Baylands Ecosystem

Species and
Community Profiles

Life Histories and Environmental Requirements of Key Plants, Fish and Wildlife
Prepared by the San Francisco Bay Area Wetlands Ecosystem Goals Project
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Life Histories and Environmental Requirements of Key Plants, Fish and Wildlife

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San Francisco Estuary Project
c/o S.F. Bay Regional Water Quality Control Board
1515 Clay Street, Suite 1400
Oakland, CA 94612
(510) 622-2465

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Michael Monroe
Carl Wilcox
Resource Managers Group Co-Chairs
San Francisco Bay Area Wetlands Ecosystem Goals Project
Credits

Editor
Peggy R. Olofson — San Francisco Bay Regional Water Quality Control Board

Authors
David G. Ainley
Joy D. Albertson
Janice M. Alexander
Peter R. Baye
Dennis R. Becker
William G. Bousman
Andrée M. Breaux
Michael L. Casazza
Steven C. Chappella
Howard L. Cogswell
Ron Duke
Jules G. Evans
Phyllis M. Faber
Leora Feeney
Steve Foreman
Stephen L. Granholm
Brenda J. Grewell
Janet T. Hanson
Laura A. Hanson
Elaine K. Harding
Bruce Herbold
Catherine M. Hickey
Kathryn A. Hieb
Glen Holstein
Mark R. Jennings
Michael L. Johnson
Robert E. Jones
Kurt F. Kline
Brita C. Larson
Robert A. Leidy
William Z. Lidicker, Jr.
Kevin Mackay
Wesley A. Maffei
Lt. Dante B. Maragni
Carolyn M. Marn
Michael F. McGowan
A. Keith Miles
Michael R. Miller
Gary W. Page

Report Production
Report Design & Layout: Peggy Olofson
Cover Design: Nina Lisowski
Computer & GIS Support: Jeff Kapellas
Text & Copy Editing: Cristina Grosso (lead)
Elisa Gill
Terra Hendrich
Harini Madhavan
Michael Monroe
Dewey Schwarzenberg
Jill Sunahara

Credits i
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Les Chibana les@auc.com
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Josh Collins
Don DesJardin
www.camacdonald.com/birding/DesJardin
Joe DiDonato, bioQuest Wildlife Photography and Consulting (510)769-9209
Dr. Richard B. Forbes
Dr. Allan Francis garter.snake@virgin.net
Rick A. Fridell nrdwr.rfridell@state.ut.us
S.H. Hinshaw
Dan Holland
Jimmy Hu
www.si.com/~njhwa/otter/otter.html
Marshall Iliff miliff@aol.com

Dr. J.L. King
Peter LaTourrette www.birdphotography.com
Denise Loving snakelady@home.com
Wesley Maffei midge@lanminds.com
Peter Moyle, from Inland Fishes of California, copyright 1976. Permission by the Regents of the University of California
Ruth Pratt
T. Douglas Rodda sawwhet@ix.netcom.com
M. Roper, Aquatic Research Organizations, Hampton, N H (1991)
Tom Rountree
Brad Shaffer
Ted Sommer
Rick Stallcup
Dr. Daniel Sudia dan.sudia@mindspring.com
U.S. Fish and Wildlife Service (USFW S)
U.S. Geological Survey, National Biological Service (USG S/N B S)
Staffan Vilcans
Jens V. Vindum, California Academy of Sciences
Bob Walker
Peter S. Weber www.wildbirdphotos.com
Zoological Society of Milwaukee County, Wisconsin www.zoosociety.org
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Introduction

The San Francisco Bay Area Wetlands Ecosystem Goals Project began in 1995 as a cooperative effort among nine state and federal agencies and nearly 100 Bay Area scientists. The Project's purpose was to develop a vision of the kinds, amounts, and distribution of habitats needed to sustain healthy populations of fish and wildlife in and around San Francisco Bay (Figure 1). This vision was presented to the public in the Goals Project's final report, the Baylands Ecosystem Habitat Goals.

Developing the habitat goals involved several steps, many of which were carried out by teams of scientists. First, each team selected "key" animal species (or plant communities, in the case of the Plants Focus Team). Then they compiled available information regarding each species' historic and modern distribution, use of habitats, migration, relationship and interaction with other species, conservation and management issues, and research needs. When time and data were available, some team members compiled additional information on life history and other relevant topics. The teams then discussed the habitat needs of their species and developed initial habitat recommendations. Ultimately, the habitat recommendations of all the teams were integrated to form the Project's final recommendations.

Compiling the information on key species and plant communities into "profiles" was a crucial step in developing the habitat goals. Sharing these profiles enabled team members to better understand the habitat needs of a large proportion of the bayland's flora and fauna. It also facilitated the development of more balanced and diverse habitat recommendations.

When Project participants began sharing the profiles, they realized that much of the information had never before been compiled. They also recognized that, although some of the profiles were not comprehensive, other researchers interested in the Bay and its watersheds might find them useful.

The intent of this report is to provide useful information to those working to restore the baylands ecosystem. However, because the profiles were compiled to inform a specific process, this report should not be considered a complete treatise. Rather, it should be seen as a reference and a starting point. Contact information for the profile authors is included in Appendix A for the reader who would like additional information or clarification, or who would just like to continue the process of scientific discussion and discovery.

Copies of the Baylands Ecosystem Habitat Goals report may be obtained from the San Francisco Estuary Project at the address indicated in the front of this report.

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1 Copies of the Baylands Ecosystem Habitat Goals report may be obtained from the San Francisco Estuary Project at the address indicated in the front of this report.
Figure 1. The Project Area of the San Francisco Bay Area Wetlands Ecosystem Goals Project – The baylands and adjacent and associated habitats of the San Francisco Bay-Delta Estuary, downstream of the Sacramento-San Joaquin Delta
Chapter 1 — Plant Communities

1

Plant Communities

Plants of Shallow Subtidal Habitat and Tidal Flats
(with an emphasis on eelgrass)
Laura A. Hanson

Introduction
There are about 200,000 acres of shallow subtidal habitat and tidal flats in San Francisco Bay, San Pablo Bay, and Suisun Bay. Of this area, approximately 171,000 acres are subtidal habitat and about 29,000 acres are tidal flats. While relatively simple in terms of species diversity, the plant communities that occur in these areas are important components of the estuarine ecosystem.

Although this paper describes the plant communities of shallow subtidal habitat and tidal flats, it focuses on the eelgrass (Zostera marina) community. For more detailed information on the other plant communities (primarily microalgae and macroalgae) that occur in the shallow subtidal areas and on tidal flats of the San Francisco Bay Estuary, please refer to Silva (1979), Nichols and Pamatmat (1988), Meiorin et al. (1991), and Herbold et al. (1992).

Environmental Setting
Shallow subtidal areas and tidal flats are defined by their elevation in relation to tidal height. The shallow subtidal range includes the areas between mean lower low water (MLLW) and the approximate bathymetric contour 18 feet below MLLW. Tidal flats generally occur between the mean tide level (MTL), or the lower elevation limit of Spartina (cordgrass) flats, to about 2.5 feet below MLLW. Tidal flat composition can include various combinations of clay, silt, sand, shell fragments, and organic debris. Daily tidal cycles submerge and expose tidal flat surfaces twice every 24.8 hours. During each tidal cycle, tidal flats are also exposed to fluctuating wave action, current velocities, and nutrient supply. Where tidal marshes still exist, incoming tides flood into the upper marsh areas. As these tidal waters recede, organic materials are transported downslope to tidal flats where they become food sources for millions of detritus-feeding invertebrates.

The environmental conditions of shallow subtidal areas and tidal flats are strongly influenced by suspended sediments. In general, the San Francisco Bay Estuary has high concentrations of suspended sediments (Hanson and Walton 1988). This suspended particulate matter is comprised of 70 - 97% non-organic sediment made up of silty clay; the remaining content is comprised of living and other organic matter (Conomos and Peterson 1977). Suspended sediment concentrations are influenced by wind speed, substrate, particle size, wave action, current velocity, tidal action, water depth and seasonal runoff (Cyrus and Blaber 1987). Human activities such as type of land use (Kemp et al. 1983), channel dredging (LaSalle 1988, Hanson and Walton 1988), construction and use of marinas and ferry terminals, and propeller wash (Walker et al. 1989, Thom and Shreffler 1995) can also affect water clarity.

Total suspended solids (TSS) in Suisun and San Pablo bays average between 50 mg/l in the summer to 200 mg/l in the winter (Nichols and Pamatmat 1988). In North Bay and Central Bay, tides can have a significant influence on sediment resuspension, particularly during spring tides and during the ebbs preceding lower low water when the current speeds are highest. Central Bay — characterized by cold, saline, and relatively clear ocean water — has the lowest TSS concentrations, at 10 to 60 mg/l. South San Francisco Bay has slightly higher TSS concentrations than Central Bay (O’Connor 1991).

Salinity levels vary depending on season, weather, amount of diverted fresh water, and location in the Bay. In general, salinity levels within the water column and within tidal flats increase along a gradient from the Delta to the Golden Gate. For example, the salinity in Suisun Bay averages about seven parts per thousand (ppt), and in Central Bay it averages about 30 ppt (Fox et al. 1991). During dry years, South Bay averages salinity levels up to 35 ppt.
Intertidal and Subtidal Plant Communities

The shallow subtidal areas and tidal flats of the San Francisco Bay Estuary support relatively few plant communities. These communities include diatoms and other microalgae, macroalgae, and eelgrass.

Microalgae form the basis for the estuarine food web. These algae, consisting of diatoms and blue-green algae, often form dense patches on tidal flats, creating a brown hue to the substrate surface during low tide. Microalgae and settled phytoplankton represent a readily available food source for creatures, such as worms and clams, within the mudflats (Nichols and Pamatmat 1988). Shorebirds and waterfowl then consume these creatures.

Macroalgae (seaweeds) are also found throughout the Estuary, particularly in the more saline areas. Few macroalgae can make the necessary adjustments in internal water and mineral content to survive at low salinity levels. The exceptions include Gracilaria sjoestedtii, Enteromorpha spp., and the closely related Ulva spp. G. sjoestedtii is usually found from the mid-intertidal to the shallow subtidal zone attached to rocks partially buried in coarse sand. It also grows attached to small bits of clam and oyster shell in muddy portions of the Bay. In such situations, the plants and associated substrata are easily moved by currents and wave action. Enteromorpha and Ulva form bright green patches and can occur in great abundance throughout the intertidal zone, often growing on any available hard substrate. Enteromorpha can be found occupying higher tidal zones where shade is available. It is especially prevalent on boat hulls, buoys, docks, and woodwork. Ulva occupies the lower tidal zones, completing its life cycle in a few weeks and varying its distribution over a short time period. These kinds of macroalgae often undergo seasonal cycles of abundance, becoming common in the warmer months and virtually disappearing in colder months. Maximum abundance occurs in late summer and early fall (Jones and Stokes Associates, Inc. 1981). Many species of Ulva are often common in heavily polluted areas because they can use ammonia as a nitrogen source and are generally tolerant of organic and metal pollution (Dawson and Foster 1982). In the absence of eelgrass, Ulva can provide a preferred habitat for several invertebrate species (Sogard and Able 1991).

Eelgrass (Zostera marina) is currently the only seagrass found in San Francisco Bay. Belying its common name, it is not a grass but is a flowering plant that has adapted to living submerged in the shallow waters of protected bays and estuaries in temperate regions of the world (Den Hartog 1970, Phillips and Menez 1988). Z. marina reproduces both sexually through pollination of seeds, and asexually through a rhizome meristem that extends through the sediments (Setchell 1929). Here abundant, Z. marina's dense, matted root and rhizome system functions to stabilize the soft bottom. Its leaves slow currents and dampen wave action, causing sediment and organic material to accumulate. Z. marina is found in intertidal areas, becoming exposed during the lower spring tides; it also occurs in subtidal areas at depths less than one to two meters below MLW (Kitting 1994).

Historic and Modern Distribution (of Eelgrass)

Information on historic distribution of Zostera marina in the San Francisco Bay Estuary is very limited. San Francisco Bay may have supported extensive Z. marina meadows in the past. (Setchell 1929, Wylie-Echeverria and Rutten 1989). Low light availability within the water column has been found to limit the development of extensive eelgrass meadows and may be the principal cause of eelgrass decline in San Francisco Bay (Alpine and Cloern 1988, Zimmerman et al. 1991).

In 1989, Wylie-Echeverria and Rutten published the first survey on the distribution of Zostera marina in San Francisco Bay (including San Pablo Bay) and mapped a total of 316 acres (Table 1.1). As Table 1.2 and Figure 1.1 show, the per area abundance of eelgrass within San Francisco Bay is much less than that of Humboldt Bay or Tomales Bay, two other northern California estuaries.

The 1989 Wylie-Echeverria and Rutten survey described the Zostera marina populations as “patchy” and some as “stressed.” Since that time a few of these beds have increased in size, and new patches have been sited (Kitting 1993 and pers. comm.).
Plants

Associated Fauna Including Rare and Sensitive Species

Tidal flats include a living system of diatoms, microalgae, and protozoa that are fed upon by suspension or surface deposit feeding invertebrates. The bottom invertebrates are in turn fed upon by larger consumers such as fish, shrimp, and crabs. During low tide, these primary and secondary consumers are exploited by millions of migratory shorebirds. The extensive intertidal mudflats of San Francisco Bay are considered a key migratory staging and refueling area for over-wintering shorebirds of the Pacific Flyway (Harvey et al. 1992).

Macroalgae and eelgrass provide food, shelter, and spawning grounds for many Bay fish and invertebrates. The major subtidal spawning areas for Clupea harengus (Pacific herring), recently the most valuable fishery in California, are Richardson Bay and the large shallow area between Richmond and Oakland. In these areas, spawning occurs predominantly on Gracilaria spp. and small patches of Zostera marina (Spratt 1981). When available, C. harengus preferentially uses Z. marina habitats for spawning (Taylor 1964, Spratt 1981).

Zostera marina beds support a variety of organisms, more than that of non-vegetated areas (van Montfrans et al. 1984, Kitting 1993, Hanson 1997). Z. marina roots and leaves provide habitat for many plants and animals. For example, the long blade-like shoots provide shelter and serve as a nursery ground for many fish species. Small plants (epiphytes) and animals (epizoites) attach to the leaves, motile animals find cover between the leaves, and burrowing animals live among the roots. Epiphytes are an important part of the eelgrass community, contributing up to 22% of the total primary productivity (Jones 1968, Marsh 1970, Penhale 1977). They, in turn, provide food for resident invertebrate grazers (Kitting et al. 1984). Within the rich organic sediment, anaerobic processes of microorganisms regenerate and recycle nutrients and carbon (Kenworthy et al. 1982).

Because Zostera marina contains noxious sulfated phenolic compounds that can inhibit bacterial degradation and animal grazing, few animals consume it (Tenore 1977, Harrison and Chan 1980, McMillan et al. 1980). Notable exceptions include several species of waterfowl such as Anas americana (wigeon), Anas strepera (gadwall), Anas acuta (pintail), Branta canadensis (Canada goose), and Branta nigricans (black brant) (Phillips 1984). Z. marina has been an obligate food for black brant along its flyway (Einarsen 1965). Black brant populations are in great decline along the Pacific Flyway, possibly due to this species' dependence on dwindling eelgrass resources (Einarsen 1965).

Some bird species also forage on the fauna associated with Zostera marina. An example is the Sterna albifrons browni (California least tern) that was listed as an endangered species in 1970. Least terns are known to forage on juvenile and small fishes (Magenheim and Rubisso 1993) that inhabit Z. marina beds, particu-

<table>
<thead>
<tr>
<th>Location</th>
<th>Acres</th>
</tr>
</thead>
<tbody>
<tr>
<td>San Pablo Bay</td>
<td>124</td>
</tr>
<tr>
<td>Point Orient</td>
<td>3</td>
</tr>
<tr>
<td>Naval Supply Depot</td>
<td>12</td>
</tr>
<tr>
<td>Point Molate Beach</td>
<td>26</td>
</tr>
<tr>
<td>Toll Plaza, East</td>
<td>0.5</td>
</tr>
<tr>
<td>Toll Plaza, West</td>
<td>0.5</td>
</tr>
<tr>
<td>Point Richmond, North</td>
<td>7</td>
</tr>
<tr>
<td>Point Richmond, South</td>
<td>4</td>
</tr>
<tr>
<td>Richmond Breakwater, North</td>
<td>18</td>
</tr>
<tr>
<td>Richmond Breakwater, South</td>
<td>7</td>
</tr>
<tr>
<td>Emeryville</td>
<td>13</td>
</tr>
<tr>
<td>Alameda</td>
<td>55</td>
</tr>
<tr>
<td>Bay Farm, North</td>
<td>2</td>
</tr>
<tr>
<td>Bay Farm, South</td>
<td>4</td>
</tr>
<tr>
<td>Coyote Point</td>
<td>1</td>
</tr>
<tr>
<td>Richardson Bay</td>
<td>13</td>
</tr>
<tr>
<td>Angel Island</td>
<td>3</td>
</tr>
<tr>
<td>Belvedere Cove</td>
<td>5</td>
</tr>
<tr>
<td>Point Tiburon</td>
<td>1</td>
</tr>
<tr>
<td>Keil Cove</td>
<td>10</td>
</tr>
<tr>
<td>Paradise Cove, North</td>
<td>4</td>
</tr>
<tr>
<td>Paradise Cove, South</td>
<td>3</td>
</tr>
<tr>
<td>TOTAL ACRES</td>
<td>316</td>
</tr>
</tbody>
</table>

Table 1.1 Acreage of Individual Eelgrass Beds in San Francisco/San Pablo Bay in 1989

Table from NMFS SW Region. Wyllie-Echeverria and Rutten 1989 Administrative Report SWE-89-05

<table>
<thead>
<tr>
<th>Location</th>
<th>Location (km²)</th>
<th>Extent of Eelgrass (Bottom coverage, acres)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Humbolt Bay</td>
<td>62.4</td>
<td>3,053</td>
<td>Phillips 1984</td>
</tr>
<tr>
<td>Tomales Bay</td>
<td>30.0</td>
<td>965</td>
<td>Spratt 1985</td>
</tr>
<tr>
<td>San Francisco Bay</td>
<td>1,140.0</td>
<td>316</td>
<td>Wyllie-Echeveria 1990</td>
</tr>
</tbody>
</table>

Table 1.2 Comparison of Three Northern California Estuaries Relative to Size of Estuary and Total Acres of Eelgrass (Zostera marina)

Table from NMFS SW Region. Wyllie-Echeveria (1990)
Plants

larly at a major nesting site near the Oakland International Airport and the Alameda Naval Air Station (Collins and Feeney 1983-6, Feeney 1988 and 1989, Harvey et al. 1992).

Invertebrates such as juvenile Cancer magister (Dungeness crab) appear to grow up most successfully in the nursery-like habitat that Zostera marina provides, particularly in the northern reaches of the Bay. The isopod, Synidotea laticauda is periodically found in high numbers (up to 200/m²) among Z. marina beds in Central San Francisco Bay (Hanson 1998). They are an important food item for economically valuable sport fishes such as young striped bass, starry flounder, steelhead trout, king salmon, white sturgeon, plus other fishes in San Francisco Bay (Morris et al. 1980).

The transport of Zostera marina fragments acts as a vector for animal dispersal (Highsmith 1985, Worces- ter 1994). Kitting (1993) found several fish species and a variety of invertebrates usually associated with Z. marina on dead blades found at depths greater than four meters below MLLW.

Conservation Issues

Exotic Plants - There is some potential for two exotic Zostera species to invade San Francisco Bay. The Asian seagrass, Zostera japonica, introduced to British Columbia, Washington, and Oregon has not yet been reported in San Francisco Bay. Z. japonica has a different life history, morphology, and preferred habitat than Z. marina (Harrison and Bigley 1982). Culture experiments determined that Z. japonica is not likely to displace existing Z. marina beds (Harrison 1982). This may not be the case in San Francisco Bay. Z. japonica favors the intertidal zones, the areas where Z. marina has been limited to in San Francisco Bay. Thus far, Z. japonica occupies only a small fraction of its potential habitat in North America, threatening significant changes in the ecology of the intertidal sediments as this seagrass spreads (Harrison and Bigley 1982).

Zostera asiatica is found from Tomales Bay in the north, to Santa Monica Bay in the south. Phillips and Wyllie-Echeverria (1990) published the first record of this species in the Eastern Pacific. It is a wide bladed Zostera that occurs sub-tidally from five meters below MLLW to 17 meters below MLLW. Z. asiatica has not yet been identified in San Francisco Bay. This is probably due to its deeper water distribution where photosynthetic processes could be limited in San Francisco Bay.


In San Francisco Bay, Zostera marina requires somewhere between three and five hours of length of irradiance-saturated photosynthesis (Hₛₑₜ) each day (Zimmerman et al. 1991). In areas with favorable light conditions, Z. marina plants have adequate carbon reserves to withstand at least 30 days of light limitation (Zimmerman et al. 1991); however, due to frequent and persistent periods of high turbidity, it is unlikely that plants at the deeper edge of eelgrass meadows in San Francisco Bay can accumulate large carbon reserves (Zimmerman et al. 1991). Average turbidity of the Bay and, more critically, brief periods of high turbidity limit Z. marina distribution in deeper water and limit establishment of seedlings and vegetative propagules (Zimmerman et al. 1991). If daily, monthly, and seasonal Hₛₑₜ requirements are not met, long-term survival of the plants may be limited (Zimmerman et al. 1991). Any activities that increase turbidity within Bay waters, whether natural or...
anthropogenic, have detrimental effects on existing eelgrass populations and associated food webs.

**Current Restoration Success** — The technology for successfully establishing seagrass beds has been unreliable (Phillips 1974, 1980; Lewis 1987), although, in 1989 Zimmerman et al. (1995) successfully transplanted Zostera marina at two locations in San Francisco Bay. According to Fonseca et al. (1988), waning interest in Z. marina restoration was due to a net loss of habitat through seagrass mitigation projects. Planting projects have often failed as a result of poor selection of planting sites or plant material and incorrect use of planting methods. Factors that limited success include a general lack of knowledge of physiological requirements and unknown local environmental factors controlling Z. marina growth (Lewis 1987, Merkel 1990). For example, in 1984, an eelgrass transplant was initiated in San Francisco Bay. Limited transplant success was attributed to a lack of data on local eelgrass autecology coupled with nearby dredging operations and diminished water quality (Fredette et al. 1988).

**Conclusions and Recommendations**

There has been considerable interest in protecting and expanding existing Zostera marina beds in San Francisco and San Pablo bays (Fredette et al. 1988). Since the 1989 survey, sites have indicated a marked change in the distribution and abundance of this species. Better conserving this species in the Estuary will require more frequent monitoring of individual populations.

It also is imperative to protect the current eelgrass beds from further decline. Because of the inherent difficulties in establishing eelgrass, plantings conducted in exchange for permitted losses (mitigation projects) could result in a greater loss of habitat and should not be allowed. The current Zostera marina populations may be the last remnants in San Francisco Bay and are extremely vulnerable to local extinction (Kitting and Wylie-Echeverria 1991); therefore, plantings should be used to enhance current beds or to create new beds.

There are several actions that should be undertaken when designing potential restoration or enhancement projects:

1. Conduct a thorough survey to assess physical conditions of the site. Collect and evaluate environmental data and/or pilot test the planting at a particular site before commitment of a full restoration project. The success of any seagrass re-vegetation effort, including long-term plant growth, is strictly dependent upon a physical environment suitable for initial establishment (Zimmerman et al. 1991).

2. Carefully evaluate light availability before proceeding with any major transplant effort. Water column turbidity is sufficiently high throughout much of the Central Bay, limiting the euphotic zone (depth where irradiance falls to 1% of surface irradiance) to less than 1 m (Alpine and Cloern 1988).

3. Use stocks for planting from a site with conditions as similar as possible to the planting site. There should be similar or equal water depths, salinity, temperature, tidal currents, wave exposure, and sediment composition (Fonseca 1994). Until we learn more about the genetic structure of this species, matching of phenotypes among restoration and donor sites remains the best guide for stock selection.

4. Limit planting to areas with small tidal ranges rather than high tidal ranges to provide greater light availability (Koch and Beer 1996), thus increasing survival success.

5. Plant in areas where parameters for deeper vertical distribution are available make the bed less vulnerable to adverse conditions (such as storm events or desiccation) due to availability of energy from the neighboring deeper shoots (Tomasko and Dawes 1989).

6. Plant in late spring and summer. Periods of high Z. marina growth and production coincide with warmer temperatures and greater light availability (Ewanchuk and Williams 1996).

**References**


Feeney, L.R. 1988. California least tern breeding season at the Metropolitan Oakland International Airport. Port of O akl and.


Plants


Tidal Marsh Plants of the San Francisco Estuary

Peter R. Baye
Phyllis M. Faber
Brenda Grewell

Introduction

The general ecology of San Francisco Bay has been reviewed by Josselyn (1983), who included a brief treatment of its tidal marsh plant community composition and structure. MacDonald (1977, 1988) reviewed the vegetation of California salt marshes, including San Francisco Bay, with emphasis on sedimentation, drainage, topography, salinity, flooding, community structure, and summaries of autecology of selected dominant species. Newcombe and Mason (1972) made descriptive accounts of the Suisun Marsh area vegetation. Atwater et al. (1979) summarized and interpreted the relationships between tidal marsh vegetation of the San Francisco Estuary and its landforms and geomorphic processes. Wells and Goman (1994) reviewed and expanded the quaternary history of the San Francisco Estuary. The purpose of this plant community profile is to supplement previous reviews, and provide additional information on historic changes in the composition, distribution, and abundance of tidal marsh plants of the Estuary.

Environmental Setting

Prehistoric Tidal Marsh Development - Tidal marshes of the modern San Francisco Estuary formed around 10,000 years ago during the Holocene submergence when the rate of sea-level rise slowed sufficiently for tidal marsh sediments to accrete near sea-level (Atwater et al. 1979). Prior to that time, during the Pleistocene epoch, the site of the modern Estuary consisted of broad stream valleys far above glacial low sea level. Pleistocene tidal marsh plant communities were probably associated with either stream mouths or backbarrier lagoons at the edge of an emergent broad coastal plain, now submerged and eroded or buried offshore from the modern Golden Gate. Tidal marsh plant species probably migrated upstream in valleys and embayments as sea level rose. Ancestral Pleistocene populations of tidal marsh plant species in today's estuaries may not have been as discontinuously distributed as they are today; coastal plain shorelines (e.g., East Coast of North America) often provide widespread tidal inlets and tidal marsh (Davia 1980). Holocene fragmentation of salt marshes from more extensive or continuous Pleistocene coastal plain salt marsh distributions may account for historic disjunct, relict populations of species in San Francisco Bay which are now found only in south-central or southern California tidal marshes (e.g., Solidago confinis (southern goldenrod), Suaeda californica (California sea-blite)).

Tidal marshes of the Estuary fluctuated in vegetation composition during the Holocene epoch, apparently in relation to changes in long-term climate. This is indicated by stratified deposits of fossil pollen and plant fragments which indicate periods of accumulation of plants associated with near-freshwater marsh conditions with species associated the more saline (brackish) conditions (Wells and Goman 1994). These findings are consistent with independent evidence on climate changes during the last 6,000 years which show prolonged periods of drought and high rainfall compared with historic conditions (Ingram et al. 1996, Peterson et al. 1989). The tidal marshes in San Francisco Bay were also not static prior to European influence. Some marsh shoreline configurations indicate long-term scarp retreat across marshes with large sinuous tidal creeks and growth of berms and sand spits (Atwater et al. 1979). Areas of rapid marsh growth in some parts of south San Francisco Bay,
outside the influence of Sierran gold mining and prior to extensive diking, were evident in maps of the Bay prepared in the 1870s (U.S. Coast Survey maps).

**Marsh Sediments and Plants** - Depositional environments of tidal marshes in the San Francisco Estuary are variable and are significant for the distribution of uncommon plant species. In most of the San Francisco Estuary, the sediments of the middle-marsh marsh plain consists of bay mud (fine silt and clay) with significant percentage of organic matter in mature marshes. Local coarse sediment deposits, often beach ridges (marsh berms, or marsh-beach ecotones) composed of sand, shell fragments, organic debris, or mixtures, create physically mobile (periodically eroded and redeposited), well-drained high marsh habitats with affinity for some common high marsh species (e.g., Grindelia stricta var. angustifolia, gumplant) and probably also species now locally extinct or rare, such as Suaeda californica, Atriplex californica (California saltbush), and Castilleja ambiguа (salt marsh owl’s clover or Johnny-nip). Marsh berms are associated with relatively high wave energy environments in the Estuary, located near coarse sediment sources such as eroding bluffs, submerged fossil sand and shell deposits, stream mouths, and eroding marsh edges. Such features were commonly represented on U.S. Coast Survey maps of the mid-1800s, but persist today in very few localities of the Estuary (e.g., Point Pinole, Redwood City area, San Leandro area, and northern San Francisco peninsula). Similar coarse-sediment features probably occurred as natural levees of upstream reaches of large tidal sloughs with significant coarse sediment loads, as is observed today in M orro Bay. Alluvial fans also create gradually sloping ecotones with uplands, with variably textured sediments and freshwater runoff and seeps. Few small alluvial fans exist at tidal marsh edges of the Estuary today (e.g., Point Pinole, Whittell Marsh), but were historically abundant in parts of the Estuary, supporting diverse ecotonal plant communities (Cooper 1926). Analogous alluvial fan-tidal marsh ecotones occur in maritime salt marshes of Point Reyes and Tomales Bay areas, where they support distinctive local plant assemblages, including uncommon to rare species.

**Comparison With Other Estuaries** - The tidal marshes of the San Francisco Estuary are the most extensive on the central coast of California, and their plant communities are distinct from other central coast tidal marshes in many respects. Most other large central coast tidal marshes are associated with shallow embayments with large open tidal inlets (e.g., Tomales Bay, Drakes Estero and Bolinas Lagoon in Marin County; Bodega Bay in Sonoma County; Elkhorn Slough in Monterey County; Morro Bay in San Luis Obispo County) which impose strong marine influence on the character of their sediments, salinities, and vegetation. Central coast tidal marshes tend to be isolated and few because of the steep modern shoreline with few valleys or wave-sheltered bays. These tidal marshes have extensive sandy substrates, relatively small, local inputs of fine sediment and freshwater discharges and brackish (mesohaline) conditions, and are inundated by water approaching marine salinity (34 ppt) during most of the growing season. Some tidal marshes associated with stream mouths have relatively more freshwater influence and brackish marsh vegetation (e.g., pre-historic Elkhorn Slough and Salinas River, Monterey County; Russian River estuary, Sonoma County), but in association with seasonal reduction in tidal influence because of partial or complete closure of coastal inlets at river mouths (dammed by sand beach ridges during periods of relatively low river discharge). In contrast, the tidal marsh plant communities of the San Francisco Bay Estuary developed under conditions of abundant and predominantly fine sediment (bay mud, clayey silts and silty clays with high nutrient-holding capacity), relatively large tidal range, and extensive brackish marshes associated with relatively large freshwater discharges, distributed over broad, fluctuating salinity gradients (Atwater et al. 1979).

Historically, salt pans (unvegetated, seasonally inundated depressions or flats within the tidal marsh) and local salt ponds (perennial deposits of crystalline salt in hypersaline ponds) were well-developed in San Francisco Bay tidal marshes (U.S. Coast Survey T-charts, 1850s), supporting distinctive vegetation (widgeongrass, Ruppia maritima, in some pans) or microalgal floras (in salt ponds). Pans are relatively infrequent in other central coast tidal marshes compared with the historic conditions of the San Francisco Estuary, and natural salt ponds were not known to occur in other central coast tidal marshes. Today, edges of high marsh pans are associated with at least two regionally rare species (Cordylanthus maritimus ssp. palustris and Castilleja ambiguа ssp. ambiguа), and may have been associated with many others in the past (e.g., Lepidium latipes, L. oxyccarpum; Table 1.3).

**Tidal Marsh Plant Communities**

The distribution of tidal marsh plants is strongly (but not exclusively) influenced by tidal elevation and salin-
Chapter 1 — Plant Communities

11

Plants and their associations (Hinde 1954, Atwater and Hedel 1976). Following Peinado et al. (1994), three elevation “zones” of the tidal marsh can be objectively distinguished (and are visually conspicuous): (1) the low marsh zone, occurs from approximately mean sea level to mean high water; (2) the middle marsh zone, occurs from approximately mean high water to mean higher high water; and (3) the high marsh zone (colloquially also called the “upland transition” or “peripheral halophyte” zone; “upper salt marsh zone” of Peinado et al. 1994), occurs near and above mean higher high water up to several meters above extreme high water line (Peinado et al. 1994). The typical species composition of these zones is described below for tidal salt marsh and tidal brackish marsh. Unlike beach, dune, and bluff communities (Barbour and Johnson 1977, Barbour et al. 1973), there is no empirical evidence of salt marsh zonation attributable to salt spray; estuaries and embayments are relatively low-energy wave environments (Davies 1980, Carter 1988). Other potentially significant influencing factors have not yet been well studied.

There are significant floristic differences between the tidal marshes of San Francisco Estuary and other central coast tidal marsh systems. These include:

1. The dominance of Spartina foliosa (Pacific cordgrass), either absent today or historically absent from most or all other central coast tidal marshes (Macdonald 1977);
2. The presence of rare species of disjunct distribution, such as Suaeda californica (native only to Morro Bay and San Francisco Bay); and
3. The presence of local endemic species such as soft bird’s-beak (Cordylanthus mollis ssp. mollis) and Suisun thistle (Cirsium hydrophilum var. hydrophilum).

Conversely, some uncommon tidal marsh species which have either declined severely or become extirpated in the San Francisco Estuary still occur in local abundance in some maritime salt marshes of the region (e.g., Atriplex californica, Castilleja ambigua, Puccinellia nutkanensis). Few species associated with high marsh zones of maritime salt marshes in the region were historically absent from the San Francisco Estuary (e.g., Astragalus pycnostachyus var. pycnostachyus (coastal marsh milkvetch), Castilleja ambigua ssp. humboldtiensis (Humboldt Bay owl’s clover), Leymus x vancouveriensis (Vancouver’s ryegrass), and Grindelia stricta var. stricta (gumplant)).

Differences exist also between the structure of vegetation found in predominantly marine-influenced salt marshes of the central coast and tidal marshes of the San Francisco Estuary. Although the middle marsh zone of San Francisco Bay salt marshes has been described as supporting “prostrate” growth forms of pickleweed (Macdonald 1977), the middle marsh plains of sandy or sandy peat salt marshes of Bolinas Lagoon (Allison 1992), Point Reyes, Tomales Bay, and Morro Bay often support very thin, low (<10 cm) turf-like vegetation mosaics with extremely short, sparse, or prostrate pickleweed as a relatively minor component, or at most co-dominant with species such as Triglochin concinna (slender sea arrowgrass; uncommon to rare in San Francisco Estuary). These salt marsh turfs often support high plant species diversity compared with San Francisco Bay salt marsh plains, which tend to be dominated by pickleweed, which often grows in dense stands (usually over 20 cm thick; up to 50-60 cm in some fringing marshes of San Pablo Bay). Low, turf-like middle marsh vegetation is very uncommon in San Francisco Bay, both in brackish and salt marshes.

Salt Marsh Plants and Their Associations — Salt marsh here refers to tidal marsh plant associations that approximate the species composition typical of near-marine salinity during the growing season (34 ppt). Few if any salt marshes in the San Francisco Estuary are ac-

Pickleweed (Salicornia virginica), a dominant within the salt marsh middle marsh zone, shown enshrouded by parasitic dodder (Cuscuta salina).

Soft bird’s-beak (Cordylanthus mollis ssp. mollis). The hairy bracts of the flowering stems are jeweled with salt-encrusted glands. (Brackish tidal marsh, Southampton Marsh, Benecia)
ualty regularly exposed to near-marine salinity, but in the upper estuarine salinity range (roughly 20 ppt and above), they are effectively salt marsh in vegetation character. The salt marsh plant community is typical of San Francisco Bay and the outer marshes of most of San Pablo Bay.

The low salt marsh zone in San Francisco Bay is usually dominated by a single species, Spartina foliosa (Pacific cordgrass), but is increasingly becoming dominated by the invasive introduced Atlantic species, Spartina alterniflora and its highly variable hybrids and novel “ecotypes” (Callaway and Josselyn 1992; Deahl and Strong 1994, 1997; Deahl et al. 1999). S. foliosa stands occur as uniform fringes along tidal creek banks or as broad uniform plains on prograding marshes at the edges of broad tidal mudflats. They extend from approximately mean high water to mean sea level (Hinde 1954, Atwater et al. 1979). On gentle elevation gradients, they intergrade with middle marsh plains in mixed stands of Salicornia virginica (pickleweed), as at Dumbarton-Mowry marsh and eastern San Pablo Bay fringe marshes. They may also occur as abrupt zones at the edge between tidal mudflats and wave-cut peat scarp.s. Pioneer colonies of Pacific and smooth cordgrasses on mudflats are abundant in some years, particularly in years of high or late rainfall. They apparently establish by seedlings and re-grown fronds, but the relative proportion of these of propagule types is unknown. Pioneer colonies of S. alterniflora were observed on open mudflats of the San Lorenzo Creek delta in 1991, and are visible in aerial photographs of the Alamada Creek area around 1980. Seedlings and pioneer colonies of S. foliosa were common on high mudflats of San Pablo Bay and its tributaries in the late 1990s. The taller S. alterniflora appears to be able to spread clonally below mean sea level, but long-term comparisons of colonial spread between native and introduced cordgrasses have not yet been conducted. The only other species of the low marsh is Salicornia europaea (annual pickleweed), which occasionally occurs in the upper edge of the zone, often in accreting high mudflats in transition between low and middle marsh zones.

The middle salt marsh zone composes the extensive salt marsh plains of San Francisco Bay (Hinde 1954, Atwater et al. 1979). Younger marshes tend to be characterized by low-diversity vegetation dominated by Salicornia virginica (Cuneo 1987), but some older marsh remnants (e.g., Greenbrae and Heerdt marsh; upper Newark slough marsh) may comprise complex and annually variable mosaics of S. virginica, Distichlis spicata (saltgrass), Cuscuta salina (salt marsh dodder), Jaumea carnosa (fleshy jaumea), Frankenia salina (alkali-heath) and Atriplex triangularis (spear scale or fat-hen). Species diversity in the middle salt marsh is not necessarily correlated with marsh age; old marshes at China Camp and Whitney marsh (Point Pinole) also support relatively low-diversity vegetation dominated by S. virginica. The parasitic Cuscuta salina (dodder) can become conspicuously co-dominant or even dominant in the middle marsh zone by mid-summer in some marshes in some years, turning the middle marsh into an orange and green mosaic visible at great distances (Dumbarton-Mowry marsh, San Pablo Bay fringe marshes). Colonization and species recovery dynamics associated with dodder-induced dieback of marsh vegetation have not been investigated.

Relatively uncommon species of the middle marsh zone of San Francisco Bay include Triglochin maritima (sea arrow-grass), Limonium californicum (sea-lavender), and Polygonum prolificum and P. patulum (non-native knotweeds). Reports of the rare Point Reyes endemic Polygonum marinense (Marin knotweed) in San Francisco Bay require taxonomic verification. Species which sel-
dom occur in the middle salt marsh zone of San Francisco Bay include Cordylanthus maritimus ssp. palustris (Pt. Reyes bird's-beak; Richardson Bay, H eredt M arsh), Puccinelia nutkaensis (Pacific alkali grass; Ravenswood fringe marshes and N ewark), Plantago maritima (sea-plantain) and Triglochin concinna (slender arrow-grass). 

These latter species are locally abundant in maritime salt marshes of M arin County. Invasive exotic species of the middle salt marsh include Spartina densiflora (Chilean cordgrass; Richardson Bay and Point Pinole), Spartina patens (saltmeadow cordgrass; near Burlingame and in brackish middle marsh at Southhampton Bay) and Cotula coronopifolia (brass buttons; early introduction, widespread but never persistent as a dominant in tidal marsh). The invasive exotic Salsola soda (M editerranean saltwort) also is spreading from high salt marsh to the middle marsh zone (D umberton-M owry marsh).

High or upper salt marsh may occur as topographic highs within the marsh plain (e.g., channel bank levees, wave-deposited ridges or mounds) or along the upland or alluvial edges of the marsh. This zone today commonly includes natives such as Grindelia stricta var. angustifolia (frequently a dominant in this zone), Distichlis spicata, erect-ascending phenotypes of Salicornia virginica, Cuscuta saliva, Frankenia saliva, Limonium californicum (sea-lavender), and Atriplex triangularis (spearscale, fat-hen). Where the upper marsh intergrades with low-lying alluvial soils and high groundwater (a condition today very rare in San Francisco Bay), the high marsh zone is dominated by dense stands of Lymnstromi-

coides or L. x multiflorus (creeping wildrye), or Juncus lesueurii (salt rush, wire rush), as still occurs commonly in maritime salt marshes of the region.

Cooper (1926) described a broad high salt marsh zone along the Palo Alto shoreline dominated by Distichlis spicata and Grindelia stricta var. angustifolia (an association still evident in reduced extent today), and a high salt marsh - alluvial transition zone which no longer exists. Cooper’s reconstructed high salt marsh ecotone community was dominated by native composites—H enizia pungens ssp. maritima, H. congesta (tarweeds), H elianthus bolanderi (Bolander’s sunflower), Aster subulatus (as “A. exilis”; slim or salt marsh aster), Aster chilensis (Chilean aster; possibly also including the rare A. lentus), Baccharis douglasii (salt marsh baccharis), Euthamia occidentalis (western goldenrod), and Iva axillaris (poverty weed). Of these, I. axillaris, A. chilensis, H. pungens, B. douglasii, and E. occidentalis still occur in high tidal brackish marsh of San Pablo Bay and Suisun M arsh. It therefore appears likely that historic upper edges of some salt marshes were at least locally brackish or subsaline rather than hypersaline in character, influenced by surface and subsurface freshwater discharges. This is also indicated by Cooper’s description of water table-dependent, salt-intolerant tall (to 9 m) thickets of willow, cottonwood, box-elder, ash, blackberry, ninebark, and California rose at the high marsh edge (Salix isolepis, Populus trichocarpa, Acer negundo, Fraxinus oregona, Rubus ursinus, Physocarpus capitatus, Rosa californica). Cooper (1926) interpreted this community from isolated remnants of what he assumed was undisturbed vegetation, but the disturbance history of the South Bay marsh edge at the time of his observations, and older reports he collected, is uncertain.

The high salt marsh zone also historically included many other native species, which are now uncommon, rare, or extirpated in San Francisco Bay (Table 1.3). Most of these still persist at other California salt marsh localities. Most high salt marsh zones in San Francisco Bay today occur on artificial slopes and substrates at the upper marsh edge, and include many non-native species that sometimes dominate the zone. Common non-native plants of the high salt marsh zone include Lepidium latifolium (broadleaf peppergrass, perennial peppergrass), Bassia hyssopifolia (bassia), Salsola soda (saltwort), Beta vulgaris (wild beet), M esembryanthemum nodiflorum (annual iceplant), Carpobrotus edulis and its hybrids (ice-plant), Atriplex semibaccata (Australian saltbush), Bromus diandrus (ripgut brome), Hainardia cylindrica and Parapholis incurva (sicklegrasses), and Polygogon monspeliensis (rabbit’s-foot grass).

Brackish Marsh Plants and Their Associations

- Brackish tidal marshes prevail over northern San Pablo Bay (slough systems of the Petaluma River, T olay Creek, Sonoma Creek, and Napa River), the Suisun M arsh area, and the Contra Costa marshes (N orth Bay marshes).
They also occur in transition with San Francisco Bay salt marshes where significant freshwater discharges occur (e.g., fringing marshes of Mud Slough, Coyote Creek, Artesian Slough, Alviso Slough, and Guadalupe Slough). The distinction between “salt marsh” and “brackish marsh” is a recent convention in descriptions of San Francisco Bay Area tidal marshes: brackish marshes were indiscriminately described as “salt marshes” by early California botanists, making it difficult to separate distinct elements of “salt” and “brackish” marsh associations. The description and demarcation of brackish marsh plant communities is essentially a matter of convenience and convention: there is no precise, stable salinity threshold at which tidal marshes are known to switch from one “type” to another (Adam 1990). Instead, brackish marsh vegetation in the San Francisco Estuary is typically a dynamic continuum between salt marshes of San Francisco Bay and freshwater tidal marshes of its major tributary rivers, fluctuating with variable influence of rainfall and freshwater discharges which alter marsh salinity and vegetation gradients geographically and over time. Associated changes in local tidal elevations (related to freshwater discharges) may also possibly interact with salinity variations in altering the character of brackish marsh vegetation. Changes in brackish marsh vegetation between dry and wet years at the same location may be dramatic: cover can change from that typical of San Francisco Bay salt marsh (dominant pickleweed) to that typical of Suisun Marsh (mosaic of rushes, bulrushes, alkali-bulrush, cattails, saltgrass, and many broad-leaved herbaceous species) in very few years. The causes of these dramatic changes in brackish tidal marsh vegetation are presumably related to plant interactions (competition, facilitation, and parasitism) which are influenced by seasonal and annual variation in salinity and drainage (Pearcy and Ustin 1984), but are poorly understood beyond descriptive observation.

The most extensive tidal brackish marshes occur in the Petaluma Marsh, but relatively large relict tidal brackish marshes also occur along the Napa River (Fagan Slough marsh) and in the Hill Slough/Rush Ranch area in Suisun Marsh. Relatively young but large and well-developed brackish marshes also occur bayward of dikes constructed after the 1870s, particularly in the Napa-Sonoma marsh complex and Suisun Marsh, including marsh islands of Suisun Bay. The Contra Costa marshes are predominantly intermediate between fully tidal marsh and diked (reduced tidal range) brackish marshes. The extensive wave-influenced, prograded pickleweed-dominated marsh plain and low natural marsh levee along northern San Pablo Bay are transitional between salt marsh and brackish marsh, exhibiting increases in brackish-associated species (particularly Scirpus maritimus at the east end of the Bay) in series of wet years.

Plant species richness and diversity markedly increase in brackish marshes of the San Francisco Estuary compared with salt marsh. Grewell (1993 et seq.) compiled extensive vascular plant species lists of the Suisun Marsh (including uplands of dikes and artificial uplands), and presented the only comprehensive and contemporary synthesis of Suisun Marsh plant ecology and its history (Grewell et al. 1999). Mason (Newcombe and Mason 1972) described plant community composition of brackish tidal marshes extending into the Sacramento-San Joaquin Delta.

The low brackish marsh zone differs from the corresponding zone in the San Francisco Estuary salt marshes in several respects: it supports multiple dominant species in variable mixtures or monospecific stands; it extends to the low end of intertidal zone, and it regularly develops tall, dense vegetation. In San Pablo Bay and western Suisun Marsh, alkali-bulrush (predominantly Scirpus maritimus around San Francisco and San Pablo bays and western Suisun Marsh, but also including S. robustus, a taxon formerly misapplied to S. maritimus in floras of the region) occurs in the upper portion of the low marsh, often dominant in the saline end of the brackish marsh gradient. The tallest graminoid species, tules and cattails, dominate where freshwater influence is relatively strong; these include Typha angustifolia, T. latifolia, T. domingensis and hybrids; Scirpus californicus (California tule), S. acutus (hardstem tule) and hybrids. These graminoid species can also establish within poorly drained portions of the middle marsh plain.

The middle brackish marsh zone was historically dominated by Distichlis spicata (saltgrass), as it commonly is today (Newcombe and Mason 1972). Other native species of the high marsh which occur in variable abun-
dance (common to co-dominant) include Salicornia virginica, Atriplex triangularis, the Juncus balticus - lesueurii complex, Jaumea carnosa, Frankenia salina and Cuscuta salina. Locally common natives include Limonium californicum (sea-lavender), Glaux maritima (sea-milkwort), and Scirpus koiolopis cernuus and S. cernuus (clubrush; also in high brackish marsh), Eleocharis macrostachya (creeping spikerush), Helenium bigelovii (Bigelow’s sneezeweed), and Deschampsia cespitosa ssp. holoformis (tufted hairgrass; especially eastern Suisun M arsh). In frequent to rare species of this zone include Lilaeopsis masonii and L. occidentalis (Mason’s and western Lilaeopsis; on exposed eroding channel bank edges as far west as T olay Creek), Triglochin maritima (locally common), T. concinna, T. striata, Sium suave (water parsnip), Onanthe sarmentosa (ditch-carrot), Cicutum maculatum ssp. bolanderi (water hemlock), Eleocharis parvula (slender spikerush), Pluchea odorata (salt marsh fleabane), and Lythrum californicum (California loosestrife; eastern Suisun M arsh and D elta). In wet years, depressions in the middle marsh plain support increased abundance of Scirpus americanus (O Iney’s bulrush) or S. maritimus (alkali-bulrush; western Suisun and San Pablo Bay) and Phragmites australis (common reed; eastern Suisun Marsh, also in the low-middle marsh zone). The dominant non-native species of the middle brackish marsh is again Lepidium latifolium, which rapidly forms dense monotypic clonal populations, spreading into the marsh plain. Other exotic species which have established in the brackish middle marsh zone include Apium graveolens (wild celery, widespread and abundant in Suisun Marsh), Lythrum hyssopifolium (annual loosestrife), Cotula coronopifolia (brass-buttons) and Chenopodium chenopodioides (fleshy goosefoot; N apa-Sonoma marshes).

The high brackish marsh zone is today typically altered by artificial dikes and invasive plants (particularly Lepidium latifolium (perennial or broadleaf peppercress), Conium maculatum (poison hemlock), Foeniculum vulgare (fennel), and Mediterranean grasses. However, many native remnants of the brackish high marsh community have regenerated on old, stable, relatively undisturbed levees, or have persisted locally along undiked tidal marsh edges. They include Achillea millefolium (yarrow), Baccharis douglasii (salt marsh baccharis), B. pilularis (coyote-brush), Leymus triticoides and L. x multiflorus (creeping wildrye), Scrophularia californica (California bee-plant), Rubus ursinus (blackberry, in the upland ecotone) Rosa californica (California rose, also in the upland ecotone), Iva axillaris (poverty-weed), Atriplex triangularis (fat-hen or sparsescal), Grindelia stricta var. angustifolia (and intermediates with G. camporum), Calystegia sepium ssp. limophila (morning-glory), Cressa truxillensis (alkali-weed), Frankenia salina (alkali-heath), Lathyrus japonii var. californicus (California tule pea), Juncus balticus - lesueurii complex (salt or wire rush), Juncus mexicanus (M exican rush), J. bufonius (toad rush), Ambrosia psilostachya (western ragweed), Euthamia occidentalis (western goldenrod), Epilobium brachycarpum, E. ciliatum (willow-herbs), Polygonum spp. (smart-weeds, knotweeds), Triglochin maritima (sea arrow-grass) and Eryngium articulatum (coyote-thistle). Uncommon to rare species such as Lathyrus japonii var. japonii (D elta tule pea), Aster lenatus (Suisun aster), A. subulatus var. ligulatus (slim aster), Plantago elongata (dwarf plantain), Rumex occidentalis (western dock), Eleocharis parvula (spikerush), and endangered Cordylanthus mollis ssp. mollis (soft bird’s beak) and Cirsium hydrophilum var. hydrophilum (Suisun thistle) typically occur locally in the lower end of well-drained high marsh gradient, often on slight topographic relief above the marsh plain. Salicornia virginica (common pickleweed) and occasionally S. subterminalis (Parish’s glasswort) can also be abundant elements of high brackish marsh near Suisun. The composition of high brackish marsh vegetation appears to vary with slope, drainage, and local surface or subsurface freshwater influence, but no studies have yet analyzed vegetation patterns or related environmental factors in brackish marshes of the region.

Invasive non-native species (weeds of mesic and wetland habitats with slight salt tolerance) of the high brackish marsh zone are numerous, particularly in years of high rainfall, but the most aggressive and successful is again Lepidium latifolium. Lotus corniculatus (bird’s-foot trefoil) and L olum multiflorum (ryegrass) are other exotics which are locally abundant along portions of the upper brackish marsh edge some years. Elytrigia pontica ssp. pontica (tall wheatgrass, currently local around Alameda Creek and M are Island), Rumex crispus and R. pulcher (curly and fiddle docks), Asparagus officinalis (locally abundant near N apa-Sonoma marshes) have also naturalized along brackish marsh edges, but are seldom invasive.
Tidal Marsh Pans and Vegetation - Poorly drained flats, depressions, and barrier-impounded areas of tidal marsh lacking emergent vascular vegetation, called pans (alternatively spelled “pannes”), range from nearly planar unvegetated marsh areas subject to shallow periodic ponding, to steep-sided or cliff-edged shallow ponds which are persistently inundated (Pestrong 1965, Pethick 1974, Atwater et al. 1979). Pans have various modes of origin and development, which have not been completely clarified (Adam 1990, Carter 1988, Pethick 1974, Chapman 1960). In San Francisco Bay Area marshes, pan variation includes nearly circular ponds between drainage channels (Pestrong 1965), historic long ponds parallel with impounding bayfront marsh berms (Atwater et al. 1979), shore-parallel pans historically present along portions of the back edge (upland or lowland margin) of tidal marsh (depicted in 1880s U.S. Coast Survey Maps), and natural historic salt ponds impounded by low estuarine ridges (Atwater et al. 1979). Some sloped to planar pans in the high marsh (bare flats, rarely submerged) may be related to wrack deposition and smothering, or local substrate conditions. Little is known of the ecology of pan types that are no longer represented in the altered modern Estuary.

Many pans are reported to become seasonally hypersaline (Pestrong 1965) or even salt-crystallizing (Atwater et al. 1979; see also salt pond profile, this volume) and lack vascular plants, but some pans along the landward edge of the tidal marsh develop marginal vegetation typical of brackish or fresh marshes (e.g., China Camp). Ponded pans within the marsh plain have been described as “unvegetated” (Pestrong 1965), but they often support a dense submerged mixed vascular and non-vascular vegetation variously composed of widgeon-grass (Ruppia maritima) and membranous green algae (particularly Enteromorpha and Ulva spp.). According to MASON (N ewcomb and M aston 1972), brackish ponds in Suisun Marsh also support Zannichellia palustris and Potamogeton pectinatus, submerged species typical of freshwater ponds. The halophilic microflora of salt ponds is discussed in the salt pond profile (this volume). Shallow, relatively planar and ephemeral pans in San Pablo Bay are either periodically or marginally colonized by pickleweed, which dies back during years of frequent flooding or high rainfall. The steep-sided edges of well-defined, nearly circular old pans sometimes develop small natural levees of locally improved drainage, and sometimes support certain species at frequencies more typical of high marsh vegetation. In the high marsh, on gently sloping alluvial fans, “dry pans” (small playa-like flats with very short flooding periods and superficial salt films) also develop, often on relatively coarse (sandy, silty, or even gravel-silt mixtures) sediments. These features are very rare today because of diking, but fine examples persist at Point Pinole (W hittell M arsh). Here, as at similar pans on alluvial fans at tidal marsh edges in maritime Marin County, the pan-marsh edges are associated with local abundance of the regionally rare salt marsh owl’s clover (Castilleja ambigua ssp. ambigua; salt-tolerant ecotypes). Salt marsh bird’s-beak (Cordylanthus maritimus ssp. palustris) also exhibits a pan-margin local distribution pattern in western San Pablo Bay (e.g., JEPS83457). Analogous artificial features (gently sloping, formerly disturbed silty to sandy high marsh fills with residual vegetation gaps) elsewhere in the Estuary have also become colonized with rare plants such as Cordylanthus mollis (Hill Slough near Lawler Ranch; B. Grewell, pers. obs.) and Cordylanthus maritimus (near M arin City). Natural and artificial high marsh pans of this type, associated with alluvial or deltaic deposition.
or erosion, have not been identified in the regional literature on salt marsh ecology, and require study.

The number of species from former alkali-subsaline vernal pools around San Francisco Bay which were historically reported from local salt marshes as well (see diked wetlands profile, this volume) suggests that ecologically equivalent habitat occurred in both ecosystems. Although there are very few intact remnants of the elongate pans which occurred along tidal marsh edges (represented clearly in historic U.S. Coast Survey maps of the 1850s), it is possible that some of these seasonally ponded depressions in the upper marsh ecotone were partial ecological equivalents of subsaline vernal pools. Strong historic evidence for this conclusion is found in Jepson’s (1911) range and habitat descriptions for the typical vernal pool species, Downingia pulchella, which he described as “abundant and of rank growth in the salt marshes near Alvarado” (now Union City). Other species indicative of vernal pools and similar seasonally ponded/desiccated alkaline/subsaline environments, such as Lasthenia conjugens (JEPS25099), L. platycarpha (DS695549, Greene 1894) and L. glabrata (CAS897444, D.S73122, DS286673) have been collected from the edges of San Francisco Bay.

Although pans are often presumed to be generally hypersaline, some appear to have occurred historically in alluvial lowlands with probable groundwater or surface discharges that could maintain brackish conditions in pans along tidal marsh edges. A number of characteristic freshwater marsh species were reported by Jepson (1911) and others from historic salt marsh habitat (e.g., Agrostis exarata, Carex aquatilis, C. densa, Lycopus asper), suggesting that freshwater sub-habitats occurred marginally along tidal salt marshes. Unpublished historic writings of southeastern San Francisco Bay marsh borders by 19th century botanist Joseph Burtt-Davy, archived at the Jepson Herbarium, University of California, describe extensive colorful wildflower meadows with species typical of vernal pools and wet grassland (R. Grossinger, pers. comm. 1999). Examples of brackish and even freshwater vegetation at edges of salt marsh with pans near zones of groundwater discharge can be observed today at China Camp (Marin County) and Point Pinole (Contra Costa County), and in maritime Marin County tidal marshes. Diked seasonal wetlands in historic tidal marsh (this volume) may also approximate this type of lost habitat, since numerous seasonal wetland species of vernal pools and alkali basins have colonized diked Baylands.

### Uncommon, Rare, Declining, and Extirpated Plant Species

There is a widespread impression, even among ecologists familiar with the San Francisco Estuary, that native plant species richness of tidal marshes (particularly salt marsh) is relatively low, and that rare species in the Estuary are principally wildlife taxa, not plants. This impression is due in part to reviews of species richness in tidal marshes based solely on modern surveys: for example, Atwater et al. (1979) reported only 15 vascular plant species for San Francisco Bay, based on modern reports. Josselyn (1983) discussed only a small representation of the San Francisco Estuary flora, and did not address either his historic or modern species richness. In addition, very few plants native to the San Francisco Estuary are federally listed as endangered or threatened, and only two of these (soft bird’s-beak, Cordylanthus mollis ssp. mollis, and Suisun thistle, Cirsium hydrophilum var. hydrophilum) currently inhabit this Estuary. The modern lack of attention to rare plants in the Estuary is probably due to unfamiliarity with plant species which were known only to early botanists, but are either now entirely extinct (or even extirpated) in the Estuary. Plant species that were historically recorded in the tidal marshes of the Estuary, or along its edges (high marsh), but have become uncommon, rare, regionally extirpated, or extinct, are summarized in Table 1.3. Most of these species were known from tidal marsh edges, transitional habitats of high ecological diversity. This is significant, because original remnants of this ecotone are almost completely eliminated from the Estuary, and their modern counterparts are mostly weedy, disturbed habitats like dikes.

Extinct species of the Estuary include California sea-blite, Suaeda californica, a federally endangered shrubby true halophyte (salt-tolerant plant) which today inhabits relatively well-drained marshy beach ridges along relatively high-energy shorelines with coarse sediment in Morro Bay. According to Jepson (1911) and Greene (1894), it was never abundant in San Francisco Bay even in the late 19th century. The distribution of its sandy marsh habitats was unfortunately in areas of the greatest urbanization: San Francisco, Oakland, Alameda, and San Leandro were its core populations,
although it was also collected in Palo Alto (where shell hash beaches today occur) and at the former San Pablo Landing (Richmond, where local beaches still persist). The species today is restricted to sandy salt marsh edges of Morro Bay, San Luis Obispo County, and also exists in cultivation. It was last collected in San Francisco Bay in 1958 in San Leandro (JEPS25020). More recent local reports are based on misidentification of the similar species, S. moquinii, in diked Baylands.

Many other salt marsh species that have affinity for high sandy salt marsh were also reported from San Francisco Bay, but are now extinct or rare in the Bay (Jepson 1911, Greene 1894). They include California saltbush (Atriplex californica), still found in Tomales Bay and Point Reyes sandy salt marshes, but extinct in the Bay, and Plantago maritima, common in sandy maritime salt marshes, uncommon to rare in the Bay. These sea-pink (Armeria maritima) a showy pink spring wildflower which still occurs locally along sandy edges of Point Reyes salt marshes, was cited by Jepson (1911) to range within San Francisco Bay. This may possibly have been along former sandy beaches, sandy salt marsh, or stabilized former bayside dunes. There are no historic herbarium specimens from San Francisco Bay salt marshes to corroborate Jepson’s report, however.) Other rare species, such as Cordylanthus maritimus ssp. palustris and Castilleja ambiguus ssp. ambiguus, are less uncommon in sandy maritime salt marshes, but are rare in San Francisco Bay. The decline or demise of these species in the Bay is very likely a result of the near-complete elimination of its sandy estuarine barrier beaches.

Two other species which are probably extinct in San Francisco Bay, but occur elsewhere, include two members of the Aster family: southern goldenrod (Solidago confinis) and Pyrrocoma racemosa (=Haplopappus racemosa). Southern goldenrod was formerly reported as rare only by Henry Bolander in 1863 (Jepson 1911), when it was misidentified as seaside goldenrod (S. sempervirens). In California tidal marshes today, S. confinis is known only locally from the high brackish marsh zone of southern Morro Bay. P. racemosa was formerly reported from the edges of salt marshes and saline soils at Cooley’s Landing and near Alviso (Thomas 1961), but has not been reported from salt marsh edges in recent decades. Another species, Adobe sanicle (Sanicula maritima), was found locally in lowlands adjacent to salt marshes at Alameda (Behr 1888, Greene 1894, Jepson 1911) and in San Francisco (Brandegge 1892). It is now extinct in the Bay Area, and is very rare elsewhere (known from fewer than 10 sites in Monterey and San Luis Obispo Counties today; Skinner and Pavlick 1994).

Two popcornflower species (genus Plagiobothrys, well represented in vernal pools) that were found in saline soils near the edge of the Estuary are now presumed to be extinct (although it is possible that buried dormant seed may persist somewhere in diked Baylands, awaiting resurrection). They include Petaluma popcornflower (Plagiobothrys mollis var. vestitus), which was probably distributed in alkaline or subsaline seasonally wet depressions (vernal pools) in grasslands and lowlands adjacent to tidal marsh in the Petaluma Valley, and Hairless popcornflower (P. glaber), a species of seasonally wet alkaline/subsaline soils of tidal marshes of the south San Francisco Bay (reported by Jepson (1911) from Alvarado [now Union City]), as well as some interior valleys. Almost nothing is known of the ecology of these species because of their early historic extinction.

Other species that are known to occur in subsaline to alkaline vernal pools, and which historically occurred in salt marshes (presumably along lowland edges), include several species of goldfields (Lasthenia spp.). Fleshy goldfields, Lasthenia platycarpa (presumed extinct in the Estuary) was known from salt marshes near Vallejo (Greene 1894), and smooth goldfields (L. glabrata ssp. glabrata) was reported from edges of salt marshes (Thomas 1961, Jepson 1911, Greene 1894). L. glabrata was recently confirmed to occur naturally at Whittell Marsh, Point Pinole, and a population of undetermined origin occurred briefly in 1998 on a hydroseeded levee at the Sonoma Baylands tidal marsh restoration project’s pilot unit. Behr (1888) listed L. glaberrima as a species occurring “near salt marshes,” but is not otherwise reported from tidal marshes in the region. The federally endangered vernal pool goldfields species, Contra Costa goldfields (L. conjugens) was reported by Jepson (1911) from “subsaline soils” near Antioch and Newark, and was recently discovered in subsaline vernal pools in Fremont near the diked edge of the at the Warm Springs Unit of the National Wildlife Refuge in Fremont and adjacent derelict fields. L. conjugens was also observed along high tidal marsh edges of Hill Slough in the early 1990s. Another well-known vernal pool species, the showy Down-
Ambigua foliage, bracts, and flowers (atypical of the subspecies Point Pinole, which supports a form with purple-tinged color in San Francisco Estuary). The only salt marsh population of this area is from extinct there now. The only salt marsh population of this is from Alameda, Bay Farm Island, Burlingame, but is nearly marshes of San Francisco Bay (Berkeley, Oakland, Central Bay, with small remnant populations probably persisting in Petaluma Marsh and near Gallinas Creek, Marin County). The remaining San Francisco Bay populations of Marin County are typically showier (usually more conspicuous, rosy purple flowers and purplish herbage) than most of the core populations of Point Reyes. It is now restricted to very few populations in the Central Bay, with small remnant populations probably persisting in Petaluma Marsh and near Gallinas Creek, Marin County. The remaining San Francisco Bay populations of Marin County are typically showier (usually more conspicuous, rosy purple flowers and purplish herbage) than most of the core populations of Point Reyes, which typically have gray-green foliage and white-and-maroon flowers. Another annual Snapdragon family herb, Johnny-nip or salt marsh owl’s clover (Castilleja ambigua ssp. ambiguа) was formerly found in the salt marshes of San Francisco Bay (Berkeley, Oakland, Alameda, Bay Farm Island, Burlingame), but is nearly extinct there now. The only salt marsh population of this colorful annual herb in the San Francisco Estuary is from Point Pinole, which supports a form with purple-tinged foliage, bracts, and flowers (atypical of the subspecies SOSC-55

Chapter 1 — Plant Communities 19
area. Sedges such as Carex subbracteata, and C. obnupta would also be expected to have occurred in former salt marsh edges, as they do in other estuaries of the Central Coast, especially northward.

Many broadleaved herbs were also more plentiful along tidal marsh edges, but have become localized or rare today. They include Aster lentus (Greene 1984, Jepson 1911), a species now generally rare in any estuarine habitat; Chilean aster (Aster chilensis) (Howell 1949, Thomas 1961), a common species of non-saline habitats which has nearly disappeared from salt marsh edges but persists occasionally in Suisun, Petaluma, and Napa Sonoma marshes. Salt marsh baccharis (Baccharis douglasii) was formerly abundant in salt marshes (Jepson 1911) but is now uncommon to rare in brackish marshes, mostly in the North Bay (Best et al. 1996, Thomas 1961). Two species which were inferred by Cooper (1926) to be major elements of his reconstructed "willow-composite" community at South Bay salt marsh edges, slim aster (Aster subulatus var. ligulatus), and spikeweed (Ehretia pungens var. maritima) are now scarce in tidal marshes, and occur mainly in the North Bay (Best et al. 1961; B. Grewell, pers. obs. 1997). Other spikeweeds, H. parryi ssp. parryi and congestii, were locally common in the South Bay salt marshes (Munz 1959), but are generally rare today. Species that were formerly frequent in North Bay brackish and salt marshes (Greene 1894) include morning-glory (Calystegia sepium var. limonoides) and sea-milkwort (Glaux maritima), which are now uncommon to rare. Other herbs which have historically declined to a significant extent in frequency, distribution, and abundance in Bay Area tidal marshes and their edges include Hutchinsia procumbens (Greene 1894, Thomas 1961), tidy-tips, Layia chrysanthoides (Howell 1949, Thomas 1961), native annual peppers cress species Lepidium dickytomum, L. latipes, and L. oxycarpum (Thomas 1961, Munz 1959, Howell 1949, Greene 1894), salt marsh fleabane, Pluchea odorata (Jepson 1911), and butterweed, Senecio hydrophilus (Greene 1894, Jepson 1911).

Conservation Issues

Exotic Plants - There are many exotic plants that have become established within, or along the edges of, the San Francisco Estuary, but only a few are aggressive invaders that have become widespread and dominant, or threaten to do so (Grossinger et al. 1998). Of these, Lepidium latifolium, Spartina alterniflora (and hybrids), and Salsola soda have demonstrated ability for rapid, extensive invasion and development of monodominant stands in the San Francisco Estuary. Spartina densiflora, an exotic cordgrass from Chile with a bunchgrass growth habit, has become a dominant species in Humboldt Bay, and is expected to be able to achieve the same dominance if its spread is unchecked in San Francisco Bay. The taller stature of S. alterniflora enables it to endure high tides with relatively little submersion of its foliage, even when rooted below mean sea level. T urf-forming S. patens (salt meadow cordgrass) and dwarf strains of S. alterniflora (Daehler et al. 1999) present in the Bay may be latent invaders of salt marsh plains. The tendency for S. alterniflora pollen to swamp the pollen of the native S. foliosa and produce hybrids and introgressants threatens to genetically assimilate the native Pacific cordgrass over a significant portion of its geographic range (D. Ayers and D. Strong, pers. comm. 1999). The higher densities, larger plant size, and greater colonizing ability of S. alterniflora at lower tidal elevations also suggest that its spread may have significant geomorphic impacts on the Estuary, particularly on channel stability, sedimentation, and mudflat colonization, and their indirect effect on wildlife habitat (Grossinger et al. 1998).

Lepidium latifolium invasion is particularly a concern for the conservation and recovery of rare or endangered plant species of the San Francisco Estuary, most of which occur in the high marsh zone where L. latifolium is dominant. L. latifolium actively encroaches on populations of endangered Cordylinthus mollis, mollis and Cirsium hydrophilum var. hydrophilum in Suisun Marsh (B. Grewell, pers. obs. 1998) and Southampton Bay (P. Baye, pers. obs. 1998). The impact of exotic plant invasions in the high marsh zone is magnified by the truncation and degradation of this habitat by widespread diking, which compresses the high marsh zone into a relatively invariant, steep slope of disturbed Bay mud.

Tidal Marsh Restoration Design - Tidal marsh restoration in the San Francisco Estuary has convention-
ally been designed for wildlife species, treating plants only as habitat for wildlife species rather than as the subject of restoration aims. Restoration designs have generally afforded little or no consideration for soils or slopes of the high marsh zone, variations in sediment texture, surface or subsurface freshwater flows, and variation in incident wave energy that influence the micro-environmental variables which are significant for plant diversity. Highly managed estuarine wetlands (e.g., artificial salt ponds, extremely microtidal or non-tidal salt marshes) generally support an artificially low diversity of native tidal marsh plant species. Plans for rare tidal marsh plant reintroduction have only recently been proposed (e.g., Pier 98, Port of San Francisco; Crissy Field, Presidio/Golden Gate National Recreation Area), and none has yet been implemented. Of the rare plant refugia in relict tidal marshes of the Estuary (e.g., Hill Slough, Fagan Marsh, Rush Ranch, Peytonia Slough, and Whittell Marsh), none has site-specific rare plant management plans or programs, despite imminent threats by invasive species. There is no Estuary-wide program to survey and map rare plant species populations; plant inventories are biased towards species with special legal status, and are typically driven by environmental impact assessment for projects rather than regional conservation. Other surveys consist of voluntary and opportunistic reports. Conservation of plant diversity in the Estuary will require both active protection of remnant rare plant refugia, active management of conserved areas, systematic inventory of the Estuary’s botanical resources, and large-scale, scientifically sound tidal marsh restoration and reintroduction projects.

Many natural resource agencies are cautious about restoration and reintroduction of rare plants, probably because this has conventionally been considered in a mitigation context (Berg 1996). Restrictive generalized policies on geographic specificity of reintroduction to documented historic localities, regardless of natural temporal and spatial scales of plant population dynamics and ecosystem processes, in some cases has narrowed opportunities for re-establishment of rare plants (White 1996). In situations where the range of rare plants is extremely reduced, historic collection data are sparse and vague (which is generally the case), and relatively few potential source populations for founders exist, an experimental approach may be most appropriate for reintroduction planning. Successful reintroduction will likely require much replication over years (variable climate conditions) and at many localities. Caution is appropriate, however, when the taxonomic interpretation or population variability is at issue when determining suitable populations for reintroduction.

**Artificial Salinity Manipulation** - In Suisun Marsh, salinity control gates on Montezuma Slough were installed to enforce standards for salinity based on the perceived needs of waterfowl marsh management in diked wetlands, aimed at maintaining low channel water salinity. The impacts of sustained low marsh salinity on the progression of exotic plant invasions and the natural dynamics of brackish tidal marsh vegetation (particularly rare and endangered species) were not considered in the design and operation of the salinity control gates, and no long-term monitoring of rare plant populations during gate operation was authorized. The reduction of periodic high salinity events during drought cycles, and

Pioneer plants (1st year seedling) of native *Spartina foliosa* and *Salicornia virginica* colonize the well-consolidated upper mudflats bayward of the marsh edge at Mare Island, eastern San Pablo Bay. The erosional scour pools and drainages adjacent to the plants indicate the relatively high wave energy estuarine environment in which they are able to establish, given stable microhabitats.
the subtle changes in tide elevations caused by gate operation, could potentially have significant adverse long-term impacts on rare plant persistence. Scientific investigations of the effects of gate operation on plant communities and rare plant populations of Suisun tidal marshes are urgently needed, as recommended by the Brackish Marsh Subcommittee of the Suisun Ecological Workshop (CWRCB 1999).

In the South Bay, perennial urban wastewater discharges in confined, diked tidal sloughs have caused conversion of salt marsh to brackish marsh (Harvey and Associates 1997). The Alviso and Milpitas area marshes were the sites of historic rare plant populations (Table 1.3) which could not be re-established naturally or be reintroduced in marsh vegetation dominated by perennial pepperweed, bulrushes and tules which are stimulated by augmented and confined freshwater flows and elevated nutrient concentrations throughout the growing season.

**Loss of Restorable Habitat** - Economic pressure to convert diked Baylands to land uses that are incompatible with potential tidal marsh restoration over large contiguous tracts (particularly in connection with uplands and alluvial areas) remains high today. Developments in diked Baylands for extensive housing (Redwood Shores, San Mateo County), golf courses (Black Point, Marin County), business parks (old Fremont Airport, Alameda County) have proceeded into the 1990s, and other large scale land use conversions for dredged material disposal and rehandling (Napa salt crystallizers) have been considered. The largest tracts of undeveloped diked Baylands are in San Pablo Bay, where vineyard expansion threatens to encroach into restorable former tidal marsh sites. Single-purpose management of other diked wetland types at large scales (salt production, waterfowl production) also restricts opportunities for tidal marsh plant community restoration. Large-scale tidal marsh restoration near centers of relict tidal marsh plant populations (e.g., Cullinan Ranch, Hamilton Airfield, Redwood Landfill, and Skaggs Island) offer some hope for long-term recovery of tidal marsh plant species in decline.

**Conclusions and Recommendations**

The San Francisco Estuary tidal marshes are poorly understood in terms of modern and historic plant species composition, the dynamics of the vegetation, and the interaction between vegetation and geomorphic and hydrologic processes. Many plant species have become extirpated or nearly so with little or no attention from botanists or ecologists, and many more species have declined significantly. The Estuary's historic and modern flora is considerably richer than has been generally recognized. Further attrition of native plant diversity in the Estuary is likely because of the uncontrolled spread of invasive exotic plants, and insufficient planning, management, and restoration of the Estuary's plant community. Carefully designed tidal marsh restoration projects that promote native plant species diversity and recovery are needed to conserve the Estuary's flora. Recommendations for the conservation of the Estuary's plant communities are presented in the Baylands Ecosystem Habitat Goals Report (Goals Project 1999, Appendix A).

An example of marsh progradation — Seedling plants of *Salicornia virginica* and *Spartina foliosa* are frequently comingled without clear zonation, as in these exceptionally firm upper mudflats in eastern San Pablo Bay, (Mare Island, north of the jetty)
<table>
<thead>
<tr>
<th>Taxon</th>
<th>Historic References</th>
<th>Contemporary Distribution</th>
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<tr>
<td><em>Agrostis exarata</em> Trin.</td>
<td><strong>Jepson 1911:</strong> Common in the San Francisco Bay region in salt marshes and other wet places: Berkeley...San Francisco; Martinez**</td>
<td>Not currently reported from tidal marsh ecotone in San Francisco Bay Estuary, but common and widespread in non-tidal moist habitats [Hickman et al. 1993].</td>
</tr>
</tbody>
</table>
*Jepson 1911:* “common on the sandy beaches or fields near the sea...or about San Francisco Bay.” | Apparently extirpated in San Francisco Bay Estuary; otherwise restricted to maritime coastal salt marshes, dunes, bluffs. |
| *Aster chilensis* Nees.      | **Cooper 1926:** [presumed species of reconstructed “willow-composite” community at salt marsh edges, Palo Alto vicinity]  
**Howell 1949:** [Marin Co.] Common and widespread from salt marshes and coastal swales to low valleys.  
**Jepson 1911:** “…edges of salt marshes.” | Few current reports known from edges of San Francisco Bay or San Pablo Bay tidal marshes; local in Suisun Marsh edges. Presumed rare from tidal marshes. |
| *Aster lentus* E. Greene [A. chilensis Nees. var. lentus] Jepson] [A. chilensis var. sonomensis (E. Greene)] Jepson | **Greene 1894:** [A.c. var. lentus] “Plentiful along tidal streams in the western part of the Suisun Marsh...”  
[A. c. var. sonomensis] “In open plains of the Sonoma Valley, in low subsaline ground.”  
**Jepson 1911:** [A.c. var. lentus] “very common and conspicuous in the Suisun Marshes”  
[A. c. var. sonomensis] “subsaline lands: Petaluma, Napa”  
**Munz 1959:** [A.c. var. sonomensis]: Coastal Salt Marsh; saline ground around San Francisco Bay. Sonoma, Napa... | Rare; restricted primarily to Suisun Marsh. Some herbarium collections known from San Francisco Estuary prior to 1960 (Berkeley, Alviso, Napa). Recent status uncertain in San Pablo Bay area tidal marshes. |
| *Aster subulatus* Michaux var. *ligulatus* Shinn. [Aster exilis Ell.][Aster divaricatus Nutt.] | **Behr 1888:** [A. divaricatus] “Salt marshes.”  
**Greene 1894:** “Borders of Suisun Marshes and elsewhere on subsaline land”  
**Jepson 1911:** “Saline soil, not common...Alvarado.”  
**Cooper 1926:** [presumed species of reconstructed “willow-composite” community at salt marsh edges, Palo Alto vicinity]  
**Thomas 1961:** “Salt marshes along San Francisco Bay and occasionally elsewhere. San Francisco, Palo Alto, Alviso...” | No current reports known from edges of San Francisco Bay. Uncommon to rare in San Pablo Bay and Suisun tidal marshes. |
| *Astragalus tener* Gray var. *tener* | **Jepson 1911:** “Alkaline fields, mostly in moist places.”  
**Thomas 1961:** Known locally only from saline areas along San Francisco Bay. San Francisco and Mayfield.” | Recently rediscovered near historic Bay edge in Fremont, Alameda County. Known in region from alkali vernal pools, Solano County. |
Table 1.3 (continued) Historic Changes in the Distribution and Abundance of Selected Native Vascular Plant Species Occurring in Tidal Marshes of the San Francisco Estuary

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<th>Taxon</th>
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<td><strong>Atriplex californica</strong> Moq.</td>
<td>Greene 1894: “... along the edges of salt marshes, from near San Francisco and Alameda, southward.” Jepson 1911: “Sandy beaches along the ocean and about San Francisco Bay.”</td>
<td>Extirpated in San Francisco Bay Estuary margins. Small relict populations occur on bluffs of Golden Gate in San Francisco. Maritime salt marsh populations occur at Limantour Estero and Tomales Bay (Marin Co.).</td>
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<td><strong>Baccharis douglasii</strong> DC.</td>
<td>Jepson 1911: “... abundant in the salt marshes about San Francisco Bay.” Thomas 1961: [SW San Francisco Bay] “... occasionally along the edges of salt marshes...Alviso.”</td>
<td>Now uncommon to rare in alluvial high marsh and upland ecotone, San Pablo Bay area and and Suisun Marsh; one colony occurs along salt pond edge at a seep in Coyote Hills, Alameda Co., possibly rare elsewhere in San Francisco Bay.</td>
</tr>
<tr>
<td><strong>Carex aquatilis</strong> Wahlenb. var. <em>dives</em> (Holm) [C. sitchensis Prescott]</td>
<td>Brewer et al. 1880: “In salt marshes, about San Francisco Bay (Bolander).” Jepson 1911: “Salt-marshes about San Francisco Bay and northward along the coast.” Munz 1959: “Rare, swampy places, usually near the coast.”</td>
<td>No current reports known from edges of San Francisco Bay or San Pablo Bay tidal marshes. Presumed rare or extirpated from tidal marshes.</td>
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<td><strong>Carex densa</strong> Bailey [C. brogniartii Kunth. var. densa Bailey]</td>
<td>Jepson 1911: [C. b. var. densa] “Salt marshes near San Francisco.”</td>
<td>No current reports known from edges of San Francisco Bay or San Pablo Bay tidal marshes. Presumed rare or extirpated from tidal marshes.</td>
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<tr>
<td><strong>Carex praegracilis</strong> W. Boott [Carex Douglasii var. brunnea Olney] [C. usta Bailey]</td>
<td>Thomas 1961: “Boggy areas along the edges of salt marshes; San Francisco, Woodside, Mayfield.”</td>
<td>Rare in Suisun area tidal marshes, west to Southampton Bay. Common in alkaline, moist places in California floristic province.</td>
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<td><strong>Castilleja ambigua</strong> Hook and Am. [Orthocarpus castilleoides Benth.]</td>
<td>Behr 1888: “Marsh near Tamalpais.” Greene 1894: “Common along the borders of salt marshes.” Jepson 1911: “Marshy ground near the coast, Alameda; W. Berkeley; Napa Valley, Sonoma Co.” Howell 1949: “Low ground along the upper reaches of the salt marshes, occasional...Mount Tamalpais, Greenbrae Marshes, Hamilton Field.”</td>
<td>Currently reported only from Point Pinole salt marsh and pan edges, other historic records at Greenbrae, Tamalpais (Mill Valley), Hamilton Field, Burlingame, Oakland. Halophytic populations rarely occur in brackish marsh and salt marsh at Rodeo Lagoon, Tomales Bay, Drakes Estero, Limantour Estero (maritime Marin Co. marshes)</td>
</tr>
<tr>
<td><strong>Centaurium trichanthum</strong> (Griseb.) Robinson [Erythrea trichantha (Griseb.)]</td>
<td>Howell 1949: “In typical form...known in Marin only from low ground bordering the salt marsh near Burdell Station.” Munz 1959: “Moist often saline places...edge of Coastal Salt Marsh...San Mateo Co. to Siskyo Co.”</td>
<td>No current reports known from edges of estuarine tidal marshes. Similar species C. muehlenbergii occurs in subaline diked wetlands, Napa-Sonoma marsh, and tidal marsh edge at China Camp.</td>
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<td><strong>Calystegia sepium</strong> (L.) R.Br. <strong>ssp. limnophila</strong> (E. Greene) Brummit</td>
<td>Greene 1894: “Plentiful in brackish marshes toward the mouth of the Napa River and about Suisun Bay; its roots within reach of tide water; its stem-twining upon rushes and sedges. Muzn 1959: “Occasional in swampy saline places: Coastal Salt Marsh; Marin, Solano and Contra Costa Cos.”</td>
<td>Occasional in Suisun Marsh area west to Southampton Bay; rare in San Pablo Bay edges.</td>
</tr>
<tr>
<td><strong>Cordylanthus maritimus</strong> Benth. <strong>ssp. palustris</strong> (Behr) Chuang and Heckard</td>
<td>Brewer et al. 1880: “Sandy salt-marshes along the coast, from San Francisco Bay to San Diego.” Behr 1888: “Salt marshes, San Francisco.” Greene 1894: “Sandy salt marshes from near San Francisco southward.” Jepson 1911: “Salt marshes near the coast from San Francisco Bay south.” Howell 1949: “Salicornia flats in salt marshes along the bay... Almonte, Greenbrae... Thomas 1961: “Salt marsh along the borders of San Francisco Bay; San Francisco, Redwood City, Palo Alto, and near Alviso.”</td>
<td>Currently reported only from Richardson Bay, Greenbrae, and Petaluma marsh (Marin Co.). Recently reported from Gallinas Creek area marsh. Extirpated in central and southern San Francisco Bay. Major populations occur in maritime tidal salt marshes of Tomales Bay, Bolinas Lagoon, and Limantour Estero (Marin Co.). San Francisco Estuary populations have purplish foliage, and rosy, well-exserted inflated flowers.</td>
</tr>
<tr>
<td><strong>Cordylanthus mollis</strong> Gray <strong>ssp. mollis</strong></td>
<td>Brewer et al. 1880: “Salt marshes of San Francisco Bay, at Mare Island, and Vallejo, C. Wright, E.L. Greene.” Behr 1888: “Salt marshes, Vallejo.” Greene 1894: “Brackish marshes about Vallejo and Suisun.” Howell 1949: [Marin Co.].... San Rafael, acc. Ferris; Burdell Station, San Antonio Creek... Best et al. 1996: [Sonoma Co.]: Rare, estuarine... Petaluma Marsh between San Antonio and Mudhen Slough...(1978):...</td>
<td>Rare (federally endangered); local in tidal brackish marsh around Napa River, Carquinez Straits tidal marsh, Suisun Marsh area. Presumed extirpated in Petaluma River marshes. Putative San Francisco (city) record is erroneous interpretation of early San Francisco Bay Area collection acc. L. Heckard.</td>
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<td>Taxon</td>
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<tr>
<td><em>Eleocharis parvula</em></td>
<td>[not reported in early floras; Munz 1959 reported only from coastal salt marshes of San Luis Obispo and Humboldt Cos.]</td>
<td>Rare in brackish tidal marshes of San Pablo and Suisun Bay area. Local in diked baylands, lower Napa River.</td>
</tr>
<tr>
<td><em>Festuca rubra</em> L.</td>
<td>[reported only from generalized habitats in early floras; halophytic populations not distinguished. Speculative likely component of historic sandy salt marsh edges of Central Bay.]</td>
<td>Not currently reported from San Francisco Bay estuarine tidal marsh edges; halophytic populations presumed extirpated. Halophytic populations occur along edges of maritime salt marsh and brackish marsh at Rodeo Lagoon, Limantour Estero, Tomales Bay (Marin Co.).</td>
</tr>
<tr>
<td><em>Glaux maritima</em> L.</td>
<td>Behr 1888: “Salt marshes.”</td>
<td>Few recent reports known from San Francisco Bay or San Pablo Bay salt marshes; reported as infrequent in Petaluma Marsh; local in tidal marsh near mouth of Tolay Creek, Sonoma Co.; occasional to locally frequent in Suisun Marsh area and Fagan Slough (Napa River).</td>
</tr>
<tr>
<td><em>Heliotropium curassavicum</em> L.</td>
<td>Greene 1894: “Borders of salt marshes about San Francisco Bay.”</td>
<td>No current reports known from San Francisco Bay. Recently reported from Suisun Marsh area.</td>
</tr>
<tr>
<td><em>Hemizonia pungens</em></td>
<td>Greene 1894: “Borders of salt marshes about San Francisco Bay.”</td>
<td>Local, infrequent species along tidal marsh edge around the San Francisco Estuary.</td>
</tr>
<tr>
<td>ssp. <em>maritima</em> (E. Greene) [Centromadia maritima Greene]</td>
<td>Jepson 1911: “Marshy shores of... San Francisco and Suisun bays”</td>
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</tbody>
</table>
Table 1.3 (continued) Historic Changes in the Distribution and Abundance of Selected Native Vascular Plant Species Occurring in Tidal Marshes of the San Francisco Estuary

<table>
<thead>
<tr>
<th>Taxon</th>
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<th>Contemporary Distribution</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Juncus effusus</strong> L. var. brunneus</td>
<td><strong>Brewer et al. 1880:</strong> “... common in the salt-marshes about San Francisco Bay.”</td>
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<tr>
<td></td>
<td><strong>Brandegee 1892:</strong> “Salt marshes about the bay shore.”</td>
<td>Rare, local at edges of salt marsh and brackish marsh ecotones in San Pablo Bay (China Camp). No known reports from San Francisco Bay tidal marshes.</td>
</tr>
<tr>
<td></td>
<td><strong>Jepson 1911:</strong> “Common in marshy ground: Monterey to San Francisco and Bolinas Bays and northward.”</td>
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<td></td>
<td><strong>Howell 1949:</strong> “Swamps and swales generally near the ocean... Tiburon; Sausalito.”</td>
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<td></td>
<td><strong>Thomas 1961:</strong> “Usually along or near the coast... Palo Alto, near Alviso.”</td>
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<tr>
<td><strong>Juncus lesueurii</strong> Boland.</td>
<td><strong>Brandegee 1892:</strong> “Salt marshes at Visitacion Bay. South San Francisco.”</td>
<td>Apparently associated with sandy salt marsh edges of maritime coast. Intermediates with <em>J. balticus</em> not uncommon in San Francisco Bay Area tidal marshes; difficult to separate. Rare in south San Francisco Bay tidal marshes; one colony in seep at salt pond edge, Coyote Hills.</td>
</tr>
<tr>
<td></td>
<td><strong>Howell 1949:</strong> “Common along the upper reaches of salt marshes... Tiburon; Tamalpais Valley,... In Marin County,... (J. balticus) is not readily distinguished from J. lesueurii...”</td>
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<tr>
<td><strong>Juncus xiphioides</strong> E. Meyer</td>
<td><strong>Jepson 1901:</strong> “A common species of salt marshes... Berkeley; Belmont... Suisun Marshes.”</td>
<td>Not recently reported; presumed rare or possibly extirpated in most tidal salt marshes of San Francisco Estuary.</td>
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<td></td>
<td><strong>Thomas 1961:</strong> “Occasional in sloughs... Palo Alto, nr. Alviso.”</td>
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<tr>
<td><strong>Lasthenia glaberrima</strong> D.C.</td>
<td><strong>Behr 1888:</strong> “Near salt marshes.”</td>
<td>No other reports, historic or current, are known from San Francisco Bay estuarine marshes.</td>
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<tr>
<td><strong>Lasthenia conjugens</strong> E. Greene</td>
<td><strong>Greene 1894:</strong> “Subsaline soil near Antioch.”</td>
<td>Occurs in alkaline/saline vernal pools bordering salt pond 22 in Fremont, Alameda Co., and in diked baylands at upper end of Hill Slough (Potrero Hills), Solano Co. Historic localities near Mt. Eden along bay shore and near Newark. Rare; federally endangered.</td>
</tr>
<tr>
<td>[<strong>Baeria fremontii</strong> (Torr.) A. Gray in part]</td>
<td><strong>Jepson 1911:</strong> “Subsaline fields in the Bay region; Antioch; Newark, etc.”</td>
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<tr>
<td><strong>Lasthenia platycarpha</strong> (A. Gray) E. Greene</td>
<td><strong>Greene 1894:</strong> “Border of salt marsh north of Vallejo; rare or local.”</td>
<td>Historic locality at Redwood City shoreline. Apparently extirpated from San Francisco Bay estuarine marshes. Occurs infrequently in alkaline vernal pools, Solano Co.</td>
</tr>
<tr>
<td>[<strong>Baeria camosa</strong> E. Greene] [<strong>B. platycarpha</strong> A. Gray]</td>
<td><strong>Jepson 1911:</strong> “Salt marshes at Vallejo (Greene).”</td>
<td></td>
</tr>
<tr>
<td><strong>Lasthenia glabrata</strong> Lindley sp. glabrata</td>
<td><strong>Behr 1888:</strong> “Common.”</td>
<td>Currently reported within San Francisco Bay Estuary only from Point Pinole (Whittier marsh) salt marsh and new seeded levee slope at Sonoma Baylands. Many historic salt marsh collections known from Burdell, Alvarado, Mt. Eden, Alameda, Mowry’s Landing, Denver. Maritime salt marsh population occurs in Limantour Estero, Marin Co.</td>
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</table>
### Table 1.3 (continued)  Historic Changes in the Distribution and Abundance of Selected Native Vascular Plant Species Occurring in Tidal Marshes of the San Francisco Estuary

<table>
<thead>
<tr>
<th>Taxon</th>
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</thead>
</table>
| *Lasthenia minor* (DC.) Omd.  
[Baeria minor (DC.) Ferris  
[Baeria uliginosa Nutt.]  
[Lasthenia uliginosa (Nutt.) E. Greene)] | **Brandegee 1891:** “About the borders of marshes, Islais Creek, Visitation Valley, Presidio, South Francisco.” | Apparently extirpated from San Francisco Bay estuarine marshes. |
| *Lathyrus jeffsonii* E. Greene var.  
*jeffsonii* E. Greene | **Greene 1894:** “Suisun marshes.”  
**Jepson 1911:** “Suisun marshes.” | Occasional to rare in Suisun Marsh. Also occurs locally in tidal brackish marshes along Napa River (Dutchman Slough). May be under-reported in drought years. |
| *Layia chrysanthemoides* (DC.) A. Gray  
[Blepharipappus chrysanthemoides Greene] | **Howell 1949:** “Locally common on flats bordering the salt marshes: Ignacio…Chileno Valley.”  
**Thomas 1961:** “occasionally in low alkaline soils of San Francisco Bay… Millbrae, Redwood City.” | No current reports known from San Francisco Estuary tidal marsh edges. |
| *Leymus triticoides* (Buckley) Pilger  
[incl. *Leymus* X *multiflorus* (Gould) Barkworth and D.R. Dewey  
[Elymus triticoides Buckley)] | [general grassland habitats reported historically. Presumed abundant or dominant species of historic tidal marsh edges.] | Occurs locally (abundant) at salt marsh edges at Newark, Alameda Co.; Rush Ranch, Solano Co; Petaluma Marsh, Marin Co.; China Camp, Marin Co.; Dutchman Slough, Solano Co. |
| *Lepidium dictyotum* A. Gray | **Greene 1894:** “Along the borders of marshes at Alameda.” | No current reports known from San Francisco Bay tidal marsh edges. Presumed extirpated or rare in Estuary. |
| *Lepidium latipes* Hook. | **Greene 1894:** “in saline soil at Martinez, Alameda, etc.”  
**Jepson 1901:** “…alkali flats…Martinez.” | No current reports known from San Francisco Bay tidal marsh edges. Reported rarely in diked baylands and tidal marsh edges, Solano Co. (Suisun Marsh area). |
| *Lepidium oxycarpum* Torrey and A. Gray | **Greene 1894:** “Borders of salt marshes at Vallejo…also in subsaline soils… near Alameda.”  
**Howell 1949:** “A rare peppercress of alkaline valley floors and of saline flats adjacent to coastal salt marshes, in Marin Co. known only from low pastures bordering San Francisco Bay near Novato.”  
**Munz 1959:** “V. Grassland and edge of Coastal Salt Marsh; largely about San Francisco Bay.”  
**Thomas 1961:** “Saline and alkaline flats along San Francisco Bay and Santa Clara Valley: Redwood City, Cooley’s Landing, Palo Alto, Mayfield.” | No current reports known from San Francisco Bay tidal marsh edges. Rare, Suisun Marsh edges. |
| *Lilaepopsis masonii* Mathias and Constance  
[Lilaepopsis lineata (Michx.) Greene, in part] | **Jepson 1911:** [as *L. lineata*, in part] “Salt marshes or brackish mud flats… Port Costa to Antioch; Robert’s Island.” | Rare in tidal brackish tidal marshes, Napa Marsh, Suisun Marsh area, to Tolay Creek, San Pablo Bay. Uncommon in western Sacramento river delta fresh-brackish marshes. |
Table 1.3 (continued)  Historic Changes in the Distribution and Abundance of Selected Native Vascular Plant Species Occurring in Tidal Marshes of the San Francisco Estuary

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<tbody>
<tr>
<td><strong>Lycopus asper</strong> E. Greene [Lycopus lucidus Turcz. ] [L. lucidus Benth. misapplied]</td>
<td>Jepson 1911: “Salt marshes: Suisun; Benicia; San Francisco.”</td>
<td>No current reports known from San Francisco tidal marsh edges; presumed rare or extirpated in Estuary there.</td>
</tr>
<tr>
<td></td>
<td>Hickman et al. 1993: Uncommon. Moist areas, marshes, streambanks… Deltaic GV, SnFrB, GB; to w Can, Great Plains.</td>
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<tr>
<td><strong>Plagiobothrys glaber</strong> (A. Gray) I.M. Johnston [Allocarya salina Jepson] [Allocarya glabra Macbr.</td>
<td>Jepson 1911: “Alvarado [Union City], margin of salt marshes.”</td>
<td>Hickman 1993: “PRESUMED EXTINCT. Wet, alkaline soils in valleys, coastal marshes…CCo, sSnFrB… Perhaps a var. of P. stipitatus.”</td>
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<tr>
<td></td>
<td>Munz 1959: “Coastal Salt Marsh; s. shore of San Francisco Bay.”</td>
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<td></td>
<td>Best et al. 1996: “salt marsh near Sears Point, Keck 1935”</td>
<td>Reported at Sonoma Baylands, high tide line, 1996. Otherwise no current reports known from San Francisco Estuary tidal marsh edges</td>
</tr>
<tr>
<td><strong>Plagiobothrys stipitatus</strong> (E. Greene) var. stipitatus</td>
<td></td>
<td>Rarely reported from San Francisco Bay Area high tidal marshes (Suisun Marsh). No recent localities in tidal marsh edges confirmed.</td>
</tr>
<tr>
<td></td>
<td>Greene 1894: “Borders of saline or brackish marshes; quite common about the Bay.”</td>
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<tr>
<td></td>
<td>Howell 1949: [Marin Co.; not reported from estuarine stations]</td>
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<td></td>
<td>Thomas 1961: [SW San Francisco Bay] “... edges of salt marshes...Mayfield, Alviso...”</td>
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<tr>
<td></td>
<td>Best et al. 1996: [Sonoma Co.] “uncommon, salt marshes...Petaluma, Congdon (1880); 5 mi. n. of Sear’s Point, Rubzoff (1970).”</td>
<td></td>
</tr>
<tr>
<td><strong>Plantago maritima L</strong> [P. maritima L. ssp. juncoides (Lamk.) Hulten, P. juncoides Lamk. var. juncoides]</td>
<td>Greene 1894: “... sandy salt marshes”</td>
<td>Infrequent to rare in San Francisco Bay tidal marshes, mostly Richardson Bay. Relatively common in maritime salt marshes, and occasional in Suisun Marsh (Hill Slough).</td>
</tr>
<tr>
<td></td>
<td>Jepson 1911: “… West Berkeley...”</td>
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<tr>
<td></td>
<td>Howell 1949: [Marin Co.] “occasional in salt marshes bordering the bay or ocean: Almonte...”</td>
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<tr>
<td></td>
<td>Thomas 1961: “occasional in salt marshes and on coastal bluffs as far south as San Mateo County: San Francisco, Redwood City, and Mayfield.”</td>
<td></td>
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<tr>
<td></td>
<td>Best et al. 1996: [Not reported from estuarine Sonoma Co. stations]</td>
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</thead>
<tbody>
<tr>
<td><em>Pluchea odorata</em> (L.) Cass [P. camphorata (L.) DC. and P. purpureascens (Sw.) DC., misapplied]</td>
<td>Behr 1888: Salt marshes.</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Greene 1894: “Borders of brackish marshes about Suisun Bay, etc.”</td>
<td>No current reports known from San Francisco or San Pablo Bays; uncommon in Suisun marshes.</td>
</tr>
<tr>
<td></td>
<td>Jepson 1911: “Common in the salt marshes about Suisun and San Francisco Bays.”</td>
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<tr>
<td></td>
<td>Howell 1949: [Marin Co.; not cited]</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Thomas 1961: [SW San Francisco Bay; not cited]</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Thomas 1961: “Plants and salt marsh along San Francisco Bay”.</td>
<td></td>
</tr>
<tr>
<td><em>Puccinellia nutkaensis</em> (J.S. Presl.) Fern. and Weath. [P. grandis Swallen]</td>
<td>Thomas 1961: “Levees and salt marsh along San Francisco Bay”.</td>
<td>Rare, local, south San Francisco Bay; No records of San Pablo Bay collections in Howell 1949, Best et al. 1996.</td>
</tr>
<tr>
<td></td>
<td>Jepson 1911: “Marshes bordering San Francisco Bay.”</td>
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<tr>
<td></td>
<td>Munz 1959: “Coastal, often brackish marshes, San Francisco Bay.”</td>
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<td></td>
<td>Hickman et al. 1993: “Uncommon. wet +/- salty places.”</td>
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<tr>
<td><em>Salicornia subterminalis</em> Parish [Arthrocnemum subterminale (Parish) Standley]</td>
<td>[Not reported from estuarine stations in early floras.]</td>
<td>Local, rare in South Bay, south Fremont, Milpitas (in diked wetlands, former Fremont Airport), and at Hill Slough, Suisun Marsh.</td>
</tr>
<tr>
<td></td>
<td>Greene 1894: “In lowlands adjacent to salt marshes near Alameda, San Francisco, etc.”</td>
<td></td>
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<tr>
<td></td>
<td>Jepson 1911: “Local species of low and wet adobe lands in the vicinity of salt marshes bordering San Francisco Bay; near Alameda...and Potrero Hills, San Francisco, the only recorded localities.”</td>
<td></td>
</tr>
<tr>
<td><em>Senecio hydrophilus</em> Nutt.</td>
<td>Brewer et al. 1880: “Salt marsh at Vallejo (Greene).”</td>
<td>Apparently extirpated in San Francisco, San Pablo Bay (incl. Petaluma R.); infrequent but locally common in Suisun Marsh area and Carquinez Strait tidal marshes; possibly Napa R.</td>
</tr>
<tr>
<td></td>
<td>Greene 1894: “Brackish marshes, formerly plentiful at West Berkeley, and on the lower Napa River; still abundant in the Suisun marshes.”</td>
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<td></td>
<td>Jepson 1911: “Abundant in the Suisun Marshes and found in other marshes about San Francisco Bay”</td>
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<tr>
<td></td>
<td>Hickman 1993: “Reduced from wetland development.”</td>
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### Table 1.3 (continued)  Historic Changes in the Distribution and Abundance of Selected Native Vascular Plant Species Occurring in Tidal Marshes of the San Francisco Estuary

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<tbody>
<tr>
<td><em>Solidago confinis</em> Nutt.</td>
<td><strong>Jepson 1911</strong>: “Salt marshes, San Francisco Bay, Bolander(1863).”</td>
<td>Rarely collected.</td>
</tr>
<tr>
<td><em>Sium suave</em> Walter</td>
<td><strong>Jepson 1911</strong>: “Suisun marshes; Stockton”</td>
<td>Rare in Suisun Marsh, primarily in wet years. Recently observed near Rush Ranch, Hill Slough, and Brown’s Island.</td>
</tr>
<tr>
<td></td>
<td><strong>Brewer et al. 1880</strong>: “In salt-marshes on the coast, about San Francisco.”</td>
<td>Extinct in San Francisco Bay Estuary, probably since ca. 1950. Restricted to one large population at Morro Bay, where it occurs primarily along sandy high marsh edges. Planned for reintroduction. Misreported occurrences often due to confusion with <em>Suaeda calceoliformis</em>, <em>S. moquinii</em>, <em>Salsola soda</em>, and <em>Bassia hyssopifolia</em>.</td>
</tr>
<tr>
<td><em>Suaeda californica</em> Wats.</td>
<td><strong>Behr 1888</strong>: “Salt marshes on an island near Alameda.”</td>
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<td></td>
<td><strong>Greene 1894</strong>: “Vicinity of sand beaches about San Francisco Bay, but seldom seen.”</td>
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<tr>
<td></td>
<td><strong>Jepson 1911</strong>: “Sandy beaches bordering San Francisco Bay, the known stations few: San Pablo Landing; Bay Farm Island.”</td>
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</tr>
<tr>
<td></td>
<td><strong>Thomas 1961</strong>: “Occasional in salt marshes along San Francisco Bay; San Francisco and Palo Alto.”</td>
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</tr>
<tr>
<td><em>Suaeda californica</em> Wats.</td>
<td><strong>Greene 1894</strong>: “Vicinity of sand beaches about San Francisco Bay, but seldom seen.”</td>
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<td></td>
<td><strong>Greene 1894</strong>: “Vicinity of sand beaches about San Francisco Bay, but seldom seen.”</td>
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<td></td>
<td><strong>Jepson 1911</strong>: “Sandy beaches bordering San Francisco Bay, the known stations few: San Pablo Landing; Bay Farm Island.”</td>
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<tr>
<td></td>
<td><strong>Thomas 1961</strong>: “Occasional in salt marshes along San Francisco Bay; San Francisco and Palo Alto.”</td>
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### References


Grewell, B. 1993 (et seq.). Unpublished vascular plant species list, Suisun Marsh. [References continued on page 29]


Peter R. Baye

Introduction

This report focuses on wetland areas within historic tidal marshes that have been isolated from tidal action by dikes (levees) and converted to non-tidal salt marsh, non-tidal brackish marsh, or subsaline to freshwater seasonal wetlands. These areas are referred to herein interchangeably as “diked wetlands” or “diked Baylands.” Because instantaneous salinity (or even average annual salinity) of diked wetland soils does not consistently correspond with plant community composition, and varies over time, these salinity categories are intended to be broadly descriptive of plant associations rather than quantitative threshold values of soil salinity. Accordingly, the marsh types described are not discrete, but intergrade continuously and may vary over time at any site. Diked wetlands as treated below do not include artificial salt ponds (treated separately) or “muted tidal” managed marshes (marshes with reduced tidal range controlled by tidegates), and cover only wetlands with non-tidal hydrologic inputs (rainfall, groundwater, surface runoff, streamflow, engineered water control structures, or very infrequent overtopping of dikes by extreme tides).

Published and unpublished sources of useful, precise data and other information on the vegetation and flora of diked Baylands are very scarce. Most usually are limited to short-term observations and coarse descriptive accounts (such as lists of dominant species) at a particular time of year, or generalized accounts of resource management plans (e.g., Eicher 1988, Hudson 1980). Vegetation was usually described for wildlife habitat evaluation, rather than for floristic analysis or quantitative plant community description. Relatively more detailed information about some individual diked Bayland sites is sometimes available for sites which are proposed for major development projects, and become the subject of detailed wetland delineations and field studies for environmental evaluations (e.g., Rugy 1991, Kaufman and Harvey 1987). The level of detail in vegetation analysis of diked Baylands even for site-specific studies was still low until the mid 1980s when technical vegetation criteria for wetland jurisdictional delineations were promulgated (WES 1997). There are no long-term studies of changes in vegetation in diked Baylands. Some coarse information about vegetation change in diked wetlands is available through inspection of historic aerial photographs, particularly color infrared photos from the 1980s to the present. Based on recent information from some of the more intensively surveyed diked wetlands (e.g., Montezuma Wetlands, Solano County; Cullinan Ranch, SOSC-55).
Solano County; Renaissance Faire site, Marin County), it appears that the diversity and dynamics of diked wetland vegetation have been substantially underestimated in past assessments.

Historic information on the diking of San Francisco Bay tidal marshes is based on numerous sources, particularly U.S. Coast Survey maps (multiple series); historic accounts of salt pond levee development (Ver Planck 1958); and field observations of modern levee maintenance and repair methods and agricultural drain systems.

Environmental Setting

The physical origins of diked wetlands are similar throughout the San Francisco Estuary. Most of the tidal marshes were reclaimed for agricultural use in the late 19th century when the use of mechanical dredges became commercially available to landowners (after ca. 1870), although many dikes were constructed manually (Madrone Associates 1977). Tidal marshes were diked for reclamation either as pasture, hayfields, salt ponds, or (rarely) cropland. Reclamation involved construction of dikes (earthen levees made of locally excavated Bay mud) along the margins of marsh plains (middle marsh between approximately MWH and MHHW) where they bordered mudflats or major tidal creeks. The borrow ditches for dike construction were typically located inside of the dike, creating narrow canals about 20 feet from the foot of the dikes. Enclosure of tidal marshes by dikes, and resulting fluctuation between winter flooding and summer desiccation of saline basins, would have rapidly killed most standing tidal marsh vegetation. When levees were stabilized after several lifts (sequential layers of dewatered dredged Bay muds) tidegates were installed to enable the enclosed basins to drain on low tides. After stabilization, dikes typically stood about 3 (to 4) feet above the marsh plain (Ver-Planck 1958).

Environmental Changes From Diking

Following the initial phase of dike construction, several changes occurred. Mature tidal marsh soils accumulated peaty organic matter under anaerobic conditions, which minimizes decomposition. Drained marsh soils high in peaty organic material underwent aerobic decomposition and dewatering, causing land elevations to subside. Dikes also caused compression of underlying plastic clayey silts and peats, and subsided (Madrone Associates et al. 1983). Differential subsidence of the marsh surface tended to exaggerate relict marsh topographic relief, causing natural levees (containing coarser silts) to stand out against isolated depressions where peat content was relatively great, and the effects of aerobic peat decomposition were greatest. Tidal creek topography typically persisted as depressional sinuous swales. Early-succession diked marsh plant communities, typically dominated by perennial pickleweed (Salicornia virginica; drier, more saline conditions) or alkali bulrush (Scirpus maritimus), bulrush (Scirpus californicus; less often S. acutus) or cattails (mostly Typha angustifolia; wetter brackish to subsaline ditches) tend to be best developed in relict swales and depressions. As salts were drained from the diked basins and lands were managed for agriculture, these pioneer diked salt marsh communities were reduced or eliminated (Madrone Associates et al. 1983, Harvey 1987).

Marsh Progradation and "Second Generation" Diked Wetlands

The strong reduction in tidal flows caused by diking all but the largest tidal creeks in the marsh system caused significant increases in sedimentation outside of diked marshes, causing rapid marsh progradation on sloping mudflats. In addition, slow migration of the pulse of hydraulic mining outwash from the Sacramento River contributed to marsh progradation in San Pablo and Suisun Bays (Doane 1999, Jaffe et al. 1998). In some areas (e.g., south of Novato Creek), marsh progradation was so extensive that a second phase of diking occurred in the newly accreted marshes. These progradational marshes are typically broad pickleweed-
dominated plains with fringes of cordgrass, cordgrass/alkali bulrush mixtures, or erosional scarp in pickweed peats. Like early-succession diked salt marsh, they support relatively low salt marsh species diversity, and low densities of narrow, sparsely branched shore-perpendicular tidal creeks. Because of the influence of wave deposition of sediment and coarse organic debris, the tidal elevations of these dike-fringing salt marshes is often above MHHW in some areas, particularly where incipient natural levees form at the edge of mudflats. These secondary prograded high marshes with little antecedent topography were readily converted to diked agricultural land, as in the Baylands of Novato (H. amilton, Bel Marin Keys).

Dike Disturbance Cycle and Vegetation - Subsidence of dikes themselves caused a need to maintain dike crest elevations by dredging borrow ditches to resupply material. This established a periodic disturbance regime to dike vegetation and adjacent ditches. In areas of high wave energy (long fetch distance, narrow mudflats), maintenance by topping dikes with dredged muds and restoring erosional slopes may occur in cycles as short as five years or less. Many bayfront dikes unsheltered by fringing marsh require aroring by placement of rock or concrete fragments. Well-protected dikes behind extensive salt marsh on firmer peats may have maintenance cycles longer than a decade or two. Repaired dike slopes provide bare mineral substrate which is gradually leached of salts and open to colonization by upland weeds.

The dike disturbance cycle has favored a ruderal flora along the upper slopes and crests of dikes (including many native and exotic halophytic weeds as well as glycophytes; e.g., mustard (H. irschi-feldia incana, Brassica spp.), radish (Raphanus sativus), fennel (Foeniculum vulgare), plantain (Plantago coronopus, P. major, P. lanceolata), annual ice-plant (Mesembryanthemum nodiflorum; mostly South Bay), sea-fig (Carpobrotus chilense), hottenot fig (Carpobrotus edulis and hybrids with C. chilense), poison hemlock (Conium maculatum), Mediterranean brome species (Bromus spp.), wild barley (Hordeum murinum ssp. gussonianum.), ryegrass (Lolium multiflorum, L. perenne). Lower portions of disturbed outboard (bayward) dike slopes are typically more saline and wetter, and support brackish marsh or salt marsh species, often with an exaggerated proportion of weedy halophytes (e.g., spearscale, Atriplex triangularis perennial peppergrass, Lepidium latifolium, sicklegrasses Parapholis incurva, Hainardia cylindrica; bassia, Bassia hysopifolia; saltwort, Salsola soda; wild beet, Beta vulgaris). Interior slopes of dikes which face salt ponds, and contiguous fringing nontidal saltmarsh, are either bare or vegetated with saltgrass (Distichlis spicata), Salicornia virginica, dodder (Cuscuta salina), and alkali-heath (Frankenia salina). Dikes with very infrequent maintenance tend to become dominated by dense stands of coyote brush (Baccharis pilularis, South Bay) or mixed coyote brush and bee-plant (Scrophularia californica; North Bay), often with sub-dominant mustard, poison-hemlock, and radish. High marsh halophytes (pickleweed, alkali-heath, gumplant, spearscale) tend to dominate the lower portion of the outboard dike slopes adjacent to salt marshes, although weedy species can persist for many years after a levee has been disturbed by maintenance and repair activities.

The ecological significance of the dike disturbance cycle for wetland plants is that it has provided corridors through tidal marshes and diked marshes for a weedy flora (both exotic halophytes and glycophytes) to disperse, and places weed seed sources along a topographically superior location for dispersal into adjacent diked and tidal wetlands. The rapid local spread of weedy halophytes on dreg spoils along recently maintained/repaired dikes (especially Salsola soda, Lepidium latifolium, Mesembryanthemum nodiflorum) can be observed throughout the Bay. Similar halophyte weed dispersal occurs along side-cast spoils in diked marshes where drainage ditches are created or maintained. Dike disturbance corridors may accelerate the spread of exotic halophyte population outposts into uninvaded wetland habitats. In particular, Lepidium latifolium's invasion of brackish marshes appears to have tracked patterns of dike disturbance, invading first along dredge spoil at levee edges, subsequently spreading into diked and tidal wetlands.

Hydrologic Changes in Diked Wetlands - Patterns of soil waterlogging and inundation in diked conditions differ fundamentally from tidal marsh. They depend principally on the efficiency of artificial drainage, the permeability of substrate (related to soil clay content), and the amount and seasonal distribution of rainfall. The efficiency of the early drainage systems in diked marshes was based on the amount of ditching and the pattern of ditching in relation to subsided marsh topography. Because of the great extent of the areas diked, density of drainage ditches was relatively low. Ditches were mostly confined to the borders of farmed parcels, but sometimes

Diked wetlands in Suisun are managed primarily for waterfowl production.
reached across extensive marsh depressions. The drains were originally driven by gravity, drawing drainage water downslope to one-way flapgates which discharged to adjacent tidal marshes at low tide. This original gravity-driven drainage system had limited efficiency. Topographic lows in the diked basins (swales of relict tidal creeks, relict marsh pans) remained poorly drained into the crop growing season, while relict creek levees and higher relict tidal marsh became better drained. Relictual tidal marsh patterns of wetland and upland are evident in black and white photographs of diked hayfields in the mid-20th century. Even with modern pump-driven drainage systems, persistent soil waterlogging and inundation in depressions occurs following rainstorms (Granholm 1986, Madrone Associates et al. 1983).

The proportions of poorly drained (waterlogged or inundated in spring) and well-drained (aerobic soils in spring) soils in diked Baylands vary with precipitation amounts and patterns. Years of normal or above normal rainfall, particularly those with large storms late in the precipitation season, cause expansion of wetland areas in diked conditions. These contract during years of below-normal precipitation, especially with a lack of spring storms. The proportions of effective wetland and upland also vary with drainage efficiency and the degree of subsidence.

Long-term Drainage of Diked Wetlands – As subsidence increased, wetland areas increased behind dikes, particularly in peaty soils. In the early 20th century, many diked farmlands failed because the costs of compensating for increased subsidence and dike degeneration at times exceeded the return on agricultural benefits. Many derelict agricultural parcels with degenerated dikes are evident in aerial photographs of San Pablo Bay in the 1940s. After abandonment of diked farmlands, partial levee and drain failures increased, causing reversion of agricultural lands to brackish or salt marsh conditions. For example, prior to conversion to salt ponds, many of the Napa Marsh area’s derelict hayfields in the 1940s had partially reverted to wetland (Madrone Associates 1977).

Contemporary Drainage of Diked Wetlands – Today, subsidence of diked active agricultural lands has increased to the point at which it cannot be compensated by passive gravity drainage through flapgates alone; drainage sufficient for oat hay farming depends primarily on active pumping of water in ditches for discharge to the Bay. It is common for elevations in diked Baylands of San Pablo Bay to average as low as -0.10 ft N.G.V.D., and some average below -3.0 ft or more over extensive areas, as at Bel Marin Keys and Hamilton Field (USACE 1988). In south San Francisco Bay, which was affected by past subsidence due to long-term groundwater extraction, diked wetland elevations may be even lower (Moffett and Nichols and Phil Williams Associates 1988). These subsided diked marsh surfaces are often very close to the groundwater surface. Accordingly, the proportion of wetland and upland in contemporary conditions depends on the intensity of pumping and ditch maintenance. These conditions vary significantly among diked parcels under different ownership and management. Therefore, the mosaic of wetland and upland in diked agricultural lands is relatively variable and unpredictable among years and between parcels.

Variability of Artificial Hydrologic Conditions – The patchiness and instability of diked wetlands is evident, for example, in recent land-use changes in San Pablo Bay. Cullinan Ranch, actively drained and farmed oat hayfield until the early 1990s, supported a matrix of upland cropland and many seasonally wet depressions with wetland weeds. After cessation of pumping by the mid-1990s (a period of above average rainfall), the Ranch rapidly (within 2 years) and spontaneously converted to a seasonal freshwater marsh dominated by cattails and flats of Eleocharis parvula (Takekawa et al. 1999). Near by, between Tolay Creek and the Petaluma River, adjacent hayfields with differing schedules of ditch maintenance changed from very similar extensive winter-ponded swale patterns to striking contrasts of ponded and drained fields. At another location near Sears Point, San Pablo Bay, cessation of pumping in relict hayfields caused conversion to seasonal wetlands dominated by annual plant species typical of vernal pool communities (many of which are present in the ephemeral weed flora of depressions within hayfields; Owna pulchella, Plagioothrys bracteatus, Eryngium aristatum, Callitriche spp., Eleocharis macrostachya). Thus, the extent of diked wetlands and their character today are very much artifacts of drainage pump activity.

Similar artificial drainage controls wetland plant communities in the diked basins of the Suisun Marsh, which is managed mostly for waterfowl production. There, relatively low-salinity tidewaters are admitted to the basins selectively to sustain fresh-brackish perennial and seasonal marshes (Jones and Stokes 1976, Mall 1969, Miorin et al. 1991). The proportion of ponded to vegetation may be controlled by modifying managed hydroperiods, so that prolonged flooding causes dieback of vegetation in areas of relatively lower substrate elevation. The seasonal variations in tidewater salinity enable the timing of flooding to control substrate salinity, also. Managed marsh hydroperiods are usually designed to favor mixtures of shallow submerged mud, bulrushes (Scirpus maritimus, S. americanus, S. pungens), tules (S. californica, S. acutus), cattail (Typha spp.) and brass-buttons (Cotula coronopifolia), and some non-native annual grasses (Echinochloa crus-galli, Polygopon monspeliensis). Also common in diked brackish marshes are baltic rush (Juncus balticus), saltgrass (Distichlis spicata) and pickleweed (Salicornia virginica). Other species have colonized these brackish managed wetlands, including goosefoot (Chenopodium chenopodioides), docks (Rumex crispus, R.
very rapidly except where drainage is very poor. Residual salinity tends to decline unless substantial maintenance efforts are applied to drainage and dikes. Diked wetlands become realigned by partial failure of tidegates and levees (Madrone Associates et al. 1983, Madrone Associates 1977). These areas are often barren of vegetation, or develop sparse, low diversity vegetation. Less extreme but low pH in diked wetlands may inhibit plant production, but the abundant phytomass of many diked wetlands (e.g., rank growth of pickleweed, cattails, peppercress) suggests that the seasonal drainage and aeration of diked wetland soils commonly has a stronger overall effect on vegetation production than low pH. Extremely low redox potential and sulfite toxicity, which often accompany low pH, are highly significant inhibitors of plant growth (Russell 1973). Soil acidity is highly variable in diked wetlands and depends on local soil conditions and prevailing hydroperiods.

Diked wetlands which have been effectively desalinized for agricultural production do not remain so unless substantial maintenance efforts are applied to drainage and dikes. Diked wetlands become realigned by partial failure of tidegates and levees (Madrone Associates et al. 1983). Leaking or ruptured tidegates allow influx of saline tidal waters in drainage ditches. Saline or brackish ditch water can recharge salts locally in groundwater, and move into the soil through evapotranspiration and capillary movement. In diked wetlands with salt marsh soils (H Arvey 1987, Madrone Associates et al. 1983, M Eiorin et al. 1991). The desalinated conditions of the substrate were maintained by drainage through ditches and tidegates. Subsidence caused (and continues to cause today) decreased efficiency of drainage, and therefore also decreased flushing of residual or reintroduced salts.

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Acidification of Diked Wetlands – Soil acidity affects plant growth primarily by altering the availability of soil nutrients, or liberating excessive amounts of otherwise low-solubility ions into the soil solution, creating toxicity problems for roots. Acid-related toxicity occurs only at very low pH (Reuss and Johnson 1986). Soil acidity is normally not highly variable in tidal salt marshes, which are buffered by cations of estuarine water and relatively stable reduction-oxidation conditions established by groundwater surface position in the marsh soil profile (Adam 1990). In diked conditions, extreme seasonal fluctuations in the soil saturation levels may occur, causing release of sulfides and free metals in marsh soils with high sulfur contents. Some depressions in diked wetlands develop very low pH (pH 4 and occasionally lower) and high concentrations of iron oxide precipitates (Madrone Associates et al. 1983, Madrone Associates 1977). These areas are often barren of vegetation, or develop sparse, low diversity vegetation. Less extreme but low pH in diked wetlands may inhibit plant production, but the abundant phytomass of many diked wetlands (e.g., rank growth of pickleweed, cattails, peppercress) suggests that the seasonal drainage and aeration of diked wetland soils commonly has a stronger overall effect on vegetation production than low pH. Extremely low redox potential and sulfite toxicity, which often accompany low pH, are highly significant inhibitors of plant growth (Russell 1973). Soil acidity is highly variable in diked wetlands and depends on local soil conditions and prevailing hydroperiods.

Disturbance in Diked Wetlands – The disturbance regimes of diked wetlands are influenced primarily by discing and flooding. Discing is performed for agriculture, suppression of weed biomass, and suppression of mosquito production. Episodes of discing have maintained a significant ruderal (weedy) element to the diked wetland flora of San Francisco Bay, creating large vegetation gaps suitable for invasion by non-native plants, particularly annuals. Extreme flooding events which are possible in non-tidal diked marshes also cause disturbances: deep, prolonged flooding causes mass dieback of most standing perennial vegetation. Following dieback events, similar or very dissimilar plant associations may establish.

Diked Bayland Plant Communities

The plant communities present in the diked Baylands can resemble those of local tidal salt marshes, tidal brackish marshes, non-tidal perennial freshwater marshes, or seasonally wet grasslands. Some also have characteristics similar to components of tidal marshes which are now regionally scarce or extirpated, such as high marsh pans and alluvial high marsh ecotones. Diked wetlands usually have lower native species richness than their analogous natural plant communities, and often a larger component of exotic plant species. The typical “weedy” character of many diked wetlands is probably more a result of past land uses rather than an intrinsic susceptibility to invasion by exotic vegetation. Some diked wetlands are managed actively to maintain community dominance by marsh plant species favored by wildlife or game manag-
Plant community composition in diked wetlands is strongly influenced by the degree of residual soil salinity or salt recharge of soils, the efficacy of artificial drainage, and the relative factors of land use history. These factors vary extremely in diked Baylands: some exhibit insignificant salinity, maximal drainage and disturbance in some intensively cropped oat hayfields in San Pablo Bay; others exhibit high salinity, poor drainage and little disturbance in diked pickleweed marshes in south San Francisco Bay. Other modifications persisting from past land uses which affect plant community composition include importation of soils or fill (e.g., former airport landing strips, dike building pads), abandoned berms and ponds of gun clubs, residual effects of past fertilizer applications; industrial waste disposal, and soil contamination.

**Relict Halophytic Vegetation** – The majority of diked wetlands in central and southern San Francisco Bay are dominated by species native to local tidal salt marshes and brackish marshes, such as Distichlis spicata and Salicornia virginica (BCDC and Harvey 1983, Madrone Associates 1983). Salt-tolerant glycophyte species have very low physiological nutritional requirements for salt, and flourish in non-saline and subsaline soils (Waisel 1972). They often co-exist with species with little affinity for saline soil, such as Polygonon monspeliensis and Lolium multiflorum (Harvey 1987, Kaufman and Harvey 1987). S. virginica and D. spicata have a significant competitive advantage over salt-intolerant plant species when substrate salinities are in the range of halophytes (over 5 ppt soil salinity), and rapidly establish dominance during episodes of high salinity conditions. Some halophytes like S. virginica are efficient colonizers of bare wet mud even when salinity is low if seed rain intensity is high. Pioneer halophytes do not necessarily decline in abundance, however, when substrate salinities decline as a result of progressive leaching and drainage of salts. Many apparent diked “salt marshes” are composed of relict vegetation halophyte vegetation which persists in relatively low salinity conditions. This condition is indicated by the presence of a minor to subdominant component of species with relatively low salt-tolerance (e.g., ruderal composites, bedstraws, mustards), growing vigorously among halophytes without indications of salt stress (stunted growth, leaf tip burn, pale leaves) in diked “salt marshes.” Examples are sometimes found in abandoned dredge disposal sites (Zentner and Zentner 1995, Huffman and Associates 1996). Some mixed halophyte-glycophyte associations may also occur where stratification of rooting zones occurs in distinct salinity horizons, caused by near-surface leaching of salts and accumulation in deeper portions of the soil profile.

Thus, apparent salt marsh vegetation in diked Baylands may indicate either current high salinity or former high salinity, and does not necessarily indicate sustained high residual salinity. It often represents inertia in plant community structure after relaxation of salinity stress. The term “non-tidal salt marsh” in the context of the San Francisco Bay Estuary should be interpreted narrowly in floristic rather than physiological terms, because dominance of halophytes in the unstable substrate salinity conditions in diked wetlands is an unreliable indicator of current substrate salinity. Some diked salt marshes are truly saline and tend to remain so because of chronically poor drainage or frequent partial dike failures. Others are in gradual succession to other vegetation types. Some diked salt marshes with low residual substrate salinity are subject to rapid conversion to other vegetation types following disturbances (e.g., discing or flooding).

**Species Richness and Composition** – The species richness and composition of diked marshes is highly variable among sites, and among different marsh types. High salinity and hypersalinity in diked marshes tend to promote low species diversity, selecting for a few tolerant species. Other extreme soil conditions, such as strong acid production and mass release of free iron (often associated with prolonged inundation followed by summer drought) minimize plant species diversity. Truly hypersaline seasonal wetlands in the Bay usually support only sparse Salicornia virginica, Distichlis spicata, and Salsola soda, with a minor component of Frankenia salina. Hypersaline seasonal wetlands are now scarce in the Bay Area, mostly scattered around South Bay salt ponds and adjacent lands. A few occur in the North Bay (e.g., parts of Gallinas Creek diked salt marshes, peripheral portions of the Napa salt ponds). Many former hypersaline diked wetlands have been altered by water management for mosquito abatement and wildlife habitat enhancement, and are now muted tidal marshes (e.g., N ew Chicago Marsh in Alviso, Oro Loma Marsh in Hayward).

Species diversity in nontidal diked brackish and salt marshes is generally much higher than in hypersaline basins, but this does not reflect relatively greater overall diversity of native plant species. Diked brackish to saline nontidal wetlands support a number of common native tidal brackish and salt marsh species (Salicornia virginica, Distichlis spicata, Frankenia salina, Cuscuta salina, Atriplex triangularis) and sometimes support relatively infrequent native species typical of the natural high tidal marsh and upland ecotype (Iva axillaris, Leymus triticoides, Baccharis douglasii). The native perennial grass Leymus triticoides, historically a dominant species of the upper transition zone of tidal salt and brackish marshes, is infrequently found in some diked brackish marshes,
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Particularly where disturbance has been infrequent. A relatively rare historic component of subsaline tidal marsh ecotones, Centaurium muehlenbergii is found in diked subsaline wetlands at Cullinan Ranch. It is currently reported known from only one tidal marsh/upland ecotone (China Camp). The sedge Scirpus maritimus, a dominant native component of tidal brackish marshes, is often abundant or dominant in brackish to saline ditches or deep, wet depressions in diked marshes ((Madrone Associates et. al. 1983, M all 1969).

Conversely, diked salt and brackish marshes generally fail to support some important species of corresponding tidal marsh communities; Spartina foliosa is excluded from non tidal conditions, and J aumea carnosa, Plantago maritima, Triglochin spp. and Limonium californicum are absent or very infrequent in non tidal salt marsh; Grindelia stricta is generally less abundant in non tidal salt marsh than tidal marsh. Diked salt marshes typically lack rare tidal marsh species (e.g., Cordylanthus spp., Castilleja ambiguca, Lathenia gabrata, Lilaeopsis masonii, Cirsium hydrophilum, Aster lentus), and also usually lack most infrequent tidal marsh species (e.g., Pluchea odorata, Sene
coa hydrophilus, Glaux maritima). The failure of these tidal marsh species in diked conditions is probably due to the relatively greater competition by robust "generalist" species with broad ecological amplitude, and physiological intolerance of extremes of inundation and dryness in diked wetlands. Diked salt and brackish marshes in some cases, however, provide refugia for tidal marsh plants of the high tidal marsh which have become (or in some cases have always been) regionally rare or infrequent in the modern tidal marsh ecosystem, such as Suaeda moquinii, H emizia pvngens spp. maritima, Salicornia subterminalis, Downingia pulchella, Juncus mexicanus). As such, they may serve to maintain genetically differentiated salt-tolerant populations of species displaced from modern tidal marshes.

