND	ND	ND	67
EK-2	ND	ND	ND	ND	60
EK-3	ND	ND	1	ND	60
EK-4	2	ND	3	ND	67
EK Composite	ND	ND	ND	ND	57

TPT Sur = Tripropyltin surrogate recovery

ND = None Detected

Detection Limit = 1 ppb

  
Laboratory Director

**Organotin Speciation**  
**µg/Kg (ppb)**  
**Dry Weight**

---

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
DATE COMPLETED: April 20-21 & May 9, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion) on a dry weight basis:

---

<u>Sample ID</u>	<u>Monobutyltin</u>	<u>Dibutyltin</u>	<u>Tributyltin</u>	<u>Tetrabutyltin</u>	<u>% TPT SUR</u>
NB-4 Rep 1	ND	ND	ND	ND	57
FL-8	ND	ND	ND	ND	56
FL-7	ND	ND	ND	ND	64
FL-6	ND	ND	ND	ND	60
FL-5	ND	ND	ND	ND	67
FL-3	ND	ND	ND	ND	61
FL-4	ND	ND	2	ND	69
FL-1	ND	ND	ND	ND	67
FL-2	ND	2	2	ND	51
FL-Composite	ND	ND	ND	ND	63
Reference Composite	ND	ND	ND	ND	57
Home Sediment	ND	ND	ND	ND	54

TPT Sur = Tripropyltin surrogate recovery

ND = None Detected

Detection Limit = 1 ppb

  
Laboratory Director

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 14, 1994  
DATE COMPLETED: April 28, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0031 Site ID: FL-3 <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	37	20	26	14
Acenaphthylene	ND	20	ND	14
Acenaphthene	ND	20	ND	14
Fluorene	ND	20	ND	14
Phenanthrene	ND	20	ND	14
Anthracene	ND	20	ND	14
Fluoranthene	28	20	20	14
Pyrene	42	25	29	18
Benzo(a)anthracene	ND	20	ND	14
Chrysene	27	20	19	14
Benzo(b)fluoranthene	37	20	26	14
Benzo(k)fluoranthene	ND	20	ND	14
Benzo(a)pyrene	ND	20	ND	14
Indeno(1,2,3-cd)pyrene	ND	20	ND	14
Dibenzo(a,h)anthracene	ND	20	ND	14
Benzo(ghi)perylene	ND	20	ND	14
2-Methylnaphthalene	45	20	31	14
Total phthalates:	180	10	120	7.0
Total PAHs:	220	20	150	14

ND = None Detected

*Phillip D. Carpenter*  
Laboratory Director *DBL*

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 14, 1994  
**DATE COMPLETED:** April 28, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

<u>Sample ID:</u> HUM94SED0033	<u>Dry Wt.</u>	<u>Dry Wt.</u>	<u>Wet Wt.</u>	<u>Wet Wt.</u>
<u>Site ID:</u> FL-4	<u>Sample</u>	<u>Detection</u>	<u>Sample</u>	<u>Detection</u>
<u>Analyte</u>	<u>Value</u>	<u>Limit</u>	<u>Value</u>	<u>Limit</u>
Naphthalene	38	20	27	14
Acenaphthylene	ND	20	ND	14
Acenaphthene	ND	20	ND	14
Fluorene	ND	20	ND	14
Phenanthrene	ND	100*	ND	70*
Anthracene	ND	20	ND	14
Fluoranthene	33	20	23	14
Pyrene	38	25	27	18
Benzo(a)anthracene	ND	20	ND	14
Chrysene	ND	20	ND	14
Benzo(b)fluoranthene	39	20	27	14
Benzo(k)fluoranthene	ND	20	ND	14
Benzo(a)pyrene	ND	20	ND	14
Indeno(1,2,3-cd)pyrene	ND	20	ND	14
Dibenzo(a,h)anthracene	ND	20	ND	14
Benzo(ghi)perylene	ND	20	ND	14
2-Methylnaphthalene	66	20	46	14
<b>Total phthalates:</b>	<b>240</b>	<b>10</b>	<b>170</b>	<b>7.0</b>
<b>Total PAHs:</b>	<b>210</b>	<b>20</b>	<b>150</b>	<b>14</b>

ND = None Detected

\* Detection Limit raised due to matrix interference.

*Philip D. Carpenter* / DBC  
 Laboratory Director

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 14, 1994  
 DATE COMPLETED: April 28, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0034 Site ID: FL-1 <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	42	20	27	13
Acenaphthylene	ND	20	ND	13
Acenaphthene	ND	20	ND	13
Fluorene	ND	20	ND	13
Phenanthrene	ND	100*	ND	64*
Anthracene	ND	20	ND	13
Fluoranthene	45	20	29	13
Pyrene	56	25	36	16
Benzo(a)anthracene	ND	20	ND	13
Chrysene	41	20	26	13
Benzo(b)fluoranthene	ND	20	ND	13
Benzo(k)fluoranthene	ND	20	ND	13
Benzo(a)pyrene	ND	20	ND	13
Indeno(1,2,3-cd)pyrene	ND	20	ND	13
Dibenzo(a,h)anthracene	ND	20	ND	13
Benzo(ghi)perylene	ND	20	ND	13
2-Methylnaphthalene	89	20	57	13
Total phthalates:	170	10	110	6.4
Total PAHs:	270	20	180	13

ND = None Detected

\* Detection limit raised due to matrix interference.

*Philip D Carpenter*  
 Laboratory Director *(DBC)*



**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

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MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 14, 1994  
DATE COMPLETED: April 28, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

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<u>Sample ID:</u> HUM94SED0035	<u>Dry Wt.</u>	<u>Dry Wt.</u>	<u>Wet Wt.</u>	<u>Wet Wt.</u>
<u>Site ID:</u> FL-2	<u>Sample</u>	<u>Detection</u>	<u>Sample</u>	<u>Detection</u>
<u>Analyte</u>	<u>Value</u>	<u>Limit</u>	<u>Value</u>	<u>Limit</u>
Naphthalene	52	20	33	13
Acenaphthylene	ND	20	ND	13
Acenaphthene	ND	20	ND	13
Fluorene	ND	20	ND	13
Phenanthrene	120	20	79	13
Anthracene	ND	20	ND	13
Fluoranthene	48	20	31	13
Pyrene	110	25	68	16
Benzo(a)anthracene	ND	20	ND	13
Chrysene	46	20	29	13
Benzo(b)fluoranthene	55	20	36	13
Benzo(k)fluoranthene	ND	20	ND	13
Benzo(a)pyrene	40	20	26	13
Indeno(1,2,3-cd)pyrene	ND	20	ND	13
Dibenzo(a,h)anthracene	ND	20	ND	13
Benzo(ghi)perylene	ND	20	ND	13
2-Methylnaphthalene	100	20	64	13
Total phthalates:	210	10	140	6.4
Total PAHs:	570	20	370	13

ND = None Detected

*Philip D. Carpenter* / DBL  
Laboratory Director

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
 $\mu\text{g/Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 14, 1994  
 DATE COMPLETED: April 28, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0036 Site ID: FL-Comp. <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	43	20	28	13
Acenaphthylene	ND	20	ND	13
Acenaphthene	ND	20	ND	13
Fluorene	ND	20	ND	13
Phenanthrene	ND	20	ND	13
Anthracene	ND	20	ND	13
Fluoranthene	42	20	27	13
Pyrene	ND	25	ND	16
Benzo(a)anthracene	ND	20	ND	13
Chrysene	ND	20	ND	13
Benzo(b)fluoranthene	ND	20	ND	13
Benzo(k)fluoranthene	ND	20	ND	13
Benzo(a)pyrene	ND	20	ND	13
Indeno(1,2,3-cd)pyrene	ND	20	ND	13
Dibenzo(a,h)anthracene	ND	20	ND	13
Benzo(ghi)perylene	ND	20	ND	13
2-Methylnaphthalene	78	20	51	13
Total phthalates:	170	10	110	6.4
Total PAHs:	160	20	110	13

ND = None Detected

*Philip D. Carpenter*  
 Laboratory Director *DBL*

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

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MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 13, 1994  
DATE COMPLETED: April 29, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

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Sample ID: HUM94SED0001 Site ID: SAM-7D <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	ND	20	ND	16
Acenaphthylene	ND	20	ND	16
Acenaphthene	ND	20	ND	16
Fluorene	ND	20	ND	16
Phenanthrene	ND	20	ND	16
Anthracene	ND	20	ND	16
Fluoranthene	ND	20	ND	16
Pyrene	ND	25	ND	20
Benzo(a)anthracene	ND	20	ND	16
Chrysene	ND	20	ND	16
Benzo(b)fluoranthene	ND	20	ND	16
Benzo(k)fluoranthene	ND	20	ND	16
Benzo(a)pyrene	ND	20	ND	16
Indeno(1,2,3-cd)pyrene	ND	20	ND	16
Dibenzo(a,h)anthracene	ND	20	ND	16
Benzo(ghi)perylene	ND	20	ND	16
2-Methylnaphthalene	ND	20	ND	16
Total phthalates:	110	10	80	8.0
Total PAHs:	ND	20	ND	16

ND = None Detected

*Philip D. Carpenter*  
Laboratory Director *DBC*

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 13, 1994  
DATE COMPLETED: April 29, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0002 Site ID: SAM-5D <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	ND	20	ND	16
Acenaphthylene	ND	20	ND	16
Acenaphthene	ND	20	ND	16
Fluorene	ND	20	ND	16
Phenanthrene	ND	20	ND	16
Anthracene	ND	20	ND	16
Fluoranthene	ND	20	ND	16
Pyrene	ND	25	ND	20
Benzo(a)anthracene	ND	20	ND	16
Chrysene	ND	20	ND	16
Benzo(b)fluoranthene	ND	20	ND	16
Benzo(k)fluoranthene	ND	20	ND	16
Benzo(a)pyrene	ND	20	ND	16
Indeno(1,2,3-cd)pyrene	ND	20	ND	16
Dibenzo(a,h)anthracene	ND	20	ND	16
Benzo(ghi)perylene	ND	20	ND	16
2-Methylnaphthalene	ND	20	ND	16
Total phthalates:	110	10	87	8.0
Total PAHs:	ND	20	ND	16

ND = None Detected

*Philip P. Carpenter*  
Laboratory Director *DBC*

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 13, 1994  
**DATE COMPLETED:** April 29, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0003 Site ID: SAM-4D <u>Analyte</u>	<u>Dry Wt. Sample Value</u>	<u>Dry Wt. Detection Limit</u>	<u>Wet Wt. Sample Value</u>	<u>Wet Wt. Detection Limit</u>
Naphthalene	46	20	36	16
Acenaphthylene	ND	20	ND	16
Acenaphthene	ND	20	ND	16
Fluorene	ND	20	ND	16
Phenanthrene	79	20	61	16
Anthracene	ND	20	ND	16
Fluoranthene	ND	20	ND	16
Pyrene	35	25	27	20
Benzo(a)anthracene	ND	20	ND	16
Chrysene	31	20	25	16
Benzo(b)fluoranthene	ND	20	ND	16
Benzo(k)fluoranthene	ND	20	ND	16
Benzo(a)pyrene	ND	20	ND	16
Indeno(1,2,3-cd)pyrene	ND	20	ND	16
Dibenzo(a,h)anthracene	ND	20	ND	16
Benzo(ghi)perylene	ND	20	ND	16
2-Methylnaphthalene	58	20	45	16
<b>Total phthalates:</b>	<b>180</b>	<b>10</b>	<b>140</b>	<b>7.8</b>
<b>Total PAHs:</b>	<b>250</b>	<b>20</b>	<b>190</b>	<b>16</b>

ND = None Detected

*Philip D. Carpenter*  
 Laboratory Director *DBc*

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 13, 1994  
 DATE COMPLETED: April 29, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0004 Site ID: SAM-3D <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	ND	20	ND	15
Acenaphthylene	ND	20	ND	15
Acenaphthene	ND	20	ND	15
Fluorene	ND	20	ND	15
Phenanthrene	ND	20	ND	15
Anthracene	ND	20	ND	15
Fluoranthene	ND	20	ND	15
Pyrene	29	25	22	19
Benzo(a)anthracene	ND	20	ND	15
Chrysene	ND	20	ND	15
Benzo(b)fluoranthene	ND	20	ND	15
Benzo(k)fluoranthene	ND	20	ND	15
Benzo(a)pyrene	ND	20	ND	15
Indeno(1,2,3-cd)pyrene	ND	20	ND	15
Dibenzo(a,h)anthracene	ND	20	ND	15
Benzo(ghi)perylene	ND	20	ND	15
2-Methylnaphthalene	ND	20	ND	15
Total phthalates:	180	10	140	7.5
Total PAHs:	29	20	22	15

ND = None Detected

*Philip D. Carpenter*  
 Laboratory Director / DBL

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 13, 1994  
**DATE COMPLETED:** April 29, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0005 Site ID: SAM-2D <u>Analyte</u>	<u>Dry Wt. Sample Value</u>	<u>Dry Wt. Detection Limit</u>	<u>Wet Wt. Sample Value</u>	<u>Wet Wt. Detection Limit</u>
Naphthalene	ND	20	ND	16
Acenaphthylene	ND	20	ND	16
Acenaphthene	ND	20	ND	16
Fluorene	ND	20	ND	16
Phenanthrene	ND	20	ND	16
Anthracene	ND	20	ND	16
Fluoranthene	39	20	32	16
Pyrene	85	25	70	21
Benzo(a)anthracene	ND	20	ND	16
Chrysene	ND	20	ND	16
Benzo(b)fluoranthene	ND	20	ND	16
Benzo(k)fluoranthene	ND	20	ND	16
Benzo(a)pyrene	ND	20	ND	16
Indeno(1,2,3-cd)pyrene	ND	20	ND	16
Dibenzo(a,h)anthracene	ND	20	ND	16
Benzo(ghi)perylene	ND	20	ND	16
2-Methylnaphthalene	ND	20	ND	16
<b>Total phthalates:</b>	<b>210</b>	<b>10</b>	<b>180</b>	<b>8.2</b>
<b>Total PAHs:</b>	<b>120</b>	<b>20</b>	<b>100</b>	<b>16</b>

ND = None Detected

*Philip D. Carpenter*  
 Laboratory Director *DBC*

Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters  
EPA Method 8270  
 $\mu\text{g/Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 13, 1994  
DATE COMPLETED: April 29, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0006 Site ID: SAM-1D <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	ND	20	ND	16
Acenaphthylene	ND	20	ND	16
Acenaphthene	ND	20	ND	16
Fluorene	ND	20	ND	16
Phenanthrene	ND	20	ND	16
Anthracene	ND	20	ND	16
Fluoranthene	21	20	17	16
Pyrene	ND	25	ND	20
Benzo(a)anthracene	ND	20	ND	16
Chrysene	ND	20	ND	16
Benzo(b)fluoranthene	ND	20	ND	16
Benzo(k)fluoranthene	ND	20	ND	16
Benzo(a)pyrene	ND	20	ND	16
Indeno(1,2,3-cd)pyrene	ND	20	ND	16
Dibenzo(a,h)anthracene	ND	20	ND	16
Benzo(ghi)perylene	ND	20	ND	16
2-Methylnaphthalene	ND	20	ND	16
Total phthalates:	130	10	100	8.1
Total PAHs:	21	20	17	16

ND = None Detected

*Philip D. Carpenter*  
Laboratory Director *DBC*



**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 13, 1994  
**DATE COMPLETED:** April 29, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

<u>Analyte</u>	<u>Dry Wt. Sample Value</u>	<u>Dry Wt. Detection Limit</u>	<u>Wet Wt. Sample Value</u>	<u>Wet Wt. Detection Limit</u>
Naphthalene	ND	20	ND	16
Acenaphthylene	ND	20	ND	16
Acenaphthene	ND	20	ND	16
Fluorene	ND	20	ND	16
Phenanthrene	ND	20	ND	16
Anthracene	ND	20	ND	16
Fluoranthene	ND	20	ND	16
Pyrene	29	25	23	20
Benzo(a)anthracene	ND	20	ND	16
Chrysene	ND	20	ND	16
Benzo(b)fluoranthene	ND	20	ND	16
Benzo(k)fluoranthene	ND	20	ND	16
Benzo(a)pyrene	ND	20	ND	16
Indeno(1,2,3-cd)pyrene	ND	20	ND	16
Dibenzo(a,h)anthracene	ND	20	ND	16
Benzo(ghi)perylene	ND	20	ND	16
2-Methylnaphthalene	ND	20	ND	16
Total phthalates:	140	10	110	8.0
Total PAHs:	29	20	23	16

ND = None Detected

*Philip D. Carpenter*  
Laboratory Director *DBC*

Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters  
EPA Method 8270  
 $\mu\text{g/Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 13, 1994  
DATE COMPLETED: April 29, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0008 Site ID: SAM-6B <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	78	20	58	15
Acenaphthylene	ND	20	ND	15
Acenaphthene	ND	20	ND	15
Fluorene	ND	20	ND	15
Phenanthrene	ND	20	ND	15
Anthracene	ND	20	ND	15
Fluoranthene	39	20	29	15
Pyrene	56	25	42	19
Benzo(a)anthracene	ND	20	ND	15
Chrysene	ND	20	ND	15
Benzo(b)fluoranthene	ND	20	ND	15
Benzo(k)fluoranthene	ND	20	ND	15
Benzo(a)pyrene	ND	20	ND	15
Indeno(1,2,3-cd)pyrene	ND	20	ND	15
Dibenzo(a,h)anthracene	ND	20	ND	15
Benzo(ghi)perylene	ND	20	ND	15
2-Methylnaphthalene	ND	20	ND	15
Total phthalates:	160	10	120	7.5
Total PAHs:	170	20	130	15

ND = None Detected

*Philip D. Carpenter*  
Laboratory Director / DBL

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

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MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 14, 1994  
DATE COMPLETED: April 29, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

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Sample ID: HUM94SED0028 Site ID: FL-7 <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	84	20	64	15
Acenaphthylene	ND	20	ND	15
Acenaphthene	ND	20	ND	15
Fluorene	ND	20	ND	15
Phenanthrene	70	20	54	15
Anthracene	ND	20	ND	15
Fluoranthene	ND	20	ND	15
Pyrene	31	25	23	19
Benzo(a)anthracene	ND	20	ND	15
Chrysene	38	20	29	15
Benzo(b)fluoranthene	ND	20	ND	15
Benzo(k)fluoranthene	ND	20	ND	15
Benzo(a)pyrene	ND	20	ND	15
Indeno(1,2,3-cd)pyrene	ND	20	ND	15
Dibenzo(a,h)anthracene	ND	20	ND	15
Benzo(ghi)perylene	ND	20	ND	15
2-Methylnaphthalene	59	20	45	15
Total phthalates:	250	10	190	7.6
Total PAHs:	280	20	220	15

ND = None Detected

*Philip D. Carpenter*  
Laboratory Director *DBL*

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

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MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 14, 1994  
DATE COMPLETED: April 30, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

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<u>Sample ID: HUM94SED0009</u> <u>Site ID: SAM-6A</u> <u>Analyte</u>	<u>Dry Wt.</u> <u>Sample</u> <u>Value</u>	<u>Dry Wt.</u> <u>Detection</u> <u>Limit</u>	<u>Wet Wt.</u> <u>Sample</u> <u>Value</u>	<u>Wet Wt.</u> <u>Detection</u> <u>Limit</u>
Naphthalene	ND	20	ND	15
Acenaphthylene	ND	20	ND	15
Acenaphthene	ND	20	ND	15
Fluorene	ND	20	ND	15
Phenanthrene	ND	20	ND	15
Anthracene	ND	20	ND	15
Fluoranthene	ND	20	ND	15
Pyrene	30	25	23	19
Benzo(a)anthracene	ND	20	ND	15
Chrysene	ND	20	ND	15
Benzo(b)fluoranthene	ND	20	ND	15
Benzo(k)fluoranthene	ND	20	ND	15
Benzo(a)pyrene	ND	20	ND	15
Indeno(1,2,3-cd)pyrene	ND	20	ND	15
Dibenzo(a,h)anthracene	ND	20	ND	15
Benzo(ghi)perylene	ND	20	ND	15
2-Methylnaphthalene	ND	20	ND	15
Total phthalates:	170	10	130	7.5
Total PAHs:	30	20	23	15

ND = None Detected

*Philip D. Carpenter*  
Laboratory Director

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

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**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 14, 1994  
**DATE COMPLETED:** April 30, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

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Sample ID: HUM94SED0010 Site ID: SAM-4(D) <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	ND	20	ND	16
Acenaphthylene	ND	20	ND	16
Acenaphthene	ND	20	ND	16
Fluorene	ND	20	ND	16
Phenanthrene	ND	20	ND	16
Anthracene	ND	20	ND	16
Fluoranthene	ND	20	ND	16
Pyrene	ND	25	ND	20
Benzo(a)anthracene	ND	20	ND	16
Chrysene	ND	20	ND	16
Benzo(b)fluoranthene	ND	20	ND	16
Benzo(k)fluoranthene	ND	20	ND	16
Benzo(a)pyrene	ND	20	ND	16
Indeno(1,2,3-cd)pyrene	ND	20	ND	16
Dibenzo(a,h)anthracene	ND	20	ND	16
Benzo(ghi)perylene	ND	20	ND	16
2-Methylnaphthalene	ND	20	ND	16
Total phthalates:	170	10	130	7.8
Total PAHs:	ND	20	ND	16

ND = None Detected

*Philip D. Carpenter*  
Laboratory Director

Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters  
EPA Method 8270  
 $\mu\text{g/Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 14, 1994  
DATE COMPLETED: April 30, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0011 Site ID: SAM-6 Comp. <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	43	20	33	15
Acenaphthylene	ND	20	ND	15
Acenaphthene	ND	20	ND	15
Fluorene	ND	20	ND	15
Phenanthrene	49	20	38	15
Anthracene	ND	20	ND	15
Fluoranthene	30	20	23	15
Pyrene	36	25	28	19
Benzo(a)anthracene	ND	20	ND	15
Chrysene	ND	20	ND	15
Benzo(b)fluoranthene	ND	20	ND	15
Benzo(k)fluoranthene	ND	20	ND	15
Benzo(a)pyrene	ND	20	ND	15
Indeno(1,2,3-cd)pyrene	ND	20	ND	15
Dibenzo(a,h)anthracene	ND	20	ND	15
Benzo(ghi)perylene	ND	20	ND	15
2-Methylnaphthalene	ND	20	ND	15
Total phthalates:	160	10	120	8.0
Total PAHs:	160	20	120	15

ND = None Detected

*Philip D. Carpenter*  
Laboratory Director

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 14, 1994  
**DATE COMPLETED:** April 30, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

<u>Sample ID: HUM94SED0022</u> <u>Site ID: EK-1</u> <u>Analyte</u>	<u>Dry Wt.</u> <u>Sample</u> <u>Value</u>	<u>Dry Wt.</u> <u>Detection</u> <u>Limit</u>	<u>Wet Wt.</u> <u>Sample</u> <u>Value</u>	<u>Wet Wt.</u> <u>Detection</u> <u>Limit</u>
Naphthalene	ND	20	ND	16
Acenaphthylene	ND	20	ND	16
Acenaphthene	ND	20	ND	16
Fluorene	ND	20	ND	16
Phenanthrene	ND	20	ND	16
Anthracene	ND	20	ND	16
Fluoranthene	ND	20	ND	16
Pyrene	ND	25	ND	20
Benzo(a)anthracene	ND	20	ND	16
Chrysene	ND	20	ND	16
Benzo(b)fluoranthene	ND	20	ND	16
Benzo(k)fluoranthene	ND	20	ND	16
Benzo(a)pyrene	ND	20	ND	16
Indeno(1,2,3-cd)pyrene	ND	20	ND	16
Dibenzo(a,h)anthracene	ND	20	ND	16
Benzo(ghi)perylene	ND	20	ND	16
2-Methylnaphthalene	ND	20	ND	16
Total phthalates:	280	10	220	7.8
Total PAHs:	ND	20	ND	16

ND = None Detected

*Philip J. Carpenter*  
Laboratory Director *DBC*

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 14, 1994  
 DATE COMPLETED: April 30, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0023 Site ID: EK-2 <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	29	20	22	15
Acenaphthylene	ND	20	ND	15
Acenaphthene	ND	20	ND	15
Fluorene	ND	20	ND	15
Phenanthrene	56	20	43	15
Anthracene	ND	20	ND	15
Fluoranthene	ND	100*	ND	76*
Pyrene	71	25	54	19
Benzo(a)anthracene	ND	20	ND	15
Chrysene	34	20	26	15
Benzo(b)fluoranthene	69	20	52	15
Benzo(k)fluoranthene	ND	20	ND	15
Benzo(a)pyrene	ND	20	ND	15
Indeno(1,2,3-cd)pyrene	ND	20	ND	15
Dibenzo(a,h)anthracene	ND	20	ND	15
Benzo(ghi)perylene	ND	20	ND	15
2-Methylnaphthalene	ND	20	ND	15
Total phthalates:	180	10	140	7.6
Total PAHs:	260	20	200	15

ND = None Detected

\* Detection Limit raised due to matrix interference.