Diked brackish and salt marshes are subject to invasion by many non-native species and species which are not typical of tidal marshes, or are typically restricted to marginal conditions in tidal marshes. Non-native pasture grasses with moderate salinity tolerance, such as Lolium multiflorum (and hybrids), Polypogon monspeliensis, Lotus corniculatus and Hordeum marismum spp. gusonaeum, and even Rumex crispus are also major components of diked salt and brackish marshes, often locally dominating either depressions (Polypogon, Hordeum) or mounds (Lolium). Exotic halophytic grasses Parapholis incurva and H ainardia cylindrica are also locally common in diked salt or brackish marsh. Cotula coronopifolia is usually only a minor component of tidal salt and brackish marsh, colonizing depressions and marsh pan edges, but is often a major component of diked brackish marshes, particularly in disturbed or winter-ponded brackish depressions where other vegetation has died back after prolonged deep flooding. Other common her
baceous non-native plant species of diked brackish and saline marshes include Lepidium latifolium, Bassia hys-
sopifolia, Beta vulgaris, Salsola kali, and Salsola soda. Le-
pidium latifolium is especially invasive in brackish diked marshes, particularly where the soil has been disturbed, but also in areas of marsh with thin or discontinuous vegetative cover.

Diked subsaline and nonsaline Baylands are very seldom the subject of careful floristic surveys (e.g., Rugt 1991, Madrone Associates 1977); vegetation descriptions usually focus on visually dominant ruderal species, often based on summer survey dates when native annual species are not identifiable (Jones and Stokes 1977, Hudson 1980, Werminski 1973). Consequently, the floristic diversity and affinities of diked subsaline to fresh seasonal wetlands has probably in many (perhaps most) cases been underestimated. Diked subsaline and non-
saline wetlands are mixtures of exotic species of ruderal seasonal wetlands and native species typical of vernal pools and swales. Diked wetlands which are mostly subsaline to nonsaline after years of agricultural drainage support a range of marsh plant associations. These are most common around San Pablo Bay and Suisun Bay, where grazing pasture and oat hayfields have been maintained for many decades in diked former tidal brackish marshes. They may have inclusions of relatively brackish indicator species where soil salinity and acidity are locally elevated (e.g., mixtures of Atriplex triangularis, Polypogon monspeliensis, Distichlis spicata) but are domi-
nated by glycophytic wetland plant species, both native and non-native. Composition of the fresh/subsaline diked wetland flora is influenced by disturbance. Annu-
ally diked hayfields support wetland "weeds" which are a mixture of native annuals (e.g., Plagiobothrys spp., esp. P. stipitatus, P. leptolatus), Juncus bufonius, Lilaea scilloides, Callitriche marginata, C. spp. Ciceria quadran-
gularis, Elati ne brachysperma, Eryngium spp., Cressa truxillensis locally, D owningia spp.; non-native annuals (Lythrum syzygosfium, Cotula coronopifolia, Polygonum aviculare, Hordeum marismum spp. gusonaeum, Polypogon monspeliensis) and non-native perennials (Lotus cornicu-
lagus, Agrostis avenacea). Grazed pasture land in diked Baylands in San Pablo Bay may also support native an-
nuals found in diked disked hayfields, as well as native perennials (Eleocharis macrostachya, Glyceria spp. Juncus effusus J. patens) and naturalized non-native perennials (Rumex crispus, R. pulcher, Cirsium arvense, Lolium multiflorum). The relative abundance of these species in diked pasture and hayfield wetlands is variable and unstable. Some diked wetlands, after relaxation of intensive agricultural manipulation, develop seasonal wetlands with plant species composition highly similar to that of regional vernal pools and swales (locally dominated by Downingia spp., Eryngium spp. Eleocharis macrostachya, Callitriche spp. Lilaea scilloides, Plagiobothrys spp., etc.)
Reference Sites

Reference sites for different types of diked wetlands would generally not be long-lived because of the prevalence of unstable vegetation conditions in diked Baylands. Droughts, wet years, changes in drainage and pumping, disturbances from agricultural practices, and succession can cause profound changes in vegetation in short periods of time. The following reference sites reflect conditions observed in the mid-late 1990s.

1. Diked non-tidal salt marsh (dominant Salicornia virginica)
   - Fremont Airport (King and Lyons site; proposed for phased tidal restoration), Alameda Co.
   - Gallinas Creek diked wetlands, Marin Co.
   - Western Marsh and Central Lowlands, Bahia Site, Novato, Marin Co.
   - Dredge pond 3E, Mare Island, Solano Co.
   - Area H, Redwood Shores, San Mateo Co.

2. Diked non-tidal brackish marsh
   - Cullinan Ranch, Solano Co.
   - Suisun Marsh managed marshes, Solano Co.
   - Huichica Unit, CDFG Napa-Sonoma Marsh, Sonoma Co.

3. Diked subsaline to nonsaline seasonal wetlands
   - Black Point/Renaissance Faire site, Novato, Marin Co. (extirpated 1999)
   - Twin House Ranch Site, Lower Petaluma River, Sonoma Co.
   - Leonard Ranch, North Point, Dixon parcels, Sonoma Co., along Hwy 37

Historic and Modern Distribution

Wetlands of diked Baylands are relatively recent historic artifacts. The plant associations they support are analogous to, but distinct from, wetlands along the margins of historic tidal marshes. Brackish non-tidal marshes somewhat similar to diked brackish marshes probably occurred within alluvial deposits at mouths of small streams which discharged into tidal marshes with locally poor drainage, such as near Ignacio (Novato), where riparian areas converged with dense marsh ponds and few or no tidal creeks. Analogous examples of brackish or subsaline marshes with marginal tidal flooding are found today along Drakes Estero and Tomales Bay, particularly near shallow backbarrier lagoons. Salt marsh with restricted tidal influence probably occurred along portions of the Bay where local sand beach ridges were likely to obstruct tidal flows. One modern example exists at Pinole Point (Whittier Marsh), where the proximal end of a sand spit episodically dams small tidal channels, causing seasonal ponding in a small salt marsh cut off from regular tidal flows. Prehistoric examples of “pocket” nontidal salt marsh probably occurred in the vicinity of Richardson Bay, Alameda, Oakland, and the San Francisco Peninsula, where sand beach ridges occurred.

Seasonal freshwater wetlands (vernal pools and swales, springs) occurred within grasslands peripheral to the Bay, particularly in the Petaluma River valley, on alluvial terraces near Fremont, portions of Richmond and Berkeley, and along much of the Suisun Marsh area. Their distribution and abundance, as suggested by soil surveys, were probably not limited to areas mapped as poorly drained; seasonal freshwater wetlands often occur as local inclusions within soil series in which wetlands are not indicated as prevalent. This is indicated by records of vernal pool endemics in locations like San Francisco, where “vernal pool” soil types are not mapped, but winter pools with typical endemic annuals were found.

The historical abundance and distribution of these wetland types is extremely difficult to quantify in terms of area. Quantitative estimates of historic abundance of seasonal wetlands displaced by urbanization depend heavily on interpretation and assumptions about early soil surveys (which were not intended to function as maps of actual or potential native vegetation), historical accounts, and fragmentary information on species occurrences in old floras. The qualitative differences in natural non-tidal wetland types and their diked Bayland analogues further obscures the relevance of quantitative comparisons between historic losses of natural seasonal wetland plant communities and their partial replacement with wetlands of diked Baylands.

Conservation Issues

Plant conservation needs for diked wetlands are dependent on larger-scale wetland management and restoration plans. Diked wetlands usually support less native plant species diversity than mature tidal marshes at equivalent locations, but may in some cases still provide important plant conservation functions. For example, in San Pablo Bay, agriculture and development have eliminated most historic natural seasonal wetlands in supratidal grasslands peripheral to the Bay. The original vernal pool flora which occurred in subsaline to alkaline depressions around the historic edge of the Bay (as in parts of northeastern Suisun Marsh today) has been largely extirpated in its original location, but persists in artificial equivalent topography and edaphic conditions in some diked seasonal wetlands. These populations maintained in subsaline conditions may provide important founder populations for opportunities to restore vernal pool and swale systems in the original soil types and topography along the margins of the Bay, in coordination with tidal restoration. Similarly, one diked salt marsh in the South Bay (former Fremont Airport) provides refugia for Suaeda moquinii, otherwise found around the Bay only in remnant alkali vernal pools ad-
adjacent to the Bay at one site (Zentner and Zentner 1996). Partial vernal pool floras have also been generated spontaneously after cessation or relaxation of agricultural manipulation at Montezuma Wetlands (Solano County), Sears Point (Sonoma County), and a construction site in Alviso (Santa Clara County). Most diked wetlands are poorly surveyed, and may act as refugia for many populations of plants of conservation significance.

Diked wetlands are also conservation threats to plant species diversity when they provide outposts, reservoirs, or dispersal corridors for invasive wetland weeds, such as Lepidium latifolium and Salsola soda. By increasing seed rain pressures on adjacent tidal marshes, or adjacent marsh restoration sites, diked wetlands may also cause degradation of tidal marshes.

Sea level rise makes long-term conservation of diked wetlands problematic. In addition to inherent tendencies of diked systems to suffer levee subsidence and erosion, sea level rise imposes increasing risks of levee failure and tidal flooding. Breached diked wetlands spontaneously revert to tidal wetlands, but usually only as low mudflat or marsh to lower middle marsh after even two decades (e.g., White Slough, Vallejo, Solano County). In addition, some high-sulfur diked marsh soils undergo long-term changes in soil chemistry which make them unsustainable for any valuable natural or artificial vegetation.

Dike maintenance and repair may cause degradation to diked and tidal marsh plant communities by favoring spread and dominance of exotic invasive marsh plant species. Dike maintenance practices currently lack any elements which facilitate recolonization by native species.

Restoration of diked marshes is somewhat self-contradictory, since true restoration would entail conversion to the original tidal marsh condition. However, diked wetlands can be significantly enhanced as non-tidal marshes by reducing or eliminating adverse land use practices. Reduction of intensive drainage efforts and elimination of high-frequency diskng can enable diked fresh/subsaline wetland plant communities to mature and accumulate greater native species. Pasture management that tolerates some winter inundation in depressions, for example, is more compatible with native wetland plant species diversity than oat crop management.

Conclusions and Recommendations

Diked wetlands considered for conversion to other marsh types, such as tidal wetlands, should be studied individually for site-specific floristic values, particularly for potential functions as refugia for species displaced from historic seasonal wetlands and tidal marsh ecotones. Diked wetlands should not be assumed to have uniformly low native wetland plant species diversity or “ruderal” status. In areas where restoration of seasonal fresh wetland systems (e.g., vernal pools, alkali basins, alluvial Juncus/Scirpus marsh, etc.) is precluded by development, some diked wetlands should be considered for modification and management to maintain regionally scarce plant communities. Generally, however, priority should be assigned to restore peripheral estuarine plant communities in their proper original soils and topographic position. Where diked wetlands support regionally rare plant populations, they should be given interim conservation priority until suitable population restoration sites are established in more natural or restored habitats. Existing diked marshes should be managed to minimize impacts of exotic invasive plants on adjacent managed or natural tidal marshes. Dike maintenance should include best management practices which favor recolonization of disturbed dike surfaces by native vegetation and suppress re-invasion by exotic species.

References


BCDC (San Francisco Bay Conservation and Development Commission) and H.T. Harvey. 1983. Aquatic and wildlife resources of Richardson Bay. Unpublished report prepared for the Richardson Bay Special Area Plan Study.


Plants of San Francisco Bay
Salt Ponds
Peter R. Baye

Introduction

The term “salt pond,” as treated in this discussion, includes both natural and artificial large-scale persistent hypersaline ponds that are intermittently flooded by Bay water, and which occur within tidal salt marsh systems of San Francisco Bay and San Pablo Bay. Historic natural salt ponds were characterized by persistent thick accumulation of salt inundated with concentrated seawater brines. They were restricted to a relatively narrow reach of San Francisco Bay near San Lorenzo Creek. They are distinguished here from related salt marsh features such as pans and which occur at smaller spatial scales, have distinctive physiographic traits, and lack strong persistent (perennial) brines and precipitated crystalline salt deposits. Artificial salt ponds (solar salterns) are diked salt marshes which are managed for the production of concentrated brine and fractional crystallization of sea salts. Natural and artificial salt ponds are presumed to share the same narrowly adapted hypersaline biota.

Information on modern artificially engineered salt pond systems is derived principally from the biological literature on solar salterns and hypersaline environments (Javor 1989, and references within), historic documentation on the salt industry in California from the State Division of Mines (Ver Planck 1958, 1951; Dobkin and Anderson 1994) and regional documentation produced by the local salt industry and government regulatory agencies (Corps of Engineers, San Francisco District, Regulatory Branch permit and compliance files; Office of Counsel files, and references within). Information on historic salt pond systems is limited to descriptive historic accounts and descriptions, detailed topographic maps of natural salt ponds prior to extensive dike construction (U.S. Coast Survey T-charts, 1956), and field investigations by the author comparing modern salt pans, marsh ponds, and artificial salt ponds.

Environmental Setting

Salt ponds are large, shallow, hypersaline impoundments or depressions in tidal salt marsh systems which undergo a sequence of infrequent flooding with saline or brackish Bay water, evaporative concentration, and formation of strong hypersaline brines and deposits of gypsum, calcium carbonate, and crystalline salt (halite; sodium chloride).

Historic salt ponds were mapped with a high degree of resolution in the 1856 U.S. Coast Survey. They were nested within particular portions of the salt marshes along the Alameda shoreline in the vicinity of San Lorenzo Creek and Mount Eden Slough. This reach of salt marsh was distinguished by a relatively straight-edge erosional marsh shoreline, little tidal drainage at the edge of the mudflats, and evidence of drowned marsh topography (mapped as emergent sinuous tidal creek levees). The upland edge was an extensive alluvial lowland, presumably with significant subsurface groundwater discharge. No major freshwater creeks were directly associated with the salt ponds. Atwater et al. (1979) suggested that natural estuarine beach ridges along outer marsh edge were responsible for the impoundments of salt marsh that created salt ponds near San Lorenzo. Some salt ponds at the northern end of the local San Lorenzo distribution were certainly associated with well-defined barrier sand spits (U.S. Coast Survey T-charts, 1850s), which were probably nourished by sand eroded from submerged Merritt sand deposits (Pleistocene marine beach and dune). Less well-defined transgressive berms of sand and coarse organic detritus may have been deposited on top of the erosional marsh edge south of the sand spits themselves. Similar transgressive beach-marsh berms today act as dams enclosing freshwater to brackish ponds and marshes in Drake’s Estero, Point Reyes and at one location in San Francisco Bay (Whittell Marsh, Point Pinole, Contra Costa County). U.S. Coast Survey T-charts also indicate numerous sandy barrier beaches which dammed (either permanently or intermittently) lagoons. The impoundment of Crystal Salt Pond by a wave-constructed swash bar or beach ridge would distinguish it morphologically, hydrologically, and topographically from more common salt marsh ponds (pans) which occurred as depressions, sometimes extensive, between tidal creeks. These were widely distributed in salt marshes in the South Bay. Extensive, elongate pans also occurred near and below the upland borders of salt marshes; these have been termed “transitional” pans, although their position and form do not necessarily indicate a gradual ecotonal relationship with alluvial or upland habitats.

Salt ponds today (solar salterns) are artificially managed and engineered diked Baylands converted from tidal salt marsh. The first artificial salt ponds began as extensions and improvements of natural salt ponds which occurred near Hayward (Crystal Salt Pond), but most of the contemporary man-made salt pond system is established in former tidal marsh that included few or no perennial hypersaline ponds. Artificial salt ponds have entirely displaced their natural forerunners; no natural true salt-crystallizing ponds remain in San Francisco Bay today, although related smaller salt pans and marsh ponds containing weak brines in summer and fall do occur.

Classification of Salt Ponds - Javor (1989) placed marine-derived hypersaline aquatic environments in four ecological salinity classes:
Plants

The first salinity class (ca. 60 - 100 ppt) contains a highly diverse, productive biota dominated by marine species. This class would correspond to “low salinity” ponds (a misnomer, since salinity exceeds seawater concentration), from intake ponds to the next one or two stages that support abundant macroalgae and fish.

The second class (ca. 100 - 140 ppt) is dominated by specially adapted halophilic species which are related to freshwater taxa, not marine taxa. The organisms include abundant cyanobacteria, unicellular green algae, brine shrimp, and various halobacteria.

The third class (ca. 140 - 300 ppt) is distinguished by marked reduction of species diversity (loss of cyanobacteria, most invertebrates other than brine shrimp), and dominance of Dunaliella and brine shrimp.

The fourth class (300 ppt to salt saturation, near 360 ppt) contains only Dunaliella and bacteria at low productivity.

The first class predominates in modern marsh ponds. The historic natural salt pond complex probably varied seasonally between Javor’s second to fourth hypersaline classes. Other natural marsh ponds were most likely predominantly in the first class only, becoming seasonally hypersaline, and supporting relatively weak brines and macroalgal cover. Natural historic salt ponds were distinguished from other types of inundated depressions in salt marshes by the persistent thick halite deposits, indicating perennial hypersaline conditions, and their large lake-like size. In these aspects, they differ from shallow marsh ponds and marsh pans, which are regularly flooded during higher spring tides, and either remain persistently ponded or develop thick algal mats which desiccate in summer (bleaching white in the sun, resembling salt deposits in aerial photographs), or only develop thin, temporary salt films on unvegetated mud and peat.

Various marsh pan features are represented in U.S. Coast Survey maps of the mid-19th century, but only a few have persisted in modern rare remnant tidal marshes, such as Petaluma Marsh, Rush Ranch and Hill Slough (Solano County). Elongate marsh ponds are evident along the upland edge of historic marshes, particularly in eastern and southern parts of San Francisco Bay. Some of these may have been influenced by surface runoff and groundwater seepage from adjacent alluvial uplands, and could have been less saline than other marsh depressions most of the year. Some historic elongate marsh edge ponds may also have been the unvegetated upper intertidal surface of alluvial fans and terraces, consistent with small modern “transitional pans” observed at Hill Slough, Solano County. These also lack brine and halite development. Modern elongate marsh ponds have formed in recently (100 year) prograded marshes adjacent to M are island dredge ponds. These ponds are about 0.3 m deep in winter and spring, and range from brackish (nearly fresh) in winter to hypersaline when ponded areas are highly reduced in summer, but no significant halite precipitation is evident in them. These and similar pans may appear white with sun-bleached dried algal mats, which resemble salt flats. High densities of true natural marsh ponds, also termed “drainage divide ponds” (owing to their position in poorly drained marsh areas between tidal creeks), also occur in the Petaluma M arsh. Marsh ponds are a variation of salt pans which are topographic depressions flooded by spring tides, and support submergent vegetation, typically macroalgae (such as Enteromorpha spp.) and beds of widgeon-grass (Ruppia maritima), indicating brackish to near-marine salinity. The beds of marsh ponds are usually a soft organic oil-like black muck composed of decayed, waterlogged organic matter.

In contrast with salt ponds in estuaries with strong marine influence, such as San Diego Bay, San Francisco Bay salt ponds are relatively nutrient-rich and sustain high primary productivity (Javor 1989). Nutrient-poor salt pond conditions promote microbial mats, while planktonic microalgae tend to dominate nutrient-rich salt pond systems (Javor 1989). Most salt ponds in San Francisco Bay support richly pigmented and somewhat turbid organic “soups” of Dunaliella, halobacteria, cyanobacteria, dissolved organics and organic particulates and, often in ponds between approximately 120 - 200 ppt salinity, large “blooms” of brine shrimp which graze primarily on Dunaliella.

Historic natural salt ponds were unlike modern artificial salt ponds in that they were not differentiated geographically into stable hypersaline classes, but varied only seasonally in salinity. Natural salt ponds went through a seasonal “intake” phase during extreme high spring tides (December-January and June-July), when Bay water flooded them and diluted them with brackish to saline Bay water, seldom exceeding 20 ppt, and typically between 2 - 10 ppt in winter. During summer-fall evaporation periods, brines formed in situ, ranging in salinity over time up to crystallization (saturation) near 360 ppt. In contrast, the modern engineered salt pond system is based on timed transfers of brines between ponds, resulting in spatial separation of brines at different stages of concentration, and fractional crystallization of various seawater salts (other than sodium chloride, halite), such as magnesium and potassium salts (bitterns), gypsum (calcium sulfate) and lime (calcium chloride) in different ponds. In this system, crystallization is restricted to relatively few ponds engineered to facilitate harvest of halite deposits, and relatively stable hypersalinity regimes are established for individual evaporator ponds in the system (Ver Planck 1958).

The sequential and spatial separation of brines in artificial salt pond systems also produces salt pond “types” which are not fully analogous to natural systems. The late stages of brine production near sodium chloride crystallization produce strong non-sodium brines called “bittern.” Bittern brines (or bittern) are a concen-
trated solution of sodium chloride, magnesium chloride and sulfate, and potassium chloride and sulfate. The ionic balance of highly concentrated bittern is toxic even to bacteria, and saturated bittern is considered sterile (Javor 1989). During winter rains, dilute bittern stratifies on top of the concentrated bittern, and brine shrimp may appear seasonally, indicating algal production (Jim Swanson, Rick Coleman, pers. comm.). Natural salt pond brines did include bittern salts; in fact, the "low quality" of early California solar salt was due to bittern, and the modern solar saltern system is principally devised as a method to fractionate sodium and bittern salts. Crystallizer ponds, which are used to precipitate halite, are also maintained near the limits of halotolerance of Dunaliella (which can nonetheless fix carbon up to salt saturation; Javor 1989), but undergo seasonal dilution during winter rains.

**Salt Pond Plant Community**

Salt ponds support a distinctive and highly specialized halotolerant to halophilic biota consisting of microalgae, photosynthetic bacteria, and invertebrates, but no vascular plants (except along the edges of natural salt pond levees). The dominant photosynthetic organisms of most hypersaline San Francisco Bay salt ponds are a single-celled green algal species, Dunaliella salina (Chlorophyta) and numerous species of blue-green bacteria (Cyanobacteria), halobacteria, and purple sulfur-reducing bacteria. The proportions of these organisms vary with salinity. Artificial eutrophic salt ponds with salinities closer to marine concentrations (near 35 ppt; "intake ponds") are dominated by marine macroalgae such as sea-lettuce (Ulva spp.,), Enteromorpha spp., Cladophora spp., and also sometimes support Fucus spp. and Codium spp., where substrate is stable and firm. They also include marine diatoms, dinoflagellates, and cryptomonads. There are no detailed studies of the species diversity, distribution or geographic variation of the halophilic microflora communities of San Francisco Bay.

Managed and engineered contemporary salt ponds are ecologically similar in many respects to their natural precursor salt ponds, and presumably share the same algal and bacterial microflora.

**Indicator Species** - There are no detailed classifications or analytic studies of salt pond algal communities. Following Javor's (1989) classification of hypersaline environments (see Classification of Salt Ponds, above), two broad hypersaline algal communities may be identified: communities dominated by free-floating marine macroalgae typical of upper tidepools near marine salinities to low-hypersaline conditions, corresponding to intake ponds and young brines in a saltern series (e.g., Ulva spp., Enteromorpha spp., Cladophora spp.; also bottom-mat forming cyanobacterial colonies); and communities dominated by motile unicellular halophilic phytoplankton (particularly Dunaliella salina), which characterize moderate to high hypersaline conditions. Macroalgae salt pond communities also correspond with fish-dominated animal communities, while phytoplankton-dominated brines are associated with brine shrimp abundance.

Dunaliella spp. is ubiquitous in salt ponds in San Francisco Bay. It is reported to survive, and can be photosynthetically active, in brines which are close to saturated (near 350 ppt), but may be absent in some extremely concentrated brines and bittern (potash-phase, or potassium-magnesium) brines (Javor 1989, Brock 1975). Its optimum salinity for growth is near 120 ppt, about four times the concentration of seawater. Dunaliella salina concentrates carotenoid and other pigments in response to various forms of physiological stress, including salinity. It can be used as a crude color-indicator of brine salinity: cells growing in 50-100 ppt are greenish, and turn yellowish-green in 150 ppt brine. Reddish hues occur in brines 200-250 ppt (Javor 1989). Purplish-red hues in brines over 200 ppt may be contributed by halophilic bacteria. A conspicuous mosaic of salt pond hues are readily visible from aerial views of San Francisco Bay, particularly in summer and fall. Dunaliella osmoregulates in hypersaline brines by concentrating glycerol as a compatible osmotic solute in its cytoplasm (Javor 1989).

**Reference sites**

There are currently no reference sites in the San Francisco Bay Estuary for true natural salt ponds (ponds which periodically or chronically produce crystalline salt deposits). The historic salt pond system near San Lorenzo Creek in Alameda was eliminated by diking in the 1850s and 1860s. All modern salt pans and marsh ponds in the Bay Area differ from these historic salt ponds. Most existing marsh ponds are only slightly hypersaline, or briefly hypersaline in late summer, and support algal mats rather than brines and halite beds. Most
existing salt pans within small modern Bay Area salt marshes are comparatively small and produce sparse and thin (few mm) salt crusts in summer and fall. In contrast, reference sites for artificial salt ponds are abundant. Examples of (relatively) low salinity intake ponds, which are saline or slightly hypersaline, are found at Pond B1/ B2 in Mountain View, Pond 1 near Mountain Slough, and Pond A9 in Alviso. Examples of intermediate hypersaline ponds (known as concentrators or evaporators) are found in ponds A10-14 in Alviso, ponds 2-8 near Coyote Hills, and ponds 2-6 between Mountain Slough and Coyote Creek. High hypersaline ponds (strong brines) approaching or reaching salt saturation, “pickle)” are found in extensive crystallizer beds near Newark and Redwood City, ponds 10 and 26 near Newark, and periodically in drained evaporators before they are re-filled.

Modern salt marsh (and brackish marsh) pans may be found in few remnant pre-historic tidal marshes at Petaluma Marsh (abundant), China Camp (scarce) and Point Pinole (Whittell Marsh; scarce). Pans vary in topography. Some upper marsh pans are similar to patches of salt flats, while pans in middle marsh zone depressions are normally shallow ponds 10-20 cm deep. Pans which become ponded, either because of depressional topography or marsh surface drainage barriers, develop algae or widgeon-grass. Salt marsh pans also occur in historically accreted marshes at Mountain Slough. Elongate marsh pans fringing uplands (“transitional” pans) have also formed in the relatively young (20th century) salt marsh at Emeryville Crescent and adjacent to Island dredge ponds. Elongate but diffuse shore-parallel marsh pans, perhaps best regarded as incipient pans, are found along the east end of the fringing salt marsh at Highway 37. Small but well-differentiated semi-circular to semi-linear salt marsh pans occur in peaty coastal salt marshes at Limantour Spit, Point Reyes; Bolinas Lagoon; Morro Bay; Elkhorn Slough; and along Tomales Bay. Morro Bay, Bodega Bay, and Bolinas Lagoon also have elongate shallow marsh salt pans fringing alluvial deposits. Most of these salt marsh pans are brackish in winter and spring, but become moderately hypersaline (usually 40-60 ppt, rarely > 90 ppt) in summer (Baye, unpub. data) when inundated.

**Historic and Modern Distribution**

The historic (pre-1860) location of natural salt ponds within San Francisco Bay was probably restricted to the Alameda shoreline in the vicinity of San Lorenzo Creek (between the historic Thompson’s Landing and Union City Creek). This area included an extensive complex of both connected and isolated large ponds in a matrix of salt marsh. The complex was labelled as “Crystal Salt Pond” on the 1856 U.S. Coast Survey T-chart of the area. The San Francisco Estuary Institute estimates the acreage of Crystal Salt Pond to be approximately 1660 acres, based on the precise pond outline represented on the 1856 T-chart (R. Grossinger, personal communication). If, however, the pond size fluctuated seasonally (as expected from winter rainfall and tidal flooding), the ponded area may have been several thousand acres from late fall to spring. Two smaller ponds with similar configuration occurred north of San Lorenzo Creek, and were clearly associated with sandy barrier beach deposits at the bayward edge of the marsh. (It is not clear whether these northern satellite ponds produced high concentration brine and halite, or were merely intermittently hypersaline lagoons). Crystal salt pond was used as a salt source by aboriginal inhabitants of the Alameda shoreline, and was exploited by early Mexican, Spanish and U.S. settlers (Ver Planck 1951, 1958). Early descriptions of Crystal Salt Pond indicate that it contained a persistent crust of crystalline salt up to eight inches thick, and the brines and salt contained “impurities” of concentrated non-sodium salts (“bitter” salts, principally magnesium chloride and sulfate; Ver Planck 1958).

The natural halite deposits of Crystal Salt Pond were exhausted rapidly by the infant salt collecting industry; by 1860 they were largely depleted. Artificial enhancement of solar evaporation of brines was initiated around 1853, when salt harvesters (farmers who used salt for tanning leather and curing meats, and expanded into the salt industry) began manual construction of low berms around natural salt ponds to enhance their capacity to retain saline floodwaters and capture and precipitate their salt loads. These artificially enhanced natural salt ponds became the nucleus of the solar salt industry.

By the end of the 19th century, the salt ponds of San Francisco Bay were still confined to the northern portion of the Alameda shoreline, from San Leandro Creek to Alvarado (Union City). They did not comprise a salt pond “system,” but were an aggregation of many independently owned and operated enterprises. Extensive conversion of salt marsh to salt ponds in south San Francisco Bay did not occur until the 20th century. This was facilitated by the consolidation of almost all the independent salt operations to a few (dominated by Leslie Salt Company) in the 1930s. Permit requests to the Corps of Engineers to dam numerous sloughs and marshes in the South Bay were not filed until the early 1920s. Actual levee construction would have taken at least several years, and new ponds take about 5 - 7 years to “seal” (become impermeable after gypsum and carbonate precipitation; Ver Planck 1958, Dobkin and Anderson 1994); therefore, the 1920s ponds were probably not fully functional salterns until around 1930. The last extensive marshes in the Alviso and Sunnyvale areas were not diked for conversion to salt ponds until the early 1950s (Pacific Aerial Photo archives). Bair Island was not converted to salt pond until the 1950s, although it had previously been diked for agricultural use. The modern
extent of salt ponds in the southern reaches of South Bay, therefore, is relatively recent compared with the northern Alameda salt ponds. The Napa salt ponds are even more recent: the diked Baylands of the Napa marshes were converted from derelict agriculture (seasonal subsaline to brackish wetlands) to salt ponds between 1953 and 1959. Salt production ceased there in the mid-1990s, but most of the system remains hypersaline.

Relative change - The minimum acreage of true natural salt pond in San Francisco Bay was less than 2,000 acres (SFEI 1998); the maximum acreage (assuming seasonal expansion of Crystal Salt Pond by flooding, and assuming that northern satellite ponds were brine/halite ponds) could have been on the order of 3,000 - 4,000 acres. Other marsh pan habitats were not likely to support persistent hypersaline algal communities and were presumably dominated by brine/macroalgae or Ruppia, as are most salt marsh pans today. However, if a significant proportion of the historic extensive elongate lake-size marsh ponds fringing uplands (Redwood City to Palo Alto, and in the Newark vicinity) were seasonally or perennially hypersaline, the total acreage of salt pond habitat could have been on the order of 5,000 - 10,000 acres. There is very weak indication that elongate upland-fringing salt marsh pans ever contained persistent strong brines supporting the narrow hypersaline algal/bacterial community, however. Today, approximately 9,500 acres of derelict salt ponds remain in San Pablo Bay, and over 29,000 acres of artificial salt pond are actively maintained in San Francisco Bay.

Conservation Issues

Exotic Species - Salt pond microbial taxa are widely spread geographically, but narrowly distributed ecologically. They are probably subject to dispersal by waterfowl and marine transport. There are no currently recognized exotic species "threats" to salt ponds as there are with vascular plants in salt marshes.

Restoration - The crude technology for creating artificial salt ponds (levée construction, wind-driven pumps, tidegates) has been well developed for over a century. There is little doubt that complete artificial salt pond systems can be created and maintained at a wide range of sizes, from as little as 20-50 acre historic "family size" or one-man operations (Ver Planck 1958), to the modern systems in the tens of thousands of acres. Low-salinity "intake" ponds can also be maintained independently, in the absence of a salt-producing system, by balancing influx of Bay water, residence time and redischarge at near-marine salinity. No new salt ponds have been constructed since the 1950s, although ponds have been interconverted from one type to another since then (evaporator ponds to bittern disposal/"storage"). Small and autonomous salt pond systems could be modified to be less "productive" of salt, and more biologically "productive," by reducing the efficiency of brine and salt production. This could be achieved by increasing the flux in intake ponds, and reducing the residence time of brines in each pond transfer. In winter, when brines are diluted by rainwater, they could also be re-mixed with intake Bay water and redischarged to the Bay at near-marine salinities.

There have been recent tidal marsh restoration designs for artificial but naturalistic ponds and pans, but no marsh restoration designs have included equivalents of salt ponds. In principle, naturalistic salt ponds could be artificially created and naturally maintained by replicating the hypothetical historic conditions of Crystal Salt Pond (as inferred by Atwater 1979). This would entail deposition of coarse sediments (sand or shell hash) at the edge of a high-energy marsh shoreline, to be reworked as beach ridges which restrict marsh drainage. In theory, beach ridges would maintain form and size as they retreat with the eroding marsh edge, given ample sediment supply and overwash processes. Under less natural geomorphic settings for salt ponds, artificial naturalistic salt ponds could be created by constructing low, broad berms made of bay mud or sand that would be set at elevations enabling highest spring tides to overtop them. Low, wide berms would be less prone to gullying and breaching than steep levees, but would require some degree of maintenance. Maintenance would be minimized by setting salt pond levees within restored marshes which would shelter them from wave erosion of the open Bay. Restored naturalistic salt ponds would undergo extreme variation of salinity within and between years, depending on rainfall variation, evaporation conditions, and storm surges.

Sea Level Change and Levee Maintenance - The modern salt pond levee system requires periodic maintenance, and levees bordering the open Bay (not sheltered by fringing salt marsh) require frequent maintenance, armoring, or both. The need for levee maintenance (topping with fresh dredged sediment) is likely to become more frequent if storm frequency increases or sea level rises, as would be expected with global warming (Moffatt and Nichol and WRA 1988). Borrow pits along the interior side of salt pond levees become depleted over time, and some old borrow ditches have been widened so much that dredges need to re-handle material to bring it within reach of levees. Dredging tidal marshes as an alternative source of sediment is unlikely, since it causes conflicts with endangered species habitat. Therefore, sea level rise is likely to cause long-term increases in costs and risk of levee failure of the existing salt pond system. Sea level rise could also make naturalistic salt pond restoration more difficult, since beach ridges or low levees are more likely to breach and allow excessive (though restricted) tidal exchange to impounded areas.
Conclusions and Recommendations

The commercial salt pond operations of San Francisco Bay are unlikely to continue indefinitely because of economic changes in the Bay region and in the salt industry, and due to physical changes in the levee and borrow ditch system. Salt ponds are not likely to regenerate spontaneously as a result of natural geomorphic processes when salt marshes are restored. Other common types of pans and ponds are unlikely to establish in young salt marshes; they are mature marsh features, associated with well-differentiated marsh topography. The environmental setting associated with salt ponds has been radically altered; the combination of steep and relatively high-energy Bay shorelines, coarse sediment supply, and extensive high salt marsh with impeded tidal drainage no longer exists. It is also likely that the feasibility of maintaining the erosion-prone levee system of the artificial salt ponds will decrease over time, as borrow ditches (sources of mud for levee repair) are depleted. Therefore, new and artificial measures will be required to conserve at least historic amounts of salt pond habitats within the Estuary in the long term. The highest priority setting for salt pond restoration of some type would be on the Alameda County shoreline, from approximately San Leandro to the Dumbarton Bridge, where the Bay shoreline profile and wave fetch may be conducive for formation of beach ridges (marsh berms), given appropriate sediment size and supply.

There is no minimal ecosystem size for salt ponds. The basic grazer food chain between Dunaliella and Artemia can be maintained in extremely small systems. However, the full microbial diversity of San Francisco Bay salt ponds, which has not been analyzed in detail, would probably not persist in small ponds. Also, since the stability of natural salt ponds is inherently low (subject to ordinary natural fluctuations as well as catastrophic changes), microbial diversity would be better conserved with a large system of semi-independent salt ponds. Pre-historic salt pond acreage was probably on the order of 2,000 acres. Aiming at this minimal acreage, in the absence of any experience at restoration of naturalistic salt ponds or “alternative” management of solar salterns, would probably be insufficient to conserve a diverse halophilic microflora.

We therefore recommend that long-term conservation of salt ponds entail the following actions:

1. Pilot projects should be undertaken that incorporate naturalistic salt pond designs as integral components of large-scale tidal marsh restoration on the northern Alameda shoreline;
2. Some existing salt ponds should be divided into smaller, autonomous units away from the open bay, preferably nested in the landward reaches of restored salt marsh areas, and managed to maintain intermediate strength brines rather than salt production;
3. Salt pond restoration and alternative management should aim for temporally variable as well as spatially variable salinity and brine depths;
4. Both artificial and naturalistic salt pond restoration should aim for designs which minimize maintenance requirements; and
5. An initial target acreage for salt ponds should reflect the uncertainty of restoring sustainable salt pond environments after commercial salt production ceases. We suggest that an initial target of approximately 10,000 acres (equivalent to late 19th century acreage) be stipulated and modified based on the results of salt pond restoration and alternative pond management.

References


Plant Communities Ecotonal to the Baylands

Glen Holstein

Introduction

The San Francisco Bay estuary wetlands ecosystem historically included vegetated and non-vegetated areas. Dominant among physical factors influencing estuarine vegetation was the semi-diurnal tidal cycle. As a consequence, vegetation exposed to tides differed dramatically from plant communities that existed above the tides. For non-estuarine vegetation diurnal factors were relatively insignificant; annual climate cycles and non-cyclic geological factors were the dominant influences. Substrates in vegetated parts of the Estuary consisted almost entirely of Bay mud (Louderback 1951, Wahrhaftig et al. 1993). Beyond it they were much more heterogenous.

Environmental Setting

Vegetation increases in structural diversity and species richness beyond the estuarine ecosystem boundary in a complex pattern caused by interactions between the physical factors of climate, geology, and hydrology.

Climate – The San Francisco Bay Area, like all the California Floristic Province (Hickman 1993), has a climate characterized by wet winters and dry summers. Such climates are called "Mediterranean" because similar climatic conditions occur in the Mediterranean Basin, but the San Francisco Bay Area's Mediterranean climate is more extreme than much of its namesake since it rarely receives any significant rainfall during the year's warmest five months (Wernstedt 1972). Despite ample water, plant growth is retarded in winter by low temperatures and day length significantly increase and reserves of soil water from winter rains are still abundant (Walter 1979).

The diversity of the San Francisco Bay Area climate is explained, to a great extent, by variation in two factors: winter precipitation and summer marine air flow. Both cause local climates to be relatively mesic, but their maxima rarely coincide and do not identically affect vegetation. High winter precipitation makes abundant soil moisture reserves available for rapid spring and early summer plant growth where low temperatures and fog brought by marine air flow do not limit it. Since rapid plant growth increases biomass, high biomass vegetation types like redwood and mixed evergreen forests are frequently dominant in the Bay Area where rainfall is highest. A popular myth contends redwoods (Sequoia sempervirens) require summer fog. What they actually require (and are limited to) are places with high precipitation that are protected from summer marine air flow and fog. By leaching mineral nutrients from surface soils, high rainfall also retards growth of herbaceous vegetation that could otherwise compete with forest tree seedlings (Holstein 1984a).

Bay Area mean annual precipitation varies from 13 inches at San Jose and Antioch to 47 inches at Kentfield in Marin County (Felton 1965). Not surprisingly, relatively undisturbed upland vegetation consists of redwood and mixed evergreen forests near Kentfield (Shuford 1993) and of grassland near Antioch and San Jose (Critchfield 1971). The Bay Area receives its precipitation from cyclonic storms with predominantly southwest winds arriving from the Pacific Ocean. Consequently stations with large mountains to the southwest receive much rainfall and marine air flow and fog mitigate summer drought sufficiently for occurrence of vegetation types like coastal scrub and prairie characterized by very complete cover and little bare ground despite relatively low biomass (Holstein 1984a).

Summer water stress causes incomplete cover and much bare ground in most Mediterranean climate vegetation. In parts of the San Francisco Bay Area, however, marine air flow and fog mitigate summer drought sufficiently for occurrence of vegetation types like coastal scrub and prairie characterized by very complete cover and little bare ground despite relatively low biomass (Holstein 1984a).

Summer marine air flow and fog arrive at the Pacific Coast predominantly from the northwest because of anticyclonic origins, but a shallow semi-permanent temperature inversion confines their movement into and through the San Francisco Bay estuary to just a few low altitude gaps in the Coast Range. By far, the most important of these is the Golden Gate (Gilliam 1962).

Since marine air flow and fog suppress summer temperatures, mean July temperature is a reliable indicator of their relative presence or absence in the San Francisco Bay Area. Not surprisingly, San Francisco's July mean of 58.8° is the lowest around the Estuary because of its location at the Golden Gate. Antioch's July...
mean of 74.0°F is the highest of any Estuary station since the low hills of northern Contra Costa County protect it from marine air flowing into the Central Valley through the Carquinez Strait. Fairfield's July mean of 72.1°F indicates more direct exposure to that air flow despite its more inland location. Mount Diablo State Park's entrance station has a July mean of 74.3°F because of its location above the inversion that limits marine air to low elevations. Most Estuary stations have July means in the sixties, but a difference of just a few degrees within that range can profoundly effect summer climate. Berkeley (61.5°F), Richmond (62.0°F), and Oakland (62.4°F) have the lowest summer temperatures in the Estuary next to San Francisco because of their location directly east of the Golden Gate. Burlingame (62.3°F) and the San Francisco Airport (62.7°F) are also relatively low because of their location at the east end of the San Bruno coast range gap (Gilliam 1962). Kentfield (65.9°F), in contrast, is relatively warm in July because Mount Tamalpais protects it from summer fog as well as inducing its high winter rainfall. Distance from the Pacific Coast is generally a poor predictor of summer marine air flow. Redwood City (67.9°F) on the west side of the Bay, for example, is warmer than Newark (64.9°F) on the east side since the latter is more directly exposed to air flow through the Golden Gate (Felton 1965, Gilliam 1962). Coastal scrub and coastal prairie, the vegetation types most associated with summer fog, are common on the outer Pacific Coast but relatively scarce in the San Francisco Bay Estuary because the parts of it most exposed to summer fog were also those settled earliest and urbanized most completely (Hover et al. 1966, Donley et al. 1979).

**Geology** - Holocene alluvium characterized by abundant clay and level topography surrounds slightly over half the Estuary and is consequently the most abundant geologic substrate beneath its adjacent non-tidal vegetation (Jennings 1977, Wahrhaftig et al. 1993). Bay Area uplands underlain by alluvium were farmed early and are now largely urbanized, but historic accounts and relic stands indicate open grassland was their overwhelmingly dominant vegetation type before settlement (Bryant 1848, Mckelvey 1955). An exception was a few oak savannas where widely spaced valley oaks (*Quercus lobata*) occurred in a grassland matrix. Such savannas were most frequent around the northern part of the Estuary where rainfall was relatively high, but even there they were most frequent in areas protected from summer marine air flow.

A specialized feature of California Holocene and older alluvium with level topography is vernal pools, small closed basins that fill in winter and dry during spring. They support a characteristic specialized flora rich in annual forbs (Holland and Jain 1977). Vernal pools were long thought to result from gopher activity (Dalquest and Schefter 1942), but are better explained as microtopographic patterns arising from ground shaking during earthquakes (Berg 1990) or interaction of localized soil processes and wind erosion (Abbott 1984).

Non-alluvial uplands around the Estuary consist of uplifted hills underlain by a variety of pre-Holocene sedimentary and volcanic rocks. These include the Mesozoic Franciscan formation and Great Valley Beds; Cenozoic sediments consisting of Paleocene, Eocene, Miocene, and Pliocene marine and non-marine deposits; and the Pliocene Sonoma volcanic deposits (Jennings 1977, Norris and Webb 1990). The influence of these rocks on vegetation is most frequently controlled by their clay content. Those with abundant clay like Paleocene, Eocene, Miocene, and Pliocene sediments weather to deep soils much like those on Holocene alluvium and predominantly support similar grassland vegetation. The Mesozoic deposits include areas where clay is abundant and others where it is scarce. As on other clay-rich substrates, deep soils and grasslands dominate the former in contrast to the thin soils and woody vegetation types predominant where clay is scarce. The Franciscan Formation, a melange of soft clay sediments and hard metamorphic rocks, has a particularly complex vegetation pattern since grass dominates the former and trees the latter. Pleistocene marine beds and the Sonoma volcanics are relatively clay poor and consequently largely support woody vegetation types like oak woodland and mixed evergreen forest (Ellen and Wentworth 1995, Critchfield 1971).

Grass is dominant on clay soils because they have a relatively high water holding capacity (Walter 1979). West of Cordelia in Solano County, for example, Dibble Los O sos and Hambright loams occur on adjacent hills in the same climate. Dibble-Los Osos soils develop on clay-rich Eocene marine sediments and consequently have B2t horizons containing accumulated clay and a water-holding capacity of 5 to 7 inches. Hambright soils, in contrast, develop on Sonoma volcanics, lack a B2t horizon, and have a water holding capacity of only 2 to 3.5 inches (Bates 1977). Despite identical precipitation, Dibble-Los Osos soils support grassland and Hambright soils support oak woodland dominated by coast live oak (*Quercus agrifolia*) because the former's B2t retains soil water that can be used by the shallow fibrous root systems of grasses. Since Hambright soils retain much less water, the excess infiltrates to the fractured rock below where it can be utilized by deep roots of trees but not grasses. In May, evidence of the Dibble-Los Osos B2t's water retention capacity is plain in the hills above Cordelia since grass stays green there several weeks longer on Dibble-Los Osos soils than it does on the Hambright despite the frequent shade of oaks. This phenomenon illustrates that two very different vegetation types can be equally "mesic" and that oak woodland and grassland are competitive enough within this region for slight soil differences to shift dominance from one to the other.
The geologic factor that most influences vegetation around the San Francisco Bay Estuary is the physical effect of clay on soil water holding capacity, but chemical effects are also locally important. Serpentinite, associated with the Franciscan Formation and occurring at the Estuary's edge in Marin and San Francisco counties, is so chemically distinctive because of its high Mg/Ca ratio and frequent heavy metals that it supports unique vegetation types and many endemic plant species (Kuckenberg 1984). Soils beyond the limits of tides are also usually much less saline than those under tidal influence, but salts can locally accumulate to high levels in non-tidal areas where drainage is poor. Salt especially accumulates in non-tidal areas where precipitation is low, relief is subdued, and Cretaceous Great Valley beds provide a salt source (Chapman 1960; Johnson et al. 1993; Harris 1991). Geology also strongly affects microclimate wherever hills have been uplifted since their south slopes receive more sunlight, warmth, and resultant evapotranspiration than their north slopes. Vegetation on Bay Area hills is consequently relatively xeric on south slopes and relatively mesic on north slopes (Bakker 1984).

**Hydrology** — The influence of geology and climate on soil water is discussed above. Streams also tend to increase in frequency and flow duration as rainfall increases. Since they provide water to plants in greater quantities and different seasons than local climates, they support distinctive riparian vegetation types not found in upland areas. Not surprisingly, riparian and upland vegetation become increasingly distinct as rainfall decreases (Holstein 1984b). Streams and their associated riparian vegetation are usually narrowly linear landscape features, but they can broaden dramatically when streams reach base level and form deltas. A broad willow-composite zone now removed by urbanization that reportedly once occurred around the southern edge of San Francisco Bay (Cooper 1926) undoubtedly represented covergent deltaic riparian vegetation of several creeks that flow into the Bay.

Alluvium in streambeds tends to be coarser and thus better aerated than interfluvial alluvium, and the running water of streams is also relatively well-aerated. Streamside vegetation consequently provide suitable environments for roots of woody riparian vegetation. In freshwater marshes, however, standing water in poorly drained interfluvial areas quickly causes anaerobic reducing conditions to develop at such shallow depths that only herbaceous vegetation with shallow, predominantly fibrous root systems can occur. The herbaceous freshwater marsh vegetation is consequently quite distinct from predominantly woody riparian vegetation (Holstein 1984a).

Freshwater marsh vegetation grades into vernal pool vegetation through a series of transitional seasonal marsh vegetation types and into moist grassland through a transitional series of lowland wet meadow and swale types. Numerous other local hydrological features around the San Francisco Bay Estuary like springs, seeps, and shallow water tables are associated with distinctive local vegetation types. The relatively shallow water table under most valley oak savannas is a notable example.

**Ecotonal Plant Communities**

Plant communities surrounding the Baylands ecosystem are here classified using the system of Holland and Keil (1995). At present the most widely used and influential classifications of California vegetation are derived from Munz and Keck's (1959) mixed system, which includes taxonomic, physiognomic, and ecological information. Barbour and Major's extensive (1977) review of California vegetation, for example, was organized around a slightly modified and expanded version of Munz and Keck's system. The units of their classification were vegetationally ill-defined, however, since the plant species lists provided for each one lacked even estimates of relative dominance. Some very important plant communities like riparian forests were also missing from both Munz and Keck's system and Barbour and Major's subsequent review. It is doubtful Munz and Keck intended their brief plant community synopsis to so profoundly influence California vegetation science, however, since the primary purpose of their book was clearly floristic. Its success at remaining California's floristic standard for decades undoubtedly strongly contributed to the influence of its community classification.

Sawyer and Keeler-Wolf (1995) have recently tried to overcome the Munz and Keck system's problems by developing a comprehensive alternative that excludes ecological information from community definitions except in the case of certain specialized habitats like vernal pools. The Sawyer and Keeler-Wolf system presents its own new problems, however, since it lumps quite different stable and successional communities when they are dominated by the same species. Excluding most ecological information also causes very different coastal and alpine communities sharing only a generic relationship between their dominant species to be lumped into catch-all groupings like “Sedge series.” Many local dominance types present in California's complex vegetation are also missing from their system despite its numerous series and apparent comprehensiveness.

Holland and Keil avoid these problems by greatly increasing the comprehensiveness and consistency of Munz and Keck's limited but fundamentally sound system. The result is a system outstanding for simplicity, ease of use, and realistic description and classification of California vegetation. Beginning with coastal sand dune vegetation and concluding with freshwater vegetation and anthropogenic environments, the plant community descriptions below follow the system developed by Holland and Keil.
1. Coastal Sand Dune Vegetation – Sand is a distinctive substrate for plants since water infiltrates it very rapidly leaving little moisture available for plants with shallow root systems (Walter 1979). Sand differs from other substrates like fractured rock which have similarly high infiltration rates, however, because of sand’s high subsurface homogeneity and lack of resistance to root penetration. Large sand deposits are characteristic landscape features of coasts and arid areas. In Holland and Keil’s (1995) system, followed here, vegetation on sand deposits of arid areas is classified as desert sand dune vegetation and consequently distinguished from vegetation on coastal sands. In California, however, some dune fields are located in areas neither coastal nor truly arid. Examples occur on the Merced River alluvial fan in Merced County and at Antioch in Contra Costa County (Wahrhaftig et al. 1993). The former was produced by outwash from glacial erosion of granite in the Yosemite Valley (Wahrhaftig and Birman 1965), but extensive Eocene to Pliocene sandstone deposits in nearby hills (Ellen and Wentworth 1995) are a likely source for the latter. Neither the Merced or Antioch dunes are discussed by Holland and Keil, but both occur in semi-arid areas and share more floristic features with their desert sand dune vegetation type than their coastal sand dune vegetation types.

Pioneer dune vegetation occurs where significant aeolian movement of sand limits development of stable soil and vegetation.

Ambrosia chamissonis is its characteristic dominant, and Abronia latifolia, Achillea millefolium, Atriplex californica, Atriplex leucophylla, Calystegia soldanella, Camissonia cheiranthifolia, Lathyrus littoralis, Leymus mollis, and Lupinus chamissonis are frequent associated species (Barbour and Johnson 1977).

Dune scrub occurs where stable soil and vegetation have developed on sand of dunes usually considerably older than those supporting pioneer dune vegetation. Artemisia ericoides is the characteristic dominant of dune scrub, and associated species include Artemisia californica, Baccharis pilularis, Lotus scoparius, Lupinus arborescens and Lupinus chamissonis (Barbour and Johnson 1977).

Sand is relatively rare around the San Francisco Bay estuary, but a significant deposit, the Pleistocene M erritt sand, is present at Alameda and adjacent parts of Oakland (Radbruch 1957). Since the local climate is marine, some areas with surface deposits of M erritt sand probably once supported pioneer dune and dune scrub communities similar to those now occurring along the outer Pacific Coast. The sandy area at Alameda and Oakland was one of the first places along the Bay to urbanize, however, and any dune vegetation present there was consequently eradicated before it could be described. A modern analogue with similar soils and climate is Elkhorn Slough (Monterey County), which is incised into Pleistocene deposits, the Aromas sand. Agricultural development has removed some natural vegetation around Elkhorn Slough, but remaining relict stands are still numerous. Topographic features recorded prior to development of the port of Oakland and Lake Merritt resemble those along Elkhorn Slough (Wahrhaftig and Birman 1965).

At Antioch, a sandy area is also present immediately east of Broad Slough. It is less urban than Alameda, but most of its dune vegetation was lost to sand mining prior to urbanization. A small protected remnant of such vegetation at Antioch supports several state and federally listed rare animal and plant species (Sawyer and Keeler-Wolf 1995). Antioch is significantly hotter and drier than the outer coast, and its sand probably originated from nearby sand deposits that extend southward along the inner Coast Range. The affinity of its distinctive sand dune flora and vegetation is consequently closer to Holland and Keil’s (1995) desert sand dune community than to either of his coastal dune communities. Because a rain shadow occurs along the inner Coast Range, the ranges of several plant and animal species with desert affinities, including the relatively well-known San Joaquin kit fox (Vulpes macrotis mutica) (Zimmer et al. 1990), extend north along the western San Joaquin Valley to near Antioch.

2. Coastal Scrub – Coastal scrub refers to communities dominated by small shrubs in non-desert areas of California. Coastal scrub typically develops on soil and friable sediments rather than conglomerate or fractured hard rock and consists of shrubs with relatively shallow root systems.

Northern coastal scrub is a dense shrub-dominated community which most frequently occurs on steep slopes receiving strong prevailing onshore winds and at least 20 inches of precipitation, but can also occur as an ecotone between northern oak woodland and southern oak woodland on slopes with less wind. Most typically, however, it occurs where precipitation and soils are adequate for development of forests, but tree growth is prevented by strong onshore winds. Since moisture is not limiting, cover is typically complete (Head et al. 1977).

Baccharis pilularis is the characteristic dominant, but Mimusops aurantiacus Rhamnus californica, and Toxicodendron diversilobum can also occasionally be locally dominant. Characteristic understory species include Achillea millefolium, Anaphalis margaritacea, Eriophyllum staechadifolium, Gaultheria shallon, Hardecia lanatum, Polygusnum munitum, Pteridium aquilinum, Rubus ursinus and Scrophularia californica. Northern coastal scrub is most common along the outer Pacific Coast but also occurs at suitable sites around the San Francisco Bay Estuary near the Golden Gate, in the Berkeley Hills, and in San Mateo County. Baccharis pilularis frequently invades disturbed grasslands and forms communities which superficially resemble northern coastal scrub but lack
most of its characteristic species. Eventually such recently invasive B. pilularis stands may develop into stable coastal scrub or oak woodland communities (Haday et al. 1977). A protected example of northern coastal scrub occurs near the estuary at China Camp State Park.

Southern coastal scrub is a relatively open shrub-dominated community occurring most frequently on steep, dry slopes. It is commonest in areas receiving under 20 inches of precipitation but can occasionally occur in wetter areas on sunny south slopes. It typically occurs where soils otherwise suitable for grassland are excessively drained because of steepness. Because water is the primary limiting factor in southern coastal scrub, its dominant shrubs tend to be widely spaced, forming relatively incomplete cover. In spring, when water stress is briefly relieved, a diverse annual forb flora develops in interstices between the dominant shrubs (Mooney 1977).

Artemisia californica is the characteristic dominant, and common associated species include Eriogonum nudum, Eriophyllum lanatum, Lotus scoparius, Lupinus albifrons, Mimulus aurantiacus and Nassella pulchra. Small stands of southern coastal sage scrub occur in hills around the Estuary and are especially frequent east of South San Francisco Bay and south of Suisun Bay, where precipitation is relatively low. An example occurs along the Estuary shore at Point Richmond.

Sea-bluff coastal scrub occurs where persistent salt-laden onshore winds suppress most other plant communities. Such climatic conditions resemble those in northern coastal scrub but are more extreme. In such sites the only communities are sea-bluff coastal scrub and northern coastal grassland. The former tends to occur on rocky sites with thin soils and the latter on deeper soils that tend to be heavier, but both frequently intermix in a complex mosaic (Holland and Keil 1995).

Eriophyllum stachadifolium is the characteristic dominant, and frequently associated species include Artemisia pyrocathapha, Baccharis pilularis, Erigeron glaucus, Eriogonum latifolium and Lessingia filaginifolia. Salt-laden winds strong and persistent enough to support this community enter the Estuary through the Golden Gate but rapidly lose their intensely marine character as they move inland. Halvor (1974) described small stands of this community at Yerba Buena Island, Brooks Island, Red Rock, Point Richmond, Point Fleming, and Potrero San Pablo, all places directly exposed to marine winds entering San Francisco Bay through the Golden Gate.

3. Chaparral - Chaparral refers to a widespread and characteristic California community dominated by large shrubs with evergreen sclerophyllous leaves. It is frequent in areas with precipitation between 10 and 20 inches per year and occasional in wetter areas on sunny south slopes. Chaparral occurs where rocky soils with little clay permit rapid infiltration of water and air to relatively great depths. Such conditions are most frequent on steep slopes but can also occur with relatively low relief on stone alluvial fans in valleys.

Holland and Keil (1995) subdivide California chaparral into 11 subclasses, of which six occur in San Francisco Bay counties. These are not separately treated here, however, since relatively little chaparral of any kind occurs in the Estuary’s immediate vicinity.

Chaparral is dominated by shrubs in the genera Adenostoma, Arctostaphylos, Ceanothus, Cercocarpus, and Quercus, which form a functional group characterized by deep root systems adapted for extracting water from deep cavities in fractured rock. The sclerophyllous leaves of chaparral shrubs are adapted for maintaining low levels of evapotranspiration and associated productivity during long growing seasons (Walter 1979, Mooney and Miller 1985). Discussions of chaparral ecology have long emphasized its adaptation to fire since its shrubs use a variety of strategies to rapidly reoccupy burns and an associated functional group of annuals has seeds that remain dormant for decades and only germinate following chaparral fires (Biswell 1974). Extensive research on Adenostoma-dominated chaparral suggesting a relatively short fire cycle may not be directly applicable to other chaparral types, however, since some other kinds of chaparral may have a much longer fire cycle (Keeley and Keeley 1988).

The nearest extensive chaparral to the Estuary occurs in Marin County on the slopes of Mt. Tamalpais two miles west of San Francisco Bay (Shuford 1993, Wieslander and Jensen 1945). While chaparral on alluvial fans is rapidly disappearing but still fairly common in parts of southern California (Smith 1980), it is virtually unknown in central and northern California. Cooper (1926), however, reported that chaparral that has since been extirpated formerly occurred near the southern end of San Francisco Bay on Los Gatos Creek’s alluvial fan.

4. Grassland – Vegetation dominated by grasses and graminoid sedges was widespread along the shores of the San Francisco Bay Estuary prior to urban development and is still fairly common there (Bryant 1848, McKelvey 1955). It occurs in non-wetlands wherever soils with clay horizons thick enough to hold significant water near the soil surface and to exclude air from deeper horizons are directly exposed to solar radiation. Clay soils are particularly favorable for grasses and other graminoids because the near-surface water they hold is preferentially available to the dense, relatively shallow fibrous root systems of such plants. In wet climates the most mesic conditions occur on soils with high clay content because of their high water holding capacity, but in arid areas that pattern is reversed. In deserts clay holds much water from scarce precipitation near the soil surface, where solar radiation quickly evaporates it (Walter 1979). Conditions intermediate between these extremes prevail in the semiarid climate surrounding most of the Estuary. Clay
soils are xeric and grass-covered on plains and south slopes, where they are directly exposed to solar radiation, but mesic and covered by forest and woodland on north slopes, where solar radiation is reduced. Grassland is most prevalent where annual precipitation is between 10 and 20 inches but becomes progressively scarcer as annual rainfall increases. Some grassland is usually present even in very wet areas, however, wherever clay is directly exposed to solar radiation. Soils with sufficient clay for grassland predominate on the recent alluvium that forms the floors of virtually all San Francisco Bay Area valleys; they are also common on hillslopes where clay-rich sediments have been uplifted (Ellen and W entworth 1995). While direct solar radiation usually keeps grassland free of woody plants on valley floors and south-facing hillslopes, similar grassland frequently dominates understoreies beneath the oak woodland that occurs on north slopes because of less intense radiation (Holstein 1984a). Native perennial grassland. Frequent relict stands and clear descriptions by early travelers leave little doubt that most native grassland near the Estuary on both valley floors and hillslopes was dominated by a rhizomatous and largely sterile hybrid between Leymus triticoides and L. condensatus (Stebbins and W alters 1949). Hybrids between these species have been called Leymus x multiflorus, but since the hybrid dominant around the Estuary is too small to match descriptions of x multiflorus (Hickman 1993), it is here included in L. triticoides. Two frequently associated rhizomatous graminoids were Carex barbara and C. praegracilis, the latter being especially frequent at upland-wetland ecotones. N asella pulchra, a non-rhizomatous bunchgrass, has received more attention than any other species as a native grassland dominant. It frequently dominated grassland but mostly did so only near ecotones with coastal scrub and oak woodland where heavy clay grassland soils had begun to thin and dissipate or where specific substrates like serpentinite prevented development of typical grassland soils (Bryant 1848, McKelvey 1955).

Native grassland had numerous local variations ranging from topographic lows where soil water and clay accumulated to topographic highs where clay was thin and water scarce. Species indicating topographic lows (locally called swales) included Juncus balticus, Juncus xiphioides, Ranunculus californicus, and Sisyrinchium bellum, while N. pulchra and a variety of forbs indicated the highs. Along the Estuary shore at ecotones with tidal marsh, D ischistis spicata, another rhizomatous grass, was particularly prominent (H eady 1977, H oiland and K ei 1995).

A scarce native grassland type especially significant for its many rare plants occurs on salt-affected soils associated with inland basins and basin rims rather than coastal tidelands (Faber 1997). These inland alkaline grasslands share features like the prominence of D ischistis spicata with the grassland-tidal marsh ecotone but often differ from it in the presence of more bare ground and many species not occurring at the Estuary shore. Cooper (1926) reported H emizonia congesta and H. pungens were formerly dominant on similar soils near the southern end of the Estuary that are now completely covered by urban development. The best presently extant examples of alkaline grassland in the Estuary’s vicinity occur near Livermore in Alameda County and near Fairfield in Solano County. Other distinctive grassland types of unusual substrates supporting rare species are serpentinite grassland and sandy soil grassland (Skin- ner and Pavlik 1994).

Native annual forbland. W ester (1981) presented evidence that the southern San Joaquin Valley, an area traditionally considered former grassland, was dominated by annual forbs prior to European settlement. California vegetation classification has traditionally called all upland vegetation dominated by herbs grassland, but W ester’s work suggests much of the area traditionally mapped as grassland (K uchler 1964) was actually native annual forbland. Since native annual forbland occurs where rainfall is insufficient for most perennial grasses, it consequently was most extensive far south of the Estuary in the southern San Joaquin Valley. Numerous relict taxa suggest, however, that a narrow native annual forbland corridor extended north from there to near the Estuary shore in Contra Costa County because of a rain shadow along the inner Coast Range’s eastern base. Forbland elements also probably occurred even more widely wherever local conditions like soil infertility and trampling by megafauna suppressed otherwise ubiquitous perennial grasses. Even today wildflower displays (i.e., annual forb dominance) are most spectacular locally where soil is relatively infertile (i.e., Bear Valley in Colusa County and T able M ountain in Butte County [Faber 1997]) and most spectacular generally in years, as in 1991, when winter drought suppression of competitive grasses is followed by forb-promoting heavy spring rains. Some forbland species like E remocarpus setigerus have adapted well to anthropogenic land use changes but others have become rare (C onvolulus simulans, M adia radiata) or extinct (E schscholzia rhombipetala).

Non-native annual grassland. Introduction of grazing and agriculture during the nineteenth century caused a dominance shift in almost all of California’s grasslands from native perennial graminoids to Eurasian non-native annual grasses. Today dominance among such annuals changes spatially in a complex pattern reflecting soil conditions. O n catenas from thick, heavy clay soils to thinner, lighter ones a typical annual grass dominance sequence L olum multiformum-B romus hordeaceus A vena fatua-A vena barbata replaces a simpler perennial sequence L eymus triticoides-N asella pulchra still occasionally extant on the same catenas. Another common dominance sequence B romus hordeaceus-B romus diandrus-H ordeum murinum reflects shifts in soil nitro-
gen content from low to relatively high. The above species are the most frequently dominant non-native annual grasses, but others also occasionally participate. Cynodon dactylon, for example, frequently dominates annual grassland where rainfall is relatively high. Several exotic forbs are also becoming increasingly important components in a vast exotic herbaceous vegetation type that may only temporarily be called grassland. Sida rhombifolia ssp. varia is increasingly planted for forage in the Avena zone; Bidens ferulifera is important in the Lolium zone; and Centaurea solstitialis, especially, is a widespread invader of the B. hordeaceae zone, where Erodium botrys is also important when soil fertility is particularly low (Holland and Keil 1995). Grazing is particularly important for maintaining replacement of native perennial grasses with exotic annual species. At numerous sites around the Estuary, for example, dominance is shifting back from exotic annual grasses to Leymus triticoides and Carex barbara where expanding urbanization has at least temporarily caused the removal of grazing.

Coastal prairie. Where clay soils are directly exposed to marine air flow, a floristically distinct grassland occurs that Holland and Keil (1995) call northern coastal grassland but is widely known in California as coastal prairie. Coastal prairie is most frequent along the outer coast, but small amounts also likely occur near the Estuary where marine air flow is particularly direct.

Much of California’s coastal prairie is now dominated by two exotic perennial grasses, Anthoxanthum odoratum and Holcus lanatus, but many distinctive native perennial grasses like Agrostis pallens, Calamagrostis nutkaensis, Danthonia californica, Deschampsia caespitosa, Festuca idahoensis and Festuca rubra can also frequently be locally dominant. Two other distinctive plant species indicative of coastal prairie are Iris douglasiana and Juncus patens (Heady et al. 1977).

5. Coastal Coniferous Forest - Forests dominated by large coniferous trees occur along the eastern Pacific Coast in a high rainfall zone extending from central California to Alaska. Holland and Keil (1995) recognize two subdivisions of coastal coniferous forest, but only one of these, redwood forest, occurs near the Estuary.

Redwood forest. Extensive forests dominated by Sequoia sempervirens, the well-known redwood and the world’s tallest tree species, occur on the southern slopes of Mt. Tamalpais within 1.75 miles of the Estuary (Shuford 1993), but individual redwoods occur in mixed evergreen forest much less than a mile from the shore of San Pablo Bay at China Camp State Park. Redwoods are common up to about 2,000 feet in the California Coast Range wherever annual precipitation is above 40 inches and soil is relatively fertile. Despite sufficient rainfall, sensitivity to cold prevents their occurrence along the Oregon coast beyond a few miles north of the border, at high elevations in the Coast Range, or on inland mountain ranges. Contrary to an enduring myth, redwoods are negatively rather than positively associated with summer fog. Consequently, even at sites protected from onshore winds they are almost completely absent along the immediate coast wherever summer fog is frequent. Redwoods survive summer drought not because of fog drip but by storing surplus water from high winter precipitation in their massive trunks, a strategy that has produced only slightly less dramatic gigantism in other conifer species where large winter water surpluses occur with summer drought. As a consequence northern California and southern Chile, both areas with unusually wet winters and dry summers located at the outer periphery of more typical Mediterranean climate zones, are the world’s two greatest centers of tree gigantism (Holstein 1984a, Zinke 1977).

Shade is so intense in the redwood forest understory that only a few plant species survive there. The trees most frequently dominating such mixed evergreen forests are Arbutus menziesii, Lithocarpus densiflorus, Pseudotsuga menziesii, and Umbellularia californica. In California the term mixed evergreen forest as presently used is somewhat anomalous, however, since it often designates forests solely dominated by Umbellularia californica, the California laurel. Such laurel-dominated forests are frequent around the Estuary where annual rainfall is between 20 and 40 inches. At the dry end of that precipitation range laurel forests are entirely confined to very shady north slopes and canyons, but they also occur on somewhat sunnier slopes as 40 inches is approached. Above that they are almost entirely replaced by redwood forests (Sawyer et al. 1977, Wainwright and Barbour 1984).

The most commonly associated tree species in central California’s laurel forests is a non-evergreen, Acer macrophyllum. Arbutus menziesii is also a frequent associate but is almost entirely confined to the rockiest slopes. Shade is so intense beneath laurel forest canopies that completely bare ground is common where drought is an added stressor, but as 40 inches is approached Polystichum munitum often dominates the understory. Holodiscus discolor, a deciduous species, is commonly dominant in shrubby openings frequent in laurel forests (Safford 1995).

6. Mixed Evergreen Forest - Central California mixed evergreen forest. Forests dominated by a mix of broadleaf and conifer evergreen trees are frequent in California where precipitation is relatively high and winter temperatures are mild. In northern California the trees most frequently dominating such mixed evergreen forests are Arbutus menziesii, Lithocarpus densiflorus, Pseudotsuga menziesii, and Umbellularia californica. In central California the term mixed evergreen forest as presently used is somewhat anomalous, however, since it often designates forests solely dominated by Umbellularia californica, the California laurel. Such laurel-dominated forests are frequent around the Estuary where annual rainfall is between 20 and 40 inches. At the dry end of that precipitation range laurel forests are entirely confined to very shady north slopes and canyons, but they also occur on somewhat sunnier slopes as 40 inches is approached. Above that they are almost entirely replaced by redwood forests (Sawyer et al. 1977, Wainwright and Barbour 1984).

The most commonly associated tree species in central California’s laurel forests is a non-evergreen, Acer macrophyllum. Arbutus menziesii is also a frequent associate but is almost entirely confined to the rockiest slopes. Shade is so intense beneath laurel forest canopies that completely bare ground is common where drought is an added stressor, but as 40 inches is approached Polystichum munitum often dominates the understory. Holodiscus discolor, a deciduous species, is commonly dominant in shrubby openings frequent in laurel forests (Safford 1995).

7. Oak Woodland - Vegetation with an overstory dominated by oak trees is common throughout California’s Mediterranean climate zone including the Estuary’s vicinity. Such oak woodlands primarily vary in species and spacing of their overstory oaks. Vegetation is called a-
vannah where oaks are widely spaced and forest where spacing is so close their canopies are closed. Woodland, as a term, describes vegetation with intermediate spacing, but tree separation is so locally variable in California's oak-tree dominated vegetation it is appropriate to use the traditional term oak woodland to refer to all of it. That generalized oak woodland can then be divided into subclasses based on its dominant species (Griffin 1977).

Since woodland oaks and grassland grasses occur in similar environmental conditions, they frequently compete directly for water and other soil resources. Specific aspects of that competition are discussed for each subclass but a few of its consequences are general. Oaks only occur where water is present in deep soil horizons, where it may arrive horizontally through shallow aquifers or vertically when precipitation is abundant enough to infiltrate past dense but relatively shallow grass root systems. Grassland grasses, in contrast, only occur where solar radiation is direct because overstory trees are either absent or so widely spaced their canopies are not contiguous (Walter 1979).

Coast live oak woodland, which is dominated by *Quercus agrifolia*, is distinctive among oak woodland subclasses because it consists almost exclusively of closed canopy forests. As a consequence it is frequently treated as a subclass of mixed evergreen forest rather than oak woodland. It is included here with oak woodland, however, because of the affinities of both its dominant tree and the majority of its fauna (Griffin 1977).

Coast live oak woodland occurs widely around the Estuary where annual precipitation is between 15 and 40 inches and continentality is at least partially moderated by marine influences. Marine air flow through Carquinez Strait even permits occurrence of coast live oak woodland with two isolated Coast Range-related mammal populations (*Sylvilagus bachmani riparius* and *Neotoma fuscipes riparia*) on the Central Valley floor near Lodi (Zander et al. 1990).

In hills on clay soils coast live oak woodland is frequently present as an extensive ecotone between grassland and mixed evergreen forest since it occurs on slopes shadier than the former but sunnier than the latter. On slopes where rockier substrates and lighter soils permit infiltration of more water to greater depths, however, coast live oak is less limited by solar exposure and can even occur on south slopes. North of Carquinez Strait, for example, adjacent ridges with identical microclimates differ only in their substrates. Ridges underlain by sediments of the clay-rich Eocene M arkle Formation are covered by grassland and have coast live oak woodland only on north slopes and in canyons, while those underlain by hard but fractured rocks of the Pliocene Sonoma volcanics are covered by coast live oak woodland on all exposures but north slopes and canyons, where *Umbellularia*-dominated mixed evergreen forest occurs. The great vegetational difference is a result of the way in which the two substrates respond to precipitation — rain rapidly infiltrates to deep levels on the fractured volcanics where it can be utilized by oak roots, whereas it is held at the surface on the clay-rich M arkle where it is more available to grass roots. Rapid infiltration on the volcanics causes such xeric conditions in its surface soils that its few stands of annual grassland cease productivity and dry two weeks earlier than M arkle grasslands dominated by the same species (Bates 1977, Ellen and Wentworth 1995).

Coast live oak woodland differs from other oak woodland subclasses in the relative rarity of annual grasses in its understory. The most frequent dominant there is *Toxicodendron diversilobum*, poison oak, but *Rubus ursinus* and *Symphoricarpus mollis* are also often important (Safford 1995).

Valley oak woodland consists of several structurally diverse communities sharing dominance by *Quercus lobata* that include savannah and woodland on clay hillslopes and savannah, woodland, and closed canopy forest on alluvial plains over shallow unconfined aquifers. Alluvial valley oak woodland often occurs on the outer edges of riparian forest corridors (see below) on relatively fine, heavy soils distinct from the coarse alluvial soils under typical riparian stream bank vegetation. Tree spacing in alluvial valley oak woodland is related to water stress since canopies closed when subsurface water is abundant become progressively more open as water stress increases, resulting first in woodland and then savannah. Much alluvial valley oak forest was removed because it coincided with highly desired agricultural soils, but a few stands are extant in the Central Valley and elsewhere. Alluvial valley oak woodland was probably always scarce near the Estuary, however, since it is better adapted to inland California's hot summers than to the outer Coast Range's relatively marine climate. One of the few examples near the Estuary is located along Green Valley Creek near Cordelia in Solano County.

Valley oak woodland is most frequent near the Estuary on clay hillslopes with annual rainfall between 15 and 40 inches, where its range overlaps coast live oak woodland and foothill woodland. It is less abundant than either but more tolerant of clay soils than the former and less resistant to water stress than the latter. The understory of valley oak woodland's savannah and woodland phases typically consists of non-native annual and occasionally native perennial grassland. Vegetation beneath closed canopy valley oak alluvial forest, however, can include both grassland and features shared with riparian forest or coast live oak forest understories. Valley oak reproduction, often low because of competition with annual grass and predation of seeds and seedlings by a variety of herbivores, can be abundant in alluvial woodland when suppression of grass by flooding coincides with large acorn crops. Urban fringes are also favorable.
sites for valley oak reproduction because their low livestock and wildlife populations result in lowered seed and seedling predation (Holstein 1984b, Holland and Keil 1995).

Foothill woodland is woodland and savannah vegetation wholly or partially dominated by Quercus douglasii, blue oak, that is widespread on hillslopes surrounding the Central Valley. Near the Estuary, foothill woodland is largely confined to the inner Coast Range. The foothill woodland environment has a relatively continental climate with cool to cold winters, very hot summers, and annual rainfall from 15 to 40 inches. Winter cold reduces understory grass growth and consequently permits infiltration of a large part of the wet season’s water surplus to deep subsoil where it can be utilized by blue oaks during spring and summer. In summer high temperatures and low humidity produce very low water potentials in blue oak leaves that permit withdrawal of water tightly held by clay-rich subsoils.

Blue oak is usually the sole foothill woodland dominant on clay hillslopes, but on slopes with more rock and thinner soils it often shares dominance with Pinus sabiniana. Blue oaks occur on a wider range of slope exposures than many other oak species, but foothill woodland dominance often shifts to Aesculus californica on shaded north slopes. In canyons and around rock outcrops Quercus wislizenii is also often a local dominant.

Because of foothill woodland’s open canopy its understory is almost universally dominated by non-native annual grassland. Native forbs like Holocarpha virgata, however, are also usually frequent there. Competition is particularly intense between annual grasses and blue oak seedlings before they develop roots long enough to reach subsoil water. Seedling mortality at this stage is so intense that much foothill woodland consists almost entirely of mature blue oaks that germinated in the 1860's, a decade when severe overgrazing reduced much presumably native perennial grassland from California’s rangelands. Subsequent increase of non-native annual grassland has severely restricted reproduction of foothill woodland developing at that time (Griffin 1977, Holland and Keil 1995).

8. Cliffs and Rock Outcrops - Vegetation of cliffs and rock outcrops is usually virtually ignored in surveys of California vegetation including that of Holland and Keil (1995) because its areal extent is small and it consists largely of non-vegetated surfaces. It is particularly important, however, as a habitat for rare plant species. Cliffs are unique environments where soil and competition with other plants is very limited and solar radiation is often abundant. Plants adapted to cliffs (chasmophytes) resemble epiphytes in producing small easily dispersed seeds in such great numbers that the likelihood of reaching rare suitable germination habitats is increased. Seeds reaching these light-rich habitats can afford to be small because they require little stored food.

Dudleya, the California genus with the most highly adapted chasmophytes, has a few taxa near the Estuary but is much more diverse in Southern California. Plants of rock outcrops are less specialized than chasmophytes but may be rare since they occur in distinctive microenvironments that consequently are free from competition with surrounding vegetation. Rock outcrops and cliffs are most likely to support rare plants when they are mineralogically different from surrounding landscapes, and one mineral receiving particular attention because of its frequent association with rare plants is serpentine. Soil development is so retarded and vegetation so distinctive on serpentine that its occurrences may be viewed as extended rock outcrops even though they occasionally cover many square miles (Bakker 1984, Skinner and Pavlik 1994, Fiedler and Leidy 1987).

9. Riparian Vegetation - Riparian vegetation refers to the distinctive plant communities of streambanks and ecologically related habitats. Its most salient environmental features are relatively coarse alluvial soils typically associated with streams and root zone water supplies greater than the local climate provides. When mature, California riparian vegetation is closed canopy forest, but early successional riparian vegetation can be shrubby.

Near the Estuary riparian vegetation is overwhelmingly dominated by three species, Acer negundo, Salix lasiolepis and Salix laevigata, but others may dominate in specialized habitats. Populus fremontii and Salix gooddingii are important where climate becomes less marine and more continental near the Central Valley; Salix exigua is important on sandbars and other habitats where early successional riparian vegetation is developing; Alnus rhombifolia and Salix lucida ssp. lasiandra are important where, as at Niles Canyon, streams with rocky beds flow perennially; and Platanus occidentalis and Baccharis viminea dominate where ones with sandy and rocky beds flow intermittently.

Typical Acer negundo-Salix lasiolepis-Salix laevigata riparian vegetation also is common where ecological conditions simulate streambank environments, as at lakeshores and a variety of places with shallow water tables. On the outer coast non-streambank riparian vegetation is frequent in dune slacks, but around the Estuary it at least formerly was most frequent in sausals, microdeltas occurring where stream channels and their subsurface water tables spread laterally as they entered tidal marsh. Most sausals have been lost to urbanization of the Estuary’s periphery, but a small example occurs at China Camp State Park.

Common riparian understory plants near the Estuary include Baccharis douglasii, Euthamia occidentalis, Rosa californica, and Rubus ursinus. For a short distance these can also replace riparian trees as dominants at the ecotone with tidal marsh where a veneer of coarse stream-
side alluvium deposited on tidal mud thins as it nears the Estuary. Like sasauals, however, such riparian-tidal marsh ecotones have almost entirely disappeared around the Estuary because of urbanization (Holland and Kel 1995, Holsten 1984b).

10. Freshwater Vegetation - Freshwater wetland vegetation occurs where land surfaces are saturated by freshwater or shallowly covered by it. Its two main phases near the Estuary, freshwater marshes and vernal pools, are very distinctive but also united by intermediate communities.

Freshwater marsh refers to vegetation dominated by plant species emergent from at least semi-permanent shallow freshwater. The most frequently dominant freshwater marsh species near the Estuary is Scirpus acutus, but Scirpus americanus, Scirpus californicus, Typha angustifolia, Typha domingensis, and Typha latifolia can also be important there as dominants. The Typha spp., in particular, are often dominant in early successional and nitrogen-enriched freshwater marshes.

Climate and geology have less influence on the distribution of freshwater marsh than they do on the occurrence of other plant communities. When vegetation is primarily limited by precipitation, temperature, and light, its distribution is controlled by climate, and when limited by mineral nutrition and soil texture, its distribution is controlled by geology. The primary limiting factor in freshwater marshes, however, is air, which, while superabundant at the marsh surface, falls to such low concentrations a short distance beneath it that environments too anoxic, reduced, and toxic for root growth are frequent there. All freshwater marsh dominants in California are consequently monocotyledons, which have shallow fibrous root systems readily supplied with air by aerenchyma-rich stems. Many dicotyledons including most trees and shrubs, in contrast, have solid stems and deep, non-fibrous root systems poorly adapted to anoxic conditions. California consequently lacks swamps, vegetation in semi-permanent shallow water dominated by woody plants, since it has no native trees or shrubs capable of completing life cycles in flooded environments. Buttonbush, Cephalanthus occidentalis, and several species of Salix can tolerate extended flooding, however, and frequently occur at the ecotone between freshwater marsh and riparian vegetation (Holland and Kel 1995, Holsten 1984a,b).

Both tidal and non-tidal freshwater marshes are frequent around the Estuary, but the former are most important in the Sacramento-San Joaquin Delta immediately upstream from the true estuary. The Delta consisted almost entirely of tidal freshwater marsh before it was largely converted to agricultural land, but a few remnant tidal freshwater marshes still occur there. Small non-tidal freshwater marshes often resulting from human alteration of hydrologic conditions are also widespread around the estuary (Bowcutt 1996).

Soils beneath freshwater marshes may be mineral or organic. Despite otherwise similar vegetation freshwater marshes with organic soils are technically fens. Since organic soils derived from Scirpus acutus rhizomes were general beneath the Delta’s tidal freshwater marshes, they once constituted a single vast fen (Atwater and Belknap 1980). Mineral soils, however, generally occur beneath the many small freshwater marshes around the Estuary. Marshes develop most readily on fine, heavy mineral soils since these exclude air and create the anaerobic conditions suitable for marsh vegetation more readily than the coarse and readily aerated sediments common on streambanks beneath riparian forests. Since waterbirds quickly transport propagules permitting establishment of freshwater marsh plants at sites with suitable hydrological conditions regardless of their climatic and geological environments, freshwater marshes are among the easiest plant communities to restore (Kusler and Kentula 1990).

Continua exist between semi-perennial marshes and both moist grassland swales (see above) and vernal pools (see below). Vegetation of areas with hydrology intermediate between freshwater marshes and vernal pools pond longer than the latter but shorter than the former. These are most frequently dominated by Eleocharis macrostachya with normal winter wet season inundation but can also be dominated by Cyperus eragrostis when ponding resulting primarily from agricultural and urban runoff occurs in the warm season. Vegetation arising from both kinds of seasonal ponding is properly called seasonal marsh, but wildlife managers also frequently use the term to describe non-tidal mudflat environments extremely important for shorebird foraging. Such non-tidal mudflats have little vegetation and once commonly occurred where flooding temporarily suppressed normal grassland development on stream terraces. Streamflow control and terrace urbanization, however, have greatly reduced traditional episodically flooded shorebird habitat around the Estuary. Most non-tidal seasonal marshes presently occurring there consequently result from seasonal drawdowns of artificial ponds and floodways (SFEP 1991a,b).

Limnetic vegetation refers to floating or submerged vegetation occurring in open freshwater too deep or otherwise unsuitable for marsh vegetation. Important native components of submerged limnetic vegetation near the Estuary include Ceratophyllum demersum, Najas guadalupensis, Potamogeton perpusitus and Potamogeton pusillus, while important floating elements are Azolla filiculoides, Lemna gibba and Lemna minor. Non-native species like Egeria densa and Myriophyllum aquatica are now also extremely significant and often predominant elements of submerged limnetic vegetation near the Estuary, but the floating component consists almost entirely of extremely widespread and readily dispersed native species except in and near the Delta, where non-na-
Vernal pool vegetation refers to a distinctive plant community dominated by annual and short-lived perennial forbs that occur in microtopographic basins flooded in the wet season and dry thereafter. Vernal pool plants are consequently adapted for beginning their life cycle like submerged limnetic species but completing it as terrestrial plants in completely dry environments. The vernal pool environment has led to adaptive radiation of numerous species primarily in the genera Downingia, Eryngium, Lasthenia, Nanneretia, Plagiobothrys, and Psilocarphus, and it is these that dominate its vegetation. Plagiobothrys bracteatus, in particular, is the most frequent dominant of vernal pool vegetation around the Estuary. Upland vegetation around vernal pools is almost invariably non-native annual grassland (Hol l and 1977).

The origin of vernal pool basins is obscure but may result from seismic activity or interaction of wind erosion and soil processes (see above). To pond water and create an environment suitable for vernal pool vegetation, however, an aquaclude or barrier to water infiltration that may be a clay horizon, duripan, or bedrock must be present immediately beneath the basin. Most vernal pools and the plants adapted to them occur entirely or almost entirely in California, and few hydrologic features resembling vernal pools occur outside North America even in otherwise similar Mediterranean climates (Thor nes 1984).

Vernal pools are at risk even in the Central Valley where they are most common because virtually all human activities except rangeland grazing destroy the microtopography and aquacludes that create the vernal pool environment. Around the Estuary they are even more threatened since they are extremely rare near southern San Francisco Bay and only slightly more frequent north of San Pablo and Suisun bays. Vernal pools north of Suisun Bay are particularly environmentally significant because they are often partially dominated by Lasthenia conjugens, a federally listed endangered species extinct throughout much of its range (Skinner and Pavlik 1994).

Artificially created basins often sufficiently resemble natural vernal pools to be colonized by a few wide-ranging and extremely tolerant pool species. More rarely a few rare species may be present in such artificial sites. The full suite of vernal pool taxa including the rarest species almost never develops in such environments, however, because soil characteristics of natural pools can rarely be replicated. As a consequence creation of artificial vernal pools has been the least successful of all wetland restoration efforts (Ferren and Ge v 1990, Kusler and Kentula 1990).

Vernal pools typically are freshwater environments since their primary water source is precipitation (Hanes et al. 1990). However, salt diffusion from underlying soils causes some to be slightly brackish. Seasonal pools hydrologically resembling vernal pools but lacking their characteristic biota because of elevated salinity also occur. They are called playas when their surrounding saline environment is inland (Chapman 1960, Waisel 1972) and pans when it is coastal (Adam 1990, Chapman 1960, Long and Mason 1983, Waisel 1972). Both occur in San Francisco Bay Area counties but only the latter near the Estuary shore (SFEP 1991a). Today vegetated pans near the Estuary are ubiquitously dominated by Cotula coronopifolia, an exotic annual that may have replaced a now extinct native annual Plagiobothrys glaber.


Agricultural environments historically surrounded much of the Estuary but have become increasingly scarce because of displacement by urbanization. Structurally and physiologically different elements like orchards, vineyards, and both irrigated and dry farmed cropland are included here, but all share low plant and animal diversity. Irrigated nursery crops are most important near the southern part of the Estuary; and vineyards, irrigated pastures, and dry farmed oats (Avena sativa) predominate near the northern part.

Urban and suburban environments now overwhelmingly dominate non-tidal uplands around the Estuary. They often structurally resemble extended and unusually diverse riparian zones since irrigated non-native trees predominantly in the genera Acacia, Eucalyptus, and Pinus are ubiquitously present above an even more diverse understory of ornamental shrubs and herbs. As a consequence, urban-suburban communities are probably the landscape unit near the Estuary with the highest plant diversity but the fewest native plants. Some native bird species have adapted to using urban areas as habitat and become common, but far fewer terrestrial species are able to do so.

Rare Plants of Ecotonal Plant Communities

Table 1.4 lists rare plant species found in the nine Bay Area counties. The table is organized using the same classification system (Hol l and K ei) as was used in the previous section. It includes, for each species, the state and federal listing status, as well as the status derived from the California Native Plant Society (CNPS) inventory (Skinner and Pavlik 1994). The CNPS inventory is more complete than the state or federal listings and is organized on the following lists:

1a. Presumed extinct
1b. Rare, threatened, or endangered in California and elsewhere.
2. Rare, threatened, or endangered in California but more common elsewhere.
3. Possibly rare, but more information is needed.
4. Distribution limited: a watch list.

Plants with the greatest need for protection are on list 1b, and 1a (presumed extinct) plants are placed there if rediscovered. The CNPS inventories rare plants by county. To prepare Table 1.4, each of the CNPS-identified rare species was assigned to a modal plant community or ecotone based on information provided by state and local floras. An effort was made to place each taxon in the plant community it most frequently (but not necessarily exclusively) occurred in, however, frequently reference materials regarding a taxon were contradictory. In these cases I sought to develop a consen sus view, and weighted local floras and my own field experience most heavily.

The greatest constraint in preparing this table was the frequent sparsity of ecological information regarding rare species. Preparation was easiest in areas with abundant local floras since these are full of observations by botanists with deep knowledge of their region’s plants and habitats. Tragically, however, a number of plants near the Estuary went extinct or became extremely rare in an older era when little or no ecological information was provided when plants were collected. We can only speculate regarding the niches of these taxa.

Many species are rare because they occur in rare ecological niches. Historically, these have not been the focus of plant community classification, which is most concerned with the commonest kinds of vegetation. While there was an effort to include some of the rarer niches occurring near the Estuary in Table 1.4, it is not comprehensive, and rare niches distant from the Estuary are not included. Rare plants that occur primarily in plant communities distant from the Estuary were not included in the narrative community descriptions.

It is hoped this table will generate discussion and suggestions for its improvement.
1. **Sand dune vegetation**
   A. Pioneer coastal dune vegetation
      1b. Abronia umbellata ssp. breviflora - FSC
   B. Coastal dune scrub
      1b. Agrostis blasdalei - FSC
      Chorizanthe cuspidata var. cuspidata - FSC
      Chorizanthe cuspidata var. villosa
      Chorizanthe robusta var. robusta - FE
      Collinsia corymbosa
      Erysimum ammophilum - FSC
      Horkelia cuneata ssp. sericea - FSC
      Horkelia marinensis - FSC
      Layia carnosa - FE, SE
      Lessingia germanorum - FE, SE
      Lupinus tidestromii - FE, SE
      +Gilia capitata ssp. chamissonis
      +Gilia millefoliata
      4. Monardella undulata
   C. Inland dune vegetation
      1b. Erysimum capitatum ssp. angustatum - FE, SE
   2. Coastal scrub
      A. Northern coastal scrub
      1b. Delphinium bakeri - FPE, SR
      Lilium maritimum - FC
      +Lupinus latifolius var. dudleyi
      3. Lupinus eximius - FSC
      4. Cirsium andrewsii
      Collinsia multicolor
      Piperia michaelii
      Sanicula hoffmannii
      B. Southern coastal scrub
      C. Sea-bluff coastal scrub
      1b. Cirsium occidentale var. compactum - FSC
      Grindelia hirsutula var. maritima - FSC
      Phacelia insularis var. continens - FSC
      Silene verecunda ssp. verecunda - FSC
      +Agrostis clivicola var. punta-reyesensis
      +Gilia capitata ssp. chamaeleonis
      +Gilia milletiota
      4. Monardella undulata
   3. Chaparral
      1b. Arctostaphylos auriculata
      Arctostaphylos densiflora - FSC, SE
      Arctostaphylos spinosa - FSC, SE
      Arctostaphylos manzanita ssp. laevigata
      Arctostaphylos montanae - FSC
      Arctostaphylos paullia - FPT, SE
      Arctostaphylos stanfordiana var. decumbens
      Ceanothus confusus - FSC
      Ceanothus divergens - FSC
      Ceanothus folius var. vineatus - FSC
      Ceanothus masonii - FSC, SR
      Ceanothus sonomensis - FSC
      Malacothamus hallii
      Plagiobothrys uncinatus - FSC
      3. Calyptdris pumy var. hesseae
      +Arctostaphylos manzanita ssp. elegans
      4. Ceanothus purpureus
      Dichonda occidentalis
      Lomatium rostratum
      Malacothamus australis
      Malacothamus helleri
      Orobanche valida ssp. howellii
      Plagiobothrys myosotoides
   A. Chaparral burns
      4. Calandrinia breweri
      +Malacothrix phaeocarpa
   4. Grassland
      A. Native perennial grassland
      1b. Amsinckia grandiflora - FE, SE
      Astragalus clarinervis - FE, ST
      Blepharidopsis plumosa ssp. plumosa
      Fritillaria plumosa - FSC
      Trypoxyla rostrata
      Trifolium amoenum - FC
      +Calochortus argillosus
      3. Lessingia hololeuca
      4. Androsace elongata ssp. acuta
      Fritillaria agrestis
      +Allium pensilifolium var. franciscanum
      +Microseris paludosa
   Alkaline grassland
   1a. Tropidocarpum capparideum - FSC
   1b. Astragalus tenuvar. pensiliformis - FSC
      Astragalus tenuvar. tener
      Atriplex cordulata - FSC
      Atriplex depressa
      Atriplex joaquiniana - FSC
      Cordylanthus mollis ssp. hispidus - FSC
      Cordylanthus palmatus - FE, SE
      Delphinium recurvatum - FSC
      Hemizonia parryi ssp. congdonii - FC
      Eriogonum argutum - FSC
      +Trifolium depauperatum var. hydropilum
      3. Hordeum intercedens
      4. Atriplex coronaria var. coronata
      Thelepodium brachycarpum

Table 1.4 Rare Plant Species* Found in the Nine Counties Adjacent to the San Francisco Bay Estuary, by Plant Community or Ecotone

<table>
<thead>
<tr>
<th>Plant Community</th>
<th>Species</th>
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<tbody>
<tr>
<td>Sand dune vegetation</td>
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<td>Dichonda occidentalis</td>
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<td></td>
<td>Lomatium rostratum</td>
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<td></td>
<td>Malacothamus australis</td>
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<td>Malacothamus helleri</td>
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<td></td>
<td>Orobanche valida ssp. howellii</td>
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<td>Plagiobothrys myosotoides</td>
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<td>Chaparral burns</td>
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<td></td>
<td>Calandrinia breweri</td>
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<tr>
<td></td>
<td>+Malacothrix phaeocarpa</td>
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<td>Native perennial grassland</td>
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<tr>
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<td>Amsinckia grandiflora - FE, SE</td>
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<td>Astragalus clarinervis - FE, ST</td>
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<tr>
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<td>Blepharidopsis plumosa ssp. plumosa</td>
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<td>Fritillaria plumosa - FSC</td>
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<td>Trypoxyla rostrata</td>
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<td></td>
<td>Trifolium amoenum - FC</td>
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<td></td>
<td>+Calochortus argillosus</td>
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<td>Lessingia hololeuca</td>
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<td>Androsace elongata ssp. acuta</td>
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<td>Fritillaria agrestis</td>
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<td>+Allium pensilifolium var. franciscanum</td>
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<td>+Microseris paludosa</td>
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<td>Alkaline grassland</td>
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<td>Tropidocarpum capparideum - FSC</td>
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<td>Astragalus tenuvar. pensiliformis - FSC</td>
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<td>Astragalus tenuvar. tener</td>
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<td>Atriplex cordulata - FSC</td>
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<td>Atriplex depressa</td>
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<td>Atriplex joaquiniana - FSC</td>
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<td>Cordylanthus mollis ssp. hispidus - FSC</td>
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<td>Cordylanthus palmatus - FE, SE</td>
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<td>Delphinium recurvatum - FSC</td>
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<tr>
<td></td>
<td>Hemizonia parryi ssp. congdonii - FC</td>
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<tr>
<td></td>
<td>Eriogonum argutum - FSC</td>
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<tr>
<td></td>
<td>+Trifolium depauperatum var. hydropilum</td>
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<td></td>
<td>Hordeum intercedens</td>
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<td></td>
<td>Atriplex coronaria var. coronata</td>
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<td></td>
<td>Thelepodium brachycarpum</td>
</tr>
</tbody>
</table>

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FSC Federal species of special concern
### Sandy soil grassland
1a. Eriogonum truncatum
4. Cryptantha hooveri 
   Linanthus grandiflorus

### Serpentinite grassland
1b. Acanthomintha duttonii - FE, SE  
   Calochortus tiburonensis - FT, ST  
   Castilleja affinis ssp. neglecta - FE, ST  
   Fritillaria biflora var. ineziana  
   Lessingia arachnoidea - FSC  
   Streptanthus niger - FE, SE  
3. Eriogonum luteolum var. caninum  
4. Astragalus breweri  
   Linanthus ambiguus

### Moist grassland
1a. Plagiobothrys hystriculus  
1b. Pleuropogon hooverianus - FSC, SR  
   Sidalcea oregana ssp. hydrophila
2. Carex californica
4. Perideridia gairdneri ssp. gairdneri - FSC

### Native annual forbland
1a. Eschscholzia rhombipetala - FSC
1b. Madia radiata
4. Convolvulus simulans  
   +Erodium macrophyllum

### Brown grassland
1b. Blennosperma nanum var. robustum - FSC, SR  
   chorisianthe valida - FE, SE  
   Erigeron supplex - FSC  
   Fritillaria lanceolata var. tristulis  
   Fritillaria lilacina - FSC  
   Holocarpha macadenia - FC, SE  
   Limnanthes douglasii ssp. sulphurea - FSC, SE  
   Plagiobothrys diffusus - FSC, SE  
   Sarcocaulon flavum - FSC  
   Tephysaria floribunda - FSC
3. Hemizonia congesta ssp. leucocephala  
   Plagiobothrys chorisianus var. chorisianus

### Coastal coniferous forest
A. Redwood forest  
4. Elymus californicus
B. Closed-cone coniferous forest
1b. Ceanothus glaucescens var. rupescens - FSC, SR  
   Cupressus goveniana ssp. arizonica - FSC, SR  
   Pinus radiata - FSC
C. North coast coniferous forest
2. Boschnia hookeri

### 6. Mixed evergreen forest
A. Central California mixed evergreen forest
1b. +Quercus panula var. tamalpaisensis  
3. +Viburnum ellipticum
4. Cypripedium montanum  
   Dirca occidentalis  
   Ribes victorius

### 7. Oak woodland
A. Coast live oak woodland
1b. Clarkia concinna var. automixia
4. Amsinckia minuta var. diabolica
   Isocoma menziesii var. diabolica
B. Valley oak woodland
C. Foothill oak woodland

### 8. Cliffs and rock outcrops
A. Cliffs
3. +Streptanthus portoriusus var. suffrutescens  
4. Arabis modesta
B. Rock outcrops
1b. Arctostaphylos virgata  
   Coreopsis hamiltonii - FSC  
   Penstemon newberryi var. sonomensis  
   Phacelia phaceloides - FSC  
   Sarcocaulon flavum - FSC  
   Streptanthus glandulosus var. hoffmanii - FSC  
   Streptanthus hispidus - FSC
3. Erigeron boletii  
   Monardella antonina ssp. antonina
4. Antirrhinum virga  
   Arabis oregona  
   Arctostaphylos hispidula  
   Navaretia subuligera

### Serpentinite outcrops
1a. Arctostaphylos hookeri ssp. franciscana - FSC
1b. Allium sharshmitae  
   Arctostaphylos bakerii ssp. bakerii - SR  
   Arctostaphylos bakerii ssp. sublaevis  
   Arctostaphylos hookeri ssp. montana - FSC  
   Arctostaphylos hookeri ssp. ravenii - FE, SE  
   Ceanothus rupicola var. rupicola  
   Calochortus raieii - FSC  
   Campanula sharshmitiae - FSC  
   Ceanothus fertilis - FE  
   Chlorogalum pomeridianum var. minus  
   Clarkia franciscana - FE, SE

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Table 1.4 (continued) Rare Plant Species* Found in the Nine Counties Adjacent to the San Francisco Bay Estuary, by Plant Community or Ecotone

<table>
<thead>
<tr>
<th>Plant Community or Ecotone</th>
<th>Species</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sandy soil grassland</td>
<td>Eriogonum truncatum, Cryptantha hooveri, Linanthus grandiflorus</td>
</tr>
<tr>
<td>Serpentinite grassland</td>
<td>Acanthomintha duttonii, Calochortus tiburonensis, Castilleja affinis, Fritillaria biflora, Lessingia arachnoidea, Streptanthus niger, Eriogonum luteolum var. caninum, Astragalus breweri, Linanthus ambiguus</td>
</tr>
<tr>
<td>Moist grassland</td>
<td>Plagiobothrys hystriculus, Pleuropogon hooverianus, Sidalcea oregana ssp. hydrophila, Carex californica, Perideridia gairdneri, FSC</td>
</tr>
<tr>
<td>Native annual forbland</td>
<td>Eschscholzia rhombipetala, Madia radiata, Convolvulus simulans, Erodium macrophyllum</td>
</tr>
<tr>
<td>Coastal prairie</td>
<td>Blennosperma nanum, Fritillaria lanceolata, Erigeron supplex, Fritillaria lilacina, Holocarpha macadenia, Limnanthes douglasii ssp. sulphurea, Plagiobothrys diffusus, Sarcocaulon flavum, Tephysaria floribunda, Hemizonia congesta, Plagiobothrys chorisianus</td>
</tr>
<tr>
<td>Coastal coniferous forest</td>
<td>Elymus californicus</td>
</tr>
<tr>
<td>Redwood forest</td>
<td>Arctostaphylos virgata, Coreopsis hamiltonii, Penstemon newberryi, Phacelia phaceloides, Sarcocaulon flavum, Streptanthus glandulosus, Streptanthus hispidus, Erigeron boletii, Monardella antonina, Antirrhinum virga, Arabis oregona, Arctostaphylos hispidula, Navaretia subuligera</td>
</tr>
<tr>
<td>Closed-cone coniferous forest</td>
<td>Arctostaphylos hookeri ssp. franciscana</td>
</tr>
<tr>
<td>Cliffs</td>
<td>Arctostaphylos hookeri ssp. bakerii, Arctostaphylos hookeri ssp. sublaevis, Arctostaphylos hookeri ssp. montana, Ceanothus rupicola, Calochortus raieii, Campanula sharshmitiae, Ceanothus fertilis, Chlorogalum pomeridianum var. minus, Clarkia franciscana</td>
</tr>
<tr>
<td>Rock outcrops</td>
<td>Arctostaphylos virgata, Penstemon newberryi, Phacelia phaceloides, Sarcocaulon flavum, Streptanthus glandulosus, Streptanthus hispidus, Erigeron boletii, Monardella antonina, Antirrhinum virga, Arabis oregona, Arctostaphylos hispidula, Navaretia subuligera</td>
</tr>
<tr>
<td>Serpentinite outcrops</td>
<td>Arctostaphylos hookeri ssp. franciscana</td>
</tr>
<tr>
<td>Cliffs</td>
<td>Arctostaphylos hookeri ssp. bakerii, Arctostaphylos hookeri ssp. sublaevis, Arctostaphylos hookeri ssp. montana, Ceanothus rupicola, Calochortus raieii, Campanula sharshmitiae, Ceanothus fertilis, Chlorogalum pomeridianum var. minus, Clarkia franciscana</td>
</tr>
</tbody>
</table>
Table 1.4 (continued) Rare Plant Species* Found in the Nine Counties Adjacent to the San Francisco Bay Estuary, by Plant Community or Ecotone

<table>
<thead>
<tr>
<th>Community</th>
<th>Species</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Cordylanthus nidularius</strong> - FC, SR</td>
<td>Volcanic outcrops</td>
</tr>
<tr>
<td><strong>Cordylanthus tenuis</strong> ssp. capillaris - FE, SR</td>
<td>1b. Enarthrum brandegeae - FSC</td>
</tr>
<tr>
<td><strong>Cryptantha clevelandii var. dissita</strong></td>
<td>4. Madia nutans</td>
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<tr>
<td><strong>Dudleya setchellii</strong> - FE</td>
<td></td>
</tr>
<tr>
<td><strong>Erigeron angustus</strong></td>
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</tr>
<tr>
<td><strong>Erigeron serpentinus</strong></td>
<td></td>
</tr>
<tr>
<td><strong>Eriogonum nelsonum</strong> - FSC</td>
<td></td>
</tr>
<tr>
<td><strong>Fritillaria falcata</strong> - FSC</td>
<td></td>
</tr>
<tr>
<td><strong>Hesperolinon bicarpellatum</strong> - FSC</td>
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</tr>
<tr>
<td><strong>Hesperolinon breviflorum</strong> - FSC</td>
<td></td>
</tr>
<tr>
<td><strong>Hesperolinon congestum</strong> - FT, ST</td>
<td></td>
</tr>
<tr>
<td><strong>Hesperolinon drymarioides</strong> - FSC</td>
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<tr>
<td><strong>Hesperolinon serpentinum</strong></td>
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</tr>
<tr>
<td><strong>Lessingia micrantha var. glabra</strong> - FSC</td>
<td>1b. Juglans californica var. hindsii - FSC</td>
</tr>
<tr>
<td><strong>Madia hallii</strong> - FSC</td>
<td></td>
</tr>
<tr>
<td><strong>Navarretia rosulata</strong></td>
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</tr>
<tr>
<td><strong>Sidalcea hickmanii</strong> ssp. viridis - FSC</td>
<td>4. Astragalus rattanii ssp. rattanii</td>
</tr>
<tr>
<td><strong>Streptanthus albidus</strong> ssp. albidus - FE</td>
<td>3. Cardamine pachystigma var. dissectifolia</td>
</tr>
<tr>
<td><strong>Streptanthus albidus</strong> ssp. peramoenus - FC</td>
<td>+Streptanthus breweri var. hesperidis</td>
</tr>
<tr>
<td><strong>Streptanthus batrachopus</strong> - FSC</td>
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<tr>
<td><strong>Streptanthus brachiatus</strong> ssp. brachiatus - FC</td>
<td>4. Acanthomintha lanceolata</td>
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<tr>
<td><strong>Streptanthus brachiatus</strong> ssp. Hoffmannii - FC</td>
<td>Aesclepias solanoana</td>
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<tr>
<td><strong>Streptanthus glandulosus</strong> ssp. pulchellus</td>
<td>Aspidotis carlotta-halliae</td>
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<tr>
<td><strong>Streptanthus montsonii</strong> ssp. elatus - FC</td>
<td>Calamagrostis ophitidis</td>
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<tr>
<td><strong>Streptanthus montsonii</strong> ssp. hirtiflorus - FSC</td>
<td>Calyptridium quadripetalum</td>
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<tr>
<td><strong>Streptanthus montsonii</strong> ssp. knuckebergii - FSC</td>
<td>Campanula exigua</td>
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<tr>
<td><strong>Streptanthus montsonii</strong> ssp. montsonii - FSC</td>
<td>Clarkia breweri</td>
</tr>
<tr>
<td><strong>+Holta strobilina</strong></td>
<td>Collyvia diversifolia</td>
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<tr>
<td>+Streptanthus breweri var. hesperidis</td>
<td>Cordylanthus tenuis ssp. bruneus</td>
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<tr>
<td><strong>3. Cardamine pachystigma var. dissectifolia</strong></td>
<td>Eriogonum argillosum</td>
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<tr>
<td>+Streptanthus glandulosus var. somomensis</td>
<td>Eriogonum tematum</td>
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<td><strong>4. Acanthonthina lanceolata</strong></td>
<td>Eriogonum tripodum</td>
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<tr>
<td><strong>Asclepias solanoana</strong></td>
<td>Fritillaria purdy</td>
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<td><strong>Aspidotis carlotta-halliae</strong></td>
<td>Galium andrewsi ssp. gatense</td>
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<tr>
<td><strong>Calamagrostis ophitidis</strong></td>
<td>Lomatium ciliolatum var. hooveri</td>
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<tr>
<td><strong>Calyptridium quadripetalum</strong></td>
<td>Navarretia leucocephala</td>
</tr>
<tr>
<td><strong>Campanula exigua</strong></td>
<td>Neostapfia colusana</td>
</tr>
<tr>
<td><strong>Clarkia breweri</strong></td>
<td>Tuctoria mucronata</td>
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<tr>
<td><strong>Collyvia diversifolia</strong></td>
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<tr>
<td><strong>Cordylanthus tenuis</strong> ssp. bruneus</td>
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<tr>
<td><strong>Eriogonum argillosum</strong></td>
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<tr>
<td><strong>Erigeron serpentinus</strong></td>
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</tr>
<tr>
<td><strong>Fritillaria purdy</strong></td>
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<tr>
<td><strong>Galium andrewsi</strong> ssp. gatense</td>
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<tr>
<td><strong>Lomatium ciliolatum</strong> var. hooveri</td>
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<tr>
<td><strong>Navarretia leucocephala</strong></td>
<td></td>
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<tr>
<td><strong>+Ceanothus jepsonii</strong> var. albiflorus</td>
<td>Neostapfia colusana - FT, SE</td>
</tr>
<tr>
<td>+Streptanthus barbiger</td>
<td>Tuctoria mucronata - FE, SE</td>
</tr>
</tbody>
</table>

**Granite and sandstone outcrops**

4. Arctostaphylos regismontana

9. **Riparian vegetation**

1b. Juglans californica var. hindsii - FSC
+Triteleia lugens

4. Astragalus rattanii ssp. rattanii

10. **Freshwater vegetation**

A. Freshwater marsh

1a. Castilleja uliginosa - FSC, SE

1b. Alopecurus aequilis var. sonomensis - FE
Arenaria paludicola - FE, SE
Campanula californica - FSC
Carex albida - FE, SE
Lilium pardalinum ssp. pitkinense - FE, SE
Potentilla hickmanii - FPE, SE
Rhyynchospora californica - FSC
Sidalcea calycosa ssp. rhizomata
Sidalcea oregana ssp. valida - FE, SE

2. Calamagrostis cassiligumis - FSC
Carex comosa
Carex leptalea
Rhyynchospora globularis var. globularis

3. Equisetum palustre

4. Calamagrostis bolanderi
Rhyynchospora alba
+Zigadenus micranthus var. fontanus

B. Limnetic vegetation

2. Potamogeton filiformis
Potamogeton zosteriformis

4. Azolla mexicana
Ranunculus lobii

C. Vernal pools

1b. Blephospemna bakeri - FE, SE
Gratiola heterosepala - SE
Lasthenia burkei - FE, SE
Lasthenia conjugens - FE
Legenere limosa - FSC
Limnanthes vinculans - FE, SE
Navarretia leucocephala ssp. bakeri
Navarretia leucocephala ssp. pauciflora - FE, ST
Navarretia leucocephala ssp. planeantha - FE, SE
Neostapfia colusana - FT, SE
Tuctoria mucronata - FE, SE

2. Downingia pusilla

3. Myosurus minimum ssp. apus - FSC
Pogogyne douglasii ssp. paviflora

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<th>Species</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Plants</strong></td>
<td></td>
</tr>
<tr>
<td>4. Eryngium aristulatum var. hooveri - FC</td>
<td>Lilium rubescens</td>
</tr>
<tr>
<td>Psilocarphus brevissimus var. multiflorus</td>
<td>Monardella viridis ssp. viridis</td>
</tr>
<tr>
<td>Psilocarphus tenellus var. globiferus</td>
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</tr>
<tr>
<td><strong>D. Thermal springs</strong></td>
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</tr>
<tr>
<td>1b. Dichanthelium lanuginosum var. thermale - FSC, SE</td>
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<tr>
<td>Psilocarphus brevissimus var. multiflorus</td>
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<tr>
<td>Psilocarphus tenellus var. globiferus</td>
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<tr>
<td><strong>11. Anthropogenic environments</strong></td>
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</tr>
<tr>
<td><strong>A. Agricultural</strong></td>
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</tr>
<tr>
<td><strong>B. Urban-suburban</strong></td>
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<tr>
<td><strong>12. Coastal marsh</strong></td>
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</tr>
<tr>
<td><strong>A. Brackish marsh</strong></td>
<td></td>
</tr>
<tr>
<td>1b. Aster lentus - FSC</td>
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</tr>
<tr>
<td>Cirsium hydrophilum var. hydrophilum - FE</td>
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<tr>
<td>Cordylanthus mollis ssp. mollis - FE, SR</td>
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<tr>
<td>Lathyrus jepsonii var. jepsonii - FSC</td>
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</tr>
<tr>
<td>Lilaepsis masonii - FSC, SR</td>
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</tr>
<tr>
<td><strong>2. Limosella subulata</strong></td>
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</tr>
<tr>
<td><strong>B. Saltmarsh</strong></td>
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</tr>
<tr>
<td>1b. Castilleja ambigua ssp. humboldtiensis - FSC</td>
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</tr>
<tr>
<td>Cordylanthus maritimus ssp. palustris - FSC</td>
<td></td>
</tr>
<tr>
<td>Suaeda californica - FE</td>
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</tr>
<tr>
<td><strong>3. Polygonum marinense - FSC</strong></td>
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<td><strong>O. Grassland-chaparral</strong></td>
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<td><strong>P. Northern coastal scrub-riparian</strong></td>
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<td>4. Veratrum fimbriatum</td>
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*Derived from the inventory of the California Native Plant Society (CNPS) (Skinner and Pavlik 1994)

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**Key to CNPS list codes:**
1a. Presumed extinct
1b. Rare, threatened, or endangered in California and elsewhere.
2. Rare, threatened, or endangered in California but more common elsewhere.
3. Possibly rare, but more information is needed.
4. Distribution limited; a watch list
+ Proposed new addition to the CNPS inventory

**Key to Federal and State List Codes:**
FE Federally listed as endangered
SE State listed as endangered
FT Federally listed as threatened
ST State listed as threatened
FC Federal listing candidate
SR State listed as rare
FPE Proposed for federal listing as endangered
FPT Proposed for federal listing as threatened
FSC Federal species of special concern

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References


Wester, L. 1981. Composition of native grasslands in the San Joaquin Valley, California. Madrono 28:
Chapter 2 — Estuarine Fish and Associated Invertebrates

**Opossum Shrimp**

*Neomysis mercedis*

Bruce Herbold

**General Information**

The opossum shrimp is a native mysid shrimp that is an important food for many estuarine fish, especially young striped bass. Since 1994, their role of dominant planktonic shrimp has been overwhelmed by the introduced species, *Acanthomysis* (Orsi and Mecum 1996).

**Reproduction**

The common name of the opossum shrimp derives from the fact that females carry their eggs and young in a pouch at the base of the last two pairs of legs. Young are released at a well-developed stage. Fecundity is related both to adult size and season (Heubach 1969).

Reproduction is continuous but the rate is strongly controlled by temperature and food supply. Thus, the rate is high during spring and summer months and slows down as temperature and insolation decline. The wintertime population is composed largely of large adults, whose greater fecundity allows rapid development of high densities as temperatures and phytoplankton densities rise. The autumn decline in density has been variously attributed to seasonal changes in high temperature, low dissolved oxygen, predation, and food supply (Turner and Heubach 1966, Heubach 1969, Siegfried et al. 1979, Orsi and Knutson 1979).

**Food and Feeding**

The diet of *N. mercedis* varies with size. At release, young shrimp eat mostly phytoplankton and rotifers. Adult diets include phytoplankton and rotifers but the diet shifts more to copepods, particularly *Eurytemora affinis* (Herbold et al. 1992).

**Distribution**

*N. mercedis* is found in greatest abundance in Suisun Bay and the western Delta, although it occurs as far upstream as Sacramento, the lower reaches of the Mokelumne River, and in the San Joaquin River to above Stockton.

**Population Status and Influencing Factors**

During most of the 1980s, the opossum shrimp population varied considerably, but remained at a lower level of abundance than existed in the early 1970s. Opossum shrimp abundance fell dramatically after 1986 and remained at very low levels from 1990 to 1993 (CDFG 1994). As a general trend, opossum shrimp populations have declined substantially in Suisun Bay, yet they have occasionally rebounded to high levels (BDOC 1993).

Reasons for the system-wide declines of several zooplankton taxa in the Bay-Delta Estuary are not known. Although the declines occurred at about the same time as declines in phytoplankton and various fish species, no cause-and-effect relationships have been established (CDWR 1992). However, several factors have been identified which are believed to have some influence on the decline of zooplankton in the Estuary.

Decrease in food supply has been associated with the decline in abundance of rotifers and the copepod, *E. affinis*. The decline of rotifers in the Delta appears to be strongly associated with declining concentrations of chlorophyll a, which formerly characterized the areas of greatest rotifer abundance (Herbold et al. 1992). However, chlorophyll and many zooplankton species have similar spatial distributions, and correlations between the two groups can arise through movement of the entrapment zone in the Estuary. Also, while it is commonly...
assumed that chlorophyll is a good measure of food availability for zooplankton. E. affinis can subsist on detrital matter and requires larger particles than those that make up total chlorophyll. In addition, small zooplankton could provide food for many of the larger zooplankton species (Kimmerer 1992). Consistently low E. affinis abundance in recent years has been named as a factor that has probably contributed to the decline of opossum shrimp (H erbold et al. 1992).

Introduced species have also been named as a potential cause for the decline in zooplankton abundance. For example, the introduction of Sinocalanus has been identified as a possible cause of the decline in abundance of E. affinis (Kimmerer 1992), although the introduced copepod does not have the same habitat requirements as the native copepods (NH I 1992). However, based on the known feeding habits of a related species of Sinocalanus, S. doerrii may prey on native copepods (H erbold et al. 1992). In addition, predation by the introduced Asian clam, Potamocorbula amurensis, has been suggested as a factor in the decline of rotifer (H erbold et al. 1992) and E. affinis populations. E. affinis abundance in Suisun Bay decreased substantially when the clam became abundant there in 1988 (CDWR 1992). Since 1994 N emysis abundance has dropped to less than that of an introduced species of mysid shrimp which has increased in abundance (Ors i 1996). Competition with both the clam and new shrimp is likely to prevent re-establishment of N emysis at the levels of their former abundance.

The decline in the abundance of opossum shrimp and other zooplankton species (e.g., E. affinis) that are found in the entrapment zone in relatively high abundances has been correlated with Delta outflow. It is presumed that low outflow reduces opossum shrimp abundance by: (1) restricting the entrapment zone to deeper, more upstream channels which are less likely to promote high densities of opossum shrimp; and (2) producing weaker landward currents along the bottom so that the ability of opossum shrimp transported downstream to return to the entrapment zone is reduced. It has also been presumed that larger numbers of opossum shrimp may be exported through the Central Valley Project and State Water Project pumps as a result of the increased proportion of inflow diverted during the years when the entrapment zone is upstream in the Estuary. The location of the entrapment zone within the lower river channels during dry years increases the vulnerability of opossum shrimp to such displacement (H erbold et al. 1992). However, analyses by Kimmerer (1992) suggest that exports by the water projects are not a major source of losses for opossum shrimp and E. affinis populations, primarily due to the small percentage of entrapment zone volume (and entrapment zone organisms) diverted. Depending on the timing, location, and quantity of withdrawals, in-Delta water diversions, whose net consumption is on the same order of export flows, may result in a higher rate of loss to resident zooplankton populations than export pumping.

Pollutants may be another factor in the decline of zooplankton in the upper Estuary. For example, rice herbicides have been shown to be toxic to opossum shrimp (CDWR 1992). However, rice herbicides are largely confined to the Sacramento River, not the entire Estuary. No Estuary-wide decline in planktonic crustaceans have been associated with the timing of herbicide occurrence in the river (NH I 1992).

**Trophic Levels**

The opossum shrimp is a primary and secondary consumer.

**Proximal Species**

**Predators:** Striped bass, longfin smelt, splittail.

**Prey:** Various copepods, various phytoplankton.

**Competitors:** Potamocorbula amurensis, Acanthomysis spp.

**Good Habitat**

Good habitat appears to be similar to that of Delta smelt; a well-dispersed area of open water with salinities in the range of 2 to 6 ppt for most of the year and clean, non-toxic over-wintering habitat in freshwater through the winter and early spring. Dead-end sloughs both in Suisun Marsh and upstream apparently serve as important refuges from predation during the annual period of low abundance and slow growth. With the advent of newly introduced competitors in the open waters of the Estuary it is possible that such refugia will become important for the year-round maintenance of opossum shrimp.

**References**


H erbold, B., P.B. M oyle and A. Jassby. 1992. Status and trends report on aquatic resources in the San Fran-
Dungeness Crab
Cancer magister
Robert N. Tasto

General Information
Dungeness crab has been the object of an immensely popular commercial and recreational fishery in the San Francisco region since 1848. The San Francisco fishery, which occurs exclusively outside the Golden Gate, was long a mainstay of statewide commercial landings. However, beginning in the early 1960s, it underwent a severe and longterm decline which persisted until the mid-1980s. The principal causes of the decline have been related to changes in ocean climate, increased predation, and possibly pollution (Wild and Tasto 1983). Landings in the past decade have rebounded to some extent and are generally able to accommodate local market demand, but the northern California fishery (Eureka and Crescent City) continues to be the major provider of Dungeness crabs throughout the rest of California. The value of the Dungeness crab resource extends beyond the traditional economic return to the fishermen, seafood processors, and retail markets, as it is an important element in the tourism industry of San Francisco.

California commercial and recreational fishing regulations pertaining to Dungeness crab have been designed to protect this species from over-harvesting. The standard commercial fishing gear is a baited 3.5-foot diameter metal trap, weighing 60 to 120 pounds (Warner 1992). California regulations set a 6.25-inch carapace width (cw) size limit, prohibit the take of female crabs, and, like most states, have established a specific fishing season to protect reproducing and egg-bearing crabs. A limited recreational fishery allows the take of female crabs and has a smaller size restriction (5.75 inches cw); a 10-crab bag limit is placed on the sportfishers. Recreational gear consists of a variety of traps, hoops, and nets of different sizes, shapes, and materials. It is currently

Actual body width to 9 inches

CDFG
illegal to catch Dungeness crab of any size in San Francisco Bay.

Reproduction

Mating occurs in nearshore coastal waters, from March through May, between hard-shelled males and recently molted, soft-shelled females. Fertilized eggs are extruded in the fall and lay protected beneath the female's abdominal flap in a sponge-like mass until hatching occurs from late December to mid-January (Wild and Tasto 1983). Fecundity ranges from 500,000 to 2,000,000 eggs, depending upon the size of the female (Warner 1992). C. magister is capable of about four broods over its reproductive life span (Hines 1991).

Growth and Development

Dungeness crab life stages include the egg, larval, juvenile, and adult. Dungeness crab eggs range in diameter from 0.016 to 0.024 inches (Warner 1992). There are a total of six larval stages (five zoeae and one megalopa) which spend about 3 to 4 months in both nearshore and offshore coastal waters; larval timing is believed to coincide with peak plankton production (Hines 1991). Late-stage megalopa, which have returned to the coast, bays, and estuaries via ocean currents and other mechanisms, settle onto relatively open sandy areas (Oresanz and Gallucci 1988) and subsequently metamorphose to the first bottom-dwelling instar stage generally between April and June. It is at this stage that the young crabs enter San Francisco Bay in large numbers, relative to year-class strength, seemingly aided by strong bottom currents (Tasto 1983). San Francisco Bay-reared crabs molt more frequently than those juveniles found in the near coastal environment and reach sexual maturity (approximately 4 inches wide) after nearly one year (Wild and Tasto 1983). This rate of growth is substantially greater than that found in open areas along the Pacific coast and may be due to increased availability of food and/or overall warmer temperatures of estuaries (Tasto 1983, Gunderson et al. 1990, Wainwright and Armstrong 1993). It is believed that the large number of molts necessary to reach sexual maturity in an estuarine environment is due, in large part, to the demands of osmoregulation (Oresanz and Gallucci 1988).

Food and Feeding

Larval Dungeness crab in the water column are planktivorous, whereas the juvenile and adult crabs are opportunistic foragers on larger bottom-dwelling organisms. In the San Francisco Estuary, juvenile crabs have been shown to feed on clams, crustaceans, and small fishes (Tasto 1983). In Grays Harbor, Washington, juvenile crab diets consisted primarily of Crangon shrimp, juvenile fish, and bivalves (Gunderson et al. 1990). Comparison to other cancrid crabs, the small chelae of C. magister are better suited for soft-bodied, mobile prey found on sandy bottoms (Oresanz and Gallucci 1988). One study has suggested that size-specific feeding on clams in the laboratory was due to an attempt to minimize handling time of the prey in a competitive situation (Palacios and Armstrong 1990).

The most common predators on juvenile crabs within the San Francisco Estuary include bottom-feeding fishes such as starry flounder, English sole, Pacific tomcod, Pacific staghorn sculpin, white croaker, pile perch, sturgeon, and several elasmobranchs (sharks, skates, and rays) (Reilly 1983a). The principal predator on young-of-the-year Dungeness crab in Gray's Harbor Estuary was found to be the Pacific staghorn sculpin (Fernandez et al. 1993a). In addition, cannibalism is reported to occur among all age groups (Warner 1992).

Distribution

Dungeness crab range from the Aleutian Islands to Santa Barbara, but are rare south of Point Conception (Warner 1992). The pelagic larval forms are found distributed widely in both nearshore and offshore waters, but return to the coast, bays, and estuaries where the juvenile and adult stages are mostly found from the intertidal zone to approximately 300 feet (Hatfield 1983, Reilly 1983b, Warner 1992). San Francisco Bay, as is the case with other coastal estuaries, is an important nursery area for the offshore stock. The vast majority of individuals in the Bay are juveniles of a single year-class, having entered in the spring of one year and exited approximately 1 year later (Tasto 1983, McCabe et al. 1988). Juveniles are often found in tidal and navigational channels early in summer, but spread out over mudflats and into protected shoreline areas as they develop over the year (Figures 2.1 and 2.2).

Population Status and Influencing Factors

Few population estimates have been made on individual Dungeness crab stocks along the Pacific coast because there is significant variation in year-class strength, purportedly due to environmental conditions and density-dependent factors (Botsford and Hobbs 1995). However, commercial crab landings, monitored annually by state and, in some instances, federal resource agencies, appear to be a reliable indicator of relative abundance.

The most important factors affecting overall population numbers in the San Francisco area (i.e., Half Moon Bay to Bodega Bay) include ocean temperatures (hatching success), ocean currents (larval drift), predation, commercial fishing, and, possibly, pollution of nursery habitat (Wild and Tasto 1983). Although labo-
Laboratory results show that cannibalism may be an important determinant in the abundance and structure of some populations (Fernandez et al. 1993b), year-class strength and recruitment to the fishery do not appear to depend upon success of any particular “critical” stage (McConnaughey and Armstrong 1990). Within the San Francisco Estuary, juvenile abundance varies considerably from year to year, but is often highest in San Pablo Bay and lowest in south Bay (Tasto 1983, CDFG 1987).

Figure 2.1 Seasonal Distribution of Juvenile Dungeness Crab Within San Francisco Bay (Tasto 1983)
**Trophic Levels**

Larvae are planktivores making them primary consumers (phytoplankton) and secondary consumers (zooplankton). Juveniles and adults are higher order consumers.

**Proximal Species**

**Predators:** Chinook and coho salmon* (prey on late larval stages); Carcinonemertes errans* (predator worm on egg masses); Dungeness crab (cannibalism by larger instars, principally females, on small juveniles), starry

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* Figure 2.2 Annual Distribution of Juvenile Dungeness Crab Within the San Francisco Bay – Caught by Otter Trawl, May-December (CDFG 1987)
flounder, English sole, Pacific tomcod, Pacific staghorn sculpin, white croaker, brown smoothhound shark, and skate (prey on juveniles); and humans (commercial and recreational fishing for adults*).

* Generally takes place outside of San Francisco Bay.

Prey: Crustaceans, bivalves (clams), small fishes.

**Good Habitat**

Juvenile crabs appear to prefer sandy or sandy-mud substrate, but can be found on almost any bottom type (e.g., shell debris). Structurally complex habitats that provide protection from predation (e.g., high relief shell, eel grass, drift macroalgae, etc.) are favored over bare mud or open sand (Fernandez et al. 1993a, Iribarne et al. 1995, Eggleston and Armstrong 1995, McMillan et al. 1995).

Chemical and physical characteristics of the water column and sediment are also important habitat features. Juvenile Dungeness crab in the San Francisco Estuary seem to be somewhat intolerant of salinities lower than 10 ppt (Tasto 1983, CDFG 1987). Maximum growth appears to occur at 15°C or above (Kondzela and Shirley 1993, McMillan et al. 1995); and studies in Washington State have shown that juvenile crab have stable metabolic rates at elevated estuarine temperatures (e.g., 14 to 16°C), whereas older crabs were more stable at colder temperatures (Gutermuth and Armstrong 1989). This is consistent with the tendency for juvenile crabs to emigrate out of estuaries into colder coastal waters as they approach sexual maturity.

Although no single pollutant, or suite of pollutants, has been shown to significantly affect Dungeness crab, various studies on different life stages have shown sensitivity to oiled sediments, dissolved oxygen levels below 5 ppm, low ammonia concentrations, pesticides, and chlorinated wastewater (Wild and Tasto 1983, Emmett et al. 1991). Juvenile crab abundance in the Bay has been shown to be negatively correlated to Delta outflow (CDFG 1987).

**References**


rock crabs (Cancer antennarius and Cancer productus)
Robert N. Tasto

General Information

The brown rock crab (Cancer antennarius) is found along the west coast of North America from Washington State to Baja California; the red rock crab (Cancer productus) has a slightly more northerly distribution, i.e., Alaska to San Diego (Carroll and Wimn 1989). A small recreational fishery exists for brown and red rock crabs in central San Francisco Bay and parts of south Bay and San Pablo Bay. Most rock crabs in this fishery are caught from piers and jetties by a variety of baited hoop nets and traps. A modest commercial fishery also occurs throughout California waters, with the vast majority of the catch taking place from Morro Bay southward (Parker 1992). Ex-vessel value for the commercial fishery approached $2 million in the mid-1980s (Carroll and Wimn 1989) and appears to be unchanged since then. Unlike their close relative, the Dungeness crab, which has a significant amount of muscle tissue in the body, rock crabs, generally, have been sought after for their claws only. In recent years, however, live whole crabs have become a larger part of the retail market. California Department of Fish and Game regulations prohibit the commercial take of crabs less than 4.25 inches carapace width (CW), require that sport-caught crabs must be 4.0 inches CW or greater, and impose a bag limit of 35 crabs per day.

Reproduction

Mating takes place between a soft-shelled (recently molted) female and hard-shelled male. Male brown rock crabs have been reported to outnumber females by a ratio of 1.6/1 (San Mateo County coast) during all seasons (Breen 1988), although studies by Carroll (1982) at Diablo Cove showed that females were more abundant in the fall, with no other seasonal trends for either sex. Unfertilized eggs remain within the female for approximately three months, following mating, and then are fertilized by the stored sperm as they are released (Parker 1992). The fertilized eggs are then carried until hatching (6 to 8 weeks) in a sponge-like mass beneath the female's abdominal flap (Parker 1992). Female body size is the principal determinant of reproductive output and fecundity, with red rock crab having 172,600 to 597,100 eggs per brood and brown rock crab having 156,400 to 5,372,000 eggs per brood (Hines 1991). Like the Dungeness crab, ovigerous female rock crabs have been observed buried in the sand at the base of rocks in shallow waters protecting their eggs (Reilly 1987). Also, some red rock crab females have been detected emigrating out of the intertidal area.
of estuaries prior to spawning to avoid osmotic stress (Oresanz and Gallucci 1988).

Hatching takes place in spring and early summer in central California (Carroll 1982). The planktonic larvae then settle to the bottom before beginning the juvenile stage. Juvenile abundance is highest in San Francisco Bay during the summer months (CDFG 1987). Much like other cancrids, larval release in spring coincides with peak plankton production, and settlement in the summer is optimal for growth (Hines 1991). The reproductive life span for the red rock crab is approximately four years with four broods, and for the brown rock crab it is approximately seven years with up to 10 broods (Hines 1991).

**Growth and Development**

Brown rock crabs are known to go through 10 to 12 molts before reaching sexual maturity at about 3 inches cw, and will likely molt one to two times per year thereafter (Parker 1992). The average number of red rock crab instars is 13 over the total life span (Oresanz and Gallucci 1988). Studies in Humboldt Bay (O’Toole 1985) found ovigerous red rock crab as small as 3.7 inches cw. Brown rock crabs have reached a maximum 6.5 inches cw and red rock crabs, the larger of the two species, at 8 inches cw (Carroll and Winn 1989, Parker 1992). Maximum life span of the brown rock crab has been estimated at 5-6 years (Carroll 1982).

**Food and Feeding**

Rock crabs are both nocturnal predators and scavengers and have been shown to feed upon hard-shelled organisms such as clams, snails, and barnacles (Parker 1992). The large chelae of these crabs is well-suited to forage on the hard shells of more sedentary prey of their rocky habitats (Oresanz and Gallucci 1988). Red rock crab feed upon intertidal mussels and barnacles (Robles et al. 1989). Juvenile rock crabs are preyed upon by other macroinvertebrates and demersal fishes, whereas adults are prey items for marine mammals (Carroll 1982). Very little is known about the specific food habits of, or predators upon, these two species of rock crabs within San Francisco Bay; however, the sportfishery within the Bay accounts for the loss of an indeterminate number of adult crabs.
Distributions

Both rock crab species inhabit the low intertidal zone to depths of 300 feet or more (Parker 1992) and, although their microhabitat utilization patterns are similar, they appear to be different in how they utilize estuaries (Oresanz and Gallucci 1988). The brown rock crab is principally a marine species and does not osmoregulate well in brackish waters, whereas the red rock crab can successfully inhabit brackish areas. All stages of the red and brown rock crab have been collected in San Francisco Bay, including larvae and ovigerous females (Tables 2.1 and 2.2). Areas of peak abundance appear to be in Central Bay, the northern portion of South Bay, and the southern portion of San Pablo Bay, with the red rock crab having a somewhat greater distribution than the brown rock crab (CDFG 1987). In general, rock crab movement is local (Breen 1988, Carroll and Winn 1989). At Fitzgerald Marine Reserve along the San Mateo County coast, studies demonstrated that juvenile brown rock crab are most abundant in July, although no seasonal trend in the settlement of early instars was evident (Breen 1988). In Santa Barbara County, Reilly (1987) found all stages of rock crabs to be most abundant in the fall.

Population Status and Influencing Factors

There are no known estimates of the overall population size or knowledge of recruitment mechanisms for San Francisco Bay rock crabs. Most studies have shown that population densities of rock crabs were well below 1/m² (Carroll 1982, Breen 1988). Small, local populations of rock crab can be overfished, although there is no evidence suggesting that overfishing occurs in the Bay. Data from the Interagency Ecological Study Program indicate that there is a negative relationship between abundance of both rock crab species and outflow from the Delta (CDFG 1987).

Trophic Levels

Rockcrab larvae are planktivores and, as such, are both primary consumers (phytoplankton) and secondary consumers (zooplankton). Juveniles and adults are higher order consumers.

Proximal Species

Predators: Marine mammals, humans (recreational fishery).
Prey: Bay mussels, barnacles.

Good Habitat

Not surprisingly, both species have been shown to prefer rocky shore, subtidal reef, or coarse gravel and sand substrate (Carroll and Winn 1989). Opportunity for concealment appears to be an important feature of red rock crab habitat in British Columbia studies (Robles et al. 1989). Juvenile brown rock crab, when settling from the last larval stage, appear to accept both sand and rock as suitable substrate (Carroll and Winn 1989), and red rock crab also tend to settle onto structurally complex substrates (Oresanz and Gallucci 1988).

### Table 2.1 Annual Abundance of Rock Crabs Caught by Otter Trawl (crabs/tow) in the San Francisco Estuary (CDFG 1987)

<table>
<thead>
<tr>
<th></th>
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<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>C. antennarius (all sizes)</td>
<td>0.101</td>
<td>0.047</td>
<td>0.010</td>
<td>0.015</td>
<td>0.071</td>
<td>0.033</td>
<td>0.007</td>
</tr>
<tr>
<td>C. antennarius (&lt;50mm)</td>
<td>0.098</td>
<td>0.037</td>
<td>0.005</td>
<td>0.015</td>
<td>0.067</td>
<td>0.024</td>
<td>0.007</td>
</tr>
<tr>
<td>C. gracilis (all sizes)</td>
<td>0.035</td>
<td>0.103</td>
<td>0.044</td>
<td>0.182</td>
<td>0.333</td>
<td>0.240</td>
<td>0.174</td>
</tr>
<tr>
<td>C. gracilis (&lt;20mm)</td>
<td>0.003</td>
<td>0.005</td>
<td>0.034</td>
<td>0.080</td>
<td>0.079</td>
<td>0.064</td>
<td>0.095</td>
</tr>
<tr>
<td>C. productus (all sizes)</td>
<td>0.014</td>
<td>0.032</td>
<td>0.005</td>
<td>0.010</td>
<td>0.055</td>
<td>0.071</td>
<td>0.088</td>
</tr>
<tr>
<td>C. productus (&lt;50mm)</td>
<td>0.014</td>
<td>0.027</td>
<td>0.002</td>
<td>0.005</td>
<td>0.040</td>
<td>0.050</td>
<td>0.081</td>
</tr>
</tbody>
</table>

### Table 2.2 Annual Abundance of Rock Crabs Caught by Ring Net (crabs/tow) in the San Francisco Estuary (CDFG 1987)

<table>
<thead>
<tr>
<th></th>
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</tr>
</thead>
<tbody>
<tr>
<td>C. antennarius (all sizes)</td>
<td>-</td>
<td>-</td>
<td>0.113</td>
<td>0.095</td>
<td>0.296</td>
<td>0.491</td>
<td>0.407</td>
</tr>
<tr>
<td>C. antennarius (&lt;50mm)</td>
<td>-</td>
<td>-</td>
<td>0</td>
<td>0.009</td>
<td>0.028</td>
<td>0.009</td>
<td>0.176</td>
</tr>
<tr>
<td>C. gracilis (all sizes)</td>
<td>-</td>
<td>-</td>
<td>0.014</td>
<td>0.019</td>
<td>0.037</td>
<td>0.009</td>
<td>0.130</td>
</tr>
<tr>
<td>C. productus (all sizes)</td>
<td>-</td>
<td>-</td>
<td>0.155</td>
<td>0.067</td>
<td>2.509</td>
<td>4.315</td>
<td>0.806</td>
</tr>
<tr>
<td>C. productus (&lt;50mm)</td>
<td>-</td>
<td>-</td>
<td>0</td>
<td>0</td>
<td>0.185</td>
<td>0.148</td>
<td>0.157</td>
</tr>
</tbody>
</table>

* Ring net survey started in May 1982
Rock crabs appear to be influenced by both temperature and salinity. In various laboratory studies, both brown and red rock crabs were adversely affected by exposure to water temperatures above 20°C (Carroll and Winn 1989, Sulkin and McKeen 1994). The brown rock crab is considered primarily a marine species, whereas red rock crabs can osmoregulate in more brackish water; although the latter have been shown to be adversely affected by salinities below 13 ppt (Oresanz and Gallucci 1988, Carroll and Winn 1989).

References


a wingspan of six feet (>2000 mm) though individuals this large are uncommon. The largest bat ray reported was a 95 kg female from Newport Bay (Baxter 1980).

**Food and Feeding**

Bat rays are opportunistic bottom feeders, feeding primarily upon benthic and epibenthic invertebrates. In Elkhorn Slough, bat rays feed primarily on clams and the echiuroid worm, *Urechis caupo*; in La Jolla kelp beds, they feed on shellfish including abalone and snails; and in Tomales Bay, they feed on polychaete worms, large clams and echiuroid worms (Karl and Obrebski 1976, Karl 1979, Talent 1982). Studies done along the southern California coast (Van Blaricom 1982) found that pits dug by feeding bat rays were an important controlling factor of infaunal community organization, opening areas for infauna recolonization and uncovering food items for other fish.

**Distribution**

The bat ray ranges from the Gulf of California to Oregon, and is found from shallow subtidal water to 46 m. It is common in bays and shallow sandy areas along the coast (Miller and Lea 1976).

**Population Status and Influencing Factors**

The current status of the bat ray in San Francisco Bay is unknown. Its distribution is likely influenced by salinity, it has occasionally been collected in San Pablo Bay at salinities lower 20 ppt (Flemming 1999).

**Trophic Levels**

Bat rays are primary consumers, feeding primarily on benthic invertebrates. They are taken by fishermen using cut fish as bait, however natural feeding on fishes has not been documented.

**Proximal Species**

**Prey:** Benthic mollusks, polychaetes, crustaceans, *Urechis caupo*.

**Good Habitat**

Sandy to muddy shallow bottoms with abundant mollusk and polychaete populations.

**References**


Leopard Shark
Triakis semifasciata
Michael F. McGowan

General Information
The leopard shark (Family: Elasmobranchs) is one of the most common sharks in California bays and estuaries (Talent 1976). It is the most abundant shark in San Francisco Bay (Ebert 1986) being found especially around piers and jetties (Emmett et al. 1991). The leopard shark is an important recreational species in San Francisco Bay and a limited commercial long-line fishery has targeted it in the bay (Smith and Kato 1979).Juveniles and adults are demersal and sometimes rest on the bottom (Feder et al. 1974). Although other elasmobranchs occur in euhaline bays and estuaries of the U.S. Pacific coast, the leopard shark was the only shark or ray included among 47 fish and invertebrate species in the life history summaries of west coast estuarine species prepared by the National Oceanic and Atmospheric Administration’s Estuarine Living Marine Resources (ELMR) program (Emmett et al. 1991). These species were selected on the basis of commercial value, recreational value, indicator species of environmental stress, and ecological importance. That the leopard shark was selected is an indication of its importance in estuaries in general and in San Francisco Bay where it is the most abundant shark.

Reproduction
The leopard shark is a live bearer with internal fertilization, but no yolk-sac placenta. Mating occurs in the spring, primarily during April and May soon after the females give birth to from 4-29 pups (Compagno 1984). Pupping can occur from March through August with a peak in April or May (Ackerman 1971). In San Francisco Bay leopard sharks pup almost exclusively in South Bay (CDFG Bay Trawl data). The center of abundance of pups <300 mm long is south of, and just north of the Dumbarton Bridge.

Growth and Development
Embryonic development is direct and internal and takes 10-12 months. At birth pups are 18-20 cm long. Females mature when 12-14 years old at a length of 110-129 cm. Males mature earlier and at smaller sizes than females. Growth rates are slow. In San Francisco Bay tagged leopard sharks grew 1.4 cm/yr (Smith and Abramson 1990).

Food and Feeding
Primary foods of the leopard shark are benthic and epibenthic crustaceans, clam siphons, echinuroid worms, and small fishes.

Distribution
The leopard shark is found from Mazatlan, Mexico including the Sea of Cortez to Oregon (Miller and Lea 1976). In California it is most common in estuaries and bays south of Tomales Bay (Monaco et al. 1990). Leopard sharks are apparently resident in San Francisco Bay, although some move out in fall and winter (Smith and Abramson 1990) and several size classes appear in the California Department of Fish and Game length data.

Population Status and Influencing Factors
The leopard shark probably has no predators except larger sharks and humans. Its broad dietary range should protect it from food limitation. Heavy fishing mortality poses a threat to the leopard shark, as it does to all sharks, because of its slow growth, long time to maturity, and low fecundity. The minimum size limit recommended by Smith and Abramson (1990) for sustainable fishing in San Francisco Bay was 100 cm (40 in). Areas of high freshwater input causing low salinity are largely avoided by leopard sharks.

Trophic Levels
Juveniles and adults are secondary and higher carnivores.

Proximal Species
Predators: Larger sharks, humans.
Prey: Yellow shore crab, Urechis caupo, ghost shrimp, rock crabs, octopus, shiner perch, arrow goby, Pacific herring, northern anchovy, topsmelt.
Cohabitors: Smoothhound sharks form mixed schools with leopard sharks.

Good Habitat
Leopard sharks are primarily a marine species which occupies bays and estuaries unless freshwater flows lower salinity excessively. Sandy and muddy bottom areas are preferred, although they may be found near rocky areas and kelp beds along the coast. Estuaries are used as pupping and rearing areas for young sharks. Shallow mud and sand flats are used for foraging during high tide (Compagno 1984).
References


Pacific Herring
Clupea pallasi
Robert N. Tasto

General Information

The Pacific herring (Family: Clupeidae) resource in the San Francisco Estuary is widely recognized for its commercial, recreational, and ecological values. The commercial fishery concentrates on ripe females for their roe (eggs) which are then exported to Japan, although there is some limited effort for the fresh fish market and for live bait by recreational salmon trollers (Spratt 1981, Lassuy 1989). Fishermen traditionally catch herring in nearshore areas of the Bay with gillnets or in deeper waters with round-haul nets, and there also is a relatively new roe-on-kelp fishery operated from rafts (Spratt 1981, CDFG 1992). The economic value of the fishery based upon ex-vessel prices paid to the fishermen in 1995-96 was approximately 16.5 million dollars (CDFG, unpub. data).

Reproduction

Adult herring congregate outside of San Francisco Bay before entering and generally spend about 2 weeks in the Bay before spawning (CDFG 1987). Spawning takes place from early November through March, with peak activity in January (Spratt 1981, CDFG 1992). The timing of spawning is believed to coincide with increased levels of plankton production as a food source for larvae (Lassuy 1989), as well as the presence of freshwater flows (Emmett et al. 1991). Pacific herring spawn primarily on vegetation, rock rip-rap, pier pilings, and other hard substrates in intertidal and shallow subtidal waters (Spratt 1981, Lassuy 1989, Emmett et al. 1991). Spawning occurs in waves of 1 to 3 days, occasionally up to a week in length, and often at night in conjunction with high tides (Spratt 1981). Waves are separated by one to several weeks over the length of the season with larger fish tending to spawn first (Lassuy 1989). The number and size of the waves is related to the distribution of the dominant year classes (CDFG 1992).

Egg-deposition is thought to be facilitated by the brushing of the female's vent up against the substrate, and, while there is no pairing of the sexes, the spawning area will be white with milt from the males so that the rate of fertilization is usually high (Hart 1973). Pa-
Pacific herring eggs adhere to the substrate in amounts ranging from a few eggs to as many as eight layers thick (Spratt 1981). The fecundity of herring is approximately 4,000 to 134,000 eggs per female, depending upon its distribution and size (Hart 1973, Emmett et al. 1991). As with spawning, most hatching takes place at night, and will occur in 10 to 15 days under 8.5° to 10.7°C temperatures; longer if the water is colder (Emmett et al. 1991). The average in San Francisco Bay is 10.5 days at 10.0°C (CDFG 1992).

Growth and Development

Pacific herring eggs are approximately 1.0 mm in diameter, and 1.2 to 1.5 mm after fertilization (Hart 1973). A newly hatched larva, with yolk sac, is about 6 to 8 mm total length (TL) and will develop swimming powers at about 20 mm TL (CDFG 1992). Metamorphosis to the juvenile stage occurs from 25 to 35 mm TL and takes place over two to three months (Emmett et al. 1991). They are free-swimming at this stage and begin to form shoreline-oriented schools (CDFG 1992). Juveniles are 35 to 150 mm TL depending upon regional growth rates, which in turn are affected by population size and environmental conditions (Emmett et al. 1991). In the Bay Area, there are no apparent differences in the growth rates of males and females (Spratt 1981). Adults range in size from 130 to 260 mm TL, and locally it takes two to three years to reach maturity (Spratt 1981, Emmett et al. 1991). The San Francisco Bay population ranges from 110 to 250 mm TL (CDFG 1992; Ken Ota, Pers. Comm.) It is possible that some Pacific herring in more northern climates may exceed 15 years in age, but few have been noted to live longer than nine years (Emmett et al. 1991).

Food and Feeding

Pacific herring larvae, juveniles, and adults are selective pelagic planktonic feeders and move toward the water’s surface to feed at dusk and dawn (Emmett et al. 1991). Generally, prey items will change with growth and geographic distribution. Larvae feed on diatoms, invertebrate and fish eggs, crustacean and mollusc larvae, bryozoans, rotifers, and copepods (Hart 1973). Juveniles consume a variety of crustaceans, as well as mollusc and fish larvae; while adults eat mostly planktonic crustaceans and fish larvae (Hart 1973, Emmett et al. 1991). In winter, there is an overall reduction in adult Pacific herring feeding as stored energy is used for ripening reproductive products and, during their spawning migration and inshore “holding” period, herring may severely limit or stop feeding entirely (Lassuy 1989).

Herring eggs are eaten by various species of fish (e.g., sturgeon), ducks (e.g., surf scoter), and gulls (CDFG 1992). Larvae are often prey for large pelagic invertebrates and various fishes, while juveniles and adults are consumed by a variety of fishes (e.g., spiny dogfish shark, Chinook salmon, Pacific staghorn sculpin, and striped bass), seabirds (e.g., Brandts cormorants, brown pelicans, and western gulls), and marine mammals, such as harbor seals (Hart 1973, Lassuy 1989, Emmett et al. 1991). Predation is considered to be the greatest source of natural mortality for juvenile and adult Pacific herring (CDFG 1992).

Distribution

Major populations exist in the eastern Pacific between San Francisco Bay and central Alaska (Hart 1973). Within San Francisco Bay, the principal spawning areas are found along the Marin County coastline (i.e., Sausalito, Tiburon Peninsula, and Angel Island), at the San Francisco waterfront and Treasure Island, on the east side of the Bay from the Port of Richmond to the Naval Air Station at Alameda, and on beds of vegetation in Richardson Bay and South Bay (Figure 2.3) (Spratt 1981, CDFG 1992). After hatching, the larvae are clumped and controlled largely by tidal factors, and following disappearance of the yolk sac and the onset of feeding, their distribution becomes patchy (CDFG 1992). Larvae and young juveniles are found in the Bay between November and April and their greatest densities are in the shallow waters of upper South Bay, Central Bay, and San Pablo Bay. Juveniles are found in the deeper areas of the Bay (peak in Central Bay) between April and August, and, for the most part, have left the Bay by late June at sizes that approach 80 mm TL (CDFG 1987). They eventually move to offshore or nearshore areas and do not return to the Bay until they are mature and ready for spawning. There is conflicting evidence of a strong correlation between juvenile abundance, as measured by young-of-the-year surveys, and recruitment to the adult spawning population two years later (Herbold et al. 1992).

Population Status and Influencing Factors

San Francisco Bay population levels fluctuate widely and have ranged between approximately 6,000 tons and 100,000 tons spawning biomass, as measured by spawn deposition surveys and hydroacoustic monitoring of fish schools (CDFG 1992). 1995-96 season estimates were approximately 99,000 tons, second highest on record (CDFG, unpub. data). Year-class strength is often determined in the first six months of life (Hart 1973, Lassuy 1989, Emmett et al. 1991). Egg mortalities can result from tidal exposure and desiccation, abrupt or severe temperature or salinity changes, low oxygen levels, wave action, suffocation by high egg densities or siltation, pollution, and predation (Lassuy 1989, Emmett et al. 1991). Factors related to natural mortality of larvae
in the Bay include competition and other density dependent mechanisms, as well as starvation during their initial feeding period and changes in dispersal patterns. Juveniles and adult survival is affected by competition, predation, disease, spawning stress, and fishing (Emmett et al. 1991).

Predation appears to be the single most important factor affecting population levels (Lassuy 1989). In addition to commercial and recreational fishing, humans influence herring survival by impacting water and habitat quality. Spawning habitat quantity and Delta outflows are not thought currently to be limiting factors in determining the Bay’s herring population size (CDFG 1987 and 1992).

**Trophic Levels**

Larvae are planktivores (primary and secondary consumers). Juveniles and adults are primary and higher order consumers.

**Proximal Species**

**Egg Predators:** Gulls, diving ducks, white sturgeon, atherinids (topsmelt and jacksmelt), surf perches, rock crabs

**Larvae predators:** Young salmonids, pelagic invertebrates.

**Juvenile Predators:** California halibut, young salmonids, harbor seals, harbor porpoise.

**Adult Predators:** California halibut, California sea lion, harbor seals, harbor porpoise.

**Habitat:** Eel grass (spawning substrate).

**Prey:** Striped bass, copepods.

**Good Habitat**

It is frequently stated that herring prefer sea grasses (e.g., Zostera marina) or algae (e.g., Gracilaria sp.) as spawning substrate (Lassuy 1989, Emmett et al. 1991); however, a variety of seemingly less attractive surfaces have proven to be very successful in the Estuary. Rigidity, smooth texture, and the absence of sediment appear to be important components of suitable substrates (Lassuy 1989). Larvae and juveniles need quiescent and productive shallow subtidal areas as rearing habitat.

In northern waters, the optimal salinity range for spawning is reported to be 8 to 22 ppt and 13 to 19 ppt for eggs and larval survival (CDFG 1987). Also in these areas, temperatures in the range of 5.5 to 8.7° C have
been shown to be best for egg development (Emmett et al. 1991); however, 10 to 12°C temperatures are about average for the spawning grounds in San Francisco Bay (Lassuy 1989). Optimal temperatures for juveniles and adults appear to be a few degrees higher than for eggs or larvae (Lassuy 1989). It has been suggested that eggs need a minimum dissolved oxygen concentration of 2.5 mg/L at the surface and, therefore, eggs elevated from the bottom on vegetation or other structures avoid siltation and receive better circulation for waste removal and oxygenation (Lassuy 1989). Water quality is an important factor as eggs are vulnerable to high levels of suspended particulate matter, particularly if the sediments are laden with contaminants (e.g., dredged material from urban ports). Additionally, larvae have been shown to be sensitive to the water-soluble fraction of hydrocarbons from spilled oil or other sources (Lassuy 1989).

References


Personal Communications

Growth and Development

Larvae grow from 2.5 to 25 mm in about two months, at which time they are considered juveniles. Growth is rapid within the warm productive bay environment. Based on analysis of length frequencies, some juveniles that were spawned late in the summer overwinter in the bay (McGowan 1986). The others apparently depart at the same time as the adults in autumn.

Food and Feeding

Larvae eat dinoflagellates and zooplankton, while adults filter-feed in dense patches of large phytoplankton or small zooplankton, but selectively pick larger zooplankters from the water (O’Connell 1972).

Distribution

The northern anchovy occurs from Queen Charlotte Islands, Canada to Cabo San Lucas, Baja California and into the Sea of Cortez. It can be found in all estuaries within this range. There is a subpopulation which occupies the Columbia River plume, an “offshore estuary.” In San Francisco Bay, they occur from Suisun Bay to South Bay, but are most abundant downstream of the Carquinez Strait (Herbold et al. 1992). There is a seasonal influx of northern anchovy into the bay in spring when water temperatures and plankton production begin to rise in the bay and when nearshore upwelling generally begins. Adults exit the bay in autumn, but some late-spawned juveniles may overwinter within the bay.

Population Status and Influencing Factors

Northern anchovy populations off California range in the hundreds of thousands of tons. Their biomass increased dramatically following the decline of the sardine stock, suggesting that competitive interactions might control population fluctuations. Historical records of fish scales in sediments suggests that large fluctuations in both anchovy and sardine populations have occurred in the past and were not strongly correlated with each other.

Variable survival of eggs and larvae due to environmental factors probably influences population size more than predation or fishing. Active research into the causes of northern anchovy population dynamics has contributed immensely to our understanding but without resolving whether starvation, predation, advection, or other cause is the key limiting factor.

Trophic Levels

First-feeding larvae may eat phytoplankters, larger larvae selectively pick copepods and other zooplankters from the water, juveniles and adults pick or filter plankton, fish eggs, and fish larvae, depending on food concentrations. Larvae and older stages should be considered as secondary and higher consumers.

Proximal Species

Predators: California halibut, Chinook and coho salmon; rockfishes, yellowtail, tunas, sharks, and almost all California current fish; harbor seal; northern fur seal; sea lions; common murre; brown pelican; sooty shearwater; cormorant spp.

Potential Competitors: Sardine, jacksmelt, topsmelt, and other schooling planktivores are potential competitors and predators on young life stages.

Good Habitat

Northern anchovy occupy near surface waters where the water temperature should be between 10° and 25° C. Eggs tend to be in water with salinities from 32-35 ppt, but juveniles and adults are abundant in fresher bays and estuaries as well as marine waters. Spawning in San Francisco Bay occurs at higher temperatures and lower salinities than spawning in coastal areas. Northern anchovy are typical species of areas with high production such as coastal upwelling regions and estuaries.

References


Personal Communications

Sacramento Splittail
Pogonichthys macrolepidotus
Ted R. Sommer

General Information

The Sacramento splittail (Family: Cyprinidae) is one of California's largest native minnows and is the only surviving member of its genus. In 1994 it was proposed for listing as a Threatened species by U.S. Fish and Wildlife Service based on concerns about reduced abundance and distribution (Meng and Kanim 1994, Meng and Moyle 1995). The species supports a small sport fishery in winter and spring, when it is caught for human consumption and live bait for striped bass angling.

Reproduction

Adult splittail generally reach sexual maturity at about 2 years of age (Caywood 1974). Some males mature at the end of their first year and a few females mature in their third year. An upstream spawning migration occurs November through May, with a typical peak from January-March. Spawning is thought to peak during February-June, but may extend from January-July. Although submerged vegetation is thought to be the preferred spawning substrate, egg samples have not yet been collected on any substrate. Reproductive activity appears to be related to inundation of floodplain areas, which provides shallow, submerged vegetation for spawning, rearing and foraging (Caywood 1974, Sommer et al. 1997). Splittail have high fecundity like most cyprinids. Reported fecundities range from 5,000 to 266,000 eggs per female, depending on age (Daniels and Moyle 1983). Generally, female splittail will have more than 100,000 eggs each year.

Growth and Development

The morphological characteristics of splittail eggs, larvae, and juveniles have been described and recent culturing studies (Bailey 1994) are providing preliminary information on early life history requirements and development. Very little is known about factors that influence splittail egg and larval development.

Mature splittail eggs are 1.3 to 1.6 mm in diameter with a smooth, transparent, thick chorion (Wang 1986 cited in CDWR and USBR 1994). The eggs are adhesive or become adhesive soon after contacting water (Bailey 1994). The eggs appear to be demersal and it is assumed that they are laid in clumps and attach to vegetation or other submerged substrates. Under laboratory conditions, fertilized eggs incubated in fresh water at 19°C (±0.5°C) start to hatch after approximately 96 hours. Asynchronous hatching of egg batches from single females has been observed in preliminary culturing tests.

Early hatched larvae are 6 mm long, have not developed eye pigment, and are physically underdeveloped. The last larvae to hatch have developed eye pigmentation and are morphologically better developed. Larvae are 7.0 to 8.0 mm total length (TL) when they complete yolk-sac absorption and become free-swimming; postlarvae are up to 20 mm (±4.2 mm) TL. First scale formation appears at lengths of 22 mm standard length (SL) or 25 mm to 26 mm TL. It is unknown when exogenous feeding actually begins, but preliminary observations indicate that newly hatched larvae may have undeveloped mouths. Well-developed mouths are observed in postlarvae between 8.1 mm and 10.4 mm TL.

Sacramento splittail are a relatively long-lived minnow, reaching ages of 5, and possibly, up to 7 years. Studies from Suisun Marsh indicate that young-of-the-year (YOY) grow approximately 20 mm per month (mm/month) from May through September and then decrease to <5 mm/month through February (Daniels and Moyle 1983). In their second season they grow at about 10 mm/month until the fall when somatic growth declined and gonadal development began. The adult growth rate ranges from 5 to 7 mm/month. During gonad development, which occurs primarily between September and February, the growth rate slows to less than 5 mm/month. The largest recorded splittail measured between 380 mm and 400 mm.

Food and Feeding

Feeding studies describe splittail as opportunistic benthic foragers. Splittail feeding appears highest in the morning and early afternoon. Studies from the Sacramento River found that their diets were dominated by oligochaetes, cladocerans, and dipterans (Caywood 1974). Samples from the lower San Joaquin River included copepods, dipterans, detritus and algae, clams (Corbicula) and amphipods (Corophium spp.). Copepods were the dominant food items. These findings were similar to results of feeding studies from Suisun Marsh (Daniels and Moyle 1983), where the diet consisted predominantly of detritus in both percent frequency of occurrence (74%) and percent volume (57%). A smaller portion of the stomach contents (41% by volume) consisted...
of animal matter, mostly crustaceans (35% by volume). Opossum shrimp (Neomysis mercedis) were the dominant crustacean food item (37% frequency; 59% volume less detritus) both daily and seasonally for splittail in Suisun Marsh. Other minor prey items included molluscs, insects, and fish.

Food selection studies from Suisun Marsh suggest that splittail specifically select Neomysis as their main prey item in the Estuary (Herbold 1987). Fullness indices data indicate that condition factors of splittail are linked to Neomysis abundance. Splittail did not switch to alternate and more prevalent food items, as was observed for other native resident species.

**Distribution**

The historical range of splittail included all low gradient portions of all major tributaries to the Sacramento and San Joaquin rivers, as well as some other freshwater tributaries to San Francisco Bay (Meng and Moyle 1995). A confounding issue is that the collection season and life stage for most of the early observations are unknown, so the relative importance of each location to different age classes of splittail cannot be established.

Splittail are presently most common in the brackish waters of Suisun Bay, Suisun Marsh, and the Sacramento-San Joaquin Delta. The data suggest that splittail inhabit much of their historical range and have been located in previously unreported sites (Table 2.3). Much of the loss of splittail habitat is attributable to migration barriers, but loss of floodplain and wetlands due to diking and draining activities during the past century probably represents the greatest reduction in habitat.

Within the San Francisco Estuary, splittail were collected from southern San Francisco Bay and at the mouth of Coyote Creek in Santa Clara County around the turn of the century. To our knowledge, no other splittail have been collected in this part of San Francisco Bay (Aceituno et al. 1976). However, splittail are caught in San Francisco Bay and San Pablo Bay in wet years. Adults and young are abundant in two tributaries to San Pablo Bay, the Napa and Petaluma rivers. The core of distribution of adult splittail during summer appears to be the region from Suisun Bay to the west Delta. Splittail are also present in some of the smaller tributaries and sloughs of Suisun Bay, including Peyton Slough, Hastings Slough, and Pacheco Creek.

**Population Status and Influencing Factors**

Abundance estimates for YOY and adult splittail were developed recently (Sommer et al. 1997) from several Interagency Ecological Program surveys. The survey equipment for the Program includes otter trawls, midwater trawls, beach seines, and toonets.

Abundance of YOY declined in the Estuary during the six-year drought, which commenced in 1987 (Figure 2.4). There was a strong resurgence in YOY in 1995, when abundance estimates were the highest on record for State Water Project, Central Valley Project, beach seine, Outflow/Bay otter trawl, and Outflow/Bay study midwater trawl. The midwater trawl index was the second highest on record. The response

<table>
<thead>
<tr>
<th>River</th>
<th>Distance (km) from Mouth of River to Collection Site</th>
<th>Distance (km) to first dam</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sacramento</td>
<td>483 (Rutter 1908)</td>
<td>331 (Sommer et al. 1997)</td>
</tr>
<tr>
<td>Feather</td>
<td>109 (Caywood 1974)</td>
<td>94 (Sommer et al. 1997)</td>
</tr>
<tr>
<td>American</td>
<td>49 (Caywood 1974)</td>
<td>19 (Sommer et al. 1997)</td>
</tr>
<tr>
<td>San Joaquin</td>
<td>435 (Caywood 1974)</td>
<td>201 (Sommer et al. 1997)</td>
</tr>
<tr>
<td>Mokelumne</td>
<td>n/a (no records)</td>
<td>63 (Sommer et al. 1997)</td>
</tr>
<tr>
<td>Napa</td>
<td>n/a (no records)</td>
<td>10 (Sommer et al. 1997)</td>
</tr>
<tr>
<td>Petaluma</td>
<td>n/a (no records)</td>
<td>8 (Sommer et al. 1997)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>16 (Sommer et al. 1997)</td>
</tr>
</tbody>
</table>

(a) For the purposes of comparing present and historical distribution, we assumed that collection of any life stage of splittail constituted evidence that a given location was part of the range of the species. The results should be considered as the minimum range only; there had not been sufficient sampling in sites farther upstream to conclusively show that they were not present. To illustrate the fact that much of the loss of channel habitat is attributable to migration barriers, the location of the first dam on each river is included.

(b) Records indicate that splittail were collected, but it is unclear where.

(c) Rutter (1908) was cited by FWS (1994) as the source of an observation of splittail at Fort Miller (km 435), near the current site of Friant Dam on the San Joaquin River. However, Rutter's distribution was based on Girard (1854), who reported two Pogonichthys species, P. symmetricus and P. inaequilobus in the San Joaquin system. P. symmetricus collected from Fort Miller is unlikely to have been a splittail (P. macrolepidotus) because Girard reported the "lobes of the caudal fin are symmetrical." Girard's description of P. inaequilobus had an asymmetrical tail and other features similar to that of splittail, but the collection location is listed as "San Joaquin River" without reference to a specific site.

(d) Dam was removed in 1994.
was not as dramatic for the Suisun Marsh, Chipps Island, or townet surveys, but there was a clear increase in abundance for each relative to the previous nine years.

There appears to be no consistent decline in adult abundance for most of the surveys (Figure 2.5). However, both the Suisun Marsh and Chipps Island surveys show significantly lower abundance in the early to mid-1980s (Sommer et al. 1997).

Floodplain inundation appears to be a key factor responsible for strong year classes, based on both statistical and limited observational data (Sommer et al. 1997). Higher flows increase inundation of floodplain areas, such as the Yolo Bypass, which provides spawning, rearing, and foraging habitat. The species has little or no stock recruitment relationship. This is best illustrated from data collected in 1995, when exceptionally large numbers of young splittail were produced by a stock.
that should have been depleted by drought conditions in seven of the previous eight years.

Attributes that help splittail respond rapidly to improved environmental conditions include a relatively long life span, high reproductive capacity, and broad environmental tolerances (Sommer et al. 1997). Additional factors that may affect population levels include habitat loss, recreational fishing, entrainment, and toxic compounds.

**Trophic Levels**

Splittail are secondary consumers.

**Proximal Species**

**Predators:** Striped bass, centrarchids.

**Prey:** Oligochaetes, zooplankton (cladocerans and copepods), terrestrial insects, opossum shrimp (Neomysis mercedis), mollusks.

**Good Habitat**

Sacramento splittail are one of the few freshwater cyprinids that are highly tolerant of brackish water. Although they have been collected at salinities as high as 18 ppt, abundance is highest in the 0-10 ppt salinity range (Sommer et al. 1997). Physiological studies show that splittail have critical salinity maxima of 20-29 ppt (Young and Cech 1996). Splittail also tolerate a wide range (7-33°C) of water temperatures in the laboratory, which fits well with thermal fluctuations associated with its habitat. Depending upon the acclimation temperature (range 12-20°C), critical thermal
maxima ranged from 22-33°C. As further evidence of the general hardiness of the species, splittail appear to be tolerant of low dissolved oxygen levels and strong water currents.

Splittail are numerous within small dead-end sloughs, those fed by freshwater streams, and in the larger sloughs such as Montezuma and Suisun (Daniels and Moyle 1983). Juveniles and adults utilize shallow edgewater areas lined by emergent aquatic vegetation. Submerged vegetation provides abundant food sources and cover to escape from predators. Shallow, seasonally flooded vegetation is also apparently the preferred spawning habitat of adult splittail (Caywood 1974).

References


Chinook Salmon

Oncorhynchus tshawytscha

Lt. D.ante B. M aragni

General Information

The Chinook salmon (Family: Salmonidae) is morphologically distinguished from other Oncorhynchus species of the northern Pacific Ocean by its large size, small black spots on both caudal fin lobes, black pigment along the base of the teeth (McPhail and Lindsey 1970 as cited in Healey 1991), and varying shades of flesh color from white through shades of pink and red (Healey 1991). The Chinook salmon life history (Figure 2.6) is characterized by adult migration from the ocean to natal freshwater streams to spawn, and juvenile migration seaward as smolts in their first year of life. During the smoltification process, juvenile Chinook salmon undergo physiological, morphological, and behavioral changes that stimulate emigration and prepare them for life in the marine environment (Healey 1991).

The Sacramento-San Joaquin Chinook salmon of California exists as four races—winter, spring, fall, and late-fall—as defined by the timing of adult spawning migration (Mason 1965, Frey 1971, Moyle 1976, Healey 1991). In 1989, the Sacramento River winter-run Chinook salmon was listed as threatened under the federal Endangered Species Act by the National Marine Fisheries Service (NMFS) (54 FR 32085). NMFS reclassified the winter-run as endangered in 1994 (59 FR 440) based on: 1) the continued decline and increased variability of run sizes since its listing as a threatened species in 1989, 2) the expectation of weak returns in certain years as a result of two small year classes (1991 and 1993), and 3) continuing threats to the population. The State of California listed the winter-run as endangered under the California Endangered Species Act in 1989. In 1995, the Oregon Natural Resources Council and R. Nawa petitioned NMFS to list Chinook salmon along the entire West Coast, including the States of California, Idaho, Oregon, and Washington, under the federal Endangered Species Act (54 FR 32085). The State of California presently includes on its list of species of special concern the late-fall (Class 2—special concern) and the spring-run (Class 1—qualified as threatened or endangered) Chinook salmon. Spring-run Chinook salmon
have also been given a special category by the state and are considered a “monitored” species.

Chinook salmon support commercial, recreational, and tribal subsistence fisheries. However, due to the state of Pacific Coast Chinook salmon populations, the U.S. Department of Commerce declared the U.S. Pacific Coast salmon commercial fishery, excluding Alaska, a disaster and has provided emergency relief funding for displaced fisherman in 1995 and 1996 (59 FR 51419, 60 FR 5908). Also, the federal Central Valley Project Improvement Act requires restoration actions to double the Chinook salmon population in the Sacramento-San Joaquin River system in California by the year 2002 estimated from average population levels from 1967 to 1991 (CDFG 1993).

Reproduction

The Chinook salmon is anadromous; that is, it spends most of its adult life in the ocean and returns to freshwater streams to spawn. Chinook salmon typically spend 3-6 years maturing in the ocean before returning as adults to their natal streams to spawn (Moyle 1976, Eschmeyer et al. 1983). Historically, most Sacramento-San Joaquin Chinook salmon returning to spawn have been four years of age (Clark 1929). The Chinook salmon is also semelparous in its reproductive strategy in that it dies after it spawns. Thus, the life span of the Chinook salmon is 3-6 years. All adults die after spawning except some “jacks” (i.e., precocious males that mature early in freshwater) (Miller and Brannon 1982).

Chinook salmon can be grouped into two types based on variations in their life histories: stream-type and ocean-type. Stream-type Chinook salmon populations are most commonly found north of 56°N latitude along the North American coast and characterized by long freshwater residence as juveniles (1+ years). Ocean-type Chinook salmon populations are most commonly found south of 56°N latitude and characterized by short freshwater residence as juveniles (2-3 months). Chinook salmon of the Sacramento-San Joaquin River system are predominantly ocean-type (Healey 1991). Adult upstream migration and juvenile downstream migration of the Sacramento-San Joaquin Chinook salmon differ among the four races. Sacramento-San Joaquin Chinook salmon populations’ migration characteristics are listed in Table 2.4 (Bryant, pers. comm.).

The Chinook salmon normally spawns in larger rivers and tributaries, and typically in deeper water and larger gravel than other Pacific salmon (Scott and Crossman 1973). In preparation for spawning, a female Chinook salmon digs a shallow depression in the gravel of the stream bottom in an area of relatively swift water by performing vigorous swimming movements on her side near the bottom (Emmett et al. 1991, Healey 1991). This depression is referred to as a “redd,” and can be 1.2-10.7 m (3.9-35.1 ft) in diameter (Chapman 1943). The female then deposits a group or “pocket” of eggs in the redd (Emmett et al. 1991, Healey 1991). From 2,000
to 14,000 eggs are laid per female, with 5,000 eggs per female being average (Rounsefell 1957, Moyle 1976, Bell 1984). The eggs are in turn fertilized by one or more males. During spawning, a female will be attended by one dominant male and occasionally other subdominant males. The female then buries the eggs by displacing gravels upstream of the redd (Emmett et al. 1991, Healey 1991).

Growth and Development

Chinook salmon eggs are spherical, non-adhesive, and the largest of all the salmonids (6.0-8.5 mm (0.24-0.33 in) in diameter) (Rounsefell 1957, Scott and Crossman 1973, Wang 1986). The incubation range is from 4-6 weeks, depending on levels of dissolved oxygen, biochemical oxygen demand, water temperature, substrate, channel gradient and configuration, water depth, water velocity and discharge (Reiser and Bjornn 1979, Alaska Department of Fish and Game 1985).

Larval sizes range from 20-35 mm (0.79-1.38 in) in length (Wang 1986). Yolk sac fry, termed "alevins," remain in the gravel from 2-3 weeks until the yolk sac is absorbed (Scott and Crossman 1973, Wydoski and Whitney 1979), whence they emerge from the gravel as fry. Fry develop into parr beginning the smoltification process as they encounter increasing salinities during their migration from freshwater to the ocean. Parr acquire a silver color as they transform into smolts during the smoltification process (Healey 1991). Fry and smolts can stay in freshwater from 1-18 months (Beauchamp et al. 1983), with residency periods differing with race (Table 2.4). Outmigration periods vary with outflow conditions. High outflows will carry fry downstream, while seasons with low outflow cause fry to rear longer in upstream areas where they grow much larger. Juvenile Chinook salmon in these two differing scenarios have substantially different habitat requirements (Kjelson et al. 1982). The fry to smolt life stages' size range is 2-152 cm (0.6-42.9 in), but is usually less than 91 cm (25.7 in), in length (Wydoski and Whitney 1979).

Juvenile Chinook salmon migration into estuaries has been reported to occur at night (Seiler et al. 1981) and during daylight (Dawley et al. 1986). Juveniles may move quickly through estuaries (Dawley et al. 1986) or reside there for up to 189 days (Simenstad et al. 1982). Juvenile Chinook salmon gain significant growth in estuarine habitats as they smolt and prepare for the marine phase of their life (Macdonald et al. 1987). The juveniles of most stocks of Chinook salmon appear to migrate north upon entering the ocean (Wright 1968, Healey 1991). Chinook salmon produced in streams from the Rogue River (Oregon) and south appear to rear in the ocean off northern California-southern Oregon (Cramer 1987). The stream-type Chinook salmon move offshore early in their ocean life, whereas ocean-type Chinook salmon remain in sheltered coastal waters. Stream-type Chinook salmon maintain a more offshore distribution throughout their ocean life than do ocean-type (Healey 1991). Chinook salmon reach maturity in 3-6 years (Moyle 1976).

Food and Feeding

Chinook salmon larvae and alevins feed on their yolk. Chinook salmon juveniles and adults are carnivorous, "opportunistic" feeders, feeding on a variety of terrestrial and aquatic insects, crustaceans, and fish (Emmett et al. 1991).


<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Winter</th>
<th>Spring</th>
<th>Fall</th>
<th>Late-fall</th>
</tr>
</thead>
<tbody>
<tr>
<td>Adult Immigration Period</td>
<td>December - July</td>
<td>March - July</td>
<td>June - December</td>
<td>October - April</td>
</tr>
<tr>
<td>Peak Immigration</td>
<td>March</td>
<td>May - June</td>
<td>September - October</td>
<td>December</td>
</tr>
<tr>
<td>Spawning Period</td>
<td>late April - early August</td>
<td>late August - late October</td>
<td>late September - December</td>
<td>late April</td>
</tr>
<tr>
<td>Peak Spawning</td>
<td>early June</td>
<td>mid September</td>
<td>late October</td>
<td>early February</td>
</tr>
<tr>
<td>Juvenile Emergence Period</td>
<td>July - October</td>
<td>November - March</td>
<td>December - March</td>
<td>April - June</td>
</tr>
<tr>
<td>Freshwater Residency Period</td>
<td>5 - 10 months</td>
<td>3 - 15 months</td>
<td>4 - 7 months</td>
<td>7 - 13 months</td>
</tr>
<tr>
<td>Juv. Emigration Period</td>
<td>July - April</td>
<td>November - January (year 2)</td>
<td>December - June</td>
<td>April - April (year 2)</td>
</tr>
<tr>
<td>Estuarine Residency Period</td>
<td>November - May</td>
<td>March - June</td>
<td>March - July</td>
<td>October - May</td>
</tr>
</tbody>
</table>
In estuaries, juveniles feed in intertidal and subtidal habitats of tidal marshes. In these habitats, juveniles prey upon insects, gammarid amphipods, harpactiid copepods, mysids, chironomids, decapod larvae, and small (larval and juvenile) fish (Levy and Levings 1978, Levy et al. 1979, Northcote et al. 1979, H ealey 1980a, Levy and Northcote 1981, H ealey 1982, Kjelson et al. 1982, Simonstad et al. 1982, Simonstad 1983, McCabe et al. 1986). In low flow years when juveniles are larger, their food source will include crab megalops, squid, and small fish (e.g., northern anchovy, Pacific herring, rockfish) (Beauchamp et al. 1983).


**Distribution**

Chinook salmon eggs and alevins are benthic and infaunal. Fry and parr are benthalpelagic. Parr become pelagic as they enter smoltification. Smolts, ocean-dwelling and maturing juveniles, and adults are pelagic (Alaska Department of Fish and Game 1985). Adults are bottom-oriented in freshwater (Emmett et al. 1991).

Chinook salmon eggs, alevins, fry, and parr occur in riverine areas from just above the intertidal zone to altitudes of 2,268 m (7,441 ft) above sea level (Allen et al. 1991). Smolts are riverine and estuarine. Ocean-dwelling juveniles are neritic and epipelagic, and found within 128 m (420 ft) of the surface (Fredin et al. 1977). Adults may be neritic and estuarine, but are riverine during their spawning migration and may travel upstream more than 4,700 km (2,920 mi) from the ocean (Emmett et al. 1991) as flows and passage allow. Most tributaries are now dammed for water supply, which limits the extent of upstream migration (USFWS 1995).

The Chinook salmon is the least abundant of the major Pacific salmon species (Emmett et al. 1991, Healey 1991). However, it is the most abundant salmon in California (McGinnis 1984). The Chinook salmon is recorded as far north as the Coppermine River in Arctic Canada, and south to northeastern Hokkaido, Japan, and southern California (Ventura River) (H art 1973, Scott and Crossman 1973). It is, however, rarely found in freshwater south of the Sacramento-San Joaquin River system of California (Eschmeyer et al. 1983).

While Chinook salmon are found in all estuaries north of San Francisco Bay in California, except Tomales Bay (Monaco et al. 1990), California’s largest populations of Chinook salmon originate in the Sacramento-San Joaquin River system (Fry 1973). Spring-run Chinook salmon are extinct in the San Joaquin River and only remnant runs remain in a few Sacramento River tributaries. Historically, spring-run Chinook salmon spawned in small tributaries that have essentially all been blocked to migration by large dams. Fall and late-fall Chinook salmon are main stem spawners. Winter-run Chinook salmon are unique to the Sacramento River and spawned in coldwater tributaries above Shasta Dam prior to its construction (Sacramento River Winter-Run Chinook Salmon Recovery Team 1996). While distribution of outmigrating juvenile Chinook salmon is not well known in the San Francisco Bay, they have been found throughout, including the South Bay on high outflow years.

**Population Status and Influencing Factors**

Chinook salmon populations have declined substantially, with winter-run at the point of near extinction and spring-run at severely depressed population levels (Table 2.5). Whereas spring-run historically outnumbered all other runs, fall-run comprises the bulk of the present Chinook salmon population. The remnant “endangered” population of winter-run now depend on cold water releases from Shasta Reservoir, and the protection of the federal Endangered Species Act.

No single impact can be attributed to the decline of Chinook salmon populations and the important Chinook salmon fishery. High mortality for Chinook salmon occurs during the early freshwater life stages (eggs, fry, parr) (Emmett et al. 1991). This mortality is caused by red tide destruction, siltation and destruction of spawning grounds, extremely high or low water temperatures, low dissolved oxygen, loss of cover, disease, food availability and competition, and predation (Reiser and Bjornn 1979). Besides the above factors, human impacts such as river flow reductions, the construction of dams and the consequent creation of reservoirs, water diversions, logging practices, and pollution have affected population abundances (Raymond 1979, Netboy 1980, Stevens and Miller 1983). Factors influencing survival of adult Chinook salmon are equally numerous. In the ocean, Chinook salmon are impacted by oceanographic conditions, disease, food availability and competition, predation, and overfishing (Fraidenburg and Lincoln 1985, Emmett et al. 1991). In freshwater, adults are subject to natural factors such as drought and flood, and human impacts including fishing, dams, road construction and other development, flood protection, dredging, gravel mining, timber harvest, grazing, and pollution (USFWS 1995).
Trophic Levels

Chinook salmon are primary and secondary consumers as juveniles and secondary consumers as adults.

Proximal Species

**Juvenile Predators:** Sacramento squawfish (*Ptychocheilus grandis*), riffle sculpin (*Cottus gulosus*), channel catfish (*Ictalurus punctatus*), steelhead trout (*Oncorhynchus mykiss*), striped bass (*Morone saxatilis*), centrarchids, rockfish (*Sebastes spp.*), kingfishers, egrets, herons, terns, grebes, pelicans.

**Adult Predators:** Pacific lamprey (*Lampetra tridentata*), harbor seal (*Phoca vitulina*), California sea lion (*Callorhinus ursinus*), killer whale (*Orcinus orca*), North American river otter (*Lutra canadensis*), American black bear (*Ursus americanus*), bald eagle (*Haliaeetus leucocephalus*).

**Juvenile Prey:** Terrestrial insects, aquatic insects, chironomids, copepods, amphipods, mysids, euphausiids, decapod larvae, bay shrimp.

**Adult Prey:** Euphausiids, decapods, squid, Pacific herring (*Clupea pallasi*), northern anchovy (*Engraulis mordax*), osmerids, rockfish (*Sebastes spp.*), Pacific sand lance (*Ammodytes hexapterus*).

**Good Habitat**

Chinook salmon eggs develop only in freshwater, but larvae can tolerate salinities of up to 15 ppt at hatching. Three months after hatching juvenile Chinook salmon can tolerate full seawater, with faster growing individuals better able to handle salinity changes (Wagner et al. 1969). Juveniles and adults occur in freshwater to euhaline waters. Successful egg incubation occurs from just above freezing to 20.0°C (68.4° F) (Olsen and Fos-

Table 2.5 Estimated Number of Sacramento-San Joaquin Chinook Salmon Returning to Spawn: 1967-1991 (Mills and Fisher 1994) (Continued on next page.)

<table>
<thead>
<tr>
<th>Year</th>
<th>Sacramento Fall-run Chinook&lt;sup&gt;1&lt;/sup&gt;</th>
<th>San Joaquin Fall-run Chinook&lt;sup&gt;2&lt;/sup&gt;</th>
<th>Sacramento Late-fall-run Chinook&lt;sup&gt;3&lt;/sup&gt;</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>grilse</td>
<td>adult</td>
<td>total</td>
</tr>
<tr>
<td>1967</td>
<td>38,410</td>
<td>104,790</td>
<td>143,200</td>
</tr>
<tr>
<td>1968</td>
<td>18,181</td>
<td>155,859</td>
<td>174,040</td>
</tr>
<tr>
<td>1969</td>
<td>48,528</td>
<td>208,289</td>
<td>256,817</td>
</tr>
<tr>
<td>1970</td>
<td>30,121</td>
<td>147,279</td>
<td>177,400</td>
</tr>
<tr>
<td>1971</td>
<td>35,775</td>
<td>140,691</td>
<td>176,466</td>
</tr>
<tr>
<td>1972</td>
<td>43,795</td>
<td>80,622</td>
<td>124,417</td>
</tr>
<tr>
<td>1973</td>
<td>40,640</td>
<td>197,193</td>
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<td>1974</td>
<td>25,364</td>
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<tr>
<td>1975</td>
<td>29,691</td>
<td>141,884</td>
<td>171,575</td>
</tr>
<tr>
<td>1976</td>
<td>21,926</td>
<td>155,767</td>
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<td>1980</td>
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<td>43,396</td>
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<td>172,784</td>
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<td>1983</td>
<td>41,714</td>
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<td>41,563</td>
<td>211,695</td>
<td>253,258</td>
</tr>
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<td>1986</td>
<td>27,356</td>
<td>212,739</td>
<td>240,095</td>
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<td>1987</td>
<td>66,364</td>
<td>150,965</td>
<td>217,329</td>
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<tr>
<td>1988</td>
<td>26,517</td>
<td>197,841</td>
<td>224,358</td>
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<tr>
<td>1989</td>
<td>26,040</td>
<td>116,726</td>
<td>142,766</td>
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<tr>
<td>1990</td>
<td>9,443</td>
<td>83,499</td>
<td>92,942</td>
</tr>
<tr>
<td>1991</td>
<td>11,546</td>
<td>87,070</td>
<td>98,616</td>
</tr>
<tr>
<td>AVERAGE</td>
<td>33,053</td>
<td>143,083</td>
<td>176,137</td>
</tr>
</tbody>
</table>

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1 Escapement data for the Sacramento River and its tributaries north of and including the American River.
2 Escapement data for the Mokelumne, Cosumnes, Calaveras, Stanislaus, Tuolumne and Merced rivers.
3 Escapement data for the main stem Sacramento River above Red Bluff Diversion Dam.
ter 1955), however, best incubation temperatures are 5.0-14.4°C (41.0-57.9°F) (Bell 1984). The upper lethal temperature for Chinook salmon is 25.1°C (77.2°F) (Brett 1952), but may be lower depending on other water quality factors (Ebel et al. 1971). Eggs and alevins are found in areas with flow of 20-150 cm/sec (0.7-5 ft/sec) and juveniles where flows are 0.5-60.0 cm/sec (0.02-2 ft/sec) (at pool edges). Adults can migrate upstream in flows up to 2.44 m/sec (8 ft/sec) (Thompson 1972). Successful egg development requires redds to have adequate dissolved oxygen (>5.0 mg/L), water temperatures (4-14°C [39-57°F]), substrate permeability, sediment composition (<25% fines, <6.4 mm [0.25 in] in diameter), surface flows and velocities, and low biochemical oxygen demand (Reiser and Bjornn 1979).

Juveniles in freshwater avoid waters with <4.5 mg/L dissolved oxygen at 20°C (68°F) (Whitmore et al. 1960). Migrating adults will pass through water with dissolved oxygen levels as low as 5 mg/L (Hallow et al. 1970). Excessive silt loads (>4,000 mg/L) may halt Chinook salmon movements or migrations. Silt can also hinder fry emergence, and limit benthic invertebrate (food) production (Reiser and Bjornn 1979). Freshwater inflow into estuaries is critical for providing adequate water temperatures, food production, and overall beneficial environmental conditions for juvenile outmigration. High freshwater flows allow for cooler water temperatures, while also stimulating and sustaining production of food. High river flows improve juvenile survival and enable active migration into estuaries and on to the ocean.

In addition to specific hydrologic components, physical habitat requirements of interrelated instream gravel, riparian, and tidal marsh habitats comprise the healthy ecosystem in which Chinook salmon spawn and rear. Chinook salmon eggs and alevins require clean, loose gravel and occur in spawning gravel or cobble that

### Table 2.5 (continued) Estimated Number of Sacramento-San Joaquin Chinook Salmon Returning to Spawn: 1967-1991 (Mills and Fisher 1994)

| YEAR | Sacramento Spring-run Chinook
girlse | adult | total | Sacramento Winter-run Chinook
girlse | adult | total | Central Valley Total Chinook Salmon
girlse | adult | total |
<table>
<thead>
<tr>
<th></th>
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<th></th>
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<td>12,297</td>
<td>23,694</td>
<td>24,985</td>
<td>32,321</td>
<td>57,306</td>
<td>81,698</td>
<td>202,245</td>
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<td>15,144</td>
<td>10,299</td>
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<td>84,414</td>
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<td>27,335</td>
<td>8,953</td>
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<td>40,409</td>
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<td>7,437</td>
<td>8,324</td>
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<td>40,409</td>
<td>50,228</td>
<td>231,297</td>
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<tr>
<td>1971</td>
<td>2,464</td>
<td>6,336</td>
<td>8,800</td>
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<td>32,225</td>
<td>53,089</td>
<td>64,366</td>
<td>229,878</td>
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<td>1,343</td>
<td>7,053</td>
<td>8,396</td>
<td>8,541</td>
<td>28,592</td>
<td>37,133</td>
<td>58,531</td>
<td>157,743</td>
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<td>1973</td>
<td>2,082</td>
<td>9,680</td>
<td>11,762</td>
<td>4,623</td>
<td>19,456</td>
<td>24,079</td>
<td>48,729</td>
<td>254,261</td>
<td>302,990</td>
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<tr>
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<td>2,538</td>
<td>5,545</td>
<td>8,083</td>
<td>3,788</td>
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<td>21,897</td>
<td>32,782</td>
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<td>4,067</td>
<td>22,006</td>
<td>26,073</td>
<td>8,634</td>
<td>26,462</td>
<td>35,096</td>
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<td>8,409</td>
<td>13,830</td>
<td>2,186</td>
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<td>17,214</td>
<td>31,372</td>
<td>172,614</td>
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<td>1,093</td>
<td>7,063</td>
<td>8,156</td>
<td>1,193</td>
<td>23,669</td>
<td>24,862</td>
<td>27,134</td>
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<td>1979</td>
<td>707</td>
<td>2,203</td>
<td>2,910</td>
<td>113</td>
<td>2,251</td>
<td>2,364</td>
<td>47,717</td>
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<td>171,746</td>
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<td>1980</td>
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<td>11,815</td>
<td>1,072</td>
<td>84</td>
<td>1,156</td>
<td>31,545</td>
<td>131,780</td>
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<td>1981</td>
<td>8,249</td>
<td>13,066</td>
<td>21,315</td>
<td>1,744</td>
<td>18,297</td>
<td>20,041</td>
<td>60,757</td>
<td>198,591</td>
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<td>1982</td>
<td>4,528</td>
<td>21,644</td>
<td>26,172</td>
<td>270</td>
<td>972</td>
<td>1,242</td>
<td>51,061</td>
<td>167,947</td>
<td>219,008</td>
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<td>1983</td>
<td>672</td>
<td>3,809</td>
<td>4,481</td>
<td>392</td>
<td>1,439</td>
<td>1,831</td>
<td>76,311</td>
<td>116,947</td>
<td>193,258</td>
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<td>4,373</td>
<td>3,988</td>
<td>8,361</td>
<td>1,869</td>
<td>794</td>
<td>2,663</td>
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<td>17,290</td>
<td>18,896</td>
<td>451</td>
<td>2,013</td>
<td>2,464</td>
<td>33,020</td>
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<td>7,330</td>
<td>11,507</td>
<td>236</td>
<td>1,761</td>
<td>1,997</td>
<td>89,826</td>
<td>180,408</td>
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<td>9,521</td>
<td>11,653</td>
<td>708</td>
<td>1,386</td>
<td>2,094</td>
<td>32,589</td>
<td>238,136</td>
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<td>6,304</td>
<td>7,188</td>
<td>53</td>
<td>480</td>
<td>533</td>
<td>25,566</td>
<td>136,587</td>
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<td>1990</td>
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<td>425</td>
<td>441</td>
<td>12,256</td>
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<td>106,603</td>
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<td>1991</td>
<td>433</td>
<td>1,208</td>
<td>1,641</td>
<td>38</td>
<td>153</td>
<td>191</td>
<td>9,112</td>
<td>94,721</td>
<td>107,833</td>
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<td>AVERAGE</td>
<td>3,276</td>
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<td>12,990</td>
<td>4,687</td>
<td>18,421</td>
<td>23,109</td>
<td>47,181</td>
<td>199,955</td>
<td>247,136</td>
</tr>
</tbody>
</table>

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1 Escapement data for the main stem Sacramento River above Red Bluff Diversion Dam.
2 Escapement data for the main stem Sacramento River above Red Bluff Diversion Dam.
is 1.3-10.2 cm (0.5-10.2 in) in diameter (Reiser and Bjornn 1979). Juveniles in freshwater are found within areas of shallow riffles and deep pools over various substrates, ranging from silt bottoms to large boulders (Chapman and Bjornn 1968). Juveniles in estuaries occur in intertidal and tidal habitats over mud, sand, gravel, and eelgrass (Zostera spp.) (Healey 1980a). Adults in marine waters show no sediment preference, but may be associated with gravel-cobble bottoms in rivers and streams during upstream migration (Alaska Department of Fish and Game 1985).

In riverine areas, both submerged cover, such as boulders, woody debris, and aquatic vegetation, and overhead cover, such as continuous riparian vegetation canopies, undercut banks, and turbulent water, provide shade, food, and protection against predation to juvenile Chinook salmon. Estuaries appear to play a vital role in Chinook salmon life history as well, and specifically, tidal marsh habitat is of great importance to juvenile salmonids (Dorcey et al. 1978, Levy et al. 1979, M eyer 1979, Levy and Northcote 1981, Healey 1982, McD onald et al. 1987, 1988). Juvenile Chinook salmon forage in the intertidal and shallow subtidal areas of tidal marsh mudflat, slough, and channel habitats, and open bay habitats of eelgrass and shallow sand shoal areas. These productive habitats provide both a rich food supply and protective cover within shallow turbid waters (McDonald 1960; D unford 1975, cited from Cannon 1991). The distribution of juvenile Chinook salmon changes tidally, with fry moving from tidal channels during flood tides to feed in nearshore marshes.

Tidal marshes are most heavily used by fry, whereas smolts tend to utilize deeper waters. Fry disperse along the edges of marshes at the highest points reached by the tide, then retreat into the tidal channels with the receding tide. Smolts congregate in surface waters of main and secondary sloughs and move into shallow subtidal areas to feed (Levy and Northcote 1981, Levings 1982, Allen and H assler 1986, Healey 1991).

In addition to good water quality, adequate flows, and productive spawning and rearing habitat, state-of-the-art barrier screens on water diversions, protection from excessive harvest, and free access to upstream migration or well-designed ladders for adult passage offer promising overall habitat for healthy Chinook salmon populations.

References


Chapter 2 — Estuarine Fish and Associated Invertebrates
west Forest Range Experimental Station, Portland, Oregon.

Personal Communications

Greg Bryant, Fisheries Biologist, National Marine Fisheries Service, Southwest Region, Protected Species Division, Eureka, California.
Steelhead
Oncorhynchus mykiss irideus
Robert A. Leidy

General Information
Steelhead (Family: Salmonidae) are the anadromous (sea-run) form of resident rainbow trout. Behnke (1992) proposed classification of steelhead on the west coast of the United States into a coastal subspecies, *O. m. irideus*, and an inland subspecies, *O. m. gairdneri*. California is considered to have only coastal steelhead (Behnke 1992). In California steelhead may be classified into two races, summer and winter steelhead, based upon the timing of upstream migration into freshwater. The San Francisco Estuary and its tributary streams support winter steelhead. Steelhead are a polymorphic species and as such populations within a stream may be anadromous, resident, or mixtures of the two forms that presumably interbreed (Titus et al., in press). Steelhead do not support a commercial fishery within the San Francisco Estuary and its tributaries. It is illegal for commercial salmon trollers to possess steelhead (McEwan and Jackson 1996). There is an inland recreational sportfishery for steelhead that is dependent largely on hatchery operations to sustain populations. The estimated net annual economic benefit of doubling steelhead stocks within the Sacramento/San Joaquin river systems is estimated at 8.0 million dollars (Meyer Resources Inc. 1988).

Reproduction
Polymorphic salmonids exhibit a high degree of life history variation (Titus et al., in press). Steelhead within the San Francisco Estuary may be classified as “ocean-maturing” or “winter” steelhead that typically begin their spawning migration in the fall and winter, and spawn within a few weeks to a few months from when they enter freshwater (McEwan and Jackson 1996). Ocean-maturing steelhead enter freshwater with well-developed gonads and spawn shortly after entering a river or stream. Steelhead begin upstream migration after one to four growing seasons at sea (Burgner et al. 1992). A small number of immature fish (i.e., grilse) may also move upstream after spending only a few months in the ocean. Releases of cold water from several large Central Valley reservoirs on the Sacramento River system may induce steelhead to begin to move into upstream tributaries as early as August and September. This means that upstream migrating steelhead may be observed within San Francisco Bay and Suisun Marsh/Bay between August and March. Ocean-maturing steelhead typically spawn between December and April, with most spawning occurring between January through March.

Steelhead are iteroparous and do not die after spawning as do other Pacific salmon; therefore, they may return to the ocean and spawn again the following year. The frequency of return spawning for a given population is generally unknown. Steelhead spawn in redds constructed by the female over a gravel/cobble substrate. Eggs are deposited in the redd and then fertilized by the male. The number of eggs produced is largely a function of the size of the female, and may range from 200 to 12,000 eggs over the geographic range of steelhead (Scott and Crossman 1973, Moyle 1976). Steelhead within the Sacramento River drainage average between 1,000 to 4,500 eggs (Mills and Fisher 1994).

Growth and Development
Steelhead eggs are spherical to slightly irregular in shape, non-adhesive, demersal, and range in diameter from 3-6 mm (Wang 1986). Incubation of eggs is dependent upon water temperature in the redd. Wales (1941) observed hatching at approximately 19 days at an average water temperature of 15.5° C and 80 days at about 4.5° C. For Waddell Creek in coastal San Mateo County, steelhead hatching time was estimated at 25 to 35 days, with emergence beginning at 2-3 weeks following hatching (Shapovalov and Taft 1954). Steelhead length at hatching ranges between 14 to 15.5 mm total length (TL), with alevis ranging between 23-26 mm TL (Wang 1986). Alevis emerge from the gravel following yolk sac absorption as fry or juveniles ready to actively feed.

Steelhead remain in freshwater for one to four years (usually two years) before downstream migration as “smolts”, at an average size ranging between 13 cm and 25 cm TL (Moyle 1976). Age at emigration is highly variable, but may occur earlier in warmer, more productive streams where juveniles can reach smolt size at a younger age (Moyle et al. 1995). Most Sacramento River juvenile steelhead emigrate as 1-year-old fish during spring and early summer (Barnhart 1986, Reynolds et al. 1993), although Shapovalov and Taft (1954) found that steelhead moved downstream in Waddell Creek during all months of the year. While steelhead may spend up to four years in the ocean, most only survive to age two. In the ocean steelhead may grow at a rate of 1.2 inches per month and reach a length of 23 inches in two years.
Food and Feeding

Rearing juvenile steelhead are primarily drift feeders utilizing a variety of terrestrial and aquatic insects, including emergent aquatic insects, aquatic insect larvae, snails, amphipods, opossum shrimp, and various species of small fish (Moyle 1976). Larger steelhead will feed on newly emergent steelhead fry. Emigrating adult and juvenile steelhead may forage in the open water of estuarine subtidal and riverine tidal wetland habitats within the Estuary, although the importance of these areas as rearing habitat for juveniles is not well documented. Apparently upstream migrating steelhead rarely eat and therefore exhibit reduced growth (Pauley and Bortz 1986).

Distribution

Steelhead populations are native to Pacific Ocean coastal drainages of the Kamchatka Peninsula and scattered mainland locations of Asia and in the western Pacific from the Kuskokwim River in Alaska to Malibu Creek in southern California (Titus et al., in press, McEwan and Jackson 1996, Moyle 1976). Although the life-history characteristics of steelhead are generally well known, the polymorphic nature of the subspecies has resulted in much confusion over the status and distribution of steelhead in San Francisco Estuary and its tributaries. Historically, the Sacramento-San Joaquin River systems supported large runs of steelhead (McEwan and Jackson 1996). Presumably, most streams with suitable habitat within the San Francisco Estuary also supported steelhead, however accurate population estimates for individual streams are not available (Skinner 1962, Leidy 1984).

Currently, small steelhead runs of unknown size are known to exist in South San Francisco Bay in San Francisquito Creek, San Mateo County; Guadalupe River and Coyote and Upper Penitencia creeks, Santa Clara County; Alameda Creek, Alameda County; and possibly San Leandro Creek, Alameda County (R. Leidy, unpub. data). Within Central San Francisco Bay steelhead runs are believed to occur in Corte Madera Creek and its tributaries, Miller Creek, Novato Creek, and possibly Arroyo Corte Madera del Presidio Creek, Marin County (R. Leidy, unpub. data). Within San Pablo Bay, steelhead make spawning runs in the Napa River and several of its tributary streams and Huichica Creek, Napa County; and the Petaluma River and Sonoma Creek and several of their tributary streams, Sonoma County (R. Leidy, unpub. data). Tributaries to Suisun Bay and adjacent drainages that support steelhead runs of unknown size include the Sacramento and San Joaquin rivers; Green Valley and Suisun creeks, Solano County; and Walnut Creek and possibly Alhambra, Pinole, Wildcat, and San Pablo creeks, Contra Costa County (R. Leidy, unpub. data). Steelhead may also be present in other tributary streams below migration barriers within the Estuary, but currently there is little or no data on their status in many streams. Steelhead adults and smolts may be found foraging in and migrating through estuarine subtidal and riverine tidal habitats within all areas of the San Francisco Estuary.

Population Status and Influencing Factors

Nehlsen et al. (1991) identified at least 43 steelhead stocks at moderate to high risk of extinction, with more than 23 stocks believed to have been extirpated, on the west coast of the United States. Steelhead in California are estimated to number roughly 250,000 adults, which is one half the adult population of 30 years ago (McEwan and Jackson 1996). As a result of this precipitous decline, the National Marine Fisheries is currently reviewing the status of steelhead to determine if they warrant listing under the Endangered Species Act. Estimates of the average annual steelhead run size for the Sacramento-San Joaquin River system, including San Francisco Bay tributaries, range between 10,000 and 40,000 adults (Hallock et al. 1961, McEwan and Jackson 1996). The California Fish and Wildlife Plan (CDFG 1965) estimated an annual run size for the Sacramento above the mouth of the Feather River of approximately 30,000 fish, and a total for the reminder of the entire Central Valley of 40,000 steelhead, including tributaries to San Francisco Bay. This likely places the size of steelhead runs in San Francisco Bay tributaries at well below 10,000 fish, however, the fact remains that reliable estimates for individual streams tributary to San Francisco Estuary do not exist.

General factors influencing steelhead population numbers during upstream migration, spawning, and incubation include barriers to passage, diversions, flow fluctuations, water temperature, and other water quality parameters, such as sedimentation of spawning habitats. Factors affecting juvenile rearing habitat and emigration within the San Francisco Estuary and its tributary streams include low summer flows combined with high water temperatures. Within Suisun Bay/Marsh the downstream migrating steelhead are adversely affected by altered flows, entrainment, and mortality associated with trapping, loading, and trucking fish at state and federal pumping facilities. In addition, dredging and dredged material disposal within the San Francisco Estuary may contribute to degradation of steelhead habitat and interference with migration, foraging, and food resources (LTMS 1996).

Trophic Levels

Larvae are primary consumers. Juveniles and adults are primary and higher order consumers.
Proximal Species

**Egg Predators:** Freshwater sculpins.

**Juvenile and Smolt Predators:** Other large freshwater, estuarine, and marine piscivorous fish.

**Juvenile and Adult Predators:** Harbor seals and other pinnipeds.

**Habitat/Cover:** Riparian, emergent, and palustrine wetland vegetation.

**Major Prey Items:** Aquatic and terrestrial insects, amphipods, snails, mysid shrimp, small fish.

Good Habitat

The preferred water depth for steelhead spawning ranges from six to 24 inches, while fry and parr prefer water depths of between two to 14 inches and 10 to 20 inches, respectively (Bovee 1978). Steelhead prefer to spawn in areas with water velocities of approximately two ft/sec (range =1.3-6 ft/sec), although optimal spawning velocity is partially a function of the size of fish; larger fish can successfully spawn in higher water velocities (Barnhart 1986). Optimal spawning substrate is reported to range from 0.2 to 4.0 inches in diameter, but steelhead will utilize various mixtures of sand-gravel and gravel-cobble (Bovee 1978, Reiser and Bjornn 1979). Optimal temperature requirements for steelhead vary as follows: adult migration, 46° to 52° F; spawning, 39° to 52° F; incubation and emergence, 48° to 52° F; fry and juvenile rearing, 45° to 60° F, and smoltification, <57° F (Bovee 1978, Reiser and Bjornn 1979, Bell 1986). While egg mortality begins to occur at 56° F and fish are known to have difficulty extracting oxygen from the water at temperatures exceeding 70° F (Hooper 1973), steelhead populations are often adapted to local environmental conditions where preferred temperature conditions are regularly exceeded for prolonged time periods (McEwan and Jackson 1996).

Some other important factors that are critical to maintaining optimal steelhead habitat include water quality and quantity, habitat heterogeneity, migration barriers, and introduced salmonids. Steelhead require relatively “good” water quality (e.g., low suspended sediment and contaminant loads and other forms of pollution), as well as sufficient flows for spawning, rearing, and migration. Diverse stream habitats consisting of shallow riffles for spawning and relatively deep pools, with well-developed cover, for rearing are important factors. The importance of estuarine or riverine tidal wetlands within the San Francisco Estuary for rearing foraging or migrating steelhead are not well understood.

References


Moyle, P.B., R.M. Yoshiyama, J.E. Williams and E.D. Wikramanayake. 1995. Fish species of special con-
General Information

The Delta smelt (Family Osmeridae) is a small, short-lived native fish which is found only in the Bay-Delta Estuary. The species was listed as threatened in 1993 under the Federal Endangered Species Act. Habitat loss is thought to be one of the most important elements in causing its decline. New water quality standards adopted by the state in 1995 are aimed in part at improving habitat conditions (SWRCB 1995).

Reproduction

The Delta smelt has low fecundity and is primarily an annual species, although a few individuals may survive a second year (Herbold et al. 1992). The location and season of Delta smelt spawning varies from year to year. Spawning, which occurs in shallow freshwater (CDFG 1992b, USFWS 1994), has been known to occur at various sites within the Delta, including the lower Sacramento and San Joaquin rivers and Georgiana Slough, and in sloughs of the Suisun Marsh (USFWS 1994). In 1996, newly emerged Delta smelt larvae were found in the Napa River, Cordelia Slough, Montezuma Slough, and in the San Joaquin River up to Stockton (CDFG unpub. data). Based on egg and larval trawls in recent low flow years, it appears that a significant portion of Delta smelt spawning now takes place in the northern and western Delta (CDWR 1992).

Spawning may occur from late winter (December) to early summer (July). In 1989 and 1990, two spawning peaks occurred, one in late-April and another early-May (USFWS 1994). Spawning has been reported to occur at about 45° to 59° F (7-15°C) in tidally influenced rivers and sloughs, including dead-end sloughs and shallow edge waters of the upper Delta. Most spawning occurs in fresh water, but some may occur in brackish water in or near the entrainment zone (Wang 1991). The demersal, adhesive eggs sink and attach to hard substrates, such as submerged tree branches and roots, gravel
or rocks, and submerged vegetation. Survival of adhesive eggs and larvae is probably significantly influenced by hydrology at the time of spawning (CDWR and USBR 1994). M. oyle et al. (1992) found no correlation between female length and fecundity. Females of 59-70 mm standard length (SL) ranged in fecundity from 1,247 to 2,590 eggs per fish, with an average of 1,907.

Spawning stock does not appear to have a major influence on Delta smelt year class success. However, the low fecundity of this species, combined with planktonic larvae which likely have high rates of mortality, requires a large spawning stock if the population is to perpetuate itself. This may not have been an important factor in the decline of Delta smelt, but it may be important for its recovery (CDFG 1992b).

Growth and Development

Newly hatched larvae are planktonic and drift downstream near the surface to the freshwater/saltwater interface in nearshore and channel areas. M. eugenei (1993) found that larvae hatched in 10 to 14 days under laboratory conditions and started feeding on phytoplankton at day four and on zooplankton at day six. Growth is rapid through summer, and juveniles reach 40 to 50 mm fork length (FL; the measure to the bottom of the fork of the tail fin) by early August. Growth slows in fall and winter, presumably to allow for gonadal development. Adults range from 55 to 120 mm FL, but most do not grow larger than 80 mm FL. Delta smelt become sexually mature in the fall at approximately seven to nine months of age. The majority of adults die after spawning.

Food and Feeding

Newly hatched larvae feed on rotifers and other microzooplankton. Older fish feed almost exclusively on copepods. Prior to 1988, Delta smelt ate almost solely the native Eurytemora affinis (Herbold 1987). During the 1980s Eurytemora affinis was displaced by the introduced copepod Pseudodiaptomus forbsi throughout Suisun Bay, and Delta smelt shifted to a diet of Pseudodiaptomus forbsi (P. Moyle, pers. comm.).

Distribution

Delta smelt are endemic to the Sacramento-San Joaquin Estuary. They have been found as far north as the confluence of the American and Sacramento rivers and as far south as Mossdale on the San Joaquin River. Their upstream range is greatest during periods of spawning. Larvae subsequently move downstream for rearing. Juvenile and adult Delta smelt commonly occur in the surface and shoal waters of the lower reaches of the Sacramento River below Isleton, the San Joaquin River below Mossdale, through the Delta, and into Suisun Bay (Moyle 1976, Moyle et al. 1992). Downstream distribution is generally limited to western Suisun Bay. During periods of high Delta outflow, Delta smelt populations do occur in San Pablo Bay, although they do not appear to establish permanent populations there (Herbold et al. 1992). Recent surveys, however, show that Delta smelt may persist for longer periods in Napa River, a tributary to San Pablo Bay (IEP, unpub. data).

Rearing and pre-spawning Delta smelt generally inhabit a salinity range of less than 2 ppt (parts per thousand), although they have been collected at salinities as high as 10 to 14 ppt (CDFG 1992b). Abundance of pre-spawning adults typically peaks upstream of the entrapment zone (CDWR and USBR 1994).

Population Status and Influencing Factors

Seven surveys, although not specifically designed to gather data on Delta smelt populations in the Estuary, have charted the abundance of Delta smelt. The summer townet survey, which began in 1959 and was primarily designed to measure striped bass abundance, is considered one of the best measures of Delta smelt abundance because it covers much of the species' habitat and represents the longest historical record. Although the abundance indices vary considerably, they generally remained low between 1983 and 1993. In recent years moderately wet conditions have produced relatively high abundances in the summer townet survey. The reduced population levels during the 1980s appear to have been consistent throughout the Delta and Suisun Bay, but declines may have occurred as early as the mid-1970s in the eastern and southern portions of the Delta (CDWR and USBR 1993).

The midwater trawl survey provides one of the best indexes of smelt abundance because it covers most of the range of Delta smelt (CDWR and USBR 1994). From 1967 through 1975, fall catches were generally greater than 10 smelt per trawl per month (in 6 of 8 years); from 1976 through 1989, catches were generally less than 10 smelt per trawl per month (in 13 of 14 years). Since 1986, catches have averaged considerably less than one smelt per trawl per month. The frequency of occurrence of Delta smelt in the trawls has also declined. Prior to 1983, Delta smelt were found in 30% or more of the fall trawl catches. In 1983-1985, they occurred in less than 30% of the catches, and since 1986, they have been caught in less than 10% of the trawls (Herbold et al. 1992). In 1993, the midwater trawl index was the sixth highest of the 25 years of record. In 1994, the index dropped to a 28-year low, but it rebounded again in 1995. Unlike the summer townet survey indices, the mean catches of Delta smelt have not declined in the midwater trawl survey. The smelt population is more dispersed in the summer than in the fall. The summer populations have decreased in average densities while the...
fall populations have decreased in numbers of schools (CDFG 1992b). Data from the Bay Study and the Suisun Marsh study show sharp declines in Delta smelt at about the same time. The exact timing of the decline is different in most of the sampling programs, but falls between 1982 and 1985 (Herbold et al. 1992).  

As a result of the sharp decline in abundance in the 1980s, the Delta smelt was listed as a federal “threatened” species by the U.S. Fish and Wildlife Service in March 1993 and as a State “threatened” species by the California Department of Fish and Game in December 1993. No single factor appears to be the sole cause of the Delta smelt decline; however, declines have been attributed primarily to restricted habitat and increased losses through entrainment by Delta diversions (CDWR 1992, Herbold et al. 1992, USFWS 1994). Reduced water flow may intensify entrainment at pumping facilities as well as reduce the quantity and quality of nursery habitat. Outflow also controls the location of the entrainment zone, an important part of the habitat of Delta smelt. A weak, positive correlation exists between fall abundance of Delta smelt and the number of days during spring that the entrainment zone remained in Suisun Bay (Herbold 1994). The number of days when the entrainment zone has been in Suisun Bay during the February through June period is one of only two parameters found so far that predicts Delta smelt abundance (Herbold 1994). Reduced suitable habitat and increased entrainment occurs when the entrainment zone moves out of the shallows of Suisun Bay and into the channels of the lower Sacramento and San Joaquin rivers as a result of low Delta outflow. The movement of the entrainment zone to the river channels not only decreases the amount of area that can be occupied by smelt, but also decreases food supply (Herbold et al. 1992).

Delta smelt in the western delta are vulnerable to entrainment by the pumps of the State Water Project and the Central Valley Project, as well as local agricultural diversions (CDWR 1992, NHI 1992, Herbold et al. 1992). Diversions in the northern and central Delta, where smelt are most abundant, are likely the greatest source of entrainment (USFWS 1994). Larvae and juveniles appear to be particularly vulnerable to pumping because screens are not effective for these life stages (CDWR and USBR 1994). Whether entrainment, estimated by salvage, affects abundance remains to be demonstrated statistically. However, the relative effects of entrainment are higher in dry years, when the abundance of Delta smelt is typically lowest and the distribution of the species shifts closer to the pumps in the interior Delta. Water diversions such as Contra Costa Canal, PG & E’s power plants, and in-Delta agricultural diversions, potentially entrain Delta smelt in numbers comparable to or greater than at the Central Valley Project and State Water Project pumps. However, initial results from Interagency Ecological Program studies have found few Delta smelt in agricultural diversions.

Although the effects of the recent high diversions of fresh water, especially when coupled with drought conditions from 1987-1992, are the most likely causes of the decline in the Delta smelt population, other contributing factors may include: the presence of toxic compounds in the water, competition and predation, food supply, disease, very high outflows, and low spawning stock.

Toxic contaminants have been identified as a factor that could affect Delta smelt survival (USFWS 1991). Possible pollutants include heavy metals, pesticides, herbicides, and polycyclic aromatic hydrocarbons. An inverse relationship has been found between copper applications to ricefields and Delta smelt abundance (Herbold, unpub. data), but no toxicity studies have been conducted to verify the degree to which pollutants in water and sediments affect Delta smelt.

Research conducted by Bennett (1995) suggests that competition with inland silversides, a non-native fish that arrived in the Bay around 1975, working synergistically with low flows, has contributed to Delta smelt decline. Inland silversides were found to be voracious predators of larval fish in both field and laboratory experiments. In addition, smelt and silversides may compete for copepods and cladocerans. Hatching and larval smelt may be extremely vulnerable to schools of foraging silversides, especially in low-outflow years when Delta smelt are forced into narrower, upstream channels, where silverside competition and predation may be increased. Evidence suggests that other non-native species, such as chameleon goby and striped bass, are either direct predators or compete with Delta smelt for food or habitat (CDWR and USBR 1994). However, it is questionable if striped bass is an important factor when both striped bass and Delta smelt were abundant in the 1960s, and the smelt was not a significant prey of the bass (CDFG 1992b).

Exact food requirements of Delta smelt are not known, but prey densities in the Estuary appear low relative to other systems in the United States, creating the potential for food limitation (Miller 1991). Moreover, there have been several changes in the species composition of zooplankton, with unknown effects on Delta smelt. The 1988 decline of Eurytemora affinis, a copepod which has been the primary food supply of Delta smelt, has been identified as a possible factor in the decline of smelt in the Estuary (CDFG 1992b). However, it may be that declines in E. affinis abundance, due to the introduction of other copepod species, is not an important factor because the smelt has shifted its diet and now consumes Pseudodiaptomus forbesi, which was introduced into the Estuary in 1986. The clam, Potamocorbula amurensis, may have an indirect effect on smelt.
populations by reducing its food supply (Herbold et al. 1992).

In some years disease is thought to cause widespread mortality of some fish species in the Estuary, but mortality of Delta smelt has not been specifically observed (Stevens et al. 1990). Mycobacterium, a genus of bacteria known to cause chronic infections in fish and other species, has been the major cause of mortality of Delta smelt held in the laboratory, and it may cause deaths among wild fish as well (Hedrick 1995).

The period of the Delta smelt decline includes unusually wet years with exceptionally high outflows. Very high outflows may be detrimental to the planktonic smelt larvae, which may be transported out of the Delta and into San Pablo and San Francisco bays with no way to get back upstream (CDFG 1992b).

It is possible that the size of the spawning stock influences population levels. However, there is not a statistically significant stock-recruitment relationship for Delta smelt, so this factor is not considered a primary factor regulating abundance (CDWR and USBR 1994).

Trophic Levels

Delta smelt are secondary consumers.

Proximal Species

Egg and larvae predators: Inland silversides, Menidia beryllina.

Juvenile and adult predators: Striped bass, Morone saxatilis (likely).

Prey: Eurytemora affinis, Pseudodiaptomus forbsii, rotifers (e.g., Trichocerca).

Good Habitat

Spawning habitat has been as widely dispersed as the Napa River to Stockton in 1996. The predominate feature appears to be shallow, freshwater conditions with some sort of solid substrate for the attachment of eggs. Spawning has been reported to occur at about 45-59°F (7-15°C) in tidally influenced rivers and sloughs including dead-end sloughs and shallow edge waters of the upper Delta.

Juvenile and adult Delta smelt commonly occur in the surface and shoal waters of the lower reaches of the Sacramento River below Mossdale, through the Delta, and into Suisun Bay (Moyle 1976, Moyle et al. 1992). Rearing and pre-spawning Delta smelt generally inhabit a salinity range of less than 2 ppt, although they have been collected at salinities as high as 10 to 14 ppt (CDFG 1992a). Analysis of the salinity preferences using midwater trawl data indicate that Delta smelt distribution peaks upstream of the entrapment zone (Obrebski 1993). It should be noted, however, that the distribution of Delta smelt is fairly broad, particularly in years when abundance levels are high (CDWR and USBR 1993). Evidence from the 1993 year class also demonstrates that salinity position does not necessarily regulate Delta smelt distribution in all years. In late 1993 and early 1994, Delta smelt were found in Suisun Bay region despite the fact that X2 was located upstream. Samples collected in this area demonstrated that high levels of the copepod Eurytemora were present, suggesting that food availability may also influence smelt distribution (CDWR and USBR 1994).

Although these results show that the Delta smelt is not an entrapment zone specialist, there is evidence that their abundance is correlated with X2. Herbold (1994) found a significant relationship between the number of days X2 was in Suisun Bay during February through June versus midwater trawl abundance. Furthermore, when the entrapment zone is in Suisun Bay and both deep and shallow water exists, Delta smelt are caught most frequently in shallow water (Moyle et al. 1992).

Results from the University of California, Davis provide an indication of environmental tolerances of Delta smelt (Swanson and Cech 1995). The study found that although Delta smelt tolerate a wide range of water temperatures (<8°C to >25°C), warmer temperatures apparently restrict their distribution more than colder temperatures.

References


Herbold et al. 1992b. Written testimony on Delta smelt. Submitted by the Ca. Dept. of Fish and Game to the

The entrapment zone, also referred to by a variety of other descriptive terms, such as the “mixing zone,” the “null zone,” and the “zone of maximum turbidity,” is the area within an estuary where the freshwater from a stream meets with the salt water of the ocean. This zone is biologically highly productive, and considered to be of critical importance to the aquatic food web of the Estuary.

2 X2 is the geographic location, measured in kilometers above the Golden Gate, of the entrapment zone. X2 is largely a function of outflow, such that when outflow is high, X2 is closer to the Golden Gate. X2 was used by U.S. Fish and Wildlife Service in defining Delta Smelt’s critical habitat under the Endangered Species Act (USFWS 1994).

Personal Communications
Peter Moyle, University of California, Davis.
**Longfin Smelt**

*Spirinchus thaleichthys*

Frank G. Wernette

**General Information**

The longfin smelt (Family: Osmeridae) is a three to seven-inch long silvery fish (Moyle 1976). Longfin smelt were the most abundant smelt species in the Bay-Delta Estuary prior to 1984 and have been commercially harvested (Wang 1986). In 1993, the U.S. Fish Wildlife Service (USFWS) was petitioned to list the longfin smelt under the federal Endangered Species Act. In January 1994, however, USFWS determined that the longfin smelt did not warrant listing because other longfin smelt populations exist along the Pacific Coast, the Bay-Delta Estuary population does not appear to be biologically significant to the species as a whole, and the Bay-Delta Estuary population may not be sufficiently reproductively isolated (Federal Register Vol. 59 No. 869, January 6, 1994). Still, longfin smelt are typically addressed in Biological Assessments because of the decline in their abundance after 1982 and the relatively small increase in abundance following a wet year in 1993. The species may also be considered in the future for listing under the California Endangered Species Act.

The longfin smelt is an euryhaline species with a 2-year life cycle. Spawning occurs in fresh water over sandy-gravel substrates, rocks, or aquatic plants. Spawning may take place as early as November and extend into June, although the peak spawning period is from January to April. After hatching, larvae move up into surface water and are transported downstream into brackish-water nursery areas. Delta outflow into Suisun and San Pablo bays has been positively correlated with longfin smelt recruitment because higher outflow increases larval dispersal and the area available for rearing. The longfin smelt diet consists of mysids, although copepods and other crustaceans also are eaten. Longfin smelt are preyed upon by fishes, birds, and marine mammals (Federal Register Vol. 59 No. 4, January 3, 1994).

**Reproduction**

Maturation of longfin smelt begins late in the second summer of their life in August and September. As they mature, the smelt begin migrating upstream from San Francisco and San Pablo bays to Suisun Bay and the Delta. Longfin smelt spawn in fresh water, primarily in the upper end of Suisun Bay and in the lower and middle Delta. In the Delta, they spawn mostly in the Sacramento River channel and adjacent sloughs (Wang 1991). During the recent drought, when saline water intruded into the Delta, larval longfin smelt were found near the Central Valley Project and State Water Project export facilities in the southern Delta (Wang 1991). Ripe adults, larvae, and juveniles are salvaged at the export facilities in every below normal or drier water year (Baxter, pers. comm.). The eggs are adhesive and are probably deposited on rocks or aquatic plants. Longfin smelt eggs hatch in 37-47 days at 45°F.

**Growth and Development**

Shortly after hatching, longfin smelt larvae develop a gas bladder that allows them to remain near the water surface (Wang 1991). The larvae do not vertically migrate, but instead remain near the surface on both the flood and ebb tides (CDFG 1992). Larvae are swept downstream into nursery areas in the western Delta and Suisun and San Pablo bays with larval dispersal farther downstream in years of high outflow than in years of low outflow (CDFG 1992; Baxter, pers. comm.). Early development of gas bladders by longfin smelt causes the larvae to remain near the surface much longer than Delta smelt larvae. That factor and earlier spawning period help explain why the longfin smelt larvae are dispersed much farther downstream in the Estuary than are Delta smelt larvae (Baxter, pers. comm.). Larval development occurs primarily in the February through May period and peaks during February-April (CDFG 1992).

Metamorphosis of longfin smelt from the larval to juvenile form begins 30-60 days after hatching, depending on temperature. Most longfin smelt growth occurs during the first summer, when length typically reaches...
6 to 7 cm. During their second summer, smelt reach 9 to 11 cm in length (NHI 1992). Most longfin smelt spawn and die at two years of age (CDFG 1992).

**Food and Feeding**

The main prey of adult longfin smelt is the opossum shrimp, *Neomysis mercedis* (NHI 1992). There is little information on food habitats of longfin smelt larvae, but fish larvae of most species, including Delta smelt, are known to feed on phytoplancton and small zooplankton, such as rotifers and copepod nauplii (H unter 1981, USBR 1993). Juvenile longfin smelt feed on copepods, cladocerans, and mysids. The mysid *Neomysis mercedis* is the most important prey of larger juveniles.

**Distribution**

Longfin smelt are widely distributed in estuaries on the Pacific Coast. They have been collected from numerous river estuaries from San Francisco to Prince William Sound in Alaska (Moyle 1976).

Longfin smelt are euryhaline meaning they are adapted to a wide salinity range. They are also anadromous. Spawning adults are found seasonally as far upstream in the Delta as Hood, M edford Island, and the Central Valley Project and State Water Project fish collection facilities in the southern Delta. Historically, before construction of Shasta Dam in 1944, saline water intruded in dry months as far upstream in the Delta as Sacramento, so longfin smelt may have periodically ranged farther upstream than they do currently (H erbold et al. 1992).

Except when spawning, longfin smelt are most abundant in Suisun and San Pablo bays, where salinity generally ranges between 2 ppt and 20 ppt (NHI 1992). Pre-spawning adults and yearling juveniles are generally most abundant in San Pablo Bay and downstream areas as far as the South Bay and in the open ocean.

**Population Status and Influencing Factors**

Abundance estimates were developed from otter trawl and midwater trawl sampling conducted by the Outflow/ Bay study as part of the Interagency Ecological Program. Fall midwater trawl surveys provide the longest index of longfin smelt abundance.

Results of the fall midwater trawl surveys indicate that, like Sacramento splittail abundance, longfin smelt abundance has been highly variable from year to year, with peaks and declines coinciding with wet and dry periods. Longfin smelt abundance has steadily declined since 1982. Abundance continued to be suppressed during the drought years beginning in 1987. Longfin abundance was very low from 1987 to 1992, with 1992 having the lowest index on record. Abundance increased moderately in 1993 following the drought during a period of improved Delta outflow.

Year-class abundance of longfin smelt appears to depend on the environmental conditions experienced by the eggs and young fish. Generally, year-class abundance is positively related to Delta outflow (i.e., high abundance follows high outflow during winter and spring). Factors possibly contributing to the recent decline in longfin smelt abundance are reduced Delta outflow, entrainment in diversions, introductions of exotic species, loss of habitat, and the recent drought.

**Delta Outflow** – Higher outflows result in higher longfin smelt survival. An index of survival computed as the ratio of the index of abundance from fall midwater trawl surveys to an index of larval abundance in previous springs was strongly correlated (r=0.95) with December-August outflow. Delta outflow or factors associated with outflow affect survival of larvae and early juveniles. Delta outflow may be the single most important factor controlling longfin smelt abundance. High outflows increase dispersion downstream, available habitat, and possibly, food availability. High outflow may also reduce predation and the effects of other adverse factors (i.e., toxin concentrations). Low outflow conditions reduce downstream dispersion and increase vulnerability to entrainment in Delta diversions.

Longfin smelt abundance (according to the fall midwater trawl survey index) is positively related to Delta outflow (Stevens and Miller 1983, CDFG 1992). Regression analysis indicated that 79% of variability in the midwater trawl survey index is explained by changes in January and February Delta outflow. The significant relationship between the index of abundance from the fall midwater trawl surveys and Delta outflow may reflect the effect of outflow on survival of larvae and early juveniles. Year-class strength may be largely determined by survival of the early life stages.

High Delta outflow may increase the amount of suitable brackish water rearing habitat; reduce salinity in the Estuary, reducing competition and predation by marine organisms; reduce predation because young smelt are more dispersed and turbulence is higher; increase phytoplankton and zooplankton production; and increase transport of larvae out of the Delta and away from diversions (CDFG 1992; Stevens and Miller 1983; Baxter, pers. comm.). Any of these mechanisms may be responsible for the observed relationship between Delta outflow and longfin smelt abundance.

The position of the entrainment zone, location of X2, and volume of critical nursery habitat are determined by Delta outflow. In addition to the relationship with outflow, the fall midwater trawl survey index has a positive relationship with the location of X2 and the volume of critical nursery habitat (J asby 1993, H ergesell 1993).

Delta smelt abundance tends to be highest when X2 has an intermediate value (i.e., X2 is located in up-
The location of X2 is also a good predictor of longfin smelt abundance. Since X2 and the volume of critical nursery habitat are largely determined by Delta outflow, the relationship between longfin smelt abundance and the location of X2 or volume of critical habitat may simply reflect effects of outflow or other correlates of outflow on longfin smelt abundance.

**Lower San Joaquin River** - Reverse flow in the lower San Joaquin River usually transports relatively fresh water drawn from the Sacramento River and may increase upstream migration of adults to the southern Delta. Reverse flow may also transport larvae to the southern Delta. In the southern Delta, adults, larvae, and juveniles are vulnerable to entrainment, predation, and other sources of mortality.

**Entrainment** - Entrainment of longfin smelt by Delta diversions affects spawning adults, larvae, and early juveniles. Older juveniles and prespawning adults generally inhabit areas downstream of the Delta. Salvage at both the Central Valley Project and State Water Project fish protection facilities has varied greatly between years. Salvage represents entrainment, but the number of fish salvaged is often much lower than total number entrained because fish, particularly those smaller than about 20-30 mm, pass through the fish screens at the salvage facilities and, therefore, are not salvaged.

With the exception of 1986, a wet year, the annual salvage of longfin smelt at the Central Valley and State Water Project pumps was much higher during 1984-1990 than during 1979-1983. The decline in abundance in 1984 may be attributable to increased entrainment by the Central Valley Project and State Water Project pumps and other diversions, but reduced Delta outflow, discussed previously, may be a more important factor affecting abundance.

Entrainment of adult longfin smelt has a potentially greater adverse effect on the population than entrainment of larvae and young juveniles because unless the adults have already spawned, their reproductive value is much greater than that of younger fish. Adult smelt are entrained at the State Water Project and Central Valley Project pumping facilities primarily during November-February. The number of adults entrained is low relative to the number of juveniles entrained.

Longfin smelt larvae have been captured in the southern Delta near the Central Valley Project and State Water Project export facilities (Spaar 1990, 1993; Wang 1991). Larval smelt are too small to be salvaged at the State Water Project and Central Valley Project fish protection facilities. Based on the high salvage rates of young-of-the-year juveniles in some years, it can be assumed that many thousands of longfin smelt larvae were also entrained, especially during February through April.

During years of high flows, most longfin smelt adults spawn in the western Delta, and their larvae are generally transported out of the Delta and therefore are unlikely to be entrained in Delta diversions in large numbers. During the 1987-1992 drought and other low flow years, however, outflows were low and exports were high. Adults, larvae, and juveniles remained in the Delta, as indicated by salvage at the Central Valley Project and State Water Project fish protection facilities. Most juveniles were entrained during April-June and averaged 30-45 mm long, with length correlated with the month of entrainment. Thus, longfin smelt suffer not only loss of larval dispersal and rearing habitat in a drought, but also from higher rates of entrainment.

Adult, juvenile, and larvae longfin smelt are vulnerable to entrainment in diversions other than exports at the Central Valley Project and State Water Project pumps, including diversions to PG&E's power generating plants, industrial diversions, agricultural diversions, and others. However, entrainment of longfin smelt in these diversions has not been extensively evaluated.

**Other Factors** - Other factors that may affect survival of longfin smelt include food limitation and presence of toxic materials and introduced species. Abundance of Neomysis and other zooplankton prey (e.g., rotifers) of longfin smelt have declined in recent years (Olbinski et al. 1992). It is not known what effect the decline in prey abundance has had on longfin smelt; however, food limitation may be important because year-class strength of many fish populations, particularly species with planktonic larvae, may be strongly influenced by feeding conditions during the larval life stage (Lasker 1981).

Agricultural chemicals (including pesticides and herbicides), heavy metals, petroleum-based products, and other waste materials toxic to aquatic organisms enter the Estuary through nonpoint runoff, agricultural drainage, and municipal and industrial discharges. The effects of toxic substances have not been tested on longfin smelt; however, bioassays indicate that water in the Sacramento River is periodically toxic to larvae of the fathead minnow, a standard EPA test organism (Stevens et al. 1990). The short life span of longfin smelt and relatively low position in the food chain probably reduce the accumulation of toxic materials in their tissues and make them less susceptible to injury than species that live longer (NHI 1992).

Many exotic species have invaded the Estuary in recent years. These species may compete with or prey on longfin smelt. No single invasion of exotic species parallels the decline of longfin smelt closely enough to suggest that competition from or predation by the species...
was a primary cause of the longfin smelt's recent decline. The effects of multiple-species invasion, which have occurred in the Estuary, are extremely difficult to assess. The effects of exotic species invasions on longfin smelt is likely not large since Delta outflow explains over 60% of the variation in abundance (Baxter, pers. comm.).

**Trophic Levels**

Longfin smelt are secondary consumers.

**Proximal Species**

**Predators:** Brown pelican, river otter, striped bass, centrarchids.

**Prey:** Zooplankton (cladocerans), opossum shrimp (Neomysis mercedis), crustaceans (copepods).

**Good Habitat**

Longfin smelt are typically pelagic and use the larger sloughs and rivers of the Delta and Bay. The optimal salinity habitat for non-spawning adults is 2 to 20 ppt. Optimal habitat for spawning adults is 0 to 2 ppt. Optimal habitat for spawning includes submergent vegetation that can be used as a substrate for the adhesive eggs. High quality habitat is also defined as having low levels of exposure to entrainment into water export facilities and agricultural or managed wetland diversions. Adjacent runoff of agricultural pesticides is minimal or does not occur in good habitat areas.

Juvenile longfin use the open water, shallow shoal areas of San Pablo and Suisun bays after being transported downstream from spawning areas in the Delta. An average X2 location in upper Suisun Bay defines good habitat conditions for longfin smelt. Adjacent tidal wetlands are important to supporting the nutrient cycling and carbon input functions which in turn support the prey species upon which longfin feed.

**References**


Personal Communications
Randall Baxter, Assoc. Fisheries Biologist, California Department of Fish and Game.

Jacksmelt
Atherinopsis californiensis
Michael K. Saiki

General Information
Although jacksmelt (Family: Atherinidae) is not an important commercial fish, it nevertheless constitutes the largest portion of “smelt” captures in California (Emmett et al. 1991). This species is also commonly caught by recreational anglers fishing from piers (Frey 1971). In an ecological sense, jacksmelt occupy an important niche in trophic pathways of nearshore coastal, bay, and estuarine ecosystems (Clark 1929, Allen and DeMartini 1983, CDFG 1987).

Reproduction
Emmett et al. (1991) describes the sexual and reproductive characteristics of jacksmelt as gonochoristic (its gender is determined by developmental rather than hereditary mechanisms) and iteroparous (it has the capacity to survive and spawn beyond one or multiple spawning seasons). Spawning occurs several times during a spawning season (Clark 1929). The eggs are demersal and adhesive, and can often be found on vegetation in shallow nearshore coastal habitats and in estuaries and bays (Clark 1929, Wang 1986).

Adults move inshore and into bays and estuaries to spawn during late winter and early spring (Clark 1929, Wang 1986). In San Francisco Bay, spawning occurs from October to early August (Wang 1986). Spawning in San Pablo Bay reportedly occurs from September to April (Ganssle 1966). Eggs are laid on substrates/vegetation (e.g., Zostera spp., Gracilaria spp., hydroids) in which they become entangled (Frey 1971, Wang 1986). Embryonic development is indirect and external, and if given a suitable environment, the yellowish-orange eggs hatch within seven days (Wang 1986). The fecundity of jacksmelt is not yet documented, but probably exceeds 2,000 eggs per female (Emmett et al. 1991). Unfertilized jacksmelt eggs are spherical in shape and 0.9-2.2 mm in diameter (Clark 1929; fertilized eggs are 1.9-2.5 mm in diameter (Wang 1986).

Growth and Development
After hatching, larvae remain on the bottom for a moment and then actively swim near the surface (Wang 1986). Larvae vary in size from 7.5 to 8.6 mm immediately after hatching, to about 25 mm long prior to the juvenile transformation (Clark 1929, Wang 1986). At eight days posthatch, they average 10.5-11.7 mm in length whereas at 24 days posthatch, they average 17.6-20.3 mm in length (Middaugh et al. 1990). Juveniles can attain 110 mm during their first year, and 180-190 mm after two years (Clark 1929). All individuals mature by their third year, but some may grow quickly and mature in their second year (Clark 1929). Adult jacksmelt have been reported to attain a length of 780 mm and an age of 11 years (Millar and Lea 1972, Frey 1971) but, more typically, the maximum size is 200 mm total length, and the maximum age is 9-10 years (Clark 1929).

Food and Feeding
The jacksmelt is omnivorous (Bane and Bane 1971, Ruagh 1976). Larvae live on their yolk-sac for about 48 hours after hatching when it is fully absorbed (Middaugh et al. 1990). Major food items for jacksmelt include algae (Ulothrix spp., Melosira moniliformis, Enteromorpha spp., and other filamentous algae), benthic diatoms, crustaceans (mysids, copepods, decapod larvae), and detritus (Bane and Bane 1971, Ruagh 1976). In addition, stomach analyses of juvenile jacksmelt show that amphipods are a common food item, indicating that juveniles may feed on the bottom (Wang 1986).
**Distribution**

Jacksmelt occur from Santa Maria Bay, Baja California, northward to Yaquina Bay, Oregon (Miller and Lea 1972, Eschmeyer et al. 1983). However, this species is uncommon north of Coos Bay, Oregon (Emmett et al. 1991).

Prior to or after the spawning season, adult jacksmelt typically occur in coastal waters near shore (Baxter 1960). Ruagh (1976) mentioned that jacksmelt are usually caught within 5 km of shore where they often school with topsmelt (Atherinopsis affinis).

Locally, jacksmelt have been reported to spawn in San Francisco Bay (Wang 1986) and San Pablo Bay (Ganssle 1966, Wang 1986). Juveniles are also present in San Francisco Bay (Baxter 1960, Aplin 1967), San Pablo Bay (Ganssle 1966), Carquinez Strait (Messersmith 1966), and occasionally in Suisun Bay (Wang 1986, Herbold et al. 1992, Jones and Stokes Assoc. 1979) and Napa marsh (Jones and Stokes Assoc. 1979). The amount of freshwater inflow seemingly affects the local distribution of jacksmelt. During years of low freshwater inflow, jacksmelt occur as far upstream as Carquinez Strait and San Pablo Bay, but during high-flow years they are seemingly restricted to Central San Francisco Bay and South San Francisco Bay (CDFG 1987).

**Population Status and Influencing Factors**

Presently, jacksmelt are particularly abundant in Tomales, Central San Francisco, South San Francisco, and San Pablo bays (Emmett et al. 1991). Midwater trawl samples performed in South San Francisco Bay between 1980-1988 showed that jacksmelt were the second most common species caught, behind northern anchovy (H erbold et al. 1992). Furthermore, jacksmelt were more abundant and occurred more frequently than topsmelt in the South Bay (Herbold et al. 1992). In San Pablo and Central San Francisco bays, Herbold et al. (1992) reported that jacksmelt were the third most common species caught. Midwater trawl samples performed in the Carquinez Strait between 1961-1962 found over 9% of the total catch consisted of jacksmelt (Messersmith 1966). Herbold et al. (1992) noted that during 1980-1988 jacksmelt numbers seemed to vary widely in the Central Bay and are seemingly unpredictable from year to year, whereas numbers in the South Bay show little variation from year to year.

Although specific studies relating fish abundance to environmental variables were not found during our search of the literature, jacksmelt may be vulnerable to pollution and habitat modifications because they depend on embayments and estuaries for spawning.

**Trophic Levels**

Omnivorous (primary and higher order consumers).

**Proximal Species**

**Predators:** Yellowtail (Seriola lalandii), sharks and other piscivorous fishes, piscivorous birds (e.g., brown pelicans and gulls).

**Prey:** Small crustaceans, algae.

**Habitat:** Kelp (cover for juveniles and adults); algae, huercoids, and eelgrass (spawning substrate).

**Parasites:** Nematodes sometimes found living in flesh.

**Good Habitat**

Bays and estuaries provide important spawning habitat for jacksmelt. In general, the preferred spawning areas are situated in shallow nearshore habitats containing submerged vegetation (Wang 1986). Water quality variables suitable for embryo development are as follows: temperature, 10-12°C; and salinity, polyhaline and as low as 5 ppt (Wang 1986). Schools of larvae occur near the water surface over a variety of substrates, but mostly sandy and muddy bottoms and in the kelp canopy (Frey 1971). Optimum larval and juvenile survival and growth appears to be at salinities of 10-20 ppt, indicating that larvae may prefer mesohaline environments (Middaugh and Shenker 1988, Middaugh et al. 1990). Juveniles and adults prefer sandy bottoms in murky water at depths of 1.5-15 m below the surface (Feder et al. 1974). Furthermore, they seem to use open waters in San Francisco Bay and sloughs in and near Suisun Marsh and Napa Marsh (Jones and Stokes Assoc. 1979). Jacksmelt are apparently more sensitive than topsmelt to fluctuations in salinity and temperature (Emmett et al. 1991).

**References**


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**Topsmelt**

* Atherinops affinis

**General Information**

On the West Coast, topsmelt (Family: Atherinidae) are represented by five recognized subspecies of which only one, the San Francisco topsmelt (*Atherinops affinis affinis*), inhabits San Francisco Bay (Wang 1986). Topsmelt are a small but tasty food fish taken from piers by recreational anglers (Emmett et al. 1991). However, commercial fishing for topsmelt is limited, with the species comprising only about 15-25% of the total “smelt” catch (Bane and Bane 1971, Frey 1971). Ecologically, topsmelt are an important prey item for many piscivorous birds and fishes (Feder et al. 1974).

**Reproduction**

According to Emmett et al. (1991), the topsmelt is gonochoristic (its gender is determined by developmental rather than hereditary mechanisms) and iteroparous (it has the capacity to survive and spawn beyond one or multiple spawning seasons). Adults move into shallow sloughs and mud flats in late spring and summer to spawn (Wang 1986). In San Francisco Bay, spawning occurs from April to October, with peaks in May and June (Wang 1986). Although eggs are deposited singly, the thick chorion bearing 2-8 filaments becomes entangled in aquatic vegetation, resulting in the formation of large clusters of eggs (Wang 1986). Topsmelt seemingly spawn in batches, laying eggs more than once during a spawning season (Fronk 1969, Wang 1986). The fecundity of topsmelt ranges from 200 eggs/fish for females measuring 110-120 mm in length to about 1,000 eggs/fish for females measuring 160 mm or more in length (Fronk 1969). Hatching time varies from 35 days at 13°C to less than 9 days at 27°C (Hubbs 1969).
Growth and Development

Topsmelt eggs are spherical in shape and approximately 1.5-1.7 mm in diameter (Wang 1986). Between nine and 35 days after fertilization, eggs hatch into planktonic larvae that measure 4.3-4.9 mm total length (TL) and 0.0011 grams wet weight (Emmett et al. 1991) or 5.1-5.4 mm standard length (SL) (Middaugh et al. 1990). Larvae measure 9.5-10.0 mm after the yolk-sac is absorbed, and begin to develop juvenile characteristics when approximately 18.5 mm long (Wang 1986). Juveniles may vary in length from 18.5 to 120.0 mm (Schultz 1933, Fronk 1969). Topsmelt mature in their second or third year, depending on subspecies, and may live six to nine years (Schultz 1933, Feder et al. 1974). Adults can attain as much as 120 mm in length for the southernmost subspecies (A. affinis littoralis) and as much as 370 mm in length for the northernmost subspecies (A. affinis oregonia) (Schultz 1933, Fronk 1969, Eschmeyer et al. 1983). In general, northern varieties grow larger than southern subspecies (Schultz 1933).

Food and Feeding

The topsmelt is characterized by an omnivorous diet (Quast 1968, Horn and Allen 1985). Topsmelt from bay and estuarine habitats consume mostly plant material (diatoms, filamentous algae, and detritus), whereas those from ocean habitats feed mainly on planktonic crustaceans (gammarid and caprellid amphipods, mysids, ostracods, copepods, and crustacean larvae) (Moyle 1976, Quast 1968, Horn and Allen 1985). Topsmelt from bay habitats feed mainly on crustacean larvae (e.g., tracods, copepods, and crustacean larvae) (Moyle 1976, Fronk 1969). Juveniles and adults forage mostly during daylight near the surface in deep water or on the bottom in shallow water (Hobson et al. 1981).

Distribution

Topsmelt can occur from the Gulf of California northward to Vancouver Island, but are usually rare north of Tillamook Bay, Oregon (Miller and Lea 1972, Hart 1973, Eschmeyer et al. 1983). The five subspecies are A. affinis oregonia (occurs from Oregon to Humboldt Bay, California), A. affinis affinis (occurs in San Francisco Bay and surrounding waters to Monterey, California), A. affinis littoralis (occurs from Monterey to San Diego Bay, California), A. affinis cedrosensis (the kelp topsmelt), and A. affinis insularum (the “island topsmelt,” occurs around the Santa Barbara Islands, California) (Schultz 1933, Feder et al. 1974).

In San Francisco Bay, spawning has been observed in the South Bay near the Aquatic Park in Berkeley and at the Dumbarton Bridge (Wang 1986). Small schools of larvae often occur near the surface of both shallow water and open water, and are particularly abundant in tidal basins (e.g., Aquatic Park in Berkeley; Lake Meritt in Oakland) and the sluggish waters of the South Bay (e.g., Robert Crown Memorial Park; Hunters Point; San Mateo Bridge; Dumbarton Bridge) (Wang 1986). Juvenile topsmelt generally move into open waters of the bay or into coastal kelp beds. Some juveniles may occur in Suisun Bay during summer and early fall as the salt wedge moves to the upper reaches of the Estuary (Wang 1986). In general, topsmelt seem to be much less common outside of the South Bay.

Population Status and Influencing Factors

Field studies indicate that topsmelt are among the most abundant fish species occurring in shallow-water sloughs of South San Francisco Bay (Jones and Stokes Assoc. 1979, Woods 1981, Herbold et al. 1992). Herald and Simpson (1955) reported that topsmelt were commonly caught in a fixed fish-collecting device located at the Pacific Gas and Electric Company power plant in South San Francisco Bay. Furthermore, Wild (1969) reported that topsmelt was the most abundant species of fish sampled at the mouth of Plummer Creek (located in South San Francisco Bay). Midwater trawls fished at several locations in South San Francisco Bay during 1980-1988 also yielded numerous topsmelt (Herbold et al. 1992). South Bay topsmelt increased in abundance during two of the recent drought years, but otherwise did not show consistent year-to-year patterns (Herbold et al. 1992).

Several factors may directly influence the abundance of topsmelt: salinity, water temperature, freshwater inflows, entrainment on intake screens at power plants and water diversions, and availability of spawning substrate. In Newport Bay, California, topsmelt abundance was significantly correlated with water temperature and salinity (Allen 1982). By comparison, no relationship was found between abundance indices and river flow in San Francisco Bay (CDFG 1987). Although this species is commonly impinged on intake screens of power plants and water diversions, this source of mortality may not be significant for bay populations (San Diego Gas and Electric 1980). In the Tijuana Estuary of southern California, abundance of topsmelt eggs and larvae was positively correlated with algal mats (Nordby 1982). In other words, topsmelt eggs and larvae were seemingly more abundant in areas with dense algal growth. Because this species uses algal mats and shallow-water eelgrass beds for spawning, destruction or removal of these types of vegetation may adversely affect topsmelt abundance.

Trophic Levels

Topsmelt are omnivorous (primary and higher order consumers).

Proximal Species

Predators: Many piscivorous birds and fishes.
**Prey:** Diatoms (major); diatoms, chironomid midge larvae, and amphipods (minor).

**Habitat:** Eel grass and micro algae (spawning substrate); kelp beds (adult and juvenile cover).

**Cohabitors:** Schools with shiner perch and jacksmelt.

**Good Habitat**

In general, topsmelt can tolerate a relatively broad range of environmental conditions during the time that they inhabit San Francisco Bay. However, for successful spawning to occur, they require submerged vegetation for egg attachment, water temperatures of 10-25°C, and salinities of less than 72 ppt (Schultz 1933, Carpelan 1955, Fronk 1969). By comparison, larvae must be able to school near the surface in shallow open-water areas, particularly tidal basins (Wang 1986). Young-of-the-year topsmelt are common in middle to low salinity portions of the Estuary (Wang 1986). Although juveniles can tolerate salinities varying from 2 ppt to 80 ppt, growth and survival are reduced at salinities above 30 ppt (Middaugh and Shenker 1988). In addition, juveniles and adults are seemingly eurythermal, but temperatures of 26-27°C or higher may cause stress (Carpelan 1955, Ehrlich et al. 1979). Recent studies indicate that embryonic and larval stages of topsmelt are sensitive to the effects of pollution (Singer et al. 1990, Anderson et al. 1991, Goodman et al. 1991, Hemmer et al. 1991). Thus, habitats used by topsmelt for spawning and rearing must not be exposed to appreciable amounts of pollution.

**References**


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**Threespine Stickleback**

*Gasterosteus aculeatus*

**Robert A. Leidy**

**General Information**

The threespine stickleback (Family: Gasterosteidae) is a small laterally-compressed fish with three spines on the dorsum and from 1 to 35 bony plates on the sides (Moyle 1976). Largely as a matter of taxonomic convenience, Miller and Hubbs (1969) suggested that there are two forms: *G. a. aculeatus* for the fully-plated, anadromous form; and *G. a. microcephalus* for the partially-plated freshwater/resident form. The threespine stickleback is a polymorphic species and as such, populations within the San Francisco Estuary and its tributary streams support resident/freshwater and anadromous/saltwater forms, as well as mixtures of the two forms that presumably interbreed (Moyle, pers. comm.). The threespine stickleback has no commercial value, but has important scientific value, especially to evolutionary biologists.

**Reproduction, Growth and Development**

The following discussion is taken largely from Moyle (1976) unless otherwise referenced. Threespine sticklebacks typically complete their life cycle within one year although some individuals may live two to three years. Individuals from freshwater populations typically do not exceed 60 mm total length (TL), while anadromous forms may exceed 80 mm TL. Adult females are usually larger than adult males.

Anadromous forms migrate into freshwater breeding areas as water temperatures increase during April through July, although some stickleback populations may remain in estuarine environments to spawn if suitable habitat is present (Moyle 1976, Wang 1986).
Anadromous forms typically spawn earlier than freshwater populations. Spawning typically occurs at 15° to 18°C (Vrat 1949, Wang 1986). Males begin to display bright green and red breeding coloration as they move away from schools to set up breeding territories and construct nests. Nests are excavated in the substrate as shallow pits. The pits are then covered with algae or other plant fragments and formed into a tunnel that is held together by a sticky renal secretion (Greenbank and Nelson 1959). Females are then courted by males into the nest where the female may lay between 50 and 300 eggs in several spawnings. Eggs are spherical and average 1.5-1.7 mm in diameter (Wang 1986). A pair can spawn up to six times within a 10-15 day period (Wang 1986). Following egg laying, the male drives away the female, fertilizes the eggs, and then begins to incubate the eggs while defending the nest from other sticklebacks and predators. The male is known to circulate water over the eggs by fanning his pectoral fins and to clean the eggs with his mouth. Immediately prior to hatching the male tears apart the nest and breaks apart the egg clusters which is thought to increase the survival of hatching young (Wang 1986). Length at hatching is between 4.2 and 5.5 mm TL (Vrat 1949, Kuntz and Radcliffe 1917).

Juveniles are most abundant in late summer, followed by drastic declines in abundance in the fall and winter (Wang 1986). It is unknown whether populations of juveniles within the San Francisco Estuary make extensive migrations into open water/subtidal habitats within the Estuary. M. oyle (1976) states that freshwater and anadromous populations range from complete ecological separation to complete interbreeding.

Food and Feeding

Threespine sticklebacks are visual feeders primarily on small benthic organisms or organisms living on submerged, rooted, or floating macrophytes such as insect larvae, chironomid midge larva, and ostracods (Hyne 1950, Beukema 1963, Hagen 1967). Anadromous forms feed mostly on free-swimming crustaceans (Barraclough and Fulton 1967, 1978; Barraclough et al. 1968). In a study of threespine stickleback diet in San Pablo Creek, a tributary to San Pablo Bay, Snyder (1984) found the diet consisted of approximately 42% insects (mainly chironomid larvae), 28% crustacea (mainly ostracods), and 10% earthworms (Lumbricidae). Fish eggs and plant material accounted for approximately 9% of the diet (Snyder 1984).

Distribution

Threespine stickleback are native to the coastal waters of Mediterranean Europe, north to Russia, and east to Japan and Korea (M. oyle 1976). In North America, threespine stickleback populations occur on the East coast south to Chesapeake Bay, and on the West coast south from Alaska to Baja California. In California, populations are found below barriers such as dams and falls in coastal streams, including the San Francisco Estuary and its tributary streams, and in the Central Valley (M. oyle 1976).

Within the San Francisco Estuary, threespine stickleback are widely distributed and often locally abundant in fresh-, brackish-, and saltwater intertidal upper marsh and riverine tidal marsh habitats (Leidy 1984; Leidy, unpub. data; Cathy Hieb, unpub. data.). Leidy (1984) recorded threespine stickleback in 43% of 457 samples of Estuary streams between elevation 0 to 123 m.

Threespine stickleback are also abundant in large areas of formerly tidal salt and brackish marsh that have been converted to salt ponds in the South Bay and San Pablo Bay (Lonzarich 1989, Herbold et al. 1992). Carpelan (1957) recorded threespine stickleback as one of the most numerous fish in the Alviso salt ponds in the South Bay. Apparently, threespine stickleback persist in these ponds, particularly near the mouth of the Napa River, until salinities become too high (i.e., salinities between 40 to 50 ppt) (Herbold et al. 1992). There are approximately 9,059 acres of salt ponds in the Napa-Solano area of the North Bay and 27,497 acres in the South Bay that may be considered available for use by threespine stickleback on a seasonal basis (M. eorin et al. 1991).

Population Status and Influencing Factors

The current status of threespine stickleback within the San Francisco Estuary may be regarded as secure. Threespine stickleback populations currently are widespread and locally abundant in suitable habitats within the San Francisco Estuary. Because sticklebacks can readily disperse through estuarine and marine environments they are able to regularly recolonize habitats from which they may have been extirpated. Important factors negatively influencing population numbers likely include excess siltation and turbidity, increased water temperatures by the removal of riparian vegetation through stream channelization, pollution, loss of nesting, feeding, and cover habitats by the removal of aquatic macrophytes, the construction of barriers such as dams or drop structures, and the introduction of exotic piscivorous fish.

Trophic Levels

Larvae are primary consumers. Juveniles and adults are primary and higher order consumers.
**Proximal Species**

**Major Predators:** Kingfisher, egrets, herons, and other wading birds.

**Other Predators:** Adult salmonids and other large freshwater, estuarine, and marine piscivorous fish terrestrial and aquatic snakes.

**Major Prey:** Aquatic insects and crustacea, earthworms, fish eggs and vegetation.

**Habitat/cover:** Riparian, submerged, floating, and emergent wetland and aquatic vegetation.

**Good Habitat**

Freshwater populations of threespine stickleback prefer clear, cool backwater and pool habitats containing submerged, floating, or emergent vegetation, with sand or small-sized gravel substrates (Moyle 1976, Leidy 1984). This species is typically uncommon in silted pools with moderate to high turbidities (Leidy 1984). Marine and estuarine populations are pelagic, although they tend to remain to close to the shore (Moyle 1976). Threespine stickleback is uncommon where water temperatures regularly exceed 24° C (Moyle 1976).

**References**


**Personal Communications**

Peter Moyle, University of California, Davis
Chapter 2 — Estuarine Fish and Associated Invertebrates

Brown Rockfish
Sebastes auriculatus
Kurt F. Kline

General Information

The brown rockfish (Sebastes auriculatus) is a member of the family Scorpaenidae, one of the largest fish families in the western Pacific. The family is dominated by the rockfishes (Sebastes spp.), a genus which is represented by over 50 species on the northwest Pacific coast.

The brown rockfish is the most common rockfish in San Francisco Bay (Alpin 1967, Wang 1986), and the Bay appears to be an important nursery area for juveniles (Kendall and Lenarz 1986, Baxter 1999). Brown rockfish are the most common rockfish caught by sport anglers in the Bay (W. Van Buskirk, pers. comm) and the third most frequently caught rockfish in the San Francisco region (Karpov et al. 1995). Most brown rockfish are caught by anglers fishing from partyboats, skiffs, piers, and the shoreline (Miller and Gotshall 1965, Karpov et al. 1995). It is also a minor, but important, component of the nearshore commercial fishery; in the San Francisco area, the majority of brown rockfish are caught by hook and line for the live or whole fresh fish markets. Since the early 1990s, the brown rockfish has been the most common species sold in the live in San Francisco markets (C. Ryan, pers. comm.).

Reproduction

All rockfishes, including the brown rockfish, are viviparous. Fertilization is internal and the larvae develop in the egg capsule within the ovarian cavity. The larvae hatch within the ovary and are released with little yolk remaining and ready to feed. The embryos develop in 40-50 days after fertilization and the larvae hatch about 1 week before extrusion (Kendall and Lenarz 1986). Brown rockfish larvae are 4.7-6.7 mm at hatching (Delacy et al. 1964) and pelagic for several months. Although brown rockfish fecundity is not known, Sebastes females typically produce 100,000 to 1,000,000 eggs per brood (Kendall and Lenarz 1986). Brown rockfish may have multiple broods within one year, with parturition from December-January and May-July in Central California (Wyllie-Echeverria 1987).

Although gravid brown rockfish have been collected in San Francisco Bay, most parturition is believed to occur in coastal waters (Kendall and Lenarz 1986, Wang 1986). In San Francisco Bay, mature females were observed in winter and spring and larvae have been collected in winter and spring (Wang 1986).

Growth and Development

Brown rockfish juveniles are pelagic until 20-30 mm, whereas older juveniles settle out of the water column and are strongly associated with some type of physical structure (Turner et al. 1969, Kendall and Lenarz 1986). Pelagic juveniles have been collected in nearshore coastal waters from April through June, while benthic juveniles are common in nearshore coastal waters and the Bay (Kendall and Lenarz 1986). In San Francisco Bay, age-0 juveniles were usually first collected from April to July and were common through summer and fall (Wang 1986, Baxter 1999).

Juvenile brown rockfish apparently spend several years in a very restricted home range in the Bay and gradually move to deeper waters and nearshore. Juvenile brown rockfish tagged in the Bay have been recaptured more than 80 km away in nearshore coastal waters (Kendall and Lenarz 1986).

Both male and female brown rockfish reach maturity as early as age 3 (260 mm TL); half reach maturity at age 5 (310 mm TL); and all are mature at age 10 (380 mm TL) (Wyllie-Echeverria 1987). Both sexes grow at similar rates and reach a maximum size of about 550 mm TL (Miller and Lea 1972). In southern California, the oldest male was 18 years, the oldest female 20 years (Love and Johnson 1998).