*Philip D. Carpenter*  
 Laboratory Director *DBc*



**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

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MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 14, 1994  
DATE COMPLETED: April 30, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

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Sample ID: HUM94SED0024 Site ID: EK-3 <u>Analyte</u>	<u>Dry Wt.</u> Sample <u>Value</u>	<u>Dry Wt.</u> Detection <u>Limit</u>	<u>Wet Wt.</u> Sample <u>Value</u>	<u>Wet Wt.</u> Detection <u>Limit</u>
Naphthalene	54	20	40	15
Acenaphthylene	ND	20	ND	15
Acenaphthene	ND	20	ND	15
Fluorene	ND	20	ND	15
Phenanthrene	59	20	44	15
Anthracene	ND	20	ND	15
Fluoranthene	ND	20	ND	15
Pyrene	42	25	31	19
Benzo(a)anthracene	ND	20	ND	15
Chrysene	ND	20	ND	15
Benzo(b)fluoranthene	28	20	21	15
Benzo(k)fluoranthene	ND	20	ND	15
Benzo(a)pyrene	ND	20	ND	15
Indeno(1,2,3-cd)pyrene	ND	20	ND	15
Dibenzo(a,h)anthracene	ND	20	ND	15
Benzo(ghi)perylene	ND	20	ND	15
2-Methylnaphthalene	44	20	32	15
Total phthalates:	180	10	130	7.4
Total PAHs:	230	20	170	16

ND = None Detected

*Philip D. Carpenter*  
Laboratory Director *DBC*

Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters  
EPA Method 8270  
 $\mu\text{g/Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 14, 1994  
DATE COMPLETED: April 30, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0025 Site ID: EK-4 <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	69	20	43	13
Acenaphthylene	ND	20	ND	13
Acenaphthene	ND	20	ND	13
Fluorene	53	20	33	13
Phenanthrene	160	20	100	13
Anthracene	ND	20	ND	13
Fluoranthene	71	20	45	13
Pyrene	92	25	58	16
Benzo(a)anthracene	ND	20	ND	13
Chrysene	54	20	34	13
Benzo(b)fluoranthene	58	20	37	13
Benzo(k)fluoranthene	ND	20	ND	13
Benzo(a)pyrene	41	20	26	13
Indeno(1,2,3-cd)pyrene	ND	20	ND	13
Dibenzo(a,h)anthracene	ND	20	ND	13
Benzo(ghi)perylene	ND	20	ND	13
2-Methylnaphthalene	76	20	48	13
Total phthalates:	200	10	130	6.3
Total PAHs:	670	20	420	13

ND = None Detected

*Philip D. Carpenter*  
Laboratory Director

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 14, 1994  
**DATE COMPLETED:** April 30, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0026 Site ID: EK-Comp. <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	37	20	28	15
Acenaphthylene	ND	20	ND	15
Acenaphthene	ND	20	ND	15
Fluorene	ND	20	ND	15
Phenanthrene	ND	20	ND	15
Anthracene	ND	20	ND	15
Fluoranthene	39	20	30	15
Pyrene	47	25	36	19
Benzo(a)anthracene	ND	20	ND	15
Chrysene	ND	20	ND	15
Benzo(b)fluoranthene	ND	20	ND	15
Benzo(k)fluoranthene	ND	20	ND	15
Benzo(a)pyrene	ND	20	ND	15
Indeno(1,2,3-cd)pyrene	ND	20	ND	15
Dibenzo(a,h)anthracene	ND	20	ND	15
Benzo(ghi)perylene	ND	20	ND	15
2-Methylnaphthalene	41	20	31	15
<b>Total phthalates:</b>	<b>160</b>	<b>10</b>	<b>120</b>	<b>8.0</b>
<b>Total PAHs:</b>	<b>160</b>	<b>20</b>	<b>130</b>	<b>15</b>

ND = None Detected

*Philip D. Carpenter*  
 Laboratory Director *DBL*

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 14, 1994  
 DATE COMPLETED: April 30, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0027 Site ID: FL-8 <u>Analyte</u>	<u>Dry Wt.</u> Sample <u>Value</u>	<u>Dry Wt.</u> Detection <u>Limit</u>	<u>Wet Wt.</u> Sample <u>Value</u>	<u>Wet Wt.</u> Detection <u>Limit</u>
Naphthalene	ND	20	ND	15
Acenaphthylene	ND	20	ND	15
Acenaphthene	ND	20	ND	15
Fluorene	ND	20	ND	15
Phenanthrene	ND	20	ND	15
Anthracene	ND	20	ND	15
Fluoranthene	34	20	26	15
Pyrene	33	25	25	19
Benzo(a)anthracene	ND	20	ND	15
Chrysene	20	20	16	15
Benzo(b)fluoranthene	ND	20	ND	15
Benzo(k)fluoranthene	ND	20	ND	15
Benzo(a)pyrene	ND	20	ND	15
Indeno(1,2,3-cd)pyrene	ND	20	ND	15
Dibenzo(a,h)anthracene	ND	20	ND	15
Benzo(ghi)perylene	ND	20	ND	15
2-Methylnaphthalene	ND	20	ND	15
Total phthalates:	190	10	140	7.7
Total PAHs:	90	20	70	15

ND = None Detected

*Philip D. Carpenter*  
 Laboratory Director *JBC*

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

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MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 14, 1994  
DATE COMPLETED: April 30, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

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Sample ID: HUM94SED0029 Site ID: FL-6 <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	33	20	23	14
Acenaphthylene	ND	20	ND	14
Acenaphthene	ND	20	ND	14
Fluorene	ND	20	ND	14
Phenanthrene	ND	20	ND	14
Anthracene	ND	20	ND	14
Fluoranthene	47	20	32	14
Pyrene	52	25	36	17
Benzo(a)anthracene	ND	20	ND	14
Chrysene	ND	20	ND	14
Benzo(b)fluoranthene	ND	20	ND	14
Benzo(k)fluoranthene	ND	20	ND	14
Benzo(a)pyrene	ND	20	ND	14
Indeno(1,2,3-cd)pyrene	ND	20	ND	14
Dibenzo(a,h)anthracene	ND	20	ND	14
Benzo(ghi)perylene	ND	20	ND	14
2-Methylnaphthalene	38	20	26	14
Total phthalates:	190	10	130	6.9
Total PAHs:	170	20	120	14

ND = None Detected

*Philip D. Carpenter*  
Laboratory Director

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 14, 1994  
DATE COMPLETED: April 30, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0030 Site ID: FL-5 <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	28	20	21	15
Acenaphthylene	ND	20	ND	15
Acenaphthene	ND	20	ND	15
Fluorene	ND	20	ND	15
Phenanthrene	ND	20	ND	15
Anthracene	ND	20	ND	15
Fluoranthene	ND	20	ND	15
Pyrene	ND	25	ND	19
Benzo(a)anthracene	ND	20	ND	15
Chrysene	33	20	25	15
Benzo(b)fluoranthene	ND	20	ND	15
Benzo(k)fluoranthene	ND	20	ND	15
Benzo(a)pyrene	ND	20	ND	15
Indeno(1,2,3-cd)pyrene	ND	20	ND	15
Dibenzo(a,h)anthracene	ND	20	ND	15
Benzo(ghi)perylene	ND	20	ND	15
2-Methylnaphthalene	42	20	31	15
Total phthalates:	130	10	99	7.4
Total PAHs:	100	20	80	15

ND = None Detected

*Philip D. Carpenter*  
Laboratory Director *DBC*

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: May 1, 1994  
DATE COMPLETED: May 6, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

<u>Analyte</u>	<u>Dry Wt. Sample Value</u>	<u>Dry Wt. Detection Limit</u>	<u>Wet Wt. Sample Value</u>	<u>Wet Wt. Detection Limit</u>
Naphthalene	ND	20	ND	15
Acenaphthylene	ND	20	ND	15
Acenaphthene	ND	20	ND	15
Fluorene	ND	20	ND	15
Phenanthrene	58	20	42	15
Anthracene	ND	20	ND	15
Fluoranthene	ND	20	ND	15
Pyrene	ND	25	ND	18
Benzo(a)anthracene	ND	20	ND	15
Chrysene	ND	20	ND	15
Benzo(b)fluoranthene	ND	20	ND	15
Benzo(k)fluoranthene	ND	20	ND	15
Benzo(a)pyrene	ND	20	ND	15
Indeno(1,2,3-cd)pyrene	ND	20	ND	15
Dibenzo(a,h)anthracene	ND	20	ND	15
Benzo(ghi)perylene	ND	20	ND	15
2-Methylnaphthalene	40	20	29	15
Total phthalates:	1200	10	910	7.0
Total PAHs:	100	20	70	15

ND = None Detected

*Philip D. Carpenter*  
Laboratory Director *03C*

**ToxScan, Inc.**Environmental Chemistry and Bioassay Laboratories  
Tel: 408/724-4522 FAX: 408/724-3188San Francisco Army Corps of Engineers  
Humboldt Maintenance Dredge Sediments  
T-10774  
C-52**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: May 5, 1994  
 DATE COMPLETED: May 10, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Site ID: HOME SED <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	ND	20	ND	16
Acenaphthylene	ND	20	ND	16
Acenaphthene	ND	20	ND	16
Fluorene	ND	20	ND	16
Phenanthrene	25	20	21	16
Anthracene	ND	20	ND	16
Fluoranthene	24	20	20	16
Pyrene	ND	25	ND	20
Benzo(a)anthracene	ND	20	ND	16
Chrysene	ND	20	ND	16
Benzo(b)fluoranthene	ND	20	ND	16
Benzo(k)fluoranthene	ND	20	ND	16
Benzo(a)pyrene	ND	20	ND	16
Indeno(1,2,3-cd)pyrene	ND	20	ND	16
Dibenzo(a,h)anthracene	ND	20	ND	16
Benzo(ghi)perylene	ND	20	ND	16
2-Methylnaphthalene	ND	20	ND	16
Total phthalates:	96	10	77	8.1
Total PAHs:	50	20	40	16

ND = None Detected

*Philip D. Carpenter*  
 Laboratory Director / DBL



Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters  
EPA Method 8270  
 $\mu\text{g/Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: June 21, 1994  
DATE COMPLETED: June 23, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0018 Site ID: NB-4 Rep 1 <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Naphthalene	ND	20	ND	15
Acenaphthylene	ND	20	ND	15
Acenaphthene	ND	20	ND	15
Fluorene	ND	20	ND	15
Phenanthrene	34	20	25	15
Anthracene	ND	20	ND	15
Fluoranthene	ND	20	ND	15
Pyrene	ND	20	ND	15
Benzo(a)anthracene	ND	20	ND	15
Chrysene	ND	20	ND	15
Benzo(b)fluoranthene	ND	20	ND	15
Benzo(k)fluoranthene	ND	20	ND	15
Benzo(a)pyrene	ND	20	ND	15
Indeno(1,2,3-cd)pyrene	ND	20	ND	15
Dibenzo(a,h)anthracene	ND	20	ND	15
Benzo(ghi)perylene	ND	20	ND	15
2-Methylnaphthalene	27	20	20	15
Total phthalates:	230	10	170	7.4
Total PAHs:	60	20	50	15

ND = None Detected

*Philip D. Carpenter*  
Laboratory Director DBL

**ToxScan, Inc.**Environmental Chemistry and Bioassay Laboratories  
Tel: 408/724-4522 FAX: 408/724-3188San Francisco Army Corps of Engineers  
Humboldt Maintenance Dredge Sediments

T-10774

C-54

**Chlorinated Pesticides  
EPA Method 8080  
µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 13, 1994  
 DATE COMPLETED: May 20-21, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0001 Site ID: SAM-7D <u>Analyte</u>	<u>Dry Wt. Sample Value</u>	<u>Dry Wt. Detection Limit</u>	<u>Wet Wt. Sample Value</u>	<u>Wet Wt. Detection Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.0
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.0
Toxaphene	ND	30	ND	24
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

*Philip D. Carpenter*  
Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
 $\mu\text{g/Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 13, 1994  
 DATE COMPLETED: May 20-21, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0002 Site ID: SAM-5D <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.0
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.0
Toxaphene	ND	30	ND	24
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected


*Philip D. Carpenter*  
 Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
**µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 13, 1994  
 DATE COMPLETED: May 20-21, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0003 Site ID: SAM-4D <u>Analyte</u>	<u>Dry Wt.</u> <u>Sample</u> <u>Value</u>	<u>Dry Wt.</u> <u>Detection</u> <u>Limit</u>	<u>Wet Wt.</u> <u>Sample</u> <u>Value</u>	<u>Wet Wt.</u> <u>Detection</u> <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	7.8
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	7.8
Toxaphene	ND	30	ND	23
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected


  
 Laboratory Director

Chlorinated Pesticides  
EPA Method 8080  
 $\mu\text{g/Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 13, 1994  
DATE COMPLETED: May 20-21, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0004 Site ID: SAM-3D Analyte	Dry Wt. Sample Value	Dry Wt. Detection Limit	Wet Wt. Sample Value	Wet Wt. Detection Limit
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.5
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	7.5
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	7.5
Toxaphene	ND	30	ND	23
PCBs:				
PCB 1242	ND	20	ND	15
PCB 1248	ND	20	ND	15
PCB 1254	ND	20	ND	15
PCB 1260	ND	20	ND	15
TOTAL PCBs	ND	20	ND	15

ND = None detected


  
Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
**µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 13, 1994  
 DATE COMPLETED: May 20-21, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0005 Site ID: SAM-2D <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.2
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.2
Toxaphene	ND	30	ND	25
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

  
 Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
**µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 13, 1994  
 DATE COMPLETED: May 20-21, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0006 Site ID: SAM-1D Analyte	Dry Wt. Sample Value	Dry Wt. Detection Limit	Wet Wt. Sample Value	Wet Wt. Detection Limit
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.1
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.1
Toxaphene	ND	30	ND	24
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

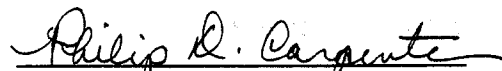
*Philip D. Carpenter*  
 Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
 $\mu\text{g/Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 13, 1994  
 DATE COMPLETED: May 20-21, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0007 Site ID: SAM-6C <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.0
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.0
Toxaphene	ND	30	ND	24
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

  
 Laboratory Director



**Chlorinated Pesticides**  
**EPA Method 8080**  
**µg/Kg (ppb)**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 13, 1994  
**DATE COMPLETED:** May 20-21, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0008 Site ID: SAM-6B <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.5
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	7.5
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	7.5
Toxaphene	ND	30	ND	23
PCBs:				
PCB 1242	ND	20	ND	15
PCB 1248	ND	20	ND	15
PCB 1254	ND	20	ND	15
PCB 1260	ND	20	ND	15
TOTAL PCBs	ND	20	ND	15

ND = None detected

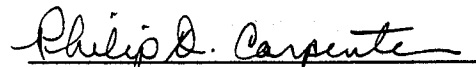
*Philip D. Carpenter*  
Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
**µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 15, 1994  
DATE COMPLETED: May 9, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

<u>Sample ID: HUM94SED0009</u> <u>Site ID: SAM-6A</u> <u>Analyte</u>	<u>Dry Wt.</u> <u>Sample</u> <u>Value</u>	<u>Dry Wt.</u> <u>Detection</u> <u>Limit</u>	<u>Wet Wt.</u> <u>Sample</u> <u>Value</u>	<u>Wet Wt.</u> <u>Detection</u> <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.5
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	7.5
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	7.5
Toxaphene	ND	30	ND	23
PCBs:				
PCB 1242	ND	20	ND	15
PCB 1248	ND	20	ND	15
PCB 1254	ND	20	ND	15
PCB 1260	ND	20	ND	15
TOTAL PCBs	ND	20	ND	15

ND = None detected

  
Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
**µg/Kg (ppb)**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 15, 1994  
**DATE COMPLETED:** May 9, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0010 Site ID: SAM-4(D) <u>Analyte</u>	<u>Dry Wt.</u> Sample <u>Value</u>	<u>Dry Wt.</u> Detection <u>Limit</u>	<u>Wet Wt.</u> Sample <u>Value</u>	<u>Wet Wt.</u> Detection <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	7.8
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	7.8
Toxaphene	ND	30	ND	23
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

*Philip R. Carpenter*  
 Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
 $\mu\text{g/Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 15, 1994  
 DATE COMPLETED: May 9, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0011 Site ID: SAM-6 Comp. <u>Analyte</u>	<u>Dry Wt.</u> <u>Sample</u> <u>Value</u>	<u>Dry Wt.</u> <u>Detection</u> <u>Limit</u>	<u>Wet Wt.</u> <u>Sample</u> <u>Value</u>	<u>Wet Wt.</u> <u>Detection</u> <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.5
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	7.7
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	7.7
Toxaphene	ND	30	ND	23
PCBs:				
PCB 1242	ND	20	ND	15
PCB 1248	ND	20	ND	15
PCB 1254	ND	20	ND	15
PCB 1260	ND	20	ND	15
TOTAL PCBs	ND	20	ND	15

ND = None detected

*Philip D. Carpenter*  
 Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
**µg/Kg (ppb)**

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**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 15, 1994  
**DATE COMPLETED:** May 10, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

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<u>Analyte</u>	<u>Dry Wt. Sample Value</u>	<u>Dry Wt. Detection Limit</u>	<u>Wet Wt. Sample Value</u>	<u>Wet Wt. Detection Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	7.8
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	7.8
Toxaphene	ND	30	ND	23
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

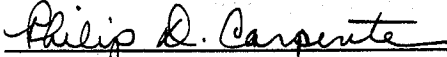
  
Laboratory Director

Chlorinated Pesticides  
EPA Method 8080  
 $\mu\text{g}/\text{Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 15, 1994  
DATE COMPLETED: May 10, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0023 Site ID: EK-2 <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.0
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.0
Toxaphene	ND	30	ND	24
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

  
Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
**µg/Kg (ppb)**


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**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 15, 1994  
**DATE COMPLETED:** May 10, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

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<u>Sample ID: HUM94SED0024</u> <u>Site ID: EK-3</u> <u>Analyte</u>	<u>Dry Wt.</u> <u>Sample</u> <u>Value</u>	<u>Dry Wt.</u> <u>Detection</u> <u>Limit</u>	<u>Wet Wt.</u> <u>Sample</u> <u>Value</u>	<u>Wet Wt.</u> <u>Detection</u> <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.0
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.0
Toxaphene	ND	30	ND	24
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

  
Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
 $\mu\text{g/Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 15, 1994  
 DATE COMPLETED: May 10, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0025 Site ID: EK-4 <u>Analyte</u>	<u>Dry Wt.</u> <u>Sample</u> <u>Value</u>	<u>Dry Wt.</u> <u>Detection</u> <u>Limit</u>	<u>Wet Wt.</u> <u>Sample</u> <u>Value</u>	<u>Wet Wt.</u> <u>Detection</u> <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	7.8
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	7.8
Toxaphene	ND	30	ND	23
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

*Philip S. Carpenter*  
 Laboratory Director



**Chlorinated Pesticides**  
**EPA Method 8080**  
**µg/Kg (ppb)**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 15, 1994  
**DATE COMPLETED:** May 10, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0026 Site ID: EK-Comp. <u>Analyte</u>	<u>Dry Wt.</u> Sample <u>Value</u>	<u>Dry Wt.</u> Detection <u>Limit</u>	<u>Wet Wt.</u> Sample <u>Value</u>	<u>Wet Wt.</u> Detection <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.5
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	7.5
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	7.5
Toxaphene	ND	30	ND	23
PCBs:				
PCB 1242	ND	20	ND	15
PCB 1248	ND	20	ND	15
PCB 1254	ND	20	ND	15
PCB 1260	ND	20	ND	15
TOTAL PCBs	ND	20	ND	15

ND = None detected

*Philip D. Carante*  
 Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
**µg/Kg (ppb)**


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MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 15, 1994  
DATE COMPLETED: May 10, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

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Sample ID: HUM94SED0027 Site ID: FL-8 <u>Analyte</u>	<u>Dry Wt.</u> <u>Sample</u> <u>Value</u>	<u>Dry Wt.</u> <u>Detection</u> <u>Limit</u>	<u>Wet Wt.</u> <u>Sample</u> <u>Value</u>	<u>Wet Wt.</u> <u>Detection</u> <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.2
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.2
Toxaphene	ND	30	ND	25
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

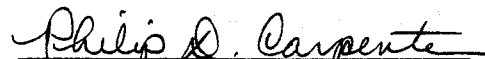
  
Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
**µg/Kg (ppb)**

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 15, 1994  
DATE COMPLETED: May 10, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

<u>Sample ID:</u> HUM94SED0028 <u>Site ID:</u> FL-7 <u>Analyte</u>	<u>Dry Wt.</u> <u>Sample</u> <u>Value</u>	<u>Dry Wt.</u> <u>Detection</u> <u>Limit</u>	<u>Wet Wt.</u> <u>Sample</u> <u>Value</u>	<u>Wet Wt.</u> <u>Detection</u> <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.1
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.1
Toxaphene	ND	30	ND	24
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

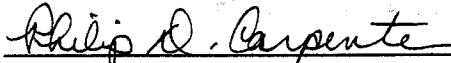
  
Laboratory Director

Chlorinated Pesticides  
EPA Method 8080  
 $\mu\text{g}/\text{Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 15, 1994  
DATE COMPLETED: May 19, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0029 Site ID: FL-6 <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.0
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.0
Toxaphene	ND	30	ND	24
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

  
Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
 $\mu\text{g/Kg (ppb)}$

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 15, 1994  
**DATE COMPLETED:** May 19, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0030 Site ID: FL-5 <u>Analyte</u>	<u>Dry Wt.</u> <u>Sample</u> <u>Value</u>	<u>Dry Wt.</u> <u>Detection</u> <u>Limit</u>	<u>Wet Wt.</u> <u>Sample</u> <u>Value</u>	<u>Wet Wt.</u> <u>Detection</u> <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.5
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	7.5
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	7.5
Toxaphene	ND	30	ND	23
PCBs:				
PCB 1242	ND	20	ND	15
PCB 1248	ND	20	ND	15
PCB 1254	ND	20	ND	15
PCB 1260	ND	20	ND	15
TOTAL PCBs	ND	20	ND	15

ND = None detected

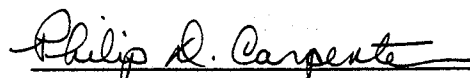
*Philip J. Carpenter*  
 Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
 $\mu\text{g/Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 15, 1994  
 DATE COMPLETED: May 19, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0031 Site ID: FL-3 <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.5
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	7.5
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	7.5
Toxaphene	ND	30	ND	23
PCBs:				
PCB 1242	ND	20	ND	15
PCB 1248	ND	20	ND	15
PCB 1254	ND	20	ND	15
PCB 1260	ND	20	ND	15
TOTAL PCBs	ND	20	ND	15

ND = None detected

  
 Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
**µg/Kg (ppb)**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 15, 1994  
**DATE COMPLETED:** May 19, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0033 Site ID: FL-4 <u>Analyte</u>	<u>Dry Wt.</u> Sample <u>Value</u>	<u>Dry Wt.</u> Detection <u>Limit</u>	<u>Wet Wt.</u> Sample <u>Value</u>	<u>Wet Wt.</u> Detection <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.0
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.0
Toxaphene	ND	30	ND	24
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected


*Philip D. Carpenter*  
 Laboratory Director

Chlorinated Pesticides  
EPA Method 8080  
 $\mu\text{g}/\text{Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: April 15, 1994  
DATE COMPLETED: May 19, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0034 Site ID: FL-1 <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	7.8
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	7.8
Toxaphene	ND	30	ND	23
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

  
Laboratory Director



**Chlorinated Pesticides**  
**EPA Method 8080**  
**µg/Kg (ppb)**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 15, 1994  
**DATE COMPLETED:** May 19, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0035 Site ID: FL-2 <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.5
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	7.5
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	7.5
Toxaphene	ND	30	ND	23
PCBs:				
PCB 1242	ND	20	ND	15
PCB 1248	ND	20	ND	15
PCB 1254	ND	20	ND	15
PCB 1260	ND	20	ND	15
TOTAL PCBs	ND	20	ND	15

ND = None detected

*Philip D. Carpenter*  
 Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
 $\mu\text{g/Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 15, 1994  
 DATE COMPLETED: May 19, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Sample ID: HUM94SED0036 Site ID: FL-Comp. <u>Analyte</u>	Dry Wt. Sample <u>Value</u>	Dry Wt. Detection <u>Limit</u>	Wet Wt. Sample <u>Value</u>	Wet Wt. Detection <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.2
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.2
Toxaphene	ND	30	ND	25
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

*Philip D. Carpenter*  
 Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
 $\mu\text{g/Kg}$  (ppb)

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**EXTRACTION DATE:** April 15, 1994  
**DATE COMPLETED:** May 19, 1994  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

<u>Analyte</u>	<u>Dry Wt. Sample Value</u>	<u>Dry Wt. Detection Limit</u>	<u>Wet Wt. Sample Value</u>	<u>Wet Wt. Detection Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.1
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.1
Toxaphene	ND	30	ND	24
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

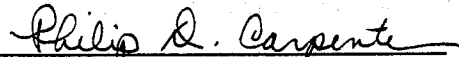
*Philip D. Carpenter*  
 Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
 $\mu\text{g/Kg}$  (ppb)

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 EXTRACTION DATE: April 13, 1994  
 DATE COMPLETED: May 21, 1994  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

Site ID: HOME-SED Analyte	Dry Wt. Sample Value	Dry Wt. Detection Limit	Wet Wt. Sample Value	Wet Wt. Detection Limit
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.1
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.1
Toxaphene	ND	30	ND	24
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

  
 Laboratory Director

**Chlorinated Pesticides**  
**EPA Method 8080**  
**µg/Kg (ppb)**


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MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
EXTRACTION DATE: June 21, 1994  
DATE COMPLETED: June 23, 1994  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows, expressed as micrograms per kilogram (parts per billion):

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<u>Sample ID: HUM94SED0018</u> <u>Site ID: NB-4 Rep 1</u> <u>Analyte</u>	<u>Dry Wt.</u> <u>Sample</u> <u>Value</u>	<u>Dry Wt.</u> <u>Detection</u> <u>Limit</u>	<u>Wet Wt.</u> <u>Sample</u> <u>Value</u>	<u>Wet Wt.</u> <u>Detection</u> <u>Limit</u>
Aldrin	ND	0.5	ND	0.4
alpha-BHC	ND	1.0	ND	0.8
beta-BHC	ND	1.0	ND	0.8
delta-BHC	ND	1.0	ND	0.8
gamma-BHC (lindane)	ND	1.0	ND	0.8
alpha-Chlordane	ND	1.0	ND	0.8
gamma-Chlordane	ND	1.0	ND	0.8
4,4'-DDD	ND	1.0	ND	0.8
4,4'-DDE	ND	1.0	ND	0.8
4,4'-DDT	ND	1.0	ND	0.8
Dieldrin	ND	0.5	ND	0.4
Endosulfan I	ND	2.0	ND	1.6
Endosulfan II	ND	0.5	ND	0.4
Endosulfan sulfate	ND	10	ND	8.0
Endrin	ND	0.5	ND	0.4
Heptachlor	ND	0.5	ND	0.4
Heptachlor epoxide	ND	10	ND	8.0
Toxaphene	ND	30	ND	24
PCBs:				
PCB 1242	ND	20	ND	16
PCB 1248	ND	20	ND	16
PCB 1254	ND	20	ND	16
PCB 1260	ND	20	ND	16
TOTAL PCBs	ND	20	ND	16

ND = None detected

  
Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 SAM-7D**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.20	0.8	0.8
1	1-0.5	0.30	1.2	2.0
2	0.5-0.25	12.95	52.1	54.1
3	0.25-0.125	10.80	43.4	97.5
4	0.125-0.062	0.17	0.7	98.2
5	0.062-0.031	0.04	0.2	98.4
6	0.031-0.016	0.03	0.1	98.5
7	0.016-0.008	0.00	0.0	98.5
8	0.008-0.004	0.09	0.4	98.9
9	0.004-0.002	0.01	0.0	98.9
>9	< 0.002	0.28	1.1	100.0
		total wt	coarse wt	fine wt
		24.9	24.4	0.4
		% sand	% silt	% clay
		98.23	0.65	1.14

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 SAM-5D**

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MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

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SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.13	0.5	0.5
1	1-0.5	1.52	5.9	6.4
2	0.5-0.25	18.60	72.5	78.9
3	0.25-0.125	5.05	19.7	98.6
4	0.125-0.062	0.11	0.4	99.0
5	0.062-0.031	0.04	0.1	99.2
6	0.031-0.016	0.00	0.0	99.2
7	0.016-0.008	0.00	0.0	99.2
8	0.008-0.004	0.03	0.1	99.3
9	0.004-0.002	0.00	0.0	99.3
>9	< 0.002	0.19	0.8	100.0
		total wt	coarse wt	fine wt
		25.7	25.4	0.2
		% sand	% silt	% clay
		99.05	0.25	0.79

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 SAM-3D**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.09	0.4	0.4
1	1-0.5	0.57	2.6	3.0
2	0.5-0.25	6.41	29.4	32.4
3	0.25-0.125	5.12	23.4	55.8
4	0.125-0.062	0.61	2.8	58.6
5	0.062-0.031	1.89	8.6	67.3
6	0.031-0.016	1.75	8.0	75.2
7	0.016-0.008	1.20	5.5	80.7
8	0.008-0.004	1.05	4.8	85.5
9	0.004-0.002	0.75	3.4	89.0
>9	< 0.002	2.41	11.0	100.0
		total wt	coarse wt	fine wt
		21.8	12.8	9.0
		% sand	% silt	% clay
		58.62	26.91	14.47

*Philip D. Carpenter*  
 Laboratory Director



**Particle Size  
 Plumb, 1981  
 (%)  
 SAM-4D**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.13	0.5	0.5
1	1-0.5	0.22	0.9	1.4
2	0.5-0.25	11.12	45.7	47.2
3	0.25-0.125	12.10	49.8	97.0
4	0.125-0.062	0.11	0.5	97.4
5	0.062-0.031	0.19	0.8	98.2
6	0.031-0.016	0.14	0.6	98.8
7	0.016-0.008	0.00	0.0	98.8
8	0.008-0.004	0.03	0.1	98.9
9	0.004-0.002	0.00	0.0	98.9
>9	< 0.002	0.28	1.2	100.0
total wt			coarse wt	fine wt
24.3			23.7	0.6
% sand			% silt	% clay
97.41			1.52	1.18

*Philip D. Carpenter*  
 Laboratory Director

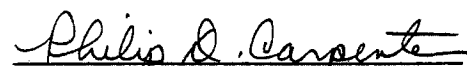
Particle Size  
Plumb, 1981  
(%)  
SAM-2D

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MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows:

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SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	1.59	6.5	6.5
1	1-0.5	3.67	15.1	21.7
2	0.5-0.25	15.73	64.8	86.4
3	0.25-0.125	3.02	12.4	98.9
4	0.125-0.062	0.02	0.1	99.0
5	0.062-0.031	0.01	0.0	99.0
6	0.031-0.016	0.00	0.0	99.0
7	0.016-0.008	0.03	0.1	99.1
8	0.008-0.004	0.01	0.0	99.2
9	0.004-0.002	0.00	0.0	99.2
>9	< 0.002	0.21	0.9	100.0
		total wt	coarse wt	fine wt
		24.3	24.0	0.3
		% sand	% silt	% clay
		98.97	0.20	0.90

  
Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 SAM-1D**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.23	1.1	1.1
1	1-0.5	0.39	1.8	2.9
2	0.5-0.25	1.20	5.6	8.5
3	0.25-0.125	18.44	86.3	94.9
4	0.125-0.062	0.80	3.7	98.6
5	0.062-0.031	0.00	0.0	98.6
6	0.031-0.016	0.00	0.0	98.6
7	0.016-0.008	0.05	0.3	98.9
8	0.008-0.004	0.03	0.2	99.1
9	0.004-0.002	0.00	0.0	99.1
>9	< 0.002	0.23	1.5	100.0
		total wt	coarse wt	fine wt
		21.4	21.1	0.3
		% sand	% silt	% clay
		98.60	0.49	1.47

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 SAM-6C**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.02	0.1	0.1
1	1-0.5	0.10	0.5	0.6
2	0.5-0.25	0.71	3.3	3.8
3	0.25-0.125	16.33	74.9	78.8
4	0.125-0.062	1.56	7.2	85.9
5	0.062-0.031	0.63	2.9	88.8
6	0.031-0.016	0.50	2.3	91.1
7	0.016-0.008	0.46	2.1	93.2
8	0.008-0.004	0.40	1.8	95.0
9	0.004-0.002	0.22	1.0	96.0
>9	< 0.002	0.88	4.0	100.0
		total wt	coarse wt	fine wt
		21.8	18.7	3.1
		% sand	% silt	% clay
		85.91	9.06	5.03

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 SAM-6B**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.52	1.9	1.9
1	1-0.5	2.10	7.6	9.5
2	0.5-0.25	12.23	44.2	53.7
3	0.25-0.125	8.28	29.9	83.6
4	0.125-0.062	0.14	0.5	84.1
5	0.062-0.031	0.92	3.3	87.4
6	0.031-0.016	0.74	2.7	90.1
7	0.016-0.008	0.69	2.5	92.6
8	0.008-0.004	0.51	1.8	94.4
9	0.004-0.002	0.31	1.1	95.5
>9	< 0.002	1.25	4.5	100.0
		total wt	coarse wt	fine wt
		27.7	23.3	4.4
		% sand	% silt	% clay
		84.11	10.28	5.60

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 SAM-6A**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.16	0.8	0.8
1	1-0.5	0.32	1.5	2.3
2	0.5-0.25	2.40	11.4	13.7
3	0.25-0.125	10.07	48.0	61.7
4	0.125-0.062	1.73	8.2	69.9
5	0.062-0.031	2.08	9.9	79.8
6	0.031-0.016	1.06	5.0	84.9
7	0.016-0.008	0.77	3.7	88.5
8	0.008-0.004	0.58	2.7	91.3
9	0.004-0.002	0.35	1.6	92.9
>9	< 0.002	1.49	7.1	100.0
		total wt	coarse wt	fine wt
		21.0	14.7	6.3
		% sand	% silt	% clay
		69.92	21.36	8.72

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 SAM-4(D)**

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MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

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SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.55	2.0	2.0
1	1-0.5	5.42	20.0	22.1
2	0.5-0.25	14.86	55.0	77.0
3	0.25-0.125	4.79	17.7	94.8
4	0.125-0.062	0.13	0.5	95.2
5	0.062-0.031	0.40	1.5	96.7
6	0.031-0.016	0.22	0.8	97.5
7	0.016-0.008	0.13	0.5	98.0
8	0.008-0.004	0.12	0.4	98.4
9	0.004-0.002	0.06	0.2	98.6
>9	< 0.002	0.37	1.4	100.0
		total wt	coarse wt	fine wt
		27.0	25.8	1.3
		% sand	% silt	% clay
		95.25	3.18	1.57

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 SAM-6 Composite**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.18	0.8	0.8
1	1-0.5	0.28	1.2	2.0
2	0.5-0.25	2.87	12.4	14.4
3	0.25-0.125	14.29	61.9	76.4
4	0.125-0.062	1.05	4.6	80.9
5	0.062-0.031	1.01	4.4	85.3
6	0.031-0.016	0.83	3.6	88.9
7	0.016-0.008	0.61	2.6	91.5
8	0.008-0.004	0.41	1.8	93.3
9	0.004-0.002	0.32	1.4	94.6
>9	< 0.002	1.24	5.4	100.0
		total wt	coarse wt	fine wt
		23.1	18.7	4.4
		% sand	% silt	% clay
		80.91	12.35	6.74

*Philip D. Carpenter*  
 Laboratory Director



Particle Size  
Plumb, 1981  
(%)  
EK-1

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MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows:

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SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.31	1.3	1.3
1	1-0.5	0.75	3.2	4.5
2	0.5-0.25	14.00	60.1	64.6
3	0.25-0.125	7.46	32.0	96.7
4	0.125-0.062	0.22	0.9	97.6
5	0.062-0.031	0.09	0.4	98.0
6	0.031-0.016	0.06	0.3	98.2
7	0.016-0.008	0.10	0.4	98.6
8	0.008-0.004	0.04	0.2	98.8
9	0.004-0.002	0.00	0.0	98.8
>9	< 0.002	0.28	1.2	100.0
		total wt	coarse wt	fine wt
		23.3	22.7	0.6
		% sand	% silt	% clay
		97.60	1.22	1.18

*Philip D. Carpenter*  
Laboratory Director

**ToxScan, Inc.**  
Environmental Chemistry and Bioassay Laboratories  
Tel: 408/724-4522 FAX: 408/724-3188

San Francisco Army Corps of Engineers  
Humboldt Maintenance Dredge Sediments  
T-10774  
C-94

**Particle Size  
Plumb, 1981  
(%)  
EK-2**

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MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows:

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SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.04	0.2	0.2
1	1-0.5	0.20	0.9	1.1
2	0.5-0.25	4.18	18.5	19.5
3	0.25-0.125	11.84	52.3	71.8
4	0.125-0.062	1.32	5.8	77.6
5	0.062-0.031	1.00	4.4	82.0
6	0.031-0.016	0.86	3.8	85.8
7	0.016-0.008	0.69	3.0	88.9
8	0.008-0.004	0.57	2.5	91.4
9	0.004-0.002	0.41	1.8	93.2
>9	< 0.002	1.55	6.8	100.0
		total wt	coarse wt	fine wt
		22.6	17.6	5.1
		% sand	% silt	% clay
		77.63	13.73	8.63

  
Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 EK-3**

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MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

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SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.05	0.2	0.2
1	1-0.5	0.20	0.9	1.1
2	0.5-0.25	3.21	13.9	15.0
3	0.25-0.125	12.18	52.7	67.7
4	0.125-0.062	1.41	6.1	73.8
5	0.062-0.031	0.95	4.1	77.9
6	0.031-0.016	1.93	8.4	86.2
7	0.016-0.008	0.38	1.6	87.9
8	0.008-0.004	1.14	4.9	92.8
9	0.004-0.002	0.03	0.1	92.9
>9	< 0.002	1.64	7.1	100.0
		total wt	coarse wt	fine wt
		23.1	17.1	6.1
		% sand	% silt	% clay
		73.78	19.02	7.20

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 EK-4**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.02	0.1	0.1
1	1-0.5	0.10	0.6	0.7
2	0.5-0.25	0.37	2.1	2.8
3	0.25-0.125	2.06	11.6	14.4
4	0.125-0.062	3.24	18.3	32.7
5	0.062-0.031	2.67	15.1	47.8
6	0.031-0.016	2.60	14.7	62.5
7	0.016-0.008	2.06	11.6	74.2
8	0.008-0.004	0.58	3.3	77.4
9	0.004-0.002	1.36	7.7	85.1
>9	< 0.002	2.64	14.9	100.0
		total wt	coarse wt	fine wt
		17.7	5.8	11.9
		% sand	% silt	% clay
		32.74	44.70	22.56

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 EK Composite**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.06	0.3	0.3
1	1-0.5	0.14	0.6	0.9
2	0.5-0.25	2.52	11.1	12.0
3	0.25-0.125	8.85	39.1	51.1
4	0.125-0.062	2.25	9.9	61.1
5	0.062-0.031	1.97	8.7	69.8
6	0.031-0.016	1.95	8.6	78.4
7	0.016-0.008	1.28	5.7	84.0
8	0.008-0.004	0.74	3.3	87.3
9	0.004-0.002	0.72	3.2	90.5
>9	< 0.002	2.16	9.5	100.0
		total wt	coarse wt	fine wt
		22.6	13.8	8.8
		% sand	% silt	% clay
		61.07	26.25	12.68

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 NB-4 Rep 1**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.33	1.4	1.4
1	1-0.5	1.30	5.6	7.0
2	0.5-0.25	6.82	29.4	36.4
3	0.25-0.125	4.76	20.5	56.9
4	0.125-0.062	1.05	4.5	61.4
5	0.062-0.031	2.37	10.2	71.7
6	0.031-0.016	1.90	8.2	79.8
7	0.016-0.008	1.30	5.6	85.4
8	0.008-0.004	0.86	3.7	89.1
9	0.004-0.002	0.66	2.8	91.9
>9	< 0.002	1.88	8.1	100.0
		total wt	coarse wt	fine wt
		23.2	14.3	9.0
		% sand	% silt	% clay
		61.44	27.64	10.92

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 FL-8**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.01	0.0	0.0
1	1-0.5	0.03	0.1	0.2
2	0.5-0.25	7.09	29.4	29.6
3	0.25-0.125	15.91	66.0	95.6
4	0.125-0.062	0.48	2.0	97.6
5	0.062-0.031	0.18	0.8	98.4
6	0.031-0.016	0.00	0.0	98.4
7	0.016-0.008	0.10	0.4	98.9
8	0.008-0.004	0.00	0.0	98.9
9	0.004-0.002	0.05	0.2	99.0
>9	< 0.002	0.25	1.1	100.0
		total wt	coarse wt	fine wt
		24.1	23.5	0.6
		% sand	% silt	% clay
		97.63	1.22	1.30

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 FL-7**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.19	0.8	0.8
1	1-0.5	0.16	0.7	1.5
2	0.5-0.25	5.20	21.6	23.0
3	0.25-0.125	11.76	48.7	71.8
4	0.125-0.062	1.08	4.5	76.2
5	0.062-0.031	1.17	4.8	81.1
6	0.031-0.016	1.26	5.2	86.3
7	0.016-0.008	0.82	3.4	89.7
8	0.008-0.004	0.67	2.8	92.4
9	0.004-0.002	0.45	1.8	94.3
>9	< 0.002	1.38	5.7	100.0
		total wt	coarse wt	fine wt
		24.1	18.4	5.7
		% sand	% silt	% clay
		76.23	16.21	7.56

*Philip D. Carpenter*  
 Laboratory Director



**Particle Size  
 Plumb, 1981  
 (%)  
 FL-6**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.35	1.7	1.7
1	1-0.5	1.65	7.9	9.6
2	0.5-0.25	5.54	26.6	36.2
3	0.25-0.125	8.64	41.5	77.7
4	0.125-0.062	1.66	8.0	85.7
5	0.062-0.031	0.76	3.6	89.3
6	0.031-0.016	0.34	1.6	90.9
7	0.016-0.008	0.61	2.9	93.9
8	0.008-0.004	0.25	1.2	95.1
9	0.004-0.002	0.24	1.1	96.2
>9	< 0.002	0.79	3.8	100.0
		total wt	coarse wt	fine wt
		20.8	17.8	3.0
		% sand	% silt	% clay
		85.69	9.39	4.92

*Philip D. Carpenter*  
 Laboratory Director

Particle Size  
Plumb, 1981  
(%)  
FL-5

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MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows:

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SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.05	0.2	0.2
1	1-0.5	0.11	0.5	0.7
2	0.5-0.25	3.35	15.3	16.0
3	0.25-0.125	13.89	63.5	79.5
4	0.125-0.062	0.84	3.8	83.4
5	0.062-0.031	0.97	4.4	87.8
6	0.031-0.016	0.28	1.3	89.1
7	0.016-0.008	0.78	3.5	92.6
8	0.008-0.004	0.38	1.7	94.3
9	0.004-0.002	0.33	1.5	95.8
>9	< 0.002	0.92	4.2	100.0
		total wt	coarse wt	fine wt
		21.9	18.2	3.6
		% sand	% silt	% clay
		83.36	10.97	5.67

  
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**Particle Size  
 Plumb, 1981  
 (%)  
 FL-3**

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MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

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SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.09	0.5	0.5
1	1-0.5	0.11	0.6	1.0
2	0.5-0.25	0.52	2.6	3.6
3	0.25-0.125	8.07	40.6	44.2
4	0.125-0.062	3.61	18.2	62.4
5	0.062-0.031	2.45	12.3	74.7
6	0.031-0.016	0.72	3.6	78.3
7	0.016-0.008	1.21	6.1	84.4
8	0.008-0.004	0.74	3.7	88.1
9	0.004-0.002	0.50	2.5	90.7
>9	< 0.002	1.86	9.3	100.0
		total wt	coarse wt	fine wt
		19.9	12.4	7.5
		% sand	% silt	% clay
		62.41	25.74	11.85

*Philip D. Carpenter*  
 Laboratory Director

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San Francisco Army Corps of Engineers  
Humboldt Maintenance Dredge Sediments  
T-10774  
C-104

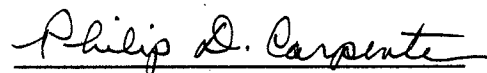
**Particle Size  
Plumb, 1981  
(%)  
FL-4**

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MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows:

---

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.04	0.2	0.2
1	1-0.5	0.10	0.5	0.8
2	0.5-0.25	0.57	3.1	3.8
3	0.25-0.125	4.01	21.7	25.5
4	0.125-0.062	3.21	17.4	42.9
5	0.062-0.031	2.54	13.7	56.6
6	0.031-0.016	1.79	9.7	66.3
7	0.016-0.008	1.94	10.5	76.8
8	0.008-0.004	1.10	5.9	82.7
9	0.004-0.002	0.76	4.1	86.8
>9	< 0.002	2.45	13.2	100.0
		total wt	coarse wt	fine wt
		18.5	7.9	10.6
		% sand	% silt	% clay
		42.90	39.79	17.31

  
Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 FL-1**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.05	0.3	0.3
1	1-0.5	0.07	0.4	0.6
2	0.5-0.25	0.20	1.1	1.7
3	0.25-0.125	2.24	12.0	13.7
4	0.125-0.062	3.18	17.0	30.7
5	0.062-0.031	3.36	17.9	48.6
6	0.031-0.016	2.33	12.4	61.0
7	0.016-0.008	2.03	10.8	71.8
8	0.008-0.004	1.16	6.2	78.0
9	0.004-0.002	1.16	6.2	84.2
>9	< 0.002	2.96	15.8	100.0
		total wt	coarse wt	fine wt
		18.7	5.7	13.0
		% sand	% silt	% clay
		30.65	47.37	21.98

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 FL-2**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.09	0.6	0.6
1	1-0.5	0.09	0.6	1.1
2	0.5-0.25	0.08	0.5	1.7
3	0.25-0.125	0.16	1.0	2.7
4	0.125-0.062	0.85	5.4	8.1
5	0.062-0.031	3.22	20.5	28.6
6	0.031-0.016	2.44	15.5	44.2
7	0.016-0.008	2.42	15.4	59.6
8	0.008-0.004	1.72	11.0	70.6
9	0.004-0.002	1.11	7.1	77.7
>9	< 0.002	3.50	22.3	100.0
		total wt	coarse wt	fine wt
		15.7	1.3	14.4
		% sand	% silt	% clay
		8.11	62.50	29.40

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 Laboratory Director

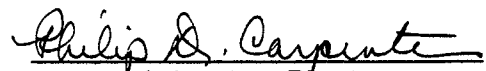
**Particle Size  
Plumb, 1981  
(%)  
FL-Composite**

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MATERIAL: Sediment samples received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows:

---

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.04	0.2	0.2
1	1-0.5	0.08	0.4	0.7
2	0.5-0.25	0.33	1.8	2.4
3	0.25-0.125	4.21	22.9	25.4
4	0.125-0.062	2.98	16.2	41.6
5	0.062-0.031	2.86	15.5	57.1
6	0.031-0.016	1.87	10.2	67.3
7	0.016-0.008	1.46	7.9	75.3
8	0.008-0.004	1.05	5.7	80.9
9	0.004-0.002	0.86	4.7	85.6
>9	< 0.002	2.64	14.4	100.0
		total wt	coarse wt	fine wt
		18.4	7.6	10.7
		% sand	% silt	% clay
		41.59	39.36	19.05

  
Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 Reference Composite**

MATERIAL: Sediment samples received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.00	0.0	0.0
1	1-0.5	0.03	0.1	0.1
2	0.5-0.25	0.06	0.3	0.4
3	0.25-0.125	1.43	6.4	6.8
4	0.125-0.062	11.67	52.5	59.3
5	0.062-0.031	3.78	17.0	76.3
6	0.031-0.016	1.98	8.9	85.2
7	0.016-0.008	1.14	5.1	90.3
8	0.008-0.004	0.62	2.8	93.1
9	0.004-0.002	0.36	1.6	94.7
>9	< 0.002	1.18	5.3	100.0
		total wt	coarse wt	fine wt
		22.2	13.2	9.1
		% sand	% silt	% clay
		59.29	33.78	6.92

*Philip D. Carpenter*  
 Laboratory Director



**Particle Size  
 Plumb, 1981  
 (%)  
 Home Sediment**

**MATERIAL:** Sediment samples received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.02	0.1	0.1
1	1-0.5	0.32	1.0	1.0
2	0.5-0.25	15.98	47.9	48.9
3	0.25-0.125	15.88	47.6	96.6
4	0.125-0.062	0.44	1.3	97.9
5	0.062-0.031	0.12	0.4	98.3
6	0.031-0.016	0.00	0.0	98.3
7	0.016-0.008	0.16	0.6	98.9
8	0.008-0.004	0.08	0.3	99.1
9	0.004-0.002	0.06	0.2	99.3
>9	< 0.002	0.30	1.1	100.0
		total wt	coarse wt	fine wt
		33.4	32.6	0.7
		% sand	% silt	% clay
		97.87	1.27	1.27

*Philip D. Carpenter*  
 Laboratory Director

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San Francisco Army Corps of Engineers  
Humboldt Maintenance Dredge Sediments  
T-10774  
C-110


**Particle Size  
Plumb, 1981  
(%)  
NB-10**

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MATERIAL: Sediment sample received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows:

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SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	4.97	18.7	18.7
-1	4- 2	4.69	17.6	36.3
0	2- 1	2.12	8.0	44.3
-1	1-0.5	2.95	11.1	55.4
2	0.5-0.25	7.99	30.1	85.5
3	0.25-0.125	3.02	11.4	96.8
4	0.125-0.062	0.19	0.7	97.6
5	0.062-0.031	0.12	0.4	98.0
6	0.031-0.016	0.08	0.3	98.3
7	0.016-0.008	0.07	0.2	98.5
8	0.008-0.004	0.04	0.1	98.7
9	0.004-0.002	0.05	0.2	98.9
>9	< 0.002	0.31	1.1	100.0
		total wt	coarse wt	fine wt
		26.6	25.9	0.7
		% sand	% silt	% clay
		78.86	1.11	1.34

  
Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 NB-9**

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MATERIAL: Sediment sample received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

---

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.52	1.9	1.9
-1	4- 2	3.26	12.1	14.0
0	2- 1	4.86	18.0	32.0
1	1-0.5	9.57	35.5	67.5
2	0.5-0.25	8.17	30.3	97.8
3	0.25-0.125	0.55	2.0	99.9
4	0.125-0.062	0.04	0.1	100.0
5	0.062-0.031	0.00	0.0	100.0
6	0.031-0.016	0.00	0.0	100.0
7	0.016-0.008	0.00	0.0	100.0
8	0.008-0.004	0.00	0.0	100.0
9	0.004-0.002	0.00	0.0	100.0
>9	< 0.002	0.00	0.0	100.0
		total wt	coarse wt	fine wt
		27.0	27.0	0.0
		% sand	% silt	% clay
		98.07	0.00	0.00

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 Laboratory Director

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San Francisco Army Corps of Engineers  
Humboldt Maintenance Dredge Sediments  
T-10774  
C-112

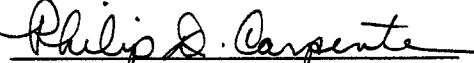
**Particle Size  
Plumb, 1981  
(%)  
NB-8**

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MATERIAL: Sediment sample received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows:

---

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	2.70	10.8	10.8
-1	4- 2	5.00	20.0	30.8
0	2- 1	3.16	12.7	43.5
1	1-0.5	3.58	14.3	57.8
2	0.5-0.25	6.67	26.7	84.5
3	0.25-0.125	2.18	8.7	93.3
4	0.125-0.062	0.19	0.8	94.0
5	0.062-0.031	0.28	1.1	95.1
6	0.031-0.016	0.30	1.2	96.3
7	0.016-0.008	0.23	0.9	97.2
8	0.008-0.004	0.15	0.6	97.8
9	0.004-0.002	0.13	0.5	98.3
>9	< 0.002	0.43	1.7	100.0
		total wt	coarse wt	fine wt
		25.0	23.5	1.5
		% sand	% silt	% clay
		83.20	3.76	2.22

  
Laboratory Director

**Particle Size  
Plumb, 1981  
(%)  
NB-7**

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MATERIAL: Sediment sample received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows:

---

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	3.30	12.4	12.4
-1	4- 2	1.54	5.8	18.2
0	2- 1	0.97	3.7	21.9
1	1-0.5	2.03	7.7	29.6
2	0.5-0.25	12.85	48.4	78.0
3	0.25-0.125	5.26	19.8	97.8
4	0.125-0.062	0.09	0.3	98.2
5	0.062-0.031	0.10	0.4	98.5
6	0.031-0.016	0.06	0.2	98.7
7	0.016-0.008	0.06	0.2	98.9
8	0.008-0.004	0.03	0.1	99.1
9	0.004-0.002	0.01	0.0	99.1
>9	< 0.002	0.24	0.9	100.0
		total wt	coarse wt	fine wt
		26.5	26.0	0.5
		% sand	% silt	% clay
		85.71	0.90	0.94

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Laboratory Director

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San Francisco Army Corps of Engineers  
Humboldt Maintenance Dredge Sediments  
T-10774  
C-114

**Particle Size  
Plumb, 1981  
(%)  
NB-6**

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MATERIAL: Sediment sample received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows:

---

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	3.74	14.9	14.9
-1	4- 2	3.23	12.9	27.8
0	2- 1	1.15	4.6	32.4
1	1-0.5	1.27	5.1	37.5
2	0.5-0.25	7.50	29.9	67.4
3	0.25-0.125	7.05	28.1	95.5
4	0.125-0.062	0.62	2.5	98.0
5	0.062-0.031	0.11	0.4	98.4
6	0.031-0.016	0.08	0.3	98.7
7	0.016-0.008	0.07	0.3	99.0
8	0.008-0.004	0.01	0.0	99.0
9	0.004-0.002	0.02	0.1	99.1
>9	< 0.002	0.24	0.9	100.0
		total wt	coarse wt	fine wt
		25.1	24.6	0.5
		% sand	% silt	% clay
		83.05	1.04	1.00

  
Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 NB-5**

**MATERIAL:** Sediment sample received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.86	3.7	3.7
-1	4- 2	0.61	2.6	6.2
0	2- 1	1.48	6.3	12.5
1	1-0.5	8.07	34.3	46.8
2	0.5-0.25	10.81	45.9	92.7
3	0.25-0.125	1.66	7.0	99.7
4	0.125-0.062	0.07	0.3	100.0
5	0.062-0.031	0.00	0.0	100.0
6	0.031-0.016	0.00	0.0	100.0
7	0.016-0.008	0.00	0.0	100.0
8	0.008-0.004	0.00	0.0	100.0
9	0.004-0.002	0.00	0.0	100.0
>9	< 0.002	0.00	0.0	100.0
		total wt	coarse wt	fine wt
		23.6	23.6	0.0
		% sand	% silt	% clay
		96.35	0.00	0.00

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 NB-3**

MATERIAL: Sediment sample received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.53	2.3	2.3
-1	4- 2	0.33	1.4	3.7
0	2- 1	1.67	7.3	11.0
1	1-0.5	6.58	28.6	39.6
2	0.5-0.25	12.09	52.5	92.1
3	0.25-0.125	1.81	7.9	99.9
4	0.125-0.062	0.02	0.1	100.0
5	0.062-0.031	0.00	0.0	100.0
6	0.031-0.016	0.00	0.0	100.0
7	0.016-0.008	0.00	0.0	100.0
8	0.008-0.004	0.00	0.0	100.0
9	0.004-0.002	0.00	0.0	100.0
>9	< 0.002	0.00	0.0	100.0
		total wt	coarse wt	fine wt
		23.0	23.0	0.0
		% sand	% silt	% clay
		97.70	0.00	0.00

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 Laboratory Director



**Particle Size  
 Plumb, 1981  
 (%)  
 NB-1**

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MATERIAL: Sediment sample received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

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SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.03	0.1	0.1
0	2- 1	0.06	0.2	0.4
1	1-0.5	0.31	1.2	1.6
2	0.5-0.25	17.24	68.3	69.8
3	0.25-0.125	7.48	29.6	99.4
4	0.125-0.062	0.14	0.6	100.0
5	0.062-0.031	0.00	0.0	100.0
6	0.031-0.016	0.00	0.0	100.0
7	0.016-0.008	0.00	0.0	100.0
8	0.008-0.004	0.00	0.0	100.0
9	0.004-0.002	0.00	0.0	100.0
>9	< 0.002	0.00	0.0	100.0
		total wt	coarse wt	fine wt
		25.3	25.3	0.0
		% sand	% silt	% clay
		100.00	0.00	0.00

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 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 NB 4 Rep 2**

MATERIAL: Sediment sample received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.06	0.3	0.3
0	2- 1	0.16	0.7	0.9
1	1-0.5	1.02	4.4	5.3
2	0.5-0.25	11.93	51.1	56.4
3	0.25-0.125	9.84	42.2	98.6
4	0.125-0.062	0.33	1.4	100.0
5	0.062-0.031	0.00	0.0	100.0
6	0.031-0.016	0.00	0.0	100.0
7	0.016-0.008	0.00	0.0	100.0
8	0.008-0.004	0.00	0.0	100.0
9	0.004-0.002	0.00	0.0	100.0
>9	< 0.002	0.00	0.0	100.0
		total wt	coarse wt	fine wt
		23.3	23.3	0.0
		% sand	% silt	% clay
		100.00	0.00	0.00

*Philip D. Carpenter*  
 Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 NB-2**

---

MATERIAL: Sediment sample received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

---

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.12	0.5	0.5
-1	4- 2	2.21	8.5	8.9
0	2- 1	0.99	3.8	12.7
1	1-0.5	2.27	8.7	21.4
2	0.5-0.25	17.95	68.7	90.1
3	0.25-0.125	2.56	9.8	99.9
4	0.125-0.062	0.02	0.1	100.0
5	0.062-0.031	0.00	0.0	100.0
6	0.031-0.016	0.00	0.0	100.0
7	0.016-0.008	0.00	0.0	100.0
8	0.008-0.004	0.00	0.0	100.0
9	0.004-0.002	0.00	0.0	100.0
>9	< 0.002	0.00	0.0	100.0
		total wt	coarse wt	fine wt
		26.1	26.1	0.0
		% sand	% silt	% clay
		99.54	0.00	0.00

*Philip D. Carpenter*  
 Laboratory Director

**ToxScan, Inc.**  
Environmental Chemistry and Bioassay Laboratories  
Tel: 408/724-4522 FAX: 408/724-3188

San Francisco Army Corps of Engineers  
Humboldt Maintenance Dredge Sediments  
T-10774  
C-120