Food and Feeding

In San Francisco Bay, smaller juvenile brown rockfish (<130 mm TL) prey primarily upon small crustaceans, including amphipods, copepods, caridean shrimp, and Cancer crabs. Larger fish (130-310 mm TL) prey upon larger crustaceans (caridean shrimp, Cancer crabs, Upogebia) and fish (Ryan 1986).

Distribution

The brown rockfish ranges from Hipolito Bay, Baja California, to southeast Alaska (Miller and Lea 1972). It most often solitary, but may be found in small aggregations (Love and Johnson 1998). In the ocean, it is most common in shallow rocky reefs (5-20 m), but also found over sand flats near eelgrass and in kelp beds while in bays and estuaries it is found near piers and over rubble (Feder et al. 1974, Matthews 1990, Love and Johnson 1998).
In San Francisco Bay the brown rockfish is found primarily in Central San Francisco Bay, to a lesser degree in South San Francisco and San Pablo bays, and occasionally in Carquinez Strait and western Suisun Bay (Ganssle 1966, Messersmith 1966, Wang 1986, Baxter 1999).

Suitable habitat and salinity are the primary factors influencing distribution of brown rockfish in the Bay. Benthic juveniles and adults are strongly associated with structure, including rocky reefs, piers and jetties, breakwaters, and riprap. In the Bay, most brown rockfish were collected at salinities >20l (median 28.3l, 90th percentile 31.8l, 10th percentile 21.5l, Baxter 1999, CDFG, unpubl. data).

Population Status and Influencing Factors

There is a modest brown rockfish population in the San Francisco Bay region. San Francisco Bay is a nursery area for brown rockfish, and most juveniles immigrate to the Bay from the nearshore coastal area soon after settlement. It is not clear if resident adult brown rockfish spawn successfully in the Bay. Juveniles rear in the Bay for several years, and the population is comprised of several year classes. But there is no reliable index or measure of year class strength in the Bay, as brown rockfish are strongly associated with structure, and are undoubtedly undersampled by trawls or other towed nets typically used by research studies.

Trophic Levels

Secondary carnivore. Feeds primarily on crustaceans and fishes.

Proximal Species

Prey: Crustaceans (caridean shrimp, Cancer crabs, Upogebia, amphipods, copepods), polychaetes, fishes, herring eggs.
Predators: Larger predatory fishes, including striped bass.

Good Habitat

Structure, including piers and rocky shores, in the higher salinity regions of the Bay

Acknowledgments

Some of the material in this report was summarized from the brown rockfish chapter in IEP Technical Report 63, which is referenced below (Baxter 1999).

References

Wang, J.C.S. 1986. Fishes of the Sacramento-San Joaquin Estuary and adjacent waters, California:

Personal Communications
Wade Van Buskirk. Recreational Fisheries Information Network Database, Pacific States Marine Recreational Fisheries Monitoring
Connie Ryan. California Department of Fish and Game, Ocean Fisheries Research Unit - Menlo Park

Pacific Staghorn Sculpin
Leptocottus armatus armatus
Robert N. Tasto

General Information
The Pacific staghorn sculpin (Family: Cottidae) is found from Kodiak Island, Alaska to San Quintin Bay, Baja, California (Miller and Lea 1972). It is the only true euryhaline species among the California cottids (CDFG 1987), and appears to move freely between fresh and saltwater environments (Moyle 1976). It is regarded as a nuisance species by many sportfishermen, but has shown some limited value as bait for gamefish (particularly striped bass) in the Estuary. Bolin (1944) recorded its maximum depth of capture offshore coastal California at 300 feet. It is a target species of the National Status and Trends Program (Emmett et al. 1991), as it is considered an indicator of stress in the estuarine environment, and may spend its entire life in Pacific coast estuaries.

Reproduction
Pacific staghorn sculpin may reach sexual maturity in their first year, and sex ratios within a population appear to favor females slightly (Boothe 1967, Tasto 1975). In northern California, spawning begins in October (Tomales Bay) or November (San Francisco Bay), peaks in January-February, and ends in March (Jones 1962). Fertilization is external. Staghorn sculpin eggs are adhesive and laid in shallow subtidal and intertidal waters. Fecundity averages 5,000 eggs per female (Jones 1962), and ranges from 2,000 to 11,000 eggs per female (Moyle 1976). Eggs range from 1.36 to 1.50 mm in diameter and hatch in 9 to 14 days at 15.5°C (Emmett et al. 1991).

Growth and Development
At hatching, Pacific staghorn sculpin larvae range from 3.9 to 4.8 mm total length (TL) (Jones 1962). Metamorphosis to the juvenile begins after about 2 months, when the larvae are 15 to 20 mm standard length (SL) (Emmett et al. 1991). The juvenile size range is approximately 20 to 120 mm TL (Jones 1962), and there appears to be considerable overlap in the length distribution of 0+ and 1+ fish, particularly in the summer and fall (CDFG 1987). The staghorn sculpin reaches maturity at about 120 mm TL its first year, and can grow to over 200 mm TL (3 years old) in California (Jones 1962). In southern California, growth was determined to be curvilinear (Tasto 1975). The largest specimen recorded was about 30 cm (Barnhart 1936).

Food and Feeding
Pacific staghorn sculpin larvae are planktivorous (Emmett et al. 1991). The juvenile and adult forms are, however, demersal predators, particularly over intertidal and shallow subtidal mudflats, and have been shown to feed on a variety of non-burrowing benthic organisms (Jones 1962, Boothe 1967, Tasto 1975). Feeding behavior of the staghorn sculpin is thought to be continuous, although there appears to be a preference for feeding at night (Tasto 1975). The principal food items for staghorn sculpin within San Francisco Bay were found to be bay shrimp (Crangon spp.), bay goby (Lepidogobius lepidus), mud crab (Hemigrapsus oregonensis), callianassid shrimp (i.e., Upogebia), and a variety of amphipods, isopods, and polychaetes (Boothe 1967). Elkhorn Slough studies showed predation on epifaunal crustaceans and infaunal and epifaunal worms (Barry et al. 1996). In southern California, major food items were similar to Elkhorn Slough and San Francisco Bay, including callianassid shrimp (i.e., Callinass sp.), mud crab, and arrow goby (Clevelandia ios) (Tasto 1975). Jones (1962) found that in Tomales Bay, staghorn sculpin fed heavily upon Upogebia and Crangon shrimp. In Grays Harbor, Washington, the staghorn sculpin’s diet consisted of amphipods, crangonid shrimp, small fish, Upogebia sp., juvenile Dungeness crab, and polychaetes (Armstrong et al. 1995). Several studies indicate that the staghorn sculpin...
is an important prey item for aquatic birds, particularly the great blue heron (Tasto 1975, Bayer 1985, Emmett et al. 1991).

**Distribution**

Pacific staghorn sculpin have been collected in all four subregions of the Bay. Larval abundance was determined to be highest from December through March, peaking in February, in various parts of the Estuary south of the Carquinez Bridge (CDFG 1987). Small juveniles are often found intertidally; catch patterns suggest that, during their first year, these early post larval forms move gradually from shallow inshore areas to deeper Bay waters (CDFG 1987, Emmett et al. 1991). In studies conducted in Yaquina Bay, Oregon, young-of-the-year first appeared in December, and were collected through April (Bayer 1985). Juveniles and adults are most frequently captured in central Bay and San Pablo Bay, and are more abundant in the channels in winter, and on the shoals in spring and summer (Figures 2.5 and 2.6). Adults experience their widest distribution during high Delta outflow, and it appears that a portion of the adult population moves out of the Estuary by late spring of their second year (CDFG 1987). In Elkhorn Slough (Monterey County), staghorn sculpin were highest in abundance, and frequently the dominant species, at sampling stations furthest inland, near sources of fresh water (Yoklavich et al. 1991). A tidal marsh population studied in Anaheim Bay, a relatively small embayment in southern California with little freshwater input, was composed almost entirely of juveniles (Tasto 1975). Pacific staghorn sculpin can also be found a mile or two up coastal streams in association with exclusively freshwater species (Moyle 1976).
Population Status and Influencing Factors

Multiple gear catch statistics from 1980-85 showed that Pacific staghorn sculpin was the most abundant of all the sculpins caught in the Estuary, and approximately 4% of all fishes caught by otter trawl and beach seine (CDFG 1987). The highest abundance of larvae noted in this study occurred during years of low Delta outflows, yet juvenile and adult numbers showed no quantifiable relationship to magnitude of flows (CDFG 1987). Larval success is thought to be the determining factor in overall recruitment to local populations (Emmett et al. 1991).

Trophic Levels

Larvae are first and second order consumers (Emmett et al. 1991). Adults and juveniles are higher order consumers.

Proximal Species

Predators: Diving ducks, great blue heron, western grebe, Caspian tern, loons, cormorants, gulls, marine mammals.

Prey: Crangon shrimp (principal prey item), bay goby (prey of large adults), mud crab, callianassid shrimp, amphipods (juvenile prey item, dominant in fresh water).

Competitor: Starry flounder.

Good Habitat

Success of local staghorn sculpin populations depends upon the quality and quantity of suitable habitat. Newly settled juveniles use intertidal and shallow subtidal mudflats for protection and feeding (Tasto 1975), although older juveniles and adults are said to prefer more sandy substrates and somewhat deeper waters (Bayer 1981, Emmett et al. 1991). Pacific staghorn sculpin are known to bury themselves in soft substrates, and have been found buried in mudflats after the tide has retreated (Tasto 1975, Bayer 1985). Staghorn sculpin have also been found associated with eelgrass (Bayer 1981).

Water quality factors are equally important for successful populations. Demersal eggs hatch most successfully at 26 ppt and larvae survive best at 10 to 17 ppt (Jones 1962). Greatest catches of larvae were in surface salinities of 18 to 30 ppt (CDFG 1987). The juvenile stage appears to be the most euryhaline, with the maturing and adult forms most likely to be found in the higher salinity waters (CDFG 1987, Emmett et al. 1991). Laboratory experiments have shown that adult \textit{L. armatus} can survive 67.5 ppt at 12°C, but gradually lose their tolerance of high salinities as temperatures rise to 25°C (Morris 1960). Since larval development is planktonic, it does not appear that, under normal conditions in the San Francisco Estuary, either temperature or salinity are very limiting to distribution.

References


Prickly Sculpin

Cottus asper

Bruce Herbold

General Information

Sculpins (Family: Cottidae) are specialized for living on the bottom, generally hiding in the nooks and crannies among rocks or rooted vegetation. Their large, flattened heads and proportionally small bodies, their fan-shaped pectoral fins, and their lack of an air bladder allow sculpins to hold their position even in wave-swept coasts or high-velocity mountain streams. The use of such habitats, combined with their secretive habits and cryptic coloration, make sculpins difficult to see by predators, prey, or inquisitive fish biologists. The large mouth relative to body size permits sculpins to consume prey almost as large as themselves. Sculpins are found in the northern Pacific Ocean and New Zealand. Most members of the family are marine but a number of species (most in the genus Cottus) occur in the fresh waters of North America.

Reproduction

Sculpins generally spawn in the late winter or early spring, although some upstream populations seem to delay spawning into the early summer (Wang 1986). Male sculpins prepare for spawning by moving downstream and establishing a nest site where they clean off some kind of overhanging structure such as a flat rock, tule root, or beer can (Kresja 1965, Moyle 1976). Females then enter the spawning area and, after a nocturnal courtship, attach their eggs to the prepared overhanging structure. Females produce between 280 and 11,000 eggs (Patten 1971), but one male may court many females and end up with up to 30,000 eggs (Kresja 1965). Males stay in the nest protecting the eggs and circulating water around them until they hatch. Hatching rates appear to improve in saltier water (Millikan 1968). After hatching, the larvae become planktonic and are carried further downstream. Young sculpins (15-30 mm SL, Broadway and Moyle 1978) settle to the bottom and begin a general upstream movement (Mclarney 1968, Mason and M. acidihi 1976). The amount of movement associated with spawning appears to vary tremendously among sculpin populations (Wang 1986). Earlier observers suggested that substantial downstream movements were only found in coastal populations, not in the Central Valley (Kresja 1967). However, very high densities of newly hatched prickly sculpins have been reported from the Delta and Suisun Bay (Turner 1966, Wang 1986), as well as in upstream sites (Wang 1986) which has led to the conclusion that the Central Valley contains both 'migratory' and 'non-migratory' populations. Recent studies suggest that the same may be true in coastal streams, such as the Eel River, where young prickly sculpins were found 100 km above the river mouth (Brown et al. 1995). Regardless of the degree to which they move for spawning, mainstem rivers appear to be an important habitat for most prickly sculpin populations. Young prickly sculpins are often found in saline water at the tributary mouths in spring months (Leidy pers. comm.).

Growth and Development

Fry at hatching average six mm total length. Newly emerged fry swim soon after hatching and appear to drift downstream as plankton for three to five weeks. This early developmental pattern leads to high concentrations in the slower waters of the Delta (Turner 1966). Young fish assume a bottom-feeding existence at sizes of 20 to 30 mm, at which time they appear to begin moving upstream (Mclarney 1968).

Food and Feeding

Sculpins have a reputation amongst anglers as predators on salmonid eggs and fry (Munro and Clements 1937, Shapirolov and Tait 1954, Reed 1967) which is probably undeserved (Moyle 1976, 1977). Diet studies generally show that sculpins prey principally on invertebrates, with younger prickly sculpins eating planktonic crustaceans and older fish eating larger, benthic animals and small fish (M. oyle 1976). In Suisun Marsh their diet is predominately benthic amphipods of the genus Gammarus (Herbold 1987).

Distribution

Prickly sculpins are found in fresh to brackish water from the Kenai Peninsula in Alaska to the Ventura River in southern California. In California's Central Valley, they can be found in the lower reaches of most foothill streams. Prickly sculpins often overlap in distribution with the similar riffle sculpin (Cottus gulosus) which is
found more in upper elevations. Neither is found in the upper Pit River drainage. Their range includes tidal habitats of brackish salinity, such as Suisun Marsh. Prickly sculpins are found from headwaters to the mouths of many of the small tributaries that flow into San Francisco Bay, (including Alameda Creek, Walnut Creek, Corte Madera Creek, Coyote Creek and the Petaluma River; Leidy 1984).

**Habitats**

Like freshwater sculpins generally, prickly sculpins use very diverse habitats from small headwater streams to coastal estuaries, and are widely distributed from Alaska to southern California (Moyle 1976). Whatever the habitat, prickly sculpins usually are found under some sort of cover: rocks in streams, vegetation in pools and marshes, or simply at depth in lakes and reservoirs (Moyle 1976, Brown et al. 1995).

**Population Status and Influencing Factors**

Many of the most recent, successful invading species of the Estuary have the potential to affect prickly sculpins. In 1986, the Asiatic clam (Potamocorbula amurensis) began a rapid and thorough domination of the benthic community. Although the decline in abundance of other benthic species has been well-documented, there is no information on the impact of these changes on the diet, distribution, or abundance of prickly sculpin. Also in the mid-1980s, the Estuary was invaded by the shimofuri goby (Tridentiger bifasciatus) which lives in the same kinds of habitats and microhabitats as prickly sculpin. However, the very small mouth of the goby reduces the likelihood of interspecific competition. Since 1996, mitten crabs (Eriochir sinensis) have become extremely abundant and are voracious and indiscriminate predators on benthic organisms. Mitten crabs undergo an annual upstream migration to spawn that results in a large overlap with the range of prickly sculpins. In the Eel River of northern California, it appears likely that the introduction of predatory pikeminnows (Ptychocheilus grandis) has resulted in a substantial change in sculpin behavior when compared to the similar Smith River (Brown and Moyle 1991, Brown et al. 1995, Whited and Harris in press). In the tributary creeks of the San Francisco Bay drainage, prickly sculpins are often associated with native species and are usually absent in areas where large non-native predatory fish are found (Leidy 1984). No work has been done to document interactions of prickly sculpin with the vastly changed benthic community of the Central Valley.

Habitat changes and degradations of water quality are associated with a restricted range of prickly sculpins in the San Joaquin River watershed (Brown 1998). Sculpins are part of an assemblage of native species that are characteristic of smaller San Joaquin tributaries that have suffered little change in habitat structure or water quality. Unfortunately, the close associations of land use practices, habitat alteration and water quality degradation in the rest of the watershed make it impossible to identify the effects of individual environmental variables on sculpin biology.

As in the San Joaquin River, prickly sculpins in Suisun Marsh tend to be found most often in association with other native fishes and in less disturbed habitats (Herbold 1987). However, the actual physical parameters of low dissolved solids and high gradient that characterize usual sculpin sites in the San Joaquin River, are absent in Suisun Marsh. This suggests that the impacts of land use and disturbance on the distribution and abundance of prickly sculpins are not simple and that the parameters that reflect disturbance in one area may not be causally connected to the parameter of importance to sculpins in that area.

California’s immense water projects appear to have had little effect on prickly sculpins. Construction of dams has isolated populations and prevented the downstream movements exhibited elsewhere, but populations have remained large in the warmwater reservoir behind Friant Dam. Prickly sculpins are also found in stream habitats upstream of impassable dams on a number of other Central Valley streams. Water export from the Delta has resulted in the establishment of new populations of prickly sculpins within the facilities of the state and federal projects, as well as within aquatic habitats in southern California outside the historic range of prickly sculpin (Wang 1986). The impacts of these introductions on the native species in southern California streams have been little studied.

**Trophic Levels**

Prickly sculpins are secondary and tertiary consumers.

**Proximal Species**

**Predators:** Centrarchids and pikeminnows.

**Prey:** Planktonic crustacea (for young); benthic invertebrates, particularly gammarid amphipods; neomysis; juvenile fish.

**Habitat:** Emergent aquatic vegetation (root masses).

**Good Habitat**

In contrast to staghorn sculpins (Leptocottus armatus), prickly sculpins larger than 20 mm are usually found in association with some kind of complex, physical cover. In upstream sites, cover consists of interstices in cobble, root wads and woody debris and even discarded soda cans and tires. In downstream sites, cover usually consists of root wads of emergent aquatic vegetation. Although
more tolerant of salinity than most other California freshwater fish, sculpins are seldom found in salinities greater than 10 ppt.

References


Personal Communications

Robert Leidy, U.S. Environmental Protection Agency
Striped Bass
*Morone saxatilis*
Ted R. Sommer

**General Information**

Striped bass (Family: Percichthyidae) were introduced into the Estuary in 1879, leading to a successful commercial fishery within 10 years (Herbold et al. 1992). The commercial fishery for striped bass was banned in 1935 following a substantial decline in abundance which appears to have begun at the turn of the century. The species are presently the principal sport fish caught in San Francisco Bay and is estimated to bring approximately $45 million per year into local economies in the Estuary.

**Reproduction**

Striped bass are present in the San Francisco Estuary throughout the year (Moyle 1976). They generally congregate in San Pablo and Suisun Bays in autumn and move into the Delta and Sacramento River system on their spawning migration during winter and early spring. The timing and location of spawning depends on temperature, flow and salinity, but typically peaks in May and early June. The annual spawning distribution appears to shift between the Sacramento and San Joaquin rivers and the Delta.

Striped bass spawn in freshwater, with optimum spawning at salinities of less than 1 ppt (Moyle 1976). The species has exceptionally high fecundity—females commonly broadcast from 0.5 to 4.5 million semi-buoyant eggs into the water column. The drifting eggs hatch in the current in about 2 days. Eggs and newly-hatched larvae are carried downstream to the Delta and Suisun Bay. Larvae show peak abundance at the upstream edge of the entrapment zone, located at a salinity of approximately 2 ppt.

**Growth and Development**

Striped bass grow to about 38 mm by late July or August (Moyle 1976). They typically reach 23 to 35 cm FL by their second year, 38 to 39 cm fork length (FL; the measure to the bottom of the fork of the tail fin) on their third year, and 48 to 50 cm in their fourth year. Growth of older adults is 1 to 3 cm annually. Most females mature at four to six years, but many are mature by the end of their third year. Males typically mature at two to three years old. Although striped bass apparently have the potential to live in excess of 30 years, most adults are three to seven years old.

**Food and Feeding**

Striped bass are gregarious pelagic predators (Moyle 1976). They begin feeding at a length of 5-6 mm on several invertebrates including cladocerans and copepods. Copepods generally dominate the diet of 7 to 11 mm larvae, but the opposum shrimp, *Neomysis*, become a more important food source in larger individuals. Young-of-the-year feed mostly on opposum shrimp, but amphipods, copepods, and threadfin shad are important alternative prey items. Fish gradually become a more important food source in juvenile bass (13 to 35 cm FL). Sub-adult and adult bass (age 2+) are primarily piscivorous, although they are highly opportunistic depending on prey availability.

**Distribution**

In contrast to the coastal Atlantic populations of striped bass, most of the local population spend their lives in the San Francisco Estuary. However, recent tagging studies suggest that striped bass are spending more time in Suisun Bay, the Delta, and surrounding freshwater areas (Sweetnam 1990). The current distribution of the species includes San Francisco Bay, San Pablo Bay, Suisun Bay, the Delta, tributaries of the Sacramento River and the Pacific Ocean (Herbold et al. 1992).

**Population Status and Influencing Factors**

Adult abundance has declined over the past 30 years, from over 1.5 million in the late 1960s to about 0.5 million in recent years (CDFG 1992). The decline was most dramatic between the beginning and the end of the 1970s, prompting the initiation of a hatchery stocking program to supplement natural production (Harris and Kohlhorst 1996). Stocking was conducted from 1981 through 1991—hatchery fish presently comprise a substantial percentage (e.g., 35% of the 1990 year class) of the adult population.

Year class abundance is assumed to depend on the environmental conditions experienced by the eggs and young fish (CDFG 1987, 1992). However, a steady decline in the survival rate of yearlings stocked into the Estuary suggests that habitat conditions for older fish also play an important role (Harris and Kohlhorst 1996). Abundance of young bass is strongly correlated with
Delta outflow and entrapment zone position, although in recent years this relationship has deteriorated. For example, in 1995 striped bass production was exceptionally poor despite wet conditions that increased the abundance of several other outflow-dependent species. Entrainment at diversions is known to be substantial, and there is statistical evidence that these losses affect abundance. Nonetheless, losses at the projects during the 1980s were at least partially mitigated using hatchery fish, yet the population decline has continued. The reduction in several invertebrate prey species has also been dramatic, particularly since the introduction of the Asian clam Potamocorbula. The decline in survival of stocked fish strongly suggests that competition for food has had an effect on the population. Other potentially important factors include toxic substances, exotic species and illegal fishing.

**Trophic Levels**

Striped bass are secondary and higher order consumers.

**Proximal Species**

**Major Prey Items:** Zooplankton (cladocerans and copepods), terrestrial insects, opossum shrimp (*Neomysis mercedis*), splittail, salmon, threadfin shad, American shad.

**Good Habitat**

Striped bass are able to tolerate a wide range of environmental conditions, illustrated by their ability to move regularly between salt- and fresh-water (Moyle 1976). Optimal temperatures for spawning appear to be from 15.6° to 20.0° C. Low oxygen (4 ppm) and high turbidity are also tolerated. Large rivers or tidal channels with moderate water velocities are required to keep the eggs and larvae suspended in the water column. Young-of-the-year striped bass show highest abundance in the entrapment zone, the region where fresh- and salt-water mix.

**References**


**White Croaker**

*Genyonemus lineatus*

Kurt F. Kline

**General Information**

The white croaker (Family: Sciaenidae) is found in small schools (Skogsberg 1939) and ranges from Magdalena Bay, Baja California, to Mayne Bay, Vancouver Island, British Columbia (Baxter, 1980, Hart 1973, Miller and Lea 1972). It is abundant in San Francisco Bay, and supports both commercial and sport fisheries in nearshore coastal waters, and a sport fishery in the Bay.

**Reproduction**

Approximately 50% of all white croakers are mature after their first year and all are mature by their fourth year (Love et al. 1984). Along the coast, spawning appears to take place in water from eight to 36 meters deep (Love et al. 1984). In San Francisco Bay spawning occurs from September through May (Wang 1986), with most yolk-sac larvae (YSL) collected from November through March (CDFG, unpub. data). Females batch spawn 18-24 times per season, with a batch consisting of 800-37,200 eggs (Love et al. 1984).

**Growth and Development**

White croaker eggs are pelagic, spherical and transparent. Under laboratory conditions (~20°C), eggs hatched in 52 hours. The newly hatched YSL are poorly deve-
oped, but by the sixth day the yolk-sac is absorbed, the swim bladder is inflated and feeding begins (Watson 1982).

Throughout their life, white croaker growth is fairly constant (Love et al. 1984). They may live to 12 years (Love et al. 1984) and reach a total length (TL) of 41.1 cm (Miller and Lea 1972).

**Distribution**

Along the coast, the greatest densities of larvae are found near the bottom between 15 and 20 meters. The smallest juveniles are common from 3 to 6 meters, and move to deeper water as they grow. Most adults are found in waters less than 30 meters, although white croakers have been recorded to 183 meters (Love et al. 1984).

Within San Francisco Bay, most of the pelagic eggs and YSL are found in Central Bay (Wang 1986; CDFG, unpub. data). As the larvae develop to the post yolk-sac stage, they move toward the bottom. Tidal currents probably transported white croaker larvae to South and San Pablo bays. High outflow events during the winter, which increases the gravitational currents, may increase the transport of larvae to San Pablo Bay (Fleming, pers. comm.). By September, most of the young-of-the-year (YOY) migrate to Central Bay and by winter, emigrate from the Bay (Fleming 1999).

Within the Bay, YOY white croaker are found at lower salinities and higher temperatures than the one year and older fish (1+), reflecting the broader distribution of YOY. The movements of older YOY and 1+ white croaker out of the Bay during the late fall and winter may be temperature related.

**Population Status and Influencing Factors**

The California Department of Fish and Game’s Bay Study has generated annual abundance indices for white croaker since 1980. The abundance of YOY white croaker has fluctuated greatly over the past 19 years (Figure 2.7). Highest abundance indices of YOY were in 1980, 1986, 1992, 1993, and 1994. White croaker 1+ indices peaked between 1988 and 1991.

From 1981 to 1986, white croaker 1+ catches were dominated by the 1980 year class and from 1987 to 1993, they were dominated by the large 1986 year class. However, the relative size of a year class as YOY is not indicative of the future abundance of 1+ fish in the Bay. For example, the 1986 year class apparently contributed to the subsequent 1+ indices more than either the 1980 or the 1993 year classes. The drought from 1987-1992 may have caused greater use of the Bay by the 1986 year class than either 1980 or 1993 year classes.

Examination of the annual indices shows no relationship between the number of mature fish and YOY, while the length frequency data shows that single year classes tend to dominate subsequent years’ 1+ catch and the monthly catch per unit effort (CPUE) shows seasonal migration patterns within and out of the Bay. From these data, one could draw the following conclusions: 1) the white croaker “population” within the Bay is an extension of the nearshore coastal population; 2) factors that influence the nearshore population of white croaker are independent of the Bay; and 3) factors that influence the Bay “population” appear to be the salinity, temperature and, perhaps most importantly, the size and distribution of the nearshore population.

**Trophic Levels**

Secondary consumers.

**Proximal Species**


**Good Habitat**

White croaker are associated with soft substrates (Love et al. 1984). In the Bay, white croaker are primarily found in areas with the most marine-like (salinity and temperature) conditions.

**Acknowledgments**

Some of the materials in this report are summarized from the white croaker chapter of IEP Technical Report 63 (Flemming 1999).

**References**


**Personal Communications**

Kevin Fleming, California Department of Fish and Game, Bay-Delta and Special Water Projects Division, Stockton, CA.

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**Shiner Perch**

*Cymatogaster aggregata*

Michael F. McGowan

**General Information**

The shiner perch (Family: Embiotocidae) is a small but abundant species common to the intertidal and subtidal zones of bays, estuaries, and the nearshore regions of California. They are commonly caught by anglers around rocks, and pilings, from shore and docks, and just about any fishing area. They are also used as live bait in the San Francisco fisheries for striped bass and California halibut.

**Reproduction**

The shiner perch, like other members of the family Embiotocidae, is a live-bearer. Mating is accompanied by elaborate courtship behavior and occurs primarily in the spring and summer in California (Shaw 1971). Females give birth during April and May (Odenweller 1975) in California. Fecundity ranges from 5-36 young per female, depending on size (Emmett et al. 1991).

**Growth and Development**

At birth, the fully developed young are 34.0-43.7 mm long (Wang 1986). Juveniles become adults at 5 cm in length. Growth is rapid the first year but slows subsequently (Odenweller 1975). Most females mature their first year. They may live 8 years and reach 20 cm long. Males mature soon after birth and rarely grow beyond 13 cm.

**Food and Feeding**

Embryos receive nutrition and gas exchange through ovarian placenta tissues and fluids. Juveniles and adults feed on plankton and benthos depending on availability. Prey items include copepods, isopods, amphipods, mussels, barnacle appendages, mysids, crab larvae, and other small invertebrates or protruding parts of invertebrates (Emmett et al. 1991).
Distribution

The shiner perch occurs near shore and in bays and estuaries from Baja California to Alaska commonly associated with aquatic vegetation. Juveniles prefer intertidal and shallow subtidal habitats in bays and estuaries (Moyle 1976). In winter they may move out of estuaries to nearshore areas and have been found as deep as 70 m (Hart 1973). In Elkhorn Slough, where they are a numerically dominant component of the fish fauna, they were classified as partial residents (Yoklavich et al. 1991).

Population Status and Influencing Factors

The availability and quality of estuarine areas for giving birth and rearing young may limit populations. Key factors are water temperature, not excessively hot (Odenweller 1975), and seagrass beds for shelter and feeding. San Francisco Bay shiner perch catches in trawl surveys declined in 1983, perhaps due to high outflow (and resulting low salinity) that year (Herbold et al. 1992). Because it uses nearshore areas, the shiner perch may have high body burdens of pesticides and other compounds (Earnest and Benville 1971), but population effects of chronic pollution have not been documented.

Trophic Levels

Shiner perch are secondary and higher level consumers. Plant matter found in some stomach analysis studies may be due to feeding on invertebrates that occur on the aquatic vegetation.

Proximal Species

Predators: Sturgeon spp., salmon spp., striped bass, California halibut, cormorant spp., great blue heron, bald eagle.
Prey: Copepods, isopods, amphipods, mussels, barnacle appendages, mysids, crab larvae, and other small invertebrates or protruding parts of invertebrates.

Good Habitat

The shiner perch appears to favor aquatic vegetation if present, but is also found over shallow sand and mud bottoms. They prefer salinities greater than 8-10 ppt and were reported in water temperatures ranging from 4 to 21°C (Emmett et al. 1991). In San Francisco Bay, they are widespread but are most abundant downstream of the Carquinez Strait. Herbold et al. (1992) considered them a euryhaline species. Eelgrass beds may be important feeding areas because shiner perch use them more at night than during the day (Bayer 1979).

References

Tule Perch
Hysterocarpus traskii
Robert A. Leidy

General Information
Tule perch are the only viviparous freshwater fish native to California and the only freshwater member of the surfperch family (Embiotocidae) (Baltz and Moyle 1982). They are deep-bodied, spiny-rayed fish found in lakes, rivers, streams, and estuaries in habitats characterized by complex cover, especially well developed beds of aquatic macrophytes (Moyle 1976). There are three recognized subspecies of tule perch, H. t. pomo from the Russian River drainage, H. t. lagunae from Clear lake, and H. t. traskii from the Sacramento-San Joaquin drainage, which includes populations found in the San Francisco Estuary (Hopkirk 1973, Baltz and Moyle 1981 and 1982). Because of their small size, tule perch have no commercial and limited sport value.

Reproduction
Tule perch breed during July through September, but fertilization of the eggs is delayed within the female until January (Bundy 1970, Bryant 1977). Embryos develop within the females ovarian compartments and are born as juveniles in May or June, at a length of between 30-40 mm standard length (SL) (Bryant 1977). The number of fish produced per female is positively correlated with the size of the female fish and ranges between 22 and 93 (Bundy 1970, Bryant 1977).

Growth and Development
Juveniles begin schooling immediately following birth within aquatic vegetation, submerged logs, or boulders (Wang 1986). It is not known whether juveniles move into tributaries following birth, but it is interesting to note that several streams feeding into Suisun Marsh and San Pablo Bay contain large numbers of juvenile tule perch (Leidy, pers. observ.). Juveniles grow rapidly and individuals in the Sacramento-San Joaquin Delta may reach 80 to 100 mm SL following the first year of growth (Moyle 1976). Maximum size for tule perch is approximately 160 mm SL, although a single individual measuring 175 mm SL was collected in Napa Slough, Napa County (Leidy, unpub. data). Tule perch rarely live longer than five years (Moyle 1976).

Food and Feeding
Within the Sacramento-San Joaquin Delta and upper Estuary, tule perch feed primarily on mysid shrimp, small amphipods, midge larvae (Chironomidae), and clams (Cook 1964, Turner 1966). Hopkirk (1962) recorded that tule perch collected in brackish water habitats near the mouth of the Napa River feed mostly on small-sized brachyuran crabs, while juvenile fish feed predominantly on midge larvae and pupae. Tule perch are also known to feed on zooplankton, aquatic insects, and a variety of benthic and plant-dwelling invertebrates in lakes and rivers (Moyle 1976, Wang 1986).

Distribution
Tule perch are native to low-elevation valley waters of the Central Valley, the Sacramento-San Joaquin Delta, including Suisun Marsh and several streams tributary to the San Francisco Estuary, Clear Lake, and the Russian, Salinas, and Pajaro Rivers (Moyle 1976). Within the San Francisco Estuary tule perch have been recorded from Suisun Marsh (Herbold et al. 1992), including Montezuma Slough, Suisun Bay (Ganssle 1966), Carquinez Strait (Messersmith 1966), the Napa River and its marshes (Moyle 1976; Leidy 1984; Leidy, unpub. data), and Sonoma, Alameda, and Coyote Creeks (Leidy 1984). Tule perch may be considered locally abundant in lower estuarine and riverine intertidal marsh and pelagic habitats of Suisun Marsh and several of its tributary streams, the Napa and Sonoma Creek marshes, and portions of San Pablo Bay (Leidy, unpub. data). Tule perch no longer occur in the Pajaro and Salinas rivers, and are rare in Alameda and Coyote Creeks (Leidy, unpub. data).

Population Status and Influencing Factors
While the historical range of tule perch within the San Francisco Estuary has been reduced, tule perch are still locally abundant in Suisun Marsh and the Napa River and Sonoma Creek and its tidal marshes. Important factors negatively influencing population numbers likely include excess siltation and turbidity, reduced freshwater flows, pollution, removal of riparian vegetation and aquatic macrophytes through stream channelization and other flood control measures, and the resultant loss of nesting, feeding, and cover habitat, and possibly the introduction of exotic centrarchids (Moyle et al. 1995). Moyle et al. (1995) identified introduced fish predators, such as smallmouth bass (Micropterus dolomieui), pond...
and dam construction, and reduced flows and poor water quality as threats to the Russian River subspecies of the tule perch. These are likely threats to the other two subspecies of tule perch as well. Interestingly, otter trawl data collected in Suisun Marsh shows a significant decline in tule perch numbers during 1983-84, a year of extremely high outflow (Herbold et al. 1992).

**Trophic Levels**

Juveniles and adults are primary and higher order consumers.

**Proximal Species**

**Juvenile predators:** Other large freshwater and estuarine piscivorous fish, egrets, herons and other wading birds. **Prey:** Aquatic and terrestrial insects, zooplankton, mysid shrimp, amphipods, clams, brachyuran crabs, midge larvae and pupae.

**Good Habitat**

Tule perch may be found in a variety of habitats from the slow-moving, turbid channels of the Delta, marshes between the mouths of Sonoma Creek and the Napa River, to relatively clear, fast-flowing rivers and streams (Moyle 1976; Leidy, unpub. data). In tidal riverine marshes, tule perch prefer slow-moving backwater and slough habitats with structurally-complex beds of floating or emergent aquatic macrophytes, overhanging banks and/or submerged woody debris. These areas serve as important feeding and breeding habitats, as well as protective rearing areas (Moyle 1976). Structurally-complex cover appears to be essential for near-term females and juveniles as refugia from predators (Moyle et al. 1995).

Although Moyle (1976) states that tule perch seldom venture into brackish water, they are present in the pelagic zone of tidal riverine and intertidal estuarine environments, such as the Napa River marshes and Suisun Marsh (Leidy, unpub. data). This suggests that some populations of tule perch may be able to tolerate brackish water conditions, or at least utilize these areas when freshwater outflows dilute surface water. In Suisun Marsh tule perch are most frequently collected in the small, heavily vegetated, dead-end sloughs where introduced centrarchids are uncommon (Moyle et al. 1985).

**References**


_References_
Arrow Goby
Clevelandia ios
Kathryn A. Hieb

General Information

The arrow goby (Family: Gobiidae) is probably the most abundant native goby in San Francisco Bay. It ranges from the Gulf of California to Vancouver Island, British Columbia (Miller and Lea 1972) and is common to intertidal mudflats and shallow subtidal areas of bays, estuaries, and coastal lagoons. It is often commensal with burrowing invertebrates. The arrow goby grows to a maximum size of 45 to 50 mm total length (TL). This small fish is an important component of the intertidal food web, as it is a common prey item for a variety of birds and fishes. It has no sport or commercial value.

Reproduction

In Elkhorn Slough, ripe females were collected from December through August, but were most common from March through June (Prasad 1948). The reproductive period occurs approximately one to two months earlier in southern California— in Mission Bay, ripe females were collected from September through June, with peak abundance from November through April (Brothers 1975), while in Anaheim Bay, ripe females were collected from December through September, with peak abundance from February through June (Macdonald 1972). Ovary development is asynchronous, as ovaries are found in various stages of maturation during the spawning season (Macdonald 1972, Brothers 1975). This indicates that each female may spawn several times during the spawning season. Fecundity ranges from 800 to 1,200 eggs per female, with clutch size ranging from 150 to 350 eggs (Brothers 1975) or from 750 to 1,000 eggs (Prasad 1948).

Some disagreement exists in the literature on the deposition of the eggs and parental care. The eggs are either deposited on surfaces with no additional parental investment (Prasad 1948, Macdonald 1972) or deposited on the wall of burrows constructed by the male and guarded by the male until hatching (Brothers 1975). In Mission Bay, all males collected in January and February were brooding clutches of eggs in burrows. Typical of most gobies, the fertilized eggs are club-shaped, with an attachment thread at one pole. Hatching occurs in 10 to 12 days and the newly hatched larvae are pelagic (Prasad 1948, Brothers 1975).

Growth and Development

Newly hatched larvae range from 2.75-3.25 mm TL (Prasad 1948). Juvenile arrow gobies settle from the plankton at approximately 8 mm standard length (SL) and are found in burrows when they are 10-14 mm SL (Macdonald 1972). The arrow goby matures at one year and a length of 30 to 40 mm SL in Anaheim and Mission bays (Macdonald 1972, Brothers 1975); in Elkhorn Slough females begin to mature at 29 mm SL and all are mature at 34 mm SL (Prasad 1948). In southern California, most arrow gobies die after spawning, with a few living to two years (Macdonald 1972, Brothers 1975). In Elkhorn Slough, arrow gobies commonly live two to three years (Prasad 1948). Fish from Elkhorn Slough apparently spawn later, grow slower, mature later, and reach a larger size than fish from southern California populations (Brothers 1975).

Food and Feeding

The arrow goby preys on a variety of small invertebrates. In Mission Bay, the major prey items (percent occurrence) of juveniles and adults are harpacticoid copepods (88%), ostracods (58%), tanacians (32%), gammarid amphipods (19%), mollusc siphon tips (11%), caprellids (8%), nematodes (7%), and polychaetes (7%) (Brothers 1975). In Anaheim Bay, the most important prey items are harpacticoid copepods, nematodes, oligochaetes, ostracods, and cyclopoid copepods (Macdonald 1972). Larvae prey primarily upon the calanoid copepod Acartia tonsa (Macdonald 1975).

The arrow goby is preyed upon by a variety of demersal fishes, including Pacific staghorn sculpin (MacGinitie and MacGinitie 1949, Brothers 1975, Tasto 1975), California halibut (Haaker 1975, Drawbridge 1990), and diamond turbot (Lane 1975). MacGinitie and MacGinitie (1949) presumed probing shorebirds, including willets, godwits, and curlews would capture arrow gobies while exploring burrows at low tides. Arrow gobies have been found in the stomachs of greater yellowlegs and dowitchers (Reeder 1951).

Distribution

The arrow goby is common on mudflats inhabited by its invertebrate commensal hosts (Brothers 1975), with densities up to 20/m² in Anaheim Bay (Macdonald 1972). It apparently utilizes invertebrate burrows as a refuge from predators and as a temporary shelter during low tides. The arrow goby primarily inhabits burrows of the ghost shrimp (Callianassa californiensis), the...
fat innkeeper worm (*Urechis caupo*), the mud shrimp (*Upogebia spp*.), and various bivalves (Prasad 1948, Brothers 1975). Males also construct burrows for reproduction (Brothers 1975). At low tides the arrow goby is also common in remnant pools of water on the mudflats (Prasad 1948).

In San Francisco Bay, larval arrow gobies are most abundant in South and San Pablo bays, with few collected upstream of Carquinez Strait in years with low freshwater outflow (Wang 1986, CDFG 1987). Juveniles and adults are common in shallow subtidal and intertidal areas of South, Central, and San Pablo bays and have occasionally been collected in Suisun Bay (CDFG 1987). The arrow goby is also common in some tidal marsh habitats from South Bay to lower San Pablo Bay. It was the second most common species collected in otter trawl samples from Hayward Regional Shoreline Marsh channel sites (Woods 1981). The arrow goby was common in weir samples collected in Plummer Creek (South Bay near Newark), although gobies were not speciated in this study, so their relative abundance is unknown (Wild 1969). In a survey of Castro Creek, Corte Madera Creek, and Gallinas Creek marshes, the arrow goby was relatively common in otter trawl samples from creek channels and mudflats adjacent to the marshes, but rare in gill nets and not collected by minnow traps set in the marsh channels (CH2M Hill 1982).

A few arrow gobies were collected in Petaluma River marshes, Napa-Sonoma Marsh, but none in Suisun Marsh (CDFG; unpublished data; ANATEC Laboratories 1981; CH2M Hill 1996; Matern et al. 1996).

Arrow goby larvae have been collected year-round in San Francisco Bay, with peak larval abundance from April through July (CDFG 1987). Peak abundance in beach seine samples from the Bay is from March through August; these catches include settled juveniles and adults (CDFG 1987). In southern California, most juveniles settle in the spring (February through May in Mission Bay, February through June in Anaheim Bay), although juveniles have been collected all but one or two months in the fall (Macdonald 1972, Brothers 1975).

Population Status and Influencing Factors

Because the arrow goby is most common in intertidal and shallow subtidal habitats, it is more effectively sampled by seines than trawls. In a beach seine survey of San Francisco Bay conducted by California Department of Fish and Game in the 1980s, the arrow goby comprised approximately 4% of the catch, ranking eighth of all fishes collected. In contrast, it comprised only 0.04% of the fishes collected by the otter trawl (Orsi 1999). As the beach seine survey has been discontinued, there is no long-term monitoring program in the Bay that effectively samples the arrow goby, and its current status is difficult to assess. From 1981 to 1986, the arrow goby beach seine annual “abundance index” varied almost 10-fold, with the highest indices in 1981 and 1986 (Figure 2.8).

Brothers (1975) hypothesized that arrow goby abundance and distribution could be controlled by the abundance and distribution of the commensal invertebrates, especially the ghost shrimp. Because the arrow goby is an annual species, devoting a large proportion of its resources to reproduction (“r-strategist”), it would be expected to undergo large population fluctuations.

Trophic Level

Arrow goby larvae, juveniles, and adults are secondary consumers, preying primarily on small benthic and epibenthic invertebrates.

Proximal Species

**Predators:** Pacific staghorn sculpin, California halibut, diamond turbot.

**Prey:** Harpacticoid copepods, ostracods, tanacians, gammarid amphipods, mollusk siphon tips, nematodes, oligochaetes.

**Commensal Hosts:** Burrowing invertebrates. Bat rays and leopard sharks impact the abundance and distribution of burrowing invertebrates.

**Figure 2.8 Annual Abundance Indices of Arrow Goby from San Francisco Bay, Beach Seine (CDFG, unpublished data)**

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Good Habitat

Good habitat for the arrow goby is shallow subtidal and intertidal mudflats inhabited by the commensal invertebrate hosts. All habitats in tidal marshes may not suitable, as the arrow goby has been collected from larger channels and adjacent mudflats, but not from smaller order channels.

References


Bay Goby
Lepidogobius lepidus
Kathryn A. Hieb

General Information

The bay goby (Family: Gobiidae) ranges from Baja California to Vancouver Island, British Columbia (Miller and Lea 1976). It is common to bays and estuaries and often commensal with burrowing invertebrates on intertidal mudflats (Grossman 1979a). Because it often occupies burrows, the bay goby is not effectively sampled by trawls and seines and its relative abundance is undoubtedly greater than indicated by most surveys. It is the most abundant native goby in larval surveys of San Francisco, Humboldt, and Yaquina bays. The bay goby grows to approximately 100 mm total length (TL) and has no commercial or sport value.

Reproduction

Females with yolk filled eggs were collected from September through March, with the peak of reproductive activity from January through March in Morro Bay (Grossman 1979b). Gonadal development is asynchronous, typical of species that spawn several times a season and have a protracted spawning period. As for many other species of gobies from temperate waters, it is assumed the eggs are laid in burrows constructed by either the males or commensal invertebrate hosts and are guarded by the male until hatching (Wang 1986). Eggs are club shaped with an adhesive thread at one pole for attachment to the burrow wall or substrate.

In San Francisco Bay, larvae were collected throughout the year, with peak abundance from June to October (CDFG 1987). The period of peak abundance is similar in other Pacific Coast estuaries—peak larval abundance is from April to September in Yaquina Bay, Oregon (Pearcy and Myers 1974) and larvae were collected from April to September in Humboldt Bay (Eldridge and Bryan 1972). In San Francisco Bay, most larvae were collected in Central Bay and northern South Bay, with relatively few collected upstream of San Pablo Bay (CDFG 1987).

Growth and Development

Bay goby larvae are approximately 2.5-3.0 mm TL at hatching (Wang 1986). The larvae are planktonic for three to four months (Grossman 1979b) and settle to the bottom as juveniles at approximately 25 mm TL (Wang 1986). Although the bay goby is reported to grow to about 87 mm TL (Miller and Lea 1976), specimens as large as 108 mm TL have been collected in San Francisco Bay (CDFG, unpub. data). Some bay gobies reach sexual maturity by the end of their first year and by the end of their second year are mature (Grossman 1979b). Bay gobies reportedly live up to 7+ years (Grossman 1979b), although based upon length frequency data from San Francisco Bay (CDFG 1987, Fleming 1999), their life span may be as short as one to two years.

Food and Feeding

The bay goby is an opportunistic predator and major prey items include polychaetes, harpacticoid copepods, gammarid amphipods, and bivalves (Grossman et al. 1980). Although larger fish (>50 mm SL) and smaller fish (<50 mm SL) consume similar prey items, larger fish include more mollusks, polychaetes, and other larger prey items in their diet.

Predators of the bay goby include the California halibut (Drawbridge 1990) and the Pacific staghorn sculpin (Boothe 1967). It is assumed that other demersal piscivorous fish prey upon bay gobies.

Distribution

In San Francisco Bay, the bay goby is common from South to San Pablo bays, and is occasionally collected in Carquinez Strait and lower Suisun Bay. Densities of young-of-the-year (YOY) bay gobies are usually highest in South or San Pablo bays while densities of older fish are usually highest in Central Bay (CDFG 1987, Fleming 1999). From 1980 to 1995, the bay goby was the most common goby and the second most common fish collected by an otter trawl survey of San Francisco Bay, comprising 14.3% of all fishes collected (Orsi 1999). Although mean densities of YOY fish were higher at shoal stations than channel stations all months, older fish appear to move from the shoals to the channels in the late summer and fall (CDFG 1987 and unpub. data).

Surprisingly, the bay goby was not common in a beach seine survey conducted by CDFG in San Francisco Bay from 1980-1987; it was the fourth most common goby and comprised only 0.06% of all fishes collected by this net (Orsi 1999). These data indicate that the bay goby may not be common in the very shallow subtidal and intertidal areas of San Francisco Bay, although Grossman (1979a) concluded it to be one of the...
numerically dominant fish species of Morro Bay lower intertidal mudflats. The bay goby inhabits burrows of the blue mud shrimp (U p o g e b i a p u g g e t t e n i s) and the innkeeper worm (U r e c h i s c a u p o) and siphon holes of the geoduck clam (Panopea generosa) in Morro Bay (Grossman 1979a). As for several other species of gobies common to San Francisco Bay, including arrow goby and longjaw mudsucker, the bay goby probably utilizes burrows as a refuge from predators and to avoid desiccation at low tides.

Few bay gobies have been collected in San Francisco Bay tidal marshes. One bay goby was reported from Gallinas Marsh and one from Corte Madera Marsh (CH2M Hill 1982). Both fish were collected by gill nets, which were used to sample the larger channels. In contrast, the bay goby was the most common species collected in otter trawl samples from Corte Madera Creek channel, adjacent to Corte Madera Marsh. No bay gobies have been collected by other San Francisco Bay tidal marsh studies (Wild 1969; Woods 1981; ANATEC Laboratories 1981; CH2M Hill 1996; CDFG. unpub. data) or by a study of fishes of Elkhorn Slough tidal marshes (Barry 1983).

Bay goby YOY are most abundant in otter trawl samples from February through June, which is a one or two months after peak abundance period for smaller juveniles from the ichthyoplankton net (CDFG 1987 and unpub. data). In several years, multiple cohorts of YOY fish have been collected; this was especially noticeable in four of the six years of the 1987-92 drought (CDFG, unpub. data). Peak abundance of older fish is usually from May through September, which corresponds with the peak period of larval abundance in San Francisco Bay.

The bay goby has been collected primarily from polyhaline salinities in San Francisco Bay, with YOY fish collected at lower salinities than older fish (Table 2.6). YOY were also collected at slightly lower temperatures than older fish (Table 2.6). These differences in salinity and temperature by age class are reflected by the distribution of YOY somewhat further upstream than older fish and by the peak abundance of YOY in the winter and spring and older fish in summer and fall.

**Population Status and Influencing Factors**

Although trawls are usually considered ineffective for gobies, the bay goby is a very common fish in San Francisco Bay otter trawl surveys. As such, the abundance indices derived from trawl data may be good indicators of population trends. California Department of Fish and Game otter trawl data from 1980-1998 is the longest data set available for the Bay. The indices from 1988 to 1997 were generally higher than the pre-1988 indices (Figure 2.9). The relatively stable salinities year-round during the 1987-92 drought may have resulted in increased nursery habitat for this species (Hieb and Baxter 1993). The multiple cohorts of YOY collected these years, which indicate successful recruitment over a period of several months, in part support this hypothesis. Additionally, high winter outflow events may carry larvæ or pelagic juveniles from the Bay.

Abundance of predators, as California halibut and Pacific staghorn sculpin, could influence the bay goby population. Additionally, factors controlling the abundance of the commensal burrowing invertebrate hosts may effect the bay goby population. This would include the abundance and distribution of intertidal and subtidal mudflat invertebrate predators, such as the bat ray and leopard shark.

**Trophic Level**

Secondary consumer.

**Proximal Species**

**Predators:** California halibut, Pacific staghorn sculpin.

**Prey:** Polychaetes, gammarid amphipods, harpacticoid copepods, bivalves.

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<th>Median</th>
<th>95th percentile</th>
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**Figure 2.9** Annual Abundance Indices of All Sizes of Bay Goby, Otter Trawl (CDFG unpublished data)
Commensal Hosts: Blue mud shrimp, inn-keeper worm, geoduck clam. Bat ray and leopard shark impact the abundance of commensal hosts.

Good Habitat

Good habitat for the bay goby is shallow subtidal areas with mud or a mud/sand mixture and possibly intertidal mudflats. The presence of burrowing invertebrates, which may serve as commensal hosts, would be beneficial. There is no evidence that this species utilizes tidal marshes in San Francisco Bay or elsewhere in its range.

References


Longjaw Mudsucker
Gillichthys mirabilis
Kathryn A. Hieb

General Information

The longjaw mudsucker (Family: Gobiidae) is the largest goby native to San Francisco Bay, reaching a size of 200 mm total length (TL). It ranges from Baja California to Tomales Bay (Miller and Lea 1972) and was successfully introduced to the Salton Sea in 1930 (Walker et al. 1961). The longjaw mudsucker is a common resident of mudflats and sloughs in estuaries and coastal streams. It is also common in salt ponds, as it can tolerate a wide range of salinities. As the tide ebbs, the longjaw mudsucker retreats to burrows or buries in the mud rather than migrate to deeper water. Due to their ability to live out of water and in freshwater for several days, mudsuckers or “mud puppies” are a sought after bait-fish; however, in recent years, the San Francisco Bay area bait fishery has targeted the yellowfin goby, a large introduced species that is very common in many shallow water habitats.

Reproduction

Male longjaw mudsuckers construct burrows for breeding, which they aggressively guard until the eggs hatch. A single female lays 4,000 to 9,000 eggs, depending on size (Weisel 1947). In southern California, spawning occurs from January through July, with peak activity apparently from February through April (Weisel 1947). In South Bay salt ponds, the spawning period is also protracted, occurring from November through June, with peak activity in February and March (Lonzarich 1989). Gonadal regression occurs from July to September, when temperatures in the salt ponds reach their maximum (de Vlaming 1972). Females were reported to spawn more than once per season in South Bay salt ponds (de Vlaming 1972) and two and possibly three times per season in the Salton Sea (Walker et al. 1961), with an interval of 40 to 50 days between spawnings (Barlow 1963). Ovarian development and spawning are asynchronous, which is typical of species that spawn more than once per season and have a protracted spawning season (de Vlaming 1972).

The eggs are club shaped, 2.8-3.4 mm long, with an adhesive thread at one pole that attaches to the burrow wall. Hatching occurs in 10 to 12 days at 18°C (Weisel 1947). Larvae have been collected year-round in the Bay, with peak abundance in May and June (CDFG, unpub. data); in South Bay salt ponds, larvae were collected at salinities up to 70‰ (Lonzarich 1989).

Growth and Development

In South Bay salt ponds, longjaw mudsuckers grow to 80-100 mm standard length (SL) by the end of year one and 120-140 mm SL by the end of year two (Lonzarich 1989). Few live more than one year and none more than two years; both sexes mature at age one (Barlow 1963, Lonzarich 1989). In the Salton Sea, longjaw mudsuckers hatched in the early spring reach 60-80 mm SL by fall and 80-120 mm SL by the next spring (Walker et al. 1961).

Food and Feeding

In Elkhorn Slough, California, the longjaw mudsucker preys primarily on gammarid amphipods, especially Orchestia traskiana, Eogammarus confervicolus, Corophium spp., and polychaetes (Barry 1983). Dipterans, harpacticoid copepods, and grapsid crabs (primarily Hemigrapsus oregonensis) are also important food items. In South San Francisco Bay salt ponds, longjaw mudsucker diet varies by salinity—in the lower salinity (20-40‰) ponds, they consume primarily polychaetes and amphipods while in the higher salinity (to 84‰) ponds they consume primarily brine shrimp and waterboatmen (Lonzarich 1989). Copepods are an important prey item in the winter, when brine shrimp are unavailable.

Distribution

In San Francisco Bay, the longjaw mudsucker has been collected in South, Central, San Pablo, and Suisun bays, although it is not common upstream of Carquinez Strait. It is the least common goby collected in trawl surveys of open water habitats and larger channels, but usually the most common goby collected in smaller marsh channels. For example, it was not collected in trawls near Castro Creek, Corte Madera Creek, and Gallinas Creek marshes, but was the most abundant goby and third most abundant species collected in minnow traps set in the marsh channels (CHM Hill 1982). Similarly, in a study of a restored marsh near Hayward, it was not common in trawls of the larger channels, but the only goby and most common species collected in minnow traps set on the mudflats (Woods 1981). It was also the second most common species collected in first and second order channels of tidal marshes in lower Petaluma River (CDFG, unpub. data). This distribution has also been
reported from Elkhorn Slough, where the longjaw mudsucker was not an important component of the otter trawl samples from deeper (>1.5 m) channels, but was the third most abundant species and most common goby in beach seine and channel net samples from shallower (<1.5 m) channels (Barry 1983).

The longjaw mudsucker is also common in salt ponds in San Francisco Bay. It was the most common goby and the second most common fish collected in South Bay salt ponds (Carpelan 1957, Lonzarich 1989). Lonzarich (1989) reported highest catches in the summer and fall.

Although longjaw mudsucker can tolerate a wide range of salinities, they are usually absent from fresh or slightly brackish water (Barlow 1963). They have been collected from salinities as high as 82.5‰ in the upper Gulf of California (Barlow 1963), and as high as 84‰ in South Bay salt ponds (Lonzarich 1989).

Although longjaw mudsuckers have been collected at temperatures as high as 33 °C (Carpelan 1957), in laboratory thermal selection studies, they preferred temperatures from 9-23°C and strongly avoided temperatures greater than 23°C (de Vlaming 1971). In another laboratory study, Courtois (1973) concluded that the longjaw mudsucker was best adapted to temperatures between 20 and 30°C.

In intertidal areas, the longjaw mudsucker often remains in the mud or burrows at low tide and is subject to fluctuating oxygen concentrations. The jaw membranes are richly vascularized and serve as an accessory respiratory apparatus (Weisel 1947). Additionally, the longjaw mudsucker will respire aerially at low (<2.0 mg/L) oxygen concentrations; they gulp air at the water surface and hold the bubbles in their large buccopharyngeal cavity (Todd and Ebeling 1966).

**Population Status and Influencing Factors**

There is no survey which routinely samples the longjaw mudsucker or its preferred habitat in San Francisco Bay, so the current status of the population cannot be assessed. With the introduction and establishment of the yellowfin goby in the 1960s, the longjaw mudsucker is no longer as sought after for bait. However, the introduction of the yellowfin goby may have had a negative impact on the longjaw mudsucker, as there is substantial overlap in the habitats of the two species.

**Trophic Levels**

The longjaw mudsucker is a secondary consumer.

**Proximal Species**

**Predators:** Bait fishers and possibly great blue herons, egrets, and larger shorebirds.

**Prey:** Gammarid amphipods, polychaetes, dipterans, copepods, *Hemigrapsus oregonensis*, waterboatmen, brine shrimp.

**Good Habitat**

The intertidal area of tidal marsh channels is the typical habitat of the longjaw mudsucker. Because this species can tolerate a wide range of environmental conditions, “good habitat” is probably defined by the complexity of these sloughs. More complex channels, with undercut banks and pools of water at low tide, would offer more protection from predators than sloughs with little incision and ponded water. These more complex channels are typical of mature marshes vs. recently “restored” marshes.

**References**


California Halibut
Paralichthys californicus
Michael K. Saiki

General Information
The California halibut (Family: Bothidae) is a large marine flatfish that is sought after in the market place because of its large size and excellent taste (Frey 1971). Commercial fishing for California halibut was historically centered in the Baja California-Los Angeles area, but has recently shifted northward to the Santa Barbara region (Barsky 1990). It is harvested by gill net, trammel net, and trawl nets (Schultze 1986). California commercial fishermen landed an average of 534 tons per year from 1983 to 1987, and received $0.64-$1.59/kg in 1987 (CDFG 1988). California halibut is also highly prized by recreational anglers and is caught primarily from piers and boats using hook, line, and live bait. Over 916,000 California halibut were caught by anglers in 1985 (USDC 1986).

Reproduction
Emmett et al. (1991) described the California halibut as being gonochoristic (its gender is determined by developmental rather than hereditary mechanisms) and iteroparous (it has the capacity to survive and spawn beyond one or multiple spawning seasons). It is a broadcast spawner whose eggs are fertilized externally (Emmett et al. 1991). The eggs of California halibut are pelagic (Allen 1988). In a laboratory tank with water depth of 2-3 m, California halibut spawned while swimming near the water surface (Allen 1990). Adults typically move into shallow (6-20 m deep) coastal waters in early spring and usually spawn over sandy substrates (Ginsburg 1952, Frey 1971, Feder et al. 1974, Hake 1975). Spawning occurs from February through August, peaking in May with a great number of mature fish (Frey 1971, Feder et al. 1974, Wang 1986). Spawning most often occurs when water temperatures are 15.0-16.5°C, and day lengths are greater than or equal to 10.5 hours (Caddell et al. 1990). However, abundant eggs and larvae have also been reported from nearshore coastal waters during winter-spring when surface temperatures are 13-15°C, and even during summer when surface waters occasionally reach 22°C (Lavenberg et al. 1986, Petersen et al. 1986).

During the spawning season, small (55.9-61.0 cm long) California halibut produce approximately 300,000 eggs every 7 days, whereas large (>114.3 cm long) halibut produce about 1 million eggs per day (Emmett et al. 1991).

Growth and Development
California halibut eggs are spherical in shape and 0.74-0.84 mm in diameter (Ahlstrom et al. 1984). Eggs hatch approximately two days after fertilization at 16°C (Emmett et al. 1991). Newly hatched larvae of California halibut measure about 2.0 mm total length (TL) (Ahlstrom and Moser 1975, Ahlstrom et al. 1984). The larval yolk sac is depleted about six days after hatching (Gadomski and Petersen 1988).

Metamorphosis occurs at a length of 7.5-9.4 mm (Ahlstrom et al. 1984) when the pelagic, bilaterally symmetrical larvae become benthic, asymmetrical juveniles. Along with other physical changes, the most visible part of this process is a change in pigmentation patterns and the migration of one eye across the top of the head to its final resting place close to the other eye (Moyle and Cech 1988).

Temperature has a major effect on survival of eggs and larvae of the California halibut. Successful hatching occurred at 12°, 16°, and 20°C, but death occurred prior to embryo formation at 8° and 24°C (Gadomski and Caddell 1991). At 17 days posthatch, all larvae died at 12°C, whereas survival varied from 23% to 46% at 16°, 20°, and 24°C. The survival of older larval stages of California halibut progressively increased as incubation temperatures rose from 16°C to 28°C. Temperature also affected the settlement rate of juveniles that had just completed metamorphosis.

Although juveniles are reported to vary in length from 8 mm to 430 mm (Emmett et al. 1991), males can mature at 200-300 mm standard length (SL) when 2-3 years old whereas females can mature at 380-430 mm SL when 4-5 years old (Fitch 1965, Fitch and Lavenberg 1986).
California halibut may reach a maximum length of 1,520 mm and a maximum weight of 33 kg (Eschmeyer et al. 1983), with certain individuals living for as long as 30 years (Frey 1971).

**Food and Feeding**

California halibut feed initially on small invertebrates, but later switch almost exclusively to feeding on fish (H aker 1975). Although the diet of larvae has not been examined, they probably feed on tiny planktonic organisms (Allen 1990). Small juveniles in three southern California embayments fed mostly on harpacticoid copepods and gammaridean amphipods, with some polychaetes, mysids, small fish, and crab megalopae also being taken (H aker 1975, Allen 1988). In Anaheim Bay, California, large juveniles and small adults ate bay shrimp, topsmelt, California killifish, and gobies, whereas subadults and adults more than 23.0 cm SL consumed mostly northern anchovy, croaker, and other larger fishes (H aker 1975). Other forage taxa in the diets included ostracods and acteonid snails. In Tomales Bay, adult California halibut (65.4-83.3 cm SL) fed on Pacific saury, Pacific herring, sanddabs, white sea perch, and California market squid (Bane and Bane 1971). The California halibut is an ambush predator (H aker 1975). During foraging it lies partially buried on the sandy bottom and waits until its prey is close enough to seize.

**Distribution**

The geographic distribution of California halibut extends from the Quillayute River, Washington, southward to Magdalena Bay, Baja California (Ginsburg 1952, M iller and Lea 1972, Eschmeyer et al. 1983). However, it is common only in bays and estuaries south of Tomales Bay, California, and reaches peak abundance in estuaries south of Point Conception (Emmett et al. 1991). Recently, large numbers of mostly female California halibut were caught by recreational anglers in Humboldt Bay, with some caught as far north as Crescent City and southern Oregon (R. Baxter, pers. comm.). A survey of carcasses suggested that the females had not developed mature eggs.

Larvae of California halibut occur primarily in the upper 30 m of coastal waters, where they apparently settle or migrate from the 0-10 m stratum to the 10-20 m stratum at night (M oser and W atson 1990). Conversely, larvae over shallow water (13 m bottom depth) tend to move downward during the day (Barnett et al. 1984). Juveniles settle in shallow water on the open coast, but are more abundant in bays (Allen 1988, M oser and W atson 1990). Juveniles remain in bays for about two years until they emigrate to the coast where they settle at water depths less than 100 m, with greatest abundance at depths less than 30 m (M iller and Lea 1972, Allen 1982). Larger juveniles (greater than 20 mm in length) may move from open coastal areas to settle in bays (Kramer 1990).

Adults move inshore during spring and summer, and offshore during winter (Ginsburg 1952, H aker 1975). Although the inshore movements are associated with spawning, they may also be influenced by seasonal patterns in forage fish abundance. For example, during spring and summer, California grunion (Leuresthes tenuis) are abundant near the surf zone (Feder et al. 1974), whereas northern anchovy (Engraulis mordax) are abundant in bays and estuaries (Tuppen 1990).

California halibut are occasionally found in Central and South San Francisco Bay (Alpin 1967, Pearson 1989) and San Pablo Bay (Ganssle 1966). Recently, eggs of a description similar to those of California halibut were collected in San Francisco Bay; however, their identity was not verified (Wang 1986). Both larval and juvenile California halibut have been captured in San Francisco and San Pablo bays (Wang 1986).

**Population Status and Influencing Factors**

Catch records indicate that the abundance of California halibut within its historic range was high in the late 1960s, declined in the 1970s, and increased in the 1980s. The intense El Niño in 1982-83 coincided with higher abundance and landings of halibut (Jow 1990). Overall, however, California halibut populations seem to be undergoing a long-term decline. This decline may be related to large-scale changes in the marine environment, overfishing, alterations and destruction of estuarine habitat, or a shift in location of population centers (Plummer et al. 1983). Pollution has been shown to reduce hatching success, reduce size of larvae at hatching, produce morphological and anatomical abnormalities, and reduce feeding and growth rates (MBC Applied Environmental Sciences 1987). By comparison, thermal effluents from California coastal power plants do not seemingly inhibit growth and may be advantageous to California halibut (Innis 1990).

Early records indicate that California halibut were uncommon in San Francisco Bay. Alpin (1967) sampled the Central Bay with bottom trawls during 1963-1966 and reported catching only three California halibut (two in the spring and one in June). Ganssle (1966) reported catching only two adult California halibut (May 1963, 1964) while fishing bottom trawls in San Pablo Bay. Recently, consistent high salinities probably have contributed to increased abundance of California halibut in the bay. Moreover, recent data suggest that successful year classes in 1983, 1987, and 1990 have contributed to increased abundance in the bay (CDWR 1991). These were years with warm water ocean events, and it is hypothesized that California halibut abundance in the San Francisco Bay increased because of increased local...
spawning, higher survival of larvae, or migration of juveniles from more southern coastal areas with warmer ocean waters (Hieb and Baxter 1994).

Abundance indices (determined from trawl samples) for California halibut in San Francisco Bay increased from 1989 to 1992 (Hieb and Baxter 1994). The 1992 index was the highest since the study began in 1980. Also, most halibut collected in San Francisco Bay are age two and older, whereas other flatfishes are caught primarily as young-of-the-year. Nevertheless, California halibut abundance indices are still very low relative to other common species of flatfish in the Bay (Hieb and Baxter 1994).

In an attempt to increase California halibut numbers, natural production has been augmented by hatchery production (Crooke and Taucher 1988). Although this effort could increase future recruitment, negative effects of the hatchery program include a possible reduction in genetic variability within natural populations and the high cost producing fish (Hobbs et al. 1990).

**Trophic Levels**

Larvae, juveniles, and adults are carnivorous (secondary and higher order consumers).

**Proximal Species**

**Predators:** Thornback (important predator on settling juveniles), California sea lions (predator on large juveniles and adults), northern sea lions, Pacific angel shark, Pacific electric, bottlenose dolphin.

**Prey:**


**Parasites:** Trematodes, cestodes, and nematodes (endoparasites); copepods and isopods (ectoparasite).

**Competitors:** Speckled sanddab (potentially important).

**Good Habitat**

Good spawning habitat for California halibut is limited to inshore waters or bays and estuaries in moderately shallow water where temperatures approximate 13-15°C, although successful spawning may also occur at temperatures approaching 22°C (Gadomski and Caddell 1991). Favorable characteristics for bays and estuaries that serve as nursery areas include productive habitats with abundant food supplies and shallow areas that allow juveniles to avoid predators, including adult halibut (Plummer et al. 1983). Juveniles and adults prefer sandy bottoms and water temperatures between 10-25°C, with a preference for 20.8°C (Ehrlich et al. 1979). Juveniles are relatively tolerant of reduced dissolved oxygen and increased water temperatures (Waggoner and Feldmeth 1971). Higher water temperatures induces faster growth rates and decreases the time to settlement for most young-of-the-year halibut (Gadomski et al. 1990). Eggs, larvae, and adults are found in euhaline waters, but juveniles often occur in oligohaline to euhaline conditions (Haaker 1975).

**References**


Starry Flounder

**Platichthys stellatus**

**Kurt F. Kline**

### General Information

The starry flounder is in the family Pleuronectidae, or right-eyed flounders. Pleuronectids are generally found in temperate marine environments, with only a few species found in the tropics or sub-tropics. There are 22 species found along the coast of California. The starry flounder is one of the few pleuronectids commonly found in brackish and freshwater (Orcutt 1950, Haertel and Osterberg 1967). While placed in the Pleuronectidae, the starry flounder is commonly right or left-eyed. However, it is quite distinguishable from other flatfishes due to the alternating dark gray and orange-yellow bands on the dorsal, anal, and caudal fins.

Many of the pleuronectids support commercial and sport fisheries. The starry flounder is a minor sport species in San Francisco Bay and most fish are taken from boats when fishing for California halibut, sturgeon, or striped bass. It common in the commercial fishery, but as a by-catch to targeted species such as petrale sole and California halibut. In recent years, nearshore gear restrictions have resulted in a decrease in starry flounder landings, as this species is most common within a few miles of shore (Haugen 1992).

### Reproduction

Spawning occurs in winter in shallow coastal areas near the mouths of rivers and sloughs (Orcutt 1950, Wang 1986, Baxter 1999). Some researchers have suggested that spawning may occur within San Francisco Bay.
(Radtke 1966, M oyle 1976); however, neither ripe fe-
male starry flounder nor mature flounder eggs or
pre-flexion larvae were collected from San Francisco Bay
in the early 1980s (B. Spies, pers. comm., W ang 1986).

Growth and Development
Eggs and larvae are pelagic and found mostly in the upper
water column (O rcutt 1950, W ang 1986). Starry floun-
der larvae are approximately 2 mm long at hatching and
settle to the bottom about two months after hatching,
at approximately 7 mm standard length (SL) (Policansky
and Sieswerda 1979, Policansky 1982). Larvae depend
upon favorable ocean currents to keep them near their
estuarine nursery areas before settlement. Transforming
larvae and juveniles migrate from the coast to brackish
or freshwater nursery areas, where they rear for 1 or more
years (H aertel and O sterberg 1967, W ang 1986, H ieb
and B axter 1993). As they grow, juvenile starry floun-
der move to higher salinity, but appear to remain in es-
tuaries through at least their second year of life (H aertel

Most males mature by the end of their second year
of life (220-276 mm SL), while females mature at 3 or
4 (239-405 mm SL) (O rcutt 1950). During the late fall
and winter, mature starry flounder probably migrate to
shallow coastal waters to spawn (O rcutt 1950). After
spawning, some adult starry flounder return to the Bay
for feeding, and are most common in the Bay from late
spring through early fall (G anssele 1966). They reach a
maximum length of 915 mm (M iller and L ea 1972).

Food and Feeding
In M onterey B ay and Elkhorn S lough, the smallest starry
flounder (10-99 mm SL) fed primarily on copepods and
amphipods. Larger juveniles (100-199 mm SL) fed on
larger amphipods, polychaetes, and bivalves (especially
siphon tips). Fish >199 SL mm fed on whole crabs and
bivalves, sand dollars, brittle stars, and occasionally fish
(O rcutt 1950). In San Francisco Bay, a large portion of
the diet of starry flounders >199 mm was bivalves (pri-
marily M ya, I schadium, T apes, S olen, M ytillus, and
Gemma), polychaetes, and crustaceans (especially
U pogebia, C ancer magister, C . gradilis, and H emigraphus
oregonensis) (C DFG , unpubl. data).

Distribution
Starry flounder range from Santa Barbara, California
northward to arctic Alaska, then southwesterly to the Sea
of Japan (M iller and L ea 1972). Adult starry flounder
inhabit shallow coastal marine water, whereas juveniles
rear in bays and estuaries (O rcutt 1950, M oyle 1976,
starry flounder are found almost exclusively in estuaries.
In San Francisco Bay, there is a shift in distribu-
tion with growth. Age-0 fish are found more commonly
in fresh to brackish water (Suisun Bay, Suisun M arsh, and
the delta), while age-1 and older juveniles are more
commonly associated with brackish to marine waters
(Suisun and San Pablo bays). Throughout their time in
the San Francisco Bay, juvenile starry flounder are com-
monly found in shallow water, including shoals, inter-
tidal areas, and tidal marshes (Woods 1981, M oyle et al

Population Status and Influencing Factors
There is evidence of a long-term decline in the San Fran-
cisco Bay starry flounder population from the Commer-
cial Passenger Fishing Vessel log book data. Both catch/
hour (C PUE) and total catch of starry flounder declined
in the mid-1970s from a peak in the late 1960s and early
1970s (C DFG 1992). This decline in CPUE and catch con-
tinued at least through the early 1990s. Additionally, juve-
nile starry flounder abundance indices from San Francisco
Bay steadily declined from the early to the late 1980s (F i-
ger 2.10). Abundance remained very low through 1994 and
increased somewhat from 1995-99. An outflow related mecha-
nisms have been proposed to control recruitment of age-0
starry flounder to the Bay (C DFG 1992, H ieb and B axter
1993). The increase in the abundance of age-0 fish from
1995 to 1999 supports this hypothesis.

Hydrologic factors and other environmental con-
ditions in San Pablo and Suisun bays are important in
determining the distribution of juvenile starry flounder.
The San Francisco Estuary is close to the southern limit
of the distribution for starry flounder and long-term

Figure 2.10 Annual Abundance Indices of Starry
Flounder: A. Age-0, May-October; B. Age-1,
February-October (C DFG Otter Trawl data)
changes in the oceanic environment (particularly temperature) may also affect recruitment. Ocean temperatures have been above average for the region for much of the 1980s and 1990s and it is possible that adult populations moved northward into cooler waters. Temperature can also influence spawning and early development, as increased temperatures may result in decreased hatching success and larval survival.

**Trophic Levels**

Primary to secondary carnivore. Feeds primarily on large benthic invertebrates and rarely on fish.

**Proximal Species**

*Prey:* Benthic invertebrates including bivalves, polychaetes, and crustaceans.

**Good Habitat**

Suitable habitat includes shallow to deep subtidal mud and sand flats. Juvenile rearing occurs in the shallow areas of Suisun and San Pablo bays. Open deeper waters with higher salinity are generally more acceptable for adults.

**Acknowledgments**

Some of the materials in this report are summarized from the Pluronectiformes chapter of IEP Technical Report 63 which is referenced below (Baxter 1999).

**References**


**Personal Communications**

Bob Spies, Applied Marine Sciences, Livermore, California.
Invertebrates

Franciscan Brine Shrimp
Artemia franciscana Kellogg
Brita C. Larsson

General Information

The Franciscan brine shrimp, Artemia franciscana (formerly salina) (Bowen et al. 1985, Bowen and Sterling 1978, Barigozzi 1974), is a small crustacean found in highly saline ponds, lakes or sloughs that belong to the order Anostraca (Eng et al. 1990, Pennak 1989). They are characterized by stalked compound eyes, an elongate body, and no carapace. They have 11 pairs of swimming legs and the second antennae are uniramous, greatly enlarged and used as a clasping organ in males. The average length is 10 mm (Pennak 1989). Brine shrimp commonly swim with their ventral side upward. A. franciscana lives in hypersaline water (70 to 200 ppt) (Maiss and Harding-Smith 1992).

In the Bay area, the optimum temperature for A. franciscana is 21-31°C. In the winter, when temperatures fall below this range, brine shrimp populations decline and their growth becomes stunted (Maiss and Harding-Smith 1992). Other environmental factors such as wind, salinity, and the quantity and quality of phytoplankton may also affect Bay area populations of A. franciscana and their effects on this species are currently being investigated (Maiss and Harding-Smith 1992).

Reproduction, Growth, and Development

Artemia franciscana has two types of reproduction, ovoviviparous and oviparous. In ovoviviparous reproduction, the fertilized eggs in a female can develop into free-swimming nauplii, which are set free by the mother. In oviparous reproduction, however, the eggs, when reaching the gastrula stage, become surrounded by a thick shell and are deposited as cysts, which are in diapause (Sorgeloos 1980). In the Bay area, cysts production is generally highest during the fall and winter, when conditions for Artemia development are less favorable. The cysts may persist for decades in a suspended state. Under natural conditions, the lifespan of Artemia is from 50 to 70 days. In the lab, females produced an average of 10 broods, but the average under natural conditions may be closer to 3-4 broods, although this has not been confirmed. Each brood contains from 30 to 100 offspring which mature in 10-25 days (Maiss and Harding-Smith 1992). The larva grows and differentiates through approximately 15 molts (Sorgeloos 1980).

Food and Feeding

Artemia franciscana feed on phytoplankton and blue-green algae that occur in Bay area salt ponds (Maiss and Harding-Smith 1992).

Distribution

Artemia franciscana occurs in highly saline waters throughout western North America, Mexico, and in the Caribbean (Bowen et al. 1985). In California, A. franciscana occurs from sea level to 1,495m and in many parts of the state, but its distribution is spotty because of this species salinity requirements (Eng et al. 1990). Historically in the Bay area they were found in salt pannes and sloughs were hypersaline conditions occurred. Currently they occur in salt ponds in the northern and southern portion of San Francisco Bay that are...
used for the commercial production of salt. Salt ponds cover approximately 111 km and in the North bay 36 Km off the Bay’s shoreline (Lonzarich 1989). The distribution of Artemia in these salt ponds is limited by the salinity of the ponds. The optimum salinity range for Artemia is 70 ppt to 175 ppt (Carpelan 1957). They do not occur where the salinity is above 200 ppt.

Population Status and Influencing Factors

Commercial salt production in San Francisco Bay is currently an active industry, so habitat for A. franciscana is not limited and populations are large due to ample amounts of habitat. Donaldson et al. (1992) sampled a 496 acre salt pond in the San Francisco Bay National Wildlife Refuge and estimated the highest winter adult population at 40 billion and the lowest winter population at 4.5 billion. Brine shrimp populations are lowest in the winter and peak in the summer months when their optimal temperatures occur so these numbers are conservative for a maximum population value for the pond. Current populations of the brine shrimp probably far exceed historic populations because the salt ponds in which they occur are manmade. Salt ponds occurred naturally and there is even some evidence that the Ohlone Indians manipulated a portion of the Bay shoreline for salt production but never was there as much salt pond habitat for brine shrimp as currently occurs in the Bay area.

Trophic Level

Artemia franciscana is a primary consumer.

Proximal Species

Anderson (1970) lists sightings of 55 bird species using salt ponds in San Francisco Bay. Mallards, California gulls, whimbrels, Wilson’s phalarope, eared grebes and American avocets are several species which feed on A. franciscana. Western and least sand pipers, willets, greater yellow legs and Bonaparte’s gulls are commonly seen roosting and feeding in the salt pond environment and most likely feed on Artemia in these ponds (Maiss and Harding-Smith 1992).

Good Habitat

Brine shrimp occur in salt ponds adjacent to San Francisco Bay that have salinities ranging from 70 to 200 ppt but are most common when the range is between 90 and 150 ppt (Maiss and Harding-Smith 1992). Harvey et al. (1988) reported that up to 46% of the 23,465 acres of active salt ponds in South Bay are within the 70-200 ppt salinity range in the summer and contain brine shrimp.

References

California Vernal Pool Tadpole Shrimp
Lepidurus packardi Simon
Brita C. Larsson

General Information
The California vernal pool tadpole shrimp is a small crustacean found in ephemeral freshwater pools that belong to the order Notostraca. They are characterized by sessile compound eyes, a shield-like carapace covering the head and much of the trunk, and a telson that is a flat and paddle-shaped protuberance. They can reach a length of 50 mm and have approximately 35 pairs of legs and two long cercopods (Pennak 1989). Tadpole shrimp are primarily benthic organisms that swim with their legs down. They can also climb or scramble over objects and plow through bottom sediments (Federal Register 1994). Information about the biology of this species is limited and incomplete (Ahl 1991).

Reproduction, Growth, and Development
Much of what is known about the reproduction, growth, and development of L. packardi comes from studies by Ahl (1991) and Longhurst (1955). Their life history is dependent on ephemeral freshwater pools. In California, vernal pools are generally hydrated during the rainy season, which extends from winter to early spring. Populations of tadpole shrimp are reestablished from diapaused eggs when winter rains rehydrate vernal pools. Once a pool rehydrates, the eggs hatch over a three week period, some hatching within the first four days. It takes another three to four weeks for the tadpole shrimp to become sexually reproductive. Populations consist of both males and females, though late in the season, pools are often dominated by males. After copulation, fertilized eggs descend into the foot capsule of the female (Desportes and Andrieux 1944). The eggs are sticky and when they are deposited they adhere to plant matter and sediment particles (Federal Register 1994). A female can have up to six clutches of eggs, totaling about 861 eggs during her lifetime (Ahl 1991). Depending on the depth and persistence of water in a pool, some eggs hatch immediately. The remainder enter diapause and lie dormant in the sediment during the dry portion of the year (Ahl 1991).

Food and Feeding
Tadpole shrimp feed on organic detritus and living organisms such as fairy shrimp and other invertebrates (Pennak 1989, Fryer 1987).

Distribution
L. packardi is endemic to vernal pools in the Central Valley, coast ranges and a limited number of sites in the Transverse Range and Santa Rosa Plateau (Federal Register 1994). The distribution of this species is not well known for the Bay area. Recently, L. packardi was collected at the Warm Springs Seasonal Wetland which is a part of the Don Edwards San Francisco Bay National Wildlife Refuge (Caires et al 1993). Other populations have been found north of the eastern half of Potrero Hills in the North Bay (S. Forman, Pers. obs.). Seasonal wetlands occur sporadically in both the North and South Bay and may provide additional habitat for this species. Surveys in seasonal wetlands surrounding San Francisco Bay may contribute and increase information on the distribution of this species.

Population Status and Influencing Factors
Current status of the population of tadpole shrimp in the Bay area is not known. Loss of seasonal wetland habitat in the Bay area may be significantly affecting the population of this species especially since distribution information for the Bay area is so limited.

Trophic Level
Lepidurus packardi is most likely a secondary consumer.

Proximal Species
Waterfowl, western spadefoot toad, and tadpoles.

Good Habitat
Lepidurus packardi inhabits vernal pools. They have been found in pools ranging in size from 5 square meters to 36 hectares. The water in the pools can be clear to turbid. The pools often have low conductivity, TDS, and alkalinity (Federal Register 1994, Eng et al. 1990). The pools dry up in the late spring and are dry in the summer and fall then fill with rain water in the winter and early spring. Vernal pool formations occur in grass bot-
tomed swales of grasslands in old alluvial soils, underlain by hardpan or in mud bottomed pools (Federal Register 1994). Pools with cobbly hardpan bottoms also serve as habitat (Gallagher 1996). Gallagher (1996) found that the depth, volume, and duration of inundation of a pool was important for the presence of L. packardi in vernal pools when compared to the needs of other branchiopods. He found L. packardi did not reappear in ponds that dried and rehydrated during the study period, while other Branchiopod species did. L. packardi needs deeper and longer-lasting pools if they are to persist over a rainy season in which both wet and dry periods occur. Temperature variation in pools where L. packardi have been found to vary from 3 to 23°C (Gallagher 1996). Salinity, conductivity, dissolved solids, and pH of the water in vernal pools are also important in determining the distribution of tadpole shrimp (Federal Register 1994).

References


Personal Communication

Steve Forman, LSA Associates, Incorporated

Reticulate Water Boatman

Trichocorixa reticulata Guerin
Wesley A. Maffei

Description and Systematic Position

Trichocorixa reticulata is a small hemipteran, approximately 3-5mm in length, that belongs to the family Corixidae. This insect, also known as the salt marsh water boatman, can be recognized by the fine network of lines on its hemelytra (outer wing covers), the 10-11 dark transverse bands on the pronotum, and the pala of front legs not exceeding two-thirds the width of an eye along the ventral margin (Figure 3.1).

Distribution

Sailer (1948) states that this insect is found along the Pacific Coast from northern San Francisco Bay south to Peru. Populations from Kansas, New Mexico, Texas, Florida, and the Hawaiian Islands have also been recorded. One isolated record was reported in China but this has been unconfirmed. Within the San Francisco Bay environs this water boatman can be found in mid to upper marsh tidal pools and man-made salt ponds. Figure 3.2 shows the locations around the Bay Area where T. reticulata have been collected, and Table 3.1 shows the collection dates.

Suitable Habitat

T. reticulata prefers saline environments. Cox (1969) found this insect in southern California coastal salt ponds with salinities ranging from brackish up to 160 ‰ and Jang (1977) states that this water boatman can occur in ponds with salinities up to 170 ‰. Carpelan (1957) found the Alviso population in Cargill salt ponds that ranged from 23 ‰ up to 153 ‰. In all instances it was found that the greatest numbers of individuals and the most reproduction occurred in saline environments with a salinity range of 35-80 ‰.

Figure 3.1 Reticulate Water Boatman – Trichocorixa reticulata
Invertebrates

Biology

Sailer (1942) believes that all species of Trichocorixa overwinter as adults. Scudder (1976) states that tones (unpub.) has found that in Saskatchewan, Trichocorixa verticalis interiores overwinters in the egg stage. Eggs are laid singly on submerged vegetation or objects on the bottom substrate. Developmental time for eggs and immatures can very considerably with temperature.

Adult water boatman are both herbivorous and predatory feeding on algal cells and various microorganisms. Although Corixids are aquatic in all life stages, the adults are capable of leaving the water and dispersing by flight. Maffei (unpub.) has noted that south San Francisco Bay populations are attracted to dark colored objects, with adults landing in large numbers on the hoods of green or burgundy colored vehicles while adjacent white vehicles had few if any specimens.

Reproduction

Cox (1969) and Carpelan (1957) have noted that peak reproduction occurs in saline environments with salinities ranging between 35% and almost 80%. Egg laying is continuous during spring, summer and fall with the greatest number of nymphs occurring during April and May. Cox (1969) has also found that crowding of adults led to increased egg production in females.

Balling and Resh (1984) have reported, that the number of generations per year for the Petaluma Marsh population was at least in part dependent on the longevity of the tidal ponds. They found that ponds which dried during late summer contained overwintering, non-reproducing adults while water filled ponds would produce another generation. Reproduction does occur year-round but Cox (1969) states that salinity and adult densities influence the number of eggs laid and the maturation rates of the immature stages. Balling and Resh (1984) noted that the time between generations of the Petaluma Marsh population was also affected by variable egg development times, variable instar development rates, and inter-pond differences in recruitment of adults. In general, it has been determined that environmental conditions can cause water boatmen to either accelerate or delay their development and production of subsequent generations.

Significance to Other Wetlands Taxa

This insect is considered an important prey item for shore birds. Howard (1983) studied the esophageal contents of 35 Ruddy Ducks, Oxyura jamaicensis, at the Alviso salt ponds and found that this water boatman comprised 12.6% of the total food volume. Howard also examined the gizzard contents of 53 Ruddy Ducks and found that 25.5% of the total food volume was water boatmen. Anderson (1970) analyzed the stomach contents of 10 Ruddy Ducks and found that water boatman, snails and Widgeon grass seeds were the primary components of their diet. He also found Least Sandpipers, Wilson’s Phalarope and Northern Phalarope’s utilized this insect as part of their diets.

Conservation Needs and Limiting Factors

Salinity and the length of time tidal marsh ponds contain water seem to be the primary driving forces affect-
ing both developmental rates and reproduction.

References


Additional Readings


Tiger Beetles

*Cicindela senilis senilis*, *C. oregona*, and *C. haemorrhagica*

Wesley A. Maffei

Description and Systematic Position

*Cicindela senilis senilis*, *C. oregona*, and *C. haemorrhagica* are moderate sized beetles, approximately 10-15mm in length, that belong to the family Cicindelidae (Figure 3.3). These beetles, also known as tiger beetles, can be easily identified by their large, bulging eyes and long, sickle-shaped mandibles that bear small teeth. Adults of *C. senilis* and *C. oregona* are usually shining metallic blue to green on the ventral surface with the dorsum dull coppery brown and bearing small yellowish-white irregular markings. *Cicindela haemorrhagica* is similar in appearance to both *C. senilis* and *C. oregona* except that the ventral surface of the abdomen is usually bright red. The larvae are S-shaped, yellowish-white, have the head and the first thoracic segment flattened, an enlarged hump on the fifth abdominal segment with hooks, and large mandibles that are similar to the adults.

Distribution

Historically the San Francisco Estuary, including the beaches just outside of the Golden Gate Bridge, was home to four species of tiger beetles. These were: *Cicindela haemorrhagica*, *C. hirticollis*, *C. oregona oregona* and *C. senilis senilis*. Only two species, *C. haemorrhagica* and *C. senilis senilis* are present today with *C. haemorrhagica* in decline within or near the tidal areas of the San Francisco Bay Estuary. *Cicindela oregona oregona* may still be present within the estuary but the last known population was destroyed in 1996.

The dominant tiger beetle, *C. senilis senilis*, is currently found throughout the south and central portions of the estuary with one population having been identified from Grizzly Island in 1991. Museum records in-
Invertebrates indicate that this beetle was also found in San Rafael, Martinez and Port Costa but these sites have not been sampled in over 40 years. *C. haemorrhagica*, has become increasingly scarce as its habitat continues to be altered for human needs. This beetle is currently found at Trojan Marsh (San Leandro), Hayward Landing (Hayward), Salt Ponds west of Newark and the Richmond Field Station (Richmond). Historically this beetle had a broader distribution with sites as far north as Martinez and south throughout most of the south San Francisco Bay. The populations at Alameda, Bayfarm Island and Oakland no longer exist and other sites identified from museum records have apparently not been sampled in at least three or more decades (Maffei, unpub.). *C. oregona* is probably no longer present within the tidal and diked marshes of the San Francisco Bay. The last known population was at Bayfarm Island and was extirpated in 1996 when the site was graded in preparation for development. Figure 3.4 shows the locations around the Bay Area where *C. senilis senilis*, *C. oregona*, and *C. haemorrhagica* have been collected, and Table 3.2 shows the collection dates.

**Suitable Habitat**

San Francisco Bay tiger beetles are commonly found along open, muddy margins of creeks and streams and also along the muddy margins of salt pannes that are occasionally inundated by high tides. High, dry banks of channels and open areas of levees associated with salt ponds and muted tidal marshes tend to be favored sites for *C. senilis senilis*. Habitat utilized by both adults and larvae can be characterized as having extensive areas of fine silt or sandy clay-like soil, exposed to full sun, with minimal to moderate vegetation, and being located near water. *C. haemorrhagica* and *C. oregona oregona* have shown a preference for wet, sandy beach-like areas that may or may not be influenced by fresh water from creeks and canals.

**Biology**

The specific biology of San Francisco Bay tiger beetles is not well known. The information that follows is a general-
Invertebrates

ized biology for these insects drawn from the studies of other species and a summary article by Pearson (1988).

Adults of these beetles are active on hot, sunny days and are exceedingly quick both in flight and on the ground. When approached these insects will run away or fly for a short distance, land, and then face their pursuer. Larvae and adults are predators, feeding on other insects. Prey items for San Francisco Bay tiger beetles include but are not limited to the Brine Flies 

### Table 3.2 Known Collection Sites for Tiger Beetle Populations

<table>
<thead>
<tr>
<th>Location</th>
<th>Date Specimen(s) Collected</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Cicindela haemorrhagica</strong></td>
<td></td>
</tr>
<tr>
<td>Martinez</td>
<td>28 Aug 1959, 21 Sep 1959</td>
</tr>
<tr>
<td>Richmond Field Station</td>
<td>24 Apr 1993</td>
</tr>
<tr>
<td>Alameda, 1930</td>
<td>23 Aug 1930, 24 Aug</td>
</tr>
<tr>
<td>San Francisco</td>
<td>4 Jul 1932</td>
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<tr>
<td>Burlingame</td>
<td>31 Jan 1944 (****)</td>
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<tr>
<td>Lake Merritt</td>
<td>7 Oct 1969</td>
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<tr>
<td>Oakland</td>
<td>15 Aug 1902</td>
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<tr>
<td>Redwood City (nr Yt. Harbor)</td>
<td>15 Jan 1952</td>
</tr>
<tr>
<td>Redwood City (Saltmarsh)</td>
<td>31 Jul 1951</td>
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<tr>
<td>Palo Alto Yacht Harbor</td>
<td>29 Jun 1969</td>
</tr>
<tr>
<td>East Palo Alto</td>
<td>31 Jul 1951</td>
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<tr>
<td>Milpitas</td>
<td>15 Jul 1966, 26 Jul 1966</td>
</tr>
<tr>
<td>Newark (2 mi west of)</td>
<td>25 Jul 1975, 24 Jul 1980</td>
</tr>
<tr>
<td>Bayfarm Island</td>
<td>21 Jun 1990</td>
</tr>
<tr>
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<td><strong>Cicindela oregona</strong></td>
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<tr>
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<td>22 May 1952</td>
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<tr>
<td>Bayfarm Island</td>
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<tr>
<td></td>
<td>21 Jun 1990, 7 Jul 1993,</td>
</tr>
<tr>
<td></td>
<td>Aug 1993, 12 Apr 1993, 1 Sep 1995</td>
</tr>
<tr>
<td>Oakland (*)</td>
<td>Jun 1906, 16 Aug 1902</td>
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<tr>
<td>Concord (*)</td>
<td>27 Apr 1935</td>
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<tr>
<td><strong>Cicindela hirticollis</strong> (data from Graves 1988)</td>
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<td>Oakland</td>
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<td><strong>Cicindela senilis senilis</strong></td>
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<td>San Rafael (*)</td>
<td>23 May 1941, 20 May 1951</td>
</tr>
<tr>
<td>Milbrae</td>
<td>1 Sep 1912, 2 Jul 1912, 3 Oct 1914</td>
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<td>San Mateo</td>
<td>24 Oct 1952</td>
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<td>Redwood City (Harbor)</td>
<td>15 Jul 1952</td>
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<td>Palo Alto (Salt Marsh)</td>
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<td>28 Aug 1955, 28 Sep 1955</td>
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<td>Lake Merritt</td>
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<td>Alameda</td>
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<td>W. End Mowry Slough</td>
<td>19 Sep 1997</td>
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<tr>
<td>Brinker Marsh</td>
<td>Mar 1989, 10 Mar 1997</td>
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<tr>
<td>E. End Albrae Sough, Fremont</td>
<td>12 Mar 1997</td>
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<td>Dixon Rd., Milpitas</td>
<td>23 Jul 1956</td>
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<td>Milpitas (wet sand)</td>
<td>1 May 1966, 12 Oct 1966</td>
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<td>21 Mar 1947, 22 Mar 1947,</td>
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<td></td>
<td>27 Mar 1947, 12 Apr 1947,</td>
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</table>

* May or may not be within the confines of the Ecosystem Goals Project.

*** Probably a dubious record, suspect mislabeled specimen.

1 Information assembled from specimens contained within the California Academy of Sciences Insect Collection, University of California Berkeley Essig Museum, University of California Bohart Museum, San Jose State University Edwards Museum, San Mateo County Mosquito Abatement District Insect Collection, and private collections of Dr. J. Gordon Edwards and Wesley A. Maffei

158 Baylands Ecosystem Species and Community Profiles
Mury (1979) found that adults of some species of tiger beetles also fed on dead organisms. Faasch (1968) and Swiecimski (1956) found the adults located live prey visually while dead prey were found tactilely. Adult beetles tend to frequent the muddy margins of their habitat where prey items are readily encountered while the immature stages tend to be found in the drier areas.

The eggs, larvae and pupae are subterranean, with the larvae living in vertical burrows and waiting near the top to seize any prey that passes by (Figure 3.5). Prey items are captured with the mandibles and pulled down to the bottom of the burrow where it is ingested. Faasch (1968) found that a dark object against a light background released the prey-catching behavior. Burrows are enlarged by loosening the soil with the mandibles and using the head and pronotum to push the soil to surface. At the surface, the soil is flicked off by flipping the head and pronotum backward (Shelford 1908, Willis 1967). The depth of larval tunnels has been found to range between 15 and 200 cm depending on the age of the larva, the species of tiger beetle, the season and soil type and conditions (Criddle 1910, Willis 1967, Zikan 1929).

Larvae undergo three molts with the time for development lasting one to four years and averaging about two years (Willis 1967). Pearson and Knisley (1985) found that the availability of food affected rate of development and was therefore a limiting resource in the life cycle of tiger beetles. They found that ample prey shortened the developmental time from egg to adult with 60 days total developmental time having been observed for some laboratory reared beetles. Prior to pupation, the last instar larva plugs the tunnel entrance and excavates a chamber or pupal cell. The period for pupation is usually short, lasting no more than 30 days.

Larvae can be found throughout the year while adults are present from March through October. Peak adult activity for the south San Francisco Bay Cicindela senilis senilis populations is from late April through June (Maffei, unpub.). Blaisdell (1912) noted that adults of C. senilis, which emerged in the fall, would hibernate. C. haemorrhagica do not emerge until mid to late June and are usually present through September.

Reproduction

Males initiate copulation by approaching a female in short sprints which is similar to the intermittent sprinting used when foraging. Once close enough to a potential mate, the male leaps onto their back, grasping the thorax with his mandibles and the elytra with his front and middle legs. The male’s hind legs remain on the ground and the coupling sulci of the female receives his mandibles. Males frequently mount both males and females of any tiger beetle species present. Females try to dislodge intruding males by rolling on their backs, lurching and then running out into bright sunshine. It is believed that the fit of the male mandibles into the female sulci may be species specific and that this feature allows other males, and females of other species, to rid themselves of unwanted mates (Freitag 1974).

Oviposition usually occurs when the female touches the ground with her antennae and bites the soil with her mandibles. The ovipositor is then extended and with a thrusting motion of the abdomen a hole up to 1 cm is excavated. One egg is deposited in the hole and it is then covered over so that no evidence of disturbance exists. The choice of soil type for oviposition has been found to be extremely critical for many species (Knisley 1987, Leffler 1979, Shelford 1912, Willis 1967).

Availability of prey has been found to directly affect female mortality and the number of eggs produced. Adult beetles in prey poor habitats were only found to approach maximum fecundity during years of high rainfall and high prey populations (Pearson and Knisley 1985). Prey availability for larvae was found to affect the size of later instars, which ultimately affected the size of the adults produced and individual fecundity (Hori 1982a, Pearson and Knisley 1985).

Significance to Other Wetlands Taxa

These beetles may be a potential prey item for shore birds. Cramp and Simmons (1983) cite a stomach content analysis study of the European race of Snowy Plover, Charadrius alexandrinus alexandrinus, in Hungary which revealed the presence of 28 tiger beetles. Swarth (1983) noted that these beetles were occasionally eaten by Snowy Plovers found at Mono Lake. Marti (1974) found tiger beetle parts in burrowing owl pellets that were studied in the northeastern part of Larimer County, Colorado.

Conservation Needs and Limiting Factors

Iwago (1982) has stated that some tiger beetles are considered to be good indicators of coastal wetlands distur-
bance, with the least disturbed habitats having the greatest species diversity. San Francisco Bay tidally influenced wetlands appear to have two species of tiger beetle, with those sites that have had minimal disturbance or that have not seen much human activity for long periods of time having the highest populations (Maffei, unpub.). Unfortunately, few sites exist that have not been subjected to human activity. This has resulted in a loss of species diversity, with potential tiger beetle habitat usually having only a single species present and having small disjunct populations. Historically, there were sites that had more than one species present within a given habitat (i.e. Lake Merritt, Bayfarm Island and Burlingame).

San Francisco Bay populations of Cicindela senilis and C. haemorrhagica prefer to be near permanent or semi-permanent bodies of water utilizing tidal pannes with sizable unvegetated flats and/or nearby minimally vegetated levees. Cicindela haemorrhagica has shown a preference for sandy beach-like sites but can utilize dry, fine silty sites as is evidenced by the population at Russell Salt Marsh, Hayward. Both species of beetles need to have fine silty clay-like or sandy clay soils, that are unvegetated or sparsely vegetated, within in which to breed. Bright sunshine and minimal flooding are also important factors.

The immature stages of other species of tiger beetles have been found to inhabit a smaller range of the habitat than the adults and are not capable of tolerating as much variation in physical factors such as soil moisture, soil composition and temperature (Hori 1982b, 1982a, Knisley 1987, Knisley 1984, Knisley and Pearson 1981, Shelford 1912, Shelford 1908). The length and duration of flooding can also be important, although what the specifics of these parameters are for San Francisco Estuary tiger beetles is not clear.

Larochelle (1977) found that many species of adults are readily attracted to lights. What impact this might have on San Francisco Bay Tiger Beetles with respect to dispersal and survival is unknown.

References


Willis, H.L. 1967. Bionomics and zoogeography of tiger beetles of saline habitats in the Central United...
Western Tanarthrus Beetle
Tanarthrus occidentalis Chandler
Wesley A. Maffei

Description and Systematic Position
Tanarthrus occidentalis is a small beetle, approximately 3-5 mm in length, that belongs to the family Anthicidae (Figure 3.6). The head, pronotum, legs and abdomen are reddish-orange and the elytra are usually brown or black with the apical and basal third sometimes reddish or yellowish in color. This beetle can be separated from similar bay area Anthicid beetles by noting the distinct medial constriction of the eleventh antennal segment. It can further be separated from Formicilla spp., a similar appearing bay area Anthicid of marshes and grasslands, by examining the posterior margin of the mesepisternum which lacks a posterior fringe of long hairs.

Chandler (1979) has indicated that this beetle is very similar to T. iselini, which is found only in central New Mexico, but can readily be separated by antennal morphology.

Distribution
Tanarthrus occidentalis was first collected in 1976 and subsequently described as a new taxon by Chandler in 1979. Specimens were collected from the Cargill salt pans, now part of the San Francisco Bay National Wildlife Refuge, adjacent to Dumbarton Bridge, Alameda County, California. Additional populations have been identified from the salt pans of the Baumberg tract, Hayward, California, and from Bayfarm Island, Alameda, California. In 1996 the Bayfarm Island population was extirpated due to modification of their habitat in preparation for anticipated development. Surveys of the south and central San Francisco Bay area have revealed no other populations at this time (Maffei, unpub.). Figure 3.7 shows the locations around the Bay Area where T. occidentalis specimens have been collected, and Table 3.3 shows the collection dates.

Figure 3.6 Western Tanarthrus Beetle – Tanarthrus occidentalis

Figure 3.7 Known Tanarthrus occidentalis Localities Within San Francisco Bay Tidal and Diked Marshes
Invertebrates

Suitable Habitat

Tanarthrus occidentalis has been found in no other locality except for abandoned crystallizer ponds and salt pannes of southern San Francisco Bay. In all instances these sites remain dry for most of the year except during late winter when temporary pools of rainwater form. Habitat can be characterized as having extensive areas of salt crystals interspersed with open areas of fine silt and very little or no vegetative cover.

Biology

The biology of this beetle is not fully understood. Maffei (unpub.) has observed the Baumberg tract population and found that the adults commonly occur out on inactive, salt encrusted crystallizer ponds. These beetles were observed feeding on the carcasses of the brine flies Ephydra cinerea and Lipochaeta slossonae (family Ephydridae) which were still in the webs of unidentified Dictynid spiders. They appeared to function as "house cleaners" being able to move freely about the web site unmolested by the resident spider. Peak adult activity is May through September. The immature stages of this beetle have not been located at this time. Larvae of other members of the beetle family Anthicidae feed on detritus and one species has been recorded as a predator.

Reproduction

Unknown.

Significance to Other Wetlands Taxa

This beetle has been identified as part of the immature Snowy Plover Diet (Page et al. 1995, Feeney and Maffei 1991). Its relationship to other taxa, other than Dictynid spiders, that utilize abandoned salt crystallizers is unknown at this time.

Conservation Needs and Limiting Factors

The conservation needs and limiting factors associated with this beetle are not very clear. Its association only with salt encrusted areas, other than the margins of salt ponds, that remain dry for most of the year appears to be the primary limiting factor.

References


Additional Readings


Inchworm Moth
Perizoma custodiata
Wesley A. M. affei

Description and Systematic Position

Perizoma custodiata is a small moth, with a wingspan of approximately 22-29mm, that belongs to the family Geometridae. This moth, commonly known as a measuring worm or inch worm moth, has an alternating pattern of vertical light and dark bands on the fore wings with plain, pale tan hind wings (Figure 3.8). The variation in width and intensity of the fore wing banding has caused different entomologists to describe this moth as a new taxon on four different occasions (Guenee 1857, Hulst 1896, Packard 1876). Wright (1923) noted the difficulty in separating examples of the “different species” of the Pacific Coast recognized at that time, stating that they intergrade so much that he found it difficult to tell one from another.

Larvae are a uniform light green or tan in color and attain a maximum size of approximately 30mm.

Distribution

Coastal areas from central northern California south along the coast of Baja California and including the Gulf of California. Found throughout San Francisco Bay tidal and diked salt marshes. Figure 3.9 shows the locations around the Bay Area where Perizoma custodiata have been collected, and Table 3.4 shows the collection dates.

Suitable Habitat

Upper middle to high marsh that has berms or levees with adequate populations of Alkali Heath (Frankenia salina).

Biology

Adults are on the wing from March through November, with peak adult populations occurring during late spring and early summer.

Larvae have been observed feeding on Frankenia salina (M. affei, unpub.) and Packard (1876) has noted that the larvae of other members of the genus Perizoma live on low growing plants with the pupa being subterranean. Caterpillars have been observed on Alkali Heath that was inundated by high tides of 6.3 or greater at the Whale’s Tail Marsh, Hayward, California. The eggs and larvae have not been found during the winter months, and it is presumed that these moths overwinter as pupae.

Reproduction

The number of generations per season and the number of eggs per female is apparently unknown for San Francisco Bay populations.

Significance to Other Wetlands Taxa

Snowy plovers have been observed consuming adult moths at the Baumberg Tract in Hayward, California (Feeney and M. affei 1991). This insect may also be a part of other shore bird and passerine bird diets.

The digger wasp, Ammophila aberti, has been observed provisioning its nests with the larvae of this moth (M. affei, unpub.).

Adult moths are pollinators of Frankenia salina and are probably pollinators for many of the other flowering plants within diked and tidal marshes.
Table 3.4 Known Collection Sites For Perizoma custodiata

<table>
<thead>
<tr>
<th>Location</th>
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</thead>
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<td>15 Feb 1957, 21 Mar 1957, 19 Sep 1957</td>
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<td>Martinez</td>
<td>30 Aug 1962</td>
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<td>Richmond</td>
<td>18 Jun 1956, 12 Apr 1959</td>
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<td>Berkeley</td>
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<td>Berkeley (Bayshore)</td>
<td>27 Jul 1916, 16 May 1955</td>
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<td>Alameda</td>
<td>12 May 1918, 13 May 1920</td>
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<td>Shoreline Int. Ctr. (Hwyd)</td>
<td>2 Jul 1990</td>
</tr>
<tr>
<td>Baumberg Tract (Hayward)</td>
<td>24 Feb 1990, 1 Apr 1990</td>
</tr>
<tr>
<td>Napa</td>
<td>5 May (no year)</td>
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<tr>
<td>Petaluma</td>
<td>13 May 1936, 15 May 1938</td>
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<td>Mill Valley (Slough)</td>
<td>17 Jul 1950</td>
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<tr>
<td>Mill Valley</td>
<td>23 Mar 1920, 5 Sep 1923, 26 Nov 1924, 3 Oct 1926</td>
</tr>
<tr>
<td>South Marin Co. Shore</td>
<td>12 Apr 1950</td>
</tr>
<tr>
<td>San Francisco (Dunes)</td>
<td>7 Apr 1961</td>
</tr>
<tr>
<td>Millbrae</td>
<td>10 Sep 1914</td>
</tr>
<tr>
<td>San Mateo</td>
<td>3 Oct 1920</td>
</tr>
<tr>
<td>E. Palo Alto</td>
<td>May 1978</td>
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<tr>
<td>Bair Island</td>
<td>1 Mar 1987, 9 Mar 1997</td>
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1 Information assembled from specimens contained within the California Academy of Sciences Insect Collection, University of California Berkeley Essig Museum, University of California Bohart Museum, San Jose State University Edwards Museum, San Mateo County Mosquito Abatement District Insect Collection, and private collections of Dr. J. Gordon Edwards and Wesley A. Maffei.

Conservation Needs and Limiting Factors

Frankenia salina has been identified as the larval host plant for this moth (Maffei, unpub.). Upper middle to high marsh areas with small dense patches of this plant support fairly high numbers of this organism. Its wide distribution along the Pacific Coast would seem to preclude this organism from any immediate danger of extirpation.

References


Additional Readings


Pygmy Blue Butterfly
Brephidium exilis Boisduval
Wesley A. Maffei

Description and Systematic Position

Brephidium exilis, also known as the Pygmy Blue, is a small butterfly, with a wingspan measuring approximately 13-20mm (Figure 3.10). Adult butterflies have the dorsal surface of the wings brown with the basal third to half light blue. The ventral surface of the wings are grayish white with pale brown bands and a row of iridescent black and silver spots along the outer edge of the hind wing. The eggs are flattened, light bluish-green in color, and have a fine raised white mesh on the surface. Larvae are pale green or cream colored and have a finely punctate surface with white tipped tubercles, a yellowish white dorsal line, and a bright yellow substigmatic line (Figure 3.11). Some specimens may lack the lateral substigmatic line but all mature larvae have a frosted appearance which resembles the ventral surface of salt bush leaves or the flower heads of pigweed. The pupae can be quite variable in color but are usually light brownish yellow, have a dark brown dorsal line, and have the wing pads pale yellowish green in color sprinkled with brownish dots.

Three subspecies of this butterfly have been recognized with Brephidium exilis noted as the western subspecies (Scott 1986).

Distribution

Brephidium exilis is found from southwestern Louisiana and Arkansas westward to California and south to Venezuela (H owe 1975, Scott 1986). Strays have been noted as far north as Kansas and Idaho. This butterfly is widely distributed throughout the San Francisco Bay, being particularly abundant in salt marshes (Tilden 1965). Figure 3.12 shows the locations around the Bay Area where B. exilis have been collected, and Table 3.5 shows the collection dates.

Figure 3.10 Adult Pygmy Blue Butterfly – Brephidium exilis.

Figure 3.11 Brephidium exilis Egg and larva (from Comstock 1927)

Suitable Habitat

Prefers lowland areas such as alkali flats, salt marshes, vacant lots, roadsides and desert prairie with various Chenopodiaceae and Aizoaceae.

Biology

The adult flight period for San Francisco Bay populations is late February through October, with peak abundance occurring in September (Comstock 1927, Garth and Tilden 1986, Tilden 1965).

Larvae feed on most parts of the host plant. Recorded larval hosts are: Atriplex canescens, A. coulteri, A. serenana, A. leucophylla, A. patula hastata, A. semibaccata, A. rosea, A. cordulata, A. hymenelytra, A. coronata, A. lentiformis breweri, Suaeda fruticosa, S. californica, S. torreyana, Salicornia virginica, Chenopodium album, C.
Table 3.5 Known Collection Sites For *Brephidium exilis*

<table>
<thead>
<tr>
<th>Location</th>
<th>Date Specimen(s) Collected</th>
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<tr>
<td>West Pittsburg</td>
<td>15 Apr 1957</td>
</tr>
<tr>
<td>Avon</td>
<td>27 Aug 1972</td>
</tr>
<tr>
<td>Richmond Point</td>
<td>3 Oct 1964</td>
</tr>
<tr>
<td>Richmond</td>
<td>10 Aug 1953</td>
</tr>
<tr>
<td>Alameda</td>
<td>12 May 1918, 17 May</td>
</tr>
<tr>
<td>Larkspur</td>
<td>20 Sep 1958</td>
</tr>
</tbody>
</table>

References


Additional Readings

Summer Salt Marsh Mosquito
*Aedes dorsalis* (Meigen)
Wesley A. Maffei

**Description and Systematic Position**

The summer salt marsh mosquito, *Aedes dorsalis*, is a medium-sized mosquito measuring approximately 5-6 mm in length. Freshly emerged adults are one of the most brightly colored marsh mosquitoes found within the San Francisco Estuary. These insects are brilliant gold in color, have a dorsal white band running the length of the abdomen and have broad white bands on the tarsal segments of the legs. Older specimens may be yellow or yellowish-brown in color and the markings on the abdomen may be incomplete if the scales have been rubbed off. The immature stages can be identified by insertion of the siphon tuft at or beyond the middle of the siphon tube, a broadly incomplete anal saddle, presence of a weak saddle hair and moderate to short anal papillae. The presence of single upper and lower head hairs has been used as an additional diagnostic feature but this can be inconsistent, especially in later instar larvae.

The similarity of this mosquito to *Aedes melaninomon* Dyar has resulted in some confusion with early efforts to identify both adults and larvae. Detailed studies of different populations of both of these mosquitoes have helped to clarify and verify the systematic position of both of these insects (Bohart 1956, Chapman and Grodhaus 1963).

**Distribution**

This mosquito can be found throughout most of the United States, southern Canada, Europe and Asia (Carpenter and LaCasse 1955, Darrie and Ward 1981). Within California, this mosquito can be found in coastal salt marshes and the brackish waters of the Sacramento and San Joaquin Delta (Bohart 1956, Bohart and Washino 1978).

**Suitable Habitat**

Larvae are found in a variety of brackish and freshwater habitats throughout their world range (Carpenter and LaCasse 1955). Within San Francisco Bay *A. dorsalis* are usually encountered in temporarily flooded tidal marsh pannes, heavily vegetated ditches and brackish seasonal wetlands. Adults prefer open habitats such as grasslands, open salt marsh and the edges of woodlands.

**Biology**

Adults are aggressive day biting mosquitoes that have been found capable of traveling distances of more than 30 miles (Rees and Nielsen 1947). Flights of adults in Alameda County have been known to disperse distances of more than five miles from their larval source (Maffei, unpub.). Garcia and Voigt (1994) studied the flight potential of this mosquito in the lab and found that the adults exhibited strong flight characteristics which they believed helped them to adapt to the strong winds encountered in their preferred open habitats. Females are readily attracted to green, grassy fields and will rest there waiting for available hosts (Maffei, unpub.).

Host studies have shown that large mammals are preferred, especially cattle and horses (Edman and Downes 1964, Gunstream et al. 1971, Shemanchuk et al. 1963, Tempelis et al. 1967). The effects of adult feeding activity on livestock can be severe resulting in reduced feeding and in some instances injury to animals attempting to evade severe attacks. Recent adult activity within the San Francisco Estuary has impacted outdoor school activities, businesses and residents, resulting in at least two instances where medical attention was required for people reacting to multiple bites (Maffei, unpub.).

Eggs are deposited individually on the mud along the edges of tidal pools or the receding water line of brackish seasonal wetlands. Winter is passed in the egg stage and hatching occurs with the first warm weather of spring. Additional hatches occur with subsequent refloodings of the larval habitat. Eggs can remain viable for many years with only part of any given brood hatching during any single flooding event.

The larval stage can last from four to fourteen days with duration being primarily dependent on temperature. Other factors that can regulate rate of larval development include competition for space and quality and availability of nutrients. Rees and Nielsen (1947) found larvae that completed their development in saline pools of the Great Salt Lake with salt concentrations as high as 120 ‰. Washino and Jensen (1990) reared larvae, from Contra Costa County salt marshes, in solutions simulating 0, 10, 50 and 100% concentrations of seawater and found that survivorship improved as salt content approached that of seawater.

Total developmental time, from egg to adult, has been observed to occur in less than one week (Maffei, unpub.).

**Reproduction**

Male mating swarms have been observed occurring over low growing bushes, prominent objects and open fields (Dyar 1917, Garcia et al. 1992). Both observations noted that swarming activity began at sunset and that the swarms were not more than two to three meters above the ground. Swarming and mating usually occurs on the marsh within a few days of adult emergence and is followed by random dispersal of host seeking adults.

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The number of gonotrophic cycles and eggs produced per female remains unclear for San Francisco Bay populations. Early work by Telford (1958) found that 12 broods and approximately eight generations occurred during one breeding season at Bolinas in Marin County. The number of generations per year does vary with respect to weather and tidal conditions.

Significance to Other Wetlands Taxa

This species of mosquito is commonly found in association with the tidal pool brine fly Ephydra millbrae and the water boatman Trichocorixa reticulata. Both the brine fly and the water boatman have been identified as food sources for shorebirds and waterfowl (Anderson 1970; Feeney and Maffei 1991; Howard 1983; Maffei, unpub.; Martin and Uhler 1939). The larvae of this mosquito may also be a food source for these birds and adults may be a food source for swallows.

Conservation Needs and Limiting Factors

This mosquito, like other species of mosquitoes, is extremely opportunistic. Care must be taken when altering or restoring seasonal or tidal wetlands. Sites that drain poorly will create habitat that can readily produce very large numbers of aggressive biting adults. Plans for long term maintenance of seasonal and tidal wetlands should include resources for mosquito control as the need arises. The dynamic nature of these types of habitats coupled with human activities can easily convert a non-breeding site into a major mosquito producing source.

References


Winter Salt Marsh Mosquito
Aedes squamiger (Coquillett)
Wesley A. Maffei

Description and Systematic Position

Aedes squamiger is a medium-sized to large mosquito, measuring approximately 6-9mm in length, that belongs to the fly family Culicidae (Figure 3.13). Adults have a distinctive black and white speckled appearance and large, flat scales along the wing veins which separates this fly from other San Francisco Bay mosquitoes. Larvae can be identified by the presence of an incomplete anal saddle, a siphon tuft distal to the pecten row, an anal saddle hair as long or longer than the anal saddle, and upper and lower head hairs that are usually branched (Figure 3.14).

This mosquito was described as a new taxon by Coquillett in 1902 from specimens collected from the cities of Palo Alto and San Lorenzo, California. Bohart (1948) differentiated the larvae and pupae of Aedes dorsalis and Aedes squamiger thereby providing a means of separating the immature stages of these two species which are very similar in appearance. In 1954, Bohart described and provided keys to the first stage larvae of California Aedes and further clarified the differences between these two mosquitoes.

Distribution

This mosquito is found along the Pacific Coast region from Marin and Sonoma counties, California, south to Baja California, Mexico (Bohart and Washino 1978, Carpenter and LaCasse 1955, Darsie and Ward 1981, Freeborn and Bohart 1951). Figure 3.15 shows the distribution of Aedes squamiger in 1950. The current distribution within the San Francisco Bay area is very similar, with additional sites having been identified along the shoreline of the East Bay.

Suitable Habitat

Preferred habitat consists primarily of coastal pickle weed tidal and diked marshes, especially salt marsh pools that are diluted by winter and early spring rains. Cracked ground of diked wetlands and old dredge disposal sites are also a favorite habitat for deposition of eggs and development of larvae. This mosquito prefers brackish or saline habitats and has not been found in truly fresh water marshes. Bohart, et. al. (1953) found larvae of various stages in pools with salinities ranging from 1.2% to 35%. Studies by Garcia and coworkers (1992, 1991) indicated that optimal larval development occurred at salinities between 5% and 15%.

Biology

Eggs hatch as early as late September and can continue to hatch with the accumulation of rainfall from each successive storm event. Maffei (unpub.) found larvae that hatched from the incidental flooding of a marsh by a duck club as early as late September. Bohart, et. al. (1953) states that three to six major hatches of eggs occur during the fall months. It is believed that only part of the eggs laid during the prior spring season hatch with a decreasing percentage of the remaining eggs hatching during successive years. Garcia, et al. (1991) found that as many as four floodings were necessary to hatch all of the eggs from field collected samples. Bohart and Washino (1978) state that the eggs are usually dormant from April through September and that this obligatory diapause is terminated by the decreasing fall temperatures that fall below 7°C. Garcia et al. (1991) found that hatching does not occur until the eggs have been exposed to temperatures that are less than 10°C. Voigt (pers comm.) believes that once the eggs have been thermally
conditioned that hatching can then occur anytime in the future following submersion. This may possibly help to explain summer hatches following flooding of sites by inadvertent human activity (Maffei, unpub.).

Larvae are principally found in salt marsh pools that are diluted by fall and winter rainfall. Bohart and coworkers (1953) found that a minimum of 48 days were required for the development of the aquatic stages before adult emergence, with the first pupae having been found during the first week of February. Under "normal" conditions pupae are usually found from the last two weeks of February through the beginning of March. Estimates of the number of larvae per acre vary from 1.65 million to 1.45 billion depending on environmental factors (Aarons 1954, Aarons et al 1951, Lowe 1932). Larvae are capable of remaining submerged for extended periods of time where they browse on vegetation and mud. Garcia, et al (1990) calculated the minimum developmental threshold for development of larvae to the adult stage to be 4.4°C. Additional studies by Garcia and coworkers (1991) found that first and second instar larvae had developmental thresholds that were 2-4°C lower than the later instars. From these data, they concluded that the lower developmental thresholds of the earlier instars allowed larvae from later hatching installments to emerge as adults in closer synchrony with those larvae that hatched earlier in the season. They also noted that larvae and pupae could survive in the mud at sites that underwent periodic draw-down of the water. Garcia, et al. (1990) also studied the diapause habit of the last instar larvae and concluded that this interesting trait probably contributed in some degree to the partial synchronous emergence of the adults.

Adults usually emerge during the last week of February through the end of March. Emergence of adults in April has occurred from unusually heavy late winter and early spring rains that have caused late egg hatches with rapid larval development. Adults usually fly to ar-

**Figure 3.15** Aedes Squamiger Distribution in the San Francisco Bay Area, 1950

From Aarons, 1954
Reproduction

Observations on mating swarms have shown that Aedes squamiger tends to swarm approximately one hour before to one-half hour after sunset (Garcia et. al. 1992). Swarms can consist of a few to several thousand individuals that hover over prominent objects such as trees or large bushes and can occur at heights ranging from six to approximately 50 feet (Bohart and W ashin 1978, Garcia et. al. 1992). Garcia et al. (1992, 1983) found that adults traveled back and forth to the marshes quite readily producing a new batch of eggs with each trip. He also found that the highest parous condition observed was seven, with average parity rates ranging between 3 and 5.4. Garcia et al. (1992) found a direct correlation between wing length and the number of eggs produced with larger females producing more eggs. The maximum number of eggs produced per female was less than 250. Garcia et al. (1990) also found that temperature played an important role in longevity, ovarian development and oviposition. Females held at 15°C were still alive 50 days after their last blood meal and average longevity was about 35 days when kept at 20°C. The minimum temperature threshold for ovarian development or oviposition was found to be about 15°C.

Significance to Other Wetlands Taxa

Aedes squamiger larvae are frequently found in association with larvae of the Summer Salt Marsh M osquito, Aedes dorsalis, and the Winter Marsh Mosquito, Culiseta inornata. The adults of these mosquitoes may be a possible food source for swallows and the larvae may be a food source for waterfowl.

Conservation Needs and Limiting Factors

This mosquito, like other species of mosquitoes, is extremely opportunistic. Care must be taken when altering or restoring seasonal or tidal wetlands. Sites that drain poorly will create habitat that can readily produce very large numbers of aggressive biting adults. Plans for long term maintenance of seasonal and tidal wetlands should include resources for mosquito control as the need arises. The dynamic nature of these types of habitats coupled with human activities can easily convert a non-breeding site into a major mosquito producing source.

References


Additional Readings


Washino’s Mosquito

Aedes washinoi Lansaro and Eldridge

Description and Systematic Position

Aedes washinoi was described as a new taxon by Lansaro and Eldridge in 1992 and was determined to be a sibling species of Aedes divis and Aedes increpitus. Prior to 1992, all three species of mosquitoes were known as Aedes increpitus. Adults of this mosquito are almost impossible to separate from its sibling species, when using morphological features, and can also sometimes be confused with Aedes squamiger. The easiest way to distinguish Ae. squamiger and Ae. washinoi is to examine the wing scales. Aedes squamiger has very broad, flat, plate-like scales on the wings whereas Ae. washinoi will have the usual thin, pointed wing scales. The wings of Ae. washinoi will also tend to be uniformly dark with a concentration of pale scales on the anterior wing veins. In all other respects, both Ae. squamiger and Ae. washinoi share a similar black and white speckled appearance. The larvae of this mosquito can be difficult to separate but Darsie (1995) has provided additions to Darsie and Ward’s 1981 keys to facilitate identification.

Distribution

This mosquito is found from Portland, Oregon south to Santa Barbara, California and eastward into the lower Sierra Nevada mountains. Populations of this mosquito have also been found along the eastern Sierra Nevada Range at Honey Lake.

Suitable Habitat

Within the San Francisco Estuary the preferred habitat is shallow ground pools and upland fresh to slightly brackish water sites that are near to salt marshes or in riparian corridors. These habitats also tend to be dominated by willow or cotton wood trees and/or black berry vines.
Biology

Larvae usually hatch during early winter after a series of successive storm events has filled ground depressions with water. Additional hatches of larvae can occur if late winter and early spring rains refill drying larval sites. Larvae of this mosquito also exhibit a late fourth instar diapause and partial synchronous adult emergence similar to that observed in *Aedes squamiger*. Adults emerge during late winter and early spring and can persist through early June, depending on weather conditions.

Females are aggressive day biting mosquitoes that tend not to travel far from their larval sources. Maffei (unpub.) found that adult mosquitoes traveled a maximum distance of one and one-half miles from their larval habitat and that local, man made canals were used as a passageway into the surrounding community.

Eggs are deposited in the muddy margins adjacent to the receding water line of the larval habitat and hatch the following winter when reflooded.

Reproduction

Adults have been observed swarming under or near the tree canopy of their larval habitat (Garcia, et al. 1992).

Significance to Other Wetlands Taxa

Unknown.

Conservation Needs and Limiting Factors

This mosquito, like other species of mosquitoes, is extremely opportunistic. Care must be taken when altering or restoring seasonal wetlands or riparian corridors. Sites that have shallow ground pools and willow or cottonwood trees or blackberry vines will create habitat that can readily produce very large numbers of aggressive biting adults. The restoration of historical willow groves should not occur if homes are within two miles of the project site.

References


Additional Readings


Western Encephalitis Mosquito

*Culex tarsalis* Coquillett

Wesley A. Maffei

Description and Systematic Position

The western encephalitis mosquito is a medium sized mosquito measuring approximately 5-6 mm in length. This fly was described in 1896 as a new taxon by Coquillett from specimens gathered in the Argus Mountains of Inyo County, California (Belkin et al. 1966).

Adults can be identified by using the following morphological features: legs with bands of pale scales overlapping the tarsal joints; femur and tibia of the hind legs with a pale stripe or row of pale spots on the outer surface; proboscis with a complete median pale band; ventral abdominal segments with v-shaped patches of darkened scales; and the inner surface of the basal antennal segment with patches of pale scales. The larvae can be recognized by the four to five pairs of ventrally located siphon tufts that are nearly in line with each other (Figure 3.16) and the 3-branched lateral abdominal hairs found on segments III to VI.

![Figure 3.16](image.png)
Distribution
This mosquito has been found in central, western and southwestern United States, southwestern Canada and northwestern Mexico (Carpenter and LAcasse 1955, Darsie and Ward 1981). Within California, this fly has been found in every county from elevations below sea level to almost 10,000 feet (Bohart and Washino 1978, M eyer and Durso 1993).

Suitable Habitat
The immature stages are found in all types of fresh water habitats except treeholes. Poorly drained pastures, rice fields, seepages, marshes and duck club ponds are especially favored as breeding habitat for this mosquito. Telford (1958) found larvae in salt marsh pools with salinities up to 10 %/o. Urban sources include poorly maintained swimming pools, ornamental ponds, storm drains, flood control canals, ditches, wastewater ponds and most man made containers (Beadle and H armston 1958, Bohart and Washino 1978, H armston et al. 1956, M eyer and Durso 1993, Sjogren 1968).

Adults rest by day in shaded or darkened areas such as mammal burrows, tree holes, hollow logs, under bridges, in caves, in eses and entry ways of residences, brush piles and in dense vegetation (M ortenson 1953, Loomis and G reen 1955, H arwood and H alfll 1960, Price et al. 1960, Rykman and Arakawa 1952).

Biology
Adult females of this species usually feed at night. Precipitin tests indicate a wide variety of hosts consisting of various birds and mammals with an occasional reptile or amphibian (Anderson et al. 1967, Edman and D owne 1964, G unstream et al. 1971, H ayes et al. 1973, Reeves and H ammon 1944, Rush and T empelis 1967, Shemanchuk et al. 1963, T empelis 1975, T empelis et al. 1967, T empelis et al. 1965, T empelis and W ashino 1967). Reeves (1971) states that host availability and season are probably the most important considerations in the adult host feeding pattern. The availability of nesting birds during spring and early summer may account for the preponderance of identified, early season, avian blood meals. With the progression of the summer season, availability and behaviour of bird hosts varies and a switch to mammal hosts occurs (H ammon et al. 1945, H ayes et al. 1973, Reeves and H ammon 1944, Reeves et al. 1963, T empelis et al. 1967, T empelis and W ashino 1967). Adults pass the winter months in facultative diapause which is triggered by short day length and low ambient temperatures. In the warmer parts of southern California adults are active year round while in San Francisco Bay populations inactivity usually occurs from December through February. Additional periods of low temperatures or unseasonably warm winters can vary the time spent in diapause.