**Particle Size  
Plumb, 1981  
(%)  
BAR**

---

MATERIAL: Sediment sample received April 4-6, 1994  
IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
TOXSCAN NUMBER: T-10774  
REPORT: Quantitative chemical analysis is as follows:

---

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.19	0.8	0.8
-1	4- 2	0.41	1.6	2.4
0	2- 1	1.78	7.1	9.4
1	1-0.5	6.84	27.1	36.6
2	0.5-0.25	11.63	46.1	82.7
3	0.25-0.125	4.34	17.2	99.9
4	0.125-0.062	0.02	0.1	100.0
5	0.062-0.031	0.00	0.0	100.0
6	0.031-0.016	0.00	0.0	100.0
7	0.016-0.008	0.00	0.0	100.0
8	0.008-0.004	0.00	0.0	100.0
9	0.004-0.002	0.00	0.0	100.0
>9	< 0.002	0.00	0.0	100.0
		total wt	coarse wt	fine wt
		25.2	25.2	0.0
		% sand	% silt	% clay
		99.25	0.00	0.00

  
Laboratory Director

**Particle Size  
 Plumb, 1981  
 (%)  
 ENT 2**

**MATERIAL:** Sediment sample received April 4-6, 1994  
**IDENTIFICATION:** Humboldt Maintenance Dredge Sediments  
**TOXSCAN NUMBER:** T-10774  
**REPORT:** Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.09	0.4	0.4
-1	4- 2	0.38	1.6	2.0
0	2- 1	1.32	5.6	7.5
1	1-0.5	4.78	20.1	27.7
2	0.5-0.25	15.43	65.0	92.7
3	0.25-0.125	1.73	7.3	100.0
4	0.125-0.062	0.00	0.0	100.0
5	0.062-0.031	0.00	0.0	100.0
6	0.031-0.016	0.00	0.0	100.0
7	0.016-0.008	0.00	0.0	100.0
8	0.008-0.004	0.00	0.0	100.0
9	0.004-0.002	0.00	0.0	100.0
>9	< 0.002	0.00	0.0	100.0
		total wt	coarse wt	fine wt
		23.7	23.7	0.0
		% sand	% silt	% clay
		99.62	0.00	0.00

*Philip D. Carpenter*  
 Laboratory Director

**ToxScan, Inc.**  
 Environmental Chemistry and Bioassay Laboratories  
 Tel: 408/724-4522 FAX: 408/724-3188

San Francisco Army Corps of Engineers  
 Humboldt Maintenance Dredge Sediments  
 T-10774  
 C-122

**Particle Size  
 Plumb, 1981  
 (%)  
 ENT 1**

MATERIAL: Sediment sample received April 4-6, 1994  
 IDENTIFICATION: Humboldt Maintenance Dredge Sediments  
 TOXSCAN NUMBER: T-10774  
 REPORT: Quantitative chemical analysis is as follows:

SIZE INTERVAL		INTERVAL WT	INTERVAL %	CUMULATIVE %
Phi	mm			
<-5	>32	0.00	0.0	0.0
-4	32-16	0.00	0.0	0.0
-3	16- 8	0.00	0.0	0.0
-2	8- 4	0.00	0.0	0.0
-1	4- 2	0.00	0.0	0.0
0	2- 1	0.04	0.2	0.2
1	1-0.5	0.12	0.5	0.7
2	0.5-0.25	10.88	46.6	47.3
3	0.25-0.125	12.18	52.2	99.5
4	0.125-0.062	0.11	0.5	100.0
5	0.062-0.031	0.00	0.0	100.0
6	0.031-0.016	0.00	0.0	100.0
7	0.016-0.008	0.00	0.0	100.0
8	0.008-0.004	0.00	0.0	100.0
9	0.004-0.002	0.00	0.0	100.0
>9	< 0.002	0.00	0.0	100.0
		total wt	coarse wt	fine wt
		23.3	23.3	0.0
		% sand	% silt	% clay
		100.00	0.00	0.00

*Philip D. Carpenter*  
 Laboratory Director

## Appendix C-1

### Dioxin Analyses Results (Alta Analytical Laboratory, Inc.)

**Please note:** The composite sample labels in this appendix are equivalent to the composite sample labels referenced in other sections and appendices of this report, as follows:

EK COMPOSITE = EKUP

SAM6 COMPOSITE = SAMTB

FL COMPOSITE = FLTB

REFERENCE COMPOSITE = REF







April 21, 1994

**Alta Batch I.D.: 13353**

Ms. Mary Lou Milazzo  
Toxscan, Inc.  
42 Hanger Way  
Watsonville, CA 95076

Dear Ms. Milazzo,

Enclosed are the results for the four sediment samples received at Alta Analytical Laboratory on April 8, 1994. This work was authorized under your Purchase Order #09680. The samples were analyzed using EPA Method 8290 for 2,3,7,8-TCDD and 2,3,7,8-TCDF. Routine turnaround time was requested for this work.

The following report consists of a Sample Inventory (Section I), Analytical Results (Section II) and the Appendix. The Appendix contains a copy of the chain-of-custody, a list of data qualifiers and abbreviations and copies of the raw data (if requested).

If you have any questions regarding this report please feel free to contact me.

Sincerely,

  
William J. Luksemburg  
Director of HRMS Services

**Alta Analytical Laboratory Inc.**

5070 Robert J. Mathews Parkway  
El Dorado Hills, CA 95762

FAX (916) 933-0940  
(916) 933-1640



Section I. Sample Inventory

Date Received: 8-Apr-94

Alta Lab ID.

Client ID.

---

13353-1-SA  
13353-2-SA  
13353-3-SA  
13353-4-SA

---

T-10774-32 SAM 6 COMPOSITE  
T-10774-33 REFERENCE COMPOSITE  
T-10774-34 FL COMPOSITE  
T-10774-35 EK COMPOSITE

**TCDD & TCDF  
EPA METHOD 8290**

**LCS RESULTS**

Lab ID: 13353-LCS1/LCS2

Matrix: Sediment

Date Received: NA

Date Extracted: 4/4/94

Sample Amount: 10.00 g

ICAL ID: I551

QC Lot: LC0404S

Units: NA

<u>Compound</u>	<u>LCS1 % R</u>	<u>LCS2 % R</u>	<u>RPD %</u>
2,3,7,8-TCDD	95	97	2.1
2,3,7,8-TCDF	92	88	4.4

**Isotopic Recovery Results**

<u>Internal Standard:</u>	<u>LCS1 % R</u>	<u>LCS2 % R</u>
<sup>13</sup> C-2,3,7,8-TCDD	87	96
<sup>13</sup> C-2,3,7,8-TCDF	87	95

**Clean-up Recovery Standard:**

<sup>37</sup> Cl-2,3,7,8-TCDD	96	98
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**Dates Analyzed:**

DB-5: 4/5/94

Analyst: JMS

Reviewer: WJZ

A

1. The first part of the document discusses the importance of maintaining accurate records of all transactions.

2. It is essential to ensure that all data is entered correctly and consistently.

3. Regular audits should be conducted to verify the accuracy of the information.

4. The final section outlines the procedures for handling discrepancies and errors.

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20.

**SECTION II.**

**TCDD & TCDF  
EPA METHOD 8290**

**METHOD BLANK**  
**Lab ID: 13353-001-MB**  
**Matrix: Sediment**

Date Received: NA  
 Date Extracted: 4/12/94  
 Sample Amount: 10.00 g

ICAL ID: I551  
 QC Lot: LC0404S  
 Units: pg/g

<u>Compound</u>	<u>Conc.</u>	<u>D.L.</u>	<u>Ratio</u>	<u>S/N Ratio</u>	<u>Qualifier</u>
2,3,7,8-TCDD	ND	0.26			
Total TCDD	ND	0.26			
2,3,7,8-TCDF	ND	0.047			
Total TCDF	ND	0.047			

**Isotopic Recovery Results**

<u>Internal Standard:</u>	<u>% R</u>	<u>Ratio</u>	<u>Qualifier</u>
<sup>13</sup> C-2,3,7,8-TCDD	95	0.78	
<sup>13</sup> C-2,3,7,8-TCDF	95	0.83	

**Clean-up Recovery Standard:**

<sup>37</sup> Cl-2,3,7,8-TCDD	101	NA	
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**Dates Analyzed:**

DB-5: 4/15/94

DB-225: NA

Analyst: Liby

Page 1 of 1

Reviewer: 



**TCDD & TCDF  
EPA METHOD 8290**

**Sample ID:** T-10774-32  
**Lab ID:** 13353-001-SA  
**Matrix:** Sediment  
**% Solid:** 67

**Date Received:** 4/8/94  
**Date Extracted:** 4/12/94  
**Sample Amount:** 10.18 g

**ICAL ID:** I551  
**QC Lot:** LC0404S  
**Units:** pg/g

<u>Compound</u>	<u>Conc.</u>	<u>D.L.</u>	<u>Ratio</u>	<u>S/N Ratio</u>	<u>Qualifier</u>
2,3,7,8-TCDD	ND	0.25			
Total TCDD	ND	0.25			
2,3,7,8-TCDF	0.51		0.86	> 10:1	
Total TCDF	0.94		0.81	> 10:1	

**Isotopic Recovery Results**

<u>Internal Standard:</u>	<u>% R</u>	<u>Ratio</u>	<u>Qualifier</u>
<sup>13</sup> C-2,3,7,8-TCDD	95	0.78	
<sup>13</sup> C-2,3,7,8-TCDF	100	0.83	

**Clean-up Recovery Standard:**

<sup>37</sup> Cl-2,3,7,8-TCDD	99	NA	
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**Dates Analyzed:**

**DB-5:** 4/15/94

**DB-225:** 4/20/94

**Analyst:** 674

Page 1 of 1

**Reviewer:** 8

**TCDD & TCDF  
EPA METHOD 8290**

**Sample ID:** T-10774-33  
**Lab ID:** 13353-002-SA  
**Matrix:** Sediment  
**% Solid:** 77

**Date Received:** 4/8/94  
**Date Extracted:** 4/12/94  
**Sample Amount:** 10.10 g

**ICAL ID:** I551  
**QC Lot:** LC0404S  
**Units:** pg/g

<u>Compound</u>	<u>Conc.</u>	<u>D.L.</u>	<u>Ratio</u>	<u>S/N Ratio</u>	<u>Qualifier</u>
2,3,7,8-TCDD	ND	0.22			
Total TCDD	ND	0.22			
2,3,7,8-TCDF	ND	0.13			
Total TCDF	ND	0.13			

**Isotopic Recovery Results**

<u>Internal Standard:</u>	<u>% R</u>	<u>Ratio</u>	<u>Qualifier</u>
<sup>13</sup> C-2,3,7,8-TCDD	96	0.79	
<sup>13</sup> C-2,3,7,8-TCDF	97	0.83	
<u>Clean-up Recovery Standard:</u>			
<sup>37</sup> Cl-2,3,7,8-TCDD	97	NA	

**Dates Analyzed:**

DB-5: 4/15/94

DB-225: NA

Analyst: Sty

Page 1 of 1

Reviewer: B





**TCDD & TCDF  
EPA METHOD 8290**

**Sample ID:** T-10774-34  
**Lab ID:** 13353-003-SA  
**Matrix:** Sediment  
**% Solid:** 67

**Date Received:** 4/8/94  
**Date Extracted:** 4/12/94  
**Sample Amount:** 10.13 g

**ICAL ID:** I551  
**QC Lot:** LC0404S  
**Units:** pg/g

<u>Compound</u>	<u>Conc.</u>	<u>D.L.</u>	<u>Ratio</u>	<u>S/N Ratio</u>	<u>Qualifier</u>
2,3,7,8-TCDD	ND	0.33			
Total TCDD	ND	0.33			
2,3,7,8-TCDF	ND	0.44			
Total TCDF	ND	0.44			

**Isotopic Recovery Results**

<u>Internal Standard:</u>	<u>% R</u>	<u>Ratio</u>	<u>Qualifier</u>
<sup>13</sup> C-2,3,7,8-TCDD	89	0.77	
<sup>13</sup> C-2,3,7,8-TCDF	97	0.81	

**Clean-up Recovery Standard:**

<sup>37</sup> Cl-2,3,7,8-TCDD	95	NA	
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**Dates Analyzed:**

**DB-5:** 4/15/94

**DB-225:** 4/20/94

**Analyst:** Bay

Page 1 of 1

**Reviewer:** B



**TCDD & TCDF  
EPA METHOD 8290**

Sample ID: T-10774-35  
Lab ID: 13353-004-SA  
Matrix: Sediment  
% Solid: 76

Date Received: 4/8/94  
Date Extracted: 4/12/94  
Sample Amount: 10.20 g

ICAL ID: I551  
QC Lot: LC0404S  
Units: pg/g

<u>Compound</u>	<u>Conc.</u>	<u>D.L.</u>	<u>Ratio</u>	<u>S/N Ratio</u>	<u>Qualifier</u>
2,3,7,8-TCDD	ND	0.30			
Total TCDD	ND	0.30			
2,3,7,8-TCDF	ND	0.45			
Total TCDF	ND	0.45			

**Isotopic Recovery Results**

<u>Internal Standard:</u>	<u>% R</u>	<u>Ratio</u>	<u>Qualifier</u>
<sup>13</sup> C-2,3,7,8-TCDD	94	0.78	
<sup>13</sup> C-2,3,7,8-TCDF	99	0.83	
<u>Clean-up Recovery Standard:</u>			
<sup>37</sup> Cl-2,3,7,8-TCDD	95	NA	

**Dates Analyzed:**

DB-5: 4/15/94

DB-225: 4/20/94

Analyst: SLY

Page 1 of 1

Reviewer: SL

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**APPENDIX**



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## DATA QUALIFIERS & ABBREVIATIONS

<b>A</b>	<b>The amount detected is below the Method Calibration Limit.</b>
<b>B</b>	<b>This compound was also detected in the blank.</b>
<b>C</b>	<b>The amount detected is less than five times the Method Quantitation Limit.</b>
<b>D</b>	<b>The amount reported is the maximum possible concentration.</b>
<b>E</b>	<b>The detection limit was raised above the Method Quantitation Limit due to chemical interferences.</b>
<b>F</b>	<b>This result has been confirmed on a DB-225 column.</b>
<b>G</b>	<b>This result has been confirmed on a SP-2331 column.</b>
<b>H</b>	<b>The signal-to-noise ratio is greater than 10:1.</b>
<b>I</b>	<b>Chemical Interference</b>
<b>Conc.</b>	<b>Concentration</b>
<b>D.L.</b>	<b>Detection Limit</b>
<b>NA</b>	<b>Not applicable</b>
<b>S/N</b>	<b>Signal-to-noise</b>
<b>*</b>	<b>See Cover Letter</b>
<b>ND</b>	<b>Not Detected</b>
<b>MPC</b>	<b>Maximum Possible Concentration</b>

**COMPANY NAME:** ToxScan Inc.  
**ATTN:** Mary Lou Wilczko  
**ADDRESS:**  
**PHONE:** 408 / 724-4522  
**FAX:** 408 / 724-3188  
**PROJECT NAME:** Humboldt Harbor  
**PROJECT NUMBER:**  
**SEND INVOICE TO:**  
**P.O. / CONTRACT NO:** 09680

**COMMENTS / SPECIAL INSTRUCTIONS:**  
 DL = 1 part per trillion  
 TAT = 3 weeks (by 29 April '94)  
 QC = 1 LCS, 1 blank

**TOXSCAN INC.**  
 42 Hangar Way  
 Watsonville, CA 95076  
**PHONE:** 408/724-4522  
**FAX:** 408/724-3188

LAB USE ONLY:	ANALYSIS(ES) REQUESTED
STORAGE LOCATION	
FREEZER #	
REFRIGERATOR #	
SHELF #	
SAMPLE CONDITION:	

Lab Use Only: ID Number	Client Sample Identification	Sample Information:		Bottle or Container Information:		No. of Bottles:
		Sampling Date	Sampling Time	Sample Matrix	Sample Preservative	
T-10774-32	SAMPLE Composite	04/01/94		sediment		1
T-10774-33	Reference Composite	04/05/94		↓		1
T-10774-34	FLU Composite	04/04/94		↓		1
T-10774-35	EK Composite	04/02/94		↓		1

**SAMPLER'S SIGNATURE AND PRINTED NAME:**  
 Sub contract Laboratory: Altra Analytical 916-933-1640

RELINQUISHED BY (SIGNATURE AND PRINTED NAME): Mary Lou Wilczko	RECEIVED BY (SIGNATURE AND PRINTED NAME): ALTA Ken Ficht	DATE: 04/07/94	TIME: 16:05
---	--	-------------------	----------------

Please return completed copy of COC to ToxScan. Prankya.

ALTA Analytical Laboratory

Batch ID: 13353

Sample Log-In Checklist		Yes	No
1. Samples Arrived by: <u>UPS Red</u>			
2. Airbill Present? Number <u>01131576516</u>		X	
3. Shipping Container is Intact?		X	
4. Custody Seals Present? Number _____			X
If yes, are they intact? <u>N/A</u>			
5. Sample Containers Intact?		X	
6. Shipping Preservation: <u>Ice/Blue Ice/None</u>			
7. Temperature: <u>18°</u>			
8. Chain of Custody Present?		X	
9. Discrepancies in Chain of Custody?			X
10. Packing Retained?		X	

Name: [Signature] Date Rcv'd: 4-8-94

Comments:



May 11, 1994

**Alta Batch I.D.: 13413**

Ms. Mary Lou Milazzo  
ToxScan Inc.  
42 Hanger Way  
Watsonville, CA 95076

Dear Ms. Milazzo,

Enclosed are the results for the one sediment sample received at Alta Analytical Laboratory on April 29, 1994. The work was authorized under your Purchase Order #09723. These samples were analyzed using EPA Method 8290 for 2,3,7,8-TCDD and 2,3,7,8-TCDF. Routine turnaround time was provided for these samples.

The following report consists of a Sample Inventory (Section I), Analytical Results (Section II) and the Appendix. The Appendix contains a copy of the chain-of-custody, a list of data qualifiers and abbreviations and copies of the raw data (if requested).

If you have any questions regarding this report please feel free to contact me.

Sincerely,

A handwritten signature in black ink, appearing to read "William J. Luksemburg".

William J. Luksemburg  
Director of HRMS Services

**Alta Analytical Laboratory Inc.**

5070 Robert J. Mathews Parkway  
El Dorado Hills, CA 95762

FAX (916) 933-0940  
(916) 933-1640





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**Section I. Sample Inventory**

**Date Received: 29-Apr-94**

**Alta Lab ID.**

**Client ID.**

---

**13413-1-SA**

---

**T-10774-89 CONTROL SEDIMENT**

**TCDD & TCDF  
EPA METHOD 8290**

**LCS RESULTS**

Lab ID: 13413-LCS1/LCS2

Matrix: Sediment

Date Received: NA

Date Extracted: 5/6/94

Sample Amount: 10.00 g

ICAL ID: I551

QC Lot: LC0506S

Units: NA

<u>Compound</u>	<u>LCS1 % R</u>	<u>LCS2 % R</u>	<u>RPD %</u>
2,3,7,8-TCDD	101	106	4.8
2,3,7,8-TCDF	95	117	21

**Isotopic Recovery Results**


<u>Internal Standard:</u>	<u>LCS1 % R</u>	<u>LCS2 % R</u>
<sup>13</sup> C-2,3,7,8-TCDD	87	107
<sup>13</sup> C-2,3,7,8-TCDF	83	96

**Clean-up Recovery Standard:**

<sup>37</sup> Cl-2,3,7,8-TCDD	93	118
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**Dates Analyzed:**

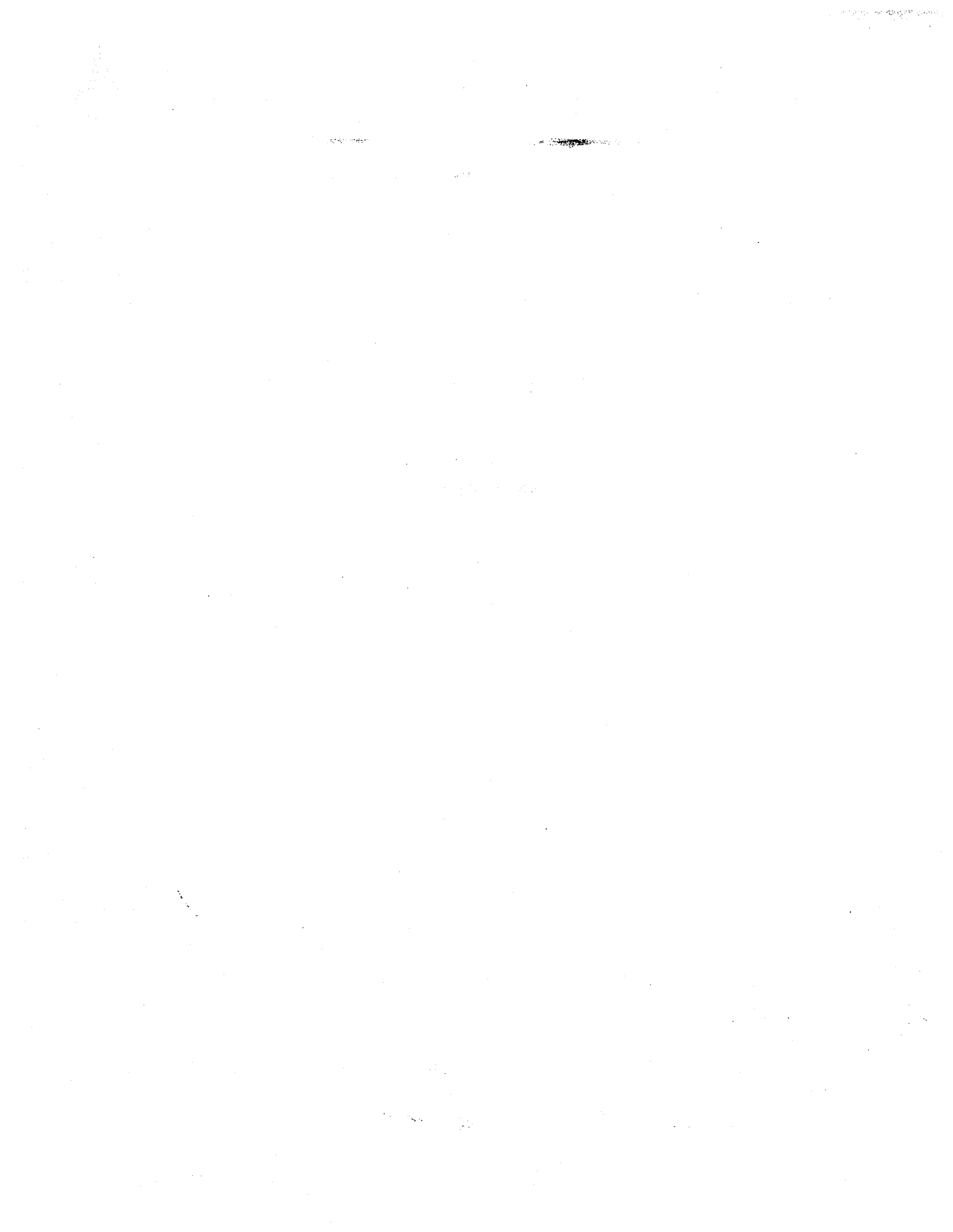
DB-5: 5/10/94

Analyst: 

Reviewer: 

---

**SECTION II.**





**TCDD & TCDF  
EPA METHOD 8290**

**METHOD BLANK**  
**Lab ID: 13413-001-MB**  
**Matrix: Sediment**

**Date Received: NA**  
**Date Extracted: 5/6/94**  
**Sample Amount: 10.00 g**

**ICAL ID: I551**  
**QC Lot: LC0506S**  
**Units: pg/g**

<u>Compound</u>	<u>Conc.</u>	<u>D.L.</u>	<u>Ratio</u>	<u>S/N Ratio</u>	<u>Qualifier</u>
2,3,7,8-TCDD	ND	0.14			
Total TCDD	ND	0.14			
2,3,7,8-TCDF	ND	0.17			
Total TCDF	ND	0.17			

**Isotopic Recovery Results**

<u>Internal Standard:</u>	<u>% R</u>	<u>Ratio</u>	<u>Qualifier</u>
<sup>13</sup> C-2,3,7,8-TCDD	109	0.83	
<sup>13</sup> C-2,3,7,8-TCDF	52	0.83	

**Clean-up Recovery Standard:**

<sup>37</sup> Cl-2,3,7,8-TCDD	114	NA	
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**Dates Analyzed:**

**DB-5: 5/9/94**

**DB-225: NA**

**Analyst: SM**

**Page 1 of 1**

**Reviewer:**



**TCDD & TCDF  
EPA METHOD 8290**

Sample ID: T-10774-89  
Lab ID: 13413-001-SA  
Matrix: Sediment  
% Solid: 82

Date Received: 4/29/94  
Date Extracted: 5/6/94  
Sample Amount: 10.06 g

ICAL ID: I551  
QC Lot: LC0506S  
Units: pg/g

<u>Compound</u>	<u>Conc.</u>	<u>D.L.</u>	<u>Ratio</u>	<u>S/N Ratio</u>	<u>Qualifier</u>
2,3,7,8-TCDD	ND	0.25			
Total TCDD	ND	0.25			
2,3,7,8-TCDF	ND	0.20			
Total TCDF	ND	0.20			

**Isotopic Recovery Results**

<u>Internal Standard:</u>	<u>% R</u>	<u>Ratio</u>	<u>Qualifier</u>
<sup>13</sup> C-2,3,7,8-TCDD	103	0.83	
<sup>13</sup> C-2,3,7,8-TCDF	114	0.86	
<u>Clean-up Recovery Standard:</u>			
<sup>37</sup> Cl-2,3,7,8-TCDD	102	NA	

**Dates Analyzed:**

DB-5: 5/10/94

DB-225: NA

Analyst: Bin

Page 1 of 1

Reviewer: BJ

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## APPENDIX





## DATA QUALIFIERS & ABBREVIATIONS



<b>A</b>	<b>The amount detected is below the Method Calibration Limit.</b>
<b>B</b>	<b>This compound was also detected in the blank.</b>
<b>C</b>	<b>The amount detected is less than five times the Method Quantitation Limit.</b>
<b>D</b>	<b>The amount reported is the maximum possible concentration.</b>
<b>E</b>	<b>The detection limit was raised above the Method Quantitation Limit due to chemical interferences.</b>
<b>F</b>	<b>This result has been confirmed on a DB-225 column.</b>
<b>G</b>	<b>This result has been confirmed on a SP-2331 column.</b>
<b>H</b>	<b>The signal-to-noise ratio is greater than 10:1.</b>
<b>I</b>	<b>Chemical Interference</b>
<b>Conc.</b>	<b>Concentration</b>
<b>D.L.</b>	<b>Detection Limit</b>
<b>NA</b>	<b>Not applicable</b>
<b>S/N</b>	<b>Signal-to-noise</b>
<b>*</b>	<b>See Cover Letter</b>
<b>ND</b>	<b>Not Detected</b>
<b>MPC</b>	<b>Maximum Possible Concentration</b>

OXSCAN CHAIN-OF-CUSTODY DOCUMENT Sub-Contract C.O.C.

COMPANY NAME: Toxscan Inc  
 ATTN: Mary Lou Malaga  
 ADDRESS: 42 Hanger Way  
Watsonville Ca 95086  
 PHONE: (408) 724-4522  
 FAX:  
 PROJECT NAME: Humboldt C.O.C.  
 PROJECT NUMBER:  
 COMMENTS / SPECIAL INSTRUCTIONS:  
DL = 1 part per trillion  
TAT = 3 weeks 05/20/94  
QC = 1 LCS, 1 blank  
 SEND INVOICE TO:  
 P.O. / CONTRACT NO: 09723

TOXSCAN INC.  
 42 Hanger Way  
 Watsonville, CA 95076  
 PHONE: 408724-4522  
 FAX: 408724-3188  
 LAB USE ONLY:  
 STORAGE LOCATION:  
 FREEZER #:  
 REFRIGERATOR #:  
 SHELF #:  
 ANALYSIS(ES) REQUESTED:  
 CHECK THE APPROPRIATE BOX BELOW:  
 SAMPLE CONDITION:  
 TDDP/TCDF only by EPA 8250  
 X

Lab Use Only ID Number	Client Sample Identification	Sample Information:		Bottle or Container Information:			No. of Bottles:
		Sampling Date	Sampling Time	Sample Type	Sample Preservative	Bottle Type:	
J-10774-89	Control Sediment	09/25/94		Sediment	On Blue Ice G/S	1L	1

SAMPLER'S SIGNATURE AND PRINTED NAME:

RECEIVED BY SIGNATURE AND PRINTED NAME:  
Mary Lou Malaga  
 RECEIVED BY SIGNATURE AND PRINTED NAME:  
Mary Lou Malaga

DATE: 4-29-94  
 TIME: 1100

# ALTA Analytical Laboratory

Batch ID: 13413

Sample Log-In Checklist	Yes	No
1. Date Samples Arrived: <u>4-29-94</u> Initials: <u>R.F.</u>		
2. <u>Samples Arrived By:</u> (circle one) Airborne Express      Federal Express <u>UPS</u> Emery      Freezer Truck      Company Courier      Other _____		
3. Shipping Documentation Present? (circle one) Shipping Label <u>Airbill</u> Tracking Number <u>02515490112</u>	X	
4. Shipping Container(s) Intact? If no, describe condition below.	X	
5. Custody Seals Present and Intact? If not intact, describe condition below. No. of Seals _____ or Seal No. _____      Type:(circle) Bottle or Container		X
6. Sample Container Intact? If no, indicate sample condition below.	X	
7. Shipping Preservation: (circle one) Ice <u>Blue Ice</u> Dry Ice      None      Temp(°C) <u>17</u>		
8. Chain of Custody (COC) or other Sample Documentation Present?	X	
9. COC/Documentation Acceptable? If no, complete COC Anomaly Form.	X	
10. Shipping Container: (circle one each) ALTA or <u>Client</u> / <u>Return</u> or Retain		
11. Container and/or Bottles Requested?		X
*12. Drinking Water Sample? If yes, Acceptable Preservation? (circle) Y or N		X

\*Required for HRMS

Name: \_\_\_\_\_ Date Samples Reconciled: \_\_\_\_\_  
(Signature Required for LCMS Only)

Comments:

# THE HISTORY OF THE UNITED STATES

## CHAPTER I

The history of the United States is a story of a young nation that grew from a small group of colonies to a powerful world superpower. It is a story of struggle, of triumph, and of the pursuit of the American dream. The story begins in the early 17th century when European settlers first arrived on the eastern coast of North America. They came seeking new opportunities, religious freedom, and a better life. Over time, these colonies developed their own unique cultures and ways of life, but they remained loyal to the British crown. However, as the colonies grew in size and power, tensions between them and the British government increased. The British imposed taxes and regulations that the colonists felt were unfair and oppressive. This led to a series of protests and acts of defiance, culminating in the American Revolution. The revolution was a struggle for independence, and it was won in 1776 when the colonies declared their independence from Britain. The new nation was born, and it set out on a path of growth and expansion. It fought wars with Britain and Spain, and it emerged as a major power in the world. The United States continued to grow and expand westward, and it became a nation of immigrants. It was a nation of diversity, and it was a nation that was always looking for a better future. The history of the United States is a story of a nation that has overcome many challenges and has achieved many great things. It is a story of a nation that has inspired the world and has shown the way to a better future.

Appendix D

QA/QC Data Plan and Report



## QA/QC PLAN AND REPORT

### 1.0 Field Survey Procedures

Sediment samples were collected from prescribed locations in Humboldt Harbor by use of a vibra-core and Smith-Macintyre grab. The vibra-core consists of a vibrating aluminum head and a ten foot long aluminum core tube. The core tube is capped with a stainless steel cutting tip and a stainless steel core catcher. The vibra-core is lowered slowly into the sediment; the vibration allows entry into the sediment from the mudline to the sample depth. If a sample was not obtained on the first attempt, core attempts were repeated until a sample was secured. The Smith-Macintyre grab consists of a set of spring-loaded galvanized steel jaws, triggered by impact with the sediment surface.

The water sample used to prepare elutriates for bioassays was collected from the disposal site using a peristaltic pump with silicon and teflon hoses which had been precleaned with soap and nitric acid, and thoroughly rinsed with deionized (DI) water.

Horizontal positioning was accomplished by use of a Trimble Global Positioning System (GPS). Water depth was measured by use of a precision Fathometer, calibrated daily according to manufacture specification. Tidal height was extrapolated from tide tables.

Sediment samples were composited in a precleaned teflon-lined container. Samples for chemical analysis were placed into pre-cleaned glass jars and sealed with teflon-lined lids. Bioassay samples were placed into one-gallon, pre-cleaned polyethylene jars with poly screw closures. Immediately after collection and compositing, samples were stored in insulated coolers with ice. Upon arrival at the ToxScan laboratory in Watsonville, CA, sediments were stored in the 4°C room until analyzed. Holding times for chemical analysis are detailed below. None were exceeded during this study.

All sampling data are documented in the field log sheets included in this report.

### 2.0 Laboratory

Laboratory QA/QC procedures for this testing program were implemented as described in the ToxScan QA/QC program. Generic QA measures are described below in an excerpt from our written program. Table 5 presents a summary of instruments used in this study for bulk sediment analyses, along with methods and schedules for calibration, maintenance, precision/accuracy monitoring and record keeping.

All sediment samples were preserved by storage at 4°C in the dark. While EPA/COE protocol allows a 6-week holding period for dredged material sampled, certain of the methods require extraction and/or analysis within a shorter time period. These restricted holding times are listed below, and were adhered to in this program.

<u>Analyte</u>	<u>Maximum Holding Time</u>
Mercury	28 days
PAH's	extraction within 14 days
Chlorinated Hydrocarbons	extraction within 14 days
Sulfides	7 days
Petroleum Hydrocarbons	28 days
Organotins	28 days

As required by the Scope of Services for this program, the frequency of duplicate analyses and spiked sample analyses has been increased over our standard practice. For this study, 10% of the analyses have been duplicated and 20% of samples have been spiked.

Following is an excerpt from our QA/QC program which details the routine QA/QC measures followed in this program.

#### Procedures for Sample Receiving

The samples, accompanied by a chain of custody form are received by the sample control officer who follows the listed procedures for receiving a sample.

All sample containers are inspected to determine if any breakage or mishandling occurred and to determine that the proper container and preservatives have been used. The sample control officer will verify that sample labels match those on the chain of custody and that all samples listed are present. If a chain of custody does not exist and one is to be generated. See section below on Chain of Custody and Documentation.

The "log-in" process is initiated by giving each sample a discrete laboratory number which is entered on the chain of custody, in the log book and on the project sheet.

The proper paperwork (Sample Analysis Request Form or SARF) indicating analyses needed, detection limits, due dates, sample description and location, and necessary QA/QC is prepared and given to the appropriate analyst. The project manager receives the project sheet, which indicates analyses to be performed and due dates, along with a copy of the original SARF.



### Sample Identification Procedure

In order to maintain sample identity, the following scheme is used:  
T-0001-01, where T = ToxScan  
0001 is the group number assigned to the set of samples  
01 is the individual container number received.

### Chain of Custody and Documentation

A chain of custody is initiated prior to sampling or at the time of sample delivery is submitted by a walk-in client. This chain of custody accompanies all samples and is given to the sample control officer along with the samples. Samples are logged in and the chain of custody is kept with the original SARF. If samples are to be subcontracted to another laboratory, a photocopy of the original chain of custody is made and will accompany those samples.

### Source and Preparation of Standards

All primary standards are purchased in concentrated solutions or as pure substances and purchased in the highest purity available from reputable manufacturers or suppliers. Liquid stock solutions of concentrated standards are accompanied by a certification as to purity and concentration. All batch numbers, catalogue numbers, supplier and date of purchase are kept in the standards log book and updated as necessary.

Stock and working standards are prepared taking into account the stability and concentration of the analyte. Thus, some standards are prepared daily, others at less frequent intervals. Those standards that are light sensitive are stored in amber or like containers. If refrigeration will maximize the lifetime of the standards, they are stored at 4°C. Included on the standards container are date of preparation, concentration of solution analyte, and weight or volume used to prepare the standard if applicable. All standards are prepared with a high quality deionized or distilled water or with known purity solvents. A blank of all dilutants is checked to determine if any contamination has been introduced.

### Calibration Procedures and Methods of Analysis

All instrument calibration methods are related to known analyte concentrations. This requires a calibration curve be prepared for each analyte. Some instruments can be calibrated directly from known concentrations of a standard; others furnish data for construction of a three-point curve.

The analyst follows the procedures specified in the operational manual for each instrument as well as those guidelines set forth by operational standard methods: Standard Methods for the Evaluation of Waters and Wastewaters, EPA Protocol SW-846, AOAC Manual of Methodologies,

etc. Calibration of instrumental parameters is further checked against standard reference materials provided by the EPA or NBS with listings of certified values. The worksheets given to the analyst have pertinent areas for calibration data to be recorded from which calibration or standard curves can be obtained.

Once the instrument has been standardized, analyte concentrations are checked against the standard curve every 10 analyses to assure continued calibration.

Samples are prepared, analyzed and reported according to those standardized procedures specified by EPA, Standard Methods, AOAC, or other recognized, documented methodologies. Sample weights, preparation, aliquots taken, and calculations are recorded on the analysis sheet furnished for each parameter to be determined and recorded in ink.

#### Method Blanks and Duplicate and Spiked Samples

A method blank is the analysis of pure organic-free water, high purity solvent or clean sample matrix after being subjected to treatment specified by the method used. Method blanks are used on all analyses to verify, qualitatively, that no false positives will occur and quantitatively, that concentrations are accurate and do not reflect contamination. A method blank is analyzed at a minimum of once for each batch of samples or after every twentieth sample, whichever is more frequent.

Spiking concentrations are dependent upon the background levels in the original sample. When spiking for a scan analysis, nominal spiking levels are used as described by the method. If a small number of specified chemicals are being measured, the sample is ideally spiked at one-half to one-and-one-half times the concentration found in the sample.

The recovery of the spiked samples is calculated and summarized in the quality control record as accuracy and gives the control chart limits.

#### Establishment of Acceptance Limits of Precision and Accuracy

Each set of samples analyzed per analyte has a blank, duplicate, spike and a standard reference material from which the precision and accuracy data are obtained.

The precision of RPD is obtained by the manipulation of duplicate sample data as follows:

$$RPD = \frac{(D1 - D2)}{(D1 + D2)/2} \times 100$$

where D1 = sample  
D2 = sample replicate

The accuracy is a measurement of the percentage of a spike recovery, %R, calculated by the formula:

$$\%R = [(SSR - SR) / SA] \times 100$$

where SSR = spiked sample  
SR = sample  
SA = spike added

Control charts are maintained to show the limits within which measurements should fall. The upper and lower control limits are calculated as follows and are based on 25 sample sets:

$$\begin{aligned} \text{Upper control limits} &= M + 3 S_m \quad (\text{UCL}) \\ \text{Lower control limits} &= M - 3 S_m \quad (\text{LCL}) \end{aligned}$$

M = the average of the RPD  
S<sub>m</sub> = standard deviation of the RPD

#### Procedures for Corrective Action

If values fall outside the ULC or LCL, the following guidelines are taken for corrective action:

1. Define the problem.
2. QA/QC officer and laboratory section leader assign the investigation responsibility to an analyst.
3. Document the action needed to correct the problem.
4. Implement and verify that corrective action is taken and the problem corrected.

In general, when QA techniques or procedures identify errors, deficiencies or an "out of control" situation, and two types of action need to be considered. The first, immediate action is generally to correct instrumentation error or malfunction, poor technique, or sample variability. Long-term action is to correct out-of-control conditions that may stem from contamination, old standards, improper spiking, or improperly calibrated equipment.

The above guidelines would be followed to correct the problem and maintain acceptable levels of confidence. No laboratory results will be reported or released until the "out of control" situation is rectified.

All worksheets given to the analyst for analyte determination are dated and initialed after major analytical procedures are completed, i.e. on date weighed, after extraction, upon completion of digestion, and on the date the sample is given to the laboratory supervisor for review. This is signed by the supervisor after review for reliability in terms of accuracy, precision, detection limits, and quantitative limits, and forwarded to data processing.

Reports submitted to clients routinely include method numbers and detection limits as well as identifying information, date received, data analyzed, etc.

#### Maintenance and Repair of Instrumentation

Instruments are maintained according to the operation manuals supplied by the manufacturer. Repairs are conducted as needed, either by manufacturer representatives or by in-house personnel (for simple problems). Routine maintenance, such as lamp replacement, is conducted as indicated by the collected QC data.

### **3.0 Bioassay**

All bioassays for this testing program (with the exception of the bivalve larval test) were conducted following methods outlined in the EPA/COE Testing Manual (1991). The bivalve larval bioassays were performed according to protocol described in ASTM (1989). Standard operating procedures (SOPs) have been written and approved for these procedures, and are accessible to all bioassay staff. Dilution water for the bioassays, collected from the ToxScan Davenport laboratory, meets all requirements outlined in ASTM (1989).

Data resulting from the bioassays were recorded in ink on laboratory data sheets, evaluated by the project manager to insure that all test conditions were within protocol limits, and incorporated into the permanent project record file.

SOPs have been developed for instrument calibration, which detail standards to be used, units for reporting data and expected performance standards for accuracy and precision. Water quality monitoring instruments (D.O. meter, pH meter, salinometer, thermometer) are calibrated at least once daily according to these SOPs, and data are recorded in logbooks at the laboratory. Backup instrumentation is available in the event of equipment failure.

Bioassay test protocols generally specify acceptable limits of water quality (pH, D.O., temperature, salinity) in test containers during test performance. They also specify certain minimum levels of organism response (survival, normal development, growth) which must be achieved in test controls in order to validate the bioassay. A reference toxicant bioassay has been requested for this program as an additional quality assurance measure. Reference toxicant tests serve to "calibrate" the sensitivity of organisms to a known toxic compound, and control charts are maintained in the laboratory

for each organism:toxicant combination. Our control charts are continuously updated as each new reference toxicant bioassay data set is incorporated. In order to be within control limits, the reftox EC50 or LC50 must fall within the range of  $\pm 2$  standard deviations of the mean of all previous reference toxicant bioassays. The following table outlines reference toxicants used by the laboratory with each test species:

TEST ORGANISM	REFERENCE TOXICANT	CONCENTRATION RANGE
Amphipod	Cadmium Chloride ( $\text{CdCl}_2$ )	0.125 - 4.0 mg/L
Mysid	Sodium Dodecyl Sulfate (SDS)	0.5 - 16.0 mg/L
Fish (Sanddab)	Sodium Dodecyl Sulfate (SDS)	0.25 - 4.0 mg/L
Bivalve (Mussel)	Copper Sulfate ( $\text{CuSO}_4$ )	2.0 - 32.0 $\mu\text{g/L}$
Worm (Polychaete)	Copper Sulfate ( $\text{CuSO}_4$ )	6.25 - 500 $\mu\text{g/L}$

Statistical analyses of bioassay data are performed using computer programs which provide not only the EC50 or LC50 calculation but also provide estimates of the precision of the data in the form of 95% confidence limits around the EC/LC50 point.

QA/QC data for chemical analyses and reference toxicant data for this test program, as well as environmental monitoring data for these bioassays are presented herein. Chains of Custody are presented in Appendix E.

Table 5. Summary of instruments, calibration methods, precision/accuracy monitoring, maintenance and record-keeping for analytical equipment utilized in this test program.

Analyte	Instruments	Calibration Method	Precision & Accuracy Standards	Maintenance Schedule	Record-keeping Methods
Metals	Varian AA5; Models 400P, 4002, 10	3-4 point standard curve	SRMs* and replicate analyses	as needed	instrument print-out, electronic meter hard copy
Oil & Grease	Perkin-Elmer IR Spectrophotometer Model 710B	4-point standard curve	spikes and replicate analyses	as needed	chromatogram charts, hard copy
Sulfides	Titration	standardized titrant	replicate analyses	clean burettes	notebook hard copy
Organotins	Hewlett-Packard GC; model 5890, series II	3-point standard curve and surrogate injection	SRMs and replicate analyses	as needed	instrument print-out, hard copy
Chlorinated pesticides and PCBs	Hewlett-Packard GC; model 5890 dual columns; ECD detectors	3-point standard curve	SRMs, matrix spikes, matrix spike duplicates, duplicate samples, surrogates	as needed	instrument printout and work sheet
PAHs, phenols, phthalates	Varian GC/MS Saturn II	5-point standard curve	SRMs, matrix spikes, matrix spike duplicates, duplicate samples, surrogates	as needed	instrument printout and work sheet

\* SRM = standard reference materials, obtained from NIST (National Institute of Standards and Technology).

Chlorinated Pesticides  
EPA METHOD 8080  
QA/QC Report  
QA/QC on sample SAM-7D

<u>Compound</u>	<u>% REC MS</u>	<u>% REC MSD</u>	<u>% RPD</u>	<u>QC LIMITS</u>	
				<u>% REC</u>	<u>% RPD</u>
Lindane	125	120	4	46-127	50
Heptachlor	220†	105	-	35-130	31
Aldrin	345†	210†	-	34-132	43
Dieldrin	124	124	0	31-134	38
Endrin	114	118	3	42-139	43
DDT	76	36	71*	23-134	50

\* Outside QC Limits; no corrective action required.

† Matrix Interference; LCS within QC limits.

MS = matrix spike

MSD = matrix spike duplicate

RPD = relative percent difference

Chlorinated Pesticides  
EPA METHOD 8080  
QA/QC Report  
QA/QC on sample EK1

<u>Compound</u>	<u>% REC MS</u>	<u>% REC MSD</u>	<u>% RPD</u>	<u>QC LIMITS</u>	
				<u>% REC</u>	<u>% RPD</u>
Lindane	100	65	42	46-127	50
Heptachlor	105	110	5	35-130	31
Aldrin	100	85	16	34-132	43
Dieldrin	72	136†	62†	31-134	38
Endrin	62	98	45*	42-139	43
DDT	76	124	48	23-134	50

\* Outside QC Limits; no corrective action required.

† Matrix Interference; LCS within QC limits.

MS = matrix spike

MSD = matrix spike duplicate

RPD = relative percent difference



Chlorinated Pesticides  
EPA METHOD 8080  
QA/QC Report  
QA/QC on sample Home Sediment

<u>Compound</u>	<u>% REC MS</u>	<u>% REC MSD</u>	<u>% RPD</u>	<u>QC LIMITS</u>	
				<u>% REC</u>	<u>% RPD</u>
Lindane	†	†	-	46-127	50
Heptachlor	†	†	-	35-130	31
Aldrin	†	†	-	34-132	43
Dieldrin	100	100	0	31-134	38
Endrin	100	140	33	42-139	43
DDT	120	130	8	23-134	50

† Matrix Interference; LCS within QC limits.

MS = matrix spike

MSD = matrix spike duplicate

RPD = relative percent difference

**Chlorinated Pesticides**  
**EPA METHOD 8080**  
**QA/QC Report**

<u>Compound</u>	<u>% REC LCS</u>	<u>QC LIMITS</u> <u>% REC</u>
Lindane	75	46-127
Heptachlor	85	35-130
Aldrin	70	34-132
Dieldrin	86	31-134
Endrin	84	42-139
DDT	54	23-134

LCS = Laboratory Control Sample

**Chlorinated Pesticides  
EPA METHOD 8080  
QA/QC Report**

<u>Compound</u>	<u>% REC LCS</u>	<u>QC LIMITS % REC</u>
Lindane	70	46-127
Heptachlor	105	35-130
Aldrin	150	34-132
Dieldrin	96	31-134
Endrin	96	42-139
DDT	112	23-134

LCS = Laboratory Control Sample

**Chlorinated Pesticides**  
**EPA METHOD 8080**  
**QA/QC Report**

<u>Compound</u>	<u>% REC LCS</u>	<u>QC LIMITS</u> <u>% REC</u>
Lindane	90	46-127
Heptachlor	95	35-130
Aldrin	95	34-132
Dieldrin	88	31-134
Endrin	92	42-139
DDT	56	23-134

LCS = Laboratory Control Sample

**Chlorinated Pesticides  
EPA METHOD 8080  
µg/Kg (ppb) dry weight  
QA/QC Report**

<u>Analyte</u>	<u>HOME-SED</u>	<u>HOME-SED Duplicate</u>	<u>Detection Limit</u>
Aldrin	ND	ND	0.5
alpha-BHC	ND	ND	1.0
beta-BHC	ND	ND	1.0
delta-BHC	ND	ND	1.0
gamma-BHC (lindane)	ND	ND	1.0
alpha-Chlordane	ND	ND	1.0
gamma-Chlordane	ND	ND	1.0
4,4'-DDD	ND	ND	1.0
4,4'-DDE	ND	ND	1.0
4,4'-DDT	ND	ND	1.0
Dieldrin	ND	ND	0.5
Endosulfan I	ND	ND	2.0
Endosulfan II	ND	ND	0.5
Endosulfan sulfate	ND	ND	10
Endrin	ND	ND	0.5
Heptachlor	ND	ND	0.5
Heptachlor epoxide	ND	ND	10
Toxaphene	ND	ND	30
PCB's	ND	ND	20

ND = None Detected

Chlorinated Pesticides  
EPA METHOD 8080  
 $\mu\text{g}/\text{Kg}$  (ppb) dry weight  
QA/QC Report

<u>Analyte</u>	<u>SAM-7D</u>	<u>SAM-7D Duplicate</u>	<u>Detection Limit</u>
Aldrin	ND	ND	0.5
alpha-BHC	ND	ND	1.0
beta-BHC	ND	ND	1.0
delta-BHC	ND	ND	1.0
gamma-BHC (lindane)	ND	ND	1.0
alpha-Chlordane	ND	ND	1.0
gamma-Chlordane	ND	ND	1.0
4,4'-DDD	ND	ND	1.0
4,4'-DDE	ND	ND	1.0
4,4'-DDT	ND	ND	1.0
Dieldrin	ND	ND	0.5
Endosulfan I	ND	ND	2.0
Endosulfan II	ND	ND	0.5
Endosulfan sulfate	ND	ND	10
Endrin	ND	ND	0.5
Heptachlor	ND	ND	0.5
Heptachlor epoxide	ND	ND	10
Toxaphene	ND	ND	30
PCB's	ND	ND	20

ND = None Detected

Chlorinated Pesticides  
EPA METHOD 8080  
 $\mu\text{g}/\text{Kg}$  (ppb) dry weight  
QA/QC Report

<u>Analyte</u>	<u>EK-1</u>	<u>EK-1 Duplicate</u>	<u>Detection Limit</u>
Aldrin	ND	ND	0.5
alpha-BHC	ND	ND	1.0
beta-BHC	ND	ND	1.0
delta-BHC	ND	ND	1.0
gamma-BHC (lindane)	ND	ND	1.0
alpha-Chlordane	ND	ND	1.0
gamma-Chlordane	ND	ND	1.0
4,4'-DDD	ND	ND	1.0
4,4'-DDE	ND	ND	1.0
4,4'-DDT	ND	ND	1.0
Dieldrin	ND	ND	0.5
Endosulfan I	ND	ND	2.0
Endosulfan II	ND	ND	0.5
Endosulfan sulfate	ND	ND	10
Endrin	ND	ND	0.5
Heptachlor	ND	ND	0.5
Heptachlor epoxide	ND	ND	10
Toxaphene	ND	ND	30
PCB's	ND	ND	20

ND = None Detected

**Chlorinated Pesticides**  
**EPA METHOD 8080**  
**µg/Kg (ppb) dry weight**  
**QA/QC Report**

<u>Analyte</u>	<u>Dry Wt.</u> <u>Sample</u> <u>Value</u>	<u>Dry Wt.</u> <u>Detection</u> <u>Limit</u>
Sample ID: Method Blank (MB062194)		
Aldrin	ND	0.5
alpha-BHC	ND	1.0
beta-BHC	ND	1.0
delta-BHC	ND	1.0
gamma-BHC (lindane)	ND	1.0
alpha-Chlordane	ND	1.0
gamma-Chlordane	ND	1.0
4,4'-DDD	ND	1.0
4,4'-DDE	ND	1.0
4,4'-DDT	ND	1.0
Dieldrin	ND	0.5
Endosulfan I	ND	2.0
Endosulfan II	ND	0.5
Endosulfan sulfate	ND	10
Endrin	ND	0.5
Heptachlor	ND	0.5
Heptachlor epoxide	ND	10
Toxaphene	ND	30
PCBs:		
PCB 1242	ND	20
PCB 1248	ND	20
PCB 1254	ND	20
PCB 1260	ND	20
TOTAL PCBs	ND	20

ND = None Detected



Chlorinated Pesticides  
EPA METHOD 8080  
 $\mu\text{g/Kg}$  (ppb) dry weight  
QA/QC Report

Sample ID: Method Blank (MB041394)

<u>Analyte</u>	<u>Dry Wt. Sample Value</u>	<u>Dry Wt. Detection Limit</u>
Aldrin	ND	0.5
alpha-BHC	ND	1.0
beta-BHC	ND	1.0
delta-BHC	ND	1.0
gamma-BHC (lindane)	ND	1.0
alpha-Chlordane	ND	1.0
gamma-Chlordane	ND	1.0
4,4'-DDD	ND	1.0
4,4'-DDE	ND	1.0
4,4'-DDT	ND	1.0
Dieldrin	ND	0.5
Endosulfan I	ND	2.0
Endosulfan II	ND	0.5
Endosulfan sulfate	ND	10
Endrin	ND	0.5
Heptachlor	ND	0.5
Heptachlor epoxide	ND	10
Toxaphene	ND	30
PCBs:		
PCB 1242	ND	20
PCB 1248	ND	20
PCB 1254	ND	20
PCB 1260	ND	20
TOTAL PCBs	ND	20

ND = None Detected

Chlorinated Pesticides  
EPA METHOD 8080  
 $\mu\text{g/Kg}$  (ppb) dry weight  
QA/QC Report

Sample ID: Method Blank (MB041594)	Dry Wt. Sample Value	Dry Wt. Detection Limit
Analyte		
Aldrin	ND	0.5
alpha-BHC	ND	1.0
beta-BHC	ND	1.0
delta-BHC	ND	1.0
gamma-BHC (lindane)	ND	1.0
alpha-Chlordane	ND	1.0
gamma-Chlordane	ND	1.0
4,4'-DDD	ND	1.0
4,4'-DDE	ND	1.0
4,4'-DDT	ND	1.0
Dieldrin	ND	0.5
Endosulfan I	ND	2.0
Endosulfan II	ND	0.5
Endosulfan sulfate	ND	10
Endrin	ND	0.5
Heptachlor	ND	0.5
Heptachlor epoxide	ND	10
Toxaphene	ND	30
PCBs:		
PCB 1242	ND	20
PCB 1248	ND	20
PCB 1254	ND	20
PCB 1260	ND	20
TOTAL PCBs	ND	20

ND = None Detected

**Chlorinated Pesticides  
EPA METHOD 8080  
 $\mu\text{g}/\text{Kg}$  (ppb) dry weight  
QA/QC Report**

Sample ID: Method Blank (MB050994)

<u>Analyte</u>	<u>Dry Wt. Sample Value</u>	<u>Dry Wt. Detection Limit</u>
Aldrin	ND	0.5
alpha-BHC	ND	1.0
beta-BHC	ND	1.0
delta-BHC	ND	1.0
gamma-BHC (lindane)	ND	1.0
alpha-Chlordane	ND	1.0
gamma-Chlordane	ND	1.0
4,4'-DDD	ND	1.0
4,4'-DDE	ND	1.0
4,4'-DDT	ND	1.0
Dieldrin	ND	0.5
Endosulfan I	ND	2.0
Endosulfan II	ND	0.5
Endosulfan sulfate	ND	10
Endrin	ND	0.5
Heptachlor	ND	0.5
Heptachlor epoxide	ND	10
Toxaphene	ND	30
PCBs:		
PCB 1242	ND	20
PCB 1248	ND	20
PCB 1254	ND	20
PCB 1260	ND	20
TOTAL PCBs	ND	20

ND = None Detected

**Chlorinated Pesticides**  
**EPA METHOD 8080**  
**QA/QC Report**  
**Surrogate Recoveries (%)**

<u>Sample</u>	<u>Tetrachloro-m-xylene</u>	<u>Decachlorobiphenyl</u>
SAM-7D	132	124
SAM-5D	214†	204†
SAM-3D	88	109
SAM-4D	156†	110
SAM-2D	98	122
SAM-1D	101	117
SAM-6C	212†	118
SAM-6B	168†	109
SAM-6A	93	96
SAM-4(D)	87	93
SAM-6 Composite	80	95
EK-1	79	90
EK-2	88	98
EK-3	106	104
EK-4	128	108
EK Composite	135	104
NB-4 Rep 1	172†	93
FL-8	119	98
FL-7	179†	97
FL-6	165†	102
FL-5	153†	92
FL-3	137	90
FL-4	166†	85
FL-1	110	81
FL-2	134	87
FL-Composite	202†	75
Reference Composite	68	75
Home Sediment	179†	123

QC Limits

(20-150)

† Matrix Interference

**Organic Compounds  
SRM QA/QC Report  
µg/Kg (ppb)**

<u>Element</u>	<u>Value Found</u>	<u>Certified Value</u>	<u>Advisory Range</u>
<b>Pesticides</b>			
Aldrin	275	246	103-300
beta-BHC	208	145	25-213
gamma-BHC (Lindane)	263	219	70-278
4,4'-DDD	421	334	104-471
4,4'-DDE	554	455	137-660
4,4'-DDT	316	287	72-459
Dieldrin	98	101	36-147
Endrin	386	297	122-598
Heptachlor	301	160	101-330
Methoxychlor	251	160	61-253
<b>Semi-volatiles</b>			
Naphthalene	757	1640	623-2180
Fluorene	2922	6060	2670-7330
Phenanthrene	1144	1570	848-1880
Anthracene	59*	1010	273-1340
Pyrene	4641	8030	4180-9230
Benzo(a)anthracene	519*	2030	670-2900
Chrysene	1727	2350	400-3950
Butylbenzylphthalate	7510	10600	1590-16100
Bis(2-ethylhexyl)phthalate	6056	7120	1140-11200

\* Outside QC limits; no corrective action required.