Flight range studies indicate that this mosquito will readily disperse from its larval source. Reeves et al. (1948) found that adults generally dispersed two miles or less, although prevailing winds helped to distribute marked females up to three miles. Bailey et al. (1965) studied the dispersal patterns of Yolo County, California populations and found that prevailing winds were important to adult dispersal with significant numbers of adults having traveled seven miles within two nights. The maximum distance traveled was recorded at 15.75 miles. From their studies they concluded that the likely dispersal distance of Sacramento Valley populations was probably about 20-25 miles. It was further concluded that most locally controlled mosquito sources are repeatedly reinfested during the summer because these mosquitoes travel so readily with the wind.

The larval stages feed on a wide variety of microorganisms, unicellular algae and microscopic particulate matter. The amount of time required to complete development from egg to adult varies depending on water temperature, availability of food and crowding. Bailey and G ieke (1968) found that water temperatures of 69°F to 86°F were optimal for larval development. Beyond 86°F, the larval stage lasted about eight days but mortality was very high. M ead and C onner (1987) found the average developmental rates from egg to adult to be 18.7 days at 67°F and 7.4 days at 88°F.

Reproduction
M ale mating swarms occur shortly before to just after sunset. H arwood (1964) found that initiation of the mating swarm was related to changes in the light intensity and that light levels of approximately 7 foot candles would initiate crepuscular flight activity. He further found that lab colonized males could be induced to swarm when abrupt changes in light intensity occurred.

Lewis and Chrestenson (1970) studied female ovipositional behaviour and found that the initial search for oviposition sites by females occurs close to the lowest available surface. Groups of eggs, also known as egg rafts, are deposited directly onto the water with the average number of eggs per raft varying between 143 to 438 (Bock and M ilby 1981, Buth et al. 1990, Reisen et al. 1984). Environmental factors such as water temperature, crowding and availability of food have been found to affect development of the immature stages, which in turn, affects the size of the female mosquito and ultimately the number of eggs and egg rafts produced. Logan and H arwood (1965) studied the effects of photoperiod on ovipositional behaviour of a W ashington strain of Culex tarsalis and found that peak oviposition occurred within the first hour of darkness and light.
Autogeny, or the development of eggs without a blood meal, does occur with this mosquito. Moore (1963) found that autogenous Culex tarsalis from Sacramento Valley, California, produced an average of 116 eggs per female with an observed maximum of 220. He also found that the level of autogeny decreased from spring to summer. Spadoni et al. (1974) also studied autogeny in Culex tarsalis populations from the same region finding similar results and detecting autogeny as early as April. They further found that no autogenous egg development was observed in overwintering females from November through February and that the mean number of eggs produced per autogenous female was 144.

Significance to Other Wetlands Taxa

This mosquito is the primary vector of Western Equine Encephalitis (WEE) and Saint Louis Encephalitis (SLE) viruses for most of the western United States (Brown and Work 1973, Longshore et al. 1960, Reeves and Hammon 1962, Work et al. 1974). Rosen and Reeves (1954) have also determined that this fly is an important vector of avian malaria.

Larvae of the Winter Marsh Mosquito, Culiseta inornata, are frequently found with the immature stages of this mosquito during fall and spring. The larvae of this insect may be a possible food source for waterfowl.

Conservation Needs and Limiting Factors

Sound water management practices should include consultations with local public health and mosquito or vector control agencies to prevent or at least minimize the production of this mosquito from managed, restored or newly created wetlands. Adequate resources need to be provided in all short and long term management plans to help protect humans and horses from the encephalitis viruses that can be vectored by this mosquito.

References


Additional Readings


Winter Marsh Mosquito  
*Culiseta inornata* (Williston)  
Wesley A. Maffei

**Description and Systematic Position**

The winter marsh mosquito was described from specimens collected in the Argus Mountains, Inyo County, California, in 1893 (Belkin, et al 1966). This insect is one of California's largest mosquitoes, measuring approximately 8-10 mm in length. Adults are generally light brown to reddish-brown in color and lack any unusual or distinctive markings. Diagnostic features of the imagines include: tip of the abdomen bluntly rounded; wings with the radial and medial cross veins nearly in line with each other; anterior wing veins with intermixed light and dark scales; and wings without distinct patches of dark scales (*Figure 3.17*). Larvae can be identified by the presence of only one tuft of hairs inserted near the base of the pecten row on the siphon and by having the lateral hairs of the anal saddle distinctly longer than the anal saddle (*Figure 3.18*).

**Distribution**

This mosquito can be found throughout the United States, southern Canada and northern Mexico over a wide range of elevations and habitats (Carpenter and LaCasse 1955). Populations of the winter marsh mosquito have been found throughout California except in Mariposa County (Meyer and Durso 1993).

**Suitable Habitat**

The immature stages can be found in a wide variety of habitats ranging from duck club ponds, ditches, seepages, rainwater pools, salt marshes and manmade containers. Telford (1958) found larvae in Marin County marshes with salinities ranging from 8 ‰ to 26 ‰.

Adults are usually found resting near their larval habitats during their breeding season while summer aestivating adults are presumed to utilize animal burrows in upper marshes and adjacent uplands (Barnard and Mulla 1977, Shemanchuk 1965).

**Biology**

Adults are present fall, winter and spring and enter facultative diapause in the summer as a means of surviving the hot, dry California summers. Aestivating females are thought to emerge from mammalian burrows and shelters in the fall following decreased temperatures and the first fall rains. Meyer, et al. (1982a, 1982b) found that optimal flight activity occurred between temperatures of 48°F and 64°F, with a sharp decrease below 43°F and above 64°F. Washino, et al. (1962) studied populations of this mosquito in Kern County, California and found that small numbers of adult females persisted throughout the summer period.

Adult female mosquitoes feed primarily on large domestic mammals although populations associated with brackish marshes have been significantly pestiferous to humans within the San Francisco Estuary (Bohart and Washino 1978; Maffei, unpub.). Precipitin tests have shown that the primary hosts are cattle, sheep, horses and pigs (Bohart and Washino 1978, Edman and Downe 1964, Edman et al. 1972, Gunstream et al. 1971, Reeves and Hammon 1944, Shemanchuk et al. 1963, Tempelis 1975, Tempelis et al. 1967, Tempelis and Washino 1967, and Washino et al. 1962).

Flight range studies have found that the maximum distance traveled was 14 miles (Clarke 1943). Adults of San Francisco Bay populations tend to stay close to their larval source, usually traveling less than two miles for a blood meal. Wind and proximity of available hosts are probably important factors affecting adult dispersal and may help account for the variability observed between different populations of this mosquito.
Adults can be attracted to lights. Bay area mosquito abatement Districts monitor adult populations of this mosquito by using New Jersey light traps. Barnard and Mulla (1977) found that the trapping efficiency of New Jersey light traps could be improved by increasing the intensity of the incandescent light bulbs used from 25W to 100W.

Studies of lab colonized females by Owen (1942) found that the average life expectancy for adults was about 97 days with a maximum of 145 days. Weather conditions, specifically temperature and humidity, and availability of nutrients will affect adult longevity.

Total developmental time from egg to adult has been studied by Shelton (1973) and Mead and Conner (1987) and both found that water temperatures above 78°F were lethal to larval development. Average developmental times ranged from 48 days at 51°F to 13 days at 74°F. Shelton (1973) also noted that as water temperature increased beyond 68°F, average body weight and adult survivorship decreased markedly.

Reproduction

Rees and Onishi (1951) found that adults usually do not swarm and that freshly emerged females are mated by waiting males. Copulation usually occurs end to end vertically, with the female above the male, and is completed in about 3.5 to 6.5 hours.

Groups of eggs, also known as egg rafts, are deposited directly on the water. Buxton and Breland (1952) studied the effects of temporary dessication and found that eggs were still viable after three to four days exposure in damp leaves at various temperatures. They also found that the eggs tolerated exposure to temperatures as low as 17.6°F and had a hatch rate as high as 98%. The survival of larvae hatched from eggs exposed at 17.6°F was low varying from 50% to 100% mortality following 24 and 48 hours exposure respectively.

Significance to Other Wetlands Taxa

Winter Marsh Mosquito larvae are frequently found in association with larvae of Aedes squamiger and the Encephalitis Mosquito, Culex tarsalis. The larvae of this mosquito may be a possible food source for waterfowl.

Conservation Needs and Limiting Factors

This mosquito, like other species of mosquitoes, is extremely opportunistic. Care must be exercised when managing, altering or restoring seasonal wetlands. Sites that pond water will produce very large numbers of adults. Care must be exercised when manipulating water levels in diked marshes. The fall flooding of these types of wetlands for waterfowl management can produce enormous numbers of adults. The proximity of human habitation or recreational facilities can be seriously affected by the biting activity of these mosquitoes.

References


Clarke, J.L. 1943. Studies of the flight range of mosquitoes. J. Econ. Ent. 36:121-122.


Brine Flies
Diptera: Ephydridae
Wesley A. Maffei

Description and Systematic Position

There are numerous species of brine flies (Diptera: Ephydridae) that can be found within the confines of the San Francisco Bay region. Three are exceptionally numerous within the bay’s tidal and diked seasonal wetlands. These are Ephydra cinerea, Ephydra millbrae (Figure 3.19), and Lipochaeta slossonae (Figure 3.20). Adults can readily be recognized by the following features: head—lacking oral vibrissae, having a swollen protruding face, and having small diverging postvertical setae; wings—with the costa broken near the subcosta and humeral crossvein; wings—with the costa broken near the subcosta and humeral crossvein, and lacking an anal cell.

Adult flies are small in size (E. cinerea 2-3 mm in length, E. millbrae 4-5 mm in length, and L. slossonae 2-3 mm in length) and have unpatterned wings. The coloration for each is as follows: E. cinerea—opaque bluish-grey with a greenish tinge and legs with knees and most tarsal segments yellow; E. millbrae—brownish grey with brown legs; and L. slossonae—whitish grey with a black-brown thoracic dorsum and legs having yellow tarsal segments.

The immature stages are small yellowish-white larvae bearing eight pairs of ventral prolegs with two or three rows of hooks. The last pair of prolegs are enlarged and have opposable hooks and the last abdominal segment bears elongate respiratory tubes with terminal spiracles. The puparium is similar in shape to the last larval stage and is generally dark yellow to brown in color (Figure 3.21).

Distribution

Ephydra millbrae is found throughout the San Francisco Bay Area in mid to upper marsh tidal pools that are infrequently affected by the tides. E. cinerea

Additional Readings


Additional Readings

is closely associated with hypersaline environments, especially salt ponds of the south and north bay. Lipochaeta slossonae is commonly found in or near crystallizer ponds of the south bay and possibly also in salt ponds with salt concentrations somewhat above that of sea water in other parts of the San Francisco Bay region. Figure 3.22 shows the locations around the Bay Area where brine flies have been collected, and Table 3.6 shows the collection dates.

**Suitable Habitat**

Saline and hypersaline environments.

**Biology**

Simpson (1976) has summarized marine Ephydrid fly biology and a modified portion of that is presented here. Eggs are deposited in the water and hatch after one to
five days. The larva immediately begins feeding and will pass through three instars. First and second instar larvae shed their cuticles in order to pass on to the next larval stage. The cuticle of the last larval instar is not shed but instead forms the protective pupal covering, also known as the puparium. Adults emerge three to ten days after the onset of pupation by inflating a balloon-like ptilinum inside their heads. The ptilinum forces a circular cap off of the front of the puparium allowing the adults to emerge. Deflation of the ptilinum and attainment of normal adult coloration occurs within 0.5 to 1.5 hours. Total developmental time from deposition of eggs to emergence of adults ranges from two to five weeks.

Adults are generally reluctant to fly and when disturbed will usually fly very close to the ground for very short distances (Simpson 1976 and Wirth 1971). Lipochaeta slossonae adults have the peculiar habit of resting with the wings and legs held very close to the body, giving the appearance of a tube or torpedo. Should the wind cause them to lose their footing, they simply roll freely across the substrate until stopped by some object such as large salt crystals of crystallizer ponds or a spiders web. Dictynid spiders frequently build webs on crystallizer ponds collecting large numbers of these flies (Maffei, unpub.).

Precise food habits have been determined for only a few species of Ephydrids with adults of E. cinerea known to feed on masses of blue-green algae and the alga Enteromorpha sp. while L. slossonae utilizes various diatoms and dinoflagellates. Cheng and Lewin (1974) observed that L. slossonae would fluidize the silt or sandy substrate by vigorously shaking their bodies, thereby freeing some of the microorganisms upon which they feed.

### Table 3.6 Known Collection Sites For Brine Flies

<table>
<thead>
<tr>
<th>Location</th>
<th>Date Specimen(s) Collected</th>
<th>Location</th>
<th>Date Specimen(s) Collected</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Ephydra cinerea</strong></td>
<td></td>
<td><strong>Ephydra millbrae</strong></td>
<td></td>
</tr>
<tr>
<td>Oakland (Tide Flat)</td>
<td>20 Jul 1937</td>
<td>Sears Pt. (Solano Co.)</td>
<td>29 Jun 1951</td>
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<tr>
<td>San Leandro</td>
<td>19 Nov 1947</td>
<td>Mill Valley (Slough)</td>
<td>17 Apr 1950</td>
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<tr>
<td>Baumberg Tract (Hayward)</td>
<td>25 May 1989, 2 Jun 1989, 8 Jun 1989</td>
<td>Tiburon</td>
<td>5 Jul 1927</td>
</tr>
<tr>
<td>Fremont (Mouth of Coyote Hills Slough)</td>
<td>15 Jul 1976</td>
<td>San Francisco</td>
<td>22 May 1915</td>
</tr>
<tr>
<td>Newark</td>
<td>13 Aug 1930</td>
<td>Milbrae</td>
<td>20 Mar 1908, 1 Sep 1912</td>
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<td>Alvarado</td>
<td>2 Aug 1931</td>
<td>San Mateo</td>
<td>3 Oct 1920</td>
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<tr>
<td>Alviso (Artesian Slough)</td>
<td>1 Jul 1980</td>
<td>Foster City</td>
<td>20 Mar 1973</td>
</tr>
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<td>Alviso</td>
<td>2 Oct 1969, 18 Nov 1971</td>
<td>Redwood City</td>
<td>Apr 1923, 10 Apr 1923</td>
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<td>Milpitas</td>
<td>29 Nov 1974</td>
<td>Menlo Park</td>
<td>31 Jul 1955</td>
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<tr>
<td>San Mateo</td>
<td>3 Oct 1920, 4 Aug 1925, 10 May 1931</td>
<td>Dumbarton Dr. (San Mateo Co.)</td>
<td>30 Dec 1947</td>
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<td>Palo Alto</td>
<td>28 Jul 1894, 6 Aug 1894, 30 Jul 1915</td>
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<td>Mountain View</td>
<td>2 Apr 1906</td>
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<td></td>
<td>12 May 1915, 18 May 1915, 12 Jul 1924</td>
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<tr>
<td><strong>Lipochaeta slossonae</strong></td>
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<tr>
<td>Oliver Salt Ponds (Hayward)</td>
<td>5 Aug 1989</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baumberg Tract (Hayward)</td>
<td>4 Jun 1989</td>
<td></td>
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</tr>
</tbody>
</table>

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1 Information assembled from specimens contained within the California Academy of Sciences Insect Collection, the University of California Berkeley Essig Museum, the University of California Bohart Museum, the San Jose State University Edwards Museum, the San Mateo County Mosquito Abatement District Insect Collection, and the private collections of Dr. J. Gordon Edwards and Wesley A. Maffei.
Larvae apparently feed on the same organisms as the adults (Brock, et al. 1969).

The known salinity tolerances for the different brine flies varies. Jones (1906) observed that E. millbrae will occur in salt water pools with salinities up to 42 ‰. Ephydra cinerea and L. slossonae seem to prefer saline environments much higher than 42 ‰ but are not entirely restricted to these hypersaline habitats (Maffeii, unpub.).

Nemenz (1960) studied the ability of immature E. cinerea to maintain proper water balance in high saline environments and found that the larvae had a normal osmotic pressure of 20.4 atmospheres in their haemolymph. He concluded that the adaptation to highly concentrated salt solutions was partly due to a relatively impermeable cuticle and probably also to active osmotic regulation.

Reproduction

Females begin laying eggs one to two weeks after they emerge. Ephydra cinerea has been observed to walk down stems of aquatic vegetation or emergent objects to oviposit underwater. The other Ephydrid flies oviposit on the water surface, where the eggs quickly sink to the bottom. Jones (1906) states that the eggs of E. millbrae are deposited on the floating mats of its puparia. Females deposit between 10 and 60 eggs and may require up to 20 days to complete deposition of their eggs.

Significance to Other Wetlands Taxa

These insects are an important prey item of shore birds and game ducks (Martin and Uhler 1939). Feeney and Maffeii (1991) observed Snowy Plovers and Maffeii (unpub.) observed California Gulls, Black Necked Stilts and American Avocets charging through large assemblages of brine flies catching disturbed adults as they attempted to fly away. Murie and Bruce (1935) have observed populations of the Western Sandpiper, Calidris mauri, feeding on Brine Flies near the Dumbarton bridge. Anderson (1970) found Lesser Scaups, Dunsins, Avocets, Western Sandpipers and Northern Phalaropes feeding on Ephydra cinerea in the salt ponds of southern Alameda County.

These flies are a common prey item of spiders, especially the Dictynidae and Salticidae. The tiger beetle, Cicindela senilis senilis, will catch these flies, and the adults of the Ant hicid beetle, Tanarthrus occidentalis, utilizes the carcasses of these flies as a food source.

Conservation Needs and Limiting Factors

Ephydra cinerea seems to prefer the hypersaline environs of salt ponds and has shown poor ability to adapt to the tidal pools of mid elevation tidal marshes. The larvae of this fly are also easily out competed by E. millbrae in salt marsh tidal pools.

The frequency of flooding and duration of flooding or drying periods limits the reproductive success of E. millbrae.

References


Additional Readings

Jamieson's Compsocryptus Wasp
Compsocryptus jamiesoni Nolfo
Wesley A. Maffei

Description and Systematic Position

Compsocryptus jamiesoni is a moderate sized wasp, approximately 15-25mm in length, that belongs to the family Ichneumonidae, tribe Mesostenini. Overall body ground color is rusty red-brown with the middle of the face, vertex and occiput of the head, apical third of the antennae, and the thoracic sutural markings black. The wings are light brownish-yellow with three dark brown transverse bands, the apical pair of bands merging near the posterior margin of the wing (Figure 3.23). Females have an ovipositor measuring approximately 6mm in length and the base of the third abdominal tergite black. Nolfo (1982) has indicated that this wasp is very similar to both Compsocryptus calipterus brevicornis and Compsocryptus aridus, which have been found within the confines of the San Francisco Bay Region exclusive of its salt marshes. Males of this wasp are very similar to Compsocryptus calipterus brevicornis but can readily be separated by the absence of any dark markings on the apex of the hind femur. Females are similar to Compsocryptus aridus but differ in having the body color rusty red-brown rather than brownish-yellow and the dark markings of the wings broader.

Distribution

This wasp was first collected in 1981 and subsequently described as a new taxon by Nolfo in 1982 from specimens collected at the salt marshes in Alviso, Santa Clara County, California. Additional populations have been identified from the salt marshes of the eastern San Francisco Bay as far north as San Leandro, California (Maffei, unpub.). Surveys for this wasp from other parts of the San Francisco Estuary have not been done at this time. Figure 3.24 shows the locations around the Bay Area where Compsocryptus jamiesoni have been collected, and Table 3.7 shows the collection dates.

Suitable Habitat

Compsocryptus jamiesoni have only been found on short grass or herbage in or near tidal and muted tidal marshes.

Table 3.7 Known Collection Sites For Compsocryptus jamiesoni

<table>
<thead>
<tr>
<th>Location</th>
<th>Date Specimen(s) Collected</th>
</tr>
</thead>
<tbody>
<tr>
<td>Trojan Marsh (San Leandro)</td>
<td>11 Sep 1997</td>
</tr>
<tr>
<td>Oliver Salt Ponds (Hayward)</td>
<td>23 Sep 1989</td>
</tr>
<tr>
<td>Baumberg Tract (Hayward)</td>
<td>4 Jul 1989</td>
</tr>
<tr>
<td>Shoreline Int. Ctr. (Hwyd)</td>
<td>1 Jul 1990, 2 Jul 1990</td>
</tr>
<tr>
<td>Ecology Marsh</td>
<td>24 Aug 1994</td>
</tr>
<tr>
<td>Hetch-Hetchy Marsh</td>
<td>16 Jul 1997</td>
</tr>
<tr>
<td>Alviso (Triangle Marsh)</td>
<td>3 Jun 1980</td>
</tr>
<tr>
<td>Santa Clara (Topotype)</td>
<td>2 Sep 1928</td>
</tr>
<tr>
<td>San Jose (Topotype)</td>
<td>16 Aug 1982</td>
</tr>
</tbody>
</table>

1 Information assembled from specimens contained within the California Academy of Sciences Insect Collection, University of California Berkeley Essig Museum, University of California Bohart Museum, San Jose State University Edwards Museum, San Mateo County Mosquito Abatement District Insect Collection, and private collections of Dr. J. Gordon Edwards and Wesley A. Maffei.
Biology

Little is known concerning the biology of this wasp. Other members of the tribe Mesostenini are known to be parasitic in cocoons of lepidoptera and other ichneumonids, puparia of diptera and other wasps, and the egg sacs of spiders (Townes 1962). Adults regularly utilize dew or rainwater from foliage and nectar from flowers when available and can be found from April through October. The peak flight period for C. jamiesoni is June through August (Maffei, unpub.).

Reproduction

Unknown.

Significance to Other Wetlands Taxa

Unknown.

Conservation Needs and Limiting Factors

Unknown.

References


A Note on Invertebrate Populations of the San Francisco Estuary

Wesley A. Maffei

The study of San Francisco Bay invertebrate populations and their interrelationships has usually been given low priority or altogether neglected during the planning and implementation of enhancement or restoration projects. Environmental assessments of habitat quality and health have frequently forgotten about the terrestrial or semi-aquatic invertebrates that are usually very sensitive to environmental changes. Arthropods, especially insects, are sensitive indicators of environmental disturbance or change (Lenhard and Witter 1977, Hellawell 1978, Hawkes 1979).

A survey of the literature shows that few studies have been done on the biology and ecology of the terrestrial and semi-aquatic invertebrates within the San Francisco Estuary. What is known about these organisms generally comes from studies of invertebrate populations well outside of this geographic area. For many of the common species, this is probably adequate. Unfortunately, little information exists about what species are found within the different wetland habitats, and less still is known about the impacts of wetlands projects on the existing invertebrate populations. Those species that are pests (i.e., mosquitoes) are fairly well known, while taxa such as Jamieson’s compsocryptus wasp or the western tanarthrus beetle, which were described as new to science within the last twenty years, have poorly known or completely unknown biologies. This lack of basic information, specifically what species exist where, coupled with an understanding of their basic biologies, warrants careful consideration and research. The fact that unknown populations of organisms, or unique, sensitive, or threatened and endangered taxa do exist within or near the tidal reaches of the Bay suggests that more care should be taken when planning enhancement or restoration projects. The relationship of some invertebrate species to the success of other organisms (i.e. plants or invertebrates) needs to be clarified.

Some invertebrates are known to play a significant role in the life cycles of other organisms. Functioning as pollinators, herbivores, scavengers, predators, and prey, terrestrial and semi-aquatic invertebrates are a significant component of any habitat or community. It became apparent through the course of the Goals Project that the experts on many of the key species of fish and wildlife were not always clear about the roles played by invertebrates with respect to the survival of their target species or communities. This prompted the construction of some graphic displays, in this case food webs, by which to illustrate what little is known about the roles performed by the largest and most...
easily overlooked group of organisms in our estuary, the invertebrates.

Food webs are frequently used to illustrate the complex relationships between organisms within a given area or habitat. Unfortunately, they cannot hope to tell the entire story. Factors such as the seasonality of the organisms, length of time and time of year the studies were performed, the limited number of organisms that can be included in the web, and the complexity of the habitat or ecosystem being studied tend to result in webs that overgeneralize what actually exists or has been observed.

The following sample invertebrate webs are undoubtedly incomplete. They have been assembled from many hours of field observation in the southern portion of the San Francisco Estuary, and from an exhaustive search of the literature. The most notable feature of all of these webs is the delicate relationships that exist between all of the organisms involved. The potential reduction or loss of one member of the web clearly illustrates how its associates could be impacted. It should be noted that not all of the organisms that have been found or studied are represented. The organisms included in these webs are those routinely found in association with the plant or plants that are indicated by the boxes with the thickened black borders. Figures 3.25, 3.26, and 3.27 are examples of partial webs developed to illustrate the relationship of some of the organisms associated with the plant species alkali heath (Frankenia salina), common pickleweed (Salicornia virginica), and willow (Salix lasiolepis), respectively. Figures 3.28 and 3.29 are examples of partial webs that illustrate the relationships of organisms within mid-marsh pans and crystallizer pond habitats. The web for the organisms associated with old crystallizer ponds was included to illustrate that even in this inhospitable habitat, webs of life can and do exist. When known vertebrate relationships for most of the webs have been included. Table 3.8 is a brief summary of the descriptions and biologies of some of the invertebrates from the alkali heath web. Table 3.9 is a listing of the scientific names associated with a major common name category. It is hoped that these tables might help the reader better visualize the nature of the relationships shown for the different organisms included in the webs.

Figure 3.25 A Partial Web of the Organisms Associated With Alkali Heath (Frankenia salina) in San Francisco Tidal Marshes
<table>
<thead>
<tr>
<th>Bees and Wasps</th>
<th>Butterflies and Moths</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Bombus vosnesenskii</strong> - A moderate to large sized bumblebee that is mostly black with a small amount of yellow on the thorax and posterior portion of the abdomen. Adults tend to nest in abandoned rodent burrows along levees and adjacent upland habitat.</td>
<td><strong>Perizoma custodiata</strong> - A moderate sized moth belonging to the family of moths known as measuring worms, or <em>Geometridae</em>. Adults are tan gray or brown in color and have dark geometric bands across the forewings. Larvae are about one inch long, light green in color and feed on the leaves of Frankenia. Adults are present throughout the year, with peak populations occurring from spring through fall.</td>
</tr>
<tr>
<td><strong>Anthophora spp.</strong> - A moderate sized native bee, belonging to the family <em>Anthophoridae</em>, that is light bee, belonging to the family <em>Anthophoridae</em>, which is closely related to ants but do have the appearance of them in opposite directions. Biology unknown.</td>
<td><strong>Bephidrium exilis</strong> - A very small brown and blue butterfly that is a frequent visitor of <em>Frankenia</em>.</td>
</tr>
<tr>
<td><strong>Osmia spp.</strong> - A bluish-black bee with smoky colored wings, that belongs to the family bees known as leaf cutting bees, or <em>Mega chilidae</em>. This genus of bees is commonly known as mason bees because of their habit of building small earthen cells on or under stones, in abandoned burrows, in holes in boards, twigs and logs, and in plant galls.</td>
<td><strong>Synanthedon bibionipennis</strong> - A small moth, belonging to the family of moths known as clear wing moths, or <em>Sesiidae</em>. Adults emerge in late May to early June and can be found through late September. These insects are frequently associated with <em>Frankenia</em> sp. It is believed that the larvae may feed on the roots and the bases of <em>Frankenia</em> sp. plants. Currently, this is the only clear wing moth known to inhabit the levees of mid to upper tidal marshes within the San Francisco Estuary.</td>
</tr>
<tr>
<td><strong>Ammophila spp.</strong> - A long, slender solitary digger wasp belonging to the family <em>Sphecidae</em>. These wasps build simple, vertical burrows, that are provisioned with moth caterpillars. Nests usually occur in fine, silty or sandy soil with minimal vegetation.</td>
<td><strong>Gymnosoma fuliginosum</strong> - A small, bright orange and black fly that is parasitic on the green stink bug, <em>Chlorochroa sayi</em>.</td>
</tr>
<tr>
<td><strong>Aporinellus completus</strong> and <strong>Aporinellus taeniatus</strong> - Small black spider wasps (family <em>Pompilidae</em>) that provision their nests with jumping spiders (<em>genus</em> <em>Phidippus</em>).</td>
<td><strong>Physcoephala texana</strong> - A bright red and black fly, about one-half an inch long, that parasitizes the bumblebee <em>Bombus vosnesenskii</em>.</td>
</tr>
<tr>
<td><strong>Chrysura pacifica</strong> - A small iridescent bluish-purple to bluish-green wasp, measuring up to 10mm in length. This wasp parasitizes the leaf cutting bee <em>Osmia</em>.</td>
<td><strong>Aplomya theclarum</strong> - A very tiny black fly, with a bright silver face, that parasitizes the larvae of the pygmy blue butterfly.</td>
</tr>
<tr>
<td><strong>Ramopse edwardsi</strong> - A moderate sized brilliant light green wasp, measuring about 10-12mm, that parasitizes the sand wasp <em>Bembix americana</em>.</td>
<td><strong>Acróstica dichroa</strong> - A small, bright green and red fly with one brown spot at the tips of the wings. This fly is frequently seen walking up and down the stems of <em>Frankenia</em> holding it’s wings outstretched and rotating them in opposite directions. Biology unknown.</td>
</tr>
<tr>
<td><strong>Bembix americana</strong> - A large sand wasp that is bluish gray in color with pale white markings on the abdomen. The eyes are usually bright yellow to yellowish-green in color. Adult wasps provision their ground nests with adult flies.</td>
<td><strong>Exoprosopa spp.</strong> and <strong>Villa spp.</strong> - Small to moderate sized, fuzzy looking flies that are commonly known as bee flies. <em>Villa</em> spp. is light brown in color with clear wings and <em>Exoprosopa</em> spp. is brown and white banded with brightly patterned brown and clear wings. Both species of flies are parasites of immature sand wasps of the genus <em>Bembix</em>.</td>
</tr>
<tr>
<td><strong>Dasymutilla aureola</strong> - A golden yellow to bright orange insect known as a velvet ant. These insects are not closely related to ants but do have the appearance of looking like an ant. Velvet ants provision their burrows with ground nesting bees and wasps.</td>
<td><strong>Eristalis aeneus</strong> - A moderate sized, shiny olive green fly that is commonly known as a hoverfly or flower fly. The larvae of this fly are known as ratted maggots and are found in somewhat saline or brackish pools of tidal marshes. Adult flies are an important food source for <em>Bembix</em> sand wasps and spiders.</td>
</tr>
<tr>
<td><strong>Fomicilla sp.</strong> - A very small, brown to tan colored beetle, known as an Ant-like flower beetle. These beetles are known to feed on decaying vegetation and can sometimes be very common at the bases of <em>Frankenia</em> sp.</td>
<td><strong>Lejops curvipes</strong> - A moderate sized flower fly, measuring about 10-15mm, that is bright reddish-orange, with a central black stripe on the abdomen and mostly black legs.</td>
</tr>
<tr>
<td><strong>Stink Bugs</strong></td>
<td><strong>Spiders</strong></td>
</tr>
<tr>
<td><strong>Chlorochroa sayi</strong> - A moderately sized (one-half inch) stink bug that is pale to deep green in color. This insect is known for releasing a foul smelling odor when disturbed or threatened.</td>
<td><strong>Phidippus spp.</strong> - Two species are common within our marshes. One is solid black with the top of the abdomen bright red and can reach a size up to one-third of an inch. The other is dark gray with variegated white lines and reaches a size of about a quarter of an inch.</td>
</tr>
</tbody>
</table>
The need for terrestrial invertebrate surveys has become more apparent with the increase in wetland enhancement and restoration projects. The conversion of one habitat type to another "more valuable" or "more improved" habitat type can and usually does have significant impacts on the often-unnoticed invertebrate populations that exist within them. In some cases these impacts can be positive, while in other instances the opposite is true. Table 3.10 lists by site and date(s) those known terrestrial and semi-aquatic invertebrate surveys or species studies.

Figure 3.26 A Partial Web of the Organisms Associated With Common Pickleweed (Salicornia virginica) in San Francisco Bay Lower High Tidal Marshes

References


Figure 3.27 A Partial Web of the Organisms Associated With Willow (Salix lasiolepis)

Figure 3.28 Partial Web of Organisms Associated With Mid-Tidal Marsh Pans
Figure 3.29 A Partial Web of the Organisms in the Baumberg and Oliver Brothers Salt Crystallizer Ponds, Hayward, California
### Table 3.9 Food Web Taxa by Major Common Name Category

<table>
<thead>
<tr>
<th>Category</th>
<th>Taxa</th>
</tr>
</thead>
</table>
| **Butterflies and Moths** | Agrius sp.  
|                   | Amara spp.  
|                   | Bembidion spp.  
|                   | Cicindela senilis senilis  
|                   | Cryptochalpus castaneus  
|                   | Enocherus eximius  
|                   | Enochrus diffusus  
|                   | Erymephala morosa  
|                   | Formicilla spp.  
|                   | Ochthebius rectus  
|                   | Pachybrachus melanostictus  
|                   | Psyllobora vigintimaculata  
|                   | Sympaphaeta guexi  
|                   | Tanarthrus occidentalis  
|                   | Tecnophilus croceicollis  
|                   | Xylotrechus insignis  |
| **Insects**       | Ammophila spp.  
|                   | Anthophora spp.  
|                   | Aporinellus completus  
|                   | Aporinellus taeniatus  
|                   | Bembix americana comata  
|                   | Bombus vosnesenskii  
|                   | Bombus occidentalis  
|                   | Ceratophila californica  
|                   | Chrysura pacifica  
|                   | Coeloxyx spp.  
|                   | Dasymutilla aureola  
|                   | Megaschile spp.  
|                   | Melissodes spp.  
|                   | Osmia spp.  
|                   | Parnopes edwardsi  
|                   | Pontania californica  |
| **Beetles**       | Agrilus sp.  
|                   | Amara spp.  
|                   | Bembidion spp.  
|                   | Cicindela senilis senilis  
|                   | Cryptochalpus castaneus  
|                   | Enocherus eximius  
|                   | Enochrus diffusus  
|                   | Erymephala morosa  
|                   | Formicilla spp.  
|                   | Ochthebius rectus  
|                   | Pachybrachus melanostictus  
|                   | Psyllobora vigintimaculata  
|                   | Sympaphaeta guexi  
|                   | Tanarthrus occidentalis  
|                   | Tecnophilus croceicollis  
|                   | Xylotrechus insignis  |
| **Hoppers and Psyllids** | Aphalara sp.  
|                   | Aphids  
|                   | Cixius praecox  
|                   | Oliarius dondonius  
|                   | Psyllids  
|                   | Scale Insects  |
| **Fies**          | Acroceras steyskali  
|                   | Acroceras fasciata  
|                   | Acrosticta dichroa  
|                   | Aedes dorsalis  
|                   | Aedes squamiger  
|                   | Aedes washinoi  
|                   | Aphalara theclairum  
|                   | Argyrocalonia californica  
|                   | Ephydra millbrae  
|                   | Eratophilus aeneus  
|                   | Exoprosopa spp.  
|                   | Gymnocarcinella ricinorum  
|                   | Gymnosoma fuliginosum  
|                   | Helophilus latifrons  
|                   | Lejope curvipes  
|                   | Lipochaeta slossonae  
|                   | Lispe approximata  
|                   | Pegomya spp.  
|                   | Peleterea sp.  
|                   | Physcocephala texana  
|                   | Ravina sp.  
|                   | Villa spp.  |
| **Bugs**          | Chlorochroa sayi  
|                   | Trichocorixa reticulata  |
| **Lacewings**     | Chrysoperla plorabunda  
|                   | Symphorophorus bifasciatus  |
| **Spiders**       | Dictynid Spider  
|                   | Lycosa spp.  
|                   | Pardosa sp.  
|                   | Phidippus spp.  |
| **Birds**         | Great Blue Heron  
|                   | Great Egret  
|                   | Salt Marsh Yellow Throat  
|                   | Snowy Egret  
|                   | Snowy Plover  
|                   | Song Sparrows  
|                   | Western Gull  |
| **Mammals**       | Feral Cat  
|                   | Red Fox  
|                   | Reithrodontomys raviventris  |
| **Plants**        | Cuscuta salina  
|                   | Dunaliella sp.  
|                   | Enteromorpha sp.  
|                   | Frankenia salina (= grandifolia)  
|                   | Salicornia virginica  
|                   | Salix lasiolepis  |
| **Fungi**         | Mildew type fungus  |


Table 3.10 Known Terrestrial or Semi-aquatic Invertebrate Surveys or Studies of Selected Invertebrate Taxa

<table>
<thead>
<tr>
<th>Locale</th>
<th>Date of Study</th>
<th>Reference(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Outer Coyote Creek Tributary</td>
<td>1980</td>
<td>J. Anderson, et. al. (1980)</td>
</tr>
<tr>
<td>Warm Springs Seasonal Wetlands, Fremont</td>
<td>1993</td>
<td>T. Caires, et. al. (1993)</td>
</tr>
<tr>
<td></td>
<td>1995</td>
<td>W. Maffei, (unpub. field notes)</td>
</tr>
<tr>
<td>Dumbarton Point Marsh, Fremont</td>
<td>1968</td>
<td>R.S. Lane (1969)</td>
</tr>
<tr>
<td></td>
<td>1996-1996</td>
<td>W. Maffei (unpub. field notes)</td>
</tr>
<tr>
<td></td>
<td>1997-1997</td>
<td>W. Maffei (unpub. field notes)</td>
</tr>
<tr>
<td></td>
<td>1979</td>
<td>V. Resh and S. Balling (1983)</td>
</tr>
<tr>
<td>Cullinan Ranch, USGS Survey</td>
<td>1995(?</td>
<td>M. Paquin (unpub.)</td>
</tr>
<tr>
<td></td>
<td>1979</td>
<td>V. Resh and S. Balling (1983)</td>
</tr>
<tr>
<td>Suisun Marsh</td>
<td>1972-1973</td>
<td>A.M. Shapiro (1975a, b, 1974)</td>
</tr>
</tbody>
</table>

1 The studies shown pertain primarily to insects and arachnids and do not include the numerous biological studies on mosquitoes.

Amphibians and Reptiles

California Tiger Salamander
Ambystoma californiense
Mark R. Jennings

General Information

The California tiger salamander (Family: Ambystomatidae) is a large (75-125 mm SVL) terrestrial salamander with several white or yellow spots or bars on a jet-black field (Stebbins 1985). Although often referred to as a subspecies of the more widespread tiger salamander (A. tigrinum; e.g., see Frost 1985, Stebbins 1985, Zeiner et al. 1988), the California tiger salamander is currently recognized as a full species (Jones 1989, Shaffer et al. 1993, Barry and Shaffer 1994). In 1992, the California tiger salamander was petitioned for listing as an endangered species (Long 1992) based on concerns about population declines due to the extensive loss of habitat, introductions of non-native aquatic predators, and interbreeding with introduced salamanders originally brought in as live fish bait (Long 1992; see also Jennings and Hayes 1994). The U.S. Fish and Wildlife Service ruled that the petition was warranted but precluded by pending listing actions on higher priority species (Sorenson 1994).

Reproduction

Most adults probably reach sexual maturity in two years, but some individuals may take longer during periods of unfavorable conditions such as annual droughts (Shaffer et al. 1993). Adults migrate during the night from subterranean refuge sites (small mammal burrows) to breeding ponds after the onset of relatively warm winter rains (late November-early March) where courtship and egg deposition occurs (Twitty 1941, Barry and Shaffer 1994, Loredo and Van Vuren 1996). Males may precede females to breeding ponds (Shaffer et al. 1993, Loredo and Van Vuren 1996) and distances travelled by adults from refuge sites to breeding sites may be up to 1.6 km (Austin and Shaffer 1992). Females lay single or small groups of 2-4 eggs (8.5-12 mm in diameter) on detritus, submerged vegetation, or on the benthos of relatively shallow rain pools (Storer 1925). The number of eggs laid per female is unknown. During periods of low rainfall, California tiger salamanders may not reproduce (Jennings and Hayes 1994). After reproducing, adults return to subterranean refuge sites, some to the same small mammal burrows they emerged from earlier in the year (Shaffer et al. 1993).

Growth and Development

Eggs hatch after 2-4 weeks (Storer 1925, Twitty 1941), and gilled aquatic larvae take a minimum of at least 10 weeks to successfully reach metamorphosis (Anderson 1968, Feaver 1971). Larvae lack legs upon hatching at 10.5 mm total length, but quickly grow four legs within 1-2 weeks. Larvae generally are about 75 mm in total length and weigh about 10 grams at metamorphosis into juveniles, although they may remain in water (for up to six months) and grow to much larger sizes with a better chance of survival after metamorphosis (Jennings and Hayes 1994). Overwintering of larvae, which is common with several species of Ambystoma (see Stebbins 1985), is unusual with A. californiense because of the temporary nature of its natural breeding habitat (Shaffer et al. 1993). All records of overwintering or aseasonal metamorphosis are based on salamanders in artificially-created habitats (Jennings, unpubl. data).

Upon metamorphosis (usually early May through July), juveniles disperse in mass at night away from desiccating breeding ponds into terrestrial habitats (Zeiner et al. 1988, Loredo and Van Vuren 1996, Loredo et al. 1996). Juveniles have also been known to disperse during periods of unfavorable conditions (e.g., August) re-
resulting in mass mortality (Holland et al. 1990). Both adults and juveniles seek refuge in small mammal burrows (especially those of California ground squirrels (Spermophilus beecheyi) and Botta’s pocket gophers (Thomomys bottae) (Barry and Shaffer 1994, Loredo et al. 1996)) and spend most of the year underground until the onset of cooler and wetter surface conditions (Jennings and Hayes 1994). Juveniles probably feed on invertebrates in subterranean mammal burrows and grow throughout the year. During the winter months however, both juveniles and adults emerge from burrows and forage at night on the surface for extended periods of time, although adults appear to do all their foraging after completing their reproductive activities (Shaffer et al. 1993).

California tiger salamanders are relatively long-lived animals, reaching ages of 20 years or more in captivity (Jennings, unpubl. data). The average life span of adults in the wild is unknown.

**Food and Feeding**

Larval California tiger salamanders subsist on aquatic invertebrates (Oligochaetes, Cladocera, Conchostraca, Ostracoda, Anostraca, Anostraca, Chironomids, etc.), as well as the larvae of western spadefoots (Scaphiopus hammondii), California toads (Bufo boreas halophilus), and Pacific treefrogs (Hyla regilla), if the latter are present in breeding ponds (Anderson 1968; Feaver 1971; Jennings, unpubl. data). Larval salamanders are also highly cannibalistic (Jennings, unpubl. data). Good numbers of food organisms in breeding ponds appear to be important for the survival and rapid growth of salamander larvae to metamorphosis (Jennings, unpubl. data).

Juvenile and adult salamanders subsist on terrestrial invertebrates (Oligochaetes, Isopoda, Orthoptera, Coleoptera, Diptera, Araneida, Gastropoda, etc.; Stebbins 1972; Morey and Guinn 1992; Jennings, unpubl. data). There is no evidence of adult salamanders feeding in aquatic environments (Jennings, unpubl. data).

**Distribution**

The historical distribution of the California tiger salamander ranged from the vicinity of Petaluma, Sonoma County and Dunnigan, Colusa-Yolo County line (Storer 1925) with an isolated outpost north of the Sutter Buttes at Gray Lodge, Butte County (Hays and Cliff 1982) in Central Valley, south to vernal pools in northwest Tulare County, and in the South Coast Range south to ponds and vernal pools between Bullerton and Lompoc in the Santa Ynez drainage, Santa Barbara County (Jennings and Hays 1994). The known elevational range extends from 3 m-1054 m (Shaffer et al. 1993). The species has disappeared from about 55% of its historic range (Jennings and Hays 1994).

In the Bay Area, California tiger salamanders have disappeared from almost all of the lower elevation areas (<50 m), save one small site on the San Francisco Wildlife Refuge near Fremont, Alameda County (Jennings, unpubl. data). There are scattered populations currently inhabiting vernal pool and stockpond habitats in hills surrounding the South Bay (Jennings, unpubl. data), to the north of Coyote Hills in Suisun, and in northern Contra Costa. A group of relict populations is also present in the North Bay region in vernal pool habitats near Petaluma (Shaffer et al. 1993) (Figure 4.1).

**Current Status and Factors Influencing Population Numbers**

Based on the data presented in Shaffer et al. (1993) and Jennings and Hays (1994), California tiger salamanders appear to have disappeared from approximately 58% and 55% (respectively), of their historic range in the state (Sorensen 1994). This salamander is most affected by land use patterns and other anthropogenic events which fragment habitat and create barriers between breeding and refuge sites (Jennings and Hays 1994). Some of the more important factors negatively influencing salamander populations include: conversion and isolation of vernal pool habitats (and surrounding oak woodland and grasslands) to agriculture and urbanization (Barry and Shaffer 1994); lowering of the groundwater table by...
overdraft (Jennings and Hayes 1994); mortality of juvenile and adult salamanders by vehicles on roads (Twitty 1941); the introduction of non-native predators such as mosquito fish (Gambusa affinis), bullfrogs (Rana catesbeiana) and crayfish (specifically Procambarus clarkii) into breeding habitats (Shaffer et al. 1993); the widespread poisoning of Californian ground squirrels and other burrowing rodents (Loredo et al. 1996); and interbreeding with introduced salamanders originally brought in as live fish bait (Shaffer et al. 1993). Juvenile and adult salamanders have also been found in a number of human-created habitats such as septic tank lines, pipes, wells, wet basements, and permanent irrigation ponds (Jennings and Hayes 1994). Such habitats may not be suitable for the long-term survival or successful reproduction of local salamander populations.

### Trophic Levels

Larval and post-metamorphic life stages are secondary consumers.

### Proximal Species

**Predators:** Common (=San Francisco) garter snake, Coast garter snake, Central Coast garter snake, California red-legged frog, bullfrog, shrews, striped skunk, opossum, herons, and egrets. Ducks and predacious aquatic insects prey on larvae only.

**Prey:** Oligochaetes, snails, and terrestrial insects. Zooplankton and aquatic insects are prey for larvae.

**Habitat:** California ground squirrel and valley pocket gopher (maintain tiger salamander’s terrestrial habitats)

### Good Habitat

The best habitats for California tiger salamanders are vernal pool complexes with colonies of California ground squirrels or Botta’s pocket gophers within the complex or nearby (Shaffer et al. 1993). Such habitats are normally associated with grasslands or oak woodlands (Barry and Shaffer 1994). Additionally, there needs to be abundant invertebrate resources and other native amphibian larvae in the vernal pools used by breeding salamanders.

### References


Reproduction

California toads breed between January and July with higher altitude populations delaying breeding until June or July (Storer 1925). At lower elevations, toads are active all year, but at higher elevations adults emerge from hibernation sites immediately before reproducing (Stebbins 1951). Males and females congregate at night around aquatic breeding sites such as stockponds, temporary roadside pools, cement water reservoirs, and the margins of flowing streams where males call, amplexus occurs, and females lay up to 16,500 eggs in two long strings wrapped around vegetation at water depths <150 mm (Storer 1925, Livezey and Wright 1947). Eggs strings are about 5 mm in diameter and the inclusive eggs 1.7 mm in diameter (Storer 1925). After reproducing, adults generally disperse back into the surrounding terrestrial habitats such as meadows and woodlands where they use almost any sort of cover (e.g., trees, low vegetation, beds of leaves, small mammal burrows, rocks, pieces of concrete, downed logs, etc.) that provides a slight amount of moisture and protection from the drying effects of the sun and wind (Storer 1925).

Growth and Development

Eggs hatch within four days to a few weeks (depending on the prevailing water temperature; Storer 1914) and the resulting larvae normally comprise schools composed of one or more clutches (Jennings, unpubl. data). Larvae grow rapidly and usually metamorphose in 2-3 months (from April-August) at 19-52 mm (Storer 1925, Wright and Wright 1949). Recent metamorphs are 12-15 mm in total length (Wright and Wright 1949) and are often observed around the immediate margin of the breeding pond under any cover that protects them from the wind and sun (Storer 1925). The number of newly metamorphosed toads at such breeding sites can number in the thousands (Storer 1925). Young toads grow rapidly and probably reach sexual maturity in two years at lower elevations and somewhat longer at mid-higher elevations (Stebbins 1951). Both juveniles and adults are largely crepuscular, although an occasional individual will be observed during the day in wet or overcast conditions (Storer 1925).

Adults may live 10 years or more in captivity (Bowler 1977) but the longevity of toads in the wild is unknown.

Food and Feeding

Larvae are thought to be algal grazers (Stebbins 1951), but the foraging ecology of larval California toads is unknown. Juveniles and adults feed on a wide variety of terrestrial and flying invertebrates including: Oligochaetes, Isopoda, Diplopoda, Orthoptera, Plecoptera,
Chapter 4 — Amphibians and Reptiles

Amphibians & Reptiles

Dermaptera, Hemiptera, Homoptera, Coleoptera, Trichoptera, Lepidoptera, Diptera, Arachnids, and Gastropoda (Storer 1914; Eckert 1934; Stebbins 1951, 1972; Moyer and Guinn 1992; Jennings, unpubl. data). Cannibalism can also occur (Stebbins 1972).

Distribution

California toads are found over most of California (except for the northernmost counties where they are replaced by boreal toads, and almost all of the Mojave and Colorado deserts where they are replaced by other toad species) from sea level to over 3,050 meters in the Sierra Nevada (Stebbins 1972). This toad is replaced at higher elevations in the central and southern Sierra Nevada by the Yosemite toad (Bufo canorus) [Karlstrom 1962]. California toads are widespread in the Bay Area (Stebbins 1959) (Figure 4.2) and are still relatively common in stockponds and other aquatic habitats in the surrounding foothills (Jennings, unpubl. data).

Current Status and Factors Influencing Population Numbers

California toads are still present throughout most of their native range in California, although they are now rare in many urban areas where they were formerly common (such as in the Los Angeles Basin; Jennings unpubl. data). The possible reasons for the localized declines are insecticides used in eradicating introduced Mediterranean fruit flies (Ceratitis capitata), changing land use patterns by agriculture and urban communities which now leave less sites containing permanent water and areas of dense vegetation (such as tule-lined canals, low ground cover, etc.), and habitat fragmentation by roads and dense regions of urbanization (Jennings, unpubl. data). In the Bay Area, California toads are still relatively abundant in natural and moderately-altered habitats (Stebbins 1959; Jennings, unpubl. data). The factors most associated with toad survival include local breeding ponds that last for at least two months, and sufficient cover (vegetative and small mammal burrows) that provide places for toads to feed and grow, as well as escape predators and desiccating conditions.

Trophic Levels

Larval stages are primary consumers and post-metamorphic life stages are secondary consumers.

Proximal Species

Predators: Common (=San Francisco) garter snake, coast garter snake, central coast garter snake, bullfrog, introduced predatory fishes, herons, egrets, raccoon, striped skunk, and opossum. Predacious aquatic insects prey on larvae.

Prey: Aquatic insects, Oligochaetes, Gastropoda, Isopoda, and terrestrial insects.

Habitat: Willows, cattails, tules, sedges, blackberries, riparian vegetation.

Good Habitat

California toads inhabit grasslands, woodlands, meadows, gardens, golf courses, and parks—in fact, anywhere where a permanent source of moisture is present and breeding ponds of at least two months duration are available (Storer 1925). The largest populations of toads seem to be found around stockponds or reservoirs that have an abundance of invertebrate prey, many small mammal burrows and objects (or vegetation) that are available for cover, and a lack of introduced predators (fishes and bullfrogs [Rana catesbeiana]) in aquatic habitats.

References


Pacific Treefrog
Hyla regilla
Mark R. Jennings

General Information

The Pacific treefrog (Family: Hylidae) is a small (19-50 mm SL) frog with toe pads and a black eye stripe (Stebbins 1985). The dorsal coloration is highly variable—green, tan, reddish, gray, brown, or black—sometimes with dark dorsal spots (Wright and Wright 1949, Resnick and Jameson 1963, Stebbins 1972); however green or shades of brown are the usual colors observed (Nussbaum et al. 1983). For a time, these treefrogs were lumped with chorus frogs of the genus Pseudacris (see Hedges 1986). However, recent work has shown that Pacific treefrogs are not chorus frogs, hence the revision to the old genus Hyla (Crocroft 1994).

This frog has the most notable voice of the frog world as its call has been used as a natural background sound in innumerable movies produced by Hollywood (Myers 1951).

Reproduction

Pacific treefrogs can become sexually mature in one year, but most become sexually mature in two years (Jameson 1956). At lower elevations, treefrogs are active all year, but at higher elevations adults emerge from hibernation sites immediately before reproducing (Stebbins 1951). From late November to July (beginning with the first warm rainfall), males congregate at night around any suitable shallow pond of water (or at the shallow edges of deep water ponds or reservoirs) and chorus to attract receptive females (Storer 1925, Brattstrom and Warren 1955, Schaub and Larsen 1978, Nussbaum et al. 1983), and also call to space themselves from one another (Snyder and Jameson 1965, Allan 1973). Groups of two or three males tend to call in sequence during these choruses and the sequence is consistently started by one frog known as the bout leader (Whitney and Krebs 1975).
The choruses may continue into daylight hours (Jameson 1957) and can be deafening if hundreds or thousands of calling males are involved (Stebbins 1959; Jennings, unpubl. data). Females attracted to these calls usually select the bout leader to mate with (Whitney and Krebs 1975). Upon amplexus, females lay approximately 20-25 packets containing 9-70 (usually 22-25) eggs on submerged aquatic vegetation or on the bottom of shallow pools (Smith 1940, Livezey and Wright 1947), generally at depths >100 mm (Storer 1925). Egg masses are normally laid close together, one against another, or separated by <25 mm (Stebbins 1951). The eggs are about 1.3 mm in diameter and females may lay from 500 to 1,250 total eggs (Storer 1925, Smith 1940). After egg deposition, males and females remain the vicinity of the breeding pond for up to one and three months respectively, and then return to surrounding terrestrial habitats (Jameson 1957, Nussbaum et al. 1983). Females may also breed up to three times during the year (Perrill and Daniel 1983).

**Growth and Development**

Eggs hatch in four days to two weeks, depending on the prevailing water temperatures and the resulting larvae (6.0-7.5 mm total length) grow rapidly (Storer 1925). Larvae are also known to aggregate into large groups of several hundred individuals (Brattstrom and Warren 1955). Metamorphosis is generally within two months at anytime between February-late August (Storer 1925; Jennings, unpubl. data), at total lengths between 45 and 55 mm (Storer 1925, Wright and Wright 1949). Post-metamorphs are about 12-15 mm and grow rapidly within the first two months often doubling their size (Jameson 1956). For the first few months, post-metamorphs remain in the immediate vicinity of the breeding pond utilizing almost any cover present (rocks, vegetation, leaves, etc.) for protection from the drying effects of the sun and wind (Jameson 1956). After several months, juveniles disperse out into surrounding terrestrial habitats and seek places that contain moisture and are protected by the elements. Such places include small mammal burrows, rock fissures, tree cavities, dense vegetation, piles of debris, buildings, artificial drains, etc., that may be 0.8 km or more from the nearest standing water (Storer 1925, Jameson 1957).

Adults may live four years or more in captivity (Jennings, unpubl. data). Longevity in the wild is apparently somewhat over three years (Jameson 1956).

**Food and Feeding**

Larvae are thought to be algal grazers (Storer 1925), but the foraging ecology of larval Pacific treefrogs is unknown. Juveniles and adults feed on a wide variety of terrestrial and flying invertebrates including: Oligochaetes, Oniscidea, Orthoptera, Hemiptera, Homoptera, Coleoptera, Lepidoptera, Diptera, Hymenoptera, Arachnids, and Gastropoda (Needham 1924; Stebbins 1951; Brattstrom and Warren 1955; Nussbaum et al. 1983; Morey and Guinn 1992; Jennings, unpubl. data).

**Distribution**

Pacific treefrogs are found over most of California (except drier parts of the Mojave and Colorado deserts) from sea level to around 3,670 m in the Sierra Nevada (Stebbins 1972, 1985). In the Bay Area they are very abundant (Stebbins 1959; Jennings, unpubl. data), and found throughout the region (**Figure 4.3**).

**Current Status and Factors Influencing Population Numbers**

Pacific treefrogs have always been abundant throughout most of their native range (e.g., see Storer 1925, Stebbins 1951, Nussbaum et al. 1983), and they still remain so even in the Sierra Nevada (Jennings 1996, contra Drost and Fellers 1996). Treefrogs are especially common in the Bay Area (Stebbins 1959) because of their ability to utilize habitats created by humans—especially in urban areas. Although populations are negatively influenced by the premature drying of breeding ponds and continued loss of many individuals through predation,
treefrogs are able to successfully reproduce in numbers to overcome these setbacks (Jameson 1956, 1957).

**Trophic Levels**

Larval stages are primary consumers and post-metamorphic life stages are secondary consumers.

**Proximal Species**

**Habitat:** Willows, cattails, tules, and sedges.

**Predators:** Common (=San Francisco) garter snake, Coast garter snake, Central Coast garter snake, California red-legged frog, bullfrog, introduced predatory fishes, herons and egrets, raccoon, striped skunk, and opossum. California tiger salamander and various predacious aquatic insects prey on larvae only.

**Prey:** Aquatic and terrestrial insects.

**Good Habitat**

Pacific treefrogs can essentially inhabit almost any place that contains sufficient moisture and protection from the wind and sun, and has suitable nearby breeding sites. They can reproduce in temporary aquatic environments as small as a jar of water as long as the water remains present for two months or more (Jennings, unpubl. data) and the water temperature is below 35-38 °C (Schechtman and Olson 1941). The largest populations seem to be present in complexes of shallow ponds (lacking fishes and other aquatic predators) surrounded by growths of tules (Scirpus sp.) and other aquatic vegetation (Jameson 1956, 1957), although Pacific treefrogs also seem to do well in golf courses, city parks, and other places that have permanent aquatic habitats and places with riparian vegetation (Jennings, unpubl. data).

**References**


California Red-Legged Frog
Rana aurora draytonii
Mark R. Jennings

General Information

The California red-legged frog (Family: Ranidae) is a large (85-138 mm SUL) brown to reddish brown frog with prominent dorsolateral folds and diffuse moderately-sized dark brown to black spots that sometimes have light centers (Storer 1925, Jennings and Hayes 1994a). The species is the largest native frog in the state and there are data to support elevation as a separate species from the northern red-legged frog (R. a. aurora) (see Hayes and Miyamoto 1984, Green 1985); however, there is also large zone of intergradation along the Pacific slope of the North Coast Range (Hays and Kremples 1986). In 1993, the California red-legged frog was petitioned for listing as an endangered species by the U.S. Fish and Wildlife Service (Sorensen 1993) based on a significant range reduction and continued threats to surviving populations (Miller 1994). The frog was subsequently listed as Threatened by the U.S. Fish and Wildlife Service (Miller et al. 1996).

Reproduction

Adults generally reach sexual maturity in their second year for males and third year for females (Jennings and Hayes 1985), although sexual maturity may be reached earlier during years of abundant food resources (Jennings, unpubl. data). During extended periods of drought, frogs may take 3-4 years to reach maturity (Jennings and Hayes 1994a). Reproduction generally occurs at night in permanent ponds or the slack water pools of streams during the winter and early spring (late November through April) after the onset of warm rains (Storer 1925, Hays and Jennings 1988, Jennings and Hays 1994a). California red-legged frogs can only successfully reproduce in aquatic environments with water temperatures \(<26^\circ C\) and salinities \(<4.5\%\) as developing embryos cannot tolerate conditions higher than this (Jennings, unpubl. data). Larvae can tolerate somewhat higher water temperatures and salinities (Jennings, unpubl. data). Males generally appear at breeding sites from 2-4 weeks before females (Storer 1925). At breeding sites, males typically call in small, mobile groups of 3-7 individuals that attract females (Jennings and Hayes 1994a). Females amplex with males and attach egg masses containing approximately 2,000-6,000 moderate-sized (2.0-2.8 mm diameter) eggs to an emergent vegetation brace at depths usually from 75-100 mm (Storer 1925). Egg masses are normally laid at the surface of the water (Lively and Wright 1947). California red-legged frogs are explosive breeders, usually depositing their egg masses within 3-4 week period after large rainfall events (Hays and Miyamoto 1984). After reproduction, males usually remain at the breeding sites for several weeks before removing to foraging habitats, while females immediately remove to these foraging habitats (Jennings, unpubl. data). There is no evidence of double clutching with this species (Jennings, unpubl. data).

Growth and Development

Eggs hatch after 6-14 days (depending on the prevailing water temperature), and the resulting larvae (8.8-10.3 mm total length) require 3.5-7 months to attain metamorphosis at 65-85 mm total length (Storer 1925; Jennings, unpubl. data). Larvae, which are solitary and almost never overwinter, typically metamorphose between July and September (Storer 1925, Jennings and Hays 1994a). Juvenile frogs are 25-30 mm total length at metamorphosis and commonly sun themselves during the day at the edge of the riparian zone next to the breeding site. As they grow, they gradually shift from diurnal and nocturnal periods of activity, to largely nocturnal activity (Hays and Tennant 1986). During periods of rainfall, both juveniles and a few adults may disperse away from breeding sites and may be found some distance (up to
0.8 km) away from the nearest water (Jennings, unpubl. data). Along the lower reaches of streams on the Central Coast of California which tend to almost completely dry up during the late summer, subadult and adult frogs have been found to occupy small mammal burrows under leaf litter or dense vegetation in the riparian zone (Rathbun et al. 1993). These frogs make overland trips every few days or so to isolated stream pools to rehydrate themselves although one frog remained in riparian habitat for 77 days (Rathbun in litt. 1994 as cited in Miller et al. 1996). Based on these observations, frogs found in coastal drainages appear to be rarely inactive, whereas those found in interior sites probably hibernate (Storer 1925).

Based on limited field data, California red-legged frogs appear to live about 8-10 years in the wild (Jennings, unpubl. data).

**Food and Feeding**

Larvae are thought to be algal grazers (Storer 1925), but the feeding ecology of larval California red-legged frogs is unknown. Juvenile and adult frogs have a highly variable animal food diet that includes: Amphipods, Isopods, Othoptera, Isopera, H emptera, H omoptera, N euoptera, Coleoptera, Lepidoptera, Diptera, H ymenoptera, Arachnids, Gastropoda, small fishes, amphibians, and small mammals (Stebbins 1972, Hayes and Tennant 1985, Baldwin and Stanford 1987). Most prey that can be swallowed that are not distasteful are eaten, with larger frogs capable of taking larger prey (Jennings and Hayes 1994a). Small red-legged frogs, Pacific treefrogs (Hyla regilla), and California mice (Peromyscus californicus) may contribute significantly to the diet of subadults and adults (Arnold and Halliday 1986, Hayes and Tennant 1985).

**Distribution**

Historically, California red-legged frogs were found throughout the Pacific slope drainages from the vicinity of Redding, Shasta County (Storer 1925), inland and at least to Point Reyes, Marin County (Hayes and Kremples 1986), California (coastally) southward to the Santo Domingo River drainage in Baja California, Mexico (Linsdale 1932). They also historically occurred in a few desert slope drainages in southern California (Jennings and Hayes 1994b). California red-legged frogs generally occurred below 1370 m in the Sierra Nevada foothills (Jennings 1996) and 1520 m in southern California, although some of the populations toward the upper limit of the range of this frog may represent translocations (Jennings and Hayes 1994a). In the Bay Area, this frog was historically abundant enough to support an important commercial fishery just before the turn of the century (Jennings and Hayes 1985) and up to the 1950s it was still considered to be present in much of the San Francisco Bay region (Stebbins 1959). However, earlier overexploitation, subsequent habitat loss from agriculture and urbanization, and the introduction of exotic aquatic predators have presently reduced red-legged frog distribution to scattered locations in the foothills and mountains of the San Francisco Bay region (Jennings, unpubl. data) (Figure 4.4).

**Current Status and Factors Influencing Population Numbers**

Based on the data presented in Jennings and Hayes (1994a, 1994b), California red-legged frogs appear to have disappeared from approximately 70-75% of their historic range in the state (Miller et al. 1996). This frog is most affected by land use patterns and other anthropogenic events which fragment high quality habitat and create environments unsuitable for the continued survival of the species (Jennings and Hayes 1994a, 1994b). Some of the more important factors negatively influencing frog populations include: conversion and isolation of perennial pool habitats (and surrounding riparian zones) to agriculture; reservoir construction projects; urbanization; lowering of the groundwater table by over-draft; overgrazing by domestic livestock; extended drought; mortality of juvenile and adult frogs by vehicles on roads; and the introduction of non-native predators such as mosquitofish (Gambusia affinis), bullfrogs (Rana...
catesbeiana) and crayfish (specifically Procambarus darkii) into breeding habitats (Miller et al. 1996). Juvenile and adult frogs have also been found in a number of human-created habitats such as irrigation canals, golf course ponds, sewage treatment ponds, gravel pits, and intermittent irrigation ponds (Jennings and Hayes 1994a; Jennings, unpubl. data). Such habitats may not be suitable for the long-term survival or successful reproduction of local frog populations, especially near urban areas where predators such as bullfrogs and raccoons (Procyon lotor) are able to build up large populations as a result of human activities (Jennings, unpubl. data).

Trophic Levels

Larval stages are primary consumers and post-metamorphic life stages are secondary/tertiary consumers.

Proximal Species

**Predators:** Common [=San Francisco] garter snake, Coast garter snake, Central Coast garter snake, bullfrog, heron, egret, raccoon, and introduced predatory fishes. Predacious aquatic insects prey on larvae only.

**Prey:** California tiger salamander, Pacific treefrog, California mouse, bullfrog, and aquatic and terrestrial insects.

**Habitat:** Willows, cattails, tules, sedges, and blackberries.

Good Habitat

Although California red-legged frogs can occur in ephemeral or artificially-created ponds devoid of vegetation, the habitats that have been observed to have the largest frog populations are perennial, deep (>0.7 m) water pools bordered by dense, shrubby riparian vegetation (Jennings 1988, Hayes and Jennings 1986). This dense riparian vegetation is characterized by arroyo willows (Salix lasiolepis) intermixed with an understory of cattails (Typha sp.), tules (Scirpus sp.), or bulrushes (Scirpus sp.) (Jennings 1988).

References


Western Pond Turtle
Clemmys marmorata
Mark R. Jennings

General Information

The western pond turtle (Family: Emydidae) is a moderate-sized (120-210 mm CL), drab brown or khaki-colored turtle often lacking prominent markings on its carapace (Bury and Holland, in press). Carapace coloration is usually dark brown or dull yellow-olive, with or without darker streaks or vermiculations radiating from the centers of the scutes (Ernst et al. 1994, Jennings and Hayes 1994).

There are two poorly differentiated subspecies of the western pond turtle (C. m. marmorata and C. m. pallida) with a wide zone of intergradation in central California (Bury 1970). Based on a morphological evaluation, Holland (1992) found three distinct evolutionary groups within this taxon. However, Gray (1995) found through DNA fingerprinting of C. marmorata samples from the extreme southern and northern edges of its range support the original designation of two distinct subspecies. Bury and Holland (in press) indicate that more comprehensive genetic studies are currently underway to determine the taxonomic status of this taxon.

In 1992, the western pond turtle was petitioned for listing as an endangered species (Sorensen and Propp 1992) based on concerns about widespread population declines due to the extensive loss of habitat, overexploitation, introductions of non-native aquatic predators (Sorensen and Propp 1992; see also Jennings and Hayes 1994). The U.S. Fish and Wildlife Service subsequently ruled that the petition was not warranted (USFWS 1993) and this turtle remains a candidate 2 species (Drewry 1994).

Reproduction

In California, sexual maturity in western pond turtles occurs at between seven and 11 years of age at approximately 110-120 mm CL with males maturing at slightly
smaller sizes and ages than females (Jennings and Hayes 1994). Sexual maturity is delayed in turtles that experience drought conditions and in more northerly populations (Jennings and Hayes 1994). Adult turtles typically mate in late April or early May, although mating can occur year-around (Holland 1985a, 1992). The nesting season is from late April to early August (Storer 1930, Buskirk 1992, Rathbun et al. 1992, Jennings and Hayes 1994, Goodman 1997a). Females emigrate from the aquatic habitats to an unshaded, upland location that may be a considerable distance (400 m or more) from riparian zones to nest (Storer 1930; Rathbun et al. 1992; Bury and Holland, in press). However, most nest locations are close to riparian zones if nesting substrates and exposures are suitable (Jennings, unpubl. data). Once a suitable site is located, females deposit from 1-13 eggs that have a thin, but hard (calcified) outer shell in a shallow (ca. 10-12 cm deep) nest (Rathbun et al. 1992, 1993)— usually in well-drained clay or silt soils (Jennings and Hayes 1994). Eggs laid in excessively moist substrates have a high probability of failing (Feldman 1982). Females can lay more than one clutch of eggs a year and may dig several “false” nests lacking eggs to deter potential predators (Rathbun et al. 1993, Goodman 1997b).

Growth and Development

Young turtles hatch at lengths of 25-29 mm CL (Ernst et al. 1994) after an incubation period of 3-4.5 months (Buskirk 1992; Bury and Holland, in press) and are thought to overwinter in the nest because there are only a few records of hatching turtle emergence in the early fall in southern and central California (Buskirk 1992, Jennings and Hayes 1994). Most hatching turtles are thought to emerge from the nest and move to aquatic sites in the spring (Buskirk 1992) where they typically double their length the first year and grow relatively rapidly over the next 4-5 years (Storer 1930, Holland 1985a). Young turtles spend most of their time feeding in shallow water dominated by relatively dense vegetation of submergents, short emergents, or algal mats (Buskirk 1992, Jennings and Hayes 1994). Juveniles and adults prefer slack- or slow-water aquatic habitats with basking sites such as rocks and logs (Bury 1972, Reese 1996). Water temperatures >15°C markedly increase turtle activity so many western pond turtles are probably active year around in coastal locations (Reese and Welsh 1998) and only active from March or April-October in interior locations (Bury and Holland, in press). Juveniles and adults seem to remain in pond environments except when ponds dry up or at higher elevations when turtles may disperse into terrestrial environments to hibernate (Jennings and Hayes 1994; Goodman 1997a, Bury and Holland, in press). In stream environments, juveniles and adults show considerable variation with regards to movements and the timing of movements into terrestrial environments (Rathbun et al. 1993, Reese and Welsh 1998). Some turtles will leave the stream during the summer when water conditions are low and water temperatures are elevated (>35°C), while others will not. However, almost turtles seem to leave streams during the winter months when large flood events are common (Rathbun et al. 1993). Additionally, some turtles will move considerable distances (e.g., 350 m) to overwinter in terrestrial habitats such as leaf litter or under the root masses of trees (Rathbun et al. 1992, 1993). Some individual turtles have displayed site fidelity for hibernation sites from year to year (Bury and Holland, in press).

Western pond turtles often move about from pool to pool in stream situations, sometimes on a daily basis during seasons of activity (Bury 1972, Reese and Welsh 1998). Distances moved along streams can be up to 5 km (Bury and Holland, in press). These turtles also have the ability to move several kilometers if their aquatic habitat dries up (Reese 1996) and they can tolerate at least seven days without water (Jennings and Hayes 1994; Bury and Holland, in press).

Western pond turtles are known to live over 42 years in the wild (Jennings and Hayes 1994) although most individuals have a much shorter life span of around 20-25 years (Bury 1972).

Food and Feeding

Juvenile and adult western pond turtles feed largely on the same items although juveniles feed more on smaller aquatic invertebrates (Bury 1986). Food items found in turtle stomachs include: algae, aquatic plants, Nematodes, Cladocera, Daphnia, Isopoda, Ephemeroptera (nymphs only), Odonata (nymphs only), Orthoptera, Hemiptera (nymphs and adults), N eumy dis (larvae only), Coleoptera (larvae and adults), T richoptera (larvae only), D iptera (larvae and adults), Araneae, Gastropoda, fishes, and amphibians (Carr 1952, Holland 1985b, Bury 1986, Ernst et al. 1994, Goodman 1997a). These turtles are dietary generalists and highly opportunistic (Ernst et al. 1994). They will consume almost anything that they are able to catch and overpower. The relatively slow pursuit of these turtles results in their diet being dominated by relatively slow-moving aquatic invertebrates and carrion, although aquatic vegetation may also be eaten (Evenden 1948, Bury 1986, Baldwin and Stanford 1987), especially by females having recently laid eggs (Jennings and Hayes 1994).

Distribution

The western pond turtle historically occurred in most Pacific slope drainages from Klickitat County, Washington along the Columbia River (Slater 1962) south to Arroyo Santa Domingo, northern Baja California,
Mexico (Jennings and Hayes 1994). Isolated populations are also known from Carson, Humboldt, and Truckee drainages in western Nevada (La Rivers 1962; Banta 1963). In California the species is known from most Pacific slope drainages between the Oregon and Mexican borders below 1430 m (Jennings and Hayes 1994). Turtle observations from above this elevation are thought to be introductions (Jennings and Hayes 1994). Western pond turtles are present throughout the Bay Area (Stebbins 1959) (Figure 4.5), although at much lower numbers and at fewer localities than previously—especially in urban areas (Jennings, unpubl. data).

**Current Status and Factors Influencing Population Numbers**

The western pond turtle is declining in population size and numbers throughout its range, particularly in southern California and the San Joaquin Valley (Bury and Holland, in press). Many turtle populations in these areas of decline are now composed almost entirely of old adults without any successful recruitment (Jennings and Hayes 1994). The reasons for these declines are largely due to urbanization, agricultural development, flood control projects, exotic diseases, exploitation for the food and pet trade, extended drought, and the introduction of exotic predatory species such as largemouth bass (Micropterus salmoides) and bullfrogs (Rana catesbeiana) which also compete for the availability of prey items—especially with young turtles (Brattstrom 1988; Buskirk 1992; Jennings and Hayes 1994; Reese 1996; Goodman 1997a; Bury and Holland, in press). Turtle nests and gravid females moving overland are especially vulnerable to predation by raccoons (Procyon lotor), whose populations have greatly increased in many rural areas due to the increase in human populations in these areas (Bury and Holland, in press; Jennings, unpubl. data). Additionally, some of the largest turtle populations in the Central Valley and southern California are found in sewage treatment ponds. Unfortunately, such habitats are probably unsuitable for the long term survival of the species because of the lack of suitable habitat for nest sites and increased vulnerability of adult turtles to predation by humans, raccoons, and other animals (Jennings, unpubl. data).

**Trophic Levels**

Young turtles are essentially secondary consumers; adults are primary and secondary consumers.

**Proximal Species**

**Predators**: Raccoon, bullfrog, black bear, humans, and introduced predatory fishes. Striped skunk and opossum prey on eggs and hatchlings, and herons and egrets prey on young turtles.

**Prey**: Aquatic insects, aquatic vegetation, and California tiger salamander larvae.

**Habitat**: Aquatic vegetation.

**Good Habitat**

The largest western pond turtle populations have been observed in warm water (15-35°C), slack- or slow-water habitats, which have abundant basking sites and underwater refugia. The presence of dense stands of submergent or emergent vegetation, and abundant aquatic invertebrate resources, as well suitable nearby nesting sites and the lack of native and exotic predators, are also important components (Bury 1972; Jennings and Hayes 1994; Bury and Holland, in press).

**References**


California Alligator Lizard
Elgaria multicarinata multicarinata
Kevin M. MacKay
Mark R. Jennings

General Information

The California alligator lizard (Family: Anguidae) is a large (100-125 mm SVL) alligator lizard with a broad head, keeled scales, and a reddish-blotched dorsum marked with nine or more dusky crossbands between the head and hindlimbs (Stebbins 1959, 1985). The top of the head is often mottled (Fitch 1938). There is a longitudinal stripe or row of dashes down the middle of each scale row on the belly (Stebbins 1985).

All alligator lizards in the western United States were formerly placed in the genus Gerrhonotus (e.g., Smith 1946; Stebbins 1958, 1959, 1972, 1985; Lais 1976). However, recently revised alligator lizard systematics places these species in the genus Elgaria (Waddick and Smith 1974; Gauthier 1982; Good 1987a, 1987b, 1988).

Reproduction

California alligator lizards are egg layers (Smith 1946) that probably reach sexual maturity in two years at about 73 mm SVL for males and about 92 mm SVL for females (Goldberg 1972). Mating apparently occurs over a relatively long period (up to 26 hours or more), but most copulation events are considerably shorter than this (Fitch 1935). Based on data from closely related E. m. webbii in southern California, adults emerge from hibernation and mate from late February-late May to August-mid September, and eggs are probably laid in small mammal burrows (Stebbins 1954) or under rocks from June-mid July (Goldberg 1972) or later into August (Stebbins 1954) or early September (Burridge 1965). Clutch sizes are 5-41 (average 13) and females can lay more than one clutch a year (Burridge 1965).

Growth and Development

Based on data from closely related E. m. webbii in southern California, incubation of the eggs probably takes 42-57 days (Atsatt 1952, Burridge 1965) and hatchlings appear from mid-August-early October at 30-36 mm SVL and 0.5-0.6 g (Burridge 1965, Goldberg 1972). Juveniles grow rapidly the next season, reaching sexual maturity about after 18 months (Goldberg 1972). The longevity of California alligator lizards in the wild is unknown, but marked lizards have been recaptured after four years (Jennings, unpubl. data).

Both juveniles and adults are active in the daytime, at dusk, and at night, and have a relatively low preferred temperature range (Brattstrom 1965, Cunningham 1956, Kingsbury 1994). Because of this, they do not bask. Instead, they prefer very dense cover and often position themselves under warmed objects such as rocks or pieces of wood during certain times of the day (Kingsbury 1994). Alligator lizards frequent riparian zones where their prehensile tails are used in climbing trees and other vegetation in pursuit of prey (Cunningham 1955; Stebbins 1959, 1972). They are also found under debris such as woodpiles, brush heaps, old logs, etc. (Stebbins 1954).

Food and Feeding

California alligator lizards probably consume the same food items taken by E. m. webbii. Food items recorded in the latter include: Isopoda, Orthoptera, Isoptera, Hemiptera, Homoptera, Coleoptera (larvae and adults), Lepidoptera (larvae and adults), Diptera (larvae and adults), Scorpionida, Araneida [including the egg cases and adults of the black widow spider (Latrodectus mactans)], Gastropoda, lizards (Sceloporus occidentalis, S. graciosus, and E. multicarinata), small mammals, and the eggs and young of small birds (Fitch 1935, Cowles 1937, Stebbins 1954, Cunningham 1956).

Distribution

California alligator lizards are found in the Sacramento Valley and surrounding foothills, from Shasta County south through the North Coast Range (Mendocino-Marin counties), the San Francisco Bay region and the South Coast Range to Ventura County (Fitch 1938). The elevational range is from sea level to around 1830 m in the Sierra Nevada (Basey and Sinclair 1980). This lizard is apparently absent from most of the San Joaquin Valley proper, but it is found on the northern Channel Islands (Fitch 1938). It intergrades with the E. m. scincicauda in Mendocino and Trinity counties in the north and E. m. webbii in Ventura County in the south and El Dorado County in the east (Stebbins 1985). They are found throughout the Bay Area (Stebbins 1959) (Figure 4.6).
Current Status and Factors Influencing Population Numbers

California alligator lizards are still present in good numbers over almost all of their historic range because of their ability to survive (and even thrive) in urban environments. The most important predator of these lizards in such modified habitats is the domestic cat (*Felis cattus*). In more natural habitats, alligator lizards are eaten by a number of reptile, avian, and mammal predators (Fitch 1935). They are still very abundant in the foothills of the Bay Area (Jennings, unpubl. data).

Trophic Levels

California alligator lizards are secondary/tertiary consumers.

Proximal Species

**Habitat:** Pickleweed, riparian vegetation, blackberries, willows.

**Predators:** Domestic cat, striped skunk, opossum, raccoon, heron, egret, hawks, coyote, red fox, bullfrog, common garter snake, and Coast garter snake.

**Prey:** Terrestrial insects, oligochaetes, and arachnids.

Good Habitat

California alligator lizards occupy many habitats from pickleweed flats to open grasslands, to oak woodlands, to mixed coniferous forest, to urban environments (Fitch 1935; Lais 1976; Stebbins 1954, 1985). However, the largest observed populations are in the riparian zones of oak woodlands and in coastal sage scrub near beaches (Jennings, unpubl. data).

References


Central Coast Garter Snake

Thamnophis atratus atratus

Mark R. Jennings

General Information

The central coast garter snake (Family: Colubridae) is a medium-sized (60-102 cm TL) garter snake with eight upper labial scales and a highly variable dorsal color throughout its range (Bellemín and Stewart 1977). Snakes usually have a dark olive to black dorsum and single yellow to orange dorsal stripe and sometimes lateral stripes of pale yellow (Stebbins 1985). The throat is a bright yellow. Both T. a. atratus and T. e. terrestris have similar dorsal and ventral colorations in habitats occupied along the central coast of California (Bellemín and Stewart 1977, Stebbins 1985). Boundy (1990) considers what is currently T. a. atratus, to be be actually composed of two different subspecies. However, his proposed subspecies from the mountains of the East Bay region and the South Coast Range (south of Santa Cruz County) has not been formally published.

Garter snake taxonomy has undergone a considerable number of revisions during this century, especially during the past 40 years. The snake T. a. atratus is often referred to as T. elegans atratus or T. couchii atratus in the literature (e.g., see Fitch 1940, 1984; Fox 1948a, 1948b, 1951; Stebbins 1954, 1972; Fox and Dessauer 1965; Lawson and Dessauer 1979). Rossman and Stewart (1987) were the first to convincingly elevate T. atratus as a separate species and this arrangement has been followed by others (e.g., see Lawson 1987).

Reproduction

Central coast garter snakes are live-bearers. Females give birth from 4-14 (or more) young (ave. 8.6) in the fall (August-September) [Fox 1948a, 1948b]. Adults probably mate annually during the spring (March-April) [Fox 1948b, 1952a], but females have the ability to store sperm for up to 53 months (Stewart 1972).
Growth and Development

Unknown. If similar to other garter snakes on the central coast of California, neonates are present from late August through November (Rathbun et al. 1993) and juveniles grow rapidly during the first year of their lives (Fox 1948a). Sexual maturity is reached in about 2-3 years (Fox 1948a). Longevity in the wild is unknown, but adults probably live for at least 4-5 years (Jennings, unpubl. data).

Food and Feeding

Juvenile and adult snakes feed almost entirely on fishes (e.g., Gasterosteus aculeatus, Hesperoleucus symmetricus, and Cottus spp.), newts (larvae and adults of Taricha torosa), toads (Bufo boreas halophilus), and frogs (e.g., larvae, juveniles, and adults of Hyla regilla, Rana aurora draytonii, R. boylii, and R. catesbeiana) [Fitch 1941; Fox 1951, 1952b; Bellemin and Stewart 1977; Boundy 1990; Barry 1994; Jennings, unpubl. data].

Distribution

Central coast garter snakes inhabit small streams, ponds, and other aquatic habitats in the San Francisco Peninsula and the East Bay Hills, Contra Costa County (south of the Sacramento River), southward through the South Coast Range to Point Conception, Santa Barbara County, and east to the western edge of the San Joaquin Valley (Fox 1951, Bellemin and Stewart 1977). Snakes north of San Francisco Bay are T. a. aquaticus (Fox 1951, Stebbins 1985). Their elevational distribution is from near sea level to 1290 m on Mount Hamilton (Fox 1951). They are relatively common East Bay and South Bay regions of the San Francisco Estuary (Stebbins 1959; Jennings, unpubl. data) (Figure 4.7).

Current Status and Factors Influencing Population Numbers

Central coast garter snakes are negatively affected by habitat alteration, especially by agriculture and urbanization which often results in intermittent aquatic habitats unsuitable for this species. These snakes are also probably negatively affected by the introduction of exotic predators such as bullfrogs (Rana catesbeiana) and largemouth bass (Micropterus salmoides), which are known to eat garter snakes (Schwalbe and Rosen 1988). However, these central coast garter snakes are still relatively abundant in aquatic habitats located in the foothills surrounding the Bay Area where urban development is less intrusive (Jennings, unpubl. data).

Trophic Levels

Central coast garter snakes are tertiary consumers.

Proximal Species

Predators: Raccoon, herons, egrets, hawks, and bullfrogs. Prey: threespine stickleback, sculpins, Pacific treefrog, California toad, foothill yellow-legged frog, California red-legged frog, bullfrog, coast range newt.

Good Habitat

Coast garter snakes are most abundant in riparian habitats with shallow ponds containing abundant numbers of native fishes and amphibians, and dense thickets of vegetation nearby (Jennings, unpubl. data). Such habitats are most common in natural sag ponds and artificial stock ponds (Barry 1994).

References

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Coast Garter Snake

Thamnophis elegans terrestris

Kevin McKay
Mark R. Jennings

General Information

The coast garter snake (Family: Colubridae) is a medium-sized (45-107 cm TL) garter snake with eight upper labial scales and a highly variable dorsal color throughout its range (Bellemin and Stewart 1977, Stebbins 1985). Snakes usually have a reddish to solid black dorsum (sometimes with a checkerboard of dark spots or bars), and single pale to bright yellow dorsal stripe, and two lateral stripes of yellow to salmon (Fitch 1983, Stebbins 1985). The throat and belly are usually tinged with orange flecks (Fox 1951). Both T. a. atratus and T. e. terrestris have similar dorsal and ven-
Reproduction

Coast garter snakes are live-bearers. Females give birth to from 4-14 young (average 8.6) in the fall (August-September) [Fox 1948, Stebbins 1954]. Adults probably mate annually during the spring (March-July) [Fox 1948, 1952a, 1956], but females have the ability to store sperm for up to 53 months (Stewart 1972).

Growth and Development

Unknown. If similar to other garter snakes on the central coast of California, neonates are present from late August through November (Rathbun et al. 1993) and juveniles grow rapidly during the first year of their lives (Fox 1948). Sexual maturity is reached in about 2-3 years (Fox 1948). Longevity in the wild is unknown, but adults probably live for at least 4-5 years (Jennings, unpubl. data).

Food and Feeding

Coast garter snakes subsist largely on slugs (Arion sp., Ariolimax columbianus, and others), California slender salamanders (Batrachoseps attenuatus), ensatinas (Ensatina eschscholtzii), arboreal salamanders (Aneides lugubris), Pacific treefrogs (Hyla regilla), western fence lizards (Sceloporus occidentalis), California voles (Microtus californicus), deer mice (Peromyscus maniculatus), young brush rabbits (Sylvilagus bachmani), harvest mice (Reithrodontomys spp.), nestling white-crowned sparrows (Zonotrichia leucophrys nuttalli), and nestling song sparrows (Melospiza melodia) [Fitch 1941; Fox 1951, 1952b; James et al. 1983; Barry 1994]. Fox (1951) also records at least one instance of cannibalism in the wild. There is a heavy preference for slugs, rodents, and nestling birds in some areas inhabited by this snake (Fox 1951, 1952b; James et al. 1983). Coast garter snakes will also eat a wide variety of fishes and amphibians if the occasion arises (see Fox 1952b).

Distribution

Coast garter snakes inhabit the North and South Coast Ranges from just north of the Oregon border, south to Point Conception, Santa Barbara County (Fox 1951, Bellemin and Stewart 1977, Stebbins 1985). They intergrade with T. e. elegans at mid-elevations of the North Coast Range (Stebbins 1985). The elevational range is from near sea level to around 350 m (Fox 1951). Coast garter snakes are widely distributed in the Bay Area (Stebbins 1959) (Figure 4.8).

Current Status and Factors Influencing Population Numbers

Coast garter snakes are negatively affected by habitat alteration, especially by agriculture and urbanization which often results in disturbed or open habitats unsuitable for this species. Because these snakes do not require permanent aquatic habitats for long-term survival like other garter snake taxa in the Bay Area, they are less affected overall by human activities. Coast garter snakes are still relatively abundant in terrestrial habitats located in the foothills surrounding the Bay Area (Jennings, unpubl. data).

Trophic Levels

Coast garter snakes are tertiary consumers.

Proximal Species

Predators: Raccoon, herons, egrets, hawks, California kingsnake, and bullfrog.
Prey: Pacific treefrog, California red-legged frog, bullfrog, Coast Range newt, oligochaetes, California mouse, California vole, white-crowned sparrow, brush rabbit (young only), shrews, slugs.

Good Habitat

Coast garter snakes inhabit meadows (such as grasslands) and clearings with second growth in the fog belt and also
chaparral (Stebbins 1972). They are often abundant in canyons with coast live oaks (*Quercus agrifolia*), California bay (*Umbellularia californica*) and numerous shrubs, as well as riparian zones or other areas of dense vegetation (such as blackberries (*Rubus discolor* and *R. ursinus*), thimbleberries (*R. parviforus*) and Baccharis (*Baccharis* spp.)) next to more open areas (Fox 1951).

**References**


received and published by the International Commission on Zoological Nomenclature to conserve the usage of T. s. infernalis and T. s. tetrataenia and designate a neotype for T. s. infernalis (Barry and Jennings 1998). Thus, the existing usage of the Fox (1951) nomenclature should be followed until a ruling is made on the case.

**Reproduction**

San Francisco garter snakes are live bearers which mate during the spring (March-April) and also during the fall (September-November), the latter often in breeding aggregations of several males and one female (Fox 1952a, 1954, 1955). Neonates (18-20 cm total length) are normally born in litters of 1-35 (average 16) during late July to early August (Fox et al. 1961; Cover and Boyer 1988; Barry 1993, 1994), although a few litters are born as late as early September (Larsen 1994). Females have the ability to store sperm for up to 53 months (Stewart 1972).

**Growth and Development**

Snakes are most active from March to September although they can be observed during any month of the year (Wharton 1989, Barry 1994, Larsen 1994). Juveniles grow rapidly during their first year, spending much of their time feeding in riparian zones or aquatic habitats (Barry 1994). Males and females probably reach sexual maturity in two years (at about 46 cm and 55 cm total length respectively), although some slower growing snakes reach sexual maturity in three years (Barry 1994). During the summer months, subadult and adult snakes may disperse away riparian areas into adjacent habitats to feed on amphibians in rodent burrows (Barry 1993). During the winter months, juvenile and adult snakes hibernate in small mammal burrows in adjacent upland habitats (Larsen 1994). Some snakes can move large distances (>2 km) over short periods of time (Wharton 1989), but limited radio tracking data indicate that most movements are considerably shorter than this distance (Larsen 1994).

**Food and Feeding**

Subadult and adult San Francisco garter snakes feed largely on the larvae and post-metamorphic life stages of Pacific treefrogs (Hyla regilla) and California red-legged frogs (Rana aurora draytonii). California toads (Bufo boreas halophilus), introduced bullfrogs (R. catesbeiana), introduced mosquitofish (Gambusia affinis), and threespine sticklebacks (Gasterosteus aculeatus) are also taken (Fox 1951, Wharton 1989, McGinnis 1984, Barry 1994). Juvenile snakes feed largely on newts (Taricha spp.), earthworms, and Pacific treefrogs (Barry 1993) and will refuse other non-amphibian items offered to them (Fox 1952b, Larsen et al. 1991). Adult snakes rarely eat California voles (Microtus californicus), even when they are abundantly available (Barry 1993).

**Distribution**

San Francisco garter snakes are a Bay Area endemic that are essentially restricted to San Mateo County, California (Stebbins 1959, Barry 1978) (Figure 4.9). Historically, they occurred in aquatic habitats and adjacent uplands along the San Andreas Rift Zone from near Pacifica, southeast to the Pulgas Water Temple, and along an arc from the San Gregorio-Pescadero highlands, west to the coast, and south to Point Año Nuevo (Barry 1978, 1994; McGinnis 1984). At least two recent records just south of Point Año Nuevo—from the mouth of Waddell Creek, Santa Cruz County—are questionable (Barry 1993, 1994). Intergrades with T. s. infernalis have been recorded in eastern San Mateo County (southeast of the Pulgas Water Temple) and extreme western Santa Clara County (Fox 1951, Barry 1994).

**Current Status and Factors Influencing Population Numbers**

San Francisco garter snakes have disappeared from significant portions of their native range due to habitat loss from agriculture and urbanization—especially from housing developments and freeway construction (Medders...
SOSC-55
1976; USFWS 1985; Barry 1978, 1993). Historically, the largest known population of snakes was at a series of sag ponds (locally referred to as the “Skyline Ponds”) along Hwy 35 in the vicinity of Pacifica, Daly City, San Bruno, and South San Francisco (Barry 1978, 1993, 1994; USFWS 1985). Today, this complex of ponds has been completely covered by urbanization. The large Bay Area population of snakes studied by Wharton (1989) has extensively declined due to the loss of several prey species from saltwater intrusion into the marsh (see Larsen 1994) and this population may now be close to extinction (Jennings, unpublished data). Besides the above, declines also resulted from large numbers of snakes being collected for the pet trade (especially overseas) and T. s. tetrataenia continues to be illegally collected for pets despite stiff penalties for doing so (e.g., see Bender 1981). Today, about 70% of the current remaining San Francisco garter snake habitat is composed of artificially constructed aquatic sites such as farm ponds, channelized sloughs, and reservoir impoundments (Barry 1993). Such habitats are often managed in ways that are detrimental to the snake and its preferred prey, the California red-legged frog (Barry 1993, 1994; Larsen 1994).

Current estimates put the number of San Francisco garter snakes at about 65 “permanent” reproductive populations of around 1500 total snakes >1 year of age (Barry 1993). About half the known populations are protected to some extent by refuges such as water preserves or state parks (Barry 1993). The key to preserving the species is to set aside adequate amounts of habitat and manage these areas for T. s. tetrataenia and its prey, especially California red-legged frogs (Barry 1993, 1994; Larsen 1994).

**Trophic Levels**

San Francisco garter snakes are tertiary consumers.

**Proximal Species**

*Prey:* Coast Range newt, California red-legged frog, threespine stickleback, Pacific treefrog, bullfrog.

*Predators:* Hawks, herons, egrets, bullfrog, striped skunk, opossum, and raccoon.

**Good Habitat**

San Francisco garter snakes are most abundant in natural sag ponds or artificial waterways that have been allowed to develop a dense cover of vegetative (Barry 1993). This is due to the presence of large amphibian populations (=prey base) and many basking sites for juvenile and adult snakes which are relatively secure from potential predators (Barry 1994). The presence of adjacent upland areas with abundant numbers of small mammal burrows are also important as hibernation sites for snakes during the winter (Larsen 1994).

**References**


Chapter 4 — Amphibians and Reptiles


Salt Marsh Harvest Mouse
Reithrodontomys raviventris
Howard S. Shellhammer

Life History

Salt marsh harvest mice (SMHM) are small, native rodents which are endemic to the salt marshes and adjacent diked wetlands of San Francisco Bay and are listed as an endangered species by the U.S. Fish and Wildlife Service and the State of California (Shellhammer 1982). They range in total length from 118 to 175 millimeters and in weight from 8 to 14 grams. They are vegetarians that can drink water ranging from moderately saline to sea water. They swim calmly and well. They do not burrow, but will build ball-like nests of dry grasses and other vegetation on the ground or up in the pickleweed (Fisler 1965). Their behavior is placid, so much so that their behavior is used as a secondary criterion in identifying them to the species level.

Historical and Modern Distribution

SMHM are composed of two subspecies. The northern subspecies, R. r. haliocoetes, is found on the upper portions of the Marin Peninsula; in the Petaluma, Napa and Suisun marshes; as well as a disjunct series of populations on the northern Contra Costa County coast. The southern subspecies, R. r. raviventris, is found in the more highly developed portions of the Bay from the Richmond area, down around the South San Francisco Bay (primarily south of a line between Redwood City and Hayward), and a disjunct series of small populations on the Marin Peninsula. Some modern distributions are indicated in Figure 5.1 and a listing of available trapping data are included in Appendix 5.1.

Their chromosome number and morphology have been studied by Shellhammer, and the two subspecies show some differences in chromosome shape indicating that genetic isolating mechanisms are beginning to form between them. No recent and modern genetic studies have been completed at the present time, hence nothing is known about the genetic variability of this species and whether or not it faces problems of inbreeding and random genetic drift as its average population size decreases.

The major threats to their habitat include filling, diking, subsidence, and changes in water salinity (Shellhammer 1982, 1989). Various estimates have been made that at least 75% of all the tidal marshes around the Bay have been filled in or otherwise destroyed over the last 150 years. Most of the remaining marshes have been back-filled or diked-off, and hence most of the remaining tidal marshes are narrow strips along the bay side of the levees. Those strip marshes and most of the remaining larger marshes have lost their upper and part of their middle zones, such that there is little escape cover from high tides available. In the southern end of the South San Francisco Bay, the combination of subsidence caused by water drawdown and the freshening of that part of the Bay by massive amounts of non-saline, treated sewage effluent has changed the saline vegetation of that area to brackish and freshwater species such as bulrushes (Scirpus sp.), cattails (Typha sp.), and peppergrass (Lepidium latifolium), species not used by SMHM (Duke et al. 1990; Shellhammer 1982, 1989).

Because of these influences, SMHM has disappeared from many marshes and is present in very low numbers in most others. The highest consistent populations are found in relatively large marshes along the eastern edge of San Pablo Bay and in old dredge spoil...
disposal ponds on former Mare Island Shipyard property; most of these marshes are in or will be included in the San Pablo Bay unit of the San Francisco Bay National Wildlife Refuge (Bias and Morrison 1993, Duke et al. 1995). Other areas supporting large populations include some parts of the Contra Costa County coastline (Duke et al. 1990, 1991), some parts of the Petaluma Marshes, and the Calaveras Point Marsh in the South San Francisco Bay (Duke et al. 1990), although the latter area is deteriorating because of the declining salinity and correlated changes in vegetation.

Diked wetlands adjacent to the Bay have grown in importance as the tidal marshes bayward of their outboard dikes have decreased in size and quality (Shellhammer 1989). Most of such diked marshes in the South San Francisco Bay are being threatened by urban and industrial development along their borders. In addition, most of these diked marshes are not managed to provide adequate vegetative cover of halophytic species or to maintain their salinity over time (Duke et al. 1990, Shellhammer 1989).

**Suitable Habitat**

SMHM are dependent on the thick, perennial cover of salt marshes and move in the adjacent grasslands only in the spring and summer when the grasslands provide maximum cover (Fisler 1965). Their preferred habitats are the middle and upper portions of those marshes, i.e., the pickleweed (Salicornia virginica) and peripheral halophyte zones, and similar vegetation in diked wetlands adjacent to the Bay (Shellhammer et al. 1982, 1988). Some areas of known suitable habitat are shown in [Figure 5.1](Figure 5.1 Salt Marsh Harvest Mouse - Some Current Locations and Suitable Habitat).

Note: Mice are likely present in areas identified as “suitable” habitat based on current information regarding habitat types. Mice may also be present in other areas.
Conservation and Management

There are many questions that need to be addressed in order to properly manage the SMH M. They include the following: (1) Little is known about the degree of genetic heterozygosity and polymorphism of this species. Is it variable, and hence is the SMH M resistant to increasing isolation, genetic drift, and potential increased inbreeding, or is it a species that has survived a series of genetic bottlenecks and become monomorphic and lacks resilience? Without information on its population genetics, the only prudent course of action is to argue for the largest possible population sizes of SMH M. Much more needs to be known about the population genetics of this species if it is to be properly managed over the long run. (2) It is not known how much upland edge constitutes enough of a buffer to protect SMH M from alien predators (especially cats) and human disturbance. The U.S. Fish and Wildlife Service Endangered Species biologists recommend 100 feet, but 100 feet of grassland, for example, may not be enough of a barrier to keep out dogs, cats, red foxes, or humans. (3) The impact of introduced red foxes is not known, but they have had a great impact on the California clapper rail, which is found in the same marshes with SMH M.

Control of red foxes is being carried out in those marshes in which there are rails and mice, but not in all marshes potentially containing SMH M alone. Actually, very little is known about the effects of predators on the SMH M, including the effects of the rail. (4) Little is known of the interactions between various species of rodents in diked marshes. Geissel et al. (1988) demonstrated seasonal displacement of SMH M from optimal habitat by California voles. Elaine Harding of U.C. Santa Cruz is studying (as of 1997-98) rodent interactions and has concerns that certain management practices in diked wetlands might work against SMH M. (5) Little is known about the impact of peppergrass on SMH M numbers. SMH M remain in mixed pickleweed-peppergrass communities (Duke et al. 1990, 1991), but no studies have been carried out in areas of 100% peppergrass, a condition that is becoming increasingly common in the southern end of the South San Francisco Bay. (6) Lastly, there is the strong possibility that youthful pickleweed marshes are more productive of SMH M than older ones. That is certainly the case reported by Bias (1994) and Bias and Morrison (1993) at Mare Island Naval Shipyard and in the marshes bordering the adjacent San Pablo Bay, a marsh that has been growing actively by accretion for decades. The effect of the relative youth of marshes (or possible the lack of their senescence), needs to be looked at along with the potential effects of toxics, the depth of buffer zones when marshes are bordered by either urban and industrial development, and other concerns spelled out previously in this document.

References


Additional Readings

### Alameda County

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Chapter 5 — Mammals

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California Vole
Microtus californicus
William Z. Lidicker, Jr.

Life History

California voles are common inhabitants of the San Francisco Bay wetlands. They are vegetarians, feeding extensively on Salicornia and other marsh vegetation. They make runways through the vegetation, burrow extensively in non-flooded areas, and often utilize driftwood for cover. They are critically important prey species for a wide variety of mammalian and avian predators.

The population dynamics of voles has been studied intensively in adjacent upland grasslands (Cockburn and Lidicker 1983; Krebs 1966; Lidicker 1973; Pearson 1966, 1971; Salvioni and Lidicker 1995), but little is known about marsh populations. It is not known, for example, if most marsh populations are merely extensions of upland ones or independent demographic units. An exception is the San Pablo Creek vole; see below. Grassland populations around the Bay exhibit annual or multi-annual cycles in numbers, but the demographic behavior of salt marsh populations is unknown. Similarly, we know that grassland voles breed mainly in the wet season, and especially intensely from February through May. Voles in marshes may well be different, perhaps breeding mostly in the summer and very little during the flood-prone winters.

California voles are keystone species in grassland communities by virtue of their importance as a major prey species (Pearson 1985) and their potentially great effect on vegetation (Lidicker 1989). Thus, if similar roles are played in San Francisco Bay wetlands, these rodents may be vital to the health of the wetland communities. Because they are known to exhibit strong fluctuations in numbers (four orders of magnitude), suitable habitat patches must be large enough for the species to survive low-density bottlenecks. These voles are also known to exhibit strong non-trophic interactions with other species of mammals. The introduced house mouse (Mus musculus) is strongly affected negatively by the presence of voles (DeLong 1966, Lidicker 1966). Interactions with Western harvest mice (Reithrodontomys megalotis) are more complex (Hesk et al. 1984). At moderate Microtus densities harvest mice are positively influenced, presumably because the harvest mice make effective use of vole runways. However, at high vole densities, the Reithrodontomys are strongly negatively impacted. It is possible that salt marsh harvest mice (Reithrodontomys raviventris) may interact in a similarly complex way with voles. Geissel et al. (1988) demonstrated seasonal displacement of salt marsh harvest mice by voles. More subtle indirect effects may also be important. For example, if voles sustain populations of red fox (Vulpes vulpes), an indirect negative effect on clapper rails (Rallus longirostris obsoletus) may be manifest.

Historical and Modern Distributions

The taxonomic status of San Francisco Bay voles is complex. Marsh inhabiting voles from Point Isabel (Contra Costa County) south on the east side of the Bay and around to the west side as far north as Redwood City have been described as the subspecies paludicola. Thaeler (1961) examined these populations in detail and concluded that at least the East Bay populations could not be distinguished from the upland subspecies californicus. Voles from the Marin County side of the Bay are placed in M.c. eximius, and those from Grizzly Island (Solano County) and eastward into the Delta represent the large, dark subspecies aestuarinus. Of special interest and concern, Thaeler (1961) described the vole population inhabiting the marshes around the mouth of San Pablo Creek (Contra Costa County) as M.c. sanpabloensis. This subspecies is viewed as a species of special concern by the State of California (Williams 1986). It is darker and yellower than the adjacent populations of M.c. californicus. Further, its palatines are deeply excavated along their posterior borders, the rostrum is narrow, and the auditory bullae relatively inflated.

Suitable Habitat

Habitat use extends from adjacent upland grasslands into both salt and freshwater marshes, at least into those where flooding does not occur regularly. Voles are good swimmers, however, and can survive occasional inundation. Some known current locations and potential suitable habitats are shown in Figure 5.2.

Conservation and Management

Efforts to conserve wetlands should be aware of the endemic form M.c. sanpabloensis and attempt to achieve
representation of the other three currently recognized subspecies in the Bay Area as well. Because of their role as a major prey species, California voles are likely keystone species in the health of Bay Area wetland communities.

References


Salt Marsh Wandering Shrew
Sorex vagrans haliocoetes
Howard S. Shellhammer

Life History

The salt marsh wandering shrew (SMWS) appears to have some of the most restrictive food and habitat requirements of any mammal inhabiting the marshes of the greater San Francisco Bay Region—far more, for example, than the endangered salt marsh harvest mouse (Reithrodontomys raviventris). While little is known of the SMWS subspecies, shrews in general are insectivores which are born in the spring and become sexually mature the following winter. SMWSs have gestation periods of about 21 days (Owen and Hoffman 1983). Many shrew species have only one litter, and adults die after the young are weaned (Jameson and Peeters 1988).

Historical and Modern Distribution

The historical range of the SMWS extended from the northern end of the San Francisco Peninsula, down through the marshes of the South San Francisco Bay, and up through the marshes of western Contra Costa County to about the Benicia Straits. Johnston and Rudd (1957) suggested that between 1951 and 1955 shrews represented about 10% of the small mammals of the marshes. They were far less numerous in the 1970s and 80s, at least in the southern part of its range (Shellhammer, pers. obs.). Known or suspected populations as of 1986 included marshes south of Foster City and Hayward and in the San Pablo marshes of the San Pablo Bay (WESCO 1986). This subspecies of vagrant shrew is currently confined to the salt marshes of the South San Francisco Bay (Figure 5.3). It exists in a narrow band of tidal salt marsh and does not seem to be present in diked marshes.
Suitable Habitat

The SMWS’s habitat is wet, medium high salt marshes. It is best described by D. Williams (1983) in a draft report for the California Department of Fish and Game using material primarily from Johnston and Rudd (1957): “[Salt marsh wandering shrews] frequent areas in the tidal marshes providing dense cover, abundant food (invertebrates), suitable nesting sites, and fairly continuous ground moisture. Their center of activity is in the ‘medium high marsh,’ about 6 to 8 feet above sea level, and in lower marsh areas not regularly inundated. Suitable sites are characterized by abundant driftwood and other debris scattered among pickleweed (Salicornia). The pickleweed is usually one to two feet in height. The detritus preserves moisture and offers refuge in dry period to amphipods, isopods and other invertebrates, and resting sites for shrews. Nesting material consists of plant material, primarily Salicornia duff. The higher marsh, 8 to 9 feet in elevation, is too dry and offers only minimal cover—few to no shrews occupy this zone. The lower cordgrass (Spartina) zone is subjected to daily tidal floods and has cover too sparse for shrews.”

Some potential suitable habitat locations are shown in Figure 5.3.

Conservation and Management

Johnston and Rudd’s 1957 paper represents the last scientific work on the subspecies, per se. The rest of the reports (Williams 1983; WESCO 1986; this present effort) are all based on that study and that of Rudd 1955.
Many changes have taken place since the early 1950s and little to nothing is known as to how such changes have affected the prey or habitat requirements of this shrew. The southern part of the San Francisco Bay has been greatly freshened by hundreds of millions of gallons of treated sewage outflows per day, and this freshening has brought about changes in plant species composition. Until point source reductions were placed on industrial sewage in the 1980s, large amounts of heavy metals, as well as polychlorinated biphenyls and petroleum hydrocarbons were poured into the Bay. In addition, the storm runoff and inflows of creeks and small rivers carried unknown amounts of pesticides, petroleum compounds, and other toxic substances. It is not known how decreased salinity and increased toxicity in the South Bay may have impacted the shrews, either directly, or indirectly, through changes in the amount and diversity of their prey. In addition to salinity, vegetation changes, and toxics, many of the marshes of the South Bay have subsided, and the Salicornia bands have become more degraded and more heavily inundated. Again little is known as to the effects of this shrew of such changes.

The Suisun Marsh Shrew (SMSW) is currently listed as "Mammalian Species of Special Concern" by the California Department of Fish and Game and as a candidate species for listing in Category 2 by the U.S. Fish and Wildlife Service. Neither classification offers legal protection to its habitat. Little recent biological information is available to support its classification as a protected species, a status it merits.

**References**


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**Suisun Shrew**

*Sorex ornatus sinuosis*

Kevin MacKay

**Life History**

The Suisun shrew is a small (95-105 mm in total length), dark, insectivorous mammal with a long, pointed nose, and a well-developed scaly tail (37-41 mm). Suisun shrews are carnivores and predators feeding primarily upon amphipods, isopods, and other invertebrate species (WESCO 1986, Hays 1990). The shrews may also occasionally serve as prey for several large predators such as the short-eared owl (*Asio flammeus*), northern harrier (*Circus cyaneus*), and black-shouldered kite (*Elanus caeruleus*) (WESCO 1986).

The reproductive period of the Suisun shrew extends from late February through September, with the majority of breeding occurring from early spring through May. A second breeding period occurs in late summer when the young born the previous spring are mature and able to mate for the first time.

Shrews typically construct domed, cup-like nests composed of small paper scraps and dead material from plants such as Pacific cordgrass (*Spartina foliosa*), pickleweed (*Salicornia virginica*), and salt grass (*Distichlis spicata*). The nests are usually placed directly on the soil surface under driftwood, planks, or wood blocks, and are situated above the high tide line to escape flooding (WESCO 1986). Runways enter from the sides and from beneath, and are not opened until two to three weeks after the birth of the young (Johnston and Rudd 1957).

After the young have dispersed, the nests may be used by other small mammals such as the endangered salt marsh harvest mouse (*Reithrodontomys raviventris*) (WESCO 1986).

There are no published data on the gestation period of the Suisun shrew, but the salt marsh wandering shrew and other small shrews have a gestation period of about 21 days (Owen and Hoffmann 1983, WESCO 1986). Litter size ranges from four to six individuals, with a survival rate of 55 to 60 percent from near birth to just after weaning (Johnston and Rudd 1957). Causes of mortality include drowning from high tides, death of the mother, starvation, exposure, and predation (WESCO 1986). The young remain in the nest for up to five weeks and then move into adjacent areas (Rudd 1955).

Suisun shrews seldom reach their maximum life expectancy of 16 months, and populations turn over on an annual basis. Populations in the early spring typically consist of adults born the previous year. These individuals gradually die off during the summer months, and by fall have been almost completely replaced by young born the previous spring (Owen and Hoffmann 1983).
Activity patterns vary according to season and reproductive condition in the Suisun shrew, but the subspecies is predominately nocturnal, especially during the breeding season. Sexually mature shrews are very active in the spring, concurrent with the breeding season, but are less active during the early summer. Young-of-the-year born in early spring become sexually mature by late summer, and their activity patterns peak during this second breeding season. Others, born later in the season are still sexually immature by late summer and remain comparatively inactive during this period (Owen and Hoffmann 1983).

Hays (1990) found that during the non-breeding season, shrews lived in loose social groups of 10 to 15 individuals. These groups contained only one adult male, and one such group occupied 0.07 ha. In the spring other adult males invade these groups, disrupting the stable structure by competing among themselves. Territorial behavior in shrews has not been well documented in the field. However, Rust (1978) noted territorial patrolling in observations of breeding captive Suisun shrews.

**Historical and Modern Distribution**

One of the nine subspecies of ornate shrew that occur in California, the Suisun shrew is a relatively rare inhabitant of the salt marsh ecosystem of San Pablo and Suisun Bays (WESCO 1986). Johnston and Rudd (1957) estimated that the shrews represent approximately 10 percent of the mammalian fauna present in marsh habitats, and were less abundant than mice (Mus sp.), rats (Rattus sp.), voles (Microtus sp.), and harvest mice (Reithrodontomys sp.).

The historical extent of the Suisun shrew distribution is unknown (WESCO 1986) (Figure 5.4). According to Rudd (1955) the subspecies historically inhabited the tidal saline and brackish salt marsh communities of northern San Pablo and Suisun bays, ranging from the mouth of the Petaluma River, Sonoma County on the west, eastward through Southampton and Grizzly Island to approximately Collingsville, Solano County (WESCO 1986, Rudd 1955, Williams 1983). The western extent of the range was redefined by Brown and Rudd (1981) as they identified the shrews inhabiting the marshes west of Sonoma Creek and Tubbs Island as S. o. californicus (WESCO 1986, Williams 1983).

However, surveys completed by Grinnell (1913) discovered Suisun shrews only at Grizzly Island. Researchers (WESCO 1986) have speculated that, at that time, the shrew was restricted to the greater Grizzly Island area because of the lack of suitable habitat throughout the rest of the historic range. The 1914 soil survey of the San Francisco Bay Area identifies most of the Napa Marsh as low tidal mud flats, a habitat that would be consistently inundated by tidal waters and thus uninhabitable by Suisun shrews or other small mammals. Once these areas were diked, and suitable habitat created, the shrew may have expanded its historic range into these adjacent areas (WESCO 1986).

There are no data available which directly measures the current densities of Suisun shrew populations. The number of individuals within a population appears to vary with season and habitat type. Newman (1970) estimated that the most favorable habitat supported shrew densities of as many as 111 individuals per hectare. A related species, the dusky shrew (Sorex obscurus), has overlapping home ranges averaging 0.037 ha in size, with a density of 37 to 42 individuals per hectare. These latter figures are probably a more accurate depiction of Suisun shrew populations as the amount of favorable habitat is limited throughout most of its range (WESCO 1986).

The Suisun shrew is currently limited in its distribution to the scattered, isolated remnants of natural tidal salt and brackish marshes surrounding the northern borders of Suisun and San Pablo bays (WESCO 1986).

Rudd (1955) identified four distinct populations of Suisun shrews: the Grizzly Island population, found throughout the marshlands east of Suisun Slough; a peripheral population found west of Suisun Slough and on Morrow Island; the Southampton population, restricted to the Benicia State Recreation Area; and the Sears Point Road population located in the Napa marshes.

No Suisun shrews were captured in either of the two most recent population studies (Williams 1983, WESCO 1986) that attempted to assess the current distribution of the shrew. This lack of trapping success can possibly be attributed to the extremely high rainfall in 1982 and 1986. Most of the low-lying marshes were flooded for extended periods of time, adversely affecting the small mammal populations. Additional trapping efforts for salt marsh harvest mice occasionally yielded Sorex captures; however, only one capture, at Cullinan Ranch on South Slough, was identified as S. o. sinuosis (WESCO 1986).

WESCO (1986) plotted all known S. o. sinuosis captures to delineate extant populations. Only two individual areas were identified that support populations of Suisun shrews: Grizzly Island and Solano Island Number 1. Nine additional marsh areas were also identified as having a high probability of supporting Suisun shrew populations: Skaggs Island, Appleby Bay/Coon Island, Steamboat Slough, Vallejo, Morrow Island, Cordelia Slough South, Hammond Island, Simmons/Wheeler Islands, and Collingsville (WESCO 1986).

**Suitable Habitat**

Suisun shrews typically inhabit saline and brackish tidal marshes characterized by Pacific cordgrass (Spartina foliosa), pickleweed (Salicornia virginica), gumplant...
Mammals

(Grindelia humulis), California bulrush (Scirpus californicus), and common cattail (Typha latifolia). However, shrew occurrence appears to be more strongly associated with vegetation structure rather than species composition. Suisun shrews prefer dense, low-lying vegetation which provides protective cover and suitable nesting sites, as well as abundant invertebrate prey species (Owen and Hoffmann 1983). Driftwood, planks, and other debris found above the high-tide line also affords the shrew with valuable foraging and nesting sites. In addition, adjacent upland habitats provide essential refuge areas for Suisun shrews and other terrestrial animals during periods of prolonged flooding (Williams 1986). Some areas of potentially suitable habitat for the Suisun Shrew are shown in Figure 5.4.

**Conservation and Management**

Williams (1986) identified the lack of an adequate elevational gradient of marsh vegetation and adjacent upland habitats as the principal obstacles to the recovery of Suisun shrew populations in San Pablo and Suisun bays. However, as the Suisun shrew does not seem to make use of upland grasslands (Hays 1990), and because of evidence of interbreeding with *S. o. californicus*, future marsh management practices should include the provision of elevated sites that flood only occasionally, but not include upland grassland, which would encourage contact with californicus.
References


Ornate Shrew
Sorex ornatus californicus
Elaine K. Harding

Life History

Ornate shrews are small insectivores weighing on average five grams and with a tail short relative to the length of the head and body (Owen and Hoffmann 1983). There are nine subspecies of ornate shrew found throughout California and Mexico, and three of these are currently considered for federal listing (USFWS 1989). Sorex ornatus californicus is not considered a sensitive species, although very little information is known about this subspecies, as is the case with most shrews. This subspecies may coexist with the Suisun shrew (Sorex ornatus sinuosus) in the marshes of San Pablo and Suisun bays. The pelage of the ornate shrew is grayish brown dorsally to a pale gray ventrally, which differentiates it from the Suisun shrew's darker pelage (Rudd 1955). Shrews reproduce from late February through September, with peaks in late spring and summer. There is little information on this species' litter size or survival, but embryo counts ranged from four to six (Owen and Hoffmann 1983). A few young-of-the-year born in spring may mature by summer and reproduce. These shrews live no longer than 12 to 16 months. Their extremely high metabolism requires that they eat high energy foods often throughout the course of a day. S. o. californicus is a predator of invertebrates and may find food and cover in low, dense, moist vegetation. In wetland areas, amphipods are known to be important sources of food for shrews, but the diet of this shrew has not been thoroughly investigated.

Historical and Modern Distribution

S. ornatus californicus' range is from the Sacramento Valley southwest to the Central Coast, including the San Francisco Bay except for the southwestern portion of San Pablo Bay. A thorough account of its current range within the San Francisco Bay is not available due to a lack of identification to the species level when found incidentally during other studies. Some known and potential habitats, however, are indentified in Figure 5.5.

The ornate shrew may hybridize with the Suisun shrew in particular parapatric zones in North San Pablo Bay marshes. Rudd (1955) described populations at Grizzly Island and Sears Points which exhibited intermediate morphological characters between S. o. californicus and S. o. sinuosus. More recently Brown and Rudd (1981) concluded that populations at other locations once considered hybrids of the Suisun shrew (S. o. sinuosus) and the salt marsh wandering shrew (S. vagrans) are only
slightly differentiated populations of S. o. californicus. However, further discrimination of the taxonomic status between S. o. californicus and S. o. sinuosus is needed as the two subspecies have identical karyotypes (Brown 1971).

**Suitable Habitat**

The ornate shrew prefers semi-arid, grassland and riparian habitats. Despite this preference, it is also found in brackish and saltwater marshes in San Pablo Bay and perhaps in other marshes throughout the Bay Area (Figure 5.5) based on records of identification only to the genus *Sorex*.

**Conservation and Management**

The ornate shrew is an uncommon inhabitant of the upland transition zones and marshes in the San Francisco Bay. Although it is not currently endangered, its local population status may be a general indicator of the health of an ecosystem, particularly as shrews are very good barometers of contaminant loads. Because they prey on a variety of invertebrates, they often bioaccumulate more rapidly than other species of similar size. It is important to monitor the status of this species and to research their potential as an indicator of wetland health.

**References**


North American River Otter
Lutra canadensis
Michael L. Johnson

Life History
Adult river otters can range in size from 900 mm to almost 1,300 mm, and can weigh from 5 to 14 kg. Clinal variation in size is present, with otters becoming smaller from north to south, especially along the Pacific Coast. Sexual dimorphism is present, with females being smaller than males. Litter sizes vary with location from 1-5, with the average litter being 2-3. Females may become pregnant every year or may become pregnant only in alternate years, depending on local conditions. Pups grow rapidly and typically emerge from the den at two months, and are weaned at about three months. Dispersal of offspring typically occurs at about one year (M. elquist and D. Ronkert 1987). A study of food habits of river otters in the Suisun Marsh performed by scat analysis (Grenfell 1974) indicated that crayfish were the most frequent prey item, and birds and fish were alternately the second most frequent prey item, depending on season. Although individual species identification of the fish was not attempted, scales and teeth indicated that the most probable prey items were the carp (Cypinus carpio), Sacramento squawfish (Ptychocheilus grandis), the tule perch (Hystrocorapustraskii), and the striped bass (Morone saxatilis). Mammals, plants, and reptiles appeared to be taken opportunistically, as their frequency of occurrence in scats was never higher than 10 percent.

Historical and Modern Distribution
The North American river otter is a member of the Family Mustelidae Subfamily Lutrinae Tribe Lutrini. Formerly abundant throughout much of Northern California, they are placed into the subspecies L. c. pacifica Rhoads (Stephens 1906, Ingles 1965, Deems and Pursley 1978). Alternate subspecies designations placed otters from California’s Sacramento and San Joaquin drainages into a separate subspecies, L. c. brevipilosus Grinnell (Anthony 1928, Grinnell 1933) with the type locality being Grizzly Island, Suisun Bay, Solano County, California (Grinnell 1933). Although there were 18 subspecies at one time, currently there are six. It was recently proposed that these be placed into a single species with the South American river otter, and placed into a New World genus Lontra (Van Zyll De Jong 1987).

Northern river otters were once found in all major drainages throughout North America, possessing one of the largest geographic ranges of all mammals. The present distribution over North America extends from 25° north latitude in Florida to over 70° north latitude in Alaska, and from eastern Newfoundland to the Aleutian Islands (Towell and Tabor 1982). In California, the distribution of river otters early in the 20th century included the Sacramento, San Joaquin and North Coast river drainages, eastward from the coast to the Sierra crest and to the Warner Mountains of Modoc County, and from the San Joaquin River east to the Sierra crest (Belfiore 1996). Grinnell placed the center of species abundance in the Sacramento-San Joaquin Delta (Grinnell 1933). Intensive trapping for pelts occurred during the latter half of the century (Ingles 1965, D. Uplaix 1978, Mason 1989, H. Albrook et al. 1994). Recent declines are also blamed on habitat destruction or alteration as well as the deterioration in water quality (Deems and Pursley 1978, Mason 1989). The California Fish and Game Commission imposed a ban on trapping in 1969. Despite meager evidence, furbearer status reports indicated that the populations were increasing throughout California (Schrempf and White 1977), and in the early
1980s the California Department of Fish and Game proposed an open river otter trapping season. However, the lack of evidence for population recovery led to maintenance of the ban in 1984 (Belfiore 1996). River otters are currently classified by U.S. Fish and Wildlife as a Class II species according to the Endangered Species Act.

The highest densities of river otters are currently found in the Klamath-Trinity River drainage, and in the Sacramento River drainage (Schrempf and White 1977). Although historical distributions do not place otters east of the Sierra crest, there are recent reports of otters in Mono and Inyo counties (Schrempf and White 1977). In the San Francisco Bay region, most river otter sitings have been in the Suisun region (Figure 5.6).

**Suitable Habitat**

Otters are found in freshwater habitats throughout northern California, as well as in brackish, salt marsh and other marine locations (Grinnell 1933, Schrempf and White 1977). Distributions are primarily associated with good river bank cover, but are not specific to any particular vegetation type (Figure 5.6). Otters are found at elevations as high as 9,000 feet, but prefer lower altitudes due to food preferences (Schrempf and White 1977).

**Conservation and Management**

Habitat loss and degradation continues to be a major problem in maintaining viable populations of river otter.
ters. Additionally, river otters are almost entirely aquatic, and therefore are at risk from contaminants that they directly contact and that may bioconcentrate through the food chain. A recent review of the association between the status of mink and otter populations and exposure to organochlorine chemicals in the Great Lakes indicates that there may be an association between higher levels of chemicals and lower harvest rates (Wren 1991), although the association needs further documentation.

Loads of chemical contaminants in the Bay-Delta system are sufficient to be toxic to invertebrates (Bailey 1993, Bennett et al. 1995). The river otter would be expected to bioaccumulate persistent, water-borne contaminants from their prey items (Ropek and Neely 1993). In fact, the San Francisco Estuary Project recommended monitoring of the river otter in the Bay-Delta system as a sentinel species for providing information about the extent of exposure incurred by wildlife in general (Bailey et al. 1995, Belfiore 1996).

References


Southern Sea Otter

Enhydra lutris nereis

David G. Ainley
Robert E. Jones

Life History

Individual adult sea otters show very little movement, being resident the year round where they occur. Males and females occupy home ranges of about 35 and 80 hectares, respectively. Subadult bachelor males disperse much more widely and comprise the vanguard as the population slowly expands to recolonize its former range. Pupping can occur year round, but mainly occurs December through March.

Sea otters feed mainly on benthic invertebrates, such as bivalves, abalone, urchins, cephalopods, and crustaceans, but they also eat fish. Lacking a layer of blubber, they must eat voraciously to maintain body heat, and consume 15-35% of their body weight daily (Leet et al. 1992). Their dense fur is their only insulation against the cold waters that they frequent.

Historical and Modern Distribution

The historic range of the sea otter in the eastern North Pacific extended along the coast from central Baja California, Mexico, north through the Aleutian Islands. Presently, however, owing to intensive hunting for fur in the 18th and 19th centuries, the range is much restricted and is centered around two populations, one in the Aleutian Islands and Southeast Alaska (south to Vancouver) and one in California. Each population is considered to be subspecifically distinct; the southern (or California) sea otter is listed as threatened under the Endangered Species Act (state and federal). The southern population, currently numbering a little over 2,200 animals, has grown from a few dozen animals that resided in a refugium near Pt. Sur early in this century.

In San Francisco Bay, sea otters once occurred abundantly in the central part at least as far inland as the mouth of Sonoma Creek, Sonoma/Marin County. The population in the Bay was likely in the low thousands but, for their pelts, all were hunted to extinction by the early-1800s (Skinner 1962). Sea otter teeth are very abundant in the middens of early Americans that are scattered around the Bay shores (e.g., Emeryville; Broughton 1999).

It has taken several decades for the population to spread from the focus near Pt. Sur north to San Mateo County (and south to Pismo Beach, Ventura County). In the San Francisco Bay Area, otters are regularly seen north to Pt. Reyes, Marin County; the northern most extent of the breeding population is at Pacifica, San Mateo County. Until recently, no documented sightings of sea otters in the Bay had been made, although sightings existed for the outer portion of the Golden Gate. During the 1990s, however, several sightings were confirmed in the Bay, including ones near Strawberry Point, north of Sausalito, and in Richardson Bay (McHugh 1998) (Figure 5.7).

Suitable Habitat

Sea otters occur in shallow, usually protected, nearshore waters to about 15 m deep. Throughout the range, which is more or less linear and continuous in California, sea otters prefer, but are not restricted to, rocky substrates near points of land (Kenyon 1969, Leet et al. 1992). The promontories provide protection from ocean swells and lush growths of kelp usually occur in these areas. The otters normally do not come to land but use the kelp for resting, support, and protection from predators (sharks). The kelp in turn is maintained by the otters through their predation of kelp grazers such as sea urchins (Dayton 1975). Thus, the invasion of rocky habitat by sea otters is followed by a recovery of the kelp forest, if not present already.

Conservation and Management

In addition to the suitable habitat described in the preceding section, the return of otters to the Bay is contingent upon the continued growth and expansion of the coastal population outside the Bay, the lack of oil pollution (oil destroys the insulating properties of sea otter pelage), and the availability of food. Other than the passage of time, as the population continues to expand, oil pollution, even low-level chronic pollution, is problematic for recolonization of the Bay by this species.

References

Broughton, J.M. 1999. Resource depression and intensification during the late holocene, San Francisco Bay: Evidence from the Emeryville shellmound
vertebrate fauna. Univ. of Ca. Press, Berkeley and Los Angeles, CA.


Important Data Sets
California Department of Fish and Game. Annual census of California sea otter population. Jack A. Ames, Monterey.
Harbor Seal
Phoca vitulina richards
William Z. Lidicker, Jr.
David G. Ainley

Life History

Harbor seals are the only marine mammals that are permanent residents in San Francisco Bay. During the recent past, California sea otters (Enhydra lutris nereis), known from numerous Indian middens, and harbor porpoises (Phocoena phocoena) were widespread in the Bay (Skinner 1962). Seasonally resident California sea lions (Zalophus californianus) use the Central Bay in the vicinity of the Golden Gate. See Ainley and Jones' accounts of the sea otter and sea lion.

Eight stomachs with food examined from individuals taken outside the Bay, but close to it, contained a variety of fish and a few octopus (Jones 1981). The most commonly taken fish were from the families Embiotocidae (surf perch) and Zoarcidae (eel pout). H arvey and T orok (1995) identified 14 species of fish and one cephalopod from 215 fecal samples collected from seals at seven haul-out sites around the Bay in 1991-92. Five species of fish made up more than 93% of the dietary weight, and one introduced species yellowfin goby (Acanthogobius flavimanus), constituted more than 54% of the total number of prey items found. Diet changed seasonally, with the goby and staghorn sculpin (Leptocottus armatus) predominating in the fall and winter, and plainfin midshipman (Porichthys notatus) and white croaker (Genyonemus lineatus), with jacksmelt (Atherinopsis californiensis) and the goby, predominating during spring and summer. Diet also differed regionally. In the extreme South Bay, the goby, sculpin, and croaker predominated; in the Central Bay, the mid-shipman predominated, comprising 91% of the diet. Curiously, no herring otoliths were found in the fecal samples.

An analysis of data from 59 radio-collared individuals revealed that frequency of diving (feeding) was greater at night (H arvey and T orok 1995). Pups are born in the spring, and a complete molt follows in early summer.

Historical and Modern Distribution

Harbor seals have been observed as far upstream as Sacramento, but little regular use is evident north of the Suisun Bay. Northerly sites are at Tubbs Island (Sonoma County) and Sister's Island (Marin). The haul-out sites associated with the Central Bay feeding area include (see Allen 1991, H arvey and T orok 1995): Sister’s Island in Suisun Bay. Northerly sites are at Tubbs Island (Sonoma County) and Sister’s Island (Marin). The haul-out sites associated with the Central Bay feeding area include: Coyote Point, Seal Slough, Belmont Slough, Bair Island, Corkscrew Slough, Greco Island, Ravenswood Point, Hayward Slough, Humbarton Point, Newark Slough, M owry Slough, Calaveras Point, Drawbridge, and Guadalupe Slough. Figure 5.8 shows the locations of known current haul-out sites plus a few potential sites.

Seals may pup at any haul-out site but, generally, pupping sites are more traditional and are the least disturbed of all sites used. During the early 1990s, there were eight known pupping sites around the Bay, although more than four pups were born at only three sites: Castro Rocks, Newark Slough, and M owry Slough (K opec and H arvey 1995). In 1992, there were a third again as many pups born at M owry Slough (67) than at all other sites combined (48; see also R iseborough et al. 1980). However, the previous year, there were only 39 at M owry Slough but double (26) the 1992 number at Castro Rocks. Current counts from 1999 include 243 seals (78 pups) at M owry Slough, 107 seals (21 pups) at Castro Rocks, and 72 seals (3 pups) at Yerba Buena Isl.
Historically, there was a large rookery near Alviso (Skinner 1962); this site most likely was Mowry Slough, given a description by Fancher (1979) (Allen, pers. comm.). If Skinner's Alviso site is not Mowry Slough, then the question becomes: did the seals move the few kilometers from "near Alviso" to Mowry Slough?

Censusing harbor seals is difficult, in the Bay or anywhere, because of the changes in numbers as a function of year, season, tide, time of day and human disturbance, and frequent movements of individuals throughout the Bay and even onto the adjacent outer coast. Nevertheless, numbers in the Bay apparently did not change significantly between 1982 and 1999 (Kopec and Harvey 1995, Green et al. 1999). A census taken in 1987 tallied 524 individuals in the Bay system (Hanen et al. 1988), and censuses in 1999 tallied 641 in the Bay (D. Green, pers. com). This population stability is in marked contrast to a steady increase during recent decades in the numbers of seals at sites along the coast, especially of Marin County (cf. Allen et al. 1989). Radio-tagged seals from San Francisco Bay have moved north to Point Reyes and south to Pescadero (Kopec and Harvey 1995). Allen (1991) believes that disturbance may be discouraging more seals from using the Bay and, thus, may be responsible for the lack of growth in the Bay. Pollutants may also be affecting the reproductive success of seals within the Bay (Kopec and Harvey 1995).

**Suitable Habitat**

Harbor seals feed in the deeper waters of the Bay. Kopec and Harvey (1995) identified two principal feeding areas. The first includes the area from the Golden Gate...
east to Treasure Island, northwest to the Tiburon Peninsula, and with a spur southward from Yerba Buena Island. Richardson’s Bay, which is adjacent to this area, has been used extensively for feeding in the past, although not as much at present; feeding in Richardson’s Bay may be contingent upon the presence of Pacific herring (Clupea pallasi; Allen 1991). This Central Bay feeding area is surrounded by nine haul-out and/or breeding sites (see above). The second major feeding area includes open Bay waters from the San Mateo Bridge southward. On the basis of the study by Kopec and Harvey (1995), this area is partitioned into five sub-areas, the largest being just west of Hayward. This South Bay feeding area is surrounded by 14 haul-out and/or breeding sites (see above).

Haul-outs must have gently sloping terrain, have deep water immediately nearby, and be free of disturbance, either by boats or by land (Allen 1991). An average of two haul-out sites are occupied by an individual seal each day, more so in the fall and winter and more so in the South Bay (Harvey and Torok 1995). Between-site movement is less frequent during the spring and summer, and is less frequent among seals in the deeper Central Bay. Haul-out sites used for pupping tend to be ones that are the most protected from disturbance. The use of such sites are persistent (traditional), and seals are slow to discover and utilize potential new pupping sites (Allen 1991).

Certain sites may be used as haul-outs at either low or high tide, e.g., seals appear to use Muzzi (Corte Madera) Marsh at high tide, but switch to Castro Rocks, 8 km away, at low tide (Allen 1991). In this case, Muzzi Marsh is separated from deep water by mud flats 2 km wide at low tide, but Castro Rocks are always surrounded by deep water.

Conservation and Management

The long association of harbor seals with humans (native Americans) in the Bay, including being actively hunted until about 1890, has made them extremely wary. They will flush from haul-out sites at 300 meters (Paulbitsky 1975, Allen et al. 1984). This makes them susceptible to harassment by persons on shore and boaters and kayakers from the Bay. Allen (1991) monitored the gradual abandonment of Strawberry Spit by seals during the 1970s and 1980s as a result of encroachment by humans and in spite of attempts for mitigation. An engineered haul-out site nearby has yet to be accepted by seals. Haul-out sites and especially pupping sites are needed that are protected from frequent human disturbances.

Allen (1991) also noted that harbor seals of the Central Bay reduced their use of winter haul-outs in and around Richardson’s Bay and used the ones farther south more frequently (i.e., Treasure and Yerba Buena islands) when the herring ceased spawning in Richardson’s Bay (and spawned more along the San Francisco waterfront). Therefore, the viability of certain prey populations may be important to the well-being of harbor seals in the Bay.

References


California Sea Lion
Zalophus californianus
David G. Ainley
Robert E. Jones

Life History

California sea lion is the pinniped ("seal") most often seen in zoos and circuses. In coastal waters of Central California, California sea lions prey mostly on schooling species, such as anchovies, Pacific whiting, midshipman and squid, as well as other fishes (Jones 1981). In the Bay, they feed mainly on anchovies, herring, surfperch, leopard sharks and spiny dogfish, and shrimp and crabs (Hanni and Long 1995). During the breeding season, bulls haul out at traditional breeding sites where they establish territories; females haul out to form groups, called harems, which each male tries to force onto his territory. In early summer (June), one pup is born to each adult female. Soon thereafter, the female mates with the harem master, but implantation is delayed for months while the mother nurses the pup. Pups form groups while their respective mothers forage at sea; some pups remain with their mother through the following winter (Jameson and Peeters 1988).

Historical and Modern Distribution

California sea lions occur along the West Coast of North America, from Vancouver to the Gulf of California; an isolated population exists on the Galapagos Islands and, formerly, another existed in Japan. The total population size of this species for the North American West Coast, as of 1990, was about 220,000; the population has been growing at about 10.2% per year since the early 1980s (Lowry et al. 1992). Population growth, following passage of the Marine Mammal Protection Act in 1972, is a recovery from former persecution.

Along the West Coast, the species breeds from Pt. Piedras Blancas, San Luis Obispo County, California,
south to the Gulf of California, Baja California, Mexico, although on rare occasions pups have been born at the Farallon Islands, San Francisco County; Ano Nuevo Island, San Mateo County; and the Monterey Breakwater, Monterey County. However, virtually all the animals that occur north of Pt. Piedras Blancas are adult and subadult males, and subadult females.

During the May-June pupping season, few adult California sea lions occur in Central and Northern California. Otherwise the species is present year round. In Central California, the largest numbers occur during the spring (April), when males that wintered in the north migrate south to breeding colonies; another smaller peak occurs in early autumn during the more leisurely migration north (Huber et al. 1981). During years of warm ocean temperatures (El Niño) much larger numbers of California sea lions visit Central and Northern California, including many more juveniles than usual (Huber et al. 1981).

In San Francisco Bay, California sea lions occur year round, but with a dynamic difference from that of the adjacent outer coast. Greatest numbers are present during the winter herring run (Dec-Feb; Hanni, pers. comm.). Following the winter peak, numbers decline to just a few animals by June and July. Numbers then increase gradually before a sudden increase in December. Known haul-out spots in San Francisco Bay are rare, and include only Pier 39, occasionally at Angel Island, and at Seal Rock just outside the Golden Gate (Figure 5.9). The largest number haul out at Pier 39, but the phe-
nomenon is only a recent one (unlike Seal Rock, which has been used by sea lions for at least the last 100 years; Sutro 1901). At first, in winter 1989-90, only a few individuals hauled out at Pier 39, but the next year they reached an average 500 (+100 SD) per day (February 1991); since then peak numbers during winter declined and now average about 200-300 animals per day (Marine Mammal Center, unpubl. data). The use of wharves at Pier 39 likely is the result of the following factors: 1) the increased size of the species’ total population (greater now than chronicled history; Lowry et al. 1992) and concomitant expansion of habitat use, 2) the construction of the wharves in the late 1980s, 3) the increasing temperatures of the California Current (which make Northern California more suitable for this species), and 4) the chance finding of this site by several individuals seeking food during the low food year of 1989-1990 (El Niño). These pioneers thus established a tradition among a group of sea lions.

Suitable Habitat

This animal uses those deep, principally marine waters that occur in the outer Bay, off Marin and San Francisco counties (e.g., Raccoon Straits to San Francisco and out through the Golden Gate). On occasion, isolated individuals, and mostly carcasses, have been found in Milpitas, Alameda, Napa, and as far upstream in the Delta as Sacramento. When salmon were netted en masse in the Delta 100 years ago, California sea lions were attracted in large numbers as far as Sacramento to take advantage of the netting operation (Sutro 1901), much as they do today in the case of herring in the Bay (and their infamous pillaging of steelhead held at the locks in Puget Sound, Washington). If free from disturbance, it is possible that sea lions would haul out on rocky peninsulas at places such as Angel Island and Alcatraz Island.

Conservation and Management

The presence of this species in San Francisco Bay is contingent upon the availability of safe haul-out sites and easily available food. Thus, its occurrence likely is tied to the fate of such fish as herring. The population in the Bay is sensitive to disturbance, to capture in gill nets (used illegally), and to certain diseases, such as leptospirosis, that result from the cattle that are grazing in coastal areas (and streams).

References


Personal Communications

Krista Hanni, The Marine Mammal Center, San Francisco

Important Data Sets

NOAA, Southwest Fisheries Science Center, La Jolla. Annual census of pinnipeds along the California coast. Jay Barlow.
Non-Native Predators: Norway Rat and Roof Rat

Rattus norvegicus and Rattus rattus
Andréée M. Breaux

Life History

Norway (or brown) rats and roof (or black) rats are similar in appearance, though the roof rat has a longer tail and can vary in color between brown and black. Norway rats have coarse fur, large naked ears, and scaly tails that are shorter than their body length (less than half of total length), pigmented venters, and tuberculate molars. Norway rats are largely nocturnal and are excellent swimmers and climbers. Roof rats have tails that are longer than their body length and pale venters, and they share the tuberculate molars that distinguish both Norway and roof rats from wood rats (Kurta 1995, Jameson and Peeters 1988).

Both the Norway and roof rats are prolific breeders, and the Norway rat tends to have slightly larger litters. Large males dominate female harems and the females actively defend resources and nest sites (Kurta 1995). Sexual maturity for both the Norway and roof rats is reached at about three to four months and breeding can occur continuously throughout the year. Litter sizes for Norway rats are between four and ten young, though litters as large as 22 young have been reported (Kurta 1995). Roof rat litters are generally between five and eight young (Jameson and Peeters 1988).

The Norway rat has been described as “the most unpleasant mammal in the world” (Jameson and Peeters 1988) as a result of its tendency to eat crops in the field as well as in storage, its tendency to eat both live and dead prey, and its ability to spread deadly diseases. It is capable of catching fish and small rodents (Kurta 1995). The Norway rat is an omnivore while the roof rat is a vegetarian.

Historical and Modern Distribution

Norway rats probably originated from the Old World tropics, but are now found globally. They are believed to have reached North America around 1775 (Kurta 1995) and are generally associated with buildings, sewers, harbors, garbage dumps, and agriculture. They reach elevations of 1,000 meters. Roof rats probably originated from the tropical Orient and can now be found in globally temperate climates. Roof rats are associated with trees, including agricultural groves, as well as with dense thickets and roofs and attics (Jameson and Peeters 1988).

The Norway and roof rats tend to occur in different habitats with the larger and presumably more powerful Norway rat occupying more urban areas, and the smaller roof rat living in more natural areas. Rats which are found in San Francisco Bay marshes are more likely to be roof than Norway rats (Jurek, pers. comm.). Where urbanization abuts natural marshes as it does in many areas of the South Bay, and garbage provides a food supply, Norway rats are likely to find the marsh habitats quite hospitable. In 1927, DeGroot noted that reclaimed land behind dikes along the San Francisco Bay shoreline was responsible for an increase in rats and a decrease in native California clapper rails (Rallus longirostris obsoletus): “No sooner is a dyke constructed than Norway rats appear in great numbers. Large gray fellows they are, on a dark night appearing to be as large as small cotton-tail rabbits....The Clapper Rail has no more deadly enemy than this sinister fellow” (DeGroot 1927).

In the Central Bay marshes, rats have been sighted at the Elsie Roemer Bird Sanctuary in Alameda, Crown Beach, the Martin Luther King Regional Shoreline and Arrowhead marsh sites, and on Brooks Island off the Richmond Habor (DiDonato, pers. comm.).

Rats are not regarded as a serious problem in the North Bay marshes, except in the Corte Madera marsh where there is inadequate buffering of the marsh from urbanization. Elsewhere, the extensive agricultural lands are probably preferred as habitat by the rats over the wetter marshlands (Botti, pers. comm.).

Suitable Habitat

Not addressed for non-native predators.

Control and Management

A 1992 report on the status of wildlife in the San Francisco Bay stated that there existed a “critical need” for research on the population dynamics and distributions of introduced mammalian predators such as the red fox, the Norway rat, and the roof rat (USFWS 1992). The report stated that techniques such as the reintroduction of the coyote to control the red fox in the South Bay, should be investigated. Control of rats has not been
implemented and continues to be a problem in the South Bay for endangered species, such as clapper rails and, quite possibly, salt marsh harvest mice (Reithrodontomys raviventris). Additional threats to other target species selected by this project as representative of wetland species in the San Francisco Bay region (e.g., California voles (Microtus californicus), ornate shrews (Sorex ornatus californicus), salt marsh wandering shrews (Sorex vagrans haliocoetes), and amphibians, reptiles, terrestrial invertebrates in general, and some ground nesting birds) probably occur.

Studies of South Bay marshes have documented predation of not only clapper rail eggs, but also of live chicks. While the primary predators may be raccoons (Procyon lotor), red foxes (Vulpes vulpes regalis), feral dogs, or feral cats, rats have been seen in the South Bay in relatively large numbers (Foerster et al. 1990; Albertson, pers. comm.; Harding, pers. comm.). Harvey (1988), in a study of clapper rails in three south San Francisco Bay marshes, attributed 24 percent of nest failures to Norway rats. A 1992 U.S. Fish and Wildlife study of hatching success and predation for 54 active clapper rail nests in south San Francisco Bay found rodents to be responsible for 90% of the eggs destroyed and 79% of the predation at monitored nests. Rodents were thought to be the predators because of the characteristic debris left behind after feeding, in this case egg shells, egg contents, and chick body parts. Other characteristics peculiar to rodent predators is the manner of leaving half of the egg shell intact with visible tooth marks, or a U-shaped notch eaten into the side of the shell (USFWS 1992 and 1997).

Negative impacts on native mammalian populations from rats in marshes include direct predation by the omnivorous Norway rat, competition for habitat, and illness or mortality resulting from diseases. While the devastation to humans from rats carrying plague, typhus, hepatitis, and trichina worms has been known for centuries in some cases (Jameson and Peeters 1988), the devastation to native wildlife is not as well-documented.

Clearly there is a need to implement the research suggested in 1992 on the distributions and population dynamics of Norway and roof rats in San Francisco Bay wetlands. While feral cats may help control the young rat populations, cats do not tend to eat the large adult rats (Jurek, pers. comm.), and the feral cats and dogs themselves are likely to prey on the small native mammals, amphibians, reptiles, and terrestrial invertebrates that are indigenous to the wetlands. Control measures are difficult, since there is no poison specific to rats that is safe for endangered mammals, such as the salt marsh harvest mouse. Given the difficulties in any control programs (e.g., public outcry against removing feral cats and the difficulty of trapping or shooting these large rodents) the most effective control measure at this time is to protect marshes with large buffers, and to keep shelter and garbage far from the wetland edge.

References


Personal Communications


Fred Botti, California Department of Fish and Game, Yountville.

Joe DiDonato, East Bay Regional Park District, Oakland.

Elaine Harding, University of California, Santa Cruz.

Ron Jurek, California Department of Fish and Game, Endangered Species Division, Sacramento.
Non-Native Predator: Red Fox

Vulpes vulpes regalis
Elaine K. Harding

Life History

The red fox is one of the most widely distributed mammals in the world, occupying a range of habitats and elevations. There are actually two red fox subspecies in California, the introduced red fox, *Vulpes vulpes regalis*, and the native, state threatened, red fox, *Vulpes vulpes necator*, which is found only in the Sierra Nevada from 5,000 to 8,400 ft (Jurek 1992). Red fox have a head and body length of about 45.5-90.0 cm and a tail length of 30.0-55.5 cm, with a weight averaging 3-14 kg (Nowak 1991). The pelage coloration is generally pale yellowish red to deep red on the upper parts and white or ashy underneath. It can be distinguished from the gray fox by its black lower legs and white tip on the tail.

Red fox family groups usually consist of a male, a female and offspring, with the territories ranging in size from 2.6 to 20.8 square kilometers (Sargeant 1972). However, urban populations may achieve even greater densities, with home ranges as small as 0.45 sq. km in Great Britain (Trewhella et al. 1988). Den sites in urban areas are often within flood control levees, freeway embankments (Sallee et al. 1992) or salt marsh levees.

Breeding occurs from December through April, with a peak in March, with the number of offspring produced (and surviving to juvenile age) averaging 3.5 (Storm 1976). Multiple dens may be used during this time, with the females often moving the litters to different locations throughout the season. The survival of juveniles to recruitment is estimated at 0.19 in midwestern populations (Storm et al. 1976) to 0.65 in southern California populations (Sallee et al. 1992). Foxes have been reported to breed at 10 months, with yearlings often breeding their first spring. Additionally, the survival of adults in urban California populations is 0.58, higher than the midwest at 0.23 (Storm et al. 1976). Adults may survive up to five years in the wild.

Red fox have extremely broad diets, including birds, small mammals, reptiles, amphibians, insects, vegetation and refuse (Foerster and Takekawa 1991). They are also known to be surplus killers, where food that is taken may be cached (buried in the ground) and never recovered. Because they are such capable predators, they are highly detrimental to native fauna which are not adapted to avoid or escape them. Foxes are known to decimate ground nesting bird populations, through predation of eggs, young and adults.

Historical and Modern Distribution

The red fox subspecies, *V. v. regalis*, is originally from the Great Plains, and was probably brought to the Central Valley for commercial fur farming in the late 1800s (Jurek 1992). The current distribution of fox throughout the state is based upon a study by Sallee, et al. (1992) which found occurrences of red fox in 36 counties. The greatest concentration of sightings were in the urban areas of Los Angeles and San Francisco Bay, with fox also found throughout the Central Valley and Monterey Bay areas. It is difficult to estimate the number of fox in California, but according to records kept by Animal Control in Orange County, there were 102 individuals in the county during the summer of 1991 (Sallee et al. 1992).

The red fox was first seen in South San Francisco Bay in 1986 (Foerster and Takekawa 1991), with subsequent sightings reported from all seven Bay Area counties (Sallee et al. 1992). Populations of red fox have established in or adjacent to tidal marshes, diked baylands, salt ponds, landfills, agricultural lands, golf courses, grasslands and urban areas. In particular, the fragmented wetlands of San Francisco Bay have become a likely source for expanding populations, as many avian and mammalian prey can be found within these habitats.

Suitable Habitat

Not addressed for non-native predators.

Control and Management

In the San Francisco Bay Area, red fox have been implicated in the population declines of the endangered California clapper rail, threatened western snowy plover, endangered California least tern, Caspian tern, and colonial nesting species, such as great blue herons and great egrets (Foerster and Takekawa 1991; Albertson 1995; USFWS, unpublish data). In response to growing evidence of the impact of red fox on the clapper rail, the U.S. Fish and Wildlife Service began a Predator Management Program in 1991 (Foerster and Takekawa 1991). The subsequent removal of red fox and other targeted predators has re-
sulted in a significant increase in the local populations of California clapper rail (Harding-Smith 1994, Harding et al. 1998). Comparable success has occurred in Southern California where removal of red fox along coastal marshes was correlated with remarkable increases in the populations of light-footed clapper rails (USFWS and U.S. Navy 1990). Additionally, predator management is becoming a common method of endangered species protection, within both government (Parker and Takekawa 1993) and private sectors.

It is imperative that all future restoration and management activities within the wetland ecosystems of San Francisco Bay consider the present and future impacts of red fox on the native wildlife. Clearly, a healthy marsh can no longer be defined by simply the quantity and composition of native vegetation and wildlife, but must include the external impacts of human urbanization which alter an ecosystem’s internal functioning through the introduction of contaminants, human disturbance and non-native species. The long-term viability of many avian and small mammal species will be impacted by expanding red fox populations in the bay area, so that no site will soon remain uninhabited nor unaffected by this wily species. Therefore, the quality and quantity of connections between sites, as well as the characteristics and extent of the surrounding matrix, will be of the utmost importance in understanding fox population dynamics and prey abundance within and between sites. Biologists and land managers must continue to study the dispersal patterns, demographics and predation impacts of red fox so that more effective methods of control can be developed.

References


Introduction

The tule greater white-fronted goose or tule goose was selected to represent the geese and swans group which also includes Pacific greater white-fronted goose (Anser albifrons frontalis), Canada goose (Branta canadensis), Aleutian Canada goose (B.c. leucopareia), lesser snow goose (Chen caerulescens), and tundra swan (Cygnus columbianus).

Tule geese are primarily associated with managed wetlands and agriculture lands. The Suisun subregion is one of only a few important wintering areas in California. The geese/swan group is of economic and recreational importance as four of the six members of this group are hunted.

Although populations are relatively low in the San Francisco Bay Area for all species in this group, at least one representative is found in all subregions designated for the Goals process. Tule geese are primarily found in Suisun Marsh and North Bay (Napa Marsh); Pacific white-fronted geese, Suisun and North Bay; Canada geese, all subregions; Aleutian geese, Suisun and Central Bay; snow geese, all subregions; and tundra swans, Suisun and North Bay.

Description

The tule goose is one of two subspecies of greater white-fronted geese that breed in Alaska and winter primarily in California (Swarth and Bryant 1917); the Pacific greater white-fronted goose is the other subspecies. Populations of the Pacific goose are far greater than the tule goose. The overall size of the tule goose is generally larger than the Pacific goose, although there may be overlap between the subspecies. The tule goose has a length to 34 inches (86 cm) and wing span to 65½ inches (167 cm) (Cogswell 1977). They are a medium-sized dark goose with the brown colors of the head and neck the same as the body and wings. Tule geese are a much darker brown than Pacifics. Adults have a white forehead and black blotched belly. In most tules, the forehead may show some orange coloration. Their feet are yellow and their bills are pink to orangish in color. Immatures do not have black blotched bellies.

Breeding Biology – Historically, nesting was known to occur at Redoubt Bay, in Cook Inlet, and suspected at Susitna Flats, Tuxedni Bay, Chinitna Bay, and Innoko National Wildlife Refuge (NWR), all in Alaska (Timm et al. 1982). Telemetry studies in 1995 showed breeding taking place on the Kahiltna and Yentna River Valleys northwest of Anchorage. Both areas were previously unreported as areas for tule geese. No radioed birds were found in the Redoubt or Trading Bay areas. The Redoubt Volcano eruption of 1989 may have made the area unfit for tule goose breeding. Few white-fronted geese have been counted in the area in the last five years. Nest initiation generally begins in early to mid-May or later depending on thaw on the breeding grounds. Incubation is 24-26 days. Clutch size averages 5-6 eggs (Timm et al. 1982, Zeiner et al. 1990). The subspecies is monogamous, solitary nester, with both parents tending the young. Breeding may occur at two years of age, but three years is more common (Bellrose 1980).

Migration Ecology – Tule geese begin to leave Alaska by mid-August. By September 1 of 1980 and 1981 only a few hundred remained in Redoubt Bay or Susitna Flats (Timm et al. 1982). Generally, tule geese fly over the open ocean from Alaska to key staging areas
in southeastern Oregon, i.e., Summer Lake Wildlife Management Area and Malheur NWR. Up to 50% of the population may be present at these Oregon sites by early September and approximately 1,000-2,000 birds may remain until late October (Mensik 1991). Birds are also present in the Klamath Basin of Oregon and California during these times (Wege 1984). The remaining 50% of the population over-fly the fall staging areas arriving at Sacramento NWR in early September (Timm et al. 1982, Mensik 1991). Field observations at Grizzly Island Wildlife Area in the Suisun Marsh during the mid-1980s showed the first tule geese arriving during the September 9 to September 16 period (CDFG, unpubl. data). Historically, the primary spring migration staging area was the Klamath Basin where numbers peaked in late March. Results of radio telemetry studies in 1994-95 showed that few tule geese used Lower Klamath NWR while Summer Lake, Chowauncan Marsh, and the Warner Valley, all in Oregon, provided the principal spring staging areas (USDI 1995). Tule geese begin leaving central California in February (Mensik 1991). By mid-February of 1989 more than 1,400 tule geese remained at the Grizzly Island Wildlife Area (CDFG, unpubl. data).

**Wintering Ecology** - Primary wintering areas in California are the Sacramento Valley and Suisun Marsh. A small number of birds use the Napa Marsh. Use is mostly at Sacramento and Delevan NWRs and adjacent rice fields. Colusa NWR also receives some use. By October and November, an estimated 90% of the tule goose population occurs in these areas (Mensik 1991). Generally, there is a winter-long interchange of geese between the three most important areas, Sacramento NWR, Delevan NWR, and Grizzly Island Wildlife Area, although for the 1995 mark-recapture survey no tule geese were observed at Grizzly Island Wildlife Area during the September ground counts (Trost and Harb 1995). Other areas where tule geese have been observed in the past include the Butte Sink, Sutter NWR, San Joaquin Valley, and Sinaloa, Mexico (Wege 1984; Ely and Takekawa 1990; Kramer, pers. comm.; Timm et al. 1982; PFSC 1991).

**Distribution and Abundance**

**North America** - In North America, tule geese have been documented in the Central Flyway, although their status there is uncertain and there are no population estimates. A specimen of this subspecies of greater white-fronted goose was collected in Texas in 1852 by Hartlaub and subsequently described as one of the greater white-fronted goose (Swarth and Bryant 1917). The bird is primarily found in the Pacific Flyway where most of the studies to locate nesting, migration, and wintering areas have been done.

**Pacific Coast** - Nesting is known to occur at Redoubt Bay and Susitna Flats in the Cook Inlet, Alaska. Nesting also takes place northwest of Anchorage, Alaska in the Kasiltna and Yentna River Valleys. Ongoing telemetry studies are attempting to document additional breeding areas. Malheur NWR and Summer Lake Wildlife Management Area, along with Klamath Basin NWRs in Oregon and California, are the most important fall and spring migration stopover sites. Sacramento and Delevan NWRs and Grizzly Island Wildlife Area in the Suisun Marsh are the major wintering areas. Other areas important to winter birds are the Butte Sink, Colusa NWR, and the Napa Marsh near where the Napa River enters San Pablo Bay.

**San Francisco Bay** - Locally, the tule goose is only found in the Suisun and North Bay subregions. Suisun Marsh is the third most important wintering area in California. In the North Bay Region (Napa Marsh), a small population uses the marshes, sloughs, and adjacent agricultural lands (Figure 6.1).

**Suisun** - Suisun Marsh is the third most important wintering area in California. The peak population index of 1,500 was in December 1980 at Grizzly Island Wildlife Area in the Suisun Marsh. Other high indices were in December 1978 (1,000), December 1981 (1,200), February 1989 (1,229), and February 1990 (1,190) (Mensik 1991). The mid-winter waterfowl survey of January 1991 showed 1,527 tule geese. Waterfowl surveys during the period October 1992 through January 1997 showed the tule goose numbers to be generally less than 500 birds (CDFG, unpubl. data).

**North Bay** - The Napa Marsh is of the North Bay region support a small wintering population of tule geese. Reports from duck hunters and hunting club owners are that the peak wintering population is less than 50 birds.

**Central Bay** - Not present.

**South Bay** - Not present.

**Historical Information**

Since the late 1960s, the following information has been gathered for tule geese on an irregular basis: population size and distribution, including fall and winter counts of national wildlife refuges and state-managed wildlife areas, and periodic leg banding and color marking; production assessment, including age composition and family size counts on staging and wintering areas; and mortality assessment and harvest management, including monitoring harvest on selected public hunting areas and disease mortality on national wildlife refuges and state-managed wildlife areas.

**Population Trends**

Information has been gathered sporadically on tule geese in conjunction with other projects since the late 1960s. There has been more intensive study of the tule goose during the last 20 years. Table 6.1 shows population
trend data for selected years between 1978-79 to 1989-90. The peak index of 8,615 was in September 1989. Observations of radioed and neck-collared tule geese in the fall and winter of 1995 (September 5 - December 31, 1995) documented approximately 6,000 birds in California and southern Oregon (Trost and Harb 1995).

**Habitat Use and Behavior**

**Foraging** - Foraging on the breeding grounds occurs on grasses, sedges, and aquatic plants in the intertidal mudflats, freshwater marshes, or poorly drained areas characteristic of the region. They are primarily grazers, but they will grub for roots and shoots (Zeiner et al. 1990). In the fall at Klamath Basin, they feed in ponds with alkali bulrush (Scirpus robustus) or harvested grain fields. Roosting occurs in open water ponds (Wege 1980). During early fall in the Sacramento Valley, they feed in harvested rice fields then shift to winter flooded uplands. Marsh units on Sacramento and Delevan NWRs with an abundance of alkali bulrush and with some open water are also used for feeding, which continues until departure in February (Wege 1980, Timm et al. 1982).

**Roosting** - Roosting and loafing generally occurs in open water ponds with emergents such as bulrush (Scirpus spp.) and cattails (Typha spp.). In the Suisun Marsh tule geese feed in ponds with alkali bulrush or in the barley/grass uplands of the sanctuary on Grizzly Island Wildlife Area. Roosting areas have shallowly flooded uplands with a grass-pickleweed (Salicornia spp.) mixture. These areas are in the closed zone to hunting. Tule geese observed feeding in the Napa Marsh were found in tidal areas fringed by emergent cattails, tules, alkali bulrush, and cordgrass (Spartina spp.) with pickleweed and Grindelia spp. in the high areas. Two tule geese taken during hunting season in December 1954 were analyzed for food habits. Results showed alkali bulrush tuber and rhizome fragments in both with forb leafage and insect fragments (Longhurst 1955).

The managed wetlands of the Suisun Marsh are the most important habitat for tule geese in the San Francisco Bay ecosystem. These wetlands, managed for alkali bulrush and other wetland wildlife food plants are critical as feeding and roosting areas as they will feed primarily on tubers and rhizomes of alkali bulrush.

**Movement** - Generally, studies of daily movement of tule geese in the winter have shown that several subflocks exist (groups with specific roost sites and movement patterns). Subflocks were readily identified utilizing Sacramento NWR during studies in 1979-80 (Wege 1980). This is also probably true in other wintering areas. Birds move between Sacramento and Delevan NWRs, Grizzly Island Wildlife Area, and in the Delta. These same studies indicated that two daily feeding flights were common. At Grizzly Island Wildlife Area, short flights were made from roosting ponds to barley fields (Wege 1980).

During waterfowl hunting seasons (late October to late January), tule geese may develop different movement patterns due to the disturbance. In the Sacramento Valley, after opening of hunting season, tule geese shift to off-refuge harvested rice fields and to closed portions of...
refuges (Timm et al. 1982). When not disturbed they generally feed early mornings and late afternoons. During hunting seasons there may be some night feeding (Cogswell 1977).

**Conservation and Management**

Since the late 1960s, information on tule geese has been gathered on population size and distribution, production, mortality, and harvest. In addition, habitat management and protection practices have been implemented that have included classifying areas as refuge or critical habitat, zoning laws to protect lands important to tule geese, enhancement of state, federal and private lands by controlled burning, grain farming, or marsh management, and using federal easements to provide incentives to private landowners to retain wetlands.

**Contaminant Risks** - No information.

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* Indices reflect trends and not absolute numbers.
† Summarized data represent contributions from cooperators of the Pacific Flyway Study Subcommittee on the Pacific Flyway Population of Tule Greater White-Fronted Geese (principal author G. Menzik). Peak monthly counts include geese recorded in immediate vicinity of indicated survey areas.
‡ Includes Sacramento NWR, Delevan NWR, and Colusa NWR.
Disturbance - Most Pacific Flyway tule greater white-fronted goose harvest occurs in California. Despite limited harvest information from band returns or hunter check stations, it appears harvest locations for tule greater white-fronted geese are similar to those for Pacific white-fronted geese. The 1979-1982 and 1987-1989 sport harvest of tule greater white-fronted geese on the Sacramento NWR Complex indicate that: (1) tule geese comprise a disproportionately high percentage of the harvest (30-60%) when compared to population composition (5-25%); (2) the majority of the harvest comes from Delevan NWR and adjacent areas; (3) harvest age ratios for tule geese (30%-40% young) more closely reflect those of the population than do those of Pacific white-fronted geese (70%-80% young).

Habitat management recommendations include protecting current and future breeding areas as “critical habitat”. Acquisition or easement of habitat areas in California and Oregon not currently under state or federal management is recommended. Management practices on state, federal, and private lands beneficial to tule geese should be maintained.

Within the San Francisco Bay Area, the areas of greatest importance to the tule goose are Suisun and the North Bay. Important habitat elements are open water, perennial and seasonal pond, high tidal ecotone (in Napa Marsh), and emergent vegetation. Fringe marsh along sloughs is also important. Regionwide, goals to support this species should include maintaining current acreages of managed marsh, managed upland habitat, and farmed upland areas that are farmed for oat hay.

Suisun - Suisun is the most important subregion for all goose species. To maintain current population levels, we need to maintain their habitat - managed wetlands and associated upland habitat.

North Bay - Tule geese do not nest in this Estuary, but an associated species, the Canada goose, nests in the Napa Marsh of the North Bay. Currently, the North Bay has a relatively small wintering population of tule geese and Pacific greater white-fronted geese, but they seem to be adjusting to the habitat gains recently made in the area (Petaluma, etc.). There is potential to increase goose populations with a further increase in managed marsh habitat. Areas of importance in the North Bay include the Napa Marsh from Sears Point to Napa River, Salt Ponds 1A and 1AN, all managed marsh areas, seasonal ponds in agricultural areas, farmed uplands, and the high tidal ecotone. Current acreages of farmed and managed upland habitat should be maintained.

Central Bay - Tule geese do not currently use this subregion in great numbers, but an increase in habitat (managed marsh and managed upland habitat) could increase the subregional population. The San Pablo Reservoir is important for Aluetian Canada geese.

South Bay - Tule geese do not currently use this subregion in great numbers, but an increase in habitat (managed marsh and managed upland habitat) could increase the subregional population.

Research Needs

Comprehensive research occurred during the late 1970s and early 1980s. Work was conducted on wintering, migration stopover, and the newly discovered breeding areas. Activities included leg banding, neck collaring, and outfitting individuals with radio transmitters. Daily and seasonal movements were monitored, sport harvest documented, and social behavior observed (Timm et al. 1982, Wige 1984).

Future research should address several aspects of the tule greater white-fronted goose ecology. Winter habitat requirements would help land managers develop strategies to protect and enhance wetlands for tule geese. Additional data is needed to develop techniques for subspecific identification. There is relatively good information on sport hunting mortality, however there is a distinct lack of data on non-sport hunting mortality due to disease, predation, subsistence hunting, and pollution. Continued improvement of fall/winter surveys will aid in obtaining concurrent peak population counts and age ratio samples on all known use areas.

Additional needs center around data gaps that are intensified by the small population size of tule geese and their physical similarity to Pacific greater white-fronted geese. Regularly scheduled surveys are needed to help answer the population status question. A better understanding of the taxonomic differences between Pacific

Recommendations

The Pacific Flyway Management Plan for tule geese contains harvest guidelines by region based on population indices. Strategies are based on the objective to maintain stable populations. These basic strategies recommend liberal harvest allowances with the population above 10,000, and with more restrictive regulations until the population index reaches 3,200 when the season would be closed. These strategies are primarily for Sacramento and Delevan NWRs and Grizzly Island Wildlife Area.

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greater white-fronted geese and tule greater white-fronted geese is needed. Estimates of production, survival, and mortality parameters are incomplete. Past research needs to be completed with analyzed and published results. Effects of some agricultural land use practices are not known. Habitat requirements are not fully delineated. There is sport harvest occurring outside of California but the magnitude and location of harvest, and thus complete wintering population size, is also unknown. (Mensik 1991).

Acknowledgments

John Takekawa of the U.S. Geological Survey, Vallejo and Greg Mensik, U.S. Fish and Wildlife Service, Sacramento National Wildlife Refuge, are the California experts on the Tule goose and their input on this report is appreciated.

References


Other Important References


Personal Communications

Mallard
Anas platyrhynchos
Steven C. Chappell
David C. Van Baren

Introduction

The mallard was selected as a representative of other dabbling ducks such as Cinnamon teal (Anas cyanoptera) and Gadwall (Anas strepera) which are found in the Suisun Marsh and the San Francisco Bay Area. All three of these species represent resident breeding populations in the San Francisco Bay Estuary, as well as migrational wintering populations from the northern breeding grounds.

The largest population of mallards occur in the Suisun subregion. Mallards were also recorded as the number one dabbling duck of the San Pablo Bay and South San Francisco Bay subregions, most often using seasonal wetlands habitats and low salinity salt ponds. The lowest numbers of mallards were recorded in the Central Bay subregion, with few mallards being recorded in the open bay habitats of all four subregions.

Description

The mallard is one of the most easily recognizable of all waterfowl species. The drake is characterized by a bright yellow bill, brilliant green head, and brown chest with a white neck ring separating the two. The drakes also have a gray body with central black tail feathers curling upward. Both sexes have white outer tail coverts, with a blue speculum bordered in white, and bright orange feet. The female is the typical mottled brown of other Anas species, and has an orange bill with a dark spot on top. Mallards are among the most vocal of all duck species. The hen mallard has a call which begins with a loud quack followed by a series of slowly diminishing quacks. The drake mallard, by comparison, has a very soft almost buzzing call. Adult male mallards typically average 24.7 inches in length and weigh 2.75 pounds, while the female tends to be a little smaller at an average of 23.1 inches and weigh 2.44 pounds (Bellrose 1980).

Breeding Biology - Mallards have one of the most widespread breeding ranges of all waterfowl species, encompassing both Canada and the United States. Loose pair bonds begin to form as early as August (Barclay 1970) with nesting beginning by early April. The pair bond generally begins to weaken with the onset of incubation, rarely lasting until pipping (pre-emergence). Nesting typically occurs on the ground, in upland fields generally in stands of dense vegetation. The nest is a shallow depression in the vegetation that is lined with down and feathers plucked from the females breast. The typical clutch size is from 7-10 eggs, but can be as high as 15. Females incubate the eggs for approximately 28 days, and are the primary care provider for the ducklings which are precocial at hatching, and move about the nest in a few hours (Batt et al. 1992).

Migration Ecology - Migration occurs along four different flyways, with the heaviest used corridors being the Mississippi Flyway in the East, and the Pacific Flyway in the West (Bellrose 1980). Birds migrating to northern breeding grounds depart the wintering areas by early February, returning as food availability becomes scarce, and arriving back in the wintering grounds by early October. The most important migration corridor in the West appears to be from Alberta to the Columbia River basin with several different routes going into the Central Valley of California (Bellrose 1980).

Wintering Ecology - Mallards primarily winter throughout the United States and along the west coast of Canada, with the Atlantic flyway attracting relatively few numbers of birds (Bellrose 1980). Suisun, San Francisco Bay, the Sacramento-San Joaquin Delta, and the Central Valley are important wintering areas for the mallards in the Pacific flyway. These areas also provide important stop-over locations for mallards migrating to and from the wintering and breeding grounds.

Distribution and Abundance

North America - Mallards are the most widely distributed species of waterfowl in North America, and are found virtually everywhere in high numbers except for the Atlantic Flyway. During the 1996-1997 waterfowl season, mallard numbers in the Suisun Marsh fluctuated from a high of 29,580 on October 16, 1996, to a low of 6,105 on January 8, 1997 (CDFG 1997). Some of the primary factors influencing mallard distribution in the San Francisco Bay Estuary is the availability of areas with low salinity water, and the necessary food resources. Accurso (1992) found that the mallards were using the salt ponds in the North and South bays at 2-3 times the expected rate based on availability. These ponds had a salinity level of around 20-33 ppt.

Pacific Coast - No information.
San Francisco Bay - In the Diked Baylands Wildlife Study (USFWS, in prep.), mallards were identified as the dabbling duck using seasonal wetland habitats most often in the greatest numbers for both the North and South bays. Figure 6.2 shows the distribution of mallards around the Bay.

Historical Information
No information.

Population Trends
Since 1960 the continental population of mallards has fluctuated widely from an overall high in 1970 of 9,986,000 birds to a low of 4,960,000 birds in 1985. During the ten year period since 1985, however, the trend in mallard numbers has been an increase to a high of 8,269,000 birds in 1995 (CDFG 1953-1997 and 1997). In 1996 and 1997, there has been a decrease in mallard numbers down to 7,643,000 (CDFG 1997). The overall Suisun Marsh mallard population has fluctuated widely since 1960 with a high of 88,885 mallards to a low of 10,876 mallards. Several years have large peaks in the total number of mallards using the marsh, which could indicate the arrival of migrants from the northern breeding grounds.

Habitat Use and Behavior
Foraging - Mallards are very opportunistic in their foraging behavior. They will feed on both natural food...
plants, as well as agricultural waste grains while on the wintering grounds. The primary natural foods eaten by waterfowl in the Suisun Marsh are alkali bulrush, fathen, brass buttons, watergrass, and smart weed (Rollins 1981). Aquatic invertebrates play an important role in mallard diets prior to and during the breeding season, due to the high energy demands of the hen for egg laying.

**Breeding** - Nest site selection begins once the flock reaches the breeding grounds, with the pairs breaking off from the flock and setting up independent home ranges. The home range will typically include one or more loafing sites consisting of bare shore surrounded by tall standing vegetation near water (Bellrose 1980). Mallards use a wide variety of vegetation types in the construction of their nests. In the Suisun Marsh the primary vegetation used is annual rye grass, lana vetch, brome, and tall wheat grass, as well as natural wetland plants within the managed wetlands. The main nesting requirement appears to be that the vegetation is dense and approximately 24 inches tall.

McLandress et al. (1996) found that the mallard nest densities in the Suisun Marsh, Central Valley, and the intermountain region of Northeastern California were higher than in the prairie breeding grounds in Canada. Canadian nest densities were found to average 10.6 mallard nests/km². In California mean nest densities ranged from 41 nests/km² in the San Joaquin Valley to 190 nests/km² in the Suisun Marsh.

Yarris et al. (1994) determined, using radiotelemetry, that some hens nesting at Grizzly Island Wildlife Area will leave the area after fledging their young, and prior to their wing molt. Yarris detected radio-tagged hens in the Delta east of Grizzly Island Wildlife Area and as far north as the southern Oregon border, locating the molting areas of 20 hens. All hens radio-tagged were detected moving in a northerly direction shortly after leaving the Suisun Marsh. Mallard ducklings are very sensitive to increasing salinity levels. Mitchum and Webster (1988) found that water with a specific conductivity of 20 mS/cm is lethal to mallard ducklings, and Wobster (1988) found that water with a specific conductivity of 190 mS/cm is lethal to mallard ducklings.

**Movement** - No information.

**Conservation and Management**

**Contaminants** - Disease and contaminants are directly responsible for a large amount of the waterfowl mortality each year, although the total number is unknown. Diseases such as botulism, cholera and duck viral enteritis historically have not been a major concern for duck populations in the Suisun Marsh. The only major event has occurred in this region during the winter of 1948-49 when approximately 40,000 ducks, geese, and coots died in an avian choleran outbreak in the San Francisco Bay Area (Bellrose 1980). As with all places that have historically used lead shot, there is still a possibility of incidental mortality due to lead poisoning. The danger of this disease is not that there is a large noticeable die-off every year, it is that no one knows how many individual birds ingest lead and die without ever being noticed. Bellrose (1959) concluded that 2 to 3 percent of the fall and winter waterfowl populations may fall victim to lead poisoning each year. With the current use of non-toxic shot loads in waterfowl hunting, this number should gradually become smaller as shot is covered by sediment and becomes unreachable.

The maintenance of good wintering and breeding habitat in the Suisun Marsh and Napa Marsh is important to the continued use of the San Francisco Bay Estuary. This can be accomplished by the protection of seasonal wetlands and the intensive management of diked managed wetland areas.

**Disturbance** - Disturbance from human activities can cause temporary changes in behavior and locally affect temporal and spatial distribution of migratory and wintering waterfowl (Madsen 1994). Disturbance by humans caused both longer duration of alert and flight behavior by pintail when compared to disturbance by raptors or other animals (Wolder 1993). Considering these disturbance impacts could be similar to mallards, activities such as wildlife viewing, urbanization, and vehicle traffic may have negative effects.

**Recommendations**

Managed wetlands are critical habitat for both resident breeding birds and for migrants, as they provide food resources and wintering habitat. Within the region, the most important areas for mallards are the managed wetlands of Suisun Marsh, some less saline areas of San Pablo Bay, and seasonal wetlands habitat around the San Francisco Bay. To support mallards, regional goals should strive to increase the acreage of managed marsh habitat; maintain or enhance current areas of lagoon (loafing and feeding habitat) and farmed baylands (critical wintering habitat); and maintain diked marsh (especially in brackish areas), ruderal baylands (breeding and nesting habitat), low salinity salt ponds, and treatment ponds. Adjacent to the baylands, grazed and managed uplands should be maintained and increased as critical breeding habitat, and riparian habitat should be improved and increased.

Important habitat elements for mallards include seasonal ponds (most critical for food production for wintering birds); perennial pond (wintering and breeding habitat, foraging); water column/open water (loafing); clay-silt substrate (foraging); mudflats (limited use); veg-
etated levees and islets (nesting); eelgrass (very minor on the open bay); pan (if brackish, important for wintering, foraging, breeding); emergent vegetation; riparian zone community; vernal pools (fresh water); and artificial vernal pools.

**Suisun** – The managed marshes of Suisun are the most important habitat for mallards in the San Francisco Bay. Riparian habitat is also important. Specific habitat goals for the Suisun subregion include increasing the acreage of managed marsh, diked marsh, ruderal baylands, and managed uplands; maintaining and enhancing farmed and grazed baylands; and maintaining grazed upland and riparian areas. In this subregion salinity levels preclude increasing riparian zones.

**North Bay** – Areas of particular importance in this subregion are the managed wetlands of the Napa River area and low salinity salt ponds. Subregional habitat goals include increasing the acreage of managed marsh, lagoon, and low salinity salt ponds; maintaining and enhancing farmed and grazed baylands; and maintaining diked marsh and ruderal baylands. Adjacent to the baylands, managed uplands and riparian zones should be increased and enhanced.

**Central Bay** – The Central Bay does not currently have a large population of mallards; to increase this population, critical mallard habitats (managed marsh and upland) should be increased. Areas of particular importance within this subregion are lagoons and the wetlands near Marin. Habitat goals for the Central Bay include increasing the acreage of managed marsh and lagoon, and maintaining areas of diked marsh and ruderal baylands. Adjacent to the baylands, managed uplands and riparian zones should be increased and enhanced.

**South Bay** – Areas of particular importance within the South Bay are managed wetlands, low salinity salt ponds, and diked wetlands. Riparian zones also show some usage by mallards. Habitat goals for the South Bay subregion include increasing the acreage of managed and diked marshes; maintaining and enhancing farmed baylands; and maintaining ruderal baylands and low salinity salt ponds. Adjacent to the baylands, managed uplands and riparian zones should be increased and enhanced.

**References**


Northern Pintail
*Anas acuta*

Michael L. Casazza
Michael R. Miller

Introduction

The northern pintail has been historically the most common puddle duck wintering in the San Francisco Bay region. Continental population declines have been severe and the declines have been even greater within the San Francisco Bay region. This disproportionate decline in pintails using the San Francisco Bay region is alarming and needs further investigation. In particular, the Suisun Marsh has seen peak numbers decline as much as 90% over the past several decades (Figure 6.3). Pintails use a wide variety of habitat types throughout the region, including managed marsh, seasonal wetlands, open bay, and salt ponds. They utilize many of the habitats used by other waterfowl species. Species which are commonly found in similar habitats as pintail are green-wing teal (*A. crecca*), northern shoveler (*A. clypeata*), and American wigeon (*A. americana*). We have grouped these three species together with pintails, but the pintail will be the focus as the key species because it is relatively abundant in the San Francisco Bay region, and it uses many different habitat types, including managed wetlands, as critical wintering areas.

Description

The northern pintail is a long slender duck with narrow, angular wings. Pintails float high on the water, offering a very elegant appearance to the casual observer. They have sexually dimorphic plumage. Drakes in nuptial plumage have a chocolate brown head with a white breast and foreneck extending upward as a stripe on each side of the head. Their backs are greyish in appearance and they have two, long black tail feathers for which they are named. Male pintails have a distinctive iridescent black-green to green speculum. The male pintail has a distinct short whistle which is heard most during winter and spring. Hens pintails are mottled brown and have a noniridescent brown to brown-green speculum. The bills of both sexes are blue-gray in color with black along the central ridge in males and black blotches in females. Both sexes have gray legs and feet. The male pintail has a total length of between 57-76 cm, and females between 51-63 cm (Austin and Miller 1995).

Breeding Biology - Under favorable wetland conditions pintails will breed in their first year. They have been known to lay as few as three and as many as 14 eggs, but their average clutch size is about eight eggs (Bellrose 1980). Pintails have been found nesting across a vast area encompassing much of the Northern Hemisphere. Their main breeding areas in North America include the prairie pothole region of Alberta, Saskatchewan, Montana, and the Dakotas, along with the arctic regions of Canada and Alaska. In California, pintails nest on the northeastern plateau, the San Joaquin Valley, and on the coastal marshes, including Suisun Marsh and San Francisco Bay. Their nests tend to be in relatively open cover and can be as far as 3 km from water (Duncan 1987). Pintails are early nesters, some initiating their nests as early as late March, depending on weather conditions and location.

Migration Ecology - The major North American migration routes range from breeding areas in northern Alaska and the prairie pothole region south to California, Mexico, Texas, and Louisiana. Pintails begin arriving on wintering areas in early August. They are also one of the first ducks to leave wintering areas, as early as mid-February, to begin migration to breeding grounds.

Wintering Ecology - California is the most important wintering area in North America, and more pintails winter here than anywhere else in the Northern Hemisphere (Bellrose 1980). Other important wintering areas include the West Coast of Mexico and the Gulf Coast regions of Texas and Louisiana.

Distribution and Abundance

North America - The 1997 breeding duck survey conducted by the U.S. Fish and Wildlife Service found 3.6 million pintails in North America, which was a significant (30%) increase over 1996, but still 19% below the long-term average (Dubovsky et al. 1997).

Pacific Coast - Pintails are known to winter throughout the Pacific Coast region, and nest here in limited numbers. Important wintering areas include the coastal marshes of British Columbia, Puget Sound, the Lower Columbia River basin, the Willamette Valley, and the northern coast of California. The inland valleys of California are the most important wintering area for pintails.

San Francisco Bay - Mid-winter surveys conducted in January each year have indicated a great de-
cline in pintail numbers within the San Francisco Bay region, with the greatest proportion of that decline occurring in the Suisun Marsh (Figure 6.3). In the 1950s, there were close to 200,000 pintails wintering in the San Francisco Bay region, whereas the 1990s have averaged under 20,000, a decline of 90 percent.

**Suisun** – Nort hern Pintails winter in significant numbers in Suisun Marsh. Some pintails may roost on the open bays in this region (Figure 6.4), but most are found on managed seasonal wetlands (CDFG 1953-97). A radio telemetry study conducted from 1991-93 found pintails widely distributed throughout the managed wetlands of Suisun, with some distinct high use areas (Figure 6.5). The Suisun Marsh consists of approximately 23,000 hectares of marshlands and 9,300 hectares of bays and waterways and is the largest contiguous estuarine marsh in the United States (Miller et al. 1975). The majority of the pintails wintering in the San Francisco Bay region can be found in this area.

**North Bay** – Waterfowl surveys conducted between 1988-90 in the San Francisco Bay area indicated that North Bay salt ponds held 13-19% of the pintails (not including the managed wetlands of Suisun Marsh) (Figure 6.4). Open waters of the North Bay accounted for 12% of the region’s pintail population in 1988-90 (Accurso 1992). Diked baylands of the North Bay had significant numbers of pintails during the winter period (Figure 6.4).

**Central Bay** – Very few pintails were observed on the Central Bay between 1988-90 (Figure 6.4).

**South Bay** – South bay salt ponds held 60-67% of the pintails wintering in the San Francisco Bay region from 1988-90 (not including the managed wetlands of Suisun Marsh), while open waters of the South Bay received very little use by pintails (Figure 6.4).

**Historical Information**

Comprehensive waterfowl surveys have been conducted since the mid-1950s which include much of the pintails primary range. Population trends and estimates are available since that time.

**Population Trends**

Nationally, current pintail population estimates have increased somewhat from the all-time low of 1.8 million in 1991, and are well below goals established by the North American Waterfowl Management plan (5.1 million) (Calthamer and Dubovsky 1997). The number of pintails wintering in California has decreased dramatically from long-term averages. Locally, pintail use of the San Francisco Bay Area has declined in even greater proportion than the overall population decline.

**Habitat Use and Behavior**

Pintails are known to use a variety of habitats within the San Francisco Bay region, including diked fresh and estuarine wetlands, salt ponds, open bays, and mudflats (Cogswell 1977, Accurso 1992, Casaza 1995). Within the managed seasonal wetlands of the Suisun Marsh, pintails prefer to feed in habitats dominated by brass buttons (Cotula coronopifolia), a perennial salt tolerant herb introduced to the Bay in the late 1800s (Casaza 1995).

**Foraging** – Several studies have been conducted on the feeding ecology and diet of northern pintails, but none have included San Francisco Bay, and studies conducted in Suisun Marsh are outdated. Pintails are adept at separating small seeds from bottom sediments in aquatic habitats, and regularly feed on small seeds (Krapu 1974). Pintails use their long necks and tipping style to feed on or near the bottom of ponds and to utilize the benthos and seeds present at shallow depths (Krapu 1974). Pintails can exploit food sources to depths of 40 cm (Thomas 1976).

The winter diet of northern pintails consists primarily of seeds and vegetative material, with important seeds including rice (Oryza), swamp timothy (Heterochloa...
Figure 6.4 Maximum Counts of Northern Pintail


Bayland data from the Diked Baylands Wildlife Study (DBWS), 1982-1989 (USFWS, in prep.).

Figure 6.5 Radio-Marked Pintail Locations in Suisun Marsh During the Fall and Winter of 1991-92 and 1992-93 (Casazza 1995)
schenoides), barnyard grass (Echinochloa crusgalli), flatsedges (Carex spp.), southern naiad (Najas guadalupensis), and smartweeds (Polygonum spp.) (Austin and Miller 1995). Miller (1987) found that plant foods accounted for nearly 100% of the diet of pintails in the Sacramento Valley in early fall, and by late winter their diet had shifted to about 40% animal matter, primarily midge larvae. In the Suisun Marsh, pintail gizzards were found to contain three main marsh seeds; brass buttons (Cotula coronopifolia), alkali bulrush (Scirpus maritimus) and fat hen (Atriplex triangularis) (George et al. 1965).

**Roosting** - Pintails roost on open water habitats, including lakes and bays, which lack extensive emergent vegetation. Pintails commonly roost on open water areas of all four regions of San Francisco Bay (Figure 6.4).

**Movement** - Pintails commonly make local and regional movements. Local movements primarily consist of evening and morning flights between roost and feeding areas, usually less than 5 km. Soon after sunset in the Suisun Marsh, radio-marked hen pintails would commonly leave open water sanctuary areas such as Joice Island, to make 1.3 km flights to feeding areas on nearby private duck clubs (Figure 6.6). Regional movements occur when pintails leave an area and establish new feeding and roosting sites, usually encompassing distances greater than 10 km. Radio-marked pintails commonly left the Suisun Marsh and established new feeding and roosting sites in the Delta, and the Sacramento and San Joaquin valleys (Casazza 1995). Only one radio-marked pintail moved to San Francisco Bay from Suisun Marsh during two years of study (Casazza 1995).

**Conservation and Management**

**Contaminant Risks** - Little information is known about exposure of pintails to contaminant risk in the San Francisco Bay region. Health warnings are published for greater and lesser scaup (Aythya marila and A. affinis) and surf scoters (Melanitta perspicillata) harvested in the San Francisco Bay region, and further investigation of contaminants in pintails is warranted. Ohlendorf and Miller (1984) found that pintails seemed to accumulate chemicals such as DDE while wintering in California, and similar accumulations of contaminants may take place in birds wintering in the San Francisco Bay region.

**Disturbance** - Disturbance of northern pintails was studied by Wolder (1993) on Sacramento National Wildlife Refuge in the Sacramento Valley of California. He found that human disturbance was a major factor in pintail distribution. Disturbances by humans resulted in longer time spent alert or flying than disturbances caused by raptors or other animals (Wolder 1993). Common types of disturbance include vehicle traffic, wildlife observation, and hunting.

**Recommendations**

Suisun currently provides critical support for pintails. North Bay and South Bay also are used by these birds. Managed marsh is the most critical habitat type for pintails in the Estuary, and for Suisun in particular, for most of winter. Uplands puddled from rain and seasonal ponding provide critical late winter/spring habitat for pintails in this region. Many of the other habitat types, such as diked marsh and muted tidal marsh, would provide even better habitat if managed for dabbling ducks. Intertidal mudflats are important feeding habitat when covered by 10-40 cm of water. Unvegetated levees and islets are important roosting habitats.

Regional goals to support pintails include increasing the acreages of managed marsh and low salinity salt ponds; maintaining and enhancing mid-tidal, muted tidal (if managed for water fowl), and diked marsh; and maintaining the acreage of intertidal flat and shallow bays. Seasonal ponds in farmed and grazed baylands should be maintained or enhanced, and mid-salinity salt ponds should be maintained but converted to low salinity. Adjacent to the baylands, irrigation ponds in farmed uplands should be maintained, and managed upland habitat should be maintained for nesting.

**Suisun** - All managed marsh habitat within the Suisun subregion is important to support pintails. Goals within this subregion should include maintaining the current acreages of muted tidal and managed marsh, intertidal flat, shallow bays or straights, and managed uplands. To best meet the needs of pintails, muted tidal marsh, diked marsh, and managed marsh should be managed for water levels, vegetation, and timing of flooding.
most beneficial to dabbling ducks. Specifically, open water
10-40 cm deep with brass button-type vegetation is best.

**North Bay** – North Bay salt ponds were an important
habitat for pintails. Survey data indicates that at least
1000 hectares (ha) of salt ponds should be maintained
to ensure at least 80% of the current use by pintails in
this region. In particular, ponds 1AN, 2, 2N, and 6N
had a significant amount of use by pintails. Other
important areas include Slaughterhouse Point; American
Canyon Marsh; the mudflats near Sonoma Creek; open,
shallow water near mudflats; Point Richmond Bay; and
Tubbs Island lagoon. Goals to support pintails within
the North Bay include maintaining current muted tidal
marshes and diked marshes and enhancing them by
managing them for ducks; maintaining mid-tidal marsh,
low salinity salt ponds; ruderal baylands (used for nest-
ing), intertidal flat, and shallow bays or straights. Pond-
ing should be enhanced in farmed or grazed baylands and
adjacent uplands. Mid-salinity salt ponds should be con-
verted to low salinity ponds, and managed upland habi-
tat should be maintained to provide nesting habitat. In-
creasing managed marsh habitat would support an in-
crease in pintail population.

**Central Bay** – Important areas include Point Rich-
mond Bay mudflats, Albany mudflats, and the Emeryville
Crescent. To support increased populations of pintails in
the Central Bay, the current acreage of managed
marsh should be increased.

**South Bay** – South Bay salt ponds held 60-67% of
wintering pintails of the San Francisco Bay (not in-
cluding Suisun) from 1988-90. Survey data indicates
that at least 2,100 hectares (ha) of salt ponds should be
maintained to ensure at least 85% of the current use by
pintails in this region. Areas of critical importance to pin-
tails include the Sunnyvale sewage pond, and Ponds A9
and A10. Other important areas include Ponds A3W,
B3C, M 1, A, and N A1; M owry Slough; Charleston
Slough; Faber Tract; the shallows near the east end of the
D umbtown Bridge; Coyote Slough; the H ayward
ponds; and the H ayward treatment ponds. Goals to sup-
port pintails would include maintaing the current acre-
age of low salinity salt ponds and increasing it by
covertting mid-salinity ponds; maintaining the current
acreage of mid-tidal marsh, intertidal flat, and shallow
bays and straights; and maintaining and enhancing the
current acreage of muted tidal marsh and diked marsh
by managing for ducks. The current acreage of managed
marsh should be maintained, or else increased to increase
pintail populations.

**Research Needs**

The disturbing decline of pintail abundance in the San
Francisco Bay region needs immediate attention. Stud-
ies need to be implemented which will identify manage-
ment practices that can be used to attract pintails to the
region and provide a solid habitat base for these water-
fowl throughout the winter. Other factors that should
be investigated include effects of disturbance and con-
taminants on pintail abundance in this region.

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**References**

Accurso, L.M. 1992. Distribution and abundance of
wintering waterfowl on San Francisco Bay, 1988-

(Anas acuta). In: A. Poole and F. Gill (eds). The
birds of North America, No. 163. The Academy
of Natural Sciences, Philadelphia, and The Ameri-
can Ornithologists’ Union, Washington, D.C.

Bellrose, F.C. 1980. Ducks, geese and swans of North

Caithamer, D.F. and J.A. Dubovsky. 1997. Waterfowl popu-
California Department of Fish and Game (CDFG).

Casaza, M.L. 1995. H abitat use and movements of north-
ern pintails wintering in the Suisun Marsh, Califor-

No. 40. 399pp.

Fish and Wildl. Serv., Office of Migratory Bird Mgmt.
Duncan, D.C. 1987. Nest-site distribution and over-
land brood movements of northern pintails in
Alberta. J. Wildl. Mgmt. 51:716-723.

An evaluation of the Suisin Marsh as a waterfowl
area. Calif Dept. Fish and Game, Admin. Rept.

Krapu, G.L. 1974. Feeding ecology of pintail hens dur-

Miller, A.W., R.S. Miller, H.C. Cohen and R.F. Schultz.
1975. Suisun Marsh study, Solano County, Cali-
ifornia. U.S. Dept. of Agriculture, Soil Cons. Serv.,
Portland, OR.

Miller, M.R. 1987. Fall and winter foods of Northern
pintails in the Sacramento Valley, California. J.
Wildl. Mgmt. 51:405-414.

Ohlendorf, H.M. and M.R. Miller. 1984. Organochlo-
rine contaminants in California waterfowl. J. Wildl.
Mgmt. 48(3):867-877.
Canvasback
Aythya valisineria
John Y. Takekawa
Carolyn M. Marn

Introduction

The canvasback is a diving duck that forages on aquatic plants or benthic invertebrates in mouths of rivers or channels, large wetlands, and brackish marshes. The continental population of canvasbacks has not increased greatly in the last 20 years, and based on mid-winter surveys (USFWS, unpubl. data), the population in the Estuary has continued to decline. Consequently, the canvasback is a species of special concern for the U.S. Fish and Wildlife Service, and protection of this species was one of the reasons for establishment of the San Pablo Bay National Wildlife Refuge. Associated species that use similar habitats in the Estuary include: common goldeneye (Bucephala clangula), greater and lesser scaup (Aythya marila and A. affinis), which also have declining continental populations, and very small populations of redhead (A. americana) and ring-necked ducks.

Description

The canvasback is one of our most distinctive waterfowl species and a member of the Tribe Aythyini. They have a steeply sloping bill with a body size similar to the mallard (0.9-1.6 kg; Cogswell 1977). Males are distinguished by their white back, underparts, and wings, black tail and breast, and red head with blood-red eyes. Females have a brown head and eyes, and a brown or dusky gray body. Redheads are similar in appearance, but have a shorter bill, and redhead males are darker. Canvasbacks are the fastest flying large duck in North America (Bellrose 1980). Their call is a weak “ik-ik-coo” (Cogswell 1977).

Breeding Biology – Canvasbacks pair during migration in March and April (Erickson 1948, Bellrose 1980). Females will breed as yearlings, but generally less successfully than in later years (Hochbaum 1944, Olson 1964, Trauger 1974). Canvasbacks are well known for their strong fidelity to natal breeding areas (Trauger 1974). Females spend one week searching for a nest site, two to three days to build the nest, and one day laying each egg. Nests occur commonly in shallow ponds of less than one acre covered by cattail and bulrush (Trauger and Stoudt 1974); however, their overlapping home ranges may reach 1,300 acres. Canvasbacks nest in deep water (greater than 6-24 inches) in ponds, marshes, sloughs, and potholes (Trauger and Stoudt 1974). They nest in late April and early May (peak, mid-May) and incubate for 24-29 days. Nest success averages 46.2%, but it is highly variable at different sites, depending on predation by raccoons, mink, skunks, coyotes, fox, weasels, crows, and magpies and desertion following flooding or parasitic laying. As much as 57% of the nests may be parasitized, primarily by redhead ducks (Trauger and Stoudt 1974). Average clutch size is 7.9 eggs, but more eggs (9.5) are found in unparasitized nests. Nest success may be as low as 17% in dry years (Serie et al. 1992), but varies from 17-62% in southwestern M anitoba (Serie et al. 1992). Renesting may occur as much as 50% of the time. Broods are reared in open, large, deep-water areas, but survival averages 25% over the first two months. The 55% of successful hens rear an average of 5.3 fledged young. Non-breeding birds form flocks in late June and July. All birds have a 3-4 week flightless wing-molt during late July and August.

Migration Ecology – Canvasbacks begin their autumn migration in September and arrive in wintering areas in early November. Their spring migration begins in February until their return to breeding areas in April. Major migration routes occur along the Upper Mississippi River and Great Lakes to the Atlantic and Gulf Coast regions, and along the Pacific Coast and Intermountain west to the west coast and M exico. Significant migration areas in the west include Puget Sound, Great Salt Lake, M alheur and Klamath Basin National Wildlife Refuges and Carson Sink in the Great Basin. During the past two decades, use of staging areas has increased at Pyramid Lake, Klamath Basin National Wildlife Refuges while use of the Great Salt Lake, M alheur and Stillwater National Wildlife Refuge has decreased.

Wintering Ecology – Birds arrive on wintering areas in late October and increase in numbers through
December. Nearly 90% of canvasbacks produced in Alaska winter in California (Lensink 1964). The largest wintering population of canvasbacks is on the Atlantic Coast (290,000), with the greatest number on the Chesapeake Bay. A large number of birds winter along the Gulf Coast in the Mississippi River Delta and Catahoula Lake. Western wintering areas include Lake Earl in Humboldt County, the Columbia and Snake river basins, Carson Sink, the Central and Imperial valleys, and San Francisco Bay. San Francisco Bay is the major wintering area for the western population with 60,000 birds counted in 1960-1971, but only about 25,000 birds counted in 1990. Canvasbacks exhibit high winter site fidelity to the Estuary (Rienecker 1985) and generally winter in more saline areas than redhead ducks.

**Distribution and Abundance**

**North America** - Canvasbacks breed in the Arctic and Subarctic, and in the prairie and parkland areas of North America. Approximately 190,000 canvasbacks are found in the parkland areas, with a peak of 10 pairs per square mile with up to 10% of the continental population near Minnedosa, Manitoba. Other breeding areas include Alaska, Yukon, Northwest Territories, and the pothole region of southern Canada and the northern United States.

**Pacific Coast** - Canvasbacks migrate along the Pacific Coast to and from their northern breeding areas. They are found in most of the major estuaries along the lower west coast during winter, including Puget Sound, Willapa Bay, and Humboldt Bay with the largest populations found in the San Francisco Bay.

**San Francisco Bay** - Within the San Francisco Bay Estuary, canvasbacks comprise 7% of the waterfowl, and 46.5% (1989) to 54% (1990) of the mid-winter population in the Pacific Flyway (USFWS unpubl. data summarized in Accurso 1992). Many birds also stage on the Estuary during migration (Cogswell 1977).

**Suisun** - About 13-16% of canvasbacks in the Estuary occur in Suisun with peak numbers of greater than 6,000 canvasbacks (Accurso 1992). Eighty percent of the waterfowl in open water habitats were found in Suisun Bay proper rather than Honker Bay or Carquinez Strait (Suisun managed marshes were not included in this survey; Figure 6.7).

**North Bay** - Between 38-59% of canvasbacks wintering in the Estuary in 1988-1990 were found in the northern salt evaporation ponds (Accurso 1992). The greatest number of canvasbacks in the Estuary are found in these ponds. An additional 9.5-25.5% of canvasbacks are found in the open bay area (Figure 6.7).

**Central Bay** - Only 1% of canvasbacks are found in the Central Bay; however, hundreds have been reported in Lake Merritt adjacent to the Bay (Cogswell 1977; Figure 6.7).

**South Bay** - Up to 17% of wintering canvasbacks are found in South Bay salt evaporation ponds with an additional 1.7-1.9% in the open bay. Up to 4-5% of waterfowl in the salt evaporation ponds (peak 6,400) are canvasbacks. Canvasbacks are the third most abundant duck found in this region (Figure 6.7).

**Historical Information**

Historically, canvasbacks were likely very abundant in parts of the Estuary. Areas mapped by Jose Canizares in 1776 in the northern reach of the Estuary were labeled “Forests of the red duck” (Josselyn 1983). Canvasbacks also were described as “abundant” in San Francisco bays and marshes in the winter in the early 1900s (Grinnell and W ythe 1927), arriving in early October and departing in early April, while peaking from late November to early March. They historically used open, deeper water for roosting, foraging in inner bays and marshes closer to shore (Grinnell and W ythe 1927). Christmas Bird Counts (National Audubon Society, unpubl. data) from Palo Alto and San Jose show a population decline of 0.27% and 1.13%, respectively, from 1969 to 1996 on the basis of the small areas surveyed. The regional population of canvasbacks actually decreased by 50% from the 1970s to the mid-1980s according to annual mid-winter surveys (USFWS, unpubl. data).

**Population Trends**

**National** - The continental canvasback population averages 534,000 birds but declined between 1955 and 1993 (ohman et al. 1995). The population is highly skewed with only 20-30% females (Trauger 1974) or 1.94 males per each female in spring (Bellrose et al. 1961) and suggests an older age structure. Females have 27% higher mortality (Geis 1959). With about 30% females, the breeding population is estimated as 140,000 pairs. There are an estimated 1.03 young per adult during the fall flight (Bellrose 1980). Adult mortality is 52%, while juvenile mortality is about 77% (Geis 1959).

**Regional** - In the 1950s, 79% of the wintering canvasbacks were found in the Atlantic or Pacific flyways. However, by the 1990s, 44% of the birds were found in the Central and Mississippi flyways. A large increase was also noted in M exico. Annual survival was found to be higher in the Pacific versus the Atlantic Flyway populations (female 56-69%, male 70-82%) (Nichols and Haramis 1980). Although the overall population estimate is 2.0-2.5 males per female, the sex ratio varies from 2.9-3.2 in the Atlantic Flyway, to 1.6-1.8 in the Mississippi (Woolington 1993), to 2.2 in the Pacific (Ohman et al. 1995).

**Local** - San Francisco Bay remains one of the top ten major wintering areas for canvasbacks in North America. The San Pablo Bay National Wildlife Refuge
was established for protection of this species. Canvasbacks are the fifth most numerous diving duck in winter in the Estuary (Accurso 1992) and account for 6-7% of all waterfowl. Their peak numbers are observed in early to mid-January. Population numbers have decreased from 60,000 canvasbacks in the 1960s to about 25,000 birds in the early 1990s.

**Habitat Use and Behavior**

Unlike most ducks, canvasbacks are dependent on aquatic habitat throughout their life cycle including the breeding period (Olmman et al. 1995). They are found in estuarine and lacustrine habitats throughout California (Zeiner et al. 1990). They are benthivores that feed in shallow waters over and near intertidal mudflats (Cogswell 1977). They prefer shallow depths with 80% of canvasbacks recorded in areas of less than 3 meters depth and 60% in areas less than 2 m depth (Accurso 1992). Their use of shallow areas of <0.99 m was twice the proportion of the availability of those areas. Canvasbacks use salt evaporation ponds of low (20-33 ppt) or medium (34-63 ppt) salinity in medium-sized ponds (2-2.25 km²). They generally roost in open water areas.

**Foraging** - Canvasbacks are strongly associated with foraging on aquatic plants, including wild celery (Vallisneria americana) with which it shares a similar scientific name. Early diet studies suggested canvasbacks foraged mostly on plants (80%) with some animal (20%) prey (Cottam 1939, Palmer 1975); food items included...
Canvasbacks are the least abundant, widely distributed game duck. They have had special hunting protection during several periods including 1936-37, 1955-74 and the present time (Anderson 1989). The goal for the continental population established by the North American Waterfowl Management Plan is 540,000 (USFWS and CWS 1994).

**Conservation And Management**

Canvasbacks are most abundant in the North Bay subregion. Ponds 1AN and 1N are used by thousands of birds, and Ponds 3N, 4N, 5N are used by many thousands of birds. White Slough is also heavily used (many thousands of birds), and the marsh near Slaughterhouse Point is moderately used by thousands. Birds also use the Point Pinole area by the many thousands. The mudflats near China Camp and Hamilton Air Field are moderately used by thousands of birds. We recommend restoring this subregional population by increasing large brackish shallow water areas. If Ponds 3-5 of the salt ponds are altered, suitable alternative habitats must be provided or the population may decrease substantially.

**Movement**

No information.

**Roosting**

Canvasbacks generally roost on open water areas. They are found in larger salt evaporation ponds.

**Disturbance**

Canvasbacks may be disturbed by boats, aircraft, people, and pets. They may avoid preferred foraging areas during the day but may use these areas at night. They will use undisturbed open water roosting areas near feeding sites when available.

**Contaminant Risks**

Studies conducted on contaminants in canvasbacks (Miles and Ohlendorf 1995, Ohlendorf et al. 1986) indicate elevated tissue concentrations in the Estuary. Recent invasions of the Asian clam (Potamocorbula amurensis) indicate an exponential increase in this exotic species which may triple the concentration of selenium (Luoma and Linville 1995) in prey likely to be consumed by canvasbacks.

**Habitat Goals for the Region**

- The shallows of northern Suisun Bay are critical for support of canvasbacks. Other areas of importance within the Suisun subregion include the middle of Honker Bay, the shallows north of the shipping channel, and Benicia State Park. The shallows in northern Suisun Bay show high use, with counts of a few thousand birds. The middle of Honker Bay is moderately used by thousands of birds. The shallow water north of the shipping channel and the area around Benicia State Park are both moderately used by hundreds of birds. To achieve the recommended increase of the subregional population, there must be an increase in the area of large brackish shallow water. Habitat goals for the Suisun subregion include maintaining the current acreage of managed marsh, shallow bay and strait, and intertidal flat, and increasing the current acreages of lagoon, treatment pond, and tidal reach.

**North Bay**

Canvasbacks are most abundant in the North Bay subregion. Ponds 1AN and 1N are used by thousands of birds, and Ponds 3N, 4N, 5N are used by many thousands of birds. White Slough is also heavily used (many thousands of birds), and the marsh near Slaughterhouse Point is moderately used by thousands of birds. Birds also use the Point Pinole area by the many thousands. The mudflats near China Camp and Hamilton Air Field are moderately used by thousands of birds. We recommend restoring this subregional population by increasing large brackish shallow water areas. If Ponds 3-5 of the salt ponds are altered, suitable alternative habitats must be provided or the population may decrease substantially.

Habitat goals for the North Bay subregion include maintaining the current acreage of intertidal flat and
shallow bays or straits, and increasing areas of lagoons, managed marsh, low salinity salt pond, treatment pond, and tidal reach.

Central Bay - The Berkeley Marina and Emeryville Crescent are both lightly used by a few hundred birds. Point Isabel and the Albany mudflats are used by a few hundred birds. Richardson Bay, Candlestick Point, and the shallows around the Oakland Airport also are used by hundreds of birds. To increase the limited population, increase large brackish shallow water areas. Habitat goals for the Central Bay subregion include maintaining the current acreage of intertidal flat and shallow bays or straits, and increasing areas of lagoon, managed marsh, low salinity salt pond, treatment pond, and tidal reach.

South Bay - Most bayside salt ponds are used by thousands of birds. Some ponds are used very heavily by many thousands of birds. Deep open water habitat is not used. The areas of Mowry Slough and Coyote Creek also receive moderate use by thousands of birds. We recommend maintaining the subregional population. An increase of large, shallow (<6 ft.), low salinity (less than 33 ppt) water areas would be beneficial. If salt ponds are altered, suitable alternative habitat must be provided.

Habitat goals for the South Bay subregion include maintaining the current acreage of intertidal flat and shallow bays or straits, and increasing areas of lagoon, managed marsh, low salinity salt pond, treatment pond, and tidal reach.

Research Needs

Information is needed on region-specific population relationships, diet effects, and cross-seasonal studies of diet and contaminants in their life cycle.

References


Serie, J.R., D.L. Trauger and J.E. Austin. 1992. Influence of age and selected environmental factors on...

Personal Communications
G. Michael Haramis, National Biological Service
Dennis G. Jorde, National Biological Service

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**Surf Scoter**

*Melanitta perspicillata*

A. Keith Miles

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**Introduction**

Surf scoters are the least studied of the North American waterfowl (Johnsgard 1975, Palmer 1976). San Francisco Bay appears to be the most important inshore habitat in the eastern Pacific, south of the Straits of Georgia and Puget Sound (Martell and Palmisano 1994, Small 1994). This species is representative of sea ducks that primarily use deeper, open water habitat. Associated species are white-winged scoters (*M. fusca*), black scoters (*M. nigra*), and red-breasted mergansers (*Mergus serrator*).

**Description**

Scoters are Anatid sea ducks of the Tribe Mergini. The surf scoter is the most common of the three North American scoters that winter at San Francisco Bay (Bellrose 1980). Adult surf scoters measure about 43 - 53 cm in length with a wing span of 76 - 86 cm, and weigh 0.7 - 1.1 kg. Adults are nearly identical in size to black scoters and slightly smaller than white-winged scoters. Male surf scoters have a distinctive hump on the bill. The coloration of all three scoters is also similar; the distinct difference is that adult male surf scoters have a white patch on the crown and nape. However, first-year males are all black and very similar in appearance to black scoters. Female and immature surf scoters have a dusky brown coloration, similar to white-winged scoters (except the latter have distinct patches of white on the wings). Surf scoters rarely vocalize, but do emit a low, guttural sound. The distinct whistling sound during flight is generated by air passing over their wings.

**Breeding Biology** - Scoters prefer fresh water, shallow, rocky, Arctic lakes for breeding. The breeding chronology begins with egg-laying in early June, and hatching the second or third week of July (Savard and Lamothe 1991). Nests are built away from water on the ground, and consist of a shallow excavation lined with...
grasses and feathers. Egg clutch and brood size are unclear because of the few observations made. Those observations have indicated clutches of about five eggs, and broods of about five young, but brood amalgamation appeared common.

**Migration Ecology** - Surf scoters migrate directly to the oceanic coast from the breeding areas. Three times more birds migrate to the Pacific Coast than to the Atlantic Coast. Their subsequent southward migratory destination appears conditioned to the preferred stopover and wintering destination of individual flocks.

**Wintering Ecology** - Surf scoters use both offshore and inshore marine and estuarine habitats during winter. Marine habitat encompasses the entire Pacific Coast from the Aleutian Islands, Alaska to central Baja California, Mexico (Root 1988). The Canadian interior passage appears crucial as habitat. Surf scoters are common along the California coast from October to May, and San Francisco Bay appears to be the most important inshore habitat in the eastern Pacific, south of the Straits of Georgia and Puget Sound (M. artell and Palmisano 1994, Small 1994).

### Distribution and Abundance

**North America** - The breeding range of surf scoters extends from patchy sites in western Alaska, extensive occupation across the Northwest Territories to Hudson Bay, and east of the Bay into Labrador (Bellrose 1980). The wintering range extends from the Aleutian Islands to Baja California on the west coast, and from the Bay of Fundy to Florida on the east coast. They also occur on the Great Lakes and on inland bodies of water along the coastal states, but are most common either on nearshore marine waters or calm estuaries. Bellrose (1980) estimated wintering populations of surf scoters in North America at about 130,000 birds.

**Pacific Coast** - The wintering range extends from the Aleutian Islands to Baja California on the Pacific Coast.

**San Francisco Bay** - Most counts of scoters lump the three species together. Surf scoters comprise the majority of scoters observed on San Francisco Bay. Accurso (1992) identified scoters as the second most abundant waterfowl on San Francisco Bay in two wintering seasons between 1988 and 1990, accounting for about 20% of the waterfowl counted. Scotters are common throughout the open waters of San Francisco Bay. Scotters can be observed close to land near China Camp and Hamilton Airfield; near shore at the Presidio cliffs, and off Point Molate in the Central Bay; and near shore on open waters off Coyote Hills, the Edwards National Wildlife Refuge fishing pier, the bayshore levee at Foster City, and the San Mateo County NWR Fishing Pier (pers. obs.; Sequoia Audubon Society 1985).

**Suisun** - Flocks numbering in the high hundreds were found on the open waters of Suisun Bay in early December. Scoter numbers increased to the upper hundreds and low thousands (<10,000) later in the winter, with their distribution shifting from north to south Suisun Bay, and east to Chipps Island and Honker Bay (Figure 6.8). Also, scoter numbers ranged from the low hundreds to low thousands (<1000) in the Carquinez Straits region east of Suisun as the winter season progressed.

**North Bay** - Beginning in mid-October, scoters were common throughout north San Francisco Bay, with the larger flocks (numbering in the mid- to upper hundreds) common near the Hamilton Airfield (Accurso 1992). By mid- to late winter, scoters were widespread in the North Bay with some flocks numbering in the thousands (Figure 6.8).

**Central Bay** - Scoters have been identified as the most abundant waterfowl in this region (Accurso 1992). Peak numbers of about 24,000 to 30,000 have been counted on the open waters of the Central Bay in the early wintering season (Figure 6.8).

**South Bay** - As the second most abundant waterfowl in this region, numbers of scoters were highest in December (about 9,500 - 11,000) in the two winters between 1989 and 1990 on the open waters in this region (Accurso 1992; Figure 6.8).

### Historical Information

No information.

### Population Trends

Mid-winter surveys (conducted in January) of scoters indicated a high of about 72,000 scoters on San Francisco Bay in 1991, and a low of 1,200 birds in 1996 (Trost 1997). Their numbers rebounded to about 28,000 in 1997. Mid-winter surveys of the western states and Mexico indicated about 69,500 scoters, which was about 20% lower than the ten year average (1987 - 1996).

### Habitat Use and Behavior

**Foraging** - Surf scoters are strong divers, and have been observed foraging in the 2-10 m depth range (pers. obs., Sequoia Audubon Society 1985, Root 1988). Scoters feed in the open waters of the Bay, and also along the cliffs at the entrance to San Francisco Bay. Their habit is to dive in the areas on the trailing side of waves breaking at the cliffs (Sequoia Audubon Society 1985). They have also been observed feeding on rock-bound intertidal or shallow subtidal mussels or scallops at high tide. Their preferred diet consists of clams inhabiting silty or sandy substrate, or mussels attached to hard substrata such as pilings or rocks (Vermeer and Bourne 1982), but are likely to opportunistically consume other molluscs and also crustaceans.
Roosting - Surf scoters roost almost exclusively on open, coastal bay, or lake waters.

Movement - Movement seems related to foraging, roosting, or disturbance. Like most diving waterfowl, surf scoters probably conduct short-distance moves from area to area as prey are depleted. Flocks have been observed at the Bay riding the high tide to shore to feed on blue mussels along the rocky shore or ribbed mussels in the cordgrass habitat.

Conservation and Management

Contaminant Risks - Several studies have indicated elevated concentrations of elemental contaminants in scoters inhabiting San Francisco Bay (e.g., Ohlendorf and Fleming 1988; CDFG, unpubl.). The consequences of elevated selenium or mercury to survival or productivity of these scoters have not been determined.

Disturbance - Surf scoters appear very intolerant of human disturbance, particularly motorboats. Their foraging movement into the intertidal zones at high tide occurs at areas secluded from human disturbance or at night. Overflights by birds of prey also were observed to elicit panic response by flocks roosting on open water. Incidental mortality of scoters was recorded from commercial fishing with gill nets (Heneman 1983). However, regulations curtailing coastal gill net fishing have probably reduced the number of birds killed to those caught in nets accidentally set adrift.

Figure 6.8 Maximum Counts of Surf Scoter


Bayland data from the Diked Baylands Wildlife Study (DBWS), 1982-1989 (USFWS, in prep.).
Recommendations

The important habitats used by scoters are the open waters throughout San Francisco Bay, and the underlying sediments for foraging. Scoters will forage in low tidal wetlands during high tide. Scoters are susceptible to bioaccumulation of contaminants. Restoration of the Bay’s shorelines to tidal wetlands should include studies of the potential for mobilization of contaminants that may be sequestered in existing soils or sediments.

Scoters primarily use the open coast. San Francisco Bay appears to be important to populations of scoters that may either be historically affiliated with the Bay or that seek refuge in the Bay during inclement weather on the coast. The Bay (all subregions) provides very crucial wintering habitat for these birds. Besides open water, other important habitat elements include clay-silt substrate, sand substrate, and rock substrate. The primary recommendation to support scoters is to maintain the current acreages of shallow bay, intertidal flat, and low tidal marsh.

Suisun – Maintain current acreages of shallow bay, intertidal flat, and low tidal marsh.

North Bay – All open waters of the North Bay are used by scoters, and particularly important are the open waters of China Camp and Hamilton Airfield (for foraging and wind protection), and the deeper water off of Wilson Point. Maintain current acreages of shallow bay, intertidal flat, and low tidal marsh.

Central Bay – All open waters near the shoreline in the Central Bay are used by scoters. The feeding habitat near the shore at the mouth of the Bay is particularly important, and eelgrass beds sporadically distributed in Central Bay may be important. Maintain current acreages of shallow bay, intertidal flat, and low tidal marsh.

South Bay – Open waters throughout the South Bay are important for scoters. Of particular importance are the open waters off of Coyote Hills area and south of Coyote Hills, and the open waters near Foster City. Maintain current acreages of shallow bay, intertidal flat, and low tidal marsh.

Research Needs

Surf scoters are the least studied of the North American waterfowl (Johnsgard 1975, Palmer 1976), but recent die-offs in Alaska has raised concern about these ducks (Bartonek 1993). Elevated concentrations of contaminants have been detected in scoters, particularly in the Suisun region. Inhabitants of this region should be radio-tagged in order to determine their wintering movements and survival, and also their survival and productivity on the breeding grounds as compared to other sub-populations of scoters.

Acknowledgments

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References


California Department of Fish and Game (CDFG). unpublished. Selenium verification studies.


Ruddy Duck  
*Oxyura jamaicensis*  
A. Keith Miles

**Introduction**

This diving duck is widespread and has one of the largest wintering concentrations in the San Francisco Bay Estuary. It uses a variety of open wetlands, including managed marsh areas, but prefers salt ponds found around the perimeter of San Francisco Bay. It is grouped with the bufflehead (*Bucephala albeola*), which uses similar habitat.

**Description**

These ducks, also known as “stifftails,” are Anatid ducks of the Tribe Oxyurini. Adult ruddy ducks are small but full-bodied, measuring about 37 - 41 cm in length, with a wing span of 53 - 61 cm, and weigh 0.3 - 0.7 kg (Bellrose 1980). The ruddy duck’s stiff, erect tail is its most pronounced attribute. During breeding season, adult males display a reddish-brown coloration, white throat patch, and exceptionally bright blue bills; both sexes have white cheek patches. Otherwise, males and females are the same dull brown color, except that males maintain a bright white cheek patch.

**Breeding Biology** - Suitable breeding habitat consists of stable, fresh or alkaline water that supports emergent vegetation (Johnsgard and Carbonell 1996). Nests are characteristically placed deep into reedbeds with channels or easy access to open water; ruddy ducks will use nest boxes placed in the reeds. Breeding birds apparently require about 0.5 to 1.5 sq. km each, but in some places the density of nests averages one every 0.6 hectares. Males do not defend territories, but will guard an area of about 3 m around a female. Unique male courtship displays consist of “rushing” (swimming or half-swimming, half-flying in hunched position) the female, and “bubbling” (beating the bill rapidly against the breast). Courtship occurs from January into July. Pair bonding is considered loose, lasting only from just before to just after egg-laying. Egg-laying occurs from early April to late August. Ruddy ducks are known to parasitize other nests. Clutch size averages about seven eggs; incubation and rearing both average about 25 days each (Gray 1980).

**Migration Ecology** - Long-distance migratory behavior of ruddy ducks is not well-defined. Substantial variations have been recorded for size of migrant flocks, and it is suspected that migration at night is common (Bellrose 1980). Ruddy ducks sometimes migrate in flocks with other species of waterfowl. The best known migration route to eastern wintering areas is from the northern prairie wetlands to the Chesapeake Bay region. Migration corridors for western populations were suggested to extend from western Canada to Utah to California, and from Utah or California to the west coast of Mexico. Fall migration occurs from about mid-September into December; spring migration occurs from about February through April.

**Wintering Ecology** - More than half of the ruddy ducks in North America winter at or near the Pacific Coast from southern Canada to Mexico. The majority (85%) of these winter in California, primarily at San Francisco Bay. Coastal wintering areas consist of shallow-depth lagoons or estuaries. Protected or managed wetland areas around San Francisco Bay (particularly salt ponds in the South and North bays) and elsewhere in California are crucial to inland wintering populations (Root 1988).

**Distribution and Abundance**

**North America** - The breeding range of ruddy ducks extends from central British Columbia east to southern Manitoba and Minnesota, south from the western portion of the central states throughout the western states. Second to the Pacific Coast, the wintering range along the Atlantic Coast is mainly Chesapeake Bay (estimated 40,000 birds), with smaller groups further south; ruddy ducks that winter along the Gulf Coast States of the U.S. are estimated at 20,000 (Johnsgard and Carbonell 1996). Current
wintering populations for North America are estimated at about 180,000.

**Pacific Coast** - About 85% of the estimated 122,000 (Trost 1997) ruddy ducks that migrated to the west coast of North America stayed in California in winter 1997. According to Bellrose (1980) the primary wintering areas are San Francisco Bay, Imperial and San Joaquin valleys, and southern California coastal bays. The remaining ruddy ducks migrate to the west coast of Mexico.

**San Francisco Bay** - The greater number of ruddy ducks that migrate to California overwinter at San Francisco Bay. Since 1986, based on mid-winter surveys, numbers have ranged from about 1,900 (1996) to 28,300 (1991) (Trost 1997). In 1997, surveys indicated about 6,200 ruddy ducks on the Bay.

**Suisun** - Ruddy ducks infrequently inhabit Suisun, and primarily have been observed as individuals on managed marshes or stop-over migrants on open waters (Figure 6.9).

**North Bay** - Salt ponds in the northern Bay support the second highest numbers of ruddy ducks in the San Francisco Bay region, with counts from about 1,500 to 10,000 ducks in the two winter seasons between 1988 and 1990 (Accurso 1992). Smaller flocks numbering in the hundreds were observed in the near shore, northern-most, and northwestern waters of the Bay (Figure 6.9).
Observations of ruddy ducks were usually limited to individuals in the near shore waters, or small flocks numbering fewer than 100; however, larger flocks numbering several hundred birds were observed in Richardson Bay on the west side of the Central Bay in mid-December to early December, indicating this area is probably an important stop-over for migrating ducks (Accurso 1992; Figure 6.9).

South Bay - Salt ponds located on the eastern and southern shores of the South Bay have supported the greatest numbers of ruddy ducks in the Bay region. These ducks numbered from about 1,300 to 19,000 ducks during the two winters between 1988 and 1990 (Accurso 1992). Smaller flocks numbering in the hundreds were observed in the near shore waters of the South Bay adjacent to the salt ponds (Figure 6.9).

Historical Information

No information.

Population Trends

Ruddy duck populations increased about 12 percent in North America between 1965 and 1992 (Trost 1997). The 1997 mid-winter survey that indicated about 122,000 ruddy ducks wintering in the western states and Mexico was 42% above the 10 year average of surveys from 1987 to 1996 (Trost 1997).

Habitat Use and Behavior

Foraging - Ruddy ducks dive about 1-3 m for their food. They apparently prefer submerged aquatic vegetation, e.g., wigeon-grass or pondweed, but consume more benthic invertebrates during the summer or when vegetation is less available (Tome 1991). Their vegetable preference may be overestimated because soft-bodied insects, mollusks, or crustaceans are indiscernible in gizzards commonly used to determine food habits (Johnsgard and Carbonell 1996). Esophageal contents in ruddy ducks collected at Chesapeake Bay were mostly amphipods and other benthic invertebrates (Tome and Miles, unpubl. data). Ruddy ducks are known to feed both diurnally and nocturnally (Tome 1991). Ruddy ducks observed foraging at the salt ponds and northern and southern San Francisco Bay probably are feeding mostly on invertebrates.

Roosting - Ruddy ducks roost almost exclusively on open waters of protected estuaries, lakes, or ponds. They are most often observed roosting on salt ponds at the northern and southern regions of San Francisco Bay.

Movement - Personal observations of ruddy ducks at Chesapeake Bay indicate local flock movements between roosting and feeding areas, and we suspected that movements occurred more commonly at night. Similar habits may occur at San Francisco Bay with these ducks moving primarily between salt ponds and nearby Bay waters.

Conservation and Management

Contaminant Risks - On the east coast, ruddy ducks have characteristically been observed wintering in heavily urbanized embayments (e.g., Baltimore Harbor), with no apparent detriment to population numbers (Tome and Miles, unpubl. data). However, when other waterfowl are considered as surrogates, then inorganic elements, such as lead, mercury, or selenium, can be toxic at low concentrations.

Disturbance - Ruddy ducks are intolerant of human presence. However, they seem tolerant of urbanization, i.e., presence of nearby roadways or highways, and moderate levels of pollution. The presence of avian predators elicits flock dispersal and flight.

Recommendations

Any efforts to restore altered wetlands to historically tidal wetlands in the San Francisco Bay region should consider the importance of human-created water impoundments, e.g., salt ponds, to wintering waterfowl populations. San Francisco Bay is the most important wintering estuary for aquatic birds on North America's Pacific Coast. The efforts of the U.S. and Canadian governments in concert with private foundations, such as Ducks Unlimited and the Nature Conservancy, have contributed to the increase in numbers of ruddy ducks and other waterfowl through the acquisition and preservation of breeding habitat. Ruddy duck populations have become dependent on San Francisco Bay's altered habitats for winter-season survival. Intensive studies must be conducted to determine the effects of tidal wetland restorations on avian populations dependent on altered habitats.

The salt ponds of the North Bay are currently the most important area for ruddy ducks, and low salinity salt ponds provide crucial wintering habitat. Other habitat types and areas are used more as migratory stop-over points, while salt ponds are the preferred, crucial habitat. Important habitat elements are clay-silt substrate and sand substrate. Regional goals to support this species would include increasing the current acreage of low salinity salt ponds, and maintaining the current acreages of tidal reach and lagoon.

Suisun - Suisun is currently little used by ruddy ducks, but an increase in the amount of low salinity salt pond may increase bird numbers in this subregion. Population is sparse and widespread in the managed marsh. Important areas include open waters (migratory stop-over) and managed marsh. Goals to support ruddy duck include maintaining the current acreages of managed marsh, tidal reach, and lagoon.
North Bay - The low salinity salt ponds are the single most important habitat for wintering ruddy ducks in the North Bay. These salt ponds supported 25 to 30% of the populations of ruddy ducks that overwintered at San Francisco Bay between 1988 and 1990 (Accurso 1992). Particularly important are Ponds 3N, 6N, 1N, 2N, 4N, 5N, 2AN, 7N, and 1AN. The first six of these ponds were crucial habitat for about 90% of the maximum counts of ruddy ducks monitored in the North Bay in 1988-1990 (Accurso 1992). The nine ponds represent 90% of the habitat (by hectares) used by ruddy ducks, and should be maintained along with the remaining ponds in the North Bay because of currently increasing numbers of these ducks. Goals to support ruddy ducks include maintaining the current acreages of tidal reach and lagoon, and increasing the amount of low salinity salt pond.

Central Bay - Ruddy ducks generally use only open water habitat in the Central Bay. Increasing the amount of low salinity salt ponds in this very urbanized subregion does not seem feasible, though this may increase the subregional population. Richardson Bay appears crucial for stop-over migrants. Goals to support ruddy duck include maintaining the current acreage of tidal reach and lagoon.

South Bay - Low salinity salt ponds are crucial wintering habitat for ruddy ducks in the South Bay region. Populations of ruddy ducks inhabiting the Bay probably are increasing (Trost 1997), and the maintenance of these salt ponds is crucial. Of particular importance are Ponds A1, A2E, A2W, A3N, A3W, A4, A9, A10, A12, A14, A18, A1B, A2B, B1, B2, B3C, B4, B5, B6, B6A, B6C, B7, B8, B8A, B9, B10, B11, B12, B13, B14, M1, M3, M4, N1, N2, N3, N3A, N4, R2, and R3. Of these, 20 ponds supported 90% of the ruddy ducks wintering in south San Francisco Bay between 1988 and 1990 (Accurso 1992). Overall, South Bay salt ponds supported from 55 to 67% of the ruddy ducks wintering at San Francisco Bay during this time period. Groups of ruddy ducks numbering from 6 to 174 ducks were observed on 39 other ponds, which are probably important for localized movements of ducks inhabiting the South Bay. Goals to support ruddy ducks in the South Bay include maintaining the current acreages of low salinity salt ponds, tidal reaches, and lagoons.

Research Needs

Any plans for habitat alteration, including the restoration of salt ponds to tidal wetlands, have to consider the potential impact on the distribution and habitat use by ruddy ducks. Studies are recommended to determine importance of salt ponds for abundance of waterfowl prey in comparison to natural ponds, and the effects of salt pond alterations on ruddy duck distribution, abundance, survival, and behavioral modification. The potential for overcrowding by waterfowl and subsequent increase in avian disease outbreaks that might result from any reduction in salt pond habitat also needs study.

Acknowledgments

Special thanks to Janice Alexander (Biological Resources Division) for compiling and synthesizing data and maps on waterfowl distribution at San Francisco Bay, and thanks also to Bob Trost, U.S. Fish & Wildlife Service, and Mike Casazza and Mike Miller, Biological Resources Division, U.S. Geological Survey.

References


Personal Communications

Introduction

The population of snowy plovers that breeds along the Pacific Coast of the United States and Baja California, Mexico was designated as Threatened by the U.S. Fish and Wildlife Service in March 1993. The Pacific Coast population has declined most likely due to habitat loss and degradation, but also suffers from poor nesting success primarily due to predation. Critical habitat for the plover has been identified and a recovery team was formed. Along the Pacific Coast, San Francisco Bay is the northernmost area supporting over 100 breeding snowy plovers (Page et al. 1991). Salt ponds, their levees, and pond edges, which may mimic historic salt pan habitat in some essential way for the plover, provide almost all known snowy plover nesting habitat in San Francisco Bay today. The potential importance of San Francisco Bay salt pond habitat to the persistence and recovery of the Pacific Coast population of snowy plovers should be considered in any comprehensive management plan for San Francisco Bay wetlands.

Description

The snowy plover is a small (15-17 cm long) shorebird, with a light brown back and cap; white underparts; brown to black forehead bar; brown to black lateral breast patches and cheek patches; short black bill; and black legs.

Breeding Biology - In coastal regions, snowy plovers nest on the ground on barren to sparsely-vegetated beaches and dunes, on salt evaporation pond levees and edges, and along lagoon margins. In inland areas, they nest on the shores of alkaline and saline lakes and on river bars. Their nest is a simple scrape in the ground lined with small pebbles, shell fragments, plant debris, mud chips, or other debris. Females incubate the three-egg clutch during most daylight hours and males incubate at night (Warriner et al. 1986). The incubation period averages 27 days with an additional 4-5 days for egg laying on the California coast. Females typically desert the male and brood at hatching. Males raise broods until the young fledge, about 30 days after hatching. If a nest fails, the female typically renests with the same mate, up to five times per season, until a clutch hatches. The most successful females may have up to three broods per season and the most successful males may have up to two.

Migration Ecology - Along the California Coast, including San Francisco Bay, part of the snowy plover population is resident year round and part is migratory. Birds, which nest along the coast, may migrate north or south for the winter or remain at their nesting site. Most birds which breed at inland locations migrate to the coast for the winter. In winter, birds which breed on the coast and birds which breed inland occur in the same flocks. Migrants begin returning to coastal breeding locations as early as late January or early February, but most migration is from early March to late April. Coastal breeders begin departing for wintering areas in early July and the exodus continues through October.

Wintering Ecology - In winter, snowy plovers are usually found roosting in flocks ranging in size from a few individuals to up to 300 birds. They often sit in footprints or other depressions in the sand. Foraging occurs on sandy beaches, salt evaporation ponds, or tidal flats. Although some individuals defend territories on beaches, most plovers usually forage in flocks.

Distribution and Abundance

North America - The snowy plover breeds along the coast of the Gulf of Mexico, and along the Pacific Coast of the United States and Baja California. Inland, it also breeds in the southern Great Plains, the Great Basin, San Joaquin Valley, and southern desert areas of California. Inland populations (except from San Joaquin Valley and Salton Sea) migrate to the coast for the winter. Current information suggests a breeding population of about 21,000 snowy plovers in the United States (Page et al. 1995).

Pacific Coast - Breeding and wintering occurs from southern Washington to Magdalena Bay, Baja California. Birds breeding in San Francisco Bay are considered part of the coastal breeding population. Currently, an estimated 1,900 snowy plovers breed along the west coast of the United States and at least another 1,900 along the west coast of Baja California (Page et al. 1995).
**Suisun** - At most, snowy plovers probably occur in Suisun only rarely.

**North Bay** - Small numbers of snowy plover have been found nesting on North Bay salt ponds, and have been seen in diked, seasonal wetlands (Feeney, pers. comm.).

**Central Bay** - Plovers nest at Alameda Naval Air Station, and also wintered at Alameda South Shore into the 1980s (Page et al. 1986).

**South Bay** - Although a few pairs have been found nesting on levees in salt ponds of San Pablo Bay (i.e. two pairs in 1989), the majority of snowy plovers in San Francisco Bay nest in the South Bay in salt evaporation ponds south of the San Mateo Bridge. Surveys of South Bay salt evaporation ponds and adjacent habitats tallied the following numbers of adult snowy plover during the peak of the breeding season: 1978 (351); 1984 (270); 1989 (216); and 1991 (176). The 1991 survey excluded a portion of the Baumberg Tract where 29 adult plovers had been recorded on the 1989 survey. Of all surveys, at least 87% of the adult plovers were found on the eastern side of the Bay using Guadalupe Slough as the division line.

**Historical Information**

There are no records of snowy plover breeding in San Francisco Bay prior to the conversion of salt ponds. This species may have bred on natural playas on the inner fringes of salt marsh which existed prior to conversion of South Bay marshes to salt ponds. Snowy plovers have been documented in San Francisco Bay during winter as early as the late 1800s (Page et al. 1986).

**Population Trends**

The breeding population along the western coast of the United States has declined and the breeding range has become increasingly fragmented during the past century. On surveys in the late 1970s, the species was absent from 33 of 53 California coastal locations with breeding records prior to 1970 and was missing from parts of San Diego, Ventura and Santa Barbara counties, most of Orange County, and all of Los Angeles County (Page and Stenzel 1981). Along the coast of Washington, Oregon and California combined, there was an approximate 20% decline in the size of the breeding population on surveys between the late 1970s and late 1980s (Page et al. 1991). Within San Francisco Bay, the population also appears to have declined as evidenced by the drop from 351 adults on the 1978 survey to 226 (216 in South Bay) on a 1989 survey (Page et al. 1991).

**Habitat Use and Behavior**

**Foraging** - Snowy plovers feed in typical plover fashion, usually pausing, looking, then running to seize invertebrate prey from the surface of a beach, tidal flat, or salt pan. Plovers also probe shallowly into the sand or mud for prey, or lower their heads and charge open-mouthed at aggregations of flies, snapping the bill at those that are flushed. Only anecdotal information on diet is available. In salt ponds and on beaches, flies (Diptera) are undoubtedly an important prey. On Pacific Coast beaches and tidal flats, the following prey have been recorded in the diet: mole crabs (Emerita analoga), small crabs (Pachygrapsus crassipes), polychaetes (N eodrilidae, Lumbrineris zonata, Polydora socialis, Scoloplos acmecps), amphipods (Corophium spp., Amphiope spp., Allorchestes spp.), sand hoppers (Orchestoidae spp.), tanadaceans (Leptocheila dubia), flies (Ephydridae, Dolicopodidae), beetles (Carabidae, Buprestidae, Tenebrionidae), clams (Transnordia spp.), and ostracods (Page et al. 1995). In San Francisco Bay salt evaporation ponds the following prey have been recorded: flies (Ephydra cinerea), beetles (Tanarthus occidentalis, Bembidion), moths (Perizoma custodiata) and lepidopteran caterpillars (Feeney and Maffei 1991).

**Breeding** - Feeney and Maffei (1991) monitored snowy plover nests in the Oliver Brothers/Baumberg region salt ponds during the 1989 breeding season. They located 66 nests, and 14 broods from nests which they did not find. Assuming the broods were from nests in locations they were studying, they located a seasonal nest total ranging from one nest per 1.1 hectare (ha) to one nest per 5.8 ha in four subregions of their salt pond study area. From all day watches of parent(s) with broods, they found the daily area covered by a brood ranged from 0.1-5.5 ha and averaged 1.6 ha. They noted that vegetation was used by chicks for hiding and for foraging. Northern harriers were observed hunting in areas where young snowy plovers were the only likely prey; an American kestrel was observed taking a snowy plover chick; and a peregrine falcon was seen taking an adult snowy plover. Ravens, ground squirrels, and California gulls also prey upon snowy plover eggs.

**Roosting** - In San Francisco Bay, roosts of snowy plovers occur on the salt pans of dry or partly dry salt evaporators, on barren to sparsely-vegetated interior salt pond levees, and on sandy tidal flats. Up to 300 snowy plovers have been found roosting in one salt evaporator in the Baumberg Tract during winter (Feeney and Cogswell, pers. comm.).

**Movement** - Snowy plovers are known to move between salt pond breeding, foraging, and roosting sites, and mudflat foraging sites during all seasons. A more detailed understanding of snowy plover movements in the San Francisco Bay would require tracking color-banded individuals.

**Conservation and Management**

**Contaminants Risks** - Little information is available. In 1996 several snowy plovers at Ocean Beach became oiled after a spill in San Francisco Bay.
Disturbance - Snowy plovers are disturbed by hikers, joggers, dogs off leashes which sometimes deliberately chase them, and by avian predators. It is not uncommon for plovers to fly back and forth over a roosting area in a tight flock for up to 20 minutes after having been attacked by a merlin or a peregrine falcon.

Protective measures - Several measures are used to reduce disturbance to nesting snowy plovers. Some coastal beaches have been closed or roped off. In other areas, a combination of informative signs and fencing of individual nests are used to protect snowy plover nests from predators and people. The U.S. Fish and Wildlife Service removes non-native predators at Monterey and San Francisco bays to improve snowy plover nesting success, as well as adult and chick survival rates.

Recommendations

This is a federally listed threatened species that cannot afford to lose habitat. Plans for tidal marsh restoration should attempt to encourage natural formation of salt pan habitat at bay's edge for potential plover use. A salt evaporation system should be preserved in the South Bay that produces medium to high salinity ponds, and that is large enough to support at least 300 breeding snowy plovers. Several salt pond sites should be provided, rather than one large contiguous salt pond area. South San Francisco Bay has recently had one of the largest breeding concentrations of snowy plovers on the western coast of the United States. The recovery plan for the Pacific Coast population will at minimum require that numbers of plovers in known high concentration sites not be reduced below recent levels.

Habitat elements important to snowy plover include mudflats and sandflats (used for feeding); salt pan (used for nesting and feeding); and unvegetated levees, islets, and beaches (used for nesting, feeding, and roosting). Important geographic regions are South Bay, Central Bay, and North Bay. Regional recommendations to support snowy plover are shown in Table 6.2.

Suisun - There is no documentation of snowy plover use of Suisun.

North Bay - Species nests in small numbers on North Bay salt ponds. Some habitat in this region of the Bay should be maintained; however, emphasis should be placed on enhancing habitat in the more heavily used South and Central bay areas. An area of particular importance in the North Bay is Little Island.

Central Bay - Areas of importance include the Alameda Naval Air Station and the Oakland Airport. Outboard tidal flats from Roberts Landing to San Mateo Bridge should be maintained as foraging habitat.

The Hayward Area Recreation District (HARD) wants to manage the Oliver Brothers' property for snowy plovers. California Department of Fish and Game and the U.S. Fish and Wildlife Service have agreed to this objective for this site. The goal of the management is approximately 50 breeding birds.

South Bay - The goal for the South Bay subregion should be to sustain at least 300 breeding snowy plovers. Areas of importance include East Bay salt ponds between the San Mateo and Dumbarton bridges; Salt pond systems south of the Dumbarton Bridge; and Oliver Brothers' property. Detailed recommendations for South Bay are provided in Table 6.3.

Research Needs

An up-to-date survey of the size of the breeding snowy plover population in San Francisco Bay is needed to determine if the population is continuing to decline. An effort to band snowy plovers in the San Francisco Bay area, as well as regular breeding and wintering season surveys for color-banded birds are needed to understand juvenile and adult snowy plover dispersal patterns between San Francisco Bay and coastal beaches. An analy-

<table>
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<th>Table 6.2 Regional Recommendations to Support Western Snowy Plover</th>
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<td>Habitat Type</td>
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<td>Salt Pond</td>
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<td>Intertidal Flat</td>
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<th>Table 6.3 Recommendations to Support Western Snowy Plover in the South Bay</th>
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<td>Habitat Type</td>
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<tr>
<td>Salt Pond</td>
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<tr>
<td>High Salinity Salt Ponds</td>
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<td>Intertidal Flat</td>
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sis of breeding habitat characteristics (e.g. soil salinity, vegetative cover) and presence of breeding birds or measures of nesting success would be useful to better understand plover habitat use and to guide future management efforts.

References


Personal Communications

Howard Cogswell, Professor Emeritus, California State University, Hayward, Calif.

Leora Feeney, Principal, Biological Field Services, 1330 8th St. Alameda, Calif. 94501.

Marbled Godwit

Limosa fedoa

Gary W. Page

Catherine M. Hickey

Lynne E. Stenzel

Introduction

San Francisco Bay holds the second largest known wintering concentration of (15,000-20,000) marbled godwits in the world. It is partly for this reason that the godwit was chosen as a focus species by the Wetland Ecosystem Goals Project's Shorebird and Waterfowl Focus Team. Additionally, although marbled godwits are more restricted to estuarine habitats than the willet (Catoptrophorus semipalmatus) and the American avocet (Recurvirostra americana), their habitat requirements may well represent those of all large shorebird species as a group (e.g. willet, long-billed curlew (Numenius americanus), whimbrel (Numenius phaeopus), black-bellied plover (Pluvialis squatarola), and American avocet). A comprehensive management plan for San Francisco Bay Area wetlands will need to recognize the importance of expansive tidal flats as foraging habitat during ebbing tides for large shorebird species and will need to identify important roosting and alternative foraging habitat during high tides.

Description

The marbled godwit is a large (about 45 cm), mottled, cinnamon-buff and black shorebird, with long dark gray legs and a distinctive long, bicolored, pink and black slightly upturned bill (Palmer 1967, Hayman et al. 1986). The long-billed curlew is of similar coloration and size, but has a long distinctive decurved bill. The willet is also similar in size, but is grayer and has a much shorter straight bill.
Breeding Biology – Although small numbers of marbled godwits breed in tundra areas of Alaska and James Bay (Ontario), most nest on the prairies of the United States and Canada. On the prairies, nesting probably begins in May and extends into August when the last young fledge. Clutch size is four with both sexes incubating for the suspected 21-23 day incubation period. Both sexes attend the precocial young which leave the nest permanently a few hours after hatching (Palmer 1967, Johnsgard 1981).

Migration Ecology – Autumn migration begins in early July and extends into October (and possibly November) with the earliest adults likely beginning to move about a month before the young of the year, as in other shorebirds (Shuford et al. 1989). Although the winter destination of some godwits is the Atlantic coast of Florida and the Gulf of Mexico, most travel to the Pacific coast of the United States and Mexico for winter (Palmer 1967, Hayman et al. 1986). Spring migration may begin in March but is most noticeable in April and May (Palmer 1967, Shuford et al. 1989). Except for the Great Salt Lake, where tens of thousands of marbled godwits stage during spring and fall, migrating godwits are rare during migration in interior wetlands of western North America (including the Central Valley of California; PRBO, unpubl. data). There is some migration of godwits along the Pacific Coast in spring and fall.

Wintering Ecology – This species is very restricted to coastal habitats in winter where it associates most commonly with willets, long-billed curlews, American avocets, and black-bellied plovers. The regions with the largest concentrations of wintering godwits are the coast of California, the western coast of Baja California, and the eastern coast of the Gulf of California (PRBO, unpubl. data; B. Harrington, unpubl data; Morrison et al. 1992, Page et al. 1997). There is no information available on how population numbers may have changed in San Francisco Bay historically. Continentally, the breeding range of marbled godwits has shrunk during the past 150 years, and the population has also likely declined (Page and Gill 1994).

Population Trends

There is no information available on how population numbers may have changed in San Francisco Bay historically. Continentally, the breeding range of marbled godwits has shrunk during the past 150 years, and the population has also likely declined (Page and Gill 1994).

Habitat Use and Behavior

Foraging – This species characteristically probes deep into sandy to muddy substrates for invertebrate prey. Tidal flats and sandy beaches are the principal feeding habitat with wet to shallowly-flooded pastures and lawns sometimes used on high tides. Some foraging also occurs in salt marshes and occasionally on rocky reefs. In San Francisco Bay, marbled godwits forage primarily on tidal flats and to a much lesser degree in salt marshes, seasonal wetlands, and possibly salt ponds. Prey of marbled godwits in San Francisco Bay include the marine polychaete (Neanthes succinea), the gastropod (Ilyanassa obsoleta), and the pelecypods (Gemma gemma, Mya arenaria, Mya acomma inconspicua; Recher 1966). At the coast of California, the western coast of Baja California, and the eastern coast of the Gulf of California (PRBO, unpubl. data; B. Harrington, unpubl data; Morrison et al. 1992). Very small numbers of godwits also winter at Willapa Bay on the Washington Coast (PRBO, unpubl. data).

San Francisco Bay – Marbled godwits occur in all regions of San Francisco Bay. The winter population size of as many as 20,000 individuals is the second largest known concentration of wintering marbled godwits in the world (Page et al. 1997). During migration, as many as 32,000 marbled godwits have been recorded in San Francisco Bay (PRBO, unpubl. data).

Suisun – Small numbers (low hundreds) of marbled godwits are found on the tidal flats of Suisun Bay.

North Bay – Typically about 30% of the Bay population of marbled godwits are found in San Pablo Bay (Figure 6.10).

Central Bay – Central San Francisco Bay typically holds only 10-20% of the Bay total of godwits.

South Bay – During all seasons, the largest numbers of marbled godwits occur in south San Francisco Bay, usually 50-60% of the Bay total (Figure 6.11).

Historical Information

Godwit populations in California were believed to have decreased markedly in number by 1910 due to market hunting and destruction of breeding habitat (Grinnell et al. 1918). Grinnell and Miller (1944) report that population recovery began about 1910 and was nearly complete by 1944.
Bolinas Lagoon, polychaetes are the godwit's chief prey although amphipods, decapods, small bivalves, fish, adult insects, tanaidaceans, ostracods, and gastropods are also taken (Stenzel et al. 1983). Elsewhere on the California coast, polychaetes, adult flies and gastropods have also been reported in the godwit's diet (Reeder 1951, Holmberg 1975). On sandy beaches, mole crabs (Emerita analoga) are also important prey (G. Page, pers. obs.).

**Roosting** - In San Francisco Bay, godwits congregate into flocks as large as 1,000 or more birds to roost on tidal flats when they are not covered by tides and during high tides in shallowly-flooded salt evaporation ponds, open areas in salt marsh, or other barren to moderately vegetated habitats such as salt pond levees or islands (PRBO, unpubl. data; Kelly and Cogswell 1979).

**Movement** - Marbled godwits forage in deeper water below the tide line compared to most other shorebirds and usually concentrate near the tide line as it advances and retreats over the flats. Roosting behavior is usually associated with high tides and feeding with low to moderately-high tides. In South San Francisco Bay, Kelly and Cogswell (1979) report the usual distance traveled between roosting and feeding areas to be about 1,000 m. They found color banded individuals consistently used the same feeding and roosting areas and some of these individuals took up residence for eight to nine months during the non-breeding season.

**Figure 6.10** Relative Use of Different Baylands Areas by Marbled Godwit, Willet, Whimbrel, and Long-billed Curlew Combined

Conservation and Management

Contaminant Risks - Little to no information is available.

Disturbance - Roosting or foraging birds may be flushed by people approaching too closely, by avian predators (particularly *Falco peregrinus*), or by unleashed dogs which chase them, as well as other shorebirds. Roosting birds have been flushed by jet skiers and wind surfers (N. Warnock, pers. comm.)

Recommendations

San Francisco Bay holds the largest wintering concentration of marbled godwits in the United States, and numerous large shorebirds use salt pond systems as high tide roosting and foraging habitat. Therefore, we need to preserve enough acreage of salt pond habitat to hold large numbers of all large shorebird species. Since alternative sites should be available, rather than one large contiguous salt pond area, multiple roosts should be located in all regions of the Bay.

Important habitat elements for marbled godwits include mudflats and sandflats (used for foraging and low-tide roosting); pan, beach, and unvegetated levees and islets (used for roosting); and seasonal wetlands (when wet, used for foraging and roosting). Birds also use man-made structures for roosting.

The geographic subregions of most importance to marble godwits are South Bay, North Bay, and Central...
Regional recommendations to support marbled godwits are shown in Table 6.4.

**Suisun** - Small number of marbled godwits forage on the tidal flats of Suisun Bay and roost in the managed wetlands. Areas of importance include the tidal flat of Honker Bay and the managed wetlands. All intertidal flat habitat should be maintained to provide foraging. Some shallow, sparsely vegetated managed marsh should be maintained for roosting.

**North Bay** - Areas of importance within the North Bay include all areas of tidal flats, seasonal wetlands, and salt ponds. Pilings extending into the Bay and the jetty in Carquinez Straits are important willet roosts. To maintain shorebird populations in the North Bay high tide roosting sites must be available. Table 6.5 provides detailed recommendations for the North Bay.

**Central Bay** - It is important to preserve all existing roosts and investigate the potential to establish more roosting sites in the Central Bay. All existing tidal flat outboard of marshes should be maintained, as should Marta’s Marsh and Shorebird Marsh in Marin County.

**South Bay** - All tidal flat is important for foraging and roosting at low tide. Maintain outboard flats and increase tidal flat along channels and in salt marsh. Mid- and high salinity ponds are important for roosting and foraging, and provide American avocet nesting habitat. Maintain some mid- to high salinity ponds as roosting and foraging habitat, and manage the former Oliver Brothers salt ponds for roosting shorebirds from September to March.

If the amount of salt pond habitat is reduced, maintain sufficient and multiple roosting sites for the current population of at least 100,000 large shorebirds. If we want to retain nesting sites for the American avocet, we need to retain some salt ponds or create alternate nesting habitat (salt pan).

### Table 6.4 Regional Recommendations to Support Marbled Godwit

<table>
<thead>
<tr>
<th>Habitat Type</th>
<th>Recommendation</th>
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<tbody>
<tr>
<td>Intertidal Flat</td>
<td>Maintain tidal flat outboard of marshes; Increase tidal flat in salt marshes by creating wide channels with exposed flat at low tide.</td>
</tr>
<tr>
<td>Salt Ponds</td>
<td>Maintain some shallowly-flooded salt ponds for high tide roosting.</td>
</tr>
<tr>
<td>Treatment Pond</td>
<td>(Possibly potential roosting areas.)</td>
</tr>
<tr>
<td>High Tidal Marsh</td>
<td>Create barren to sparsely vegetated areas above high tide for roosts.</td>
</tr>
<tr>
<td>Farmed/Grazed Bayland</td>
<td>(When wet, could provide high tide foraging habitat in winter.)</td>
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### Table 6.5 Recommendations to Support Marbled Godwit in the North Bay

<table>
<thead>
<tr>
<th>Habitat Type</th>
<th>Recommendation</th>
</tr>
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<tbody>
<tr>
<td>Intertidal Flat</td>
<td>Maintain all outboard flats; Increase channel tidal flat in salt marshes.</td>
</tr>
<tr>
<td>Salt Pond</td>
<td>Manage former Cargill intake pond for roosting and foraging shorebirds.</td>
</tr>
<tr>
<td>Unvegetated Shore</td>
<td>Maintain present acreage</td>
</tr>
<tr>
<td>Muted Tidal Marsh</td>
<td>Manage muted tidal diked marsh west of intake ponds and south of Napa Slough for roosting and foraging shorebirds.</td>
</tr>
<tr>
<td>Farmed/Grazed Bayland</td>
<td>Enhance seasonal wetlands as high tide foraging and roosting areas.</td>
</tr>
</tbody>
</table>

**References**


Personal Communications
Brian A. Harrington, National Biological Service.
Nils Warnock, Point Reyes Bird Observatory, Stinson Beach, California.

Black Turnstone
Arearia melanocephala
Stephen L. Granholm

Introduction
In the San Francisco Estuary, black turnstone is the most numerous of a group of uncommon shorebirds that typically use rocky unvegetated shores. Other species in this group include ruddy turnstone (Arenaria interpres), surfbird (Aphriza virgata), spotted sandpiper (Actitis macularia), black oystercatcher (Haematopus bachmani), and wandering tattler (Heteroscelus incanus). These species occur in the Estuary as migrants and winter residents. They do not breed here, except for a few pairs of oystercatchers and an occasional pair of spotted sandpipers.

Description
The black turnstone is a short-legged, short-billed shorebird, about 23 cm long, with a blackish back, chest, and legs, and a white belly. In feeding plumage it has some white speckling and a white dot in front of the eye. It displays a striking black and white harlequin pattern in flight.

Distribution and Abundance
North America/Pacific Coast – Black turnstones are found exclusively along the Pacific coast of North America. They breed in western and southern Alaska and winter from southeastern Alaska south to southern Baja California and central Sonora, Mexico (Johnsgard 1981). A few occur inland during migration.

Black turnstones are present along the California Coast from about mid-July to mid-May (Cogswell 1977, McCaskie et al. 1979), and small numbers of nonbreeders remain through the summer (Zeiner et al. 1990).
San Francisco Bay – Total counts of black turnstone for the Estuary have ranged from 40-137 birds in fall, 69-144 in winter, to 212 birds in spring (PRBO, unpubl. data).
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Probably these counts underestimated black turnstone numbers, however, because the counts focused on intertidal mudflats and sandflats, rather than rocky unvegetated shores. Black turnstones do not breed in the Bay Area.

**Suisun** - Apparently, this species does not occur regularly east of the Carquinez Bridge.

**North Bay** - Black turnstones are found along the eastern shore of San Pablo Bay, primarily south of Hercules. Few are found on the western and northern shores of San Pablo Bay (north of Point San Pedro; Figure 6.12).

**Central Bay** - This species occurs widely throughout the Central Bay, in suitable habitat (Figure 6.12).

**South Bay** - In South Bay, black turnstones are found primarily north of the San Mateo Bridge, but smaller numbers are found farther south (Figure 6.12).

**Historical Information**

No information.

**Population Trends**

No information.

**Habitat Use and Behavior**

Foraging - On their wintering grounds, black turnstones feed primarily on rocky unvegetated shores, including rocky breakwaters and riprap, as well as natural rocky shorelines. They also feed on intertidal mudflats, sandflats, and sandy beaches.
Black turnstones pick food from the surface or turn over seaweed, rocks, or shells to search for prey. They feed mostly on barnacles, limpets, and other small mollusks and crustaceans (Johnsgard 1981, Paulson 1993). Black turnstones usually forage and roost in small flocks of a few birds to a few dozen (Paulson 1993).

**Roosting** - They roost primarily on rocky unvegetated shores above the tide level, on relatively undisturbed sites such as breakwaters, islets, and inaccessible shorelines.

**Movement** - Black turnstones move from their foraging grounds to nearby roosting sites at high tides.

### Conservation and Management

**Contaminant Risks** - No specific information.

**Disturbance** - Frequent disturbance by people and dogs could reduce this species' ability to accumulate sufficient energy reserves for migration.

### Recommendations

The black turnstone population in the San Francisco Estuary is probably limited by the availability of rocky intertidal habitat with an adequate food supply. Natural rocky shorelines are very limited in extent, and riprapped shorelines probably have less abundant invertebrate prey than natural shorelines. Like other shorebirds, this species also requires undisturbed roost sites.

Regionwide goals should include maintaining or increasing population levels within the species' current range in the San Francisco Estuary. This will require preservation of natural and semi-natural rocky shorelines and other important feeding and roosting areas, especially areas with an abundant invertebrate population. Riprap habitat is also used by these birds, but a loss of this habitat is acceptable if unavoidable as part of tidal marsh restoration. Preservation of roosting areas must include protection from disturbance by people and dogs.

Areas of importance to black turnstones are primarily found in Central Bay, North Bay (eastern portion), and South Bay (northern portion). Important habitat elements are rocky shore and unvegetated levees and islets (used for roosting). Some of the shorebirds associated with rocky substrates, in particular, ruddy turnstones, also forage on mudflat and sandflat. Regionwide goals thus should include maintaining current acreages of unvegetated shore and intertidal flats.

**Suisun** - Maintain current acreage of unvegetated shore and intertidal flat.

**Central Bay** - Maintain current acreage of unvegetated shore and intertidal flat.

**South Bay** - Maintain current acreage of unvegetated shore and intertidal flat.

### Research Needs

Future surveys should map rocky intertidal habitats in the Estuary and identify significant feeding and roosting areas for black turnstones and other rocky intertidal shorebirds. Foraging studies should be conducted to determine the relative importance of natural and semi-natural rocky shorelines, compared to riprap and other habitats.

### Acknowledgments

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### References


### Important Data Sets


Red Knot
Calidris canutus
Catherine M. Hickey
Gary W. Page
Lynne E. Stenzel

Introduction
The red knot is a high-arctic breeder and a long distance migrant. Knots are most abundant on the Pacific coast of North America during spring migration and less abundant during fall (Page et al. 1979, Paulson 1993). In winter, significant numbers (hundreds to thousands) of knots appear to be localized in distribution into three areas on the Pacific coast of North America: San Francisco Bay, San Diego Bay, and Laguna Ojo de Liebre, Baja California, Mexico (PRBO, unpub. data). In San Francisco Bay Area wetlands, knots are also very localized in distribution. The importance of San Francisco Bay, and of specific sites in the region, to the red knot should warrant special concern for this species in any comprehensive management plan for San Francisco Bay Area wetlands. Such a plan should include a combination of extensive intertidal flats as foraging habitat for the knot, and adequate, undisturbed high tide roost sites. Because red knots frequently associate with dowitchers (Limnodromus spp.), dunlins (Calidris alpina) and black-bellied plovers (Pluvialis squatarola), management plans to preserve, enhance, or restore habitat for red knots may also benefit these other species.

Description
The red knot is the second-largest calidrid, about the same size of dowitchers. Knots look heavy and rounded in shape (Hayman et al. 1986). The bill is blackish and faintly downcurved, the iris is dark brown, and the legs are rather short. In nonbreeding or juvenile plumage, knots have mostly white underparts, rather plain gray upperparts, gray streaking on the breast and upper belly, and greenish legs. Breeding-plumaged adults have dark gray legs, chestnut-red face and underparts, gray and black speckled backs, and white undertail coverts (Paulson 1993). Knot flight feathers are blackish, with distinct but narrow white wing bars, the rump is pale gray, and the tail shows broad brown bars and narrower white ones (Hayman et al. 1986).

Breeding Biology - Red knots breed inland on moist tundra and upland glacial gravel (Hayman et al. 1986). Nesting begins in late May and extends into August when the last young fledge (Harrington 1996). Knots lay four eggs at approximate one day intervals. Incubation lasts 21-23 days. Both sexes incubate, but the males primarily tend the brood. The fledgling period is approximately 18 days (Johnsgard 1981).