SRM = Environmental Resource Associates Lot #324

**Polynuclear Aromatic Hydrocarbons (PAHs)**  
**EPA METHOD 8270**  
**QA/QC Report**  
**QA/QC on sample HUM94SED0001**

<u>Compound</u>	<u>% REC MS</u>	<u>% REC MSD</u>	<u>% RPD</u>	<u>QC LIMITS</u>	
				<u>% REC</u>	<u>% RPD</u>
Acenaphthene	79	88	11	31-137	19
Pyrene	62	64	3	35-142	36

MS = Matrix Spike

MSD = Matrix Spike Duplicate

RPD = Relative Percent Difference

**Polynuclear Aromatic Hydrocarbons (PAHs)  
EPA METHOD 8270  
QA/QC Report  
QA/QC on sample HUM94SED0028**

<u>Compound</u>	<u>% REC MS</u>	<u>% REC MSD</u>	<u>% RPD</u>	<u>QC LIMITS</u>	
				<u>% REC</u>	<u>% RPD</u>
Acenaphthene	94	87	8	31-137	19
Pyrene	110	93	17	35-142	36

MS = Matrix Spike

MSD = Matrix Spike Duplicate

RPD = Relative Percent Difference

**Polynuclear Aromatic Hydrocarbons (PAHs)**  
**EPA METHOD 8270**  
**QA/QC Report**

<u>Compound</u>	<u>% REC LCS</u>	<u>QC LIMITS</u> <u>% REC</u>
Acenaphthene	67	31-137
Pyrene	70	35-142

LCS = Laboratory Control Sample



Polynuclear Aromatic Hydrocarbons (PAHs)  
EPA METHOD 8270  
QA/QC Report

<u>Compound</u>	<u>% REC LCS</u>	<u>QC LIMITS</u> <u>% REC</u>
Acenaphthene	80	31-137
Pyrene	71	35-142

LCS = Laboratory Control Sample

Polynuclear Aromatic Hydrocarbons (PAHs)  
EPA METHOD 8270  
QA/QC Report

<u>Compound</u>	<u>% REC LCS</u>	<u>QC LIMITS</u> <u>% REC</u>
Acenaphthene	79	31-137
Pyrene	61	35-142

LCS = Laboratory Control Sample

**Polynuclear Aromatic Hydrocarbons (PAHs)**  
**EPA Method 8270**  
**µg/Kg (ppb) dry weight**  
**QA/QC Report**

<u>Analyte</u>	<u>SAM-7D</u>	<u>SAM-7D DUPLICATE</u>	<u>Detection Limit</u>
Naphthalene	ND	ND	20
Acenaphthylene	ND	ND	20
Acenaphthene	ND	ND	20
Fluorene	ND	ND	20
Phenanthrene	ND	20	20
Anthracene	ND	ND	20
Fluoranthene	ND	ND	20
Pyrene	ND	ND	25
Chrysene	ND	ND	20
Benzo(a)anthracene	ND	ND	20
Benzo(b)fluoranthene	ND	ND	20
Benzo(k)fluoranthene	ND	ND	20
Benzo(a)pyrene	ND	ND	20
Indeno(1,2,3-CD)pyrene	ND	ND	20
Dibenzo(a,h)anthracene	ND	ND	20
Benzo(ghi)perylene	ND	ND	20
2-Methylnaphthalene	ND	ND	20
Total PAH's	ND	20	20
Total phthalates	110	120	10

ND = None Detected

Polynuclear Aromatic Hydrocarbons (PAHs)  
EPA Method 8270  
 $\mu\text{g/Kg}$  (ppb) dry weight  
QA/QC Report

<u>Analyte</u>	<u>FL-7</u>	<u>FL-7 DUPLICATE</u>	<u>Detection Limit</u>
Naphthalene	84	37	20
Acenaphthylene	ND	ND	20
Acenaphthene	ND	ND	20
Fluorene	ND	ND	20
Phenanthrene	70	62	20
Anthracene	ND	ND	20
Fluoranthene	ND	ND	20
Pyrene	31	27	25
Chrysene	ND	ND	20
Benzo(a)anthracene	38	ND	20
Benzo(b)fluoranthene	ND	ND	20
Benzo(k)fluoranthene	ND	ND	20
Benzo(a)pyrene	ND	ND	20
Indeno(1,2,3-CD)pyrene	ND	ND	20
Dibenzo(a,h)anthracene	ND	ND	20
Benzo(ghi)perylene	ND	ND	20
2-Methylnaphthalene	59	55	20
Total PAH's	280	180	20
Total phthalates	250	180	10

ND = None Detected

Polynuclear Aromatic Hydrocarbons (PAHs)  
EPA Method 8270  
 $\mu\text{g/Kg}$  (ppb) dry weight  
QA/QC Report

Analyte	MB050194 Method Blank	Detection Limit
Naphthalene	ND	20
Acenaphthylene	ND	20
Acenaphthene	ND	20
Fluorene	ND	20
Phenanthrene	ND	20
Anthracene	ND	20
Fluoranthene	ND	20
Pyrene	ND	25
Chrysene	ND	20
Benzo(a)anthracene	ND	20
Benzo(b)fluoranthene	ND	20
Benzo(k)fluoranthene	ND	20
Benzo(a)pyrene	ND	20
Indeno(1,2,3-CD)pyrene	ND	20
Dibenzo(a,h)anthracene	ND	20
Benzo(ghi)perylene	ND	20
2-Methylnaphthalene	ND	20
Total PAH's	ND	20
Total phthalates	940*	10

\* Suspected laboratory contaminant

ND = None Detected

Polynuclear Aromatic Hydrocarbons (PAHs)  
EPA Method 8270  
 $\mu\text{g}/\text{Kg}$  (ppb) dry weight  
QA/QC Report

<u>Analyte</u>	<u>MB050594 Method Blank</u>	<u>Detection Limit</u>
Naphthalene	ND	20
Acenaphthylene	ND	20
Acenaphthene	ND	20
Fluorene	ND	20
Phenanthrene	ND	20
Anthracene	ND	20
Fluoranthene	ND	20
Pyrene	ND	25
Chrysene	ND	20
Benzo(a)anthracene	ND	20
Benzo(b)fluoranthene	ND	20
Benzo(k)fluoranthene	ND	20
Benzo(a)pyrene	ND	20
Indeno(1,2,3-CD)pyrene	ND	20
Dibenzo(a,h)anthracene	ND	20
Benzo(ghi)perylene	ND	20
2-Methylnaphthalene	ND	20
Total PAH's	ND	20
Total phthalates	59*	10

\* Suspected laboratory contaminant

ND = None Detected

Polynuclear Aromatic Hydrocarbons (PAHs)  
EPA Method 8270  
 $\mu\text{g}/\text{Kg}$  (ppb) dry weight  
QA/QC Report

<u>Analyte</u>	<u>MB041494 Method Blank</u>	<u>Detection Limit</u>
Naphthalene	ND	20
Acenaphthylene	ND	20
Acenaphthene	ND	20
Fluorene	ND	20
Phenanthrene	ND	20
Anthracene	ND	20
Fluoranthene	ND	20
Pyrene	ND	25
Chrysene	ND	20
Benzo(a)anthracene	ND	20
Benzo(b)fluoranthene	ND	20
Benzo(k)fluoranthene	ND	20
Benzo(a)pyrene	ND	20
Indeno(1,2,3-CD)pyrene	ND	20
Dibenzo(a,h)anthracene	ND	20
Benzo(ghi)perylene	ND	20
2-Methylnaphthalene	ND	20
Total PAH's	ND	20
Total phthalates	190*	10

\* Suspected laboratory contaminant

ND = None Detected

Polynuclear Aromatic Hydrocarbons (PAHs)  
EPA Method 8270  
 $\mu\text{g/Kg}$  (ppb) dry weight  
QA/QC Report

<u>Analyte</u>	<u>MB041394 Method Blank</u>	<u>Detection Limit</u>
Naphthalene	ND	20
Acenaphthylene	ND	20
Acenaphthene	ND	20
Fluorene	ND	20
Phenanthrene	ND	20
Anthracene	ND	20
Fluoranthene	ND	20
Pyrene	ND	25
Chrysene	ND	20
Benzo(a)anthracene	ND	20
Benzo(b)fluoranthene	ND	20
Benzo(k)fluoranthene	ND	20
Benzo(a)pyrene	ND	20
Indeno(1,2,3-CD)pyrene	ND	20
Dibenzo(a,h)anthracene	ND	20
2-Methylnaphthalene	ND	20
Total PAH's	ND	20
Total phthalates	90*	10

\* Suspected laboratory contaminant

ND = None Detected



Polynuclear Aromatic Hydrocarbons (PAHs)  
EPA Method 8270  
 $\mu\text{g/Kg}$  (ppb) dry weight  
QA/QC Report

<u>Analyte</u>	<u>MB062194 Method Blank</u>	<u>Detection Limit</u>
Naphthalene	ND	20
Acenaphthylene	ND	20
Acenaphthene	ND	20
Fluorene	ND	20
Phenanthrene	ND	20
Anthracene	ND	20
Fluoranthene	ND	20
Pyrene	ND	25
Chrysene	ND	20
Benzo(a)anthracene	ND	20
Benzo(b)fluoranthene	ND	20
Benzo(k)fluoranthene	ND	20
Benzo(a)pyrene	ND	20
Indeno(1,2,3-CD)pyrene	ND	20
Dibenzo(a,h)anthracene	ND	20
2-Methylnaphthalene	ND	20
Total PAH's	ND	20
Total phthalates	150*	10

\* Suspected laboratory contaminant

ND = None Detected

**Polynuclear Aromatic Hydrocarbons (PAHs) and Phthalate Esters**  
**EPA Method 8270**  
**QA/QC Report**  
**Surrogate Recovery (%)**

<u>Sample ID</u>	<u>S1</u>	<u>S2</u>	<u>S3</u>	<u>S4</u>	<u>S5</u>	<u>S6</u>
SAM-7D	54	62	62	70	51	82
SAM-5D	52	61	66	71	50	86
SAM-4D	48	57	53	66	69	81
SAM-3D	52	58	59	67	53	80
SAM-2D	60	71	78	80	48	85
SAM-1D	53	58	39	75	44	71
SAM-6C	60	57	62	71	68	84
SAM-6B	66	68	69	73	86	96
SAM-6A	41	49	57	61	34	78
SAM-4(D)	56	68	68	85	58	84
SAM-6 Composite	70	69	63	81	83	86
EK-1	52	63	72	89	34	89
EK-2	61	65	59	78	86	89
EK-3	63	67	61	79	88	84
EK-4	73	77	67	81	95	91
EK Composite	67	69	62	76	89	89
NB-4 Rep 1	70	65	65	76	85	80
FL-8	62	70	63	79	72	83
FL-7	55	66	67	74	57	89
FL-6	76	82	71	88	101	92
FL-5	72	67	52	78	78	91
FL-4	68	78	72	88	87	74
FL-3	64	71	67	91	94	82
FL-2	73	85	76	89	93	77
FL-1	66	73	71	85	94	74
FL-Composite	59	72	69	88	98	78
Reference Composite	39	57	46	75	76	85
Home Sediment	25	28	39	49	42	47

<u>Surrogate ID</u>	<u>QC Limits</u>
S1 = 2-Fluorophenol	(25-121)
S2 = Phenol-d5	(24-113)
S3 = Nitrobenzene-d5	(23-120)
S4 = 2-Fluorobiphenyl	(30-115)
S5 = 2,4,6-Tribromophenol	(19-122)
S6 = Terphenyl-d14	(18-137)

**Organotin Speciation**  
**µg/Kg (ppb) dry weight**  
**QA/QC Report**  
**QA/QC on sample SAM-6 Composite**

<u>Sample ID</u>	<u>Monobutyltin</u>	<u>Dibutyltin</u>	<u>Tributyltin</u>	<u>Tetrabutyltin</u>	<u>% TPT SUR</u>
Amount of Spike (µg/Kg) = 100					
Spike 1 (%)	3	85	88	74	60
SRM (%)	188	122	102	-	53
Blank	ND	ND	ND	ND	54

TPT Sur = Tripropyltin surrogate recovery  
 Detection Limit = 1 ppb  
 ND = None Detected

NOTE: As stated in TBT methodology protocol<sup>1</sup>, the analytical method has been optimized to tributyltin at the decreased efficiency of monobutyltin extraction and recovery of analyte

	<u>SRM Value Found</u>	<u>SRM Certified Value</u>	<u>% Recovery</u>
Dibutyltin	1.42	1.16	122
Tributyltin	1.30	1.27	102

SRM = National Research Council Canada PACS-1, marine sediment

<sup>1</sup>Battelle Project No. N-0519-6100, Measurement of Butyltin Species in Sediment by n-Pentyl Derivatization with Gas Chromatography/Flame Photometric Detection.

**Organotin Speciation**  
**µg/Kg (ppb) dry weight**  
**QA/QC Report**  
**QA/QC on sample EK-1**

<u>Sample ID</u>	<u>Monobutyltin</u>	<u>Dibutyltin</u>	<u>Tributyltin</u>	<u>Tetrabutyltin</u>	<u>% TPT SUR</u>
Amount of Spike (µg/Kg) = 100					
Spike 1 (%)	7	74	99	83	64
SRM (%)	165	101	88	-	85
Blank	ND	ND	ND	ND	70

TPT Sur = Tripropyltin surrogate recovery  
 Detection Limit = 1 ppb  
 ND = None Detected

NOTE: As stated in TBT methodology protocol<sup>2</sup>, the analytical method has been optimized to tributyltin at the decreased efficiency of monobutyltin extraction and recovery of analyte

	<u>SRM Value Found</u>	<u>SRM Certified Value</u>	<u>% Recovery</u>
Dibutyltin	1.17	1.16	101
Tributyltin	1.12	1.27	88

SRM = National Research Council Canada PACS-1, marine sediment

<sup>2</sup>Battelle Project No. N-0519-6100, Measurement of Butyltin Species in Sediment by n-Pentyl Derivatization with Gas Chromatography/Flame Photometric Detection.

**Metals**  
**µg/g (ppm)**  
**QA/QC Report**  
**QA/QC on sample SAM-7D**

Analyte/ Sample ID	Amount of Spike µg/mL	% Recovery of Spike*	Rep 1	Rep 2	RPD**	Method Blank
Arsenic	0.29	107	5.9	6.0	2	ND
Cadmium	0.029	92	ND	ND	NA	ND
Chromium	2.86	117	77	96	22	ND
Copper	2.86	86	5.7	6.3	10	ND
Lead	1.43	95	6.1	5.7	7	ND
Mercury	0.029	98	0.020	0.025	22	ND
Nickel	1.43	107	41	43	5	ND
Selenium	0.14	79	ND	ND	NA	ND
Silver	0.29	96	ND	ND	NA	ND
Zinc	2.86	94	31	33	6	ND

ND = None Detected

NA = Not Applicable

\*\* ToxScan acceptance limits: 30% (at 5 times detection limit)

Element	Value Found µg/g	Certified Value µg/g	+/-	Percent Recovery*
Arsenic	21.2	23.4	0.8	91
Cadmium	3.69	3.45	0.2	107
Chromium	95.6	135	5.0	71
Copper	83.0	89.6	5.0	84
Lead	155	161	17	96
Mercury	1.20	1.47	0.07	82
Nickel	31.5	44.1	3.0	71
Zinc	373	438	12	85

SRM = National Institute of Standards and Technology Estuarine Sediment, # 1646

\* ToxScan acceptance limits: 65-130% recovery

**Metals**  
**µg/g (ppm)**  
**QA/QC Report**  
**QA/QC on sample FL-2**

<u>Analyte/ Sample ID</u>	<u>Amount of Spike µg/mL</u>	<u>% Recovery of Spike*</u>	<u>Rep 1</u>	<u>Rep 2</u>	<u>RPD**</u>	<u>Method Blank</u>
Arsenic	0.29	104	8.9	8.4	6	ND
Cadmium	0.029	106	0.3	0.2	40	ND
Chromium	2.86	106	120	120	0	ND
Copper	2.86	94	38	36	5	ND
Lead	1.43	96	12	11	9	ND
Mercury	0.029	107	0.10	0.10	0	ND
Nickel	1.43	118	110	110	0	ND
Selenium	0.14	73	ND	ND	NA	ND
Silver	0.29	99	ND	ND	NA	ND
Zinc	2.86	101	82	79	4	ND

ND = None Detected

NA = Not Applicable

\* ToxScan acceptance limits: 65-130% recovery

\*\* ToxScan acceptance limits: 30% (at 5 times detection limit)

### Inorganic Method Numbers

Percent Solids .....	EPA Method 160.3
Sulfides .....	EPA/SW846 Method 9030
Total Organic Carbon .....	Gaudette et al. <sup>1</sup>
Arsenic .....	EPA/SW846 Method 7060
Cadmium .....	EPA/SW846 Method 7131
Chromium .....	EPA/SW846 Method 7191
Copper .....	EPA/SW846 Method 7210
Lead .....	EPA/SW846 Method 7421
Mercury .....	EPA/SW846 Method 7471
Nickel .....	EPA/SW846 Method 7521
Selenium .....	EPA/SW846 Method 7741
Silver .....	EPA/SW846 Method 7761
Zinc .....	EPA/SW846 Method 7950
Organotins .....	GC/FPD
Oil & Grease .....	Standard Method 5520C
Total Petroleum Hydrocarbons .....	Standard Method 5520F

<sup>1</sup> Henri E. Gaudette, Wilson R. Flight, Lois Toner and David W. Folger, *Determination of Organic Carbon in Recent Sediments*; Journal of Sedimentary Petrology, Vol. 44, No. 1, p. 249-253; 1974.

Summary of Environmental Monitoring Data: Bivalve Larvae Suspended Particulate Phase Bioassay

Sample ID	Parameter	Initial	Final
Seawater Control	pH value (units)	7.9	8.1
	Temperature (°C)	16.9	15.6
	D.O. (mg/L)	7.6	7.4
	Salinity (‰)	32.8	—
Humboldt Reference Sediment	pH value (units)	7.7	8.1
	Temperature (°C)	16.5	15.5
	D.O. (mg/L)	7.8	7.5
	Salinity (‰)	33.0	—
SAMTB  100%	pH value (units)	7.8	8.1
	Temperature (°C)	16.7	15.4
	D.O. (mg/L)	7.8	7.4
	Salinity (‰)	32.8	—
FLT  100%	pH value (units)	7.8	8.2
	Temperature (°C)	16.7	15.4
	D.O. (mg/L)	7.7	7.5
	Salinity (‰)	32.8	—
EKUP  100%	pH value (units)	7.8	8.2
	Temperature (°C)	16.7	15.5
	D.O. (mg/L)	7.8	7.5
	Salinity (‰)	32.8	—



Summary of Environmental Monitoring Data: *Rhepoxynius abronius* Solid Phase Static Bioassay

Sample ID	Parameter	Mean	Std.Dev.	Maximum	Minimum
Control	D.O. (mg/L)	7.69	0.11	8.0	7.3
	Temperature (°C)	15.34	0.37	15.6	14.5
	pH value (units)	8.06	0.07	8.1	7.9
Humboldt Reference Sediment	D.O. (mg/L)	7.68	0.09	7.8	7.5
	Temperature (°C)	15.32	0.38	15.6	14.5
	pH value (units)	8.10	0.07	8.2	7.9
EKUP	D.O. (mg/L)	7.64	0.09	7.8	7.5
	Temperature (°C)	15.31	0.35	15.5	14.5
	pH value (units)	8.07	0.06	8.1	7.9
SAMTB	D.O. (mg/L)	7.65	0.11	7.9	7.5
	Temperature (°C)	15.32	0.37	15.5	14.5
	pH value (units)	8.07	0.06	8.2	7.9
FLTB	D.O. (mg/L)	8.07	0.05	8.1	7.9
	Temperature (°C)	15.26	0.38	15.7	14.6
	pH value (units)	8.07	0.05	8.1	7.9

Summary of Environmental Monitoring Data: *Holmesimysis costata* Solid Phase Flow Through Bioassay

Sample ID	Parameter	Mean	Std Dev	Maximum	Minimum
Control	D.O. (mg/L)	8.59	0.18	8.9	8.2
	Temperature (°C)	13.35	0.73	14.6	12.1
	pH value (units)	8.02	0.11	8.2	7.7
Humboldt Reference Sediment	D.O. (mg/L)	8.59	0.18	9.0	8.2
	Temperature (°C)	13.32	0.70	14.4	12.1
	pH value (units)	8.03	0.10	8.1	7.7
EKUP	D.O. (mg/L)	8.59	0.17	8.9	8.3
	Temperature (°C)	13.30	0.72	14.3	12.1
	pH value (units)	8.02	0.11	8.1	7.6
SAMTB	D.O. (mg/L)	8.61	0.20	9.0	8.0
	Temperature (°C)	13.33	0.71	14.4	12.0
	pH value (units)	8.02	0.10	8.1	7.7
FLT B	D.O. (mg/L)	8.61	0.19	9.0	8.3
	Temperature (°C)	13.34	0.70	14.4	12.0
	pH value (units)	8.03	0.11	8.1	7.6

Summary of Environmental Monitoring Data: *Nephtys caecoides* Solid Phase Flow Through Bioassay

Sample ID	Parameter	Mean	Std.Dev.	Maximum	Minimum
Control	D.O. (mg/L)	8.16	0.20	8.6	7.6
	Temperature (°C)	12.96	0.40	14.0	12.4
	pH value (units)	8.03	0.10	8.1	7.7
Humboldt Reference Sediment	D.O. (mg/L)	8.15	0.22	8.5	7.5
	Temperature (°C)	13.04	0.43	14.1	12.5
	pH value (units)	8.02	0.10	8.1	7.7
EKUP	D.O. (mg/L)	8.18	0.19	8.6	7.5
	Temperature (°C)	12.95	0.40	14.1	12.4
	pH value (units)	8.01	0.12	8.1	7.6
SAMTB	D.O. (mg/L)	8.19	0.17	8.5	7.7
	Temperature (°C)	12.97	0.38	14.1	12.4
	pH value (units)	8.00	0.13	8.1	7.6
FLTB	D.O. (mg/L)	8.12	0.18	8.4	7.7
	Temperature (°C)	13.03	0.40	14.2	12.5
	pH value (units)	8.02	0.10	8.1	7.7

Summary of Environmental Monitoring Data: *Citharichthys stigmaeus* Suspended Particulate Phase Bioassay

Sample ID	Parameter	Mean	Std.Dev.	Maximum	Minimum
Control	D.O. (mg/L)	7.73	0.12	7.9	7.4
	Temperature (°C)	14.71	0.06	14.8	14.6
	pH value (units)	8.02	0.05	8.1	7.9
Humboldt Reference Sediment	D.O. (mg/L)	7.61	0.31	7.9	6.1
	Temperature (°C)	14.72	0.07	14.8	14.6
	pH value (units)	8.04	0.10	8.2	7.8
EKUP	D.O. (mg/L)	7.65	0.23	7.9	6.3
	Temperature (°C)	14.70	0.08	15.0	14.6
	pH value (units)	7.90	0.24	8.1	6.9
SAMTB	D.O. (mg/L)	7.71	0.19	7.9	6.4
	Temperature (°C)	14.71	0.09	15.0	14.6
	pH value (units)	8.08	0.07	8.2	7.8
FLT B	D.O. (mg/L)	7.69	0.22	7.9	6.6
	Temperature (°C)	14.66	0.07	14.8	14.5
	pH value (units)	8.13	0.17	8.4	7.8

Summary of Environmental Monitoring Data: *Holmesimysis costata* Suspended Particulate Phase Bioassay

Sample ID	Parameter	Mean	Std.Dev.	Maximum	Minimum
Control	D.O. (mg/L)	7.75	0.29	8.1	7.3
	Temperature (°C)	14.80	0.11	15.0	14.7
	pH value (units)	8.02	0.04	8.1	8.0
Humboldt Reference Sediment	D.O. (mg/L)	7.64	0.30	8.0	7.0
	Temperature (°C)	14.76	0.11	15.0	14.6
	pH value (units)	8.01	0.10	8.1	7.7
EKUP	D.O. (mg/L)	7.64	0.28	8.0	7.1
	Temperature (°C)	14.73	0.07	14.9	14.7
	pH value (units)	8.04	0.07	8.1	7.9
SAMTB	D.O. (mg/L)	7.62	0.37	8.0	7.0
	Temperature (°C)	14.71	0.05	14.8	14.6
	pH value (units)	8.06	0.05	8.2	8.0
FLT B	D.O. (mg/L)	7.60	0.33	8.0	7.0
	Temperature (°C)	14.71	0.07	14.9	14.6
	pH value (units)	8.13	0.15	8.6	7.8

REFERENCE TOXICANT BIOASSAY  
*Mytilus edulis* (Copper)