Migration Ecology - Red knots migrate long distances and concentrate in fewer areas than most other arctic breeding shorebirds. They are primarily coastal migrants, occurring only rarely inland in spring and fall. They are strictly coastal in winter (Hayman et al. 1986).

Wintering Ecology - Like the majority of shorebirds in San Francisco Bay, red knots feed primarily on tidal flats. At high tide they roost in flocks, particularly in salt ponds in South San Francisco Bay.

Distribution and Abundance
North America - There are three recognized subspecies of red knots that occur in North America. C. c. islandica breeds in the high arctic of Canada and Greenland and winters in Great Britain, France, and the Wadden Sea. C. c. rufa breeds in the central Canadian arctic and winters in southeastern North America, the Caribbean, parts of the northern coast of South America, and in the southern parts of South America. Harrington et al. (1988) estimate a total population of 100,000-170,000 for this subspecies. Little is known about the third subspecies, C. c. roselaari, which is believed to breed in northern Siberia on Wrangel Island and in Alaska and to migrate and winter along the Pacific coast of North America (Tomkovich 1992). Currently, there are no population estimates for this subspecies. A fourth subspecies, C. c. rogersi, breeds in northeastern Siberia and perhaps in Alaska, and winters primarily in Australia and New Zealand but possibly also on the Pacific coast of South America.

Pacific Coast - Red knots are significantly more abundant along the Pacific coast of North America during spring migration than during fall migration. Grays Harbor and Willapa Bay in Washington appear to be important staging areas for the knot in spring migration. The fall migration route of the large numbers of knots that migrate along the coast during the spring is not well-known (Paulson 1993). In winter, significant numbers of knots appear to have a localized distribution into three main areas: San Francisco Bay, San Diego Bay and La-
guna Ojo de Liebre, Baja California, Mexico. Knots do not winter regularly in the Pacific Northwest.

San Francisco Bay - Small numbers of red knots occur scattered in various wetland habitats around San Francisco Bay. Larger numbers of knots (hundreds to over one thousand) have a more localized distribution in the Bay. Within San Francisco Bay, 18 to 2,100 red knots were found on Point Reyes Bird Observatory (PRBO) counts.

Suisun - Probably uncommon to rare in Suisun.

North Bay - Large numbers of knots were recorded on intertidal flats just north of Pinole Point, Contra Costa County, but were rarely found elsewhere in San Pablo Bay on PRBO counts (Figures 6.13 and 6.14; PRBO counts).

Central Bay - Knots were reported on intertidal flats just north and south of the San Mateo Bridge, Alameda County. Up to 100% of the San Francisco Bay population was found in the Central Bay on PRBO counts (Figure 6.13; PRBO counts).

South Bay - Knots have been reported in two salt pond complexes in Alameda County (PRBO, unpubl. data). Up to 100% of the San Francisco Bay total was found in the South Bay (Figures 6.13 and 6.14; PRBO counts).

Historical Information

There is no historical population estimate for red knots in San Francisco Bay. Red knot populations in North
America suffered from intense hunting. Since legal protection was afforded, their status has improved.

**Population Trends**

No information.

**Habitat Use and Behavior**

**Foraging** – Knots are specialized feeders, requiring concentrated, rich food resources (Harrington 1996). Knots typically forage in cohesive flocks, feeding mainly by probing, but also by pecking at surface foods (Hayman et al. 1986). In Bolinas Lagoon, Marin County, California, Stenzel et al. (1983) found that several bivalve species (Transennella spp., Protothaca spp., Gemma spp., Macoma spp., Mytilus spp., and Clinocardium spp.) were the knot’s primary prey. Other prey items included errant polychaetes (Lumbrinerius spp. and Glycinde spp.), gastropods, alga-dwelling and tube-dwelling amphipods, and trace amounts of small crustaceans, Foraminifera, tanaidaceans, and sedentary polychaetes. Other shorebird diet analyses have also determined molluscs to be the primary prey item of red knots on the western coast of North America (Sperry 1940, Recher 1966). On the Atlantic Coast, eggs of horseshoe crabs (Limulus polyphemus) are a staple. Adult and larval insects, as well as plant material are the primary prey items on the breeding grounds.

**Roosting** – Red knots are known to form dense, sometimes mixed, roosting flocks. In San Francisco Bay,
salt evaporator ponds in the Hayward area appear to be important roosting habitat for knots.

Movement – Little is known about red knot movement between habitat types in the San Francisco Bay Area.

Conservation and Management

Contaminant Risks – Little or no information is available, but because red knots are localized in distribution, they are particularly vulnerable to contaminants and potential oil spills.

Disturbance – Red knots, like most other shorebirds, may be flushed by people approaching too closely, by avian and terrestrial predators, or by unleashed dogs that chase them. Knots should warrant special concern in development and management plans in the San Francisco Bay region.

Recommendations

San Francisco Bay is one of three main red knot wintering areas along the western coast of the United States. Since this species is relatively rare compared to other species occurring in the Bay, special consideration should be given to its habitat requirements and to specific areas of known use. Its habitat requirements should not be expected to be an umbrella for the associated species. Salt ponds currently used by red knots for roosting may also be used as alternate foraging habitat.

Within the Bay, areas of importance to red knot are found in North Bay, Central Bay, and South Bay. Mudflat and sandflat are important for roosting and foraging, and unvegetated levees, islets, beaches, and pans are used for roosting. Goals for this species should include maintaining tidal flat outboard of marshes for foraging habitat, and some salt ponds on the eastern side of the South Bay for roosting sites.

Suisun – There is no documentation of red knots using Suisun.

North Bay – The tidal flats between Point Pinole and Carquinez Straight are important and should be maintained. Investigation is needed to identify the important roosting areas in the North Bay subregion.

Central Bay – Central Bay tidal flats along the Hayward shoreline appear to be the most important site for red knots in the entire Bay. Roosting areas for red knots in the Alameda area need to be identified. All tidal flats on the eastern side of the Bay should be maintained and improved. The Oliver Brothers salt ponds should be maintained as roosting habitat.

South Bay – Baumberg/Hayward area salt ponds are known to hold roosting red knots. Though salt ponds are primarily for roosting, they also may be used as alternate foraging areas when intertidal flats are covered at high tide.

All tidal flat should be maintained for foraging, and the amount of flat along channels in tidal marsh should be increased. Some salt ponds should be preserved for roosting sites, especially in the Baumberg area. The Oliver Brothers salt ponds should be maintained as roosting habitat.

Research Needs

Substrate and invertebrate sampling from various intertidal flats around San Francisco Bay and San Pablo Bay would help determine whether known areas of consistent red knot concentration have a unique combination of substrate and/or food resources.

References


Western Sandpiper
*Calidris mauri*
Nils Warnock
Sarah E. Warnock

**Introduction**

The western sandpiper is the most abundant shorebird of California during fall and spring migration and the second most abundant during the winter. Largest concentrations of western sandpipers occur at coastal estuaries, but during the spring and to a lesser extent the fall, interior sites such as the western Great Basin, the Central Valley, and the Imperial Valley may host tens to hundreds of thousands of migrants. In winter, this species is more restricted to marine habitats than associated species, including least sandpiper, dunlin, and semipalmated plover. All the above species use tidal flats, salt ponds, managed wetlands, and seasonal wetland habitats in the Bay.

**Description**

Western sandpipers are marginally the largest of the stints. The species is sexually dimorphic with males typically smaller in size than females (Page and Fearis 1971). In basic plumage, they are pale grey with brown streaking on the crown, with white underparts. Fine, blackish streaks often join across the breast, usually heavier streaking than in semipalmated sandpipers with whom they share the special feature of partial webbing between the toes. Their typically longer, heavier, and decurved bill is also a helpful distinguishing factor. During breeding season, they have a distinctive dark streaking on their breasts with “V’s” or arrowheads on flanks, and usually much more rufous in upper scapulars and head than other species likely to be encountered.

**Breeding Biology** – Western sandpipers are monogamous, bi-parental sub-Arctic and Arctic breeders. At least some begin breeding after their first year (Holmes 1971). Generally, they only lay one clutch per season, but will lay a replacement clutch if the first is lost early in the breeding season. Mean clutch completion in western Alaska occurs between 25 May and 5 June (Holmes 1972). Western sandpipers generally lay four eggs. Favored nesting habitat is moist to wet graminoid tundra, but they will also occasionally breed on lower mountain slopes (Wilson 1994). They nest in depressions in the ground typically lined with grass and under a dwarf birch (Wilson 1994). Mean incubation period is 21 days (Holmes 1973). Hatching success is 84% at the Yukon-Kuskokwim Delta, Alaska (Holmes 1972) and 55% at Nome, Alaska (Wilson 1994). Major nest predators include jaegers and foxes.

**Migration Ecology** – The global distribution of western sandpipers is skewed by sex. Female birds are more likely to winter at more southerly sites than males (Page et al. 1972, Harrington and Haase 1995). In California and San Francisco Bay, winter populations of western sandpipers are skewed towards males (Page et al. 1972).

At San Francisco Bay, during fall migration, adults arrive about a month earlier than juveniles (late June, July); followed by juveniles (August through October). During spring migration, peak numbers occur from 20 April though 5 May; males are followed by females (N. Warnock, unpubl. data). The length of stay at San Francisco Bay of northward migrating western sandpipers radio marked there was 9.1 ± 4.6 days (n = 58 birds), although these stays may have been prolonged due to temporary effects of capturing the birds (Bishop and Warnock 1998, Warnock and Bishop 1998). Birds radio marked at San Francisco Bay have been detected at all major coastal estuaries between the Bay and the Yukon-Kuskokwim Delta (with the exception of sites in Oregon, a state not monitored). Breeding destination of many of the birds migrating through the Bay appears to be the Yukon-Kuskokwim Delta (Bishop and Warnock 1998). Breeding birds from the Seward Peninsula also have been detected at San Francisco Bay, as well as wintering birds from Panama and Mexico (Butler et al. 1996). Spectacular concentrations of western sandpipers are found during the northward spring migration with hundreds of thousands of birds concentrating at such sites as San Francisco Bay, CA, Grays Harbor, WA, Fraser River Delta, BC, Stikine River Delta, Alaska and the Copper River Delta, Alaska (Iverson et al. 1996). During spring, exchange of western sandpipers between Bolinas Lagoon (an estuary approximately 14 km northwest of San Francisco Bay) and North San Francisco Bay was found (Warnock, unpubl. data).

**Wintering Ecology** – First-year birds (26.6 ± 3.6 km²) have larger winter home ranges than adults (17.2 ± 2.5 km²), but no significant differences in size of home...
range were found between sexes (Warnock and Takekawa 1996). Individual birds are strongly site-specific within San Francisco Bay.

**Distribution and Abundance**

**North America** - Western sandpipers breed in western Alaska from the Alaska Peninsula up to the North Slope of northern Alaska. Small numbers also breed in northeastern Russia. They commonly winter from Washington through the southeastern United States, and south through Mexico and Central America to northern Peru.

**Pacific Coast** - The largest winter concentrations are found from San Francisco Bay south to Panama. Areas of largest concentrations include San Francisco Bay, Lagunas Ojo de Liebre and Guerro Negro in Baja California, Mexico (Page et al. 1997), western coast of Mexico in the Gulf of California, and coast of Panama.

**San Francisco Bay** - Western sandpipers are found in all parts of the San Francisco Bay Estuary. A comprehensive April count of shorebirds counted over 555,000 western sandpipers (Stenzel and Page 1988). As many as 707,000 were found during five additional spring counts. Largest numbers were found in areas of the South Bay with large expanses of mudflats at low tide backed by salt pond complexes (Figures 6.15 and 6.16).

**Suisun** - Point Reyes Bird Observatory (PRBO) surveys indicate as many as 5,000 western sandpipers in

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**Figure 6.15** Maximum counts of Western Sandpiper, Least Sandpiper, and Dunlin Combined

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North Bay – Sixteen percent of the San Francisco Bay total were found in San Pablo Bay (north of the Richmond Bridge). The North Bay accounted for up to 30% of the Bay total on PRBO counts.

Central Bay – Up to 30% of the Bay total for western sandpipers were found between the Richmond and San Mateo bridges on PRBO counts.

South Bay – Up to 79% of the Bay total were found south of the San Mateo Bridge on PRBO counts.

Historical Information

No information.

Population Trends

Rigorously estimated global population counts for western sandpiper are not available, but the West Coast population is estimated to be a few million birds (Page and Gill 1994). Population trends are currently unknown.

Habitat Use and Behavior

Based on a sample of 106 radio marked western sandpipers in South San Francisco Bay, birds showed seasonal differences in habitat preference both on local and regional scales (Warnock and Takekawa 1995). During winter at low tide, salt pond levees followed by mudflats...
were most preferred, and salt marsh plains were used the least (Warnock and Takekawa 1995). At high tide, saltpond levees were ranked as the most preferred habitat, followed by drained salt ponds, tidal salt ponds and seasonal wetlands, and salt marsh plains were the least preferred habitat. In spring at low tide, use of habitats was significantly different from the winter. Western sandpipers preferred tidal sloughs over tidal salt ponds, drained salt ponds, and mudflats. At high tide, they preferred drained salt ponds and seasonal wetlands. At both tides, salt marsh plains were the least preferred habitat. At low tide, in all seasons, the majority of western sandpipers occur on tidal flats.

**Foraging** - Western sandpipers feed mainly on invertebrates but occasionally will also feed on small fish and plant matter. They are tactile feeders that typically probe in the mud for prey. On the breeding grounds at the Yukon-Kuskokwim Delta, Alaska, common food items include Diptera larvae, adult Coleoptera, and arachnids depending on the time in the breeding season (Holmes 1972). On the winter grounds, common food items include amphipods, small bivalves, annelids, and insects (Stenzel et al. 1983, Wilson 1994). At San Francisco Bay, they have been found to feed on a diverse diet of amphipods, bivalves, polychaetes, ostracods, and gastropods (Recher 1966). In salt ponds, they also feed on brine flies, insects, and seeds (Murie and Bruce 1935, Carpenter 1957, Anderson 1970). In the winter, birds have been found out on low tide mudflats during day and night (Warnock and Takekawa 1996).

**Roosting** - Roosting flocks sometimes number tens to thousands of individuals. Roosting sites tend to be barren to sparsely vegetated and include salt pond levees, dry to very shallow salt ponds, islands, beaches, diked baylands, etc.

**Movement** - Radio-tracking studies show species to be very limited in its movements within the Bay, but birds are known to move back and forth between Bolinas Lagoon and North San Francisco Bay during spring. Birds radio marked in South San Francisco Bay were not found outside the South Bay despite extensive search efforts in surrounding areas; birds typically move between Bay mudflats at low tide to salt pond roost areas at high tide regardless of time of day (Warnock and Takekawa 1996).

**Conservation and Management**

**Contaminant Risks** - Due to their small home ranges within the Bay, there is an increased potential for repeated exposure to environmental contaminants from industrial and municipal discharge (Warnock and Takekawa 1996).

**Disturbance** - Feeding and roosting flocks take flight in response to hunting raptors, sudden loud noises, and close approach by humans, pets, jet skis, and boats.

**Introduced Species** - Introduced invertebrates such as Potamocorbula amurensis (Nichols et al. 1990) could significantly influence the distribution and abundance of western sandpipers within the Bay.

**Recommendations**

The dynamics of the active salt industry provide a varied habitat for these small shorebirds. They need plenty of intertidal foraging habitat at low tide, and supertidal roosting and foraging areas at high tide. Seasonal wetlands, diked wetlands, muted tidal wetlands and salt ponds provide these supertidal functions. Salt ponds, especially when drained, provide habitat for huge numbers of birds. Any kind of habitat with muted tidal flow and less than 50% vegetative cover is important foraging habitat. Allowing these sites to become revegetated will likely negatively affect western sandpipers. Farmed and grazed baylands are crucial in extreme events such as severe winter storms or unusually high bird numbers, but an increase in this kind of habitat should not be at the expense of marsh habitat. Any expansion of tidal or supertidal habitats should not be done at the expense of intertidal mudflat habitat, which remains the most important feeding area for these birds.

To support the western sandpiper, the current acreage of tidal marsh should be increased. Mid-tidal and low tidal marsh should have no more than 60-70% vegetative cover. The current acreages of agricultural baylands, intertidal flat, treatment pond, and mid- and high salinity salt ponds should be maintained. Lagoons are important during low tide, and the acreage should be increased. Current acreages of low salinity salt pond and tidal reaches in streams should be increased. These general recommendations apply throughout the sub-regions.

**Suisun** - General recommendations given above.

**North Bay** - Areas of importance in the North Bay include all areas of intertidal flat, the exposed banks of Sonoma Creek and Napa and Petaluma rivers, tidal marsh channels, drained and shallow salt ponds, salt crystalizers, and Ponds 1A, 2A, and 4A. The general recommendations given above also apply to this subregion.

**Central Bay** - Areas of importance in the Central Bay include all areas of intertidal flat, roosting islands, and the Hayward marshes. The general recommendations given above also apply to this subregion.

**South Bay** - Areas of importance in the South Bay include all of the salt ponds (especially when drained), intertidal areas, seasonal wetlands, levee roost sites, diked marsh, and duck clubs. Particular locations include the Knapp property; Mowry, Newark, and Coyote sloughs; the upland marsh of tract 102 marshes, the Alviso Ponds, and Ponds RSF 1 and RSF 2. The general recommendations given above also apply to this subregion.
Research Needs

Currently, we have a poor understanding of where migrants that pass through San Francisco Bay in the spring are coming from, as well as where southward moving birds are going. The use of salt ponds by western sandpipers ought to be examined in more detail, as well as how invasive invertebrate species impact the distribution of western sandpipers within the Bay. The impact of revegetation of reclaimed salt ponds on western sandpipers using the ponds should be studied.

References


Additional Readings


**Long-Billed Dowitcher**

*Limnodromus scolopaceus*

John Y. Takekawa
Sarah E. Warnock

**Introduction**

The San Francisco Estuary supports large wintering populations of the long-billed dowitcher numbering in the low tens of thousands. More than most other abundant shorebirds, this species concentrates in fresh and brackish water wetlands. The key habitat for this bird is managed wetlands. It associates with dunlin (*Calidris alpina*), greater and lesser yellowlegs (*Tringa melanoleuca* and *T. flavipes*), black-necked stilt (*Himantopus mexicanus*), and American avocet (*Recurvirostra americana*), which also use these habitats.

**Description**

The long-billed dowitcher is a medium-sized shorebird (28-32 cm) with short legs and a long snipe-like bill (5.7-7.6 cm). In breeding plumage, long-billed dowitchers have cinnamon underparts with bars of black on the sides of the breast and flanks; a white wedge on the rump; black, buff, and white feathers above; and dull olive legs (Hayman et al. 1986). Although difficult to distinguish from the similar-looking short-billed dowitcher (*L. griseus*), it was identified as a separate species on the basis of morphological (Pitelka 1950) and genetic data indicating their relatively large congeneric divergence (Avise and Zink 1988). About 15% of the long-billed dowitchers are separable from short-billed dowitchers by a bill more than two times longer than the head (Richards 1988), but more consistent characteristics distinguishing the species include the “keek” vs. the “tu-tu-tu” call (Miller et al. 1984), white tail bars narrower than or equal in width to the dark tail bars, darker unspeckled throat and breast sharply distinct from underparts, foreneck densely spotted, center of breast barred, and belly lightly spotted (Wilds and Newlon 1983). Juveniles are distinguished by the buff appearance of their underparts and a grayer head and neck with narrow rusty fringes on their coverts.

**Breeding Biology** - Long-billed dowitchers do not breed in the San Francisco Bay Area. They are arctic breeders that nest in grassy or sedgy marshes, or near small lakes (Hayman et al. 1986). They arrive during late April and May, and their breeding displays include low-level hovering flights and a distinctive “pee-ter-wee-too” call. They defend comparatively small territories, and nest in small clumps of low sedges, in mounds of moss and sedge, or on the ground. They lay four eggs (olive with many elongated spots), and both sexes incubate the nest for the 20-21 day incubation.

**Migration Ecology** - Females form postbreeding flocks as early as late June, while males depart their breeding areas in late July or early August. Juveniles migrate 1-2 months later (Campbell et al. 1990). They peak in autumn migration in British Columbia during September and October (Campbell et al. 1990). The long-billed dowitchers are generally 5-6 weeks behind the short-billed dowitchers during migration. Long-billed dowitchers generally migrate in smaller flocks than short-billed dowitchers. Flocks of 10-100 birds are commonly observed (Campbell et al. 1990). They migrate southeast to the Atlantic Coast and along the Pacific Coast or through the midcontinent. Groups of more than 30,000 have been counted at Malheur National Wildlife Refuge in interior Oregon (Gilligan et al. 1994). Their peak spring migration lasts from late March to mid-May, and they are rare east of the Mississippi River. They are seen in Oregon during late February and early March (Gilligan et al. 1994). They are rare migrants to Japan and Europe and are vagrant to Nova Scotia, Bali, Brunei, and Thailand.

**Wintering Ecology** - Adult long-billed dowitchers begin arriving at Point Reyes, Marin County, California in July (Shuford et al. 1989), while juveniles arrive in mid-September. They winter in the southern United States and in Central America, as far south as Panama (Hayman et al. 1986). Those observed in inland areas in California are usually long-billed dowitchers (Cogswell 1977), but both species are found in coastal areas. They disappear during rainy periods on the coast, perhaps to the Central Valley (Shuford et al. 1989, Shuford et al. 1998).

**Distribution and Abundance**

**North America** - Long-billed dowitchers breed in coastal northeast Siberia, western and northern Alaska, the northwest McKenzie River Delta (Cogswell 1977), and the Northern Yukon (Richards 1988). They winter in northwest and north central California, western Nevada, southern Arizona, and east to South Carolina and Florida, as far south as Guatemala and Panama, and as
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Pacific Coast – Long-billed dowitchers are found in both coastal and interior regions, including the Central Valley and in California lakes. Few birds are known to migrate along the western Pacific, but the San Francisco Bay Estuary is used during both migration and the winter. Estuaries in Sinaloa, Mexico also include large populations of this species (Engilis et al. 1998).

Suisun – Point Reyes Bird Observatory (PRBO) recorded up to 11,200 dowitchers in August, 6,000 in November, and 7,900 in April. This includes both Limnodromous species, but collections in San Francisco Bay (Takekawa and Warnock, unpubl. data) suggest most are long-billed dowitchers (Figures 6.17 and 6.18)

North Bay – Present, common.
Central Bay – Present, low abundance.
South Bay – Present, common.

Historical Information

There was a decline in numbers of dowitchers in eastern North America and California in the late 1880s and early 1900s because of hunting (Page and Gill 1994), until the passage of the Migratory Bird Treaty Act of 1918. The size of the populations have likely followed the abundance of mudflat habitats with benthic invertebrate prey, and this species may have benefitted by the development of salt evaporation ponds which provided high tide roost sites.
Population Trends

In Florida, there has been an upward trend in recent years (Stevenson and Anderson 1994), but it is not known whether this reflects shifting populations or true population increases. No information is reported about the long-term population trends of the West Coast.

Habitat Use and Behavior

Long-billed dowitchers prefer freshwater habitats over brackish and intertidal areas (Campbell et al. 1990, Cogwell 1977, Pitelka 1950, Takekawa and Warnock in press). They use seasonal wetlands, flooded fields, reservoir edges, sewage lagoons, small ponds, lake and pond muddy shores, river banks, and sewage ponds (Campbell et al. 1990). They use intertidal flats for foraging and roosting, and roost on offshore rocks and islands (Campbell et al. 1990). At Nelson Lagoon, Alaska, long-billed dowitchers were found in muddier areas than short-billed dowitchers, which were found in sandier areas (Gill and Jorgenson 1979), but in Bolinas Lagoon, California, they were found in areas of moderately to well-sorted fine sand (Page et al. 1979). Flight surveys in the Central Valley of California found 66% of dowitchers in managed wetlands, 28% in agricultural lands, 3% in sewage and evaporation ponds, and 3% in other habitats (Shuford et al. 1993).

On the East Coast, they primarily use freshwater habitats, especially impoundments, salt pans (Veit and
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Petersen 1993), or shallow impounded waters with scant vegetation (Tomkins 1961). They are most commonly found on "barrow-pit habitats" of soft dredge material in the Midwest, usually disturbed sites, often successional areas with short life spans (Tomkins 1961).

Foraging - Long-billed dowitchers are probing feeders on muddy substrates. Sand may reduce the success in prey capture or detection in comparison to muddy sites (Quammen 1982). They feed with exploratory jabs (depth 1/3 bill length) and acquire prey with prolonged and vigorous probing (depth to bill length) (Burton 1972) with 99% of their foraging in the sediment (Young 1989). The appearance of their foraging has been described as "sewing-machine" activity (Richards 1988).

They commonly forage in salt water during migration (Cogswell 1977), but they are most commonly reported as associated with freshwater wetlands. They often forage in water up to their belly, and were found in depths of 2-8 cm in the Central Valley, California (Elphick and Oring 1998), 4-5 cm on South Island, South Carolina (Weber and Haig 1996), and 0-16 cm at Playa Lakes of Texas (Davis and Smith, 1998). On Seward Peninsula, Alaska, dowitchers are often found in salt grass meadow (Kessel 1989). In pastures along coastal California, dowitcher abundance is negatively correlated with vegetation height (Colwell and Dodd 1995). Their time-activity budget includes 79.6% feeding, 17.4% resting or preening, and 1.9% alert (Young 1989). Their breeding area diet includes meltwater pool insects, seeds, moss, and cranely larvae, while migration and wintering diets include midge and fly larvae, worms, and burrowing crustaceans.

Roosting - Long-billed dowitchers probably roost in shallow water or on barren to sparsely vegetated islands and levees. They have also been seen roosting in salt marsh (G. Page, pers. comm.). Roosting flocks sometimes exceed 1,000 birds. They commonly use intertidal flats for foraging and roosting, and roost on offshore rocks and islands (Campbell et al. 1990).

Movement - Long-billed dowitchers generally follow a pattern of feeding on intertidal flats during low tide and roosting in adjacent wetlands or uplands during high tides.

Conservation and Management

Contaminant Risks - Pesticide and other contaminant levels are available for dowitchers, but interpretation of results is difficult since no experimental work on the effects of these toxins on either breeding or migrating and wintering birds is available. Compared with birds in Texas, dowitchers collected at San Francisco Bay had higher selenium levels and elevated aluminum levels (C. Hui, J.Y. Takekawa, and S.E. Warnock, unpubl. data).

Disturbance - Long-billed dowitchers feed in intertidal flats and roost on nearby uplands or shallow wetlands. They are likely to be disturbed regularly by raptors, loud noises, and close approach by humans and their pets. Page and Whitacre (1975) determined that sixteen percent of dowitchers wintering at Bolinas Lagoon, California were killed by raptors.

Recommendations

Information suggests that these birds prefer fresher water habitats over brackish and intertidal habitats. Seasonal wetlands and freshwater ponds are therefore more important to this species. These species are commonly found on soft, dredged-material habitats or disturbed sites.

Unvegetated levees and islets, pans, and seasonal and perennial ponds are important roosting habitat. Mudflats, pans, and seasonal and perennial ponds are important for feeding. General regional goals to support dowitchers include increasing mid- and high tidal marsh for roosting, maintaining the current acreage of intertidal flat and salt pond, maintaining the current acreage of agricultural baylands (or increasing the areal extent of seasonal wetlands on a smaller area), and increasing the acreage of diked and managed marsh.

Suisun - Within Suisun, these birds are found primarily in managed marshes, but the timing and numbers are not well known. The goals for Suisun should include maintaining current acreages of muted tidal marsh and intertidal flat, and increasing mid-tidal marsh, diked marsh, and managed marsh to increase shallow water habitat.

North Bay - Seasonal wetlands and freshwater ponds are more important to this species in the North Bay. There is limited information available on the use of North Bay salt ponds, and also farmed and grazed bayland. Ponds SF1 and SF2; all low salinity salt ponds; salt ponds and mudflats south of the San Mateo Bridge, and the Knapp property. The current acreages of muted tidal marsh and intertidal flat should be maintained, and shallow marsh, mid-tidal marsh, and diked marsh should be increased.

Central Bay - Within the Central Bay there are limited areas of use, and unknown roost areas. The general regional goals above apply wherever applicable.

South Bay - The South Bay is the most important area in the Estuary for the long-billed dowitcher. Particular areas of importance include Ponds R1, R2, SF1, and SF2; all low salinity salt ponds; salt ponds and mudflats south of the San Mateo Bridge, and the Knapp property. The current acreages of low and mid-salinity salt ponds, muted tidal marsh, and intertidal mudflat should be maintained; and acreages of mid-tidal marsh, shallow marsh, diked marsh should be increased.
**References**


Wilson's Phalarope
Phalaropus tricolor
Janet T. Hanson

Introduction

Wilson’s phalarope is representative of the group of shorebird species concentrated in salt ponds, including red-necked phalarope, American avocet, and black-necked stilt. Phalaropes forage by swimming in tight circles, or “spinning” on the ponds’ surface to stir up prey items, a difference that sets them apart from other shorebirds. Wilson’s and red-necked phalaropes are present on the Bay during fall and spring migration.

No precise information exists on local numbers of these species in the early part of this century, but current surveys suggest that they increased significantly with the development of the salt pond system, which they utilize as both foraging and roosting habitat. In addition, stilts and avocets are present year-round and make use of the levees and islands in the pond system for breeding habitat. Wilson’s phalarope has suffered loss of breeding habitat in the inland wetlands it favors. On a global scale, this species appears to be declining in numbers.

Description

Wilson’s phalarope is a medium-sized (22-24 cm) shorebird with mostly white plumage, short legs and a mid-length straight bill. This species is characterized by its reverse sexual dimorphism: the female is larger and more colorful in breeding plumage. In non-breeding plumage, both sexes have gray upperparts, pure white underparts, and yellow legs. There is a white superciliary line with a strong gray post-ocular line above. In breeding plumage, the females’ crown and nape are pale gray. A strong black stripe runs from the bill, through the eye and down the neck. The breast is cinnamon or rusty color with a white belly and the legs are black. The breeding plumage of the male is a duller version of female (Colwell and Jehl 1994).

Breeding Biology - Wilson’s phalarope breeds on seasonal wetlands and freshwater marshes in the North American grasslands and prairies. This species is one of 13 species of shorebirds reported to practice polyandry. Females arrive first on the breeding grounds in late April-early May, followed by mixed flocks (Bent 1927). Several females often court a single male. Eggs are laid by mid-May to early June; a single nest typically contains four camouflaged eggs, buff colored with brown blotching. The male incubates while the female defends the territory. Incubation lasts approximately 23 days. The male broods and cares for the precocial young until they fledge.

Migration Ecology - Females depart as early as mid-June, followed by males and then juveniles in July (Colwell and Jehl 1994). Adults and juveniles congregate mostly on large interior alkali lakes, including Lake Abert, Oregon, Great Salt Lake, Utah, and Mono Lake, California to molt into basic plumage and accumulate fat for southbound migration (Reed et al. 1994). Smaller numbers stop on the West Coast, with San Francisco Bay’s salt pond system being the major coastal staging area (Colwell and Jehl 1994). Individuals are rarely seen away from these staging areas. By mid-September, the birds have headed south to feeding grounds in South America, apparently flying non-stop from the central Pacific Coast and Great Basin staging areas to wintering grounds in South America.

Wintering Ecology - Wilson’s phalaropes winter on saline lakes and freshwater marshes in southern South America, mainly in the Andean highlands of Bolivia, Peru, Chile, and Argentina (Jehl 1988).

Distribution and Abundance

North America - During breeding season, this species is found in the shallow ephemeral wetlands of interior western North America. Fall populations are estimated at 1.5 million birds on major staging areas (Jehl 1988).

Pacific Coast - The only major coastal site is San Francisco’s salt pond system in the summer months.

San Francisco Bay - Wilson’s phalaropes are seen almost entirely on the South Bay’s salt ponds and their islands, primarily from June to August (Jurek 1973; Swarth et al. 1982, Harvey et al. 1988). Peak numbers occur in July.

Standard spring and fall shorebird surveys have fallen either too early or too late to capture peak numbers of Wilson’s phalaropes. Point Reyes Bird Observatory’s (PRBO) baywide counts detected maximums of 1,642 in the fall, zero in the winter, and 213 in the spring.

Suisun - PRBO fall count: 225; not seen on winter or spring counts.

North Bay - Not seen on PRBO’s spring 1988 count, but see Figure 6.19.

Central Bay - Not seen on PRBO’s spring 1988 count, but see Figure 6.19.
South Bay - PRBO spring count: 213. Reportedly can reach a fall peak of 40,000 birds (Jehl 1988; see Figures 6.19 and 6.20).

Historical Information

No information.

Population Trends

It has been suggested that the global population of Wilson’s phalaropes has declined due to loss of prairie wetland habitat in their breeding range (Dahl and Johnson 1991; Page and Gill 1994).

Habitat Use and Behavior

During spring high tide censuses in south San Francisco Bay, Wilson’s phalaropes were most commonly found on the salt ponds and on islands in the salt ponds; they have also been observed in freshwater treatment ponds (Hanson and Kopec 1994). Harvey, et al. (1988) documented a preference for high salinity (75-200 ppt) ponds, probably due to the invertebrates that are present in more abundance at these higher salinities.

Foraging - Wilson’s phalaropes forage most commonly on open water habitats, either by “spinning,” or by standing in shallow water and lunging, or by scything like an avocet. Unlike other phalarope species present

Figure 6.19 Maximum Counts of Phalarope Species

on the Bay, they also probe mudflats; foraging strategy is probably dictated by prey availability. Prey items found in five birds collected on South San Francisco Bay salt ponds included water-boatmen, pupae of brine flies, brine shrimp, seed of round stem bulrush, and larvae of brine flies and other insects (Anderson 1970).

Roosting - Wilson’s phalaropes roost at night on open water. In the San Francisco Bay, they roost nocturnally on the salt ponds. On inland lakes, they also form diurnal roosting flocks on shore or on boulders (Jehl 1988). They may possibly use islands or levees in the Bay’s salt pond system for this purpose but this has not been documented.

Movement - There is no recorded observation of local movements of Wilson’s phalaropes. Because this species has been observed foraging on mudflats in other locations, they may be moving from the ponds to exposed mud during low tides.

Conservation and Management

Wilson’s phalarope is threatened by loss of its breeding habitat, the seasonal wetlands of the interior. Many of its post-breeding locations (the salt pond system, Mono Lake, the Salton Sea, and Great Salt Lake) are undergoing change, some created by human management and some outside of that control. Threats to wintering habitat in South America are poorly understood.

Recommendations

The South Bay is the area of greatest importance to this species, and salt ponds and treatment ponds, both of which are used for foraging and roosting, are the most important habitat elements.

Wilson’s phalaropes use the San Francisco Bay most heavily for two weeks in late July during their post-breeding migration, the time when they are most dependent on the Bay. Most bird surveys do not capture the true numbers of Wilson’s phalaropes in the Bay because of the short duration of their visit, but the importance of the Bay to the population should not be underestimated. Wetlands could possibly be managed for this species if more was known about its requirements. Maintain medium to high salinity ponds, especially in June-August and particularly in the South Bay, where mild climatic conditions may be favored by shorebirds in general.

Suisun - Wilson’s phalaropes have only been reported in small numbers from the Suisun subregion, but this may be due to the timing and nature of bird surveys in that area. Maintain or increase the current acreage of managed wetlands to support this species.

North Bay - Though pond habitat exists in the North Bay subregion, Wilson’s phalaropes have not been seen. This may be due to the timing of the bird surveys, however. Maintain some of the existing salt ponds to support this species.

Central Bay - Wilson’s phalaropes have not been seen in the Central Bay subregion. Promote inclusion of ponds and managed wetlands in any bayland restoration projects that may be undertaken in this highly developed area.

South Bay - Maintain the current acreages of low, mid, and high salinity salt ponds, and of treatment ponds. Particularly important within this region are the

Figure 6.20 Relative Use of Salt Ponds by American Avocet, Snowy Plover, Black-necked Stilt, and Phalaropes

Hayward area sewage treatment ponds; Turk Island area ponds (particularly 1C-6C); Coyote Hills Ponds 2A-4A; Coyote Creek waterbird pond; Sunnyvale sewage treatment ponds; Mountain View Pond A3N; and Ravenswood Ponds 2 and 4A. Many more ponds in the South Bay subregion have the potential for use by Wilson’s phalarope (Dumbarton; Mowry ponds; Alviso ponds; Ponds A3E and A3W in Mountain View), but birds have not been seen on these ponds yet. This may be due to the timing of the surveys.

Research Needs

Future research should investigate optimum salinities for the invertebrate prey base this species is dependent on.

References


Danson, J.T. and D. Kopec. 1994. Habitat location and utilization by wintering and migrating shorebirds during high tides in south San Francisco Bay. A draft report of the San Francisco Bay Bird Observatory.


the proposed listing or protection of more than one hundred species, many associated with tidal salt marsh habitats. Many projects to rehabilitate or restore wetlands, especially tidal salt marshes, have been proposed to benefit listed species in the Estuary. For example, the San Francisco International Airport recently proposed restoration of salt ponds in the South Bay, used heavily by both waterfowl and shorebirds, to tidal marsh as mitigation for runway expansion. However, results of such wetland restoration efforts are highly variable (Race 1985), and the efforts to complete successful salt marsh restorations for certain species may come at the expense of shorebird and waterfowl populations that use the existing habitats, including salt evaporation ponds. Critical habitats for waterfowl and shorebirds include tidal flats; sparsely vegetated wetland elements (levees, islets, beaches); managed wetlands; large, persistent seasonal ponds with lots of open water; and active and inactive salt evaporation ponds.

Unfortunately, we lack specific information relating abundance of current populations to the amount of their habitats. We are unable to predict how reduction of present wetland habitat used by these species may affect their populations. Thus, we recommend care in implementing large-scale changes and encourage further study of critical habitats and better delineation of the regional populations present in the ecosystem. We also offer several principles to guide management efforts while considering the habitat needs of waterfowl and shorebirds (Goals Project 1999: Appendix C), and emphasize that these species are unlikely to benefit from tidal marsh conversions when the conversion is from another wetland type. Finally, we support an ultimate goal of accepting no net loss of shorebird and waterfowl habitats and populations in the ecosystem while conducting restoration or enhancement projects.

Waterfowl

The San Francisco Bay region is identified as one of the 34 waterfowl habitat areas of major concern in the North American Waterfowl Management Plan (USFWS 1989). More than 30 species of waterfowl are found in the San Francisco Bay ecosystem. These species are commonly divided into dabbling ducks, which feed at the surface or in shallow water to the depth of their body length, diving ducks, which forage underwater, and swans and geese, which feed on plants by grubbing in sediments of wetlands or fields. Mid-winter waterfowl surveys (USFWS, unpubl. data) of the San Francisco Bay and Delta include more than 700,000 waterfowl, and surveys of the open bays and salt ponds (Accurso 1992) include more than 300,000 individuals, a 25% decrease from the earliest surveys in the 1950s. In 1988-1990, dabbling ducks comprised up to 57,000 of the waterfowl in the open bays and ponds of the Estuary, while diving ducks comprised up to 220,000 of the total. For this review, we selected six species as representative taxa of the waterfowl and the habitats they use in the ecosystem.

Dabbling ducks represent 8-30% of total waterfowl numbers. Northern pintail (Anas acuta) use a wide variety of habitats, including managed marsh, seasonal wetlands, open bay, and salt ponds (see Northern Pintail profile). They were historically the most common dabbling ducks in the ecosystem, but recent population declines of this species have been severe (90% decrease in Suisun Marsh). Pintails in the South Bay subregion have little interchange with birds in the Central Valley (M. Miller, pers. comm.) and may comprise a distinct population with a unique breeding area. Mallards (Anas platyrhynchos) have large economic and recreational importance as a hunted species, and are the most abundant dabbling duck in diked baylands, and especially seasonal wetlands, low salinity salt ponds, and managed marshes of the San Pablo and South Bay subregions.

However, the Estuary is most recognized for the large populations of diving ducks, both bay ducks and sea ducks. Canvasbacks (Aythya valisneria) are bay ducks that were identified as a species of special concern because of declining numbers, but the Estuary still supports the largest population of canvasbacks in the Pacific Flyway and represents one of the largest wintering areas in North America (see Canvasback profile). Protecting their open bay habitats was part of the reason that the San Pablo Bay National Wildlife Refuge was established in the 1970s. They traditionally foraged on aquatic plants in mouths of rivers or channels, but now primarily consume nonindigenous mollusks in open bays or salt ponds. The Estuary also is a major wintering area for up to 140,000 greater and lesser scaup (A. marila and A. affinis), a species we associate with canvasbacks that comprises more than 40% of the waterfowl in the open bays and salt ponds. However, scaup have suffered an unexplained continental decline in the past decade. Smaller diving ducks of the Estuary include the ruddy duck (Oxyura jaimaiensis) and bufflehead (Bucephala albeola), which use a variety of managed marsh areas and

Mallards have significant economic and recreational importance as a hunted species. Hunting proponents lead many efforts to preserve and restore waterfowl habitat.
salt ponds in the baylands. The wintering population of ruddy ducks is one of the largest in North America (see Ruddy Duck profile), and as many as 7,000 bufflehead also are found in the Estuary.

Many sea ducks have declining populations which resulted in organization of a Sea Duck Joint Venture under the North American Waterfowl Management Plan. Sea ducks use open-water marine habitats, and surf scoters (Melanitta perspicillata) are one of the least studied of the North American waterfowl (see Surf Scoter profile). San Francisco Bay appears to be the most important inshore habitat for this species in the eastern Pacific, south of the Straits of Georgia and Puget Sound. Surf scoters are the second most numerous species in the ecosystem, with estimates as high as 73,000 birds in 1991 (Trost 1997).

Geese and swans are of economic and recreational importance, as four of the six members of this group are hunted, and overpopulation of geese may cause large urban and agricultural damage. These species are associated primarily with managed wetlands and agricultural lands in the region. Tule geese (Anser albifrons gambelii) were chosen to represent geese and swans, because they are recognized as one of the smallest goose populations in the world (> 7,000), and Suisun Marsh is one of the few wintering areas where it is found. Formerly, geese were present in larger numbers in the San Francisco Bay Estuary, but they are now down to a remnant few, primarily in Suisun Marsh. A population of what was perhaps a few hundred greater white-fronted geese (and possibly the tule subspecies) in the North Bay now number less than 20 individuals (L. Allen, pers. comm.), and thousands of lesser snow geese (Anser caerulescens caerulescens), which were once reported in the South Bay, no longer occur in the region.

Waterfowl Habitat Considerations

Salt Evolution Ponds - In one of the only studies to examine wetland conversion effects on ducks, the body condition of mallards decreased significantly following loss of salt pans and fish ponds in Spain (Rodrigues and Fabiao 1997). Alternate roosting areas were 13 km farther from feeding areas, which may have resulted in higher energy costs for travel. Salt evaporation ponds supported 30-41% of the waterfowl in the San Francisco Bay Estuary, 9-14% in the former North Bay ponds, and 21-27% in the South Bay ponds (Accurso 1992). Many of the birds found in the Estuary during migration (September-October, March-April) were found in these areas. Up to 42,000 diving ducks have been counted in the North Bay ponds, including 30% of the ruddy ducks in the Estuary, 59% of the canvasbacks, and 38% of the bufflehead. As many as 15% of the dabbling ducks were also found in these ponds, including 19% of the northern pintail and 47% of the mallards. Eighty-three percent of waterfowl were found in 54% of the salt pond area with salinities of 20-93 ppt, with most birds preferring 20-33 ppt areas. Pond size explained much of the variation in counts, with less than 2% of the use on small ponds < 150 ha, and most diving duck use on ponds 200 to 550 ha.

South Bay salt ponds supported up to 76,000 or 27% of the Estuary’s total waterfowl. This area provided the largest haven for ruddy ducks (up to 67% of the population), and supported 17% of the canvasbacks, 50% of the bufflehead, and up to 86% (47,000) of dabbling ducks, including the majority (90%) of northern shovelers. Waterfowl were concentrated in lower salinity (20-63 ppt) ponds, with few birds present in ponds above 154 ppt. Most waterfowl used ponds of moderate size, from 50 to 175 ha.

Open Bay Areas - Up to 50% or 140,000 of the diving ducks surveyed in the Estuary during the winter were counted in the North Bay subregion. Densities were as high as 653 birds/100 ha. The populations include up to 35% of the scoter, 26% of the canvasbacks, and 12% of the scaup. Most of the use was in water depths < 4 m, although much of the open bay area was less than 6 m. The Central Bay supported 17% of the waterfowl, or up to 53,000 birds including 20% of the diving ducks. This area was important for scoter (up to 50%), scaup (16%), and bufflehead (13%), but only 1% of the dabbling ducks. The South Bay supported 9-11% or 36,000 of the waterfowl in the Estuary, and was important for scaup (18%) and scoter (16%). The open waters of Suisun Bay supported only 12% of waterfowl in the Estuary, including up to 15% of the diving ducks (17% of scaup, 16% of scoter, and 16% of canvasbacks).

Shorebirds

Shorebirds are aquatic birds with cylindrical bills varying considerably in length and curvature, reflecting different foraging strategies. Among the 31 species encountered regularly on San Francisco Bay, a wide range of sizes is evident from the sparrow-sized least sandpiper (Calidris minutilla) to the duck-sized long-billed curlew (Numenius americanus). They feed primarily on invertebrates obtained on tidal flats, salt ponds, managed wetlands and other habitats. Most tidal flat specialists are found concentrated in the North and South bays (Figure 6.21; G. Page, unpubl. data). Recent survey information indicates that San Francisco Bay supports very high numbers of shorebirds of most species during migration and winter compared with other wetlands along the Pacific Coast of the United States (Page et al. 1999). San Francisco Bay has been recognized as a site of hemispheric importance to shorebirds by the Western Hemisphere Shorebird Reserve Network.

The Waterfowl and Shorebirds Focus Team selected seven “key” shorebird species as a basis for defin-
ing regional wetland habitat goals for shorebirds. The key shorebird species were selected based on their taxonomic grouping, population status, and habitat use (Goals Project 1999). The Pacific Coast population of western snowy plover was selected because it is federally listed as a threatened species, and about 10% of the listed population (over 100 pairs) has been recorded breeding in the Estuary, primarily in South Bay salt evaporation ponds (see Snowy Plover profile). The red knot (Calidris canutus) was selected because the Estuary is one of only three wetlands on the Pacific Coast supporting as many as several hundred wintering individuals, and they are found foraging in tidal flats of the Central and South bays and roosting in salt ponds.

The western sandpiper is the most abundant shorebird in the Estuary (see Western Sandpiper profile), and represents small sandpipers and plovers, while the marbled godwit (Limosa fedoa) was selected to represent large sandpipers and plovers. Tidal flats are the most important foraging habitat of all these species, and they roost at high tides in salt ponds, managed wetlands, seasonal wetlands, and other habitats above the high tide line. The long-billed dowitcher (Limnodromus scolopaceus) and its associates are singled out as potentially deriving more benefit from managed brackish water wetlands and seasonal wetlands than other shorebirds (see Takekawa and Warnock, in press). The black turnstone (Arenaria melanocephala) represents shorebirds that use gravel to rocky intertidal habitat (see Black Turnstone profile), although none of these rocky intertidal species are abundant in the Bay, numbering at most in the low hundreds.

The Wilson’s phalarope (Phalaropus tricolor) was chosen to represent those shorebirds that, in addition to the snowy plover, are most dependent on the salt ponds for foraging habitat. These salt pond specialists are found patchily distributed (Figure 6.22; G Page, unpubl. data), especially in the South Bay salt ponds, depending on water level and salinity. Some members of this group, including the Wilson’s and red-necked phalarope (Phalaropus lobatus), occur only during spring and fall migration, while the others, including black-necked stilt and American avocet (Recurvirostra americana), are resident and nest primarily in South Bay salt ponds.

**Shorebird Habitat Considerations**

Except for anecdotal information suggesting an increase by shorebird species using salt ponds as their primary foraging or breeding habitat, there are no consistent historic data on changes in abundance of shorebirds in the Estuary during the past 150 years when most human-induced habitat alterations have occurred. The most recent mapping of historic and current habitats by the San Francisco Estuary Institute (SFEI) indicates that tidal flats outboard of the salt marshes have increased in the North Bay and South Bay subregions, but that the total amount of tidal flat has decreased in all subregions, primarily due to loss of tidal flats along slough channels in salt marshes. Since the majority of the shorebirds in the Bay use tidal flats as their primary foraging habitat, the amount of foraging areas, and possibly the abundance of these species, may have decreased in the past 150 years, unless artificial salt ponds and managed wetlands have compensated for the tidal flat losses.

For the majority of shorebirds that forage primarily on tidal flats, conversion of salt ponds might be mitigated (by an unpredictable degree) by creating wide, gently-sloped tidal flats along large channels in restored tidal marsh. Tidal salt marsh and slough channels do not, however, provide high tide roosting habitat for most shorebird species, which require barren to sparsely vegetated sites above the high tide line. Thus, in projects where tidal marshes replace existing habitats, suitable roosting areas in reasonable proximity to tidal flats will need to be constructed to replace roosting areas that are lost.

For the salt pond specialists, substantial areas of salt pond habitat should be maintained in both the North Bay and South Bay subregions. If portions of the existing salt pond systems are converted to tidal marsh and managed salt ponds, it will not be feasible to set aside ponds with important shorebird habitat in a piecemeal fashion. Instead, smaller salt pond systems should be retained and activity managed to produce salinities and water depths most favorable to shorebirds and the other aquatic species targeted for protection. Low, wide, barren to sparsely-vegetated internal levees with fine scale topographic relief should be incorporated into the pond design as nesting and roosting substrate. In addition, salt marsh restoration efforts should attempt to recreate plays that occurred in historic salt marshes.

Since the success of marsh restoration efforts are likely to be highly unpredictable and the value of slough mudflats and salt marsh playas for shorebirds is not well understood, incorporation of these habitats into restored marshes should not be counted as replacement habitat for shorebirds. Further research must be undertaken to...
estimate the amount of salt pond habitat that should be intensively managed for shorebirds and other target species. The maintenance of at least the current numbers of shorebirds relying extensively on salt pond habitat will require an adequate acreage of suitable ponds for 25,000 wintering American avocets, 5,000-7,000 wintering black-necked stilts, tens of thousands of migrating Wilson’s and red-necked phalaropes in the fall, and 300 breeding snowy plovers.

General Recommendations for Waterfowl and Shorebirds

It is important to maintain existing populations of waterfowl and shorebirds in the Bay while increasing habitat for other species that are dependent on salt marsh. Increasing the acreage of salt marsh will come at the expense of other habitats, especially salt ponds and managed wetlands that are also important for waterfowl and shorebirds. Maintaining current shorebird and waterfowl populations will thus require increasing the carrying capacity of remaining salt ponds and managed wetlands or recreating their function in new locations.

Suisun - Although these wetlands are managed primarily for waterfowl habitat by private landowners, populations of one of the major target species, northern pintail, have decreased by as much as 90%. Thus, despite the best management efforts, populations of waterfowl in the Suisun Marsh have decreased. Any conversion of managed wetland habitats will result in a loss of waterfowl. Conversion of this area should proceed gradually to provide time to evaluate population changes and the effects of the loss of habitat. Conversion or loss of this habitat type must be offset by enhanced manage-
Waterfowl & Shorebirds

Figure 6.22 Relative Use (High, Medium, Low) of Different South Bay Salt Pond Areas by Salt Pond Specialists as Indicated from the Proportion of Shorebirds Counted in Different Survey Areas


Management of existing areas or mitigation with alternative areas. Shorebirds are present in the tens of thousands. Management should be promoted to improve areas for their populations.

North Bay - The former salt evaporation ponds in this region are a critical area for waterfowl and shorebirds. Ongoing conversion should be linked to enhanced management of existing areas or mitigation. In this subregion, conversion of 50% of the former salt ponds may result in loss of 24% of the 42,000 waterfowl that are counted in these ponds (Figure 6.23; J. Takekawa, unpubl. data). Change in salt pond areas may already be resulting in reduction of waterfowl numbers (USFWS mid-winter surveys, unpubl. data). Thus, there is an immediate need to develop alternative managed marsh areas in this subregion. Although mudflat habitats seem abundant in the North Bay, shorebird roosting habitats may be limiting and should be increased.

Central Bay - This subregion is highly urbanized and is used least by both waterfowl and shorebirds. Any additional roosting habitat that can be protected from disturbance would be beneficial in maintaining or improving existing populations. Restoration of any large, shallow ponds would likely benefit waterfowl and shorebirds. Wetland rehabilitation in urban areas should be encouraged.

South Bay - The majority of the waterfowl and shorebirds in the South Bay use the salt evaporation ponds for roosting or feeding habitat during the winter. Conversion or loss of this habitat type must be offset by enhanced management of existing areas or mitigation with alternative areas, including created salt ponds, managed wetlands, and seasonal wetlands. For example, analysis of waterfowl survey data from 1988-1990 suggests that if 50% of the salt ponds are converted, 15% of the 76,000 waterfowl may be lost (Figure 6.24; J. Takekawa, unpubl. data). An increasing number of waterfowl would be displaced if more area was converted.

Although mudflat foraging habitat seems adequate, with salt pond conversion, suitable roosting habitat for shorebirds may become limiting. Little is known about how salt ponds and seasonal wetlands provide food for shorebirds and protected microclimate areas during adverse weather. Thus, we recommend not more than 50% or 15,000 acres of salt ponds in the South Bay be converted to other habitats without careful planning for habitat mitigation for shorebird and waterfowl populations. We also recommend an increase in seasonal wetlands as migration habitat and roosting areas.

Enhancing Tidal Marsh Restoration Projects for Waterfowl and Shorebirds

Waterfowl and shorebirds may use several elements in tidal salt marshes. As restoration or rehabilitation is undertaken, these elements should be provided when possible.

1. Larger channels with large mudflats are often used by shorebird and waterfowl species and should be encouraged in tidal marsh design.
2. Muted tidal areas provide temporal diversity which may provide good habitat, especially for diving ducks.
3. Unvegetated levees and islets with gradual slopes that are durable, and bare areas that remain unvegetated with limited management should be constructed as roosting sites.
4. A diverse mix of pans and ponds should be retained in marsh plains for high tide roosting and foraging areas.
5. Designs should be made to minimize disturbance by people, pets, and predators.
6. Surveys of waterfowl and shorebirds should be conducted prior to restoring areas to tidal salt marsh so losses may be evaluated and suitably mitigated.

**Research Needs**

Relationships among habitat change and change in populations of waterbirds have been studied in other estuaries (see Goss-Custard et al. 1997). We should learn from these efforts and develop a research program in the San Francisco Bay Estuary to examine questions raised during the Habitat Goals Project, including the following topics:

1. Determine the feasibility of designing ponds or systems from the existing salt evaporation ponds which can support the current populations of waterfowl and shorebirds.
2. Evaluate what constitutes a good roosting area for different species of shorebirds, including distance from feeding areas. Areas used within tidal salt marshes should be included.
3. Estimate the size and composition of shorebird populations in Suisun subregion.
4. Determine the importance of non-mudflat habitats such as salt ponds and seasonal wetlands as foraging areas, especially during inclement weather.
5. Examine seasonal wetland use and extent (not currently shown in the EcoAtlas), including diked farmland, grazed baylands, diked marsh, managed marsh, and ruderal baylands through wet and dry years.
6. Test differences in shorebird and waterfowl response to different actions in managed wetlands by measuring use-days and numbers.
7. Relate diving ducks use of wetlands by area size and water depth.
8. Quantify shorebird foraging and roosting in wetlands other than intertidal flats, including intertidal pans, low and medium salinity ponds, managed marsh, diked marsh, muted tidal, and seasonal ponds. Include factors such as tidal cycle, salinity, vegetation, and distance to intertidal flats.
9. Describe use of wetlands by salinity and prey differences for waterfowl and shorebirds.
10. Provide more information about the effects of disturbance on waterfowl and shorebirds to develop suitable habitat buffer zones.
11. Determine the effects of channelization, levee alteration, and use of dredged-spoil on mobilization of contaminants sequestered in soils or sediments and bioaccumulation in waterfowl and shorebirds.
12. Characterize hydrology, biology, and chemistry of salt ponds heavily- and lightly-used by waterfowl and shorebirds to examine the differences.
13. Determine habitat values and use by waterfowl and shorebirds of managed wetlands versus tidal wetlands.
14. Investigate the effect non-native invertebrates and plants (e.g. Potamocorbula amurensis, Spartina alterniflora) on waterfowl and shorebirds.
15. Evaluate methods to reduce effects of non-native predators on waterfowl and shorebirds.
16. Examine the effects of contaminants on breeding birds.
17. Pilot Projects - encourage monitored experiments in wetland restoration or mitigation. Pilot Projects should:
a. Include repeatable waterbird surveys before and after project actions.
b. Examine maintenance or creation of salt pond systems, including low to mid-salinity ponds in the absence of commercial production. Habitat values and use should be maximized while minimizing maintenance costs.
c. Test methods of constructing habitat elements with low maintenance requirements such as bare roosting islands, intertidal pans, and non-tidal seasonal ponds.
d. Examine differences in use of different wetland unit sizes.
e. Test methods of increasing shorebird and waterfowl use of managed marshes.
f. Increase monitoring efforts on existing projects with habitat elements valuable for waterfowl and shorebirds.
g. Employ adaptive management by applying earlier findings to change design elements through time.
h. Conduct preliminary sampling for contaminants of areas designated for salt marsh restoration.
i. Conduct preliminary sampling of salt ponds for invertebrate community, salinity, and other water quality characteristics.

References


Personal Communications

Janet Kjelmyr, Staff Biologist, Point Reyes Bird Observatory, Stinson Beach, California.

Michael Miller, U.S. Geological Survey, Biological Resources Division, Dixon Field Station, California.

L. Allen, Canvasback Duck Club, Napa, California.
Other Birds of the Baylands Ecosystem

Chapter 7

Eared Grebe
Podiceps nigricollis
Howard L. Cogswell

Introduction
The eared grebe is a small, stonky-bodied bird that may be found in a variety of habitats within the San Francisco Estuary, and particularly in the salt ponds of the South Bay. The eared grebe acquires its name from the golden tufts of feathers that fan out behind the eyes of the adult in breeding plumage.

Description

Breeding Biology - Eared grebes nest primarily on medium-sized to large lakes with marshy borders. They build a floating nest attached to underwater or emergent vegetation, as is typical of grebes. Where successful, they typically nest in colonies from a few pairs to many hundreds. In California, most breeding occurs at lakes east of the Sierra Nevada-Cascade mountain ranges. However, at least in wet years, nesting colonies have been found in the Sacramento and San Joaquin valleys, inland valleys of coast-slope southern California, and at such mountain lakes as Lake Tahoe and the Big Bear-Baldwin lakes in San Bernardino County (Grinnell and Miller 1944; miscell. records).

In the Bay Area, a colony of 70 or more eared grebes nested in 1983 in northwest Pleasanton on out-of-use sewage ponds with marsh at one corner. There were at least 65 young produced at this site that were still flightless when the pond was drained in July, and the adults all left. Most of the young that survived in the remaining border ditch were captured, banded, and released at Coyote Hills or Lake Chabot (personal field notes and Amer. Birds 37:1022). An apparently larger colony (101 nests in use or being built on July 15) the same year was successful in Crittenden Marsh near Moffett Field, 39 juveniles being seen there on August 19 (Bousman, pers. comm.). At least 12 adults and 15 "immatures" were noted in the same marsh in August 1986, and 10 nests (seven with eggs) were also there in June 1992, but these disappeared by July 25. On May 11-24, 1993, Peter Metropulos found 12+ nests, many with eggs, in a pond east of Crittenden Marsh. In 1994, as many as 10 adults were seen in the same area, at least six of them obviously paired, but no nesting evidence was obtained. Irregular, but at least sometimes successful nesting is thus demonstrated in a bay-related habitat in our area of interest. Bousman (pers. comm.) also reported that Gloria Heller discovered at the Western Foundation of Vertebrate Zoology (Camarillo, California) a "nest card for eared grebe on an unnamed lake on Stanford University on May 14, 1908 with seven eggs (not collected)." This was apparently the earliest record of breeding anywhere near the Bay. Neither Grinnell and Wythe (1927) nor Grinnell and Miller (1944) mentioned this early record, although they indicated summer occurrence on Lake Merced, San Francisco, as indicative of possible nesting there.

1 This was an out-of-use salt pond, and so may have contained brackish rainwater.
2 These records were summarized from the Santa Clara County breeding season records of eared grebes, supplied by William Bousman.
Migrant Ecology - The Great Salt Lake, Utah, and Mono Lake in central-eastern California are well-known major concentration points for post-breeding migrant eared grebes. Adults engage in a "molt-migration" to reach such areas and begin to put on weight from the abundant food (brine shrimp, etc.) before beginning their annual molt, which involves loss of all flight feathers at the same time. The birds are thus flightless for more than a month. Many aspects of the Mono Lake populations - the weight changes, progress of the molt, mortality, arrival, departure, and distribution on the surface of the lake - are detailed by Jehl (1988). Total eared grebe populations on Mono Lake, per Jehl's extensive sampling and careful analysis, reached peaks of 500,000 to 800,000 in September through October. He also showed a summary map of sizes of populations at this and other (all lesser) migration stopover points for August through October 1985. However, Jehl's work did not include San Francisco Bay salt ponds, despite the fact that they undoubtedly held far more than the median number of grebes (6,000) among his sites from Saskatchewan to southeastern California.

Wintering Ecology - According to Jehl (1988) the only truly major concentrations during mid-winter are on the Salton Sea in southeastern California, and the Gulf of California, Mexico; but small numbers persist through some winters even at Mono Lake and other inland waters not subject to freezing. At Salton Sea, estimates of the total population in February and March have been as high as 205,000 to 700,000 (1977 and 1953 aerial counts; AOU 1998 and Audubon Field Notes). Christmas Bird Counts in 1981-82 through 1985-86 for only the south end of the Sea ranged from 3,510 to 24,140 (Jehl 1988).

Distribution and Abundance

North America - The regular breeding range of the eared grebe extends from southwestern Canada to western Minnesota and eastern Illinois south to northwestern Mexico and south-central Texas, with local populations south to central Mexico. In winter, numbers are found regularly along the Pacific Coast from southern British Columbia to Central America, and inland from central California, Utah, and central Texas south. Non-breeding birds of this species occur casually in the eastern United States. In the Old World, the eared or "black-necked" grebe is found locally across central and southern Eurasia and parts of Africa (AOU 1998, map in Palmer 1962). Several closely related species (classified as subspecies of the eared grebe by some authors) are resident in the Andes, high plateaus, or southern part of South America (Sibley and Monroe 1990).

San Francisco Bay Area - Eared grebes occur rather widely as migrants and more narrowly through the winter on waters of many sorts in the Bay Area. They have been seen on lakes, ponds, marsh sloughs, the open bay, and (especially during spring migration) even the ocean as far offshore as the Farallon Islands.

The larger counts or estimates of numbers of eared grebes tallied from published reports or field notes pertaining to San Francisco Bay or vicinity by Cogswell (1977) range from 10 to 170 on freshwater lakes (September through January), five to 50 on brackish lakes (Lake Merritt in Oakland and Berkeley Aquatic Park, October through January), 13 to 90 on eastern mid-San Francisco Bay or harbors connected to it (October through January), and 25 to 600 on the ocean near South Farallon Island (October through May, but mostly Christmas Bird Counts). By comparison, the peak numbers reached on medium-salinity salt evaporators from October through April regularly range in the hundreds or thousands per one to several large ponds. Counts in the southwestern Hayward evaporators were from 113 to 751 in September through December 1965-69, 490 on February 14, 1965, and 7,500 on April 17, 1965 - but only 83 on May 4, 1965 (H. Cogswell, field notes). The 7.5-mile radius Hayward-Fremont Christmas Bird Count circle (centered at Hyd. 880 and Whipple Rd.) includes all of the same salt ponds and others to the south as far as Dumbarton Point. Over 1,000 eared grebes are tallied in many years of that count, nearly all of which are on salt evaporators. The same is probably true for the occasional thousands (max. 13,615 in 1973) on the San Jose Christmas Bird Count, although a smaller area of salt ponds is in that circle.

Habitat Use and Behavior

Eared grebes may occasionally be seen in a variety of habitats within the San Francisco Estuary, but they most frequently are found in medium to high-salinity salt evaporator ponds, where they rest and forage. The deciding preferred habitat from late August through April or early May is the medium or medium-high salinity ponds, where counts may range up to several thousand birds per pond. These ponds show high concentrations of brine shrimp (Artemia salina) and/or water boatmen (Hemiptera: Corixidae), which are prime prey for these small grebes. When on fresh water, they also take many kinds of aquatic insects and crustaceans but apparently few fish (Palmer 1962).

Two special studies of the use of salt-pond habitats by birds deserve mention. Anderson (1970) studied the series of ponds lying north of Mowry Slough, which then ranged from low-salinity (intake pond M1) through medium-high salinity. He found a maximum of 6,330 eared grebes in November on his "ponds of high salinity." In a

3 These ponds were actually of lower salinity than two or three other ponds that lay between his study area and the final crystallizers.
more thorough, two-year study of the 11 ponds lying west of Coyote Hills and north of the Dumbarton Bridge approach, Swarth, et al. (1982) reported a maximum count of 5,565 eared grebes in April 1980, but numbers the following spring were below 3,200. They had average counts of 500 and about 1,950 in November of the two successive years, and a very few were found through the summer. The November through April eared grebe numbers per 10 hectares in each of the 11 ponds in the same study were strongly correlated (at p<0.02 level) with the grams dry weight of invertebrate biomass in the same ponds. That biomass was calculated from samples obtained by hundreds of plankton hauls thru the upper ¼ meter of water within three meters of a canoe. Brine shrimp (Artemia salina) and water-boatsmen (Hemiptera: Corixidae) constituted nearly all of these samples. The grebes may, however, also have been eating brine-fly (Diptera: Ephydra sp.) larvae and pupae which spend most of the time below the ¼-meter depth, or even adult brine flies on the water surface which are quite able to escape the hauled net.

In the baylands of the San Francisco Estuary, eared grebes will also use subtidal and tidal habitats (including large marsh channels) for foraging and resting, although this is more common on offshore islands during migration than on the Bay proper. A relatively few birds use freshwater marsh for breeding and foraging. Seasonal wetlands are used for foraging and less commonly for breeding, particularly when water ponds for long periods and there is at least some emergent or near-surface vegetation. For the purposes of the Goals Project (and the Project’s Habitat Matrix), it should be considered that eared grebes do not significantly use those areas classified by the Project as tidal flats, tidal marsh, riparian woodland, adjacent uplands, unvegetated supratidal shores, rocky islands or cliffs, and towers or other human-built structures.

Conservation and Management

On the South Bay, each set of medium- to medium-high salinity ponds presumably hosts numbers of eared grebes similar to those cited above for the Mowry Slough, Coyote Hills, and southwest Hayward areas. Peaks appear to be in October or November on some ponds (at which time Mono Lake still harbors about a half-million or more grebes), while on other ponds the peak does not occur until spring migration in April. For example, during the April migration period in 1980, over 5,000 birds were reported west of Coyote Hills (Swarth et al. 1982), and on April 14, 1996, 2,000 birds were counted on one salt pond (H 9) in Hayward (J. & F. Delfino, pers. comm.). There are at least seven sets of such medium-salinity ponds in the South Bay system. If it was estimated that each set of ponds harbors just 3,500 birds (the mean of the two figures cited) during the spring migration and in the fall, then it could be hypothesized that the South Bay salt ponds may support as many as 24,500 eared grebes. This would constitute an additional migration stop for this species that is well above the median for the entire range shown by Jehl. While midwinter numbers here are somewhat lower, there are still thousands of eared grebes to be found on the salt ponds, and the area as a whole may well serve as wintering or migration habitat for 50,000 to 100,000 birds—a significant portion of the total species population, even though far below the major magnets of Mono Lake and Salton Sea.

This species is the most partial to use of the saltpond habitat of any of the birds found there. It should be recognized in all plans for future habitat management that this species would thus suffer a marked impairment of its total available high-quality habitat if the saline ponds were eliminated or sharply reduced in extent. Nearly the same dependence on saline lakes or salt ponds for migration stopovers is also likely true of the Wilson’s and red-necked phalaropes, even though their wintering area is far to the south of that of the eared grebe.

References


Grinnell, J. and A.H. Miller. 1944. The distribution of the birds of California. Pacific Coast Avifauna no.27. 608pp. (Cooper Ornithological Club, Berkeley, Ca.)


Western and Clark’s Grebes
Aechmophorus occidentalis and A. darkii
David G. Ainley

Introduction
The western and Clark’s grebes are very closely related; until recently, the Clark’s grebe was thought to be a light morph of the western grebe. Still more recent work indicates that the two soon may be re-merged taxonomically. In the San Francisco Bay region, the western grebe outnumbers the Clark’s grebe by at least 9 to 1 (Shuford et al. 1989). The biology of the two species is virtually identical. Unless indicated otherwise, the information elsewhere in this account applies to both species and is from Storer and Nuechterlein (1992).

The western/Clark’s grebe is the largest grebe in North America and one of the largest in the world; they are the size of a medium-sized duck (weighing about 1,430 g). These species are basically piscivores and the stiletto-shaped beak is well-suited for spearing fish. The bill also is used like a forceps to grasp fish (and occasionally crustaceans).

Distribution and Abundance
Western/Clark’s grebe is found throughout the western portion of North America, except in the deserts and tundra. These grebes frequent lakes, large rivers, tidal sloughs, bays, and coastal marine waters (greater than 15 m deep). They breed on inland bodies of fresh and saline waters or protected tidal waters, from the Pacific Coast east to Colorado and from Saskatchewan south to Mexico. The breeding season extends from February to September (Cogswell 1977), and they vacate inland breeding areas, flying to the west coast, before freeze-up. Wintering birds occur in Pacific Coast waters from British Columbia south to Mexico.

These species do not breed in habitats that are directly part of San Francisco Bay. Around the Bay Area, these grebes breed among the tall emergent vegetation on a number of isolated reservoirs (e.g., Calaveras Reservoir, Santa Clara County; Bousman, pers. comm.; Lake Merced, San Francisco County: Grinnell and Miller 1944). The largest breeding population close to San Francisco Bay is at Clear Lake, Lake County, and at Lake San Antonio, Monterey County.

No area-wide counts for western/Clark’s grebes are available either historically or in recent years. The longest records of local censuses come from various Christmas Bird Counts. Counts that have provided data from the late 1960s at localities reporting few grebes (circa five or fewer grebes reported per party-hour) — Crystal Springs Reservoir, Benicia, Oakland, Hayward-Fremont, and Palo Alto — have shown no trends in numbers. The southern Marin County count, however, began at levels of 30-70 grebes/party-hour in the early 1970s, but declined gradually to vary around 10 grebes/party-hour by the late 1980s or early 1990s.

Most of the grebes of these species seen on San Francisco Bay are non-breeding individuals. In the Bay Area, peak numbers occur October through April (Ainley and DeSante 1980, Briggs et al. 1987, Shuford et al. 1989), the non-breeding period. The majority of such wintering birds come from breeding sites well inland (Great Basin, etc.). A few individuals, perhaps including local breeders, can be seen on San Francisco Bay in any month of the year, however.

Habitat Use and Behavior
Within the San Francisco Estuary, western/Clark’s grebes can be found in the waters of sheltered coves, and sparsely in sloughs. Rarely are they found in the open Bay, except along tidal rips in the vicinity of Racoon Straits and Angel Island.

Western/Clark’s grebes are entirely aquatic and never come to land, unless ill. Their nests float, but are attached to emergent reeds. When foraging, these grebes dive by jumping up and forward. They use their feet for propulsion. Many of the fish consumed are near-bottom dwellers. Herring (Clupea harengus) are an important component of the diet of these grebes in bays of the Pacific Coast, such as Puget Sound (Palmer 1962). This fish is likely an important part of the grebe diet in San Francisco Bay.
Conservation and Management

The presence of these species in San Francisco Bay is contingent upon the availability of forage fish, such as herring. The decline in grebe numbers in southern Marin may be due to changes in the herring population size or distribution. Herring, which occur most densely in the central and outer part of San Francisco Bay (waters of Marin and San Francisco counties), declined in spawning biomass from the 1960s through the mid-1980s, and during the 1990s, they have shifted somewhat from spawning in waters off Marin to waters off San Francisco (CDFG 1995).

The western/Clark's grebes and other grebe species typically seek sheltered waters, where in San Francisco Bay they are constantly displaced by the boats of human fishers, which also seek these localities. The prohibition of boats in the inner part of Richardson's Bay provides sanctuary. In fact, very large numbers of these grebes occur in Richardson's Bay, which is also in proximity to herring spawning areas.

References

Grinnell, J. and A.H. Miller. 1944. The distribution of the birds of California. Pacific Coast Avifauna No. 27.

Personal Communications


American White Pelican

Pelecanus erythrorhynchos

David G. Ainley

Introduction

The American white pelican is one of the larger birds of North America, and certainly the largest piscivore (22-35 kg). The species is very gregarious, being both a colonial breeder and a group-forager.

Distribution and Abundance

The species nests exclusively on islands within large saline lakes in western North America, from just south of the tundra in central Canada to Texas, and from the Pacific Coast to the Mississippi River Valley. During winter, breeding populations move to traditionally established sites in California and Mexico as well as areas along the Gulf of Mexico (Palmer 1962).

Occurrence in the San Francisco Bay Area is very localized and is confined to the non-breeding season, generally from June through December (Shuford et al. 1989, Bousman 1993). The American white pelican frequents very shallow water and is seen (rarely) in the open parts of the Bay only in transit. They are almost exclusively gregarious and roost in flocks on dikes. One wintering population can be found at White's Slough, Contra Costa County (pers. obs.), another in the Hayward area, and another frequents salt evaporation ponds of the South Bay (Bousman 1993).

A few thousand likely spend their non-breeding season in the San Francisco Bay Area. No trend in numbers has been apparent during recent decades (Bousman 1993).
Habitat Use and Behavior

American white pelicans feed on small, rough fish; in San Francisco Bay this would include sticklebacks (Gasterosteus lineatus) (Palmer 1962). They capture prey by swimming in large groups, coralling them, and then scooping them up with their large beaks.

Conservation and Management

The presence of this species in San Francisco Bay results from its well-being at inland breeding sites and the presence of quiet waters, such as salt evaporation ponds.

References


Brown Pelican

Pelecanus occidentalis
David G. Ainley

Introduction

The brown pelican is one of the largest piscivorous birds of coastal and estuarine waters of North America (weighing about 17 kg). The species breeds colonially, constructing its stick nests on the ground or, more commonly, in trees or shrubs. Pelicans lay two eggs per nesting attempt.

Distribution and Abundance

In western North America, the brown pelican breeds on islands in marine waters on either side of Baja California, Mexico, north to the Channel Islands of southern California and to Florida. In the West, following the breeding season, many thousands move north to "winter" from central California north to the Columbia River. Peak numbers in central California, including the San Francisco Bay and surrounding area, occur from July through November (Shuford et al. 1989, Bousman 1993, Jacques 1994). During years when pelicans do not breed, such as during El Niño years, large numbers (in the thousands) occur throughout the year in northern California, including San Francisco Bay (Anderson and Anderson 1976). The highest counts in central and northern California occur during those warm-water periods (Ainley and DeSante 1980, Jacques 1994). Choice of wintering areas has to do with the availability of food and to tradition (Jacques 1994).

There are no current or historical Bay-wide censuses of brown pelican. The number of birds found over the waters of San Francisco Bay in a given year varies according to the well being of this species at its breeding grounds and the numbers in coastal waters of central California. In years of high breeding productivity or years of non-breeding, more pelicans can be found here. The fall peak in brown pelican numbers in central California has ranged from about 7,000 (in 1987) to 21,000 (in 1981; Jacques 1994). Currently, on average, several hundred occur within the Bay each summer and fall. As the species recovers from effects of DDT on its breeding productivity in the 1950s and 1960s (Anderson and Gress 1983), numbers seen in the Bay Area have slowly increased (e.g., Ainley 1972, Baldridge 1973).

Habitat Use and Behavior

In San Francisco Bay, brown pelicans frequent all the deeper waters, including some salt evaporation ponds and the mouths of the larger creeks (e.g., Corte Madera Creek, Marin County). Significant numbers are not found much farther inland than San Pablo Bay. They roost in numbers on small islands (e.g., Red Rocks) and breakwaters (e.g., Alameda Naval Air Station).

Brown pelicans feed on schooling fish. In waters of the San Francisco Bay, their diet includes such species as anchovies (Engraulis mordax) and smelt (e.g., Hypomesus spp.; Pers. obs. and Palmer 1962). Their technique of feeding—plunging beak first from altitude into the water to grasp fish up to a meter or so deep—requires deep water.

Conservation and Management

Except on nesting grounds, brown pelicans are not intimidated by the presence of humans. The species occurs in close proximity to humans and forages very close to human fishers. As long as forage fish are available, the population of brown pelicans will do well. When forage...
fish are not available, brown pelicans scavenge fish offal discarded by humans.

Because this species is a higher order consumer, populations suffered considerably due to the effects of DDT on breeding productivity in the 1950s and 1960s (Anderson and Gress 1983).

Currently, the California population of this species is listed as endangered on the Federal Endangered Species List, but may (or should) be down-listed or delisted soon (e.g., CEQ 1986, Ainley and Hunt 1990).

References


Double-Crested Cormorant
Phalacrocorax auritus
David G. Ainley

Introduction

Cormorants are found the world over from the Arctic to the Antarctic. The family is large, and its members are mostly confined, with exception, to coastal marine waters. Cormorants are foot-propelled divers and feed mostly on fish, although they take mid-water swimming crustaceans (such as shrimp) as well (Ainley 1984).

Cormorants construct their nests in colonies. Most nest on the ground, although some colonies occur in trees or on man-made structures. Among marine birds, cormorants are the most prolific, with their clutches averaging three to four eggs per nest. The capacity to lay so many eggs (most marine birds lay one egg only) allows their populations to respond positively to periodic conditions of food abundance. On the other hand, having so many chicks to feed often leads to food stress among parents when food is sparse and, consequently, to high mortality of chicks (Ainley 1984).

Distribution and Abundance

In North America, the double-crested cormorant is the only cormorant species associated with inland bodies of fresh, brackish, and saline water. They also occur close to ocean shores in protected waters. All other North American cormorant species are strictly marine, and in San Francisco Bay, except for vagrants, these other species occur only in the vicinity of the Golden Gate and Angel Island.
In the early part of the 20th century, almost all double-crested cormorants that occurred in San Francisco Bay likely nested on the offshore Farallon Islands, but commuted to the Bay for foraging. Since the late 1970s, they began to nest in small numbers around the Bay, especially on power transmission towers, bridges and, rarely, trees.

This species now is widespread in San Francisco Bay and the Delta. Since the species is a colonial breeder, breeding birds are concentrated in only a few locations – one major concentration is in the North Bay salt evaporators near Napa, two are in the Central Bay at the Richmond and Oakland-Bay bridges, and another is in the South Bay at the Dumbarton Bridge. The birds radiate outward from these colonies to forage at distances of 20 or more miles away. Double-crested cormorants often forage in flocks (see Barlow 1942, 1943), but also do so singularly.

In the 1800s through the 1940s, many thousands of these birds occurred in San Francisco Bay and were associated with schools of sardines (Sardinops caerulea), upon which they fed (Barlow 1942, 1943; Ainley and Lewis 1974). Owing to persecution by humans and perhaps the decline of sardine populations, the numbers of double-crested cormorants in San Francisco Bay declined rapidly, reaching a low during the 1960s and early 1970s (Ainley and Lewis 1974, Carter et al. 1995). Since then populations have been recovering. As of 1991, about 2,800 birds nested around San Francisco Bay in 12 colonies: Russ, Knight, Wheeler, and Donlon islands; San Pablo Bay radar station and beacon; the Richmond, Bay, and San Mateo bridges (and associated electric towers); and electric towers along the very southern shore of the Bay (Carter et al. 1995; SF Bay Bird Observatory, unpbl. data). The largest colonies in the Bay are on the Oakland-San Francisco Bay Bridge and Richmond-San Rafael Bridge (846 and 1,116 birds, respectively, in the mid-1990s; Carter et al. 1995). On the basis of sightings of banded birds, as well as a decline at the Farallones and simultaneous increase on the Richmond Bridge, the Farallon colony has supplied many recruits to these populations (Stenzel et al. 1995). A few pairs also nest at several localities in counties bordering San Francisco Bay; for example, several pairs nest on transmission towers near the mouth of Stevens' Creek, Santa Clara County (Bousman 1993).

This species is most prevalent in waters of the San Francisco Bay and Delta during winter – November through March. The increasing summer breeding population is the result of the arrivals of yearlings from the previous breeding season and birds from the colder, interior parts of North America. Although no Bay-wide census has been conducted during winter, their numbers likely reach 10,000 or more.

Habitat Use and Behavior

Breeding cormorants typically are very sensitive to disturbance from the intrusion of humans. Double-crested cormorants are among the least sensitive among all cormorants, which is not to say, however, that the species is oblivious to the presence of humans. They will flee their nests, leaving the contents to scavenging gulls or corvids when people approach within a couple hundred feet.

The double-crested cormorant forages in shallow waters overlying bottoms of flat relief. Such foraging areas may include rivers and sloughs tributary to San Francisco Bay, as well as salt evaporation ponds and areas such as San Pablo Bay. Large numbers are found in the tidal rips associated with Angel Island and Raccoon Straits. Double-crested cormorants feed mainly on fish. Herring (Clupea harengus) is an important prey in many coastal areas, and likely also in San Francisco Bay during winter. Midshipmen (Porichthys notatus) are an important food item during spring and summer (Palmer 1962, Ainley et al. 1981, Ainley, pers. obs. for San Francisco Bay).

Conservation and Management

For many years, the species was eradicated throughout North America because it was accused of foraging on favorite sport fishes. Protection from persecution and disturbance, and the increased availability of man-made structures on which to nest, has contributed greatly to the increase in numbers during recent decades. Even today there is pressure to control numbers, especially in cases where they forage on stupid, hatchery raised trout introduced to urban reservoirs (e.g., Lagunitas Reservoir, Marin County). Another factor that may have “allowed” the recent increases has been control of pesticides. This species is particularly sensitive to these compounds (Gress et al. 1973, Fry 1994).

References


Snowy Egret

Egretta thula
William G. Bousman

Introduction

The snowy egret is a member of the family of herons and egrets (Ardeidae) that occur in wetlands throughout the world’s avifaunal regions. Within the New World, the snowy egret is widespread in its distribution and is a counterpart of the little egret (Egretta garzetta) of the Old World. Two subspecies are recognized (Palmer 1962), E. t. thula, that breeds in the eastern United States through Mexico and into South America as far south as Chile and Argentina, and E. t. brewsteri, that breeds in the western United States including California south to Baja California and western Mexico. Within its breeding range it is generally common, although strongly dependent upon wetlands for foraging both during and outside of the breeding season. In the San Francisco Bay, it is a resident species.

Snowy egrets are generalists in their feeding habits, foraging on small fishes, frogs, lizards, snakes, crustaceans, worms, snails, and insects. As with most generalists, they are opportunistic in their feeding and benefit from drying periods in seasonal wetlands and fish blooms that occur in salt ponds or other impoundments. Males establish breeding territories, and then, after pair formation, the pair normally defends a smaller nesting territory (Palmer 1962). Foraging territories are also defended. Breeding is normally colonial with one brood per year. Snowy egrets normally lay three to five eggs, but the young hatch asynchronously, and the smallest young survive only when food is plentiful. Nests are constructed on the ground, in trees, or marsh vegetation. On West Marin Island, they nest on the ground, in coastal scrub, in buckeye, and in live oaks. Birds that occasionally nest at Audubon Canyon Ranch use redwood trees and nest 60 to 70 feet above the ground (Shuford 1993). In Alviso, they nest in tules along Artesian Slough just barely above the surface of the water.

Distribution and Abundance

Modern Distribution – McCaskie et al. (1979) described the snowy egret as common to abundant in the seashore, coastal, interior, and Great Basin districts in northern California, although they noted that it is much less common inland, as well as on the coast north of Sonoma County. They considered beaches, mudflats, and marshes to be the primary habitat for this egret.

In Southern California, Garrett and Dunn (1981) considered snowy egrets to be common residents at the Salton Sea and along the Colorado River Valley, but only common as a winter visitor along the coastal slope where some birds are found uncommonly in the summer. It has occurred as an uncommon transient anywhere in the region. This species has nested along the Salton Sea, but has declined because of competition with cattle egrets. Along the coastal slope, recent nesting records included Sandyland Slough in Santa Barbara County and Buena Vista Lagoon and the Tijuana River Valley in San Diego County.

Today this species is a common, year-around resident in the San Francisco Bay. Christmas Bird Count (CBC) data from the late 1960s to the present, shown in Figure 7.1, indicate that this species is found regularly on the CBCs in Benicia, Oakland, Hayward-Fremont, San Jose, Palo Alto, Crystal Springs Reservoir,
Marin County (southern), and Arroyo Cheap Thrills. For all of these counts, the numbers are comparable and show no substantial changes in the last 25 years. The aggregate number of birds counted in the nine CBC circles shown in Figure 7.1 can be approximated by summing the mean number recorded on each count. This aggregate mean, 1,112 birds, represents a lower bound of the birds present in San Francisco Bay, as not all estuarine habitats are sampled within these circles, and it is likely that some birds were missed during these counts. Comparisons of Palo Alto CBC and Summer Bird Count (SBC) data obtained in the same count circle allow a comparison of summer and winter numbers using identical census protocols. These data show essentially the same population in both the summer and winter seasons, and it appears that this species is permanently resident in this portion of the Bay.

Historical Distribution – Within California, Grinnell and Miller (1944) considered the snowy egret to be a year-round resident below the 1,000-foot elevation level in the southern three-fourths of the State, which includes the San Francisco Bay Area. Elsewhere it was found only in the summer or as a vagrant. They described the Sacramento, San Joaquin, and Colorado River valleys as the chief location for this egret, but noted that it was found coastally from Marin County south to San Diego County.

Prior to 1880, this species was considered locally common in the State (Grinnell and Miller 1944). Plume hunting, however, was as devastating to this species on the Pacific Coast as it was in the East, and starting in the 1880s, this species was nearly wiped out in the State. By the early 1900s, it was thought to be extinct within California. By the 1920s, it was considered a rare straggler to the Bay Area with only two locations noted (Grinnell and Wythe 1927). By the early 1940s, however, this species had started to recover and in favored places was locally common. Even by 1940, however, the only known breeding site was in Los Banos (Grinnell and Miller 1944).

By the early 1950s, in the South Bay, this species was considered an uncommon winter vistor (Sibley 1952). Emily Smith (Audubon Field Notes 9:51) considered ten birds at Alviso on 30 August 1954 to be notable, and this was the largest number reported for the Middle Pacific Coast Region in that season. A survey of South
Bay breeding birds in 1971 (Gill 1977) recorded 340 pairs on Bair Island, near Redwood City and this colony apparently was first started in 1969. This colony was still active at least through 1975 (Gill 1977), but it is no longer extant, and the reasons for its abandonment have not been described.

The species now appears to have recovered to its carrying capacity in the Bay Area, as noted above by the CBC population trends. It appears that most of this recovery occurred in the period from the mid-1950s to the late 1960s. No census data are available, however, that can accurately characterize the numbers present prior to European settlement.

Habitat Use and Behavior

From Palmer (1962), this species uses fresh, brackish, and salt-water habitats throughout its range. Within the San Francisco Estuary, it uses all of these habitats for foraging, although for breeding, it is rarely far from brackish or salt water. The densest concentrations of snowy egrets are found either where drying ponds concentrate suitable fish species or where fish blooms occur, and by inference, seasonal wetlands and impoundments are an important source of prey. Nonetheless, this species feeds widely along the tidewater margin, in nearby freshwater streams, and in lakes and reservoirs. In all cases, it depends upon healthy fish habitats for its prey base.

It uses a wide variety of substrates for nesting, and it seems clear that the actual substrate is of little importance compared to the security that the nesting locality provides from predation. Nearly 500 pairs have been noted at West Marin Island (Shuford 1993) and this is the largest concentration in the Bay Area. At Alviso, nests are built only slightly above the water in dense tules and 150 pairs were noted here in 1980 (Am. Birds 34: 811). Away from the Bay Area, this egret was first found breeding in Sonoma County in 1991 in the midst of an active black-crowned night heron colony in Penngrove. Seven active nests were found that year (Burrell 1995), and this shows the flexibility this species exhibits as long as a satisfactory prey base exists and there are secure nesting sites.

No quantitative data are available on the use of estuarine habitats for foraging by this species, either during the breeding season or at other times of year. In the South Bay, this species is observed in a wide range of habitats; birds leaving the Alviso heronry fan out to forage on the mudflat tidal edge, along streams flowing into the Bay, and the salt ponds. At times, unusual fish concentrations occur in seasonal wetlands or salt ponds, and, at these times, unusual concentrations of herons result. Some representative high counts from the South Bay include 390 counted by Alan M. Eisner in Charleston Slough on 3 August 1992, and 340 censused by Stephen C. Rottenborn in the vicinity of the Sunnyvale Water Pollution Control Plant ponds on 24 July 1993 (Bousman 1994).

The two breeding population centers of this species in the North and South bays suggest the plasticity of this species in its use of all major estuarine habitats. South Bay observations clearly indicate the importance of salt pond habitats, as well as the tidal edge of mudflats and riparian areas, whereas in the North Bay, use of salt ponds and other impoundments is less important than foraging in tidal areas.

Conservation and Management

The basic needs of this species are secure areas for nesting, adequate wetlands for foraging, and continued protection from direct persecution by man. However, this species is still vulnerable in its limited nesting colonies as indicated by the killing of snowy egrets, along with many great egrets in the West Marin Island colony in October 1955 (Audubon Field Notes 10: 51). The protection of the two large colonies, the one on West Marin Island in the North Bay and the other along Artesian Slough in the South Bay, is the most important need for this species within the estuarine system. As a foraging generalist tied directly to numerous habitats within the Estuary, the continued health of this population depends upon the general health of the Estuary and the various prey stocks.

Population surveys of the West Marin Island colony (Shuford 1993) are shown below in Table 7.1. Although these numbers demonstrate fairly wide fluctuations in breeding numbers, they do not indicate any long-term changes. Comparable data from the Alviso heronry have not been published. Both of these major Bay colonies are presently protected, but each is vulnerable to natural hazards, as well as direct and indirect acts of man.

The Bair Island colony near Redwood City was successful from 1969 into the 1980s and was then abandoned. Except for Gill’s records (Gill 1977), data documenting the growth and decay of this colony have not been published, and there has been only limited discus-

Table 7.1 Estimated Breeding Pairs of Snowy Egrets on West Marin Island (Shuford 1993)

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sion of why the colony was abandoned. San Francisco Bay Bird Observatory records indicate that the decline of the colony is probably linked to severe predation by red fox (Cogswell, pers. comm).

The Baire Island colony example of a fairly recent birth, growth, and decay of a major colony within the estuarine system, with little documentation, remains a warning for the stewards of our estuarine system. It is unclear how the Goals Project is to set goals for estuarine health without sufficient data to examine the pathology of specific population failures or shifts.

It is believed that the greatest hazard now for this species is the continuing population increase of the non-native red fox in the South Bay. The Alviso heronry nests are largely in tules slightly above the tidal line, and although the water offers some protection from predators, the red fox has shown in its depredations on the clapper rail its willingness to overcome water barriers. As this population increases it appears only a matter of time before this colony is extirpated. It is possible that colony protection against this predator’s burgeoning population could be obtained through a carefully designed barrier that enhances the effect of the present water barrier. It is possible that this colony could be re-established on Bair Island; however, protection of that area may be even more difficult.

References

Sibley, C.G. 1952. The birds of the south San Francisco Bay region (mimeo).

Black-Crowned Night Heron

Nycticorax nycticorax
William G. Bousman

Introduction

The black-crowned night heron is a member of the family of herons and egrets (Ardeidae) and is found worldwide. In the New World, the subspecies N. n. hoactli is recognized and breeds from Oregon and southern Canada, south to Chile and Argentina. In winter, the northern populations withdraw to the southern United States, although they linger on the West Coast to Oregon and the East Coast to Massachusetts (AOU 1957). Within its breeding range, it is generally common and is dependent largely on wetlands for foraging. It is a resident species in the San Francisco Bay Area.

The black-crowned night heron generally forages at the margins of lakes and streams, on brackish and on salt waters. Its chief prey items are fish, crustaceans, insects, and amphibians normally obtained by stalking or waiting for prey from a stationary position. It normally feeds at night, dawn, or dusk on individual feeding territories and roosts during the day. When trees are available it will frequently use them for roosting, otherwise it uses tules and cattails. It tends to use less open habitats than other egrets and herons (Egretta and Ardea), but is not so secretive as the bitterns (Palmer 1962, Cramp 1977).

Locally, in the South Bay, this species is nicknamed the "night raven" for its tendencies to prey on recently hatched ducklings. On 26 June 1988, Phyllis Browning (pers. comm.) watched two herons take six Class 1 gadwall ducklings in a half-hour period in the Palo Alto...
Flood Control Basin. At the same time, a western gull made 17 attempts on a gadwall family without success.

The black-crowned night heron nests in trees and shrubs, or less frequently in tules. Although a solitary feeder, this species is gregarious at roosts and is a colonial breeder. Males establish territories within the breeding colony and will bring twigs to a nesting site as part of advertising displays. Once a pair-bond is established, the male will bring sticks to the site where the female remains. A number of social behaviors are associated with nesting pairs (Palmer 1962, Cramp 1977). The species is single-brooded and will normally lay three to five eggs. The young hatch asynchronously, and the younger (smaller) nestlings will survive only when food is plentiful.

Distribution and Abundance

**Modern Distribution** - McCaskie et al. (1979) described the black-crowned night heron as uncommon to fairly common in Northern California within the seashore, coastal, interior, and Great Basin districts, but as a vagrant in the mountain district. They characterized its habitat use as beaches, mudflats, marshes, rocky shores, and riparian areas. Garrett and Dunn (1981) noted the species as fairly common in Southern California along the coastal slope, at the Salton Sea, and along the Colorado River Valley. They recorded recent breeding from Morro Bay, Santa Barbara, and San Diego.

Current Christmas Bird Count (CBC) data from the late 1960s to the present, shown in Figure 7.2, indicate that this species is found regularly on CBCs in Benicia, Oakland, Hayward-Fremont, San Jose, Palo Alto, Crystal Springs Reservoir, San Francisco, Marin County (southern), and Arroyo Cheap Thrills. For all of these counts, the numbers are comparable. Winter populations in terms of birds/party-hour have increased for Benicia (+6.0%, p<0.005), San Jose (+3.7%, p<0.025), and Arroyo Cheap Thrills (+19.5%, p<0.025), while declines are noted on Crystal Springs Reservoir (-5.8%, p<0.005) and Palo Alto (-3.7%, p<0.025). This species is less common in the Palo Alto count circle during the summer based on Summer Bird Count (SBC) data with a mean of 0.33 birds/party-hour (std. dev. = 0.47, n=15) compared to the winter average of 0.87 birds/party-hour (std. dev. = 0.61, n=27). Interestingly, the Palo Alto SBC also shows a significant decline in the population (-2.6%, p<0.005). The black-crowned night heron appears to be common throughout the Bay Area, and is perhaps best considered a resident species, with some augmentation in winter by non-resident northern or interior birds. The aggregate number of birds counted in the nine CBC circles, calculated by summing the means of the CBCs, is 838 birds. This represents a lower bound on the wintering population of black-crowned night heron in the San Francisco Bay estuarine system.

**Historical Distribution** - Grinnell and Miller (1944) considered the black-crowned night heron to be somewhat common throughout the State in the summer with fewer birds present in the winter. They noted that it occurred on both sides of the Sierran crest and bred from the lower Sonoran to the Transition life zones. They commented that this heron was formerly abundant in some localities, but numbers had been greatly depleted in historical times. Within San Francisco Bay, they cited breeding records from Belvedere Island in Marin County, and Alameda and a location near Alvarado in Alameda County.

Sibley (1952) noted that the Alameda County nesting colonies were no longer active by the 1920s and that there were no longer any active colonies in the South Bay. Emily Smith (Audubon Field Notes 8: 359) considered 27 birds counted on 11 July 1954 and 21 on 25 July 1954 at Alviso to be an unusual concentration. A nesting colony was established on Bair Island near Redwood City in 1967, and this included at least 684 nests in 1971 (Gill 1977). This colony was later abandoned because of red fox incursions (H. Cogswell, pers. comm.).

This species has clearly recovered in recent decades to where there are relatively stable populations, although it is unclear whether these are as large as existed prior to European settlement. Unlike the snowy egret, it is not apparent that this bird was hunted for its plumes at the end of the last century, and the down turn in its population must be related to other factors. Without knowing what these factors were, it is difficult to determine with any certainty why the species has recovered. It is possible, however, that the decline of duck hunting in the South Bay has benefited this species, as well as a number of other non-game species that were targets of casual hunters.
Habitat Use and Behavior

The black-crowned night heron is a generalist in its foraging, as with many of the herons, and uses a variety of habitats. Unlike some of the other herons, however, it is a solitary feeder and does not normally concentrate at drying ponds and fish blooms. Suitable habitat requires numerous foraging opportunities in either fresh, brackish, or salt water. A suitable roost site for this crepuscular species is required as well.

The black-crowned night heron uses a variety of substrates for nesting. On West Marin Island, it uses coastal scrub and sometimes buckeye for their nests (Shuford 1993). At Alviso, they nest in tules along Artesian Slough. There is some evidence that introduced eucalyptus provide particularly safe nest locations because of the smooth bark (Roberson and Tenney 1993). The substrate per se appears to be relatively unimportant for this species, and the primary need is for a nest site that is safe from predation. Up to 300 pairs have nested on West Marin Island (Shuford 1993), and this is the densest concentration in the Bay Area. Up to 150 pairs were counted nesting in the Artesian Slough colony in the South Bay in 1980 (Am. Birds 34: 811). Outside of the Bay Area, breeding is less common in coastal areas. A small colony exists in Monterey County at Carmel Point in eucalyptus. A maximum of 13 nests were counted in 1992 (Roberson and Tenney 1993). At least three small colonies are located in Sonoma, largely in urban areas where they create some difficulties for the local (human) residents (Burridge 1995).

Conservation and Management

The basic needs of this species are secure areas for nesting, adequate wetlands for foraging, and the continued protection from direct persecution by man. The protection of the two large colonies, one on West Marin Island in the North Bay and the other along Artesian Slough in the South Bay, is the most important need for this species within the estuarine system. As a foraging generalist tied directly to numerous habitats within the Estuary, the continued health of this population depends upon the general health of the Estuary and the various prey stocks. Population surveys of the West Marin Island colony (Shuford 1993) are shown in Table 7.2 and,

Figure 7.2 Christmas Bird Count data for Black-Crowned Night Heron — Relative geographic location of Count circles indicated by dashed lines

National Audubon Society's Audubon Field Notes, and its successor publications, American Birds, and Field Notes, Volumes 24-51.
Although these data demonstrate fairly wide fluctuations in breeding numbers, they do not indicate any long-term population changes. Comparable data from the Alviso heronry have not been published. Both of these major San Francisco Bay colonies are presently protected, but each is vulnerable to natural hazards, as well as direct and indirect acts of man. We have, at best, only a poor understanding of the factors that determine whether a nesting colony will succeed or fail. As an example, the Bair Island colony near Redwood City was successful from 1969 into the 1980s and was then abandoned. Except for Gill's records (Gill 1977), data that document the growth and decay of this colony have not been published, nor has there been a detailed discussion as to why the colony was abandoned. The fairly recent history of birth, growth, and decay of a major colony within the estuarine system, with little documentation, remains a warning for the stewards of our estuarine system.

It is believed that the greatest hazard now for the black-crowned night heron is the increasing population of the non-native red fox in the South Bay. The Alviso heronry nests are largely in tules, slightly above the tidal line and, although the water offers some protection from predators, the red fox has shown in its depredations on the clapper rail its willingness to overcome water barriers. As the fox population increases, it appears only a matter of time before this colony is extirpated. It is possible that colony protection against this predator's burgeoning population could be obtained through a carefully designed barrier that enhances the effect of the present water barrier. It is possible that this colony could be re-established on Bair Island; however, protection of that area may be even more difficult.

No quantitative data are available on the use of estuarine habitats for foraging by this species, either during the breeding season or at other times of year. In the South Bay, this species is observed in a wide range of habitats, and birds leaving the Alviso heronry fan out to forage on the mudflat tidal edge, the salt ponds, and along streams flowing into the Bay. This species does not concentrate at prey resources, such as fish blooms or drying ponds as some of the other Ardeids, but it does congregate at secure day roosts. A representative day roost was of 145 birds tallied at the Palo Alto Baylands on a high tide on 22 November 1984 (pers. obs.).

References


Sibley, C.G. 1952. The birds of the South San Francisco Bay region (mimeo).

Personal Communications

Phyllis Browning, birdwatcher and Santa Clara Valley Audubon member, Palo Alto, California.

Howard Cogswell, Professor Emeritus, California State University, Hayward, California.

Table 7.2 Estimated Breeding Pairs of Black-Crowned Night Herons on West Marin Island (Shuford 1993)

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California Clapper Rail  
*Rallus longirostris obsoletus*

Joy D. Albertson  
Jules G. Evens

**Introduction**

Populations of clapper rails along the Pacific Coast and the Colorado River have been considered variously as races of *Rallus longirostris*, races of *Rallus elegans*, or separate species (AOU 1983). Currently, *R. longirostris* and *R. elegans* are treated as superspecies (Taylor 1996).

Three subspecies of clapper rail (*Rallus longirostris*) occur in California: the 'California' clapper rail (*R. l. obsoletus*), the 'light-footed' clapper rail (*R. l. levipes*), and the 'Yuma' clapper rail (*R. l. yumanensis*) (AOU 1957). Each of the three subspecies is classified as 'endangered' by the U.S. Department of the Interior (Federal Register 50 CFR 17.11; USFWS 1973), primarily due to habitat loss. Both *obsoletus* and *levipes* are listed as 'endangered' by the State of California; *yumanensis* is listed as 'threatened' (CCR Title 14, Section 670.5).

The California clapper rail is a secretive, hen-like waterbird, indigenous to estuarine marshlands in San Francisco Bay (Grinnell and Miller 1944, Gill 1979, AOU 1983). Though *R. l. obsoletus* may still occur as a transient in outer coast marshes, its status there is precarious. No breeding populations have been detected at Morro Bay and Elkhorn Slough in recent years. (AOU 1983). Though *R. l. obsoletus* and *R. l. levipes* are known to inhabit the marshes of the Pacific Northwest (AOU 1957), the nests are listless as accurately as 'resident at Tomales Bay and Monterey Bay' (AOU 1957) are no longer accurate.

**Description**

The clapper rail is one of the largest species of the genus *Rallus*, measuring 31-40 cm in length and weighing approximately 250-350 grams, with the males slightly larger (Taylor 1996). It has a rusty or rufous breast, orange bill, white and black feathers on the flanks, as well as white undertail coverts, creating effective camouflage within the marsh vegetation (Ridgeway 1880, Grinnell et al. 1918).

**Breeding Biology** – Clapper rail pairs are monogamous and will fiercely defend overlapping, year-round territories (Applegarth 1938, M assey and Zembal 1987, Zembal et al. 1989, Albertson 1995). Courtship is initiated by the male and involves the male approaching the female with an uplifted tail, pointing his bill to the ground and swinging it from side to side. Courtship feeding of the female is also common. Males normally build the nest, which occurs on or near the ground, usually on a slight rise (Ehrlich et al. 1988). The nest consists of a platform of dead plant material arched over by surrounding live vegetation to form a roof. In the South Bay, nests have primarily been found in gumplant bushes (Grindelia humilis), pickleweed clumps (*Salicornia virginica*), cordgrass stands (*Spartina foliosa*), saltgrass patches (*Distichlis spicata*), and wrack (DeGroot 1927, Applegarth 1938, Zucca 1954, Harvey 1988, Foerster et al. 1990). In the North Bay, nests have been found in *Scirpus robustus*, *Salicornia virginica*, or *Grindelia humilis*. Nests tend to be located less than two meters from first-order channels and at least 100 meters upstream from the marshland shoreline (Evans and Page 1983, Evans and Collins 1992). The marshland beneath the nests ranges in elevation from 15 cm below MHHW (Mean High Water) to about five cm above MHHW, and the nests themselves are constructed entirely above MHHW (Evans and Collins 1992, Collins et al. 1994).

Eggs are laid March through July (DeGroot 1927, Harvey 1980, Evans and Page 1983). A clapper rail can lay between five and 14 eggs, with the average being seven eggs per clutch (DeGroot 1927, Zucca 1954). Incubation is shared by both adults and is variously reported as 23 to 29 days (Applegarth 1938, Zucca 1954) and 18 to 29 days (Taylor 1996). The peak nesting period for clapper rails is April through May, and a majority of hatching occurs mid-April through early June (Applegarth 1938, Zucca 1954, Harvey 1988, Foerster et al. 1990). DeGroot (1927) states that clapper rails may “double clutch,” or produce two broods per year. However, Applegarth (1938) attributes late nesting attempts to renesting, occurring when the first nest has failed, rather than to production of a second clutch after a successful hatch.

Clapper rail chicks are precocial and will leave the nest soon after hatching (Applegarth 1938). One adult will tend the newly hatched chicks, while the other parent continues incubation until all eggs have hatched (Applegarth 1938, Meany 1985). Young rails accompany the parents for approximately eight weeks, learning to forage for food on their own (DeGroot 1927, Zembal 1991). Juveniles fledge at ten weeks (Johnson 1973) and may breed in the spring following hatch.

Survivorship is low, 0.49-0.52 (Albertson 1995), similar to that of the Yuma clapper rail (Eddleman 1995).
Much predation takes place during high winter tides and is likely due to the ease of capture by predators at this time. This increased predation is likely enhanced by the increased movement of rails within this season, similar to other clapper rail subspecies (Eddleman 1989, Zembal et al. 1989). Raptors, in particular, gain advantage during high tide in marshes that do not have sufficient high vegetation to provide aerial cover for rails. In one study, most (64%) of the rails killed were taken by raptors, primarily during the winter season (Albertson 1995). In another study, an estimated 25% of the population of rails in one 35 ha marsh was taken by raptors from April through November; circumstantial evidence indicated that the barn owl (Tyto alba) was the primary predator at that site (Evens and Page 1983). In that study, predation of nests and eggs was also attributed to rats, ravens, and high tides (Evens and Page 1983).

**Migration Ecology** - California clapper rails are considered non-migratory residents of San Francisco Bay salt marshes, but post-breeding dispersal has been documented during the fall and early winter (Orr 1939, Wilber and Tomlinson 1976). Harvey (unpubl. data) reported three of 54 banded birds moving approximately one km across a slough and one moving about 10 km, from Dumbarton Point to Alameda Creek. Most birds, however, did not move from the marsh in which they were banded: 48% were found 100 m or less from the capture sight, and 78% were less than 500 m away. Albertson (1995) reported one of 29 monitored rails moving approximately three kilometers in early breeding season and successfully establishing a breeding territory. Old records from the Farallon Islands, outer coast marshes, and a variety of extralimital locations (Grinnell and Miller 1944; Evens, unpubl. field notes; American Birds notebooks) suggest that there is a fairly regular fall dispersal period from August through November. This dispersal may be irruptive in nature, occurring in some years, not in others.

**Food and Feeding** - The primary diet of clapper rails consists of various invertebrate species, including mussels, clams, crabs, snails, amphipods, worms, spiders, insects, and fish (Williams 1929, Applegarth 1938, Moffitt 1941). In addition, clapper rails will opportunistically take small birds (Jorgensen and Ferguson 1982) and rodents (pers. obs.), as well as carrion (Moffitt 1941). A majority of foraging occurs during low tide when mudflats and tidal sloughs are exposed, and food is more readily available (Applegarth 1938, Foerster and Takekawa 1991).

**Distribution and Abundance**

**Historical Distribution** - The historical distribution of the California clapper rail was restricted to the tidal marshlands of coastal California from Humboldt Bay in the north to Morro Bay in the south (Grinnell 1915, Grinnell and Wythe 1927, Grinnell and Miller 1944, AOU 1957, AOU 1983, Gill 1979). It occurred formerly at Humboldt Bay (Grinnell and Miller 1944); Tomales Bay (Storer 1915, Brooks 1940, Grinnell and Miller 1944, AOU 1957); Elkhorn Slough, Monterey County (Grinnell and Miller 1944, Varoujian 1972); and Morro Bay, San Louis Obispo County (Brooks 1940, AOU 1957). Present distribution is restricted to the tidal marshes of San Francisco Bay (Evens 1985, Baron and Takekawa 1994). Recent records from coastal estuaries outside of San Francisco Bay are sporadic and represent presumed dispersants or vagrants.

The historical distribution within San Francisco Bay was apparently restricted to tidal marshlands downstream from Suisun Bay (Grinnell 1915, DeGroot 1927, Grinnell and Wythe 1927, Moffitt 1941, Grinnell and Miller 1944). The literature suggests that the populations have fluctuated widely in historic times. A decline noted in the 19th century was attributed to predation by hunters (Taylor 1894). DeGroot (1927) implies that numbers declined around the turn of the century in the North Bay, and Grinnell and Wythe (1927) reported that although still common in the South Bay, they had become rare elsewhere around the Bay. By 1944, Grinnell and Miller stated that clapper rails had recently recolonized former habitat “in marshes on northern and eastern sides of the Bay.” This wording, along with a distribution map delimiting the range at Carquinez Strait (Grinnell and Miller 1944), suggests the North Bay population was limited to the shores of San Pablo Bay. It seems clear that, historically, clapper rails were restricted to the tidal marshes of San Francisco and San Pablo bays, but were absent from Suisun Bay and associated marshlands (Collins et al. 1994).

**Modern Distribution** - Data on current population levels is somewhat equivocal and may indicate fairly wide population fluctuations, or partial knowledge of abundance, among sub-regions of the Bay. Based on surveys conducted from 1971 through 1975, Gill (1979) estimated a population of 4,200-6,000 rails with 55% in the South Bay, 38% in the Napa marshes, and the remaining 8% in other North Bay and outer coast marshes. There is some indication that Gill overestimated; however, the weight of the evidence suggests that the decline in the population has been real, at least since the mid-1980s (J. Takekawa, pers. comm.). By the mid-1980s, on the basis of breeding and winter population estimates, approximately 1,200-1,500 California clapper rails remained, with greater than 80% of the population found in the South Bay (H Harvey 1980, H. Harvey 1988, T. H. Harvey and P. Kelly, unpubl. data). By 1988, populations were estimated to have declined to 700 rails (Foerster and Takekawa 1991), with one of the primary causes of this decline being predation caused by the introduction of the red fox. One estimate suggested that...
the South Bay supported up to 90% of the total rail population (SFEP 1992), however, the most recent update suggests a more even distribution between the South and North bays (see below).

In 1990-91, the Bay-wide population was estimated as 300-500 individuals, followed by a rebound in 1993 to over 800 individuals (USFWS unpubl. data). Increases in South Bay rail populations during this time period are largely attributable to ongoing predator management, initiated in 1991 (Harding et al. 1998). Winter surveys conducted in 1997-98 estimated the South Bay populations to be 650-700 individuals (USFWS unpubl. data). The most recent comprehensive surveys (1992-93) in the northern reaches of the Bay (San Pablo and Suisun bays) resulted in conservative estimates of 195-282 breeding pairs, or 390-564 individuals (Collins et al. 1994). Given these data (and their limitations) the most recent estimates indicate a total population of 1,040-1,264 rails in San Francisco Bay. Figure 7.3 shows the known distribution of California clapper rails around the San Francisco Bay.

Carquinez Strait - Habitat is limited along the Strait, but a small population (estimated at one to three pairs) has persisted at least since 1948 at Southhampton Marsh (Evans and Collins 1992).

Suisun - Clapper rails are scattered at several sites around Suisun Bay and Marsh, with an indication that populations are present some years but not others. Areas where rails have been found with some regularity since 1978 include the shoreline marshes from Martinez east to Point Edith, bayshore marshes near the mouth of Goodyear Slough, the upper portions of Suisun and Hill sloughs (B. Grewell, pers. comm.), and the western reaches of Cutoff Slough and associated tributaries (H Arvey 1980). Winter records appear to be more numerous in this region than breeding season records. An apparent range expansion into this area probably indicates habitat changes resulting from conversion of these marshes into more brackish condition with substantial decreases in freshwater flow from the Sacramento-San Joaquin Delta (Rozengurt et al. 1987, Evans and Collins 1992, Leipsic-Baron 1992.)

North Bay - The marshlands along the North Bay shore and associated rivers and sloughs support clapper rails, with concentrations near the mouths of the larger tributaries (e.g., Gallinas Creek, Novato Creek, Petaluma River, Black John Slough, Sonoma Creek, and Napa River). Gill (1979) identified the Napa River as a North Bay population center which supported “40% of the entire population.” Subsequent field work (Evans and Collins 1992, Collins et al. 1994) suggests a decline at that site, but concentrations still exist at White Slough (Vallejo) and Coon Island (Evans and Collins 1992).

Central Bay - A relatively small extent of appropriate habitat occurs in the Central Bay. Primary areas that support clapper rails are: Corte Madera (aka ‘H eerdt’) and Muzzi marshes (30 pair in 1992-93; Collins et al. 1994); San Leandro Area (Arrowhead and Elise Romer marshes); and inner Richmond Harbor (Collins et al. 1994). Muzzi Marsh is of particular interest because it is a restored marsh that was not colonized until 1984, and the population was estimated at 15 rails in 1987 (Evens and Page 1987). Other sites include Richardson Bay and Creekside Marsh, Marin County.

South Bay - Foerster (1989) indicated that California clapper rail numbers on the western side of the Bay were stable, but the East Bay population (primarily in Ideal, Dumbarton, and Mowry marshes) had decreased substantially during the past decade, from 400-500 individuals in the early 1980s, to 50-60 in 1991-1992 (H Arvey 1980, USFWS unpubl. data). Recent surveys show strong recovery of East Bay populations following implementation of a predator management program to control red foxes (Harding et al. 1998), with over 330 counted in 1997-98 winter surveys. Currently, the largest populations of rails occur in Dumbarton and Mowry marshes on the East Bay, and Palo Alto and Greco marshes on the West Bay. The most recent survey data indicate that rail populations on the east and west sides of the South Bay are approximately equal.

Habitat Use and Behavior

Habitat Requirements - The California clapper rail occurs primarily in emergent salt and brackish tidal marshlands of San Francisco Bay. Preferred habitat is subject to direct tidal circulation and is characterized by predominant coverage by pickleweed (Salicornia virginica) with extensive stands of Pacific cordgrass (Spartina foliosa), and, in the North Bay, Scirpus robustus, abundant high marsh cover, and an intricate network of tidal sloughs which provide abundant invertebrate populations (Grinnell et al. 1918, DeGroot 1927, H Arvey 1988, Collins et al. 1994) as well as escape routes from predators (Zembal and Massey 1983, Foerster et al. 1990). Generally, the upper marsh zone is dominated by pickleweed, with saltgrass (Distichlis spicata), alkali heath (Frankenia grandifolia), and jaumea (Jaumea carnosa) occurring at the highest elevations, as well as gumplant (Grindelia spp.) along the upper edge of some tidal sloughs. The lower marsh zone along the Bay is dominated by stands of Pacific cordgrass, which also occurs along the banks of tidal sloughs within the marsh (DeGroot 1927, H Arvey 1988). Low marsh areas with sparse vegetation, mudflats, and tidal sloughs are important foraging areas for rails (Applegarth 1938, Albertson 1995). Higher marsh areas with dense vegetation are used for nesting and high-tide refugia (DeGroot 1927, H Arvey 1988, Foerster et al. 1990, Evans and Collins 1992, Collins et al. 1994).

Past studies (Applegarth 1938, Zucca 1954, Jorgensen 1975, Massey et al. 1984, H Arvey 1988) re-
ported on the importance of cordgrass as a canopy and nesting material. This apparent preference for cordgrass may be tied to the fact that cordgrass grows primarily along tidal sloughs and at the marsh edge, where rails prefer to forage. Weather-influenced changes in Spartina biomass and productivity may account for variations in nesting habitat preference (Gill 1979).

California clapper rails also occur in brackish wetlands consisting of bulrush (Scirpus spp.) (Gill 1979). In these areas, rails use bulrush plant materials for nest building and cover, but nests are still associated with tidal channels, as in pickleweed dominated marshes (Evens and Collins 1992). This type of habitat occurs along the larger creeks in the South Bay, in some areas of Napa Marsh, Petaluma River, and Sonoma Creek in San Pablo Bay, and in Suisun Bay (Gill 1979).

In the North Bay, natural habitat for obsoletus is the saline and marginally brackish tidal marshland with small channels that extend through or into patches of tall monocot vegetation. The ecological functions of salinity and tidal action are unknown. The tidal channels serve an important function as areas for foraging and as protected pathways. The monocot vegetation is used as nesting material. At marshland elevations near Mean High Water (MHW), the vegetation must be at least 50 cm tall to permit the construction of a nest that is low enough to be concealed by the natural plant canopy and yet high enough that it will not be inundated by the maximum high tides of the breeding season. At marshlands of higher elevation, shorter vegetation may be utilized.

Rail density seems to be positively correlated to channel density, although minimum and maximum values of channel density are not obvious from the data collected thus far. Suitable habitat is provided by most of the youthful marshlands that have evolved since the middle of the last century, as well as the remaining frag-
ments of historical, mature marshlands. Local populations of breeding California clapper rails are most dense where patches of habitat are at least 100 ha in size. Such patches typically comprise some historical and youthful marshlands together. These marshlands may support relic populations of obsoletus. Fewer than fifteen such patches exist within the northern Estuary. One third of these adjoin the mouths of major tributaries downstream from Carquinez Strait. Small parcels of marshland along the immediate margin of a major tributary seem more likely to support obsoletus than similar sized parcels that are more isolated. In general, the density of rails decreased upstream toward the headward extent of the major tributaries of the Estuary (Collins and Evens 1992, Collins et al. 1994).

Other physical attributes of a marsh that influence rail use and may contribute to creating a self sustaining population of rails include size of the marsh, location relative to other marshes, buffer areas between marsh and upland, marsh elevation, and hydrology (Collins et al. 1994, Albertson 1995). These “high quality” characteristics play an integral part in the everyday survival of the clapper rail, providing food resources, cover from predators, breeding and nesting habitat, and refuge areas at high tides. Hence, the quality of a marsh will determine how many rails can be supported in a particular marsh (Garcia 1995, Albertson 1995).

There are few records of breeding rails utilizing diked marshes or other non-tidal habitat, but one observer (K. Rambo, pers. comm., in Orton-Palmer and Takekawa 1992) documented a successful breeding pair in a sewage oxidation pond, and Orton-Palmer and Takekawa (1992) documented use of a diked marsh by a breeding pair at Moffett Field. One pair was noted breeding in a small diked marsh adjacent to a larger undiked wetland at Muir Marsh, Marin County in 1992, and a pair apparently bred in a sewage pond adjacent to Richardson Bay in Mill Valley in 1997. Close proximity of tidal marshes supporting other breeding rails are thought to contribute to the use of these non-tidal areas.

Movement - Results of recent radio-telemetry studies indicated that most rails showed strong site tenacity, with very little movement between seasons and small core-use area (x=0.87 ha) which was defended throughout the year (Albertson 1995). This same observation has been made for Yuma clapper rails (Tod 1987) which were found to vocalize only in core-use areas, demonstrating probable territorial defense areas. Home ranges sizes vary by individual bird, but significant within-season differences are apparent among marshes, particularly in core-use areas (Albertson 1995). Core-use areas are defined as the highly defended portion of the territory (per Hinde 1954) and contain the nest site. Although the basis for differences in rail home range size between marshes has not been absolutely determined, differences in predation pressure, quality of habitat, and size/orientation of the marsh may account for most of these differences (Albertson 1995).

Conservation and Management

Numerous human-related factors, including commercial and sport hunting during the late 1800s (DeGroot 1927, Wilber and Tomlinson 1976, Gill 1979), have led to rail population declines over the last 150 years (Harey 1988, Foerster et al. 1990). The Migratory Bird Treaty Act (1913), which restricted harvest on game species including the clapper rail, is believed to have led to a substantial recovery of populations in many remaining marshes (Bryant 1915, Moffitt 1940, Grinnell and Miller 1944). During the early to mid-1900s, commercial and urban development destroyed over 85% of the primary tidal marshes of San Francisco Bay, resulting in severe rail population declines, range contraction, and fragmented distribution (Gill 1979, USFWS 1984, Nichols et al. 1986, Foerster et al. 1990, Leipsic-Baron 1992).

Presently, California clapper rail populations are restricted to fragmented salt marshes in San Francisco Bay. Remaining marshes are geographically disjunct, and characterized by lack of a significant transition zone to terrestrial habitat, relatively small size, a large edge to area ratio, and close proximity to urban and industrial development. Several factors have previously been identified as negatively affecting current rail populations, including predation by non-native red fox (Foerster et al. 1990, Albertson 1995), contaminants (Ohlendorf and Fleming 1988, Ohlendorf et al. 1989, Lonzarich et al. 1990, Leipsic-Baron 1992), and marsh conversion and degradation (Foerster and Takekawa 1991). Predation is likely their most immediate threat for survival.

Clapper rail (Rallus sp.) populations are subject to predation by a number of species. At least ten native and three non-native predators are known to prey on California clapper rails and their eggs (Albertson 1995). However, recent evidence suggests that the non-native red fox may pose the most serious threat to adult clapper rails (USFWS and USN 1990, Foerster et al. 1990, Foerster and Takekawa 1991, Zembal 1992, Albertson 1995).

During the late 1800s, the red fox (Vulpes fulva) was introduced to western California by hunters. In addition, some individuals escaped from commercial farms (Lewis et al. 1993). Red foxes are well adapted to urban environments, and thus their populations have rapidly expanded along the coast in such areas as San Francisco Bay (Albertson 1995).

In addition, free-roaming and feral cats (Felis domesticus) prey on rails in marshes adjacent to housing and landfill areas (Albertson 1995). The Norway rat (Rattus norvegicus), raccoons (Procyon lotor), and non-native foxes prey on eggs and may cause low annual recruitment (DeGroot 1927, Applegarth 1938, Harey 1988, Foerster et al. 1990). Harey (1980) found that...
rungs took 24% of eggs in 50 nests, and a 1992 study in the South Bay showed that rats preyed on 31% of eggs in 54 monitored nests (USFWS unpubl. data).

Extensive conversion of tidal lands resulting from historic and ongoing pressures of agricultural production, urbanization, and salt production has drastically reduced California clapper rail habitat in the San Francisco Estuary. The remnant tidal marshlands of the Estuary, the largest and last refuge of obsoletus, occupy only about 15% of their historic extent (D edrick 1989), yet even in such diminished capacity comprise greater than 90% of all remaining California tidal marshlands (MacDonald 1977).

Fragmentation of habitat involves the construction of dikes and levees that serve as corridors for terrestrial predators. The devastating effect of red fox on obsoletus in the southern reaches of the Estuary is well-documented (Foerster et al.1990, Burkett and Lewis 1992), and has been noted in regard to R. l. levipes in southern California (Zembal in Foerster et al. 1990). In the northern reaches of the Estuary, the distribution of red fox is disjunct. They are reported west of Benicia and east of Dixon, but not in the Suisun system (Bob Smith, pers. comm.). We observed red fox at Wildcat, Point Pinole, China Camp, M are Island, Second N apa Slough, and Dutchman Slough. N one were detected in the Suisun Marsh, where coyote (Canis latrans) was quite common. We also noted sign of raccoon (Procyon lotor), striped skunk (Mephitis mephitis), feral cat, and rats (Rattus spp.). Raccoon sign was noted in virtually every tidal marsh, and we suspect that, as in the southern reach (Foerster et al. 1990), raccoon populations as well as those of other mesopredators have increased dramatically over the last decade. In addition, river otter (Lontra canadensis) is quite common in the Suisun Marsh area and should be considered as a potential predator of birds that nest on the ground near water (Ingles 1965).

According to MacArthur (1972), colonization among isolated patches of habitat requires very high fecundity of source populations. Given the effects of fragmentation, increased predation rates, possibly lowered fecundity (Foerster et al. 1990), and a low rate of dispersal, California clapper rails probably can not colonize or survive in all fragments of their habitat. Mortality due to predation could be so severe that immigration from source populations is infrequent, and the rate of survival of pioneering individuals is low. This might explain their absence in ostensibly suitable habitat observed in this study.

As with the California black rail (Laterallus jamaicensis coturniculus) and other tidal marsh dependent species (Evens et al. 1991, N ur et al. 1997) subject to similar pressures, simulation models have demonstrated that populations of fewer than 10 pairs (Richter-Dyn and Goel 1972, Roth 1974), or perhaps 25 pairs (Shaffer 1981), are inherently unstable and tend toward extinction due to stochastic events. Due to these factors, the persistence of subpopulations may depend on contiguity of marsh parcels and ability of rails to disperse among sites.

In addition to habitat fragmentation and the concomitant threat of predation, other pressures that threaten to alter or degrade the habitat and impact rail populations include: continued diversion of freshwater inflow from the North Bay (Rozengurt et al. 1987, Williams and Josselyn 1987); a progressive rise in sea level (Williams 1985, Moffatt and Nichol et al. 1987); and contamination. Contamination of other species of estuarine birds has been documented in this estuary (Ohlendorf et al. 1986, Ohlendorf and Fleming 1988, Phillips and Spies 1988), and other studies have detected dangerous levels of contaminants in clapper rail eggs (Lonzarich et al. 1990). The threat of toxic contamination of the substrate is ongoing, cumulative, and poorly understood.

**Recommendations**

The quality of restored habitat will dictate the potential rail population density. High quality habitat should include:

- **Direct tidal circulation sufficient to allow the full tidal cycle,**
- **A predominant pickleweed marsh with cordgrass, gumplant, and other high marsh plants,**
- **Abundant, dense high marsh cover, and**
- **An intricate network of tidal sloughs.**

In addition to these qualities, other physical characteristics of marshes that should be taken into account when planning a restoration project include size of marsh, location relative to other marshes, buffer area between marsh and upland, and type/extent of interface with humans and human-made structures.

Small sites may not provide enough habitat for a viable population of breeding birds. Ideally, restored areas should be able to potentially support viable rail populations and not be subject to wide population fluc-
tations. Therefore, marsh restoration sites should be at least as large as existing sites, such as Dumbarton Marsh (118 ha) or Mowry Marsh (164 ha). Marsh restoration should focus on significantly expanding marsh acreages in areas currently supporting high populations of clapper rails, such as the Dumbarton, Mowry, Greco, and Palo Alto areas in the South Bay. In the North Bay, historic wetland acreage in close proximity to rail populations provides opportunities for restoration, for example White Slough and Coon Island on the Napa River, diked areas adjacent to Sonoma Creek and the Petaluma River, and in large diked areas associated with Suisun Slough in Suisun Bay. This would allow existing populations to increase, reducing the probability of local extinctions.

Location of marshes with respect to one another should also be taken into consideration when planning restoration. Areas to be managed for clapper rails should be in close proximity to facilitate dispersal without risk of excessive predation. Rails have been found to disperse between 1-3 km, therefore primary marshes should be within this distance from one another. Intervening marsh corridors need to be of sufficient quality and width (at least several hundred feet) to provide adequate cover and food resources for dispersing rails.

Buffer areas between marsh and upland are critically important to rails for escape cover from predators during high tides. Absence of higher transitional areas adjacent to a marsh could result in high mortality during periods of tidal inundation. In a telemetry study (Albertson 1995), much of the predation was found to occur during high tides, when cover was scarce and little refugial area was available. Buffer areas need to provide cover of sufficient height and density to protect rails during extreme high tides. Marsh restoration projects should incorporate gradual transition areas from marsh into upland whenever possible, rather than an abrupt elevation change from marsh to levee.

Human-made structures, such as power lines, poles, and buildings, provide raptor perches. If these structures are in close proximity to marshes, predation by raptors can be high (Albertson 1995). Marsh restoration projects should take this into account and plan to remove structures if possible, or plan larger restoration projects to minimize the effects of such structures. Human dwellings, landfills, and rubble piles can harbor Norway rat and feral cat populations, therefore marshes in these areas may be subject to high levels of predation from these species. Human disturbance from recreational use, utilities maintenance, and high-intensity adjacent uses can disturb rails and cause homing abandonment with subsequent nesting failure. Proposed use of adjacent land and public access to marshes should, therefore, be carefully evaluated prior to being permitted.

Habitat restoration should include management of non-native plant species, such as smooth cordgrass (Spartina alterniflora) and pepperweed (Lepidium spp.), in order to protect existing and future rail habitat from degradation. In particular, invasion of smooth cordgrass causes excessive sedimentation, which will clog tidal sloughs important to rails for foraging. A Bay-wide effort must be undertaken to control smooth cordgrass.

Continued predator management will be needed to maintain viable clapper rail populations and prevent extinction. Control efforts should target red fox, feral cats, and selected known native predators in areas of prime rail use. Structures and debris that harbor rats (nip-rap, decrepit buildings and vessels, etc.) should be removed from areas adjacent to rail habitat.

Predator management is not the solution to increasing future populations, however. In the long-term, only restoration of high quality tidal marsh habitat will ensure future survival and recovery of the California clapper rail. The current amount and configuration of suitable habitat is insufficient to substantially increase rail densities and population sizes. Several tidal restoration projects around the Bay are being undertaken by the San Francisco National Wildlife Refuge complex and others. One 1,500 acre parcel, Cullinan Ranch, formerly farmland along the Napa River area of the Nort Bay is being restored to full tidal action by means of levee breaching and sedimentation. The Knapp property, a 452 acre former salt pond in Alviso and Guadalupe sloughs, will be restored to tidal marsh. Bair Island, a 1,600 acre former salt pond will be restored to tidal action. This area holds much promise for rail recovery due to its large size and close proximity to another fairly large rail population on nearby Greco Island. Additional opportunities exist in San Pablo and Suisun bays.