Sample ID	Number		Total Recovered per 1 mL	Resuspended Volume	Total # Normal Larvae Recovered	% Survival	Mean % Survival $\pm$ S.D.	% Normal Development	Mean % Normal Development $\pm$ S.D.	Survival			Normal Development		
	Normal	Abnormal								Abotts Corrected Value	Mean Corrected Value	Abotts Corrected Value	Mean Corrected Value		
Control	1	126	2	128	50.5	6363	132.7	98.4	98.4	89.4	93.5	101.0	98.4		
	2	91	3	94	51	4641	96.8	96.8	99.0	78.0	±	95.1	±		
	3	84	0	84	51	4284	89.3	100.0	±	113.1	17.91	99.2	3.02		
	4	104	0	104	49	5096	106.3	100.0	1.42	57.8	80.1	94.6	96.8		
	5	78	0	78	51.5	4017	83.8	100.0		76.1	±	98.5	±		
2	1	89	0	89	49	4361	90.9	100.0	97.5	89.4	93.5	101.0	98.4		
	2	81	5	86	47	3807	79.4	94.2	±	78.0	±	95.1	±		
	3	115	2	117	48	5520	115.1	98.3	2.99	113.1	17.91	99.2	3.02		
4	1	60	4	64	47	2820	58.8	93.8	95.9	57.8	80.1	94.6	96.8		
	2	79	2	81	47	3713	77.4	97.5	±	76.1	±	98.5	±		
	3	106	4	110	49	5194	108.3	96.4	1.94	106.4	24.57	97.3	1.95		
8	1	23	56	79	47	1081	22.5	29.1	25.2	22.2	23.3	29.4	25.4		
	2	40	73	113	46	1840	38.4	35.4	±	37.7	±	35.7	±		
	3	10	81	91	48.5	485	10.1	11.0	12.67	9.9	13.92	11.1	12.80		
16	1	0	74	74	48	0	0.0	0.0	2.7	0.0	2.3	0.0	2.8		
	2	0	60	60	51	0	0.0	0.0	±	0.0	±	0.0	±		
	3	7	78	85	49	343	7.2	8.2	4.75	7.0	4.06	8.3	4.80		
32	1	0	10	10	51	0	0.0	0.0	0.0	0.0	0.0	0.0	0.0		
	2	0	9	9	49.8	0	0.0	0.0	±	0.0	±	0.0	±		
	3	0	0	0	55.5	0	0.0	0.0	0.00	0.0	0.00	0.0	0.00		

LC<sub>50</sub> = 5.72 ppb (5.21, 6.27); EC<sub>50</sub> = 6.66 ppb (6.21, 7.15)

Reference Toxicant Bioassay

Species: *Rhepoxynius abronius*  
Toxicant: Cadmium chloride

Date: 25 April 1994  
T-10774

Concentration (mg/L)	Replicate	Number Surviving		Mean % Survival
		Observation Time (hours)		
		0	96	
Control	1	10	10	100
	2	10	10	
0.125	1	10	10	90
	2	10	8	
0.25	1	10	8	85
	2	10	9	
0.50	1	10	8	75
	2	10	7	
1.00	1	10	6	40
	2	10	2	
2.00	1	10	1	5
	2	10	0	
4.00	1	10	0	0
	2	10	0	

96-hour  $LC_{50}$  (Spearman) = 0.76 ppm

95% confidence limits = 0.56 ppm - 1.02 ppm

Reference Toxicant Bioassay

Species: *Holmesimysis costata*  
Toxicant: Sodium Dodecyl Sulfate

Date: 4 May 1994  
T-10774

Concentration (mg/L)	Replicate	Number Surviving		Mean % Survival
		Observation Time (hours)		
		0	96	
Control	1	10	10	100
	2	10	10	
0.5	1	10	10	100
	2	10	10	
1.0	1	10	8	90
	2	10	10	
2.0	1	10	9	95
	2	10	10	
4.0	1	10	10	100
	2	10	10	
8.0	1	10	8	65
	2	10	5	
16.0	1	10	1	10
	2	10	1	

96-hour  $LC_{50}$  (Spearman) = 9.28 ppm

95% confidence limits = 7.61 ppm - 11.3 ppm



Reference Toxicant Bioassay

Species: *Holmesimysis costata*  
Toxicant: Sodium Dodecyl Sulfate

Date: 11 May 94  
T-10774

Concentration (mg/L)	Replicate	Number Surviving		Mean % Survival
		Observation Time (hours)		
		0	96	
Control	1	10	10	100
	2	10	10	
0.5	1	10	10	100
	2	10	10	
1.0	1	10	10	100
	2	10	10	
2.0	1	10	8	90
	2	10	10	
4.0	1	10	7	80
	2	10	9	
8.0	1	10	3	25
	2	10	2	
16.0	1	10	0	0
	2	10	0	

96-hour  $LC_{50}$  (Spearman) = 5.46 ppm

95% confidence limits = 4.45 ppm - 6.71 ppm

Reference Toxicant Bioassay

Species: *Citharichthys stigmaeus*  
 Toxicant: Sodium Dodecyl Sulfate

Date: 10 May 1994  
 T-10774

Concentration (mg/L)	Replicate	Number Surviving		Mean % Survival
		Observation Time (hours)		
		0	96	
Control	1	10	10	100
	2	10	10	
	3	10	10	
	4	10	10	
	5	10	10	
0.25	1	10	9	97
	2	10	10	
	3	10	10	
0.5	1	10	8	93
	2	10	10	
	3	10	10	
1.0	1	10	10	100
	2	10	10	
	3	10	10	
2.0	1	10	9	90
	2	10	10	
	3	10	8	
4.0	1	10	1	13
	2	10	2	
	3	10	1	

96-hour LC<sub>50</sub> (Spearman) = 2.87 ppm

95% confidence limits = 2.66 ppm - 3.10 ppm

Reference Toxicant Bioassay

Species: *Nephtys caecoides*  
Toxicant: Copper Sulfate

Date: 28 April 1994  
T-10774

Concentration ( $\mu\text{g/L}$ )	Replicate	Number Surviving		Mean % Survival
		Observation Time (hours)		
		0	96	
Control	1	10	8	90
	2	10	10	
6.25	1	10	8	90
	2	10	10	
12.5	1	10	9	85
	2	10	8	
25	1	10	8	70
	2	10	6	
50	1	10	9	90
	2	10	9	
100	1	10	6	60
	2	10	6	
500	1	10	0	0
	2	10	0	

96-hour  $LC_{50}$  (Spearman) = 108 ppb

95% confidence limits = 71.5 ppb - 162 ppb

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## Appendix E

### Appendix E: LPC Calculation and Test Organism Handling Logs



Table E1. Calculations of the Limiting Permissible Concentrations (from EPA/ACOE 1977) for disposal depths of **50, 20 and 10** meters, Humboldt Baseline Survey II, Composite FLTB: *Citharichthys stigmaeus*.

Mixing Zone Estimation ( $V_m$ )

Depth of disposal site (m) =	<u>50</u>	<u>20</u>	<u>10</u>
Pi =	3.1416	3.1416	3.1416
Width of vessel (m) =	11*	11*	11*
Length of vessel (m) =	35*	35*	35*
Speed of vessel (m/sec) =	0.5*	0.5*	0.5*
Time of discharge (sec) =	<u>15*</u>	<u>15*</u>	<u>15*</u>
Mixing Zone Volume ( $m^3$ ) =	2139075	855630	427815

Volume of Liquid Phase ( $V_w$ )

Bulk density (constant) =	1.5	1.5	1.5
Particle density (constant) =	2.6	2.6	2.6
Density of liquid phase (constant) =	1.0	1.0	1.0
Volume of disposal vessel ( $m^3$ ) =	<u>1155*</u>	<u>1155*</u>	<u>1155*</u>
Liquid phase volume ( $m^3$ ) =	794	794	794

Concentration of Suspended Phase ( $C_{sp}$ )

Percent Silt =	39.4	39.4	39.4
Percent Clay =	<u>19.1</u>	<u>19.1</u>	<u>19.1</u>
Volume ( $m^3$ ) of suspended phase ( $V_{sp}$ ) =	211	211	211
Projected Percent Concentration ( $C_{sp}$ ) =	0.0099	0.024	0.049
LC <sub>50</sub> from bioassay (% elutriate) =	71.7	71.7	71.7
Factor LC <sub>50</sub> X 0.01 =	0.717	0.717	0.717

The factored LC50s are higher than the projected concentrations; therefore the Limiting Permissible Concentration (LPC) is not exceeded for composite FLTB: *Citharichthys stigmaeus* for disposal depths of 50, 20 and 10 meters.

\* These values are estimations; actual values were not available.





TOXSCAN, INC.  
 Receiving, Evaluation and Acclimation Log Worksheet  
 REAL

Species: <u>Citharichthys stamineus</u>	Supplier: <u>Bretina &amp; Assoc.</u>	# Ordered: <u>960</u>
Date Shipped: _____	Date/Time Received: <u>06MAY94 1130</u>	Shipped via: <u>Fed Ex</u>

**INITIAL EVALUATION**

Package Intact? <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No	Initials: <u>clw</u>	pH: <u>6.8</u> units
Temperature: <u>14.7</u> °C	Dissolved Oxygen: <u>720</u> mg/l	Sal./Cond.: <u>31.9</u> ‰/µmhos
Estimated # received: <u>800 200/4 boxes</u>		# Dead: <u>4</u>
Organism condition/comments: <u>Arrived in 4 boxes, 1 pack of blue ice per box</u> <u>Divided into Holding Tanks 3, 7, &amp; 8</u>		

**HOLDING/ACCLIMATION**

Date	Initials	pH (units)	Temp (°C)	D.O. (mg/l)	Sal/Cond (‰/µmhos)	Food	# Dead	Comments
06MAY94	clw	8.0	11.7	9.1	33.3	—	—	Holding Tank Conditions
07MAY94	ASB	7.9	12.3	8.6	33.2	✓	1	
08MAY94	ASB	7.9	13.4		33.0	✓	1	
09MAY94	clw	8.1	13.7	8.6	33.0	Brine ✓	6	
10May 94	TS	8.0	14.1	8.3	33.0	—	8	Test ↑ today
		used	entire lot of animals - clw 11MAY94					

**TEST INITIATION**

Date	Initials	Test ID	# Organisms used	Comments
10MAY94	clw	T-10774	300	ACOE - Humboldt
		T-		
		T-		
		T-		
		T-		



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 To (Recipient's Name) Please Print: Pete Schaffer  
 Recipient's Phone Number (Very Important): (408) 724-4522

Company: REGINA AND ASSEC  
 Department/Floor No.:  
 Company: TORSCAN  
 Department/Floor No.:

Street Address: 520 OCEANA DR  
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Priority Overnight (Delivery by next business morning) <input checked="" type="checkbox"/> OTHER PACKAGING <input type="checkbox"/> FEDEX LETTER <input type="checkbox"/> FEDEX PAK* <input type="checkbox"/> FEDEX BOX <input type="checkbox"/> FEDEX TUBE  Standard Overnight (Delivery by next business afternoon, no Saturday delivery) <input type="checkbox"/> OTHER PACKAGING <input type="checkbox"/> FEDEX LETTER <input type="checkbox"/> FEDEX PAK* <input type="checkbox"/> FEDEX BOX <input type="checkbox"/> FEDEX TUBE  Economy Two-Day (Delivery by second business day) <input type="checkbox"/> ECONOMY* <input type="checkbox"/> OVERNIGHT FREIGHT**  Government Overnight (Restricted for authorized users only) <input type="checkbox"/> GOVT LETTER <input type="checkbox"/> GOVT PACKAGE  Two-Day Freight (for packages over 150 lbs.) <input type="checkbox"/> TWO-DAY FREIGHT**	1 HOLD AT FEDEX LOCATION WEEKDAY (Fill in Section H) <input checked="" type="checkbox"/> DELIVER WEEKDAY Saturday Service 31 HOLD AT FEDEX LOCATION SATURDAY (Fill in Section H) <input type="checkbox"/> DELIVER SATURDAY (Extra charge) (Not available to all locations) 9 SATURDAY PICK-UP (Extra charge) 4 DANGEROUS GOODS (Extra charge) 6 DRY ICE (Dangerous Goods Shipper's Declaration not required) 12 HOLIDAY DELIVERY (if offered) (Extra charge)	1 15 100	1 15 100	100			<input type="checkbox"/> Cash Received <input type="checkbox"/> Return Shipment <input type="checkbox"/> Third Party <input type="checkbox"/> Chg. To Del. <input type="checkbox"/> Chg. To Hold Street Address City State Zip Received By: X Date/Time Received FedEx Employee Number Release Signature:

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 Your Phone Number (Very Important): (707) 878-2853  
 To (Recipient's Name) Please Print: Pete Schaffer  
 Recipient's Phone Number (Very Important): (408) 724-4522

Company: REGINA AND ASSEC  
 Department/Floor No.:  
 Company: TORSCAN  
 Department/Floor No.:

Street Address: 520 OCEANA DR  
 Exact Street Address (We Cannot Deliver to P.O. Boxes or P.O. Zip Codes.): 42 Hanger Way

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 5 Cash/Check

SERVICES (Check only one box)	DELIVERY AND SPECIAL HANDLING (Check services required)	PACKAGES	WEIGHT In Pounds Only	YOUR DECLARED VALUE (See note)	Emp. No.	Date	Federal Express Use
Priority Overnight (Delivery by next business morning) <input type="checkbox"/> OTHER PACKAGING <input type="checkbox"/> FEDEX LETTER <input type="checkbox"/> FEDEX PAK* <input type="checkbox"/> FEDEX BOX <input type="checkbox"/> FEDEX TUBE  Standard Overnight (Delivery by next business afternoon, no Saturday delivery) <input type="checkbox"/> OTHER PACKAGING <input type="checkbox"/> FEDEX LETTER <input type="checkbox"/> FEDEX PAK* <input type="checkbox"/> FEDEX BOX <input type="checkbox"/> FEDEX TUBE  Economy Two-Day (Delivery by second business day) <input type="checkbox"/> ECONOMY* <input type="checkbox"/> OVERNIGHT FREIGHT**  Government Overnight (Restricted for authorized users only) <input type="checkbox"/> GOVT LETTER <input type="checkbox"/> GOVT PACKAGE  Two-Day Freight (for packages over 150 lbs.) <input type="checkbox"/> TWO-DAY FREIGHT**	1 HOLD AT FEDEX LOCATION WEEKDAY (Fill in Section H) <input checked="" type="checkbox"/> DELIVER WEEKDAY Saturday Service 31 HOLD AT FEDEX LOCATION SATURDAY (Fill in Section H) <input type="checkbox"/> DELIVER SATURDAY (Extra charge) (Not available to all locations) 9 SATURDAY PICK-UP (Extra charge) 4 DANGEROUS GOODS (Extra charge) 6 DRY ICE (Dangerous Goods Shipper's Declaration not required) 12 HOLIDAY DELIVERY (if offered) (Extra charge)	1 15 100	1 15 100	100			<input type="checkbox"/> Cash Received <input type="checkbox"/> Return Shipment <input type="checkbox"/> Third Party <input type="checkbox"/> Chg. To Del. <input type="checkbox"/> Chg. To Hold Street Address City State Zip Received By: X Date/Time Received FedEx Employee Number Release Signature:

REVISION DATE 12/92  
 PART #137204 FXEM 12  
 FORMAT #158  
 158

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Receiving, Evaluation and Acclimation Log Worksheet

REAL

Species: <u>Macoma nasuta</u>	Supplier: <u>Bireina &amp; Assoc.</u>	# Ordered: <u>800</u>
Date Shipped: <u>19APR94</u>	Date/Time Received: <u>20APR94:1000</u>	Shipped via: <u>fed ex 8637040274</u>

INITIAL EVALUATION

Package Intact? <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No	Initials: <u>FB</u>	pH: <u>7.0</u> units
Temperature: <u>15.0</u> °C	Dissolved Oxygen: <u>2.6</u> mg/l	Sal/Cond.: <u>32.5</u> (‰)umhos
Estimated # received: <u>800</u>	# Dead: <u>5</u>	
Organism condition/comments: <u>Very low DO level Mostly died by 1645</u> <u>Split between two tanks #7 and #8</u>		

HOLDING/ACCLIMATION

Date	Initials	pH (units)	Temp (°C)	D.O. (mg/l)	Sal/Cond (‰)umhos	Food	# Dead	Comments
20APR94	FB	8.1	13.2	8.6	33.2	NA	—	Holding tank conditions
21APR94	FB	8.1 / 8.1	13.4 / 12.9	8.7 / 8.9	33.2 / 33.2	1 / 1	—	
22APR94	CRW	8.1 / 8.1	12.6 / 12.2	8.5 / 9.1	33.3 / 33.3	NA	4	
23 Apr 94	ASB/CRW	8.1 / 8.1	12.3 / 12.3	8.6 / 7.2	33.1	NA	2	
24APR94	FB/Ass	8.1 / 8.1	11.5 / 11.2	8.5 / 8.9	33.2	NA	1 / 0	
25APR94	FB	8.1 / 8.1	13.0 / 12.0	8.8 / 9.3	33.1	NA	1 / 1	
26APR94	FB	8.0 / 8.0	12.2 / 11.6	8.0 / 8.9	32.9	NA	0 / 0	
27 Apr 94	TS	8.1 / 8.1	12.1 / 12.2	8.6 / 8.9	32.8	—	1 / 0	
28APR94	CRW	8.0 / 7.9	12.5 / 12.3	8.0 / 8.4	32.8	—	2 / 0	

TEST INITIATION

Date	Initials	Test ID	# Organisms used	Comments
28APR94	CRW	T-	750	COE - Humboldt project
		T-		
		T-		
		T-		
		T-		

11/1091474

ToxScan, Inc.  
Receiving, Evaluation and Acclimation Log Worksheet  
REAL

Species: <u>Mytilus edulis</u>	Supplier: <u>Carlsbad Aquafarm</u>	# Ordered: <u>1 pound</u>
Date Shipped: <u>13 APR 94</u>	Date/Time Received: <u>14 APR 94 0943</u>	Shipped via: <u>FedX 8985447365</u>

INITIAL EVALUATION

Package Intact? <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No	Initials: <u>FB</u>	pH: <u>NA</u> units
Temperature: <u>14.5</u> °C	Dissolved Oxygen: <u>NA</u> mg/l	Sal./Cond.: <u>NA</u> ‰/μmhos
Estimated # received: <u>100+</u>	# Dead: <u>0</u>	

Organism condition/comments: Arrived dry on blue ice. Cracked to open to set gonads  
5 ♀: 1 poor, 4 good, 1 ♂ fair. ~2 ♂ spawned during acclimation, spawners removed from tank  
Tank temperature turned down to prevent further spawning. Monitored closely for spawning  
activity. Lots of spawning. floated tobs till spawning stopped. Discarded gonads and rinsed  
animals before submerging tobs again. Monitored closely. No more spawning observed

HOLDING/ACCLIMATION

Date	Initials	pH (units)	Temp (°C)	D.O. (mg/l)	Sal/Cond (‰/μmhos)	Food	# Dead	Comments
4 APR 94	FB	7.7	15.2	7.6	33.2	-	0	holding tank conditions
15 APR 94	JC	7.5	14.3	7.7	33.8	-	0	
16 APR 94	JC	7.5	14.3	7.7	33.8	-	0	
17 APR 94	CRW	7.5	14.4	7.7	33.9	-	0	
19 APR 94	FB	7.5	14.4	7.8	33.9	-	0	
20 Apr 94	TS	7.7	14.3	7.9	33.8	-	0	
↳ Termination								

TEST INITIATION

Date	Initials	Test ID	# Organisms used	Comments
4/20/94	PL	T-10774	≈ 40	Successful Spawn
		T-		
		T-		
		T-		
		T-		



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 USE THE INTERNATIONAL AIR WAYBILL FOR SHIPMENTS TO PUERTO RICO AND ALL NON U.S. LOCATIONS.  
 QUESTIONS? CALL 800-238-5355 TOLL FREE.

AIRBILL  
 PACKAGE  
 TRACKING NUMBER

8785447365

8785447365

**RECIPIENT'S COPY**

From (Your Name) Please Print [Redacted]		Date 4/10/94		To (Recipient's Name) Please Print [Redacted]		Recipient's Phone Number (Very Important) [Redacted]	
Your Phone Number (Very Important) [Redacted]		Department/Floor No. [Redacted]		Company TASCAN		Department/Floor No. [Redacted]	
Company [Redacted]		Street Address [Redacted]		Exact Street Address (We Cannot Deliver to P.O. Boxes or P.O. Zip Codes.) 12 HARRISON WAY		State FL	
City [Redacted]		State FL		City WATSONVILLE		State CA	
ZIP Required [Redacted]		ZIP Required [Redacted]		ZIP Required [Redacted]		ZIP Required [Redacted]	
YOUR INTERNAL BILLING REFERENCE INFORMATION (optional) (First 24 characters will appear on invoice.) [Redacted]						IF HOLD AT FEDEX LOCATION, Print FEDEX Address Here Street Address [Redacted]	
CITY [Redacted]		STATE FL		CITY [Redacted]		STATE CA	
ZIP Required [Redacted]		ZIP Required [Redacted]		ZIP Required [Redacted]		ZIP Required [Redacted]	
PAYMENT 1 <input type="checkbox"/> Bill Sender 2 <input checked="" type="checkbox"/> Bill Recipient's FedEx Acct. No. 3 <input type="checkbox"/> Bill 3rd Party FedEx Acct. No. 4 <input type="checkbox"/> Bill Credit Card		5 <input type="checkbox"/> Cash/Check		SERVICES (Check only one box)		DELIVERY AND SPECIAL HANDLING (Check services required)	
11 <input type="checkbox"/> OTHER PACKAGING 16 <input type="checkbox"/> FEDEX LETTER 12 <input type="checkbox"/> FEDEX PAK * 13 <input type="checkbox"/> FEDEX BOX 14 <input type="checkbox"/> FEDEX TUBE		Standard Overnight (Delivery by next business afternoon No Saturday delivery) 51 <input checked="" type="checkbox"/> OTHER PACKAGING 56 <input type="checkbox"/> FEDEX LETTER * 52 <input type="checkbox"/> FEDEX PAK * 53 <input type="checkbox"/> FEDEX BOX 54 <input type="checkbox"/> FEDEX TUBE		Weekday Service 1 <input type="checkbox"/> HOLD AT FEDEX LOCATION WEEKDAY (Fill in Section H) 2 <input type="checkbox"/> DELIVER WEEKDAY Saturday Service 31 <input type="checkbox"/> HOLD AT FEDEX LOCATION SATURDAY (Fill in Section H) 3 <input type="checkbox"/> DELIVER SATURDAY (Extra charge) (Not available to all locations) 9 <input type="checkbox"/> SATURDAY PICK-UP (Extra charge)		Special Handling 4 <input type="checkbox"/> DANGEROUS GOODS (Extra charge) 6 <input type="checkbox"/> DRY ICE (Dangerous Goods Shipper's Declaration not required) DIM SHIPMENT (Chargeable Weight) L x W x H [Redacted]	
Economy Two-Day (Delivery by second business day **) 30 <input type="checkbox"/> ECONOMY * * Economy Letter Rate not available. Minimum charge: One pound. Economy rate.		Government Overnight (Restricted for authorized users only) 46 <input type="checkbox"/> GOV'T LETTER 41 <input type="checkbox"/> GOV'T PACKAGE		Freight Service (for packages over 150 lbs.) 70 <input type="checkbox"/> OVERNIGHT FREIGHT ** (Confirmed reservation required) 80 <input type="checkbox"/> TWO-DAY FREIGHT **		Received At 1 <input type="checkbox"/> Regular Stop 3 <input type="checkbox"/> Drop Box 2 <input type="checkbox"/> On-Call Stop 4 <input type="checkbox"/> B.S.C. 5 <input type="checkbox"/> Station	
** Delivery commitment only, be held in former office.		** Declared Value Limit \$500.		Release Signature: [Signature]		Emp. No. _____ Date _____ <input type="checkbox"/> Cash Received <input type="checkbox"/> Return Shipment <input type="checkbox"/> Third Party <input type="checkbox"/> Chg. To Del. <input type="checkbox"/> Chg. To Hold Street Address _____ City _____ State _____ Zip _____ Received By: X Date/Time Received _____ FedEx Employee Number _____	

REVISION DATE 12/92  
 PART #137204 EXEM.  
 FORMAT #158

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ToxScan, Inc.  
Receiving, Evaluation and Acclimation Log Worksheet  
REAL

Species: <u>Rhepoxynius abronius</u>	Supplier: <u>NAS</u>	# Ordered: <u>900</u>
Date Shipped: <u>4-18-94</u>	Date/Time Received: <u>19 APR 94 1000h</u>	Shipped via: <u>FED. EX.</u>

**INITIAL EVALUATION**

Package Intact? <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No	Initials: <u>PS</u>	pH: <u>N/A sediment</u> units
Temperature: <u>12.5</u> °C	Dissolved Oxygen: <u>N/A</u> mg/l	Sal./Cond.: <u>30</u> %/µmhos
Estimated # received: <u>900 + 10%</u>	# Dead: <u>4 after immersing in SW</u>	
Organism condition/comments: <u>Animals acclimated to full-strength S.W. at 14.8°C. in 30 minutes.</u>		

**HOLDING/ACCLIMATION**

Date	Initials	pH (units)	Temp (°C)	D.O. (mg/l)	Sal/Cond (%/µmhos)	Food	# Dead	Comments
21 APR 94	AB	8.1	15.7	7.8	33.2	—	3	
22 APR 94	AB	8.2	15.4	7.8	33.9	—	0	lowered SW. to 33% using 16°C MH water.
23 APR 94	CRW	8.0	15.5	7.7	33.0	—	6	
24 APR 94	FB	8.1	15.5	7.7	33.1	—	4	
25 APR 94	CRW	8.1	15.6	7.7	33.0	—	2	
26 APR 94	CRW	8.1	15.4	7.7	33.5	—	3	
27 APR 94	PS	8.0	15.4	7.8	34.0	—	0	
28 APR 94	PS	8.1	15.5	7.6	34.1	—	0	
29 APR 94	ASB	8.0	15.6	7.7	34.0	—	0	
29 APR 94	PS	1534h	Animals (few) terminated after successful Ref. Tox. test.					

**TEST INITIATION**

Date	Initials	Test ID	# Organisms used	Comments
25 APR 94	PS	T-10774	600	Set up Home, Ref, SAM6 & EK <del>and</del> and Cd ref tox but not FL Home-Renew or NH <sub>3</sub> Ref. Tox.
26 APR 94	TS	T-10774	220	FL/Home-Renew ↑ NH <sub>3</sub> Ref. Tox. not run — PS

29 APR 94



SUBJECT: Animal Collection Data Sheet			
SOLD TO: <i>TOSLAN</i> <i>42 HANCOCK WAY</i> <i>WATSONVILLE, CA 95076</i>		ATTN: <i>Peter Shafer</i> <i>(408) 724-4522</i>	
DATE OF SHIPMENT: <i>4-18-94</i>			
ANIMAL HISTORY			
Species	Age/Size	Number Shipped	
<i>Rleporxymus abronius</i>	<i>adult</i>	<i>900 (+10%)</i>	
WATER QUALITY AT TIME OF SHIPMENT			
<input checked="" type="checkbox"/> Temperature	<i>13.8°C</i>	Hardness	_____
<input checked="" type="checkbox"/> Salinity	<i>30ppt</i>	Alkalinity	_____
Conductivity	_____	pH	<i>7.9</i>
<input checked="" type="checkbox"/> DO	<i>8.8 mg/L</i>	other:	_____
PACKAGED BY: <i>Carri Bullock</i>		DATE: <i>4-18-94</i>	
FIELD COLLECTION/CULTURE NOTES: <i>Collected on 4-11-94 at West Beach, whidbey IS. wt. No collection site data available.</i>			
ADDITIONAL COMMENTS: <i>2 Liters of sediment included.</i>			

Please call Linda Garrison at (503) 265-7225 if you have any questions.



## Appendix F

### Chains of Custody



COMPANY NAME: Kinnetic Laboratories Inc  
 ATTN: Tasha Granda  
 ADDRESS: 307 Waskington St.  
 Santa Cruz Ca 95060  
 PHONE: (408) 457-3950  
 FAX:   
 PROJECT NAME: Humboldt  
 PROJECT NUMBER:   
 SEND INVOICE TO:   
 P.O. / CONTRACT NO:   
 COMMENTS / SPECIAL INSTRUCTIONS:  
 → ToxScan Biosci  
 Division

TOXSCAN INC.  
 42 Hangar Way  
 Watsonville, CA 95076  
 PHONE: 408/724-4522  
 FAX: 408/724-3188

LAB USE ONLY:  
 STORAGE LOCATION  
 FREEZER #  
 REFRIGERATOR # 11  
 SHELF # 12

ANALYSIS(ES) REQUESTED	CHECK THE APPROPRIATE BOX BELOW:	SAMPLE CONDITION
Sediment PS Donly Sediment Chloristry	<input checked="" type="checkbox"/>	T-10774-01
	<input checked="" type="checkbox"/>	T-10774-02
	<input checked="" type="checkbox"/>	T-10774-03
	<input checked="" type="checkbox"/>	T-10774-04
	<input checked="" type="checkbox"/>	T-10774-05
	<input checked="" type="checkbox"/>	T-10774-06
	<input checked="" type="checkbox"/>	T-10774-07
	<input checked="" type="checkbox"/>	T-10774-08
	<input checked="" type="checkbox"/>	T-10774-09
	<input checked="" type="checkbox"/>	T-10774-10

Lab-Use-Only ID-Number Sample I.D.	Client-Sample Identification Site I.D.	Sample Information:			Bottle or Container Information:			No. of Bottles:
		Sampling Date	Sampling Time	Sample Matrix	Sample Preservative	Bottle Type:	Bottle Size:	
UM1945ED0001	SAM-7D	03/31/94		Sediment		Glass	1 liter	1
UM1945ED0002	SAM-5D	03/31/94						
UM1945ED0003	SAM-4D	03/31/94						
UM1945ED0004	SAM-3D	03/31/94						
UM1945ED0005	SAM-2D	03/31/94						
UM1945ED0006	SAM-1D	03/31/94						
UM1945ED0007	SAM-6C	03/31/94						
UM1945ED0008	SAM-6B	03/31/94						
UM1945ED0009	SAM-6A	04/01/94						
UM1945ED0010	SAM-4(D)	04/01/94						

SAMPLER'S SIGNATURE AND PRINTED NAME:

RELINQUISHED BY (SIGNATURE AND PRINTED NAME):	RECEIVED BY (SIGNATURE AND PRINTED NAME):	DATE:	TIME:
	<i>Michael Seiberg</i>	03/04/94	

Internal C.O.C.

T-10774

COMPANY NAME: Kinneth Laboratories Inc.  
 ATTN: Tasha Grand.  
 ADDRESS: 307 Washington St.  
 Santa Cruz Ca 95060  
 PHONE: (408) 457-3950  
 FAX:  
 PROJECT NAME: Humboldt  
 PROJECT NUMBER:  
 SEND INVOICE TO:  
 P.O. / CONTRACT NO:

TOXSCAN INC.  
 42 Hangar Way  
 Watsonville, CA 95076  
 PHONE: 408/724-4622  
 FAX: 408/724-3188

LAB USE ONLY  
 STORAGE LOCATION  
 FREEZER #:  
 REFRIGERATOR #:  
 SHELF #:

LAB USE ONLY ID Number Sample I.D.	Client Sample Identification Site I.D.	Sample Information:			Bottle or Container Information:			No. of Bottles:
		Sampling Date	Sampling Time	Sample Matrix	Sample Preservative	Bottle Type:	Bottle Size:	
HUM945ED0011	SAM-6 Composite	04/01/94		Sediment		Glass	1L-ev	1
HUM945ED0012	NB-10	04/01/94						
HUM945ED0013	NB-9	04/01/94						
HUM945ED0014	NB-8	04/01/94						
HUM945ED0015	NB-7	04/01/94						
HUM945ED0016	NB-6	04/01/94						
HUM945ED0017	NB-5	04/01/94						
HUM945ED0018	NB-4 Rep 1	04/01/94						
HUM945ED0019	NB-3	04/01/94						
HUM945ED0020	NB-1	04/01/94						

ANALYSIS(ES) REQUESTED: Sediment Chemistry, Sediment PSD only

CHECK THE APPROPRIATE BOX BELOW:

SAMPLER'S SIGNATURE AND PRINTED NAME:

RELIQUISHED BY (SIGNATURE AND PRINTED NAME):  
 RECEIVED BY (SIGNATURE AND PRINTED NAME):  
 DATE: 04/01/94  
 TIME:

COMPANY NAME: Kinnethick Laboratories Inc  
 ATTN: Tasha Gravel  
 ADDRESS: 307 Washington St.  
 Santa Cruz, Ca 95060  
 PHONE: (408) 457-3950  
 FAX:

COMMENTS / SPECIAL INSTRUCTIONS:  
 In Internal C.O.C.  
 ANALYSIS(ES) REQUESTED  
 Sediment Chemistry  
 Sediment PSD only

TOXSCAN INC.  
 42 Hangar Way  
 Watsonville, CA 95076  
 PHONE: 408/724-4522  
 FAX: 408/724-3188

LAB USE ONLY:  
 STORAGE LOCATION  
 FREEZER #  
 REFRIGERATOR #  
 SHELF # 12

LAB USE ONLY	STORAGE LOCATION	FREEZER #	REFRIGERATOR #	SHELF #	SAMPLE CONDITION
					10774-21
					10774-22
					10774-23
					10774-24
					10774-25
					10774-26
					10774-27
					10774-28
					10774-29
					10774-30

Lab Use Only	ID Number	Client Sample Identification	Sample Information:			Bottle or Container Information:			No. of Bottles
			Sampling Date	Sampling Time	Sample Matrix	Sample Preservative	Bottle Type	Bottle Size	
	Sample I.D.	Site I.D.	04/01/94		Sediment	Glass	1-Liter	1	
	HUM94SE0001	NBY rep 2	04/02/94						
	HUM94SE0002	EK-1	04/02/94						
	HUM94SE0003	EK-2	04/02/94						
	HUM94SE0004	EK-3	04/02/94						
	HUM94SE0005	EK-4	04/02/94						
	HUM94SE0006	EK-Composite	04/02/94						
	HUM94SE0007	FL-8	04/03/94						
	HUM94SE0008	FL-7	04/03/94						
	HUM94SE0009	FL-6	04/03/94						
	HUM94SE0010	FL-5	04/03/94						

SAMPLER'S SIGNATURE AND PRINTED NAME:

RELINQUISHED BY (SIGNATURE AND PRINTED NAME):	RECEIVED BY (SIGNATURE AND PRINTED NAME):	DATE:	TIME:
	Michael Sealey Michael Sealey	04/04/94	





**COMPANY NAME:** ToxScan Inc.  
**ATTN:** Mary Lou Milazzo  
**ADDRESS:**  
**PHONE:** 408 / 724-4522  
**FAX:** 408 / 724-3188  
**PROJECT NAME:** Humboldt Harbor  
**PROJECT NUMBER:**

**COMMENTS / SPECIAL INSTRUCTIONS:**  
 DL = 1 part per trillion  
 TAT = 3 weeks (by 29 April '94)  
 QC = 1 LCS, 1 blank

**SEND INVOICE TO:**  
**P.O. / CONTRACT NO:** 091680

**TOXSCAN INC.**  
 42 Hangar Way  
 Watsonville, CA 95076  
**PHONE:** 408/724-4522  
**FAX:** 408/724-3188

**LAB USE ONLY:**  
**STORAGE LOCATION:**  
**FREEZER #:**  
**REFRIGERATOR #:**  
**SHELF #:**

**ANALYSIS(ES) REQUESTED:**  
 TCDD/TCDF only by EPA 8290

Lab Use Only OXS ID Number Reference ID	Client Sample Identification Site I.D.	Sample Information:		Bottle or Container Information:			No. of Bottles:	CHECK THE APPROPRIATE BOX BELOW:
		Sampling Date	Sampling Time	Sample Matrix	Sample Preservative	Bottle Type:		
10774-32	SAM Co Composite	04/01/94		sediment		1L	1	X
10774-33	Reference Composite	04/05/94		↓		↓	1	X
10774-34	FLV Composite	04/04/94		↓		↓	1	X
10774-35	EK Composite	04/02/94		↓		↓	1	X

**SAMPLER'S SIGNATURE AND PRINTED NAME:**  
 Sub contract Laboratory: Altra Analytical 916-933-1640

**RECEIVED BY (SIGNATURE AND PRINTED NAME):**  
 Mary Lou Milazzo via UPS Red Label  
**DATE:** 04/07/94  
**TIME:** 16:05

Please return completed copy of COC to ToxScan. Thank you.



**COMPANY NAME:** Kinnelich Laboratories Inc  
**ATTN:** Tasha Granel  
**ADDRESS:** 307 Washington St.  
 Santa Cruz, Ca 95060  
**PHONE:** (408) 457-3950  
**FAX:**  
**PROJECT NAME:** Humboldt  
**PROJECT NUMBER:**  
**COMMENTS / SPECIAL INSTRUCTIONS:**  
 TOXSCAN BIOASSAY DIVISION  
 →  
**SEND INVOICE TO:**  
**P.O. / CONTRACT NO.:**

**TOXSCAN INC.**  
 42 Hangar Way  
 Watsonville, CA 95076  
**PHONE:** 408/724-4622  
**FAX:** 408/724-3188

**LAB USE ONLY:**  
**STORAGE LOCATION:**  
**FREEZER #:**  
**REFRIGERATOR #:**  
**SHELF #:**

ANALYSIS(ES) REQUESTED	CHECK THE APPROPRIATE BOX BELOW:	SAMPLE CONCENTRATIONS
Sediment Bivalve collection	<input checked="" type="checkbox"/>	70.0000 I.D.
	<input checked="" type="checkbox"/>	T-10774-45-46
	<input checked="" type="checkbox"/>	T-10774-48-50
	<input checked="" type="checkbox"/>	T-10774-51-52
		T-10774-53-54-55

Sample Information:		Bottle or Container Information:		
Lab-Use City: 2014	Offent-Sample - 2014	Sample Matrix	Sample Preservative	No. of Bottles:
IP-Number 2014	Identification - mt			
Sample I.D.	Site I.D.	Sediment	Boiler 68gt	3
HUM94SEED0036	FL-Composite			↓
HUM94SEED0042	Ref-Composite			2
HUM94SEED0048	SAM 6 Composite			3
HUM94SEED0056	EK-Composite			

**SAMPLER'S SIGNATURE AND PRINTED NAME:**  
 \* Under Shelf 6 Coolers 03/02 \* Under Shelf 31 Boiler 53 Under Shelf 6 Cooler 45-46  
 Under Shelf 12 Coolers 03/04 Under Shelf 45 Coolers 54-55 Under Shelf 12 Cooler 47-48  
 Under Shelf 18 Coolers 05/06 Under Shelf 51 Cooler 31-52 Under Shelf 18 Cooler 49-50

**RELINQUISHED BY (SIGNATURE AND PRINTED NAME):**  
 Tasha Granel / Tasha Granel

**RECEIVED BY (SIGNATURE AND PRINTED NAME):**  
 Tasha Granel / Tasha Granel

**DATE:** 04/04-06/94  
**TIME:**



**COMPANY NAME:** Kimmich Laboratories Inc  
**ATTN:** Tasha Grant  
**ADDRESS:** 307 Washington St  
 Santa Cruz Ca 95060  
**PHONE:** 408-457-3950  
**FAX:**  
**PROJECT NAME:** Humboldt  
**PROJECT NUMBER:**  
**SEND INVOICE TO:**  
**P.O. / CONTRACT NO.:**

**COMMENTS / SPECIAL INSTRUCTIONS:**  
 → TOXSCAN BIOASSAY DIVISION  
 Diss Solids

**TOXSCAN INC.**  
 42 Hangat Way  
 Watsonville, CA 96076  
**PHONE:** 408724-4622  
**FAX:** 408724-3188

**LAB USE ONLY:**  
**STORAGE LOCATION:**  
**FREEZER #:** 4  
**REFRIGERATOR #:**  
**SHELF #:**  
**SAMPLE CONDITION:**  
 INTACT

Lab Use Only: ID Number	Client Sample Identification	Sample Information:			Bottle or Container Information:			No. of Bottles:
		Sub Sampling Date	Sampling Time	Sample Type	Sample Preservative	Bottle Type:	Bottle Size:	
T-10774-59	EK-1	04/14/94	10:00	Sediment	Frozen	Glass	125ml	1
T-10774-60	EK-2							
T-10774-61	EK-3							
T-10774-62	EK-4							
T-10774-63	EK Composite							
T-10774-64	SAM-1D							
T-10774-65	SAM-2D							
T-10774-66	SAM-3D							
T-10774-67	SAM-4D							
T-10774-68	SAM-4(D)							

**SAMPLER'S SIGNATURE AND PRINTED NAME:** Michael Selberg  
**RECEIVED BY (SIGNATURE AND PRINTED NAME):** Douglas Clark  
**DATE:** 04/14/94  
**TIME:** 1520

Sub-Sample C.O.C

COMPANY NAME: Kinnestick Laboratories Inc  
 ATTN: Tasha Greenel  
 ADDRESS: 307 Washington St.  
Santa Cruz Ca 95060  
 PHONE: (408) 457-3950  
 FAX:  
 PROJECT NAME: Humboldt  
 PROJECT NUMBER:  
 COMMENTS / SPECIAL INSTRUCTIONS:  
TOXSCAN BIOASSAY DIVISION  
 SEND INVOICE TO:  
 P.O. / CONTRACT NO.:

TOXSCAN INC.  
 42 Hangar Way  
 Watsonville, CA 96076  
 PHONE: 408/724-4522  
 FAX: 408/724-3188  
 LAB USE ONLY:  
 STORAGE LOCATION  
 FREEZER # 4  
 REFRIGERATOR #  
 SHELF #  
 ANALYSIS(ES) REQUESTED  
Diss Sulfides  
 CHECK THE APPROPRIATE BOX BELOW:

Lab Use Only ID Number	Client Sample Identification	Sample Information:			Bottle or Container Information:			No. of Bottles:	SAMPLE CONDITION:
		Sub Sampling Date	Sampling Time	Sample Type	Sample Preservative	Bottle Type:	Bottle Size:		
T-109774-69	SAM-5D	04/14/94	10:00	Sediment	Frozen	Glass	125ml	1	INTACT
T-109774-70	SAM-6A								
T-109774-71	SAM-6B								
T-109774-72	SAM-6C								
T-109774-73	SAM-6 Composite								
T-109774-74	SAM-7D								
T-109774-75	FL-1								
T-109774-76	FL-2								
T-109774-77	FL-3								
T-109774-78	FL-4								

SAMPLER'S SIGNATURE AND PRINTED NAME: Michael Sedberg

RELINQUISHED BY (SIGNATURE AND PRINTED NAME): <u>Michael Sedberg</u>	RECEIVED BY (SIGNATURE AND PRINTED NAME): <u>Douglas Clark</u>	DATE: <u>04/14/94</u>	TIME: <u>1520</u>

COMMENTS / SPECIAL INSTRUCTIONS:

→ TOXSCAN BIOASSAY DIVISION

COMPANY NAME: Kinnierich Laboratories Inc  
 ATTN: Tasha Girard  
 ADDRESS: 307 Washington St  
Santa Cruz Ca 95060  
 PHONE: 408-457-3950  
 FAX:  
 PROJECT NAME: Humboldt  
 PROJECT NUMBER:

SEND INVOICE TO:

P.O. / CONTRACT NO:

ANALYSIS(ES) REQUESTED

Diss Sol/Files

TOXSCAN INC.  
 42 Hangar Way  
 Watsonville, CA 96076  
 PHONE: 408/724-4622  
 FAX: 408/724-3188

LAB USE ONLY:  
 STORAGE LOCATION  
 FREEZER # A  
 REFRIGERATOR #:  
 SHELF #:

Lab Use Only: ID Number	Client Sample Identification	Sample Information:		Bottle or Container Information:			No. of Bottles:	SAMPLE CONDITION:
		Sampling Date	Sampling Time	Sample Type	Sample Preservative	Bottle Type:		
T10774-79	FL-5	04/14/94	10:00	Sediment	Frozen	Glass	125ml	INTACT
T10774-80	FL-6							
T10774-81	FL-7							
T10774-82	FL-8							
T10774-83	FL-Composite							
T10774-84	Reference Composite							
T10774-85	NB-4 Repl							

CHECK THE APPROPRIATE BOX BELOW:

X								
X								
X								
X								
X								
X								
X								

SAMPLER'S SIGNATURE AND PRINTED NAME: Michael Schlegel

RELINQUISHED BY (SIGNATURE AND PRINTED NAME): <u>Michael Schlegel</u>	RECEIVED BY (SIGNATURE AND PRINTED NAME): <u>Douglas Clark</u>	DATE: <u>04/14/94</u>	TIME: <u>1520</u>





SAMPLE LOG-IN CHECKLIST		(CHECK ONE)		YES	NO	N/A
1. Shipping service: UPS Red / UPS Blue / UPS Ground / Federal Express (circle one) courier / delivered by hand / <u>other</u> (describe): <u>GREYHOUND</u>						
Airbill number (write the number in this space):						
2. Is the shipping container intact? <span style="float: right;"><input checked="" type="checkbox"/></span>						
Type of shipping container received? cooler / box / none / <u>other</u> (describe): (circle one) <u>BUCKET</u>						
Are custody seals present on the shipping container? <span style="float: right;"><input type="checkbox"/></span>						
If yes, are they intact? <span style="float: right;"><input type="checkbox"/></span>						
Shipping preservation (circle one): water ice / dry ice / blue ice / <u>none</u> / other (describe):						
Shipping container temperature (record on this line in °C): <u>A/Dc 15.0°</u>						
3. Is a chain of custody (COC) form present? <span style="float: right;"><input checked="" type="checkbox"/></span>						
Are there discrepancies between the COC and the samples? (If yes, describe on COC or below.) <span style="float: right;"><input type="checkbox"/></span>						
4. Are the sample containers intact? (If no, describe on COC or below.) <span style="float: right;"><input checked="" type="checkbox"/></span>						
Are custody seals present on the sample containers? <span style="float: right;"><input type="checkbox"/></span>						
If yes, are they intact? (If no, describe on COC or below.) <span style="float: right;"><input type="checkbox"/></span>						
5. Liquid samples only: Are the samples appropriately preserved? <span style="float: right;"><input checked="" type="checkbox"/></span>						
Liquid sample pH verification (10% of samples):		(circle one or write-in the pH)				
ToxScan ID:		pH<2	pH>12	pH=		
ToxScan ID:		pH<2	pH>12	pH=		
ToxScan ID:		pH<2	pH>12	pH=		
ToxScan ID:		pH<2	pH>12	pH=		
ToxScan ID:		pH<2	pH>12	pH=		
6. Date shipping container returned (write-in date or N/A): <u>N/A</u>						
NOTES / COMMENTS:						
For Samples Preserved by Sample Acquisition Staff (write-in the appropriate information):						
Chemical Preservative:	Sample ID #'s preserved:	Lot # of Chemical Preservative:				
Signature: <u>Peter Schafer</u>						Date Received: <u>27 APR 94</u>

COMPANY NAME: TOXSCAN BIOTASSAY DIV.  
 ATTN: RAY MARICEL  
 ADDRESS: 42 HANGAR WAY  
 WATSONVILLE CA 95076  
 PHONE: 724-4522  
 FAX:  
 PROJECT NAME: HUMBOLDT C.O.C.  
 PROJECT NUMBER:  
 COMMENTS / SPECIAL INSTRUCTIONS:  
 SUB-SAMPLED FROM A COMPOSITE  
 OF T-10774-86 + -87  
 \* SUBCONTRACTED TO ALTA ANALYTICAL  
 SEND INVOICE TO:  
 P.O. / CONTRACT NO:

ANALYSIS(ES) REQUESTED  
 TIKR II CHEMISTRY \*  
 TCDD / TCDF \*  
 TOXSCAN INC.  
 42 Hangar Way  
 Watsonville, CA 95076  
 PHONE: 408/724-4522  
 FAX: 408/724-3188  
 LAB USE ONLY:  
 STORAGE LOCATION  
 FREEZER #:  
 REFRIGERATOR # 11  
 SHELF # 47

Lab Use Only ID Number	Client Sample Identification	Sample Information:		Bottle or Container Information:			No. of Bottles:	CHECK THE APPROPRIATE BOX BELOW:	SAMPLE CONDITION:
		Sub. Sampling Date	Sub. Sampling Time	Sample Type	Sample Preservative	Bottle Type:			
T-10774-88	HOME SEDIMENT	27 APR 94		SEDIMENT	NONE	G/J	1 LITER	✓	INTACT
T-10774-89	HOME SEDIMENT							✓	

SAMPLER'S SIGNATURE AND PRINTED NAME: Peter Schafar Peter Schafar Peter Schafar

RELINQUISHED BY (SIGNATURE AND PRINTED NAME):	RECEIVED BY (SIGNATURE AND PRINTED NAME):	DATE:	TIME:

COMMENTS / SPECIAL INSTRUCTIONS:

COMPANY NAME: TOXSCAN BIOASSAY DIV.  
 ATTN: RAY MARTEL  
 ADDRESS: 42 HANGAR WAY  
 WATSONVILLE CA 95076  
 PHONE: 724-4522  
 FAX:

PROJECT NAME: HUMBOLDT C.O.E.  
 PROJECT NUMBER:

SEND INVOICE TO:  
 P.O. / CONTRACT NO:

ANALYSIS(ES) REQUESTED

DISOLVED SULFIDES

TOXSCAN INC.  
 42 Hangar Way  
 Watsonville, CA 95076  
 PHONE: 408/724-4522  
 FAX: 408/724-3188

LAB USE ONLY:  
 STORAGE LOCATION  
 FREEZER # 4  
 REFRIGERATOR #  
 SHELF #

Lab Use Only ID Number	Client Sample Identification	Sample Information:		Bottle or Container Information:			No. of Bottles:	CHECK THE APPROPRIATE BOX BELOW:	SAMPLE CONDITION:
		Sampling Date	Sampling Time	Sample Type	Sample Preservative	Bottle Type			
F-10774-90	HOME SEDIMENT	03 MAY 94	1130	SEDIMENT	FROZEN	G/J	125 ml	1	INTACT

SAMPLER'S SIGNATURE AND PRINTED NAME: *Sut Sampled by M. Lee R. Kelly* 05/03/94

RELINQUISHED BY (SIGNATURE AND PRINTED NAME):	RECEIVED BY (SIGNATURE AND PRINTED NAME):	DATE:	TIME:





TEL (408) 724-4522  
 FAX (408) 724-3188

Show this Purchase Order Number  
 on all correspondence, invoices,  
 shipping papers and packages.

09723

TO

Alta Analytical

916.933.1640

DATE 04/28/94	REQUISITION NO. T-10774
SHIP TO Marylou Milazzo.	

QUANTITY	DESCRIPTION	UNIT	PRICE	TOTAL
1	EPA 8290 for TCDD/TCDF only		800 -	800 -
	D.L. = 1 part per trillion			
	QC = LCS, blank.			
	TAT = 3 weeks (by <del>05/19/94</del> ) 05/20/94			

- Please send \_\_\_\_\_ copies of your invoices.
- Order is to be entered in accordance with prices, delivery and specifications shown above.
- Notify us immediately if you are unable to ship as specified.

*M. Schlegel*  
 AUTHORIZED BY  
 RECYCLED PAPER  
 Contents: 40% Pre-Consumer • 10% Post-Consumer

ORIGINAL

**COMPANY NAME:** ToxScan, Inc.  
**ATTN:** Mary Lou Malagz  
**ADDRESS:** 42 Hangar Way  
 Watsonville, CA 95076  
**PHONE:** (408) 724-4522  
**FAX:**  
**PROJECT NAME:** Humboldt C.O.E.  
**PROJECT NUMBER:**

**COMMENTS / SPECIAL INSTRUCTIONS:**  
 DL = 1 part per trillion  
 TAT = 3 weeks 05/20/94  
 QC = 1 LC, 1 blank

**SEND INVOICE TO:**  
**P.O. / CONTRACT NO:** 09723

**TOXSCAN INC.**  
 42 Hangar Way  
 Watsonville, CA 95076  
**PHONE:** 408/724-4522  
**FAX:** 408/724-3188

**LAB USE ONLY:**  
**STORAGE LOCATION:**  
**FREEZER #:**  
**REFRIGERATOR #:**  
**SHELF #:**

**ANALYSIS(ES) REQUESTED:**  
 LCDD/TDF only by EPA 8090

Lab Use Only ID Number	Client Sample Identification	Sample Information:		Bottle or Container Information:			No. of Bottles:	CHECK THE APPROPRIATE BOX BELOW:
		Sampling Date	Sampling Time	Sample Type	Sample Preservative	Bottle Type		
T-10994-89	Control Sediment	04/25/94		Sediment	ON Blue Ice	G/S	1K	X

**SAMPLER'S SIGNATURE AND PRINTED NAME:**

Via UPS Next Day 04/28/94  
 M. Lou Malagz  
 Watsonville, CA

**RELIQUISHED BY (SIGNATURE AND PRINTED NAME):**  
 M. Lou Malagz  
**RECEIVED BY (SIGNATURE AND PRINTED NAME):**  
 M. Lou Malagz

**DATE:** 4-29-94  
**TIME:** 1100

# ALTA Analytical Laboratory

Batch ID: 13413

Sample Log-In Checklist	Yes	No
1. Date Samples Arrived: <u>4-29-94</u> Initials: <u>R.F.</u>		
2. <u>Samples Arrived By:</u> (circle one)    Airborne Express      Federal Express <u>UPS</u> Emery      Freezer Truck      Company Courier      Other _____		
3. Shipping Documentation Present? (circle one)    Shipping Label <u>Airbill</u> Tracking Number <u>02515490112</u>	X	
4. Shipping Container(s) Intact? If no, describe condition below.	X	
5. Custody Seals Present and Intact? If not intact, describe condition below. No. of Seals _____ or Seal No. _____      Type:(circle) Bottle or Container		X
6. Sample Container Intact? If no, indicate sample condition below.	X	
7. Shipping Preservation: (circle one) Ice <u>Blue Ice</u> Dry Ice    None    Temp(°C) <u>17</u>		
8. Chain of Custody (COC) or other Sample Documentation Present?	X	
9. COC/Documentation Acceptable? If no, complete COC Anomaly Form.	X	
10. Shipping Container: (circle one each)    ALTA or <u>Client</u> / <u>Return</u> or Retain		
11. Container and/or Bottles Requested?		X
*12. Drinking Water Sample? If yes, Acceptable Preservation? (circle) Y or N		X

\*Required for HRMS

Name: \_\_\_\_\_ Date Samples Reconciled: \_\_\_\_\_  
(Signature Required for LCMS Only)

Comments:



42 Hangar Way  
WATSONVILLE, CA 95076-2404

TEL (408) 724-4522  
FAX (408) 724-3188

Show this Purchase Order Number  
on all correspondence, invoices,  
shipping papers and packages.

09680

TO

Alta Analytical

916.933.1640

DATE 04/07/94	REQUISITION NO. T-10774
SHIP TO Mary Lou Milazzo	

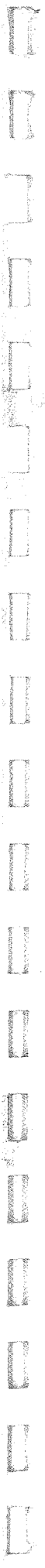
QTY ORDERED	QTY RECEIVED	DESCRIPTION	UNIT PRICE	TOTAL
4		EPA 8290 for TCDD/TCDF only. DL = 1 part per trillion QC = LCS, blank TAT = 3 weeks (by 29 April 1994)	800	3200

1. Please send \_\_\_\_\_ copies of your invoice.
2. Order is to be entered in accordance with prices, delivery and specifications shown above.
3. Notify us immediately if you are unable to ship as specified.

*Mary Lou Milazzo*  
AUTHORIZED BY  
RECYCLED PAPER  
Contents: 40% Pre-Consumer • 10% Post-Consumer

ORIGINAL





1960

1961

1962

1963

1964

1965

**CHEMICAL ANALYSIS, TOXICITY EVALUATION  
AND BIOACCUMULATION EXPOSURE  
OF SEDIMENTS FROM  
HUMBOLDT BAY:**

**BASELINE SURVEY II**

Fiscal Year 1994

**FINAL REPORT**

Prepared for:

**U.S. ARMY ENGINEERING DISTRICT  
SAN FRANCISCO CORPS OF ENGINEERS  
San Francisco, California**

Prepared by:

**TOXSCAN INC. and KINNETIC LABORATORIES, INC.  
Watsonville, California**

**NOVEMBER 1994**