Recovery of California clapper rail populations will require the preservation of existing habitat and restoration of large acreages of high quality tidal marshes. In order to afford this species the best chance for recovery, restoration of former salt and brackish tidal marsh areas should be maximized in all subregions of San Francisco Bay. Restoration should focus on areas that have the greatest potential for developing into high quality salt marsh habitat.

References


Applegarth, J.H. 1938. The ecology of the California clapper rail in the south arm of the San Francisco


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Grinnell, J. 1915. A distributional list of the birds of California. Pacific Coast Avifauna No. 11.

[Other Birds]
California Black Rail

Laterallus jamaicensis coturniculus

Lynne A. Trulio
Jules G. Evens

Introduction

Two subspecies of the black rail breed in North America, the eastern black rail (Laterallus jamaicensis jamaicensis) and the California black rail (L. j. coturniculus). The bulk of the population (>90%) is associated with the tidal marshlands of the San Francisco Estuary. The species' reliance on tidally influenced, completely vegetated, high elevation salt marsh habitat makes it a valuable indicator species of mature, upper tidal marsh habitat.

The California subspecies is a sparrow-sized bird, approximately five to six inches long. Cogswell and Christman (1977) note that this secretive bird can be positively identified by its "dark slate color, with faint white bars on the sides and chestnut nape and the prominent white spotted back." The California black rail is found primarily in three locations on the West Coast: the San Francisco Estuary and local coastal marshes, the lower Imperial Valley and the lower Colorado River at the border of California and Arizona, and northwestern Baja California (Eddleman et al. 1994).

The discovery in 1994 of a black rail population in freshwater marsh habitat in the Sierra foothills east of Marysville (Aigner et al. 1995) suggests that other unknown populations also may exist. Indeed, in 1997, systematic surveys of potential habitat in the vicinity of this 1994 site detected small populations scattered throughout the foothills, mostly between 100 and 200 meter elevations. Of more than 100 sites surveyed, approximately one quarter supported black rails in 1997 (J. Tecklin, pers. comm.). Subsequent field work detected rails at 71 sites in the lower foothills of three counties, extending from north of Chico on Butte County, south through Nevada County (J. Tecklin, in press).

In the San Francisco Bay Area, this rail is primarily a bird of tidally influenced marshes and is most often seen during very high tides when it is forced out of the lower elevation pickleweed marsh.
Other Birds

Description

Because the California black rail is so furtive, very little is known about its ecological requirements, although recent studies have begun to reveal some of these mysteries (Flores and Eddleman 1993). Old records from San Diego Bay and more recent surveys in San Francisco Bay indicate that birds may begin the breeding season (as evidenced by calling) as early as mid-February at coastal locations, later in the interior. During breeding season, birds call primarily during twilight hours. Males and females are distinguished by their very different calls, although much remains to be learned about the vocalizations of this species.

Breeding Biology - Nesting occurs in tall grasses or marsh vegetation, and nests consist of a small, woven cup of marsh plants, reeds, or grasses constructed by the male and female. In San Francisco Bay, nests with eggs have been found in April (J. Evens, pers. obs.), in San Diego Bay (where the species no longer breeds) in May (Cogswell and Christian 1977). Limited data suggest that San Francisco Bay rails lay 6 ± 1.4 eggs (Wilbur 1974). Data from non-tidal Arizona habitat showed that both males and females may incubate, that incubation is 17 to 20 days, and that second and replacement clutches are possible (Flores and Eddleman 1993). Behavior in tidal marshes in San Francisco Bay may differ from Arizona, however.

The first known record of the species in California was of a single individual on the Farallon Islands, 48 km west of San Francisco, October 18, 1859 (Brewster 1907). There are no reliable records of birds breeding in San Francisco Bay before 1970 (Evens et al. 1991), but undoubtedly this species has been resident since before European colonization. Recent evidence confirms breeding in the North Bay at China Camp, Black John Slough and Day Island, Marin County, and Sonoma Creek, Sonoma County (J. Evens, 1986-94 unpubl. field notes), however, presence of territorial birds during the breeding season implies breeding throughout the tidal marshlands of San Pablo Bay, Carquinez Strait and Suisun Bay, and at sporadic sites in the Delta and on the outer coast.

Single nest or juvenile observations have been recorded at Alviso, Newark, Richardson Bay, Benicia, and Pinole between 1910 and 1970. Very few nests or juveniles have ever been recorded from the Central or South Bay, none in recent years and breeding status there is uncertain. Records compiled by S. Rottenborn from Santa Clara Valley Audubon Society dating back to 1927 suggest that black rails have been very rare to non-existent outside the winter months in the South Bay during this period. A recent late-season record of a black rail calling was on April 26-27, 1993 at the Palo Alto Baylands. Evens et al. (1991) also report territorial calling at the Dumbarton Bridge in April 1989.

Migration Ecology - Migration is commonly believed to be an autumn (August through October) dispersal, probably comprised mostly of birds of the year (juveniles). Extralimital records support this view. Grinnell and Wytte (1927) noted that this species was a "fairly common fall and winter visitant" in the San Francisco Bay Area. Although black rails winter in the United States, their migration pattern in California is not clear, and the extent of their winter dispersal is not known (Ehrlich et al. 1992). Based on the known distribution within San Francisco Bay, it seems likely that the post-breeding season dispersal from North Bay marshes accounts for winter numbers in the South Bay.

Although there are numerous non-breeding season (August-March) records of black rails in these Central and South Bay marshes, their numbers are apparently not sustained through the spring (Evens et al. 1991). Very little is known, however, about spring movement. Reasons for the breeding season absence in the South Bay may include several interrelated factors, including lower elevation of marshes, less peripheral high marsh or transitional habitat, and increased predation rates (Manolis 1978, Evens et al. 1991).

The recent discovery of numerous populations in freshwater marshes and seepages in the Sierra Foothills (Aigner et al. 1995; J. Tecklin, pers. comm.) indicates that the species has the ability to colonize isolated habitat patches. The Rallidae in general disperse themselves effectively and the colonization of several disparate and isolated sites in California in recent years, some of which are newly created (Evens et al. 1991, Aigner et al. 1995, Nur et al. 1997), points to the importance of any marsh habitat for black rails.

Food and Feeding - This species feeds by ground gleaning on terrestrial insects, aquatic invertebrates, and perhaps seeds. Sampling of rail habitats in North Bay marshes indicate Arachnida and Amphipoda as likely prey items, although these findings are largely speculative (Evens, pers. obs.).

Distribution and Abundance

Historically, the black rail occurred from Central California south to San Diego and Baja. Several breeding season surveys of the bird's distribution in California provide current information on the abundance, distribution, and habitat choice of this species in the San Francisco Bay (Manolis 1978, Evens et al. 1991, Nur et al. 1997). Eddleman et al. (1994) stated that in "coastal California during the breeding season, the California black rail is presently found at Bodega Bay, Tomales Bay, Bolinas Lagoon, San Francisco Estuary, and Morro Bay." Although once more widely distributed, the bulk of the population is now restricted to the tidal marshlands of the northern reaches of the San Francisco Estuary (San Pablo Bay) ... at relatively few sites" (Evens et al. 1991).
Figure 7.4 shows the approximate current distribution and relative abundance of black rails in the San Francisco Bay region (Evens et al. 1991). Evens et al. (1991) described the species as rare throughout most of its California range, except in those areas noted by Eddleman, where it is fairly common. Currently, the coastal Southern California breeding population is extirpated.

In his spring and summer surveys, Manolis (1978) found no birds in Central and South Bay marshes, but did find them in San Pablo Bay and Suisun marshes. The extensive breeding season survey by Evens et al. (1991) in San Francisco Bay marshes confirmed distributional patterns found by Manolis. The survey also found numbers concentrated in the northern reaches; birds were largely missing from the Central Bay (except Corte Madera marsh) and were very rare in the South Bay.

Areas of highest concentration are "Petaluma River Wildlife Management Area, along Black John and Fagan sloughs and Coon Island in Napa Marsh, and in some bayshore marshes of San Pablo Bay" (Evens et al. 1991, SFEP 1992). "In these northern reaches rail numbers were much higher in tidal marshes than in marshes with restricted tidal flow, generally higher in marshes along large tributaries than in smaller tributaries or along the bayshore; much higher in bayshore marshes located at the mouths of creeks, rivers, or sloughs than in bayshore marshes not bisected by water courses" (Evens et al. 1991). Nesting birds are also patchily distributed in Suisun Bay marshes and the Delta (SFEP 1992). Recent field work suggests that black rails are more widely distributed in Suisun Marsh than previously understood, particularly in the northern most undiked marshes (B. Grewell, pers. comm.).

A survey of the entire Estuary, conducted from 1986-1988, detected 608 rails at 1,168 stations (Evens et al. 1991). All but two rail detections occurred in the northern reaches of the Bay, including San Pablo Bay, the Carquinez Strait, Suisun Bay, and the Delta. This survey detected rails at 87% of the San Pablo Bay stations, 20% of the Carquinez Strait and Suisun Bay stations, 5% of the Delta stations, <1% of the South Bay stations, and none of the Central Bay stations. Evens (pers. obs.) notes that the population at the Corte Madera Ecological Reserve is believed to now be extirpated.

A follow-up study in 1996 of those parcels surveyed in 1986-1988 (Nur et al. 1997) found stable populations in San Pablo Bay and Suisun Bay and detected no black rails in the Central and South bays. Although there was no overall trend for decrease in the San Francisco Bay population, in the outer coast marshes (smaller and more isolated than the Bay marshes), numbers of black rails were low and appeared to have decreased over the past decade and are considered in danger of extirpation. While this study found that individual marsh size was
not significantly related to overall black rail density, it
did find a significant tendency for black rails to be ab-
sent from small marshes.

There are many records of adults and juvenile
black rails in Central and South bays during the non-
breeding season, but no breeding is known to occur in
these areas (SFEP 1992). The lack of high tide refugia
for birds and low marsh elevation in the Central and
South Bay may explain why breeding populations are not
found there.

**Habitat Use and Behavior**

This species prefers Salicornia dominated marsh habi-
tat (Cogswell and Christman 1977). It is also known to
occur in fresh, brackish, and salt marshes (Erlich et al.
1988). In their survey of the San Francisco Bay during
breeding seasons from 1986-1988, Evens et al. (1991)
found the birds occurred almost exclusively in marsh-
lands with unrestricted tidal influence. This study found
very few birds associated with diked, impounded, or par-
tially tidal marshes. Moreover, Evens and his colleagues
found there.

Peripheral vegetation at and above mean high
higher water is necessary to protect the birds during
periods of extremely high tides. They may be able to use
a range of cover species; at Corte Madera they used fen-
nel growing at the edge of the marsh (Evens and Page
1983). The birds are very vulnerable at this time and, if
not hidden, are subject to predation by northern harri-
ers (Cirrus cyaneus), great egrets (Casmerodius albus),
great blue herons (Ardea herodias), and probably many
other predators (Evens and Page 1986). Observations at
Tomales Bay suggest that bird abundances may be de-
pressed by lack of sufficient, quality upland for refuge
during high tides (Evens and Page 1986). Uplands can
also be degraded by having pedestrians too close to the
marsh, which inhibits the escape of birds to the upland
during high tides (Evens and Page 1983).

**Conservation and Management**

This taxon is acknowledged to be declining (Evens et al.
1991, SFEP 1992) and is listed as threatened and as a
‘California Fully-protected Species’ under the Californ-
ia Endangered Species Act, and as a ‘Species of Spe-
cial Concern’ (formerly ‘Category 2 Candidate Species’)
under the Federal Endangered Species Act. Fragmenta-
tion and habitat loss associated with historic and ongo-
ing pressures of agricultural practices, salt production,
and urbanization are identified as the primary causes of
the decline. The remnant tidal marshlands of San Fran-
cisco Bay occupy only 15% or less of their historic area
(Dedrick 1989), yet even in this diminished capacity
comprise 90% of all remaining California tidal marshes
(MacDonald 1977).

Marshlands of San Francisco Bay and the outer
coast still occupied by black rails have been degraded
by the loss of the zone of peripheral halophytes that form a
natural vegetative transition between the marsh and up-
land and provide high tide refugia for rails. Livestock gra-
ing and diking have reduced or eliminated this transition
zone in most of the marshes around the Bay and the outer
coast, resulting in rail susceptibility to heavy predation by
herons, egrets, and raptors during high tides (Evens and
Page 1986). Predation by rats, feral cats, and red foxes are
also likely to contribute to the problem (SFEP 1992).
Other pressures that threaten to alter or degrade San Francisco Bay habitat include continued diversion of freshwater inflow from the North Bay, a progressive rise in sea level, and contamination by toxic agents shown to have had adverse biological effects on other birds in the Estuary (Evens et al. 1991). The patchy distribution of black rails makes the population susceptible also to metapopulation dynamics and stochastic variables (Evens et al. 1991, Nur et al. 1997).

Recommendations

Increases in black rail populations will require the protection of existing habitat and the restoration of good quality rail breeding habitat. This habitat should be undiked (fully-tidal) salt marsh, with dense stands of pickleweed, and other halophytes characteristic of the salt marsh community. Upland refugium that provides cover during highest tides is critical. Formerly diked marshes that are restored to tidal influence may provide additional habitat for black rails if they encompass elevations at or above mean high water, and are adjacent to extensive tidal marshes with full tidal influence, and include high tide refuge.

Control of non-native predators is also expected to benefit the rail. The largest, most resilient rail populations may not occur in newly established marshes, but may require the development of a mature salt marsh community. The age of the marsh necessary to support black rails is unknown. To ensure the continued viability of this taxon, it is critical that, as marshes are restored, current rail habitat be protected, and transitional vegetation be allowed to become established around the perimeter of existing habitat. It may be many years before restored marshes offer ideal habitat for black rails.

Rail habitat shares many similar features with salt marsh harvest mouse (Reithrodontomys raviventris) habitat, although the rail occupies a narrower band within the elevational marsh gradient. Whatever habitat improvements benefit the rail are likely to benefit the mouse, and vice versa.

References


Personal Communications

Jerry Tecklin, Sierra Foothill Research and Extension Center, University of California, Davis.

Brenda Grewell, Department of Environmental Science and Policy, University of California, Davis.
Common Moorhen
Gallinula chloropus
William G. Bousman

Introduction

A member of the rail family (Rallidae), approximately 12 subspecies of common moorhen are found worldwide, ranging through Europe, Asia, Africa, North America, and numerous islands in the Pacific and Indian oceans. The North American subspecies, G. c. cachinnans, breeds widely in North America and winters south to Mexico (AOU 1957).

In general, the common moorhen requires open fresh water with plant cover. An omnivorous forager, it consumes varying proportions of plant and animal materials. It feeds while swimming or while walking on land or floating plants (Cramp 1980).

A nest is normally constructed in vegetation above water. The typical clutch size is six to seven eggs; pairs are often double-brooded and experienced birds may even raise three broods in a season (Cramp 1980). Birds defend breeding territories and, where resident, reduced winter territories. Generally, moorhens are monogamous and the pair-bond is normally for just a single year. However, an extended bond may occur for resident birds (Cramp 1980).

Distribution and Abundance

Modern Distribution – McCaskie et al. (1979) reported moorhens to be uncommon to fairly common in the interior (Central Valley) of Northern California and rare to very uncommon along the coast. Moor birds were reported to occur along the coast in the winter, but only stragglers were found along the northern coast. McCaskie et al. (1979) noted the primary habitats used were lakes and marshes. Garrett and Dunn (1981) reported that in Southern California, moorhens are uncommon to fairly common along the Salton Sea and in the Colorado River Valley, but occur primarily as a winter visitor along the coastal slope. They noted that the moorhen was formerly more common along the coast, but it now breeds there only rarely and is largely extirpated because of the destruction of freshwater marshes.

In the last 25 years, comparable numbers have been found on the Benicia, Hayward-Fremont, and San Jose Christmas Bird Counts (CBCs), as shown in Figure 7.5. Elsewhere in the Bay, this species is at best a straggler. Aggregate numbers, based on the means of the nine CBCs shown in Figure 7.5, are 48, and this represents a lower bound on the winter population. The Benicia CBC has been showing a long-term decline in numbers (-10.1%, p<0.005), while San Jose CBC has shown an increase (+5.8%, p<0.05). The Palo Alto CBC, on the other hand, reported few birds until about 1980 when the Mountain View Forebay was constructed. Currently, within the Palo Alto count circle, equal numbers of birds are found on the CBC and the Summer Bird Count (SBC), indicating that the species is largely resident. This species is still considered uncommon in the South Bay, although it is regularly found in freshwater areas and is known to breed at the brackish Warm Springs Marsh near Newark. It is much less commonly found on the Marin CBCs, and, in this respect, its distribution appears to be little different than that noted by Grinnell and Miller (1944).

Historical Distribution – In the earlier part of the century, Grinnell and Wythe (1927) noted records from San Francisco and Alameda but did not believe that the species bred within the estuarine system. Grinnell and Miller (1944) considered the common moorhen a summer resident of the Sacramento and San Joaquin valleys and the coastal district southwards from Alameda County. They noted that it did not occur north along the coast beyond San Francisco, and some birds were reported to remain through the winter. Within its preferred habitat of freshwater marsh, they considered it to be locally common. They noted a reduction in numbers in direct proportion to the reduction in available habitat, but pointed out that the species does take advantage of habitat created by irrigation, and this has offset the destruction of marshland to some degree.

Sibley (1952) considered this species to be an uncommon summer resident and noted records from W. E. Unglish of eggs taken from “Soap Lake.” Sibley’s map shows that his “Soap Lake” is what is presently called San Felipe Lake, just south of the Santa Clara line in San Benito County. Ken Schulz (Audubon Field Notes 11:427) reported a high count of six adults and 12 immature birds at a ‘colony’ near the intersection of Singleton Road and Coyote River [sic] on 4 July 1957. Very small numbers of common moorhens were found throughout the 1970s in various areas along Coyote Creek in the southern Santa Clara Valley, but not in any locations along the Bay. Gill (1977), in his survey of South Bay breeding avifauna, did not include the common moorhen on his
list, suggesting that he failed to find it in any areas he checked, including sites such as Coyote Hills Regional Park.

The historic abundance of the common moorhen is difficult to determine, not only because of the lack of historical data, but also because it appears that the San Francisco Bay estuarine system is near the northern limit of this species’ range. The historical atlas clearly shows that there was more freshwater habitat available 200 years ago than there is today and, in the South Bay, there were a number of freshwater marshes associated with willow thickets and natural artesian systems. However, there are no records of moorhens having used these marshes, and it may be that this species is a recent colonizer of man-made freshwater habitats along the Bay edge.

**Habitat Use and Behavior**

The common moorhen appears to be tied closely to freshwater impoundments within the San Francisco estuarine system and, less often, along freshwater streams. These freshwater areas are usually dominated by cattails. For the most part, these freshwater areas are constructed for other uses, and the moorhen invades once the vegetation becomes suitable for foraging and breeding.

The species breeds in small numbers in Marin County and was found in two atlas blocks near Novato during that county’s breeding bird atlas. There are prior breeding records from Olema Marsh in 1967 as well and recent records from the Bolinas sewage ponds and Novato (Shuford 1993). It nests, as well, on Joice Island (C. Wilcox, pers. comm.) in the Suisun Marsh – this site being more typical of natural sites found in the Sacramento Delta than the highly modified man-made sites which this species uses along the Bay. The common moorhen also breeds in small numbers in Santa Clara County, but it is found more regularly there than in the North Bay. It nests every year in freshwater areas alongside the bay, such as the Mountain View Forebay and the Moffett channel at the Sunnyvale Water Pollution Control Plant. It also uses a number of the freshwater percolation ponds on the upper reaches of the Guadalupe River and, apparently, portions of the river itself. In wetter years, it extends its range to suitable percolation ponds with sufficient growth of cattails and tules, such as the small ponds at Coyote. Draining and maintenance

**Figure 7.5** Christmas Bird Count data for Common Moorhen — Relative geographic location of Count circles indicated by dashed lines

National Audubon Society’s Audubon Field Notes and its successor publications, American Birds, and Field Notes, Volumes 24-51.
activities along Coyote Creek during the atlas period prevented breeding in a number of areas where the species nested previously (and may still do so in wetter years).

**Conservation and Management**

The common moorhen, perhaps more than any other single species, is directly tied to freshwater marshes with good cover of cattails. This species is an active prospector for new territories (Cramp 1980) and, in this limited sense, is adaptable to change. Examples of this adaptability include the development of a new breeding population following construction of the Mountain View Forebay in the late 1970s. As cattails started to grow in this freshwater area, birds moved in and breeding was detected by 1983. Peak population counts in this area include 20 birds counted by David Suddjian on 24 November 1985 (pers. comm.) and 20 there as well reported by Peter Metropulos on 13 August 1988 (pers. comm.). Similarly, a suitable freshwater marsh developed at Moffett Field in the early 1980s due to cooling water runoff and a blocked drain, and by 1983 at least three pairs were nesting in this area. With the repair of the drain and a different treatment of the cooling water effluent, this area is no longer suitable for common moorhen and they are not present.

The moorhen's inflexibility in habitat needs is balanced by its adaptability in finding new habitats when and where they occur. In this sense, this species can respond successfully to carefully designed freshwater management programs. At the present time, within the bounds of the San Francisco estuary, almost every location used by this species has been constructed to serve the needs of local communities in one way or another. As a consequence, these habitats are not designed to benefit the common moorhen and, in almost none of these situations, do the agencies that oversee these habitats include within their management plans an oversight and stewardship responsibility for the moorhen. This lack of responsibility by local agencies is regrettable and in most cases where their actions discourage or destroy this species' local populations, alternate approaches can be envisioned that would enhance the local population without significantly impacting local agency goals. There are numerous opportunities, both through education and legal mandates, to improve the management of freshwater habitats in the San Francisco Bay Area to benefit this species.

**References**


Sibley, C.G. 1952. The birds of the South San Francisco Bay region (mimeo).

**Personal Communications**


David Suddjian, Biological Consulting Services, Capitola, California. Dsuddjian@aol.com.

Carl Wilcox, California Department of Fish and Game, Yountville.
California Gull
Larus californicus
Thomas P. Ryan

Introduction
California gulls are a recent addition to the breeding avifauna of San Francisco Bay, drawn here by the availability of remote nesting grounds and rich food sources provided by the salt ponds and local refuse dumps. They have steadily increased in number since their first breeding attempt in 1980. Associated species which use similar habitats are Forster’s tern (Sterna forsteri), Caspian tern (Sterna caspia), black-necked stilt (Himantopus mexicanus), American avocet (Recurvirostra americana), killdeer (Charadrius vociferus), and Wilson’s phalarope (Phalaropus tricolor).

Description
California gulls attain adult plumage in their fourth year. The adult California gull has a white head, chest, and underparts. The mantle (upperwings) is grey, their mantle is darker than similar appearing ring-billed gulls and herring gulls (Garrett 1985). Their wing-tips are black, with white mirrors. In breeding plumage their legs are bright yellow-green, this fades during the winter. Their bill is yellow with black and red subterminal spots (gonys). Juvenile birds are very dark with mottled underparts, pale undertail coverts, and pink legs (Garrett 1985). First year birds are a lighter grey-brown mottled with white, and have dull pink legs. The rump is barred. Second and third year birds have increasingly white underparts and more grey on the mantle, and the bill may have a black ring around it (Garrett 1985). Adult California gulls can be told from western gulls, herring gulls, and glaucous-wing gulls by their smaller size and yellow-green leg color. They can be distinguished from ring-billed gulls by a darker mantle, yellow-green leg color, and black and red subterminal spots.

Breeding Biology - California gulls begin to return to their breeding colonies three to seven weeks prior to breeding (Winkler 1996). The nest is a scrape in the substrate with bones, feathers, and vegetation placed within the cup (Winkler 1996). Nests are sometimes used in subsequent breeding seasons (Winkler 1996). Two to three, and occasionally four eggs are laid at daily intervals approximately one week after nest building begins (Winkler 1996, Baicich and Harrison 1997). Larger clutches (four or more) may be the result of female-female pairs. Eggs are laid in early to mid-May. Eggs at a given colony are laid within 20 days of each other, later clutches are often second attempts (Vermeer 1970, Winkler 1983, Jones 1986). The incubation period lasts 23-27 days (Winkler 1996); Jones (1986) reported a mean of 26.6 days at South Bay colonies. Both parents incubate the eggs (Winkler 1996). Chicks hatch at South Bay colonies in late May to early June. Chicks stay in or near the nest for the first week, from nine to 20 days old they have been found to run as far as seven meters from the nest for cover (Winkler 1996). Most chicks abandon the nest area entirely at 40-60 days, and fledge by 48 days (Winkler 1996). There is little association between adults and juveniles after fledging (Winkler 1996). California gulls are long lived with high survivorship, banded adults of 27 and 30 years of age have been reported (Winkler 1996).

Migration Ecology - California gulls appear to move continuously throughout the fall, winter and spring (Winkler 1996). Younger birds are found farther south in the fall and farther north in the spring. Individuals on the West Coast move farther north after breeding, and are driven farther south as winter storms increase in number and intensity.

Wintering Ecology - California gulls winter from central California, south to Guerrero, Mexico (AOU 1983). In California, concentrations are found in the Central Valley and along the coast. Local populations are found as far north as British Columbia. Birds banded as chicks at San Francisco Bay colonies have been observed at Pismo Beach in San Luis County and at Doheney State Beach and Seal Beach in Orange County.

Distribution and Abundance
In North America, California gulls breed at inland lakes from the Great Basin northward to the Northwest Territories (Canada) and southward to Utah, east to the Dakotas. They winter in smaller numbers in Utah, in Nevada through the Colorado River Valley, and at the Salton Sea in California.

On the Pacific Coast, California gulls breed at inland lakes east of the Sierra-Cascade axis from British Columbia through central California and in San Francisco Bay. The largest breeding population in California is at Mono Lake, Mono County (Winkler 1996).
California gulls winter along the Pacific Coast from British Columbia to Mexico. They are also found in the Central Valley and at the Salton Sea in California. Small numbers are found in the Columbia River Valley of Washington and Oregon and Willamette Valley in Oregon (Winkler 1996).

Approximately 5,000 California gulls nested at six sites in southern San Francisco Bay in 1997 (Table 7.3). They breed in the hundreds at Alameda Naval Air Station; this colony has been active since 1982 (L. Collins, pers. comm.). There are no known active colonies in the North Bay. They roost in large numbers during the winter in mixed flocks on the levees and in the salt ponds in the South Bay.

**Population Trends** - In 1982, Conover (1983) estimated the United States population at 276,000 California gulls. The worldwide population of this species is likely between 500,000 and one million individuals (Winkler 1996). Winkler (1996) notes apparent declines across prairies south and east from Alberta. However, he notes that patchy distribution of this species makes it difficult to determine any actual trend. The overall population appears to be increasing since the turn of the century, although there is debate about the rate of increase (Winkler 1996). This increase has been attributed to an increase in farmlands, increased availability of nesting sites at reservoirs, decreased harvesting of feathers and eggs, and increased winter food availability, especially at refuse dumps (Conover 1983).

California gulls first nested in the San Francisco Estuary in 1980 when 12 nests were encountered at Pond A6, near Alviso in Santa Clara County. We believe this was the first time California gulls nested in a coastal region. This colony grew each year, peaking in 1994 with 4,363 nests. Between 1994 and 1997, nest counts at Pond A6 decreased 26% to 3,241 nests. Beginning in 1984, California gulls began breeding at other sites within the South Bay. At Newark, 22 nests were found in 1984, increasing to 277 in 1985; this site was abandoned in 1989. In 1984, 33 nests were found at Pond A9 near Alviso, 2.2 km from Pond A6. Low numbers of California gulls nested here from 1984 to 1990, except in 1985 when 187 nests were found. In 1991, the colony was abandoned. It was re-established in 1992, but failed in 1994 and 1995. However, in 1996 and 1997 the colony increased to 702 and then to 878 nests. In 1990, a colony established on Pond A1. This colony grew to 86 nests in 1994, and currently has an estimated 40 nests. In 1992, a colony was established near Mowry Slough in Alameda County, 3.5 km from Pond A6. This site has been characterized by year-to-year fluctuations in nest numbers. In 1996, this colony moved to Pond M 1/2, 1 km west of the original site. Red fox had been found at Pond M 4/5 prior to this move. In 1993, California gulls nested on a rafted levee and a series of small dredge spoil islands at Pond B2 near Mountain View, Santa Clara County, 2.4 km from Pond A6. Initially, colony size fluctuated, but increased from 1995 to 1997.

Currently approximately 10,000 California gulls nest in South Bay. California gulls are abundant in the San Francisco Bay in the winter, although no reliable estimates of wintering numbers exist (Harvey et al. 1992).

**Habitat Use and Behavior**

California gulls are opportunistic feeders, and their diet varies greatly at different locations. In 1983-84, Jones (1986) found the diet at colonies in the South Bay included arthropods (brine fly larvae, brine shrimp, insects, etc), garbage, and fish. California gulls are often seen foraging at garbage dumps, and at the edge of salt ponds on rafts of brine fly larvae and brine flies. They are also observed feeding in fields and schoolyards, presumably on insects and human refuse.

California gulls roost in mixed species flocks on salt pond levees, salt ponds, and schoolyards. There is a large daily movement from the local refuse dumps to the roosting areas on the levees and salt ponds.

**Conservation and Management**

California gulls are documented to abandon colonies following predation events, and observations of feral cats and red fox in close proximity to their nesting grounds. There have been partial and complete nesting failures at all sites that are attached or become land-bridged to the mainland early in the nesting season.

California gulls in the South Bay require remote, insular, abandoned levees and abandoned islands for nesting. The continued presence of such levees and islands will provide them with adequate nesting grounds. They require high saline salt ponds for their primary natural food source, brine fly larvae, brine flies, and brine shrimp. These are also food sources for many other species that use salt ponds.

The largest colony of California gulls is at the original point of colonization, Pond A6, where in recent years 3,000 to 4,600 pairs have nested annually. These levees

### Table 7.3 California Gull Breeding Sites in the South Bay

<table>
<thead>
<tr>
<th>Colony Site</th>
<th>Status</th>
<th>1997 Nest Count</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alameda NAS</td>
<td>Active</td>
<td>100+</td>
</tr>
<tr>
<td>Pond A1</td>
<td>Active</td>
<td>22</td>
</tr>
<tr>
<td>Pond B2</td>
<td>Active</td>
<td>328</td>
</tr>
<tr>
<td>Pond A6</td>
<td>Active</td>
<td>3,128</td>
</tr>
<tr>
<td>Pond A9/A10 levee</td>
<td>Active</td>
<td>878</td>
</tr>
<tr>
<td>Mowry M1/M2 levee</td>
<td>Active</td>
<td>620</td>
</tr>
<tr>
<td>Newark</td>
<td>Historic</td>
<td>0</td>
</tr>
</tbody>
</table>
should be maintained as nesting habitat for this species. If this colony is flooded as has been suggested, approximately 7,000 adult, breeding California gulls will be displaced. This could also negatively impact the tern populations in the South Bay. It is unknown exactly what the result would be of a major disruption of this colony. The possible consequences should be studied in greater depth before such an action is undertaken.

California gulls provide us with a robust subject to study the reproductive biology of a salt pond associated species. We are studying the effects of predation on species nesting on insular versus non-insular levees. We are studying the re-colonization of an area by a colonially nesting species after major predation events. At Pond B2, nesting occurs on a series of different sized islands, providing a natural experiment for studying the effects of island size and colony size on the reproductive success of larids.

In the 1980s, thousands of chicks were banded by San Francisco Bay Bird Observatory. Currently, 20-25% of the adults at these colonies carry U.S. Fish and Wildlife Service bands, and 5-10% carry color bands identifying them to cohort. As these birds are long lived, they provide an excellent subject for studies of survival, population dynamics, fall and winter movements and ecology, and eventually senescence, in addition to other banding related studies.

Acknowledgments

I would like to thank the Peg Woodin, Mike Rigney, and the late Richard Mewaldt for their meticulous efforts in studying the South Bay colonies in their early years; Laura Collins for her information regarding the Alameda Naval Air Station colony; and the many dedicated volunteer observers of the San Francisco Bay Bird Observatory.

References


Personal Communications

Laura Collins, biologist, Berkeley, California.

Forster's Tern

Sterna forsteri

Thomas P. Ryan

Introduction

The Forster's tern is a mid-size tern found in open water, salt pond, marsh, and estuarine habitats within the San Francisco Estuary. It nests and roosts on dredge spoil islands and degraded, insular levees. Forster's tern forage in salt ponds, open bay, slough channels and marshes. Associated species which use similar habitats are California gull (Larus californicus), Bonaparte's gull (Larus philadelphia), Caspian tern (Sterna caspia), American avocet (Recurvirostra americana), black-necked stilt (Himantopus mexicanus), and killdeer (Charadrius vociferus).

Photo by Ed Talfl
**Description**

Forster's terns are a medium-sized tern. In breeding plumage, they have an all white body, a dark cap, black eyes, grey upperwings, orange bill with a black tip, and orange legs. Forster's terns are distinguished from other terns by size, a black cap and nape, deeply forked grey tail with white outer feathers, and uniformly light-colored upperwings (Peterson 1990). In August, they begin to molt into winter plumage: the black cap becomes a mask, with black coloration around and behind the eye. The crown becomes white, the nape varies from grey to black, and the bill turns from a bright orange color to a uniform black (Peterson 1990).

Forster's terns occur in freshwater and salt marshes, seacoast, estuaries, and inland rivers and lakes (AOU 1983). They feed on small fish and arthropods. The call is a harsh, nasal “za-a-ap” and a nasal “kyarr” (Peterson 1990).

**Breeding Biology** - Forster's terns nest alone and in colonies ranging in size between two and 600 birds. Their breeding season spans from April until August. They usually begin breeding in their second to third year. First-year birds remain on their wintering grounds year-round. During courtship, the males pass fish to the females. The pair will remain together for the duration of the breeding season (Bent 1921). The nests are usually scraped made in the dirt, with vegetation, small stones, sticks, and bones placed in the cup (Baicich and Harrison 1997, SFBBO unpubl. data). They are placed both in vegetation and on bare dirt. Birds at the South Bay colonies lay two to three eggs from mid-May to mid-June. Both parents incubate the eggs; incubation lasts 23 to 25 days (Baicich and Harrison 1997). Chicks begin to appear in mid-June. The young hatch in a semi-precoital state, and fledge at approximately 28 days (Baicich and Harrison 1997). Most young have fledged by the end of August.

**Migration Ecology** - During September and November, most Forster's terns begin their southward migration to spend the winter in locations from central California to Mexico, and possibly as far south as Costa Rica (Gill and Mewaldt 1979, AOU 1983). Of 2,943 Forster's tern young banded by Gill and Mewaldt (1979) at South Bay colonies, five were recovered during migration in the Los Angeles-San Diego area, and the sixth was recovered in Sinaloa, Mexico. It appears that their migration route follows the coastline of California; whether they cross Baja into the Sea of Cortez or move around the peninsula is unknown.

**Wintering Ecology** - It is unknown exactly where the San Francisco Bay population of Forster's terns spends the winter. Circumstantially, Forster's terns are common along the coast of western Mexico during the winter months (Arbib 1974, 1975, 1976). These sightings and the six band recoveries noted above led Gill and Mewaldt (1979) to state that, "nearly all Forster's terns leave the San Francisco Bay area each winter and that most juveniles and adults winter from coastal southern California well down the west coast of Mexico." Their habits and ecology are poorly known on their wintering grounds.

**Distribution and Abundance**

In North America, Forster's terns breed in the interior of the continent from central British Columbia to central Ontario, and south from Oregon to northwest Indiana. Along the East Coast, they breed from southern New York to South Carolina, and along the Gulf Coast from Tamaulipas, Mexico to Alabama. They winter from the Virginia coast to Florida and western Texas south to Mexico, casual to Costa Rica (AOU 1983).

On the Pacific Coast, breeding occurs from the coast of British Columbia south to Baja California. They winter from central California south to Oaxaca and Guatemala, casual to Costa Rica (AOU 1983).

In the North Bay, nesting occurs in the Napa River Marsh salt ponds, but there are no recent population summaries available for all colonies (Carter et al. 1990). Colonies were documented during the Napa County Breeding Bird Atlas Project at Russ Island (1989), Island #2 at the Can Club Duck Club (1989, 1991), Little Isle (1991), Knight Island (1990), and White Slough (1987, 1988, 1989, 1992, 1996) (R. Leong, pers. comm.) (Table 7.4). From counts done at these sites in various years, it appears that the total number of nests at these colonies is in the low hundreds in any given year. These colonies should be surveyed more completely to determine their actual size.

In 1997, Forster's terns bred at 21 colonies in the South Bay; two colonies were not surveyed. Since 1992, they have bred at 28 sites. These sites extend from Belmont Slough south to Alviso on the western side of the Bay, and from Baumberg south to Alviso on the eastern side (Table 7.5). They breed on dredge spoil islands, and degraded levees in current or former salt ponds, slough channels (Belmont), and diked marshes (Ravenswood).

Small numbers are found locally throughout the winter (W. Bousman, CBC data). Numbers increase during the spring, as migrants begin to arrive in April.

<table>
<thead>
<tr>
<th>Colony Site</th>
<th>Status</th>
</tr>
</thead>
<tbody>
<tr>
<td>Coburn</td>
<td>Unknown</td>
</tr>
<tr>
<td>Island #2</td>
<td>Unknown</td>
</tr>
<tr>
<td>Knight Island</td>
<td>Unknown</td>
</tr>
<tr>
<td>Little Isle</td>
<td>Unknown</td>
</tr>
<tr>
<td>Russ Island</td>
<td>Unknown</td>
</tr>
<tr>
<td>White Slough</td>
<td>Unknown</td>
</tr>
</tbody>
</table>

**Table 7.4** Forster's Tern Breeding Sites in the North Bay
Other Birds of the Baylands Ecosystem  

Chapter 7 — Other Birds of the Baylands Ecosystem

Other Birds (Harvey et al. 1992). Migrants and local breeders are found here until late October through November (Harvey et al. 1992).

**Population Trends** - Currently there are no estimates available for either the total North American population or the total Pacific Coast population. The only population estimates available from other coastal colonies are from the Bolsa Chica Ecological Reserve, where a colony established in the late 1980s and has increased to over 200 pairs (C. Collins, pers. comm.).

Forster’s terns first bred in the San Francisco Estuary in 1948, when roughly 100 nests were found near the eastern end of the San Mateo Bridge (Sibley 1952). Gill (1977) found 935 nesting pairs at six South Bay colonies in 1971; this would be 1,200 nesting pairs if colonies “found outside the study area” were to be included. In 1972, 10 colonies were present, containing an estimated 2,000 pairs (Mansfield in Gill 1977). Rigney and Rigney (1981) estimated 2,500 pairs at six sites in 1981. Woodin (1988) summarized San Francisco Bay Bird Observatory data from 1984 to 1988 and reported that counts ranged from 2,183 adults present in 1987 to 3,610 in 1984. In 1988-89, 3,550 breeding birds were estimated at 21 colonies for all Bay Area colonies (Carter et al. 1990). From 1992 to 1997, between 1,842 and 2,365 individuals were observed, and between 1,012 and 1,754 nests were counted at peak breeding season counts at all South Bay colonies. Data collected in 1993 were not used in these figures as surveys were not as thorough as in other years. A decrease from 1,754 nests to 1,012 nests was seen between 1992 and 1994, a 42% decline. The number of nests increased to 1,362 in 1997.

Historically, the largest nesting site was at Pond B2 near Mountain View, with approximately 600 nests. The second largest site was at the Baumberg salt pond islands and the former Knapp Property, near Alviso (Rigney and Rigney 1981, Harvey et al. 1992). In 1997, the largest colony was at Turk Island (300 nests), followed by Hayward Shoreline (226 nests), and Pond B2 (127 nests).

California gulls have colonized both B2 and the Knapp property. At B2, California gulls have expanded from zero to 328 pairs from 1992 to 1997. At the Knapp property, California gulls increased in number from 2,750 pairs to 3,241 pairs between 1992 and 1997. Additionally, since 1992, red fox have been noted preying Forster’s tern colonies in both areas.

Forster’s tern populations steadily decreased between 1971 and 1997 (the time of this writing). Population estimates place the number of Forster’s terns between 1,870 and 5,000 individuals (x=3,623; SD=1,599) between 1971 and 1981; 2,183 and 3,610 (x=2,707; SD=536) between 1984 and 1988; and 1,842 and 2,365 (x=2,137; SD=202.9) between 1992 and 1997. Although the population of Forster’s terns in San Francisco Bay decreased across these periods, the large year-to-year fluctuations in the local breeding population make it difficult to determine if any significant trend exists.

**Habitat Use and Behavior**

Forster’s terns forage on the open bay, slough channels, freshwater and salt marshes, and on salt ponds. During the breeding season, they are generally observed foraging singly and in groups. They are occasionally observed foraging with pelicans, egrets, gulls, and other terns in large foraging flocks on certain salt ponds, particularly in the late summer and fall. Forster’s terns roost prior to, during, and after breeding on the dredge-spoil islands and levees. They also have been observed on docks,
Archimedes' screws, duck blinds, pilings, algal mats, floating debris, and other suitable structures surrounded by water. During the breeding season Forster's terns move singly and in groups between their nesting sites within the salt ponds and the aforementioned foraging areas throughout the day. Nocturnal movements, if any, are unknown.

**Conservation and Management**

In the San Francisco Bay Area, feral red fox (*Vulpes vulpes*) and domestic cat (*Felis domesticus*) are known predators of active Forster's tern colonies (SFBBBO unpubl. data). Predation by great egrets and other ardeids is likely; other possible avian predators include raven, peregrine falcon, northern harrier, and red-tailed hawk.

It is unknown what effects, if any, contaminants have on Forster's terns in the San Francisco Estuary. Ohlendorf et al. (1988) found measurable levels of mercury, DDE, organochlorines, and PCBs in the eggs of Forster's terns at Bair Island in 1982.

In a few cases, colonies have been abandoned in association with off season levee maintenance and fluctuating water levels within ponds. High water levels can flood islands. Low water levels can land-bridge islands, making them more susceptible to mammalian predation. Most colonies are in areas of restricted human access, and at these colonies, disturbance does not appear to be a significant problem.

The continued presence of isolated, insular islands is crucial to the continued presence of Forster's terns in the San Francisco Estuary. In all cases, colonies are found within or in close proximity to former and current salt ponds. This habitat provides suitable nesting substrate isolated from human disturbance and makes access more difficult for mammalian predators. We recommend the continuation of programs to control mammalian predators, as this will likely decrease the impact of predation on these colonies.

A schedule of levee maintenance that will minimize disturbance to breeding colonies should be implemented. We recommend initiation of a management program that will: 1) minimize work near colonies during the breeding season (April 1 to September 1); 2) maintain islands by placing fresh dredge material on colonies after the breeding season is over (September 1st); 3) minimize the impact to the population in a given area by disturbing as few colonies within a given pond system as possible within a given year; and 4) construct more islands within salt ponds when possible.

The status of North Bay colonies is unknown due to a lack of available surveys, but it is likely that these colonies are still active. It should be noted that when South Bay Caspian terns were predated and disturbed in the South Bay, they shifted their nesting activities to the Central Bay and North Bay. These North Bay colonies could increase in importance in the event of a natural or man-made disturbance at the South Bay colonies.

Our recommendations are to maintain or increase local nesting and foraging habitats. These areas include salt ponds, levees and islands associated with salt ponds, lagoons, shallow bay/strait, and marsh.

**Research Needs** - The reasons behind the decline of the South Bay Forster's tern population requires further study. Possible causes of this decline include encroachment by California gulls, predation by red fox and feral cats, disturbance by levee maintenance, and fluctuating water levels within salt ponds. Continued population monitoring is needed to document future population trends. Studies of reproductive success and fledgling success would be valuable in documenting problems with the year-to-year reproductive effort. Combining these studies with studies of predation, the presence and effects of contaminants would be valuable in detecting causes of the current decline.

Studies of their natural history and ecology on their wintering grounds in southern California and the west coast of Mexico are needed as well. The general decline in the total number of individuals returning and year-to-year fluctuations in the total numbers of individuals present may indicate problems elsewhere.

**References**


Caspian Tern
Sterna caspia
Thomas P. Ryan

Introduction

This large tern forages on the open bay, salt ponds, marshes, freshwater ponds, rivers, reservoirs, and in the open ocean near San Francisco Bay. They nest on sandy beaches and on salt pond levees and islands. The continental and Pacific Coast populations have both increased. However, declines have been seen at southern San Francisco Bay breeding colonies in recent years. Associated species include Forster’s tern (Sterna forsteri), least tern (Sterna antillarum), and California gull (Larus californicus).

Description

The Caspian tern is the largest of the North American terns. The body and underwings are white, the underside of the tips of the primaries are black, the upperwings are a light silvery-grey, and the crown is black extending below the eye. The tail is shallowly forked. The bill is a deep red to orange-red with black on the tip. The legs are black in adults, and vary between orange, red, and black in the fall immature birds. Immature and winter plumage birds have white streaks in the cap (Peterson 1990).

The Caspian tern is a cosmopolitan species. They occur at lakes, bays, estuaries, marshes, and rivers, on all continents except Antarctica (AOU 1983). They forage by “plunge diving”—the bird hovers over the water before diving to or below the surface to catch their prey. They feed on fish, amphibians, and arthropods. Their call is a hoarse, low “kraa-uh or karr” (Peterson 1990).

Breeding Biology – Caspian terns breed on sandy beaches, dredge spoil levees, and islands. They are colonial, but will nest singularly (Baicich and Harrison 1997). They often nest with or near other larids. The nest is generally a hollow depression in the substrate, with plant and other debris placed in it (Baicich and Harrison 1997). In the San Francisco Bay Area, eggs are laid from May to July. They lay two to four eggs asynchronously at intervals of two to three days (Soikkeli 1973). Chicks hatch in June and July, and are present through August. The chicks are semi-precocial and move about the nesting colony after a few days; they fledge at 25-30 days (Baicich and Harrison 1997), but many stay near the colony as long as it is active.

Migration Ecology – Caspian terns which breed in San Francisco Bay migrate along the Pacific Coast to and from southern California and western Mexico (Gill and Mewaldt 1979).

Wintering Ecology – During the winter months, Caspian terns are found from central California, southward along the western coast of M exico, to northern Colombia. Band recoveries indicate that Caspian terns that breed in the Bay Area winter along the west coast of M exico, going as far south as southern Chiapas (Gill and M ewaldt 1979). Little is known about their winter ecology other than they are found at both coastal and inland sites (Gill and M ewaldt 1979). Christmas Bird Counts indicate that small numbers of Caspian terns winter locally throughout the Bay Area (W. Bousman, pers. comm.).

Distribution and Abundance

In the interior of N orth America, Caspian terns breed east of the Sierra-Cascade axis in the Great Basin from eastern Washington to California, and from Nevada through Utah to northwestern Wyoming (AOU 1983). In the M id-West, they breed in northwestern Alberta, and central Saskatchewan east through the Great Lakes region south to N orth D akota and central M ichigan (AOU 1983). On the East Coast, they breed from Newfound-
Island to South Carolina. They breed along the Gulf Coast from Texas to Florida (AOU 1983). Caspian terns winter from North Carolina south to Venezuela (AOU 1983).

On the Pacific Coast, Caspian terns breed from British Columbia, Canada south to islands off the coast to Sinaloa, Mexico. They winter from coastal central California to northern Colombia (AOU 1983).

The first breeding accounts in the San Francisco Bay Area are from the South Bay, where the majority of Caspian terns nested prior to 1990. Since 1990, the majority of birds nest at colonies in the Central Bay and North Bay. Former colonies in the South Bay exist at Baumberg, Turk Island, Bair Island, and Drawbridge/Mowry. Current colonies exist at Pond A7 near Alviso (104 pairs) and Coyote Hills (30 pairs). Single pairs have recently nested among Forster’s terns at Ravenswood Open Space Preserve, and Hayward Shoreline.

The only North Bay breeding colony exists at Knight Island at the Napa River Marsh. This colony was first detected in 1989 (R. Leong, pers. comm.). In 1977, a small colony was detected at the Can Club, Island #2 (Gill 1977; R. Leong, pers. comm.).

In the Central Bay, colonies are found at Brooks Island and at the Alameda Naval Air Station (NAS).

**Population Trends** - The largest continental population of Caspian terns is found in North America, with a minimum of 35,000 pairs nesting throughout North America in the 1980s and 1990s (Cuthbert and Wires 1999). The North American population has steadily increased since at least the 1960s (Cuthbert and Wires 1999).

Prior to 1970, the largest reported breeding colony of Caspian terns on the Pacific Coast was in San Diego, California (Gill and Mewaldt 1983). In the early 1980s, the largest colony was at Gray’s Harbor in Washington, where there were 2,157 pairs in 1981 (Gill and Mewaldt 1983). This colony was abandoned in the late 1980s, and relocated at nearby Rice Island, Oregon. Currently, the Rice Island colony is believed to be the largest breeding colony, with 8,000+ pairs (Cuthbert and Wires 1999). Other coastal colonies exist or have existed at Gray’s Harbor and Willapa Bay, Washington; Columbia River, Oregon; Humboldt Bay, Elkhorn Slough National Estuarine Research Reserve, Salinas River Mouth, Bolsa Chica Ecological Preserve, Newport Back Bay Ecological Preserve, Salton Sea, and South San Diego Bay, California (Gill and Mewaldt 1983; Cuthbert and Wires 1999); and Laguna Figueroa (Palacios and Alfaro 1992), Scammon’s Lagoon (Bancroft 1927), and Laguna San Ignacio (Danemann 1992), Mexico. Overall, the Pacific Coast population has shown an increase from at least 5,661 pairs estimated by Gill and Mewaldt (1983) to at least 12,263 pairs estimated by Cuthbert and Wires (1999). Most of this increase is accounted for in the Gray’s Harbor/Rice Island colonies where, roughly 2,300 pairs were estimated in the early 1980s (Gill and Mewaldt 1983), and over 8,000 pairs were estimated in 1996-97 (Cuthbert and Wires 1999).

In 1922, Caspian terns were first documented nesting in San Francisco Bay on a levee near the Dumbarton Bridge in the Coyote Hills salt ponds. This small colony of about seven nests increased over the next two decades, with a total of 287 nests in 1931 (DeGroot 1931), and 378 nests in 1943 (Miller 1943). This colony numbered approximately 299 pairs in 1966; however, because of levee maintenance between 1968-69, the colony was abandoned.

Several new colonies formed and disappeared in other regions of the Bay. A colony established itself in 1968 near the town of Drawbridge, adjacent to Mowry Slough, in Alameda County, which contained over 100 nests; 200 nests were counted in 1971 (Gill 1977). In 1968, colonies were also established at Baumberg and Turk Island. The Turk Island colony rose to 540 nests in 1985, but was abandoned in 1986, after levee maintenance in 1985 (SFBBBO 1988). In 1971, a colony was discovered with 304 nests on a salt dike on outer Bair Island. This colony grew to between 500-600 nests by 1975 (Gill 1977) and 800-850 nests by 1981 (Rigney and Rigney 1981).

In the ensuing years, the activity and success at the outer Bair Island colony was erratic. After decreasing to 200 nests in 1983 and being abandoned in 1984 and 1985, the colony was reestablished and included 1,700 adults (850 pairs) by 1988. The colony was again abandoned in 1989 and then re-colonized in 1992 and 1993, but no young were produced. In recent years, both the Turk Island and outer Bair Island colonies have been deserted.

South Bay populations were estimated at 1,000-1,200 individuals in 1971 (Gill 1973) and 2,350 individuals in 1981 (Rigney and Rigney 1981). From 1984 to 1988 South Bay populations ranged from 1,120 to 2,111 adults (Woodin 1988), but in 1990 only 100 pairs were reported (Woodin in Harvey et al. 1992). A Bay Area-wide estimation of 2,818 individuals at five colonies in 1989-90 was made by Carter et al. (1990). In 1997, an estimated 1,400 to 1,500 pairs nested in the Bay Area, although only 136 pairs nested in the South Bay.

Nesting colonies have been growing in the Central Bay at Brooks Island and Alameda Naval Air Station (NAS), concurrent with the colony abandonments in the South Bay. The colony at Brooks Island began in the early 1980s, grew to 400 adults (200 nesting pairs) in 1990 (Carter et al. 1990), and had reached 500 nesting pairs in 1997. Six hundred pairs of Caspian terns were recorded at the Alameda NAS colony in the early 1990s (Harvey et al. 1992), and approximately 350 pairs nested there in 1977.
In addition to the colonies at Brooks Island and Alameda NAS, there are two other smaller colonies: one near Alviso with approximately 104 pairs, and one, a recently reoccupied (1997) historical colony at Coyote Hills, with approximately 30 pairs. The overall population of Caspian terns in the San Francisco Estuary has increased in the past 26 years. However, in part due to predation and levee maintenance, the population has decreased in the South Bay. Table 7.6 shows some of the historic and existing colony locations around the Bay.

The only other nearby colonies are in Monterey County at Elkhorn Slough, which was active until 1995; and the Salinas River Mouth, which was abandoned in 1997 (J. Parkin, pers. comm.).

Habitat Use and Behavior

Caspian terns forage on the open bay, salt ponds, marshes, freshwater ponds, rivers and reservoirs, and at sea (Sibley 1952, SFBBO unpubl. data). They feed on small to medium sized fish, crustaceans, and amphibians. In California, they have been reported to feed on fish and crayfish (Horn et al. 1996; Loeffler 1996; J. Parkin, pers. comm.). Bent (1921) reported that they feed on shrimp, water mussels, and eggs and young of other birds. Ewins et al. (1994) added insects and larid eggshells from Caspian terns in the Great Lakes region.

Caspian terns roost on salt pond levees, sandy beaches, mudflats, and on islands in salt ponds, slough channels, marshes, and in the Bay. During the breeding season, the majority are observed roosting locally, near nesting colonies, although some are seen at local reservoirs (Almaden Lake Park, Los Gatos Creek Park, Calero Reservoir, Lexington Reservoir, Anderson Reservoir, Stevens Creek Reservoir) and along the outer coast. After breeding season, they disperse and roost in association with each other and with other larids at sites throughout the Bay. The post-breeding season aggregations may be augmented by individuals dispersing from colonies elsewhere.

<table>
<thead>
<tr>
<th>Colony</th>
<th>Site Status</th>
</tr>
</thead>
<tbody>
<tr>
<td>Knight Island, Napa River Marsh</td>
<td>Active</td>
</tr>
<tr>
<td>Brooks Island</td>
<td>Active</td>
</tr>
<tr>
<td>Coyote Hills, west levee</td>
<td>Active</td>
</tr>
<tr>
<td>Baumberg</td>
<td>Historic</td>
</tr>
<tr>
<td>Turk Island</td>
<td>Historic</td>
</tr>
<tr>
<td>Drawbridge</td>
<td>Historic</td>
</tr>
<tr>
<td>Bair Island</td>
<td>Historic</td>
</tr>
<tr>
<td>Alviso, Pond A7</td>
<td>Active</td>
</tr>
<tr>
<td>Alviso, Pond A3</td>
<td>Historic</td>
</tr>
<tr>
<td>Hayward Shoreline</td>
<td>Active</td>
</tr>
<tr>
<td>Ravenswood OSP</td>
<td>Active</td>
</tr>
</tbody>
</table>

The majority of their foraging likely occurs in the Bay and surrounding marshes and salt ponds. However, Caspian terns are occasionally seen over urban areas carrying fish. Presumably, they are hunting in local reservoirs and ponds, bringing this prey to their nesting colonies in the South Bay (Cogswell 1977, SFBBO unpubl. data). A trout tag from a lake 16 miles away was found at a nesting colony in San Francisco Bay (Cogswell 1977). Additionally, local observers have noted small numbers of Caspian terns flying, perhaps daily, on north-south routes over the Santa Cruz Mountains to the Pacific Ocean. In 1987, 64 individuals were observed making such crossings, and 88 were observed in 1988 (W. Bousman, pers. comm.). After breeding, some adults and juveniles remain near the nesting areas, while others disperse within the Bay and beyond. Little is known about these post-breeding movements.

Conservation and Management

Caspian terns numbers have declined in recent years in the South Bay, in part due to predation, flooding, routine levee maintenance, and levee erosion. Routine levee maintenance is a threat to South Bay colonies because of the small number of larger colonies, and because of the tern’s tendency to nest on levees rather than on dredge spoil islands. The colonies at Coyote Hills and Turk Island were deserted after levee maintenance. Alviso and Coyote Hills have been subject to flooding. Their tendency to nest on attached levees also increases their exposure to predation. Predators have been observed to dig under and otherwise defeat barriers erected to protect nesting birds. The desertion of the Drawbridge/Mowry colony was associated with an increase in observations of predators, such as red fox and feral cats, in the area, with direct evidence of predation (SFBBO unpubl. data). At Bair Island, the colony was deserted following episodes of predation and erosion of levees leading to tidal inundation of the colony (SFBBO unpubl. data, Harey et al. 1992).

Contaminants may also pose a threat to local Caspian tern populations. Ohlendorf et al. (1988) found high levels of DDE, mercury, and PCBs in the eggs of Caspian terns nesting at Bair Island. Additionally, the colony at Brooks Island may be exposed to contaminated prey from the Levin Richmond Terminal Superfund Site (Harey et al. 1992). The impact of these contaminants on the reproductive success and populations of Caspian terns in the San Francisco Bay is unknown at this time. At Alameda NAS, the current plans for development and usage of the existing runways could adversely impact the nesting colony there.

Human disturbance is a potential threat at Brooks Island. It is important to restrict public access to the site prior to and during the breeding season, which spans from early April to the end of August. Other
Other Birds

...colonies are in areas of restricted access where human disturbance is minimal, but they should be carefully monitored.

Caspian terns are known to re-occupy historical nesting areas many years after they have been abandoned, as was most recently demonstrated by the re-occupation of the Coyote Hills site. As predation is reduced and levees become suitable again after maintenance, Caspian terns may return to former nesting areas in the South Bay. Therefore, it is important to conserve and protect these areas even though there are no Caspian terns currently occupying them.

Research Needs - Colonies in the South Bay have declined from a mean of 1,626 individuals (SD = 491) between 1971 and 1988, to 136 nests in 1997. Possible causes of this decline include predation by red fox and feral cats, disturbance by levee maintenance, flooding, and environmental contamination. Continued population monitoring is needed to document future population trends. Studies of reproductive success and fledgling success would be valuable in documenting problems with year-to-year reproduction. By combining these studies with studies of predation and the presence and effects of contaminants, we may better understand the causes of the current decline. Studies of nest site selection coupled with studies of reproductive success will provide information on which features of the habitat are most important to the reproductive success of these birds. This information can assist in lessening the impact of levee maintenance on the Caspian tern by allowing improved maintenance techniques with more rapid recovery of disturbed nesting areas.

Acknowledgments

I would like to thank Jennifer Parkin for her assistance with the text; Linda Wires for assistance with national and regional population trends; Howard Cogswell for supplying records from his field notes and a helpful review of the text; William Bousman and the Santa Clara County Breeding Bird Atlas for numerous records; and the many dedicated volunteer observers at the San Francisco Bay Bird Observatory.

References


**Personal Communications**

Charles Collins, Department of Biology, California State University, Long Beach.
Leong@community.net.
Jennifer Parkin, San Francisco Bay Bird Observatory, Alviso, California.

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**California Least Tern**
Sterna antillarum browni
Leora Feeney

**Introduction**

The California least tern was first described from a specimen collected in San Diego County (Mearns 1916). It is currently one of three subspecies of least terns in the United States — S. antillarum antillarum is found on the East Coast (Lessons 1847), and S. antillarum athalassos is associated with the great interior river systems of the United States (Burleigh and Lowery 1942). The California least tern was listed as an endangered species by the Federal government in 1970 and by the State of California in 1971.

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**Description**

The length of the least tern averages 23 cm. (9 in.), and its wingspan is about 51 cm (20 in.). The rump and upper tail of adults are a pale gray, concolorous with the back and upper wings. The outer primary feathers are black, creating narrow black outer wing edges. The tail is relatively short and forked. During the breeding season, the adult head is crowned in black, with a white triangular patch on the forehead. The bill is yellowish, and tipped in black. The short legs are a varying yellow-orange color. The basic plumage of some adults is sometimes observed in California late in the breeding season. These birds will have dark bills and legs. The definition of the black cap and triangular head patch is lost to a white face with sooty cap. It takes two to three years for least terns to mature. There sub-adult plumages are similar in complexity to other members of the gull group and are therefore not described here. It is impossible, for the most part, to separate least tern subspecies in the field, and races are usually identified by distribution.

The least tern bears a close physical resemblance to the little tern (Sterna albifrons), which is found seasonally along coastal waters of Great Britain, Europe, Africa, Asia, and Australia. During a period when combining or “lumping” species was a trend, the two accepted New World least terns, S. antillarum antillarum and S. antillarum browni, became subspecific (Hartert 1921) to the worldwide species. This was supported later (Peters 1934). Studies defining differences in morphology, behavior, and vocalizations (M assey 1976) provided the foundation needed to support the original separate specification, and in 1983, the American Ornithological Union split the American least tern from the more cosmopolitan little tern; the New World tern was once again S. antillarum. The least tern is slightly smaller than the little tern, making it the smallest member of the gull family, Laridae (Olsen and Larsson 1995).

Two other subspecies of least terns are in the literature, but are not found in the United States; S. a. mexicana is found along the east coast of the Gulf of California or the Sonoran coast (Van Rossem and Hachisuka 1937) and S. a. staebleri is found in southern Chiapas (Brodkorb 1940).

**Breeding Biology** — Least terns typically arrive at California breeding areas in middle or late April. Courtship is observed from the time birds arrive. Nesting is reported in “two waves,” the first from early May through early June, and the second from mid-June through early July. The species is a colonial nester, although single pairs are sometimes found.

Least tern nests are simple depressions in the substrate, called scrapes. One to three (usually two) eggs require about 21 days of incubation. The young are downy and able to walk soon after hatching. Least terns with adequate food resources fledge from about 17 to
21 days. Flight skills that allow young birds to follow their parents to foraging areas typically take longer (Laura Collins, pers. comm.). Both parents tend the young and share at some portion incubating, brooding, and feeding responsibilities. Young, well-fledged, least terns eventually leave breeding sites and disperse to localized post-breeding foraging areas where fish are plentiful and waters are calm. These post-breeding foraging areas, which offer young birds opportunities to develop foraging skills and provide all terns the food to build reserves for migration, are considered by some to be as important to the survival of juvenile terns as the nesting areas (M assey and Atwood 1984). Several post-breeding sites in the Bay Area are located at South Bay “intake” salt ponds. Shallow tidal areas are also used, such as at the E. B. Roemer Bird Sanctuary in Alameda and at Roberts Landing in San Leandro. California least terns most often finish breeding activities by late August and are usually absent from California breeding and post-breeding areas by late September.

Distribution and Abundance

The California least tern is migratory. Winter distribution is largely unknown, although least terns banded as chicks in California have been found as far south as southern Colima (M assey 1981) and Guatemala (Charles Collins, pers. comm.). Least terns have been found along the coast of Peru (Schulenberg et al. 1987), Panama (Vacher 1988), and Costa Rica (Stiles and Skutch 1989, Barrantes and Pereira 1992), but these accounts do not specify S. a. browni. A better knowledge of winter locations and migratory routes for California least terns would greatly improve our understanding of this bird.

During the breeding season (spring and summer), California least terns are found nesting along the Pacific Coast as far north as Pittsburg, Contra Costa County, California (Collins 1988) and as far south as Bahia M agdalena (Palacios and Alfaro 1993). In addition, there are reports of S. antillarum nesting at the northern Gulf of California on the Colorado River delta (Palacios and M ellink 1994) and in northwestern Sonora on the Mexican mainland (M ellink and Palacios 1993), without reference to subspecies.

In the State of California, least terns nest annually at about 35 sites from San Diego County to Contra Costa County. Some colonies, though reported separately, are so close as to be considered related sites. For instance, the Mission Bay area in San Diego County has three small colonies. The breeding locations shift somewhat due to annual conditions; nearly 40 colony sites are monitored (Caffrey 1995a).

It was once thought by some that California least terns nested from the Mexican border north only as far as Monterey County (Wilbur 1974). However, records show the bird’s presence further north in Santa Cruz County from 1939 through 1954 (Wilbur 1974). Accounts of least tern numbers in California prior to 1970 are sketchy, however, colony numbers described as “abundant,” in the “thousands,” “good-sized,” “1,000,” “600 pairs,” and “large numbers” were reported at numerous sites along California’s coast at the turn of the century (as in Caffrey 1995b). By 1971, Craig (1971) reported less than 300 pairs over only 15 sites. Craig’s work was limited and may have overlooked some sites. In 1973, Bender (1973) located 624 pairs statewide. After state and federal listings, recovery efforts and sometimes intense management strategies were put into place. Recovery efforts succeeded. Surveys in recent years have indicated fluctuating numbers, but in 1995, approximately 2,536 pairs of least terns were estimated to have nested at about 35 California nesting locations (Caffrey 1995a).

Sightings in the San Francisco Bay Area date back to 1927. Curiously, the earliest Bay Area sighting was in the city of Alameda (Grinnell and Miller 1944), where the current largest northern California colony breeds, with over 200 pairs in 1996 (Laura Collins, pers. comm.). Although least terns, including groups with over 50 birds and juveniles, had been sighted in the Bay Area for decades (Allen 1933, Allen 1934, Chase and Paxton 1965), it was not until 1963 that nesting was confirmed at the Oakland Airport (Roemer 1963) and at another Alameda County location soon thereafter (Chandik and Baldridge 1967). These nesting sites caused some to speculate that breeding least terns had expanded their range. However, the fact that California least terns, including young, have been seen in the Bay Area since the 1920s could also support the contention that an infrequently seen population of the bird may have bred undetected here for decades.

At the present time, Alameda’s least tern colony and two to three least tern pairs nesting at the Pittsburg Power Plant are the only known Bay Area nesting sites producing fledglings. In 1995, one to six pairs nested at the Oakland Airport, but all failed due to predation (Feeney 1996). In the past, least terns were documented to nest on Bair Island (CDFG 1981, Anderson 1970), and on various salt pond levees (CDFG 1981).

Although the history of the least tern in the San Francisco Bay Area is not clear, the Bay Area birds are today considered a critical population – vital to the statewide species recovery effort. In 1995, California Department of Fish and Game preliminary numbers showed that the Alameda Colony was the State’s fourth largest producer of fledglings (CDFG, Unpubl. data).

There are currently large gaps between breeding colonies throughout the modern range of the California least tern, probably due primarily to disturbance and habitat loss. A particularly significant gap of 330 km (178 miles) occurs between the breeding Bay Area least terns and the nearest breeding colony to the south (Pismo Dunes) (Caffrey 1995b).
Habitat Use and Behavior

California least terns forage by hovering over shallow to deep waters and diving or, less often, dipping onto the surface of the water to catch prey. Least terns also make short skimming approaches onto pools of water left on mudflats during low tide to catch trapped prey items. Although California least terns have been known to consume a wide variety of fish species, they appear partial to northern anchovy (Engraulis mordax), and silversides (Atherinidae sp.) (Atwood and Kelly 1984, Collins 1985).

To a much lesser extent there is evidence that least terns may take small invertebrates such as the water borne larvae of drone flies, Eristalis tenax (Laura Collins, pers. comm.; Leora Feeney, pers. obs.). For nesting, least terns require tracts of open sand or fine gravel substrate with sparse vegetation. Loss of natural habitat has caused these birds to become opportunistic, using areas such as newly filled or graded lands and airports for nesting. Nesting areas must be located near open water, usually along coastal beaches and estuaries, and they must host adequate numbers of small elongate fishes to sustain adults and growing young.

Conservation and Management

Human development of least tern habitat, highway access to the coast, and summertime beach recreation have caused destruction of breeding sites and resulted in least tern breeding failures. Although recovery efforts have brought about increased least tern numbers in California, some problems continue to challenge these efforts. It appears that for colonies to have guaranteed successes, they require intense management policies to protect nest sites, including regular monitoring of breeding activities, adequate barriers or supervision to restrict public access, persistent predator control, and vegetation management. These measures can be costly and funds are not always available for known breeding sites to be properly protected.

Predator management has become more difficult due to the recent introduction of red fox (Vulpes vulpes) on California’s coast. Feral cats and the establishment of cat feeding stations in the State have added to least tern reproductive failures (Edwards 1919, Caffrey 1995b, Feeney 1996). Public support for feral animals has created additional problems with predator management programs. In recent years, there has been concern over reduced fish availability at some sites, which may be related to “El Niño” weather patterns or other phenomena (Caffrey 1995a).

To assure the future of a healthy least tern population in the San Francisco Bay Area, adequate habitat must be set aside and properly managed to support nesting and post-breeding foraging. These protected areas should be established at several locations around the Bay to allow for alternative safe sites during potential localized habitat crises.

References

Craig, A.M. 1971. Survey of California least tern nesting sites. Ca. Dept. Fish and Game Project W-54-
Western Burrowing Owl

*Athene (Speotyto) cunicularia hypugaea*

Lynne A. Trulio

**Introduction**

The western burrowing owl is a small, semi-fossorial bird of prairie and grassland habitats. It is the only owl that routinely lives and nests underground. Burrowing owls in the western United States rarely dig their own burrows, but take over burrows dug by ground squirrels (*Spermophilus* spp.), prairie dogs (*Cynomys* spp.), badgers (*Taxidea taxus*) or other burrow digging species (Zarn 1974). The western subspecies of the burrowing owl lives west of the Mississippi to the Pacific Coast and from southern Canada into northern Mexico.

The species was listed by the California Department of Fish and Game as a Species of Special Concern in 1979. In November 1994, the U.S. Fish and Wildlife Service listed the species as a federal Category 2 Candidate for listing as endangered or threatened. The revised and shortened candidate species list in the February 28, 1996 Federal Register does not include the burrowing owl. In California, owl numbers have declined 50-60% in the last 10 years.

**Personal Communications**

Charles Collins, Department of Biology, California State University, Long Beach.

Laura Collins, biologist, Berkeley, California.
Description

The burrowing owl is a small, brown and white mottled owl, approximately 230-280 mm (9-11 inches) tall. It is not easily confused with any other owl due to its semidorsorial nature. Males and females are often difficult to distinguish in the field and, unlike many other raptors, the male is slightly larger than the female. Adults weigh an average of 150g (Zarn 1974). Chicks less than three months of age are distinguished from adults by their completely buffy breast and white collar. Chicks often emerge from the burrow weighing approximately one-half to two-thirds adult weight, and they reach adult weight within a month of emergence (S. N eudecker, pers. comm.; T rulio, pers. obs.).

Western burrowing owls are migratory throughout much of their U. S. range and leave their breeding grounds in the fall. Owls often return to the same or nearby burrows the next spring. This site fidelity is well documented (M artin 1973, G reen 1983). In most parts of its range in central and southern California, the owl is a year-round resident.

Breeding Biology – Burrowing owls are primarily monogamous for the nesting season. They produce one clutch per year, but may lay a second clutch if the first is lost. Burrowing owls lay between two to eleven eggs (average four to six) in a chamber of the nest burrow. Eggs are laid between M arch and A ugust depending upon location. T he female incubates the eggs for approximately 26 to 29 days (S. N eudecker, pers. comm.; Z arn 1974). After the chicks hatch, they remain in the burrow for approximately three weeks. Just before or just after they emerge, chicks lose their natal down and gain juvenile plumage. An average of two to four chicks emerge, although clutches of six or eight are not uncommon. Generally, an average of two to four chicks survive to fledging (age at which they can fly), which occurs about one month post-emergence. Fledglings remain with their parents until fall and then disperse. They molt by this time and gain their adult plumage. Some young remain with their parents until fall and then disperse. A typical juvenile plumage. An average of two to four chicks emerge, although clutches of six or eight are not uncommon. Generally, an average of two to four chicks survive to fledging (age at which they can fly), which occurs about one month post-emergence. Fledglings remain with their parents until fall and then disperse. They molt by this time and gain their adult plumage. Some young remain with their parents through their first winter (P. D elvoyas, pers. comm.).

Demographic parameters and their relationship to populations are not well known. T homsen (1971) found a juvenile survival rate of 0.3 and an adult survivorship of 0.81 based on two years of study at the Oakland Airport, Alameda County. T he longevity of free-living owls has not been documented, although an average of five years is informally used (P iest, pers. comm.). Fecundity is better known. T homsen found an average of 3.9 chicks survived to fledging. At M offett Field, Santa Clara County, T rulio (1994) found an average fledging success of 2.6 chicks per reproductive pair (SD =1.4) and an average of 1.8 chicks per pair (SD =1.7). An average of 73% of pairs produced emergent chicks. T his level of fledging success is the second lowest reported in the literature.

Food and Feeding – During the day, owls may be visible at their burrows, but tend to do little hunting. They become active at dusk and do most of their feeding at night (H aug and O liphant 1990). T hey use multiple feeding strategies, including running along the ground, but most often they can be seen hovering over fields and diving at prey. H aug and O liphant (1990) found burrowing owls at their site in Canada had an average home range size of 2.4 sq. km, and owls confined 95% of their movements to within 600 m of their nest burrows.

Burrowing owls are mid-level carnivores preying primarily on large insects and small rodents, but they will take a wide variety of prey. M any studies have found that important food items include vole species (L agurus spp., M icrotus spp.), mice (P eromyscus spp., M us spp., R ethromyomys spp., Zapus spp.), pocket mice (P erognathus spp.), pocket gophers (T omomys spp.), and young ground squirrels (S permophilus spp.), as well as a wide array of beetles, grasshoppers, crickets, reptiles, amphibians, small birds, fish, and crustaceans (Z arn 1974). T hey forage in ruderal, manicured, or natural grasslands. Burrowing owl predators include the great horned owl, harrier, and the red fox.

Distribution and Abundance

Burrowing owls inhabit flat, dry, open grasslands in prairie and arid habitats throughout California, exclusive of the humid, northwest coastal areas and the forested and shrub-covered mountains. Burrowing owls are most abundant in wide, low, interior valley bottoms and in flat coastal lowlands (G rinnell and M iller 1944). D eSante and R uhlen (1995) found that fully 92% of the breeding owls occurred in such lowland areas, generally below 60 to 300 meters in elevation.

Once a widely distributed and relatively common grassland bird, the burrowing owl has been declining significantly in California for at least the last 40 years (G rinnell and M iller 1944). At least 50% of the population has been lost in the last 10 years. D eSante and R uhlen (1995) estimate that approximately 9,450 pairs of birds remain in the State. O ver 71% of these pairs live in the Imperial Valley, an area subject to rapid human population growth and development in the near future (D eSante and R uhlen 1995). In the San Francisco Bay Area, nearly all the owls, approximately 170 pairs, are found in the South and East Bay between Palo Alto and the Fremont-N ewark area.

Researchers have noted burrowing owl declines, especially since the 1950s (Z arn 1974, A rlib 1979, J ames and E thier 1989), although there was little quantitative data to support this impression until just recently. T he Institute for Bird Populations completed an extensive, cooperative three-year study (1991-1993) of the burrowing owl population in California, exclusive of the Great

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Other Birds
Basin and desert areas (DeSante and Ruhlen 1995; DeSante et al., in press). Overall, in the entire census area, nearly 60% of the breeding groups of owls known to have existed during the 1980s had disappeared by the early 1990s (DeSante and Ruhlen 1995).

The species has disappeared as a breeding bird from substantial portions of its former range. DeSante and Ruhlen (1995) showed that the burrowing owl apparently has been extirpated as a breeding species within the last 10 to 15 years from Marin, San Francisco, Santa Cruz, Napa, coastal San Luis Obispo, and Ventura counties, as well as from the Coachella Valley. It has been very nearly extirpated from Sonoma, Santa Barbara, Orange, coastal Monterey, and San Mateo counties. Perhaps only one to two breeding pairs still exist in most of these counties.

The basic threat to burrowing owls in California is the annual, methodical loss of breeding and foraging habitat to development by humans. Habitat loss to urban development and destruction of ground squirrels (DeSante and Ruhlen 1995, Trulio 1995) are two primary reasons for the decline. Other factors include soil disturbances such as disking, grading, and blading, vehicular strikes, and predation by non-native or feral species. In agricultural areas, where the majority of owls live, chemical spraying may be contributing to population declines.

**Habitat Use and Behavior**

Historically, owls were found in natural areas of open prairies or open shrub-steppe habitat (Butts 1971, Coulombe 1971). Human population growth and continuous land use changes have resulted in burrowing owls utilizing human-altered habitats ranging from agricultural irrigation ditches (Coulombe 1971) to urban habitats (Collins and Landry 1977, Trulio 1995, Thomsen 1971). Burrowing owls have become quite tolerant of human presence, as long as suitable nesting and foraging habitat exist. Florida burrowing owls (S. c. floridiana) readily use suburban areas up to a density of approximately 60% development; when development densities exceed 60%, owl numbers drop (Wesemann and Rowe 1987).

Good burrowing owl habitat is open, dry, and sparsely vegetated with available burrows (Zarn 1974). However, several other subtle characteristics make some burrows more suitable than others. These characteristics include percent vegetative cover, height of vegetation surrounding the burrow, the presence of colonial fossorial mammals, soil texture, and presence of perches for horizontal visibility.

The California Department of Fish and Game’s “Wildlife Habitat Relationships System” database lists 18 major habitat types that support burrowing owls. In most of these habitats, burrowing owls are generally found in open country, where tree or shrub canopies cover less than 30% of the habitat. Typical habitats include annual and perennial grasslands, open agricultural areas, deserts, and vacant lots.

Burrowing owls are able to adapt to some human-altered landscapes. The land uses of sites where owls live include dry open grassland, the perimeters of agricultural fields, irrigation ditches, fallow agricultural fields, open fields prepared for development, airports, golf courses, military bases, and parks.

These owls can be found adjacent to the San Francisco Bay on levees next to salt ponds, open unmanicured grasslands, or manicured fields near the Bay’s edge where ground squirrel numbers and foraging area are adequate. These birds are primarily terrestrial predators and in these locations still focus on mice and insects. However, they are opportunistic and will eat species associated with wetlands, including amphibians and crustaceans (L. Yuen, pers. comm.).

**Conservation and Management**

Increasing burrowing owl numbers will require adding more nesting and foraging habitat. Burrowing owls are an indicator of the marsh-upland edge of the San Francisco Bay. Within the structure of the San Francisco Estuary Goals Project, burrowing owl habitat may be increased by adding upland transition zones between the high marsh and lands converted to human use. These zones should include short grass habitat capable of supporting a healthy population of ground squirrels. Trees should be kept to a minimum.

Increasing habitat for burrowing owls should also provide upland refugia for marsh species that must escape high tides, such as salt marsh harvest mice, as well as black and clapper rails. Since burrowing owls are predators and since this habitat will also benefit marsh hawks, adequate cover for mice and rails must be provided.

**References**


DeSante, D.F., E.D. Ruhlen, S.L. Adamany, K.M. Burton and A. Sabrina. 1997. A census of burrowing...


Personal Communications

Penelope Delevoryas, H. T. Harvey and Associates, Alviso, California.

Joan Priest, Silicon Valley Wildlife Rehabilitation, San Jose, California.

Dr. Stephen Neudecker, Chula Vista Nature Center, Chula Vista, California.

Lois Yuen, Berkeley, California.

Other Studies and Information Sources


Expert Sources

Jack Barclay, Albion Environmental, Inc., Santa Cruz, California. Jbarclay@albionenvironmental.com.

Janis Buchanan, California Burrowing Owl Consortium, San Jose, California. Jbuchanan@gaia.arc.nasa.gov.

Penelope Delevoryas, H.T. Harvey Associates, Alviso, California.

David DeSante, Institute for Bird Populations, Point Reyes Station, California. Ddesante@birdpop.org.

Brenda Johnson, University of California, Davis. Bjohnson@mother.com.

Joan Priest, Wildlife Center of Silicon Valley, San Jose, California.

Brian Walton, University of California, Santa Cruz, Predator Bird Group.
Salt Marsh Common Yellowthroat
Geothlypis trichas sinuosa
Scott Terrill

Introduction

The common yellowthroat (Geothlypis trichas spp.) is a small, insectivorous warbler common in most of North America. Twelve subspecies of common yellowthroat have been recognized in the United States (AOU 1957), however, yellowthroat taxonomy remains complicated and there is room for further work (e.g., see Howell and Webb 1995). Grinnell and Miller (1944) listed three yellowthroat subspecies that breed in California: the "western yellow-throat" (G. t. occidentalis), the "tule yellow-throat" (G. t. scirpicola), and the "San Francisco yellowthroat" (G. t. sinuosa), currently known as the salt marsh common yellowthroat. Based on Grinnell and Miller (1944), occidentalis is the most widespread and breeds over much of California (excluding the higher Sierra Nevada mountains). The breeding distribution of occidentalis surrounds the breeding distribution of sinuosa, which is restricted to San Francisco Bay wetlands and adjacent riparian areas. However, the American Ornithologists' Union (AOU) considers occidentalis as restricted to central eastern California and considers the subspecies that surrounds sinuosa as G. t. arizela. Thus, the AOU considers four subspecies to breed in California. The breeding range of scirpicola comprises the western portion of southern California and the Imperial and Lower Colorado River valleys. Arizela and occidentalis occur in the Bay Area in winter (Evens et al. 1997).

The salt marsh common yellowthroat was first identified as a distinct subspecies by Grinnell (1901). He described this subspecies as being darker dorsally and laterally and smaller than the other two subspecies of yellowthroats found in the State. Mewaldt and Rigney attempted to repeat Grinnell's results and were unable to do so with respect to coloration (Rigney, pers. comm.), however, size (specifically wing chord) did appear to hold up, and thus small size may be the primary characteristic for distinguishing this taxon (Rigney, pers. comm.). Foster (1977a,b) compared specimens of all three subspecies and found wing length difference to be significant at the 95% confidence level. Raby (1992) found that 81% of sinuosa and arizela populations were distinguishable from one another by song. Marshall and D edrick (1994) indicated that sinuosa can be distinguished by dark coloration and small size. These authors found that birds in prealternate molt (July through September) can be distinguished by the darker coloration of the emerging feathering on the back and flanks relative to pale "occidentalis (often called arizela)." Raby (1992) found that the Grizzly Island population represented a zone of intergradation between sinuosa and arizela, and more work is needed to clarify the taxonomic situation in Suisun Bay.

Associated species that use similar habitats include the marsh wren (Cistothorus palustris), red-winged blackbird (Agelaius phoeniceus), and the salt marsh song sparrows (Melospiza melodia samuelis, M. m. pusillula, M. m. maxillaris).

Description

The name "salt marsh common yellowthroat" is somewhat of a misnomer, since sinuosa occurs in salt marsh only in winter (Foster 1977a,b). Rather, this subspecies breeds in fresh and brackish marsh associated with and close to Bay wetlands. Thus, this taxon has also been referred to as San Francisco yellowthroat (Ray 1916, Schussler 1918, Sibley 1952).

Breeding Biology – Male salt marsh common yellowthroats begin establishing territories by mid-March and the nesting season extends from early March through late July (Hobson et al. 1986). Females construct the nest relatively close to the substrate (ground or water). Yellowthroats lay three to five eggs, which are incubated for 12 days. The young remain in the nest for 10 days and are fed by both parents for at least two weeks following fledging (Hobson et al. 1986).

Food and Feeding – Yellowthroats are primarily insectivorous and glean insects on or near the ground (to about five or six feet above the ground or water) from low herbaceous vegetation, bushes, and small trees, or from the surface of mud — although they will forage substantially higher during the non-reproductive period (Shuford 1993). Yellowthroats in California eat 99.8% animal matter (Shuford 1993). The main dietary items in a sample of 114 were ants, wild bees and wasps, true bugs, beetles, caterpillars and moths, flies, grasshoppers, and spiders.
Predators – There is little direct information on predators of salt marsh common yellowthroats. However, likely predators include species that typically prey on passerines, including feral cats, raccoons and red foxes (eggs and nestlings) and raptors such as Cooper’s and sharp-shinned hawks.

Distribution and Abundance

Salt marsh common yellowthroats have been collected in San Luis Obispo, Los Angeles, and San Diego counties (Grinnell and Miller 1944). Sibley (1952) referred to sinuosa as a resident species, although the collection of specimens outside the breeding range indicates at least a migratory element to the population. It should be added that there is an influx of other races of common yellowthroats into the San Francisco Bay Area during migration and in winter. The relative numbers of “western” common yellowthroats and salt marsh common yellowthroats in Bay wetlands at this time of year is entirely unknown. I assume that some specimens of sinuosa were collected from salt marshes during winter and that these specimens provided the basis for the statement that salt marshes provide wintering habitat for salt marsh common yellowthroats (Foster 1977a,b). However, Foster (1977) stated that it seemed likely that the birds that winter in Salicornia marshes of San Francisco and San Pablo bays breed in adjacent brackish marshes, and Hobson et al. (1986) indicated that some populations of sinuosa appear to be any substantiation that any race other than sinuosa.

Foster (1977a,b) found populations to be at critically low numbers in the South Bay and Peninsula areas and greatly reduced from historic abundance throughout the breeding range. During the 1975-76 season, Foster identified breeding habitat at O’lema Marsh, Limantour Estero, San Pablo Bay, Napa Marsh, Lake Merced, Sharp Park, Searsville Lake, Palo Alto Marsh, “Alviso” Marshes (including the San Francisco Bay National Wildlife Refuge in the vicinity of Artesian Slough), Coyote Hills Regional Park, Alameda Creek, San Gregorio Creek, and Pescadero Marsh. Foster (1977b) estimated that 200 pairs were present in the areas occupied in the 1976 breeding season. This estimate increased to 230 pairs the following year with more thorough coverage. However, 1976 and 1977 were severe drought years with significant impact on marsh vegetation, and it was recognized that Foster’s yellowthroat status reports during those years might not have been representative of normal conditions. This assumption was supported by surveys conducted in 1985 in which breeding populations of salt marsh common yellowthroat in many areas were higher than in 1977 (Hobson et al. 1986). Foster (1977b) assumed the density of yellowthroats in suitable habitat was the same before alteration of marsh areas began, and estimated that the population under pristine conditions was probably on the order of 2,000 to 2,300 pairs (but see below for more recent, higher estimates). Finally, Foster (1977b) calculated that the population of salt marsh common yellowthroats had been reduced by 80 to 95% in the past 100 years based on calculated loss of suitable habitat. Hobson et al. (1986) estimated that fewer than 900 breeding birds occurred in all habitats.

The estimates of both Foster (1977b) and Hobson et al. (1986) may have been low. In a very recent study (Evans et al. 1997), 239 pairs were estimated to breed at Point Reyes alone. An earlier study (Hobson et al. 1986) estimated 135 pairs from the Point Reyes Peninsula. Evans et al. (1997) attributed the increase in estimated population size to increased coverage, increased habitat values due to changing land-use practices, and to more favorable weather patterns in 1996 relative to 1986. The most recent estimate on population size in tidal marsh alone, presented in Nur et al. (1997), was 6,000 to 11,000 breeding birds.

No salt marsh common yellowthroats have been collected during the breeding season outside the range described by Grinnell and Miller (1944), which is bounded by Tomales Bay on the north, Carquinez Strait on the east, and Santa Cruz County on the south (Foster 1977a,b). Within this range, all specimens collected between March and August were sinuosa (Foster 1977a,b). Sibley (1952, p. v), evidently, erroneously distinguished between common yellowthroats breeding in South Bay freshwater marshes as “yellowthroats,” and yellowthroats breeding in salt marshes around the shores of San Francisco Bay as “race sinuosa.” There does not appear to be any substantiation that any race other than sinuosa breeds in South Bay marshes.

Habitat Use and Behavior

Common yellowthroats form a complex of superspecies and subspecies that inhabit North and Central American dense, brushy habitats, generally associated with wetlands or moist areas. In California, yellowthroats are found in freshwater marshes, coastal swales, swampy riparian thickets, brackish marshes, salt marshes, and the edges of disturbed weed fields and grasslands that border soggy habitats (Shuford 1993). In the San Francisco Bay region as a whole, about 60% breed in brackish marsh, 20% in riparian woodland/swamp, 10% in freshwater marsh, 5% in salt marsh, and 5% in upland vegetation (Hobson et al. 1986, Shuford 1993).

Yellowthroats frequently use borders between these various plant communities, and territories often straddle...
the interface of riparian corridors and ecotones between freshwater or tidal marsh and the upland vegetation of weedy fields or grassland (Shuford 1993).

Breeding habitat has been divided into three main categories (Foster 1977a,b): (1) woody swamp (e.g., Olema Marsh, Searsville Lake, etc.); (2) brackish marsh (e.g., Napa Marsh); and (3) freshwater marsh (e.g., Coyote Hills Regional Park). Nests are well concealed and are primarily found on or near the ground in grass tussocks, low herbaceous vegetation, cattails, tules, and bushes to approximately five feet above the ground (Kendeigh 1945, Gross 1953, Stewart 1953, Shuford 1993). Breeding generally starts in mid-March to April, and second clutches take the breeding season into August (Foster 1977b).

Conservation and Management

Foster (1977b) attributed an estimated salt marsh common yellowthroat population decline of 80-95% over the past 100 years to increasing urbanization of the Bay Area and consequential loss of habitat.

Based on Foster (1977a), California Department of Fish and Game (Fish and Game) staff recommended that suitable salt marsh common yellowthroat habitat be maintained and protected in parks and refuges. Other specific recommendations included the preparation of a management plan for Olema Marsh and Limantour Estero, development of fresh and brackish water marsh areas in a portion of New Chicago Marsh, protection and enhancement of habitat in Coyote Hills Regional Park, and incorporation of salt marsh common yellowthroat habitat protection into management of Napa Marsh and Skaggs Island Naval Base. The staff report also recommended that planning agencies in Marin, San Francisco, San Mateo, Santa Clara, and Alameda counties, as well as other agencies and entities that manage or otherwise influence yellowthroat breeding habitats, be advised of yellowthroat breeding habitats in their respective counties and be encouraged to assist in efforts to preserve and enhance these areas. Fish and Game staff concluded that further study should be conducted to determine the location, quality, and extent of salt marsh common yellowthroat wintering grounds; seasonal movement patterns; minimum size of marsh habitat that will support breeding birds; and relative productivity of the various marsh habitat types used by breeding yellowthroats.

Foster (1977b) made the following additional management recommendations:

- **Encourage water treatment plant operations that allow treated discharge to flow into the Bay at places where it will support marsh vegetation, rather than discharging it in the deeper parts of the Bay by means of pipes.**
- **Encourage parks and other agencies that have yellowthroat breeding habitat in their jurisdiction to ensure the water supply to these areas, by artificial flooding if necessary. Foster (1977a) noted that the yellowthroat habitat least affected by a drought (1975-76) was habitat in which water levels were artificially maintained. Such areas included Coyote Hills Regional Park and outfalls of sewage treatment plants.**
- **Research by Hobson et al. (1986) concluded that continued loss of habitat (due primarily to development), poor habitat management, and drought or flood could seriously affect the future of this subspecies. They recommended further study involving systematic banding, recovery, and resighting; a study of dispersal corridors, buffer zones, and nonreproductive season habitat requirements; and more taxonomic work on the Suisun Bay population to determine the eastern boundary of sinuosa.**

More recently, Nur et al. (1977) recommend extensive surveys of salt marsh common yellowthroats to derive reliable estimates of population densities in a diversity of potential habitats. Nur et al. (1977) also recommend a molecular genetic analysis to clarify the genetic relationship of this “taxon.”

References


Foster, M.L. 1977b. A breeding season study of the salt marsh yellowthroat (Geothlypis trichas sinuosa) of the San Francisco Bay Area, California. M.A. Thesis, San Jose State Univ., San Jose, Calif.


Grinnell, J. and A.H. Miller. 1944. The distribution of the birds of California. Pacific Coast Avifauna No. 27.

Gross, A.O. 1953. Northern and Maryland yellowthroats, Florida yellowthroat, western yel-


**Personal Communications**

Michael Rigney, Coyote Creek Riparian Station, Alviso, California.

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**Savannah Sparrow**

_Passerella sandwichensis_

**Howard L. Cogswell**

**Introduction**

The savannah sparrow is a small (12-15 cm), inconspicuous, and unobtrusive bird whose usual perch is a weed stalk in a meadow and seldom anything higher than a fence post. Although this bird bears some resemblance to the song sparrow, it is grayer and smaller than its cousin, and its tail is shorter and slightly notched, that of the song sparrow being somewhat rounded. Adults have a dark brown crown with a distinctive pale yellow mark in front of the eye. The savannah sparrow's song is a weak, buzzy trill, preceded by two introductory notes of differing pitches.

Some 17 subspecies of savannah sparrow are currently recognized (Wheelwright and Rising 1993), all of the northern ones being migratory. The southernmost subspecies in the west are all resident or nearly so, including _P. s. beldingi_ of the salt marshes from Morro Bay, California, south into northwestern Baja. That subspecies, now officially listed as threatened, occupies a very similar habitat within its range as do the three salt marsh races of the song sparrow in the San Francisco Bay Region (see report on song sparrows, this publication). The "Coast" subspecies of the savannah sparrow, _P. s. alaudinus_, breeds from Humboldt County south to San Luis Obispo County where it intergrades with _beldingi_ at Morro Bay. Through much of its range, _alaudinus_ occupies some salt marsh areas, but also the more moist sorts of grasslands of the Coast Ranges. Unlike the very
dark beldingi, which is easily distinguishable in the field from other races, that winter in its range, birds of the alaudinus subspecies (which includes the former P. s. bryanti of narrower range) are not readily told from those other races except by close comparison and often measurements in hand. So data on relative numbers of birds of the different wintering subspecies within our area of interest are nearly non-existent.

Description

The 28-page account for savannah sparrow in the new "Birds of North America" series (Wheelwright and Rising 1993) should be a primary source for anyone investigating this species. It includes some information about the various subspecies, particularly the "well-marked" ones of southern California and west Mexico, as well as the pale P. s. princeps that breeds on one island off Nova Scotia. The account includes distinguishing characteristics, distribution, migration, habitat (but this lacks any quantification - see below), food habits, sounds, behavior, breeding (including development of young, parental care, etc.), demography and populations, conservation and management, appearance (including molts and geographic variation), and measurements. A few items about the species of special interest for resource managers, as gleaned from this account, seem worth mentioning here.

Male savannah sparrows defend territories (announced by song from frequently used perches), which usually include or may be adjusted to include the much smaller territory defended by nesting females. Polygyny has been noted in a number of populations in good habitat (thus, a census of just singing males may under-report the size of a breeding population). Nests are normally on the ground, well hidden under tussocks of grass or low shrubs and often with an approach tunnel of up to several inches.

From studies in eastern Canada and eastern United States to the Great Lakes area, annual survivorship of adults ranges from 28 to 70%, varying both by location and by years. As with most small birds, mortality in the first year is high, but once a savannah sparrow is a year old the rate remains relatively constant for five to six years, after which it rises abruptly. So an expected maximum life span would be about that number of years.

Although the population dynamics of beldingi and alaudinus seem not to be reported in the literature, studies from other areas may provide some insight. According to the reference by Wheelwright and Rising (1993), young birds from island populations (especially princeps) show a much higher natal philopatry (tendency to return to the area where they were hatched/raised) than do the young from mainland areas. The mere fact that beldingi is so highly differentiated in its markings while alaudinus is not, would indicate a similar relationship between these races in California.

During the breeding season, savannah sparrows feed primarily on insects of various orders and all stages of development, spiders, and small crustaceans. In the non-breeding period, the diet is predominantly small seeds and fruits gleaned from the ground or low vegetation - the shift occurring as the young sparrows are fledged in mid-summer. Even when breeding, these sparrows do considerable amounts of foraging off their territory. In the non-breeding period, they may appear to be gregarious, but the "flocks" are probably primarily aggregations drawn together by attraction to a good food supply and have no ongoing social structure.

Predators that take savannah sparrows or their eggs and young are extremely varied; an instance of a clapper rail doing so is cited.

Effects of human activity on savannah sparrow populations have probably been beneficial, over-all, due to extensive clearing of forests and maintenance of agriculture and grazing lands. Pesticides applied to habitats occupied by the species are probably detrimental, but this seems not to have been documented. (These authors do not mention filling or draining of wetlands as a factor - perhaps because of the northeastern emphasis in the research that has been carried out on the species.)

Distribution and Abundance

Savannah sparrows are found nearly continent-wide at the appropriate season, wintering commonly from northern California, Missouri, and New Jersey south to Central America, and irregularly or sparsely as far north as New England and coastal British Columbia. The overall breeding range extends from Arctic shores of Alaska and Canada, south to northern Georgia and Illinois to Colorado, northeastern Arizona, and through California west of the southern deserts, with additional populations in western Baja California, Mexico and on the Mexican tableland.

In various parts of its range, the savannah sparrow is found occupying open, mostly un-wooded habitats of many sorts - from arctic tundra and mountain meadows to hayfields (particularly old or unkempt ones) to cultivated croplands, wet (but not flooded) meadows, marsh borders, and near-desert grasslands in some locations. Wintering savannah sparrows of various subspecies are also to be found in any such open habitat in the Bay Area (more on non-breeding numbers, below). However, the habitats selected by the breeding form, alaudinus, were summarized by Johnston (1968) as follows. "It maintains populations in two main types of habitat in coastal California: the Salicornia association of tidal marshes and the grassland associations of the coastal fog belt." Comparing its habitat niche with that of the salt marsh races of the song sparrow, he also wrote that the savannah "on salt marshes is limited to the broad expanses of low-lying salicornia (Salicornia ambigua) on...
the older and higher parts of marshes...[that] lie back of that salt marsh vegetation (cordgrass, Spartina foliosa) best suited to frequent submersion by tidal flooding” (Johnston 1968). Eight nests were found in 1971 by Gill (1977) in his survey of breeding birds around the Bay south of San Mateo Bridge, the preferred nesting habitat being “levee tops grown to annual grasses and high pickleweed growing on the levee banks.” Gill (1977) further estimated the overall nesting population of savannahs in his study area to be from 800 to 1,000 pairs; but the large size of area he surveyed precluded any calculation of densities for any type of habitat.

Savannah sparrows have been reported in at least 17 breeding-bird censuses1 on measured plots in California, although on several of these they were listed only as “visitors” to the plots. Five of these censuses were in bayside marsh or adjacent grasslands and, thus, sampled the gradation of habitats – from marsh to unwooded uplands – which this species exemplifies (Table 7.7). Only one of the five surveys listed in Table 7.7, the San Pablo Creek survey, was from within the geographic limits of the Goals Project. A similar census on a plot of brackish marsh and adjacent grassland at Southhampton Bay did not find any savannah sparrows (Stoner et al. 1963), nor did one of the somewhat brackish “diked coastal saltmarsh” three miles northwest of Alviso (apparently part of what is now called Triangle Marsh) in 1983 (Anderson and Jennings 1981). However, two census plots including habitat of breeding beldingi in southern California showed much higher densities for those strictly marsh-inhabiting birds: 60.8 territories per 100 acres along Ballona Creek in western Los Angeles (Dial 1978), and 104 territories per 100 acres in the Bolsa Chica Ecological Reserve in Orange County (Alexander 1974).

Winter population densities of savannah sparrows have been obtained for a few discrete plots on Humboldt Bay, near Marshall on Tomales Bay, and in coastal southern California. However, I know of no winter population counts for San Francisco Bay, except for some of intertidal mudflats that had a remnant bit of marsh at the edge – hardly a useful sampling for this species.

Some indication of typical numbers (subspecies not distinguished) may be gleaned from counts made two or three times a month for one year in the north-Newark and northwest-Fremont area (Figure 7.6). In these studies, several assistants and I made counts of all bird species on three properties of the Leslie Salt Company and three other “comparison” plots in the period of October 1984 through October 1985 – although songbirds were not a targeted subject for the counters until March 1985. Some of the plots were too large for the counts to be considered total censuses, thus the numbers counted are very likely well below the true densities.

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1 A full listing of these California breeding-bird censuses may be obtained from the author at CA BIRDS@aol.com.

### Table 7.7 Censuses in California Bayside Marsh or Adjacent Grasslands Where Savannah Sparrows Were Reported

<table>
<thead>
<tr>
<th>Location</th>
<th>Habitat Description</th>
<th>Date</th>
<th>Population Density (territories/acre)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>San Pablo Creek, Contra Costa County</td>
<td>Tidal marsh (Salicornia, Spartina, Grindelia, etc.)</td>
<td>(1979)</td>
<td>5T/70 ac. [=7T/100 ac]</td>
<td>Johnston 1952</td>
</tr>
<tr>
<td>Humboldt Bay, 3 mi. So. of Eureka, Humboldt County</td>
<td>“marsh” (but with much grass and some subshrubs)</td>
<td>(1980)</td>
<td>5.5T/26 ac. [=21.5T/100 ac]</td>
<td>Sorenson and Springer 1977</td>
</tr>
<tr>
<td>Humboldt Bay, 1 mi. So. of Eureka, Humboldt County</td>
<td>“marsh” (brackish, with various herbs and coyote bush)</td>
<td>(1980)</td>
<td>1+ visitor only/20.3 ac.</td>
<td>Base 1981</td>
</tr>
<tr>
<td>Coastal Prarie, Cypress Grove ACR property, Marshall, Marin County</td>
<td>Marine terrace grassland adjacent to bayside freshwater marsh</td>
<td>(1988)</td>
<td>1+ visitor/31.0 ha.</td>
<td>Kelly 1989</td>
</tr>
</tbody>
</table>
Figure 7.6 Summary of Data from Six Bird Count Studies Conducted Between Late Fall 1984 and Winter 1985. Data from Cogswell (1986) and author’s personal raw data records.

<table>
<thead>
<tr>
<th>Location</th>
<th>Data Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hickory Tract, Newark</td>
<td>65 acres, partly barren old salt ponds, diked pickleweed marsh, old head of Newark Slough, several acre patch of partly filled land with grass and forbs.</td>
</tr>
<tr>
<td>Coyote Tract, Newark</td>
<td>About 174 acres, only about 60 of which were vegetated, mostly used as pasture.</td>
</tr>
<tr>
<td>Coyote Tract, Fremont</td>
<td>About 100 acres, some 60 acres being nearly barren former salt ponds, the remainder well vegetated former gun club ponds re-grown to various herbs, including dock, spike-rush, grass, and local cattails; two small ponds much enlarged in winter.</td>
</tr>
<tr>
<td>Coyote Hills Regional Park</td>
<td>37 acre plot marked off with posts in 209-ft. squares. Seasonal wetlands with Salicornia, Scirpus robustus, Frankenia, local Rumex and other tall forbs, grasses in western 1/4, a belt of tall Typha marsh along a shallow ditch in eastern and northern parts; all in floodwater storage basin with water covering varying parts of the plot shallowly (many plants protruding) in winter.</td>
</tr>
<tr>
<td>Don Edwards S.F. Bay National Wildlife Refuge Headquarters Flat (LaRiviere Marsh)</td>
<td>Approximately 105 acres west of Thornton Ave. and south of route 84. Former salt crystallizers and ditches and dikes, plus smaller ponds becoming more marsh than barren; western 1/4 opened to tidal action in previous year, and large central part to muted tidal action (or diked off when gate closed) beginning in spring 1985. Dikes in north-central to northeastern and northwestern parts were usually not walked, so some savannah sparrow habitat was missed. Note: there was no singing or other evidence of breeding in this plot.</td>
</tr>
<tr>
<td>Newark Slough</td>
<td>Tidal marsh from the southwest corner of the south tip of Coyote Hills to the slough-head at Thornton Ave. Approximately 60 acres, including the main channel, which was here only about 2-10 feet wide. A few old dikes are in the eastern part, including one at the actual Mayhews Landing site that has tall forbs on it, the others fully covered by pickleweed and various high-mars plants. Many of the savannah sparrows noted were along the levee between the marsh and the adjacent salt evaporator on the southwest, where there was a narrow, interrupted fringe of upland plants. Note: Larger numbers in Mar and Oct ‘85 were found by walking the very narrow shoulder of Thornton Ave. fringing the marsh.</td>
</tr>
</tbody>
</table>
Conservation and Management

The local population data for the savannah sparrow are so meager that they provide no indication of any local historical population changes. For the continent as a whole, there may be notably larger populations now than occurred before European settlement and expansion (Wheelwright and Rising 1993); however, this can be presumed not to be the case on the periphery of San Francisco Bay. The filling in of the upper parts of many tidal marshes and the urban development that has taken place on uplands so created would have decidedly decreased habitat acreage and quality for savannah sparrows, at least in the central section of the Bay. The conversion of tidal marsh to salt evaporators also eliminated much of the marsh areas in the South Bay – particularly when this conversion is considered in combination with the subsequent urban expansion to the very edge of those salt ponds. Only the conversion of former tidal marshes to bayside or near-bay upland fields - such as the diked farmlands near San Pablo Bay and the gun club/grazing lands established years ago around parts of Suisun Bay - may have been favorable to savannah sparrows. Modern data on the distribution and numbers of the species, especially during the breeding season, are sorely needed from those areas. Such data would be of most value to resource managers if they were obtained with reference to the different vegetation and the changes in it emanating from various land-use practices over all seasons and for years of different rainfall patterns.

The savannah sparrow (especially its subspecies alaudinus) is an ideal target species to represent those birds dependent upon both Bay-related marshes of most kinds and the adjacent upland grasslands and fields of various sorts. The relative densities of its populations in the grasslands of higher elevations of the Coast Ranges and in the apparently relatively few areas where it breeds in valleys between these ranges are unknown. Maintenance or restoration of as much vegetation that is transitional from marsh to upland, in various parts of the Bay system where it is still possible, should be a goal until the comparison of qualities of this habitat combination to strictly upland types can be made. I suspect that the transitional marsh-upland habitat will be found to support far higher densities.

References


**Song Sparrow**

*Melospiza melodia samuelis*

- *M. m. pusillula*
- *M. m. maxillaris*

Howard L. Cogswell

**Introduction**

As a breeding species, the song sparrow (*Melospiza melodia*; Emberizidae/Emberizinae) is found across North America from southern Alaska and central and eastern Canada to northern Florida and Mexico, in the drier regions being restricted to riparian or other wetland habitats. As of 1957, some 31 subspecies were recognized as valid on morphological grounds (AOU 1957), including a very pale one in the southwestern desert area, and several quite large ones resident along the Aleutian-Alaska coast area. A population of “ordinary” song sparrows (regular-sized at 6.25 inches total length) breeding in Cincinnati, Ohio was the subject of intensive study using colored leg bands for a number of successive years. The resultant monographs (Nice 1937, 1943) made the song sparrow, for many years thereafter, the best known of any species of American song bird. The study provided details on the bird’s home range and territory, mating system (mostly monogamous), tendency to migrate (males there included some that left for the winter, but others that stayed), nesting, production, and survival of young, etc. Nice (1937, 1943) also set a high standard for others in her exhaustive search of relevant literature from throughout the world.

Tidal marshlands along the Atlantic Coast from Long Island to Virginia are occupied by a distinct subspecies of song sparrow (*M. m. atlantica*), but it has apparently been little studied. At least the northern populations of that race are migratory (AOU 1957). In the tidal marshlands about San Francisco Bay, however, three distinctly separate subspecies have evolved, and all of them are apparently quite resident year-round within those marshlands or immediate vicinity. These are:

- *Melospiza melodia samuelis* of San Pablo Bay and northern San Francisco Bay (south to Sausalito and north Richmond); it was first recognized as distinct in 1858.
- *Melospiza melodia pusillula* of the balance of San Francisco Bay shores (breeding originally from San Francisco and southeast Richmond south to Alviso); first described in 1899.
- *Melospiza melodia maxillaris* of the Suisun Bay marsh complex and west to include South Hampton Bay; first described in 1909.

Marshall (1948a, 1948b) studied all of these subspecies from the standpoint of the habitats occupied, and the very tenuous connections their populations had with adjacent upland forms of the song sparrow. The gist of his findings was that all are quite distinct in size or proportions of bill, wings, tail (etc.) and/or coloration from the upland subspecies. Only one of these subspecies, *samuelis* has been studied in detail using banded birds (Johnston 1954, 1956a, 1956b); but findings from that study that pertain to the birds’ adaptations for life in an intertidal area have been supported by incidental observations made on both *pusillula* and *maxillaris* and are used in this account as applicable to all three of these forms. The same assumption was a key aspect of the California Department of Fish and Game staff report to the Fish and Game Commission (Larsen 1989) when a petition to list *maxillaris* as Endangered was forwarded with the recommendation that Threatened status was warranted. A general survey of the status of all three San Francisco Bay races of song sparrows was done by Walton (1975), and Marshall and Dedrick (1994) presented an updated review that ties the estimated populations to the acreages of remaining tidal marshes. The last paper also has a color figure that illustrates diagnostic features of each of these subspecies compared to their adjacent upland relatives.

The presence within such a small, overall region of three such narrow endemics is in itself a highly valuable sample of evolution at the critical “not-yet-species” level – the case histories of which serve to illustrate early stages of the gradual process of speciation.

**Description**

Song sparrows of most of the United States are “typical” Emberizine sparrows about six to 6.5 inches in total length, with rather average body, neck, head, bill, and leg ratios. They have somewhat shorter wings (approxi-
Birds of the *maxillaris* race are slightly smaller, and considerably less rusty-toned in dorsal color than *gouldii* of the adjacent Marin and Sonoma County uplands. The South Bay *pusillula* birds are still smaller, especially the bill, and with a tail averaging 10% shorter than the upright form there, santacruceis (included in gouldii by the AOU). Marshall and Dedrick (1994) show *pusillula* as being generally grayer in background tone, with a distinct light gray collar (but brown-streaked) on the hind-neck; but at the Museum of Vertebrate Zoology in Berkeley, H. Cogswell surveyed over 50 spring-summer specimens of *pusillula* (many collected by Marshall himself) and could not distinguish this grayness on at least half of them. The degree to which differences persist or disappear with wear of the feathers apparently remains to be worked out. The Suisun Bay birds, *maxillaris*, are more nearly equal to typical upland song sparrows in size, and show much rusty coloration in the lighter areas above and on the back of the neck and the tail; but their most distinctive feature is a somewhat swollen basal half or so of the bill (noticeable only on very close inspection). This last feature should be the easiest clue to distinguish them from *mailliardi* of the adjacent Sacramento and lower San Joaquin valleys, but Marshall and Dedrick (1994) do not illustrate that subspecies.

**Breeding Phenology** — Territories of salt marsh song sparrows are apparently "held" all year, even if not actively defended in fall and early winter. Singing by males is prevalent by February, however, and one of the major adaptations discovered by Johnston (1954, 1956a) for *samuelis* is that the peak date for completion of the first clutch of eggs is more than two weeks earlier than that of song sparrows of the nearby upland race. The mode of this laying comes about March 27 with fair numbers of birds completing first clutches up to mid-April. The advantage of early laying is that the nests are less vulnerable to being flooded by the gradually increasing height of high tides that occurs annually in late April and even more in late May and June, the young of the early nesters fledging before then. In some years, the first clutch layings were delayed by bad weather, and loss of eggs or nestlings to flooding was widespread. Nests are not always placed in the highest vegetation available, a trait that Johnston attributes to the selective pressure of nest-predators. He also found a lower peak of clutch-completion in mid-May and a third small one (seven nests out of 111 total) in June, when spring-phase high tides would almost certainly flood most of the marsh. These may include re-nestlings by birds that lost their first brood, but many were apparent attempts to raise a second brood even if a higher percentage are then flooded out. Indeed Johnston (1968: 1548) says "almost all pairs nest twice in a season. If replacement nests are considered, each pair will nest on the average 2.5 to (rarely) three times each season."

The same seasonal time pattern for clutches of eggs is seen in *pusillula*, of which many sets of eggs were taken by collectors prior to 1940, and presumably also for *maxillaris*, for which relative few data are at hand. Of 80 egg-sets of *pusillula*, 50 were taken in April, 25 in May, five in March, and only one in June. There were 14 sets of *samuelis* eggs, all of which were taken between April 8 and May 7, and seven sets of *maxillaris* eggs, all of which were taken between the first and 23rd of May. Whether there is a difference in timing for individual birds of any of these subspecies that hold territories adjacent to a suitable dike or other bit of upland that permits nesting above the level of the June high tides has not been investigated. Johnston’s study area had a dike only along its northeastern side, with channels attractive to sparrows only at its two ends, so the question was not addressed.

Nests of *samuelis* were found by Johnston (1956a, 1968) to be placed on average 9.5 inches above ground, but averaging 12 inches in the marsh lower in the tide range. Any of the four main marsh plant species (Salicornia, Spartina, Grindelia, Distichlis) were used for nest support; but the uppermost parts of the tallest (Grindelia) were avoided, which Johnston attributed to the selective effect of predators finding nests there more readily.

**Productivity** — The mean clutch size of 157 nests found in Johnston’s study varied among years from 2.91 to 3.42 eggs, with the mid-season (April 6 to May 25) nests having a slightly larger mean number (3.23) of eggs than the earlier or later ones. However, the mid-season group for 1953, the year with highest success of early nestings, was almost identical to the later nest group which Johnston (1956a: 37) suggests might be a response to the saturated population, including the many juveniles still on hand from the early nests.

Basic natality (number of eggs/pair/year) ranged from 7.5 to 9.1 in the years 1952-55 of Johnston's study (1956b: 260). Since his studies of dispersal (see below) indicate a very strong tendency of the song sparrows to remain in the area where they were hatched and raised, he assumed that a color-banded individual that disapp...
Mortality factors applicable to the egg and nestling phases of the birds in the study by Johnston (1956b): 266 show predation and high tides accounting for 20% and 11%, respectively, of the losses. Storms were irregularly important, as in 1955 for 13% of the deaths. Brood-parasitism by the brown-headed cowbird is listed as a factor in five deaths (5% of the 1955 total of eggs and nestlings), but is not discussed. In more recent years, at least at the Hayward Shoreline, pairs of pusilla have been often noted feeding cowbird fledglings, so the increased population size of the cowbird now may be a negative factor of significance for these marsh-inhabiting sparrows. Johnston (1956b) also recorded about 49% mortality of eggs attributable to rodent predation (mostly Norway rats), and 59% loss of nestlings from the same factor. These were higher figures than all losses from high water and desertion. Other predators of minor influence were thought to be northern harriers and short-eared owls - but only four song sparrow skulls were found in 491 pellets he examined from the owls in a four-year period (Johnston 1968: 1550).

Dispersal - Johnston's (1956a) study of the subspecies population on San Pablo Creek Marsh in the 1950s included the distances from hatching site to breeding site of 34 juveniles, in 23 of which being 200 meters or less, and in only four birds was it more than 600 meters. Some authors have objected that his single study area was too small to really check on this. However, there was a pronounced exponential drop-off in distance from the peak at 100-200 meters, and nearly 15% of the 241 nestlings he banded were found subsequently breeding in the study area - a very good number when one takes into account the 50% or higher nestling and early fledgling mortality. Furthermore, the Poisson statistical test for randomness in the observed distances showed that while most birds moved very little, a small percentage of individuals had a tendency to move a distance much greater than the mean - beyond the 500-600 meter distance where none were found. This pattern was true also with Nice's song sparrows in Ohio, so is probably a genetic feature in the species - only the absolute distance being shorter in samuelis and presumably also the other salt marsh subspecies. The significance for conservation of the populations of these subspecies is that continuity of habitat is of very great importance in ensuring their ability to repopulate local areas where all or most individuals are eliminated by graphically and time-limited factors, i.e., local ecological disasters.

Trophic Relationships - The salt marsh races of the song sparrow, where present in the fully tidal marshes to which their evolutionary history fits them, are the most abundant of the passerine birds to be found there. Therefore, they must constitute an important segment of the food web in such communities. But just where should that segment be placed? On a year-round basis, they must be classed as omnivores; but in most of their breeding period from March through June or so, their mostly animal food would shift them upward trophically into the carnivore blocks - to some extent even into a secondary or tertiary carnivore position because they eat spiders and many carnivorous insects. But what does one do with the carnivore or omnivore that eats detritus-feeders on a large scale from the intertidal mud, which these birds do at least at times? The simple producer/consumer trophic picture is complicated by large quantities of detritus being recycled into the same trophic chains as are the organic products of green plants. Until a more thorough analysis of the percentages of the various types of food taken by birds of these three subspecies has been done, any detailed diagram could have no quantification of the relative importance of the connections shown.

Nearly the same lack of quantification is true for the known trophic relationships that would show song sparrow biomass passing on into predators, parasites, and scavengers, or for the reduction in the sparrow population success that is attributable to the brood-parasitic brown-headed cowbird. The behavior of the song sparrows on a salt marsh when a northern harrier or short-eared owl flies within its view indicates that these predators do indeed take some of the birds. However, the few song sparrow skulls Johnston (1956a) found in pellets from these owls indicates that the transfer of biomass to that species would be a very minor link in the food web. On a population basis, the most important predators of the salt marsh song sparrows are no doubt the Norway rat (Rattus norvegicus) and garter snakes (Thamnophis sp.), with now, in recent years, the non-native red fox (Vulpes fulva) added to the list, and in the parts of the marsh near a dike or upland, feral cats as well.

Habitat Use and Behavior

Except as otherwise indicated, this section is essentially summarized from Johnston's (1956a,b) results for a population of samuelis north of Richmond. He had some difficulty in attracting adults into traps, but was able to
color-band 33, plus 13 juveniles; most of his data, however, came from the 241 nestlings that he so marked. The marsh in his whole study area covered about 200 acres in 1950-55, but the song sparrows did not inhabit the bayside one-tenth or so that was dominated by Spartina, nor the 60% or so that was relatively unbroken Salicornia lacking small curving channels. Instead, they were in the parts of the Salicornia zone next above the Spartina zone and along Wildcat Creek itself and smaller channels with frequent small branch channels or sloughs where scattered to numerous Grindelia subshrub “bushes” provided both song perches and usual nest placement above the level reached by most spring tides. Other plants found in this and the upper marsh zone are also listed by Johnston (1956a: 27). During lower tide periods, the mud along the same small side-channels (areas where marsh plants are typically overhanging) provided primary forage areas for these birds.

The territories of salt marsh song sparrows are much smaller, at least in this optimum habitat, than the average of one acre or a little more found by Nice in Ohio, and by many others elsewhere if one judges from the densities reported in various breeding-bird censuses even in “good” riparian situations. Johnston (1956b: 256) showed the linear arrangement of the 14 to 22 territories along one particular tidal channel through six successive years. He reported (p.254) that “thirty feet represents the average width of territories along sloughs within which the birds perform practically all their activities [emphasis added] ... With this as definition of the area in which these song sparrows live, the actual density in a year of high numbers [1953] is seen to run from 8 to 10 pairs per acre. This is not as unreasonable as it may seem, for 10 pairs of song sparrows per acre would give each pair about 4,350 square feet per territory.” — about a 30- by 150-foot swath along one of the small sloughs, as he in fact found to be true. The density per unit area based on the whole marsh, including the parts not inhabited by song sparrows, would of course be much lower. For example, 74 territorial males were found in 70 acres of this same marsh (105 per 100 acres) and reported by Johnston (1952) in a census that included only two other breeding bird species: five territorial male savannah sparrows and four clapper rails. Year-to-year variation in a 100-acre part of this marsh (presumably including the 70-acre 1953 plot) was estimated to range from 87 to 124 pairs. Four different small sloughs supported song sparrows at “linear” densities along their length of a pair every 130 to 170 feet in 1953. The extremely sedentary nature of the territory holders is shown by data in Johnston (1956a), summarizing those birds with territories mapped in two or more successive years. Of the 48 instances of possible movement (24 birds) where movement could have happened, the only cases where territory centers were more than 16 meters from the previous year were two for males and three for females, and none was moved more than 35 meters.

Johnston’s two papers (1956a,b) contained but the barest mention of what happens to song sparrows that hold territories in the salt marsh when the tides are high enough to flood most of the vegetation in the Salicornia zone — as they are on the high high tides of May through June and again in November through December. He stressed the shift in breeding times, but did not report on the behavior of post-breeding birds. A selection of records from my own notebook (unless otherwise indicated) and a few other sources indicate that there is often temporary aggregation at the upper fringe of the marsh and particularly on any adjacent dike or fill with vegetation; for example:

- About 50 (5+ juveniles) were at the upper end of Plummer Creek, south-southwest of Newark, at high water on August 19, 1956;
- A similar number was seen while driving the dikes along Mt. Eden Creek, southwest Hayward, on September 10, 1964;
- Thirty to 50 were seen along the railroad fill that crosses the large Dumbarton Marsh on various summer and fall dates, and 100 or more on October 10, 1953 when the railroad was walked eastbound and the nearby San Francisco Aqueduct back westbound (Cogswell 1953, 1956);
- About 120 were seen from a boat plus a short distance afoot (along the railroad) in this same marsh on October 26, 1969 spanning a 7.5 foot high-water;  
- About 185 were estimated in walking from near Dumbarton Bridge to Newark Slough along the railroad on January 27, 1979;
- An estimated 75 were seen in two miles of dike through a salt marsh near Alviso on November 19, 1949 at high water (Sibley 1955);
- From 51 to 86 were counted in the marsh fringes along Alvarado Channel (Old Alameda Creek) and/or the northern side of the new Alameda Creek Channel on Christmas Bird Counts from 1967 to 1981 at various times of tide, including a “flock” of 18 feeding on driftwood massed in a salt pan in the marsh at high water on December 31, 1967.

Elsewhere in the Bay, high estimates have been:

- 200 along Sears Point Road (=amudis) on January 28, 1938 by Graham and Stoner (publ. in The Gull), and 150 by myself on November 19, 1950, during an Audubon group trip by boat down Suisun Slough with walking about on a part of Joice Island (=maxillaris). I believe that these aggregations include many territory holders, perhaps mostly in areas without significant number of Grindelia or other plants that would still provide cover at the higher high tides, and that these birds return to their territories as the tide ebbs. Johnston (1968: 1552) later reported some limited
investigation of such aggregations during winter high tides, at one time having 17 birds perched on the raft he was using to explore the marsh just after a northern harrier flew by.

**Food and Foraging** - Johnston (1968: 1551-2) gives considerable detail of the foraging behavior of song sparrows in the salt marshes, presumably based on his four years of study of samuelis. Preferred forage sites are the muddy edges of small channels, but they also obtain food on the firmer ground under and from the marsh plant surfaces themselves. While they engage in the double-scratch bouts separating periods of pecking at the ground surface, like other sparrows, they also specialize at times on the small molluscs and other marine invertebrates in this intertidal mud. They also feed at the maturing heads of the Grindelia flowers, and in autumn (when fresh water from insect foods is at a low ebb) eat the fleshy fruits and tiny seeds of the Salicornia. In winter, after the seeds of the latter have been released from the dried spikes, they are washed up in the drift from high tides, and the salt marsh song sparrows spend much time picking at such accumulations, taking both seeds and various invertebrates. In spring and early summer, the young are fed almost entirely on insects [and other invertebrates], some of which are obtained by short hops or flutters approaching flycatching.

Quantitative study of the diet of these subspecies has been very minimal, although Beal is said in the literature to have reported in 1910 on some stomach contents of amuelis. Without regard to subspecies, the varied diet of song sparrows in general as reported in various early accounts is summarized by Nolan (1968).

**Quality Habitat** - The intensive study of a samuelis population by Johnston (1954, 1956a,b), the follow-up surveys of maxillaris populations from Southampton Bay through the Suisun Marsh complex as summarized by M. Marshall and Dedrick (1994), and numerous but more casual observations of pusillula populations in the Hayward, Newark, Alviso, Palo Alto, and Redwood City areas all lead to the general picture of highest populations of each of these subspecies being found in fully tidal marshes. This is true even though the vegetation differs very significantly in the marshes about Suisun Bay compared to San Pablo Bay and that in turn differs somewhat from the marshes of the southern arm of San Francisco Bay. The partiality shown for foraging along the banks of the sinuous minor channels within the tidal marsh, and the greater availability of nest sites and song perches in the Grindelia “bushes” that tend to grow along these same channels are probably the prime factors in supporting higher populations there. Johnston found no song sparrows with territories in the pure Spartina marsh adjacent to the mudflat along the open bay, but at Palo Alto they are moderately numerous in mixed Salicornia-Spartina marsh with a channel or adjacent dike and along the boardwalk that traverses the marsh. Various subsequent searches of open Salicornia flats have confirmed Johnston’s finding that the two subspecies that inhabit adjacent more complex marshes avoid at least breeding season residence where there are no small channels or adjacent mudflats or dikes.

The extent to which birds of each of these subspecies utilize, and indeed occupy for breeding purposes, the various types of diked marshlands near the tidal marshes is poorly understood. For maxillaris, the petition for that race to be listed as Endangered (M. arshall and M. ealدت 1988) and the California Department of Fish and Game staff report that accompanied it (Larsen 1989) both indicated that diked-off, managed marshes were not inhabited by birds of that race. However, surveys of many parts of the North Suisun Marsh in 1990 by M. arshall and several assistants located 79 pairs in non-tidal situations compared to 334 pairs or singing males on the 621 acres of tidal brackish marsh surveyed. H abitats mentioned for the non-tidal birds included coyote-bush (Baccharis pilularis) and roses close to a tidal slough (where those birds might have foraged at times). Seven birds were found in “hedges” of giant Atriplex and/or dry grass along a ditch with only water at the ends, and even a few were seen in dry fields with clumps of non-emergent tules. Cogswell’s South Bay field notes have for years regularly noted a few pusillula living, foraging, and singing along outer-levee rip-rap with only traces of salt marsh vegetation, and others at least foraging frequently in diked-off Salicornia with various herbs and sometimes a few shrubs admixed. However, all or most of these sightings were within a few hundred feet from a tidal channel or the outer bayshore. It is not clear from observation with binoculars whether the several birds that regularly sing from coyote-bushes or tall clumps of sweet fennel (Foeniculum vulgare) on the western side of Coyote Hills, Fremont, are pusillula or upland-race birds. The only mud available to them by May is in the adjacent tidal or diked-off marshes.

Table 7.8 shows the relevant Goals Project “key” habitats, and summarizes their usage by each of the three salt marsh song sparrow races.

**Distribution and Abundance**

Because there is insufficient data on the actual locations and population sizes of song sparrows, this section outlines the historical changes in the extent (and quality) of suitable habitat, and provides estimates of population sizes based on habitat availability.

**M. m. pusillula** - Dedrick (1993) measured the total historic (pre-diking) suitable habitat within the range of this subspecies at 65,871 acres and the present habitat area as only 10.2% of that, or 6,678 acres (M. arshall and Dedrick 1994: table 1). I performed a separate evaluation to estimate the remaining tidal marsh habitat in the range of pusillula, and to rank the quality...
of this habitat as well. For this work, I measured on 1977-84 aerial photographs (courtesy H. T. Harvey Associates) every habitat block or portion of block that could be classified by close inspection (using some magnification) as of “high,” “medium,” or “poor” quality for this subspecies. Areas of high quality were those containing tidal marsh with numerous small channels and complex vegetation structure. Areas of medium quality were those with fewer channels or small parcels remote from major parcels. Poor quality was assigned to those tidal marsh areas that did not have the features indicated for high or medium quality. I then considered the position of each habitat block within the landscape and rated each block in overall quality categories of A, A-, A/B, B, B-, B/C, and C quality categories (with a few even of D level). I then summed the total acreage in the highest three categories (A through A/B) and the next three highest categories (B through B/C), and got 3,989 acres and 2,511 acres, respectively. Together these total 6,500 acres, compared to Dedrick’s 6,678 total. The acreages of the eight largest single blocks rated as A, A-, or A/B were:

- Dumbarton Marsh (incl. adjacent “Aqueduct Marsh”) - 836.9 acres
- Greco Island (Redwood City) - 740.4 acres
- Outer Bair Island (incl. 474.9 ac. reopened to tide) - 603.5 acres
- Mowry Slough mouth to Newark Slough Mouth - 326.1 acres
- Mundy Marsh (Palo Alto) & Faber Tract to Cooley Landing - 316.2 acres
- Whaley’s Tail Marsh (N & S of Alvarado Channel, Hayward) - 271.1 acres
- Corkscrew Slough (in several segments) - 200.4 acres
- “Ideal Marsh” (Ideal Basic Industries, west of Coyote Hills) - 128.3 acres

* The presence of 17 territories of M. m. pusillula in a 14.7 acre plot of tidal salt to brackish marsh (Spartina foliosa, Salicornia virginica, Scirpus robustus, Frankenia, Distichlis) NNW of Alviso (Anderson and Jennings 1981) with two nests found in S. robustus is the best indication of breeding by this subspecies in brackish tidal marsh.

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<th>Table 7.8 Key Habitats Usage by Three Salt Marsh Song Sparrow Races</th>
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<td><strong>M. m. pusillula</strong></td>
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<td><strong>Tidal Salt Marsh</strong></td>
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<td><strong>Tidal Brackish Marsh</strong></td>
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<td><strong>Seasonal Wetlands</strong></td>
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<td><strong>Salt Ponds: dikes or levees</strong></td>
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<td><strong>Intertidal Mudflats</strong></td>
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<td><strong>Adjacent Uplands</strong> (mixed grass and tall forbs or shrubs)</td>
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* The presence of 17 territories of M. m. pusillula in a 14.7 acre plot of tidal salt to brackish marsh (Spartina foliosa, Salicornia virginica, Scirpus robustus, Frankenia, Distichlis) NNW of Alviso (Anderson and Jennings 1981) with two nests found in S. robustus is the best indication of breeding by this subspecies in brackish tidal marsh.

2 Marshall and Dedrick (1994) give the current size of the Dumbarton Marsh as 906.1 acres, but from their text it appears they included all of the marsh belts along Newark Slough up to its head, whereas I kept them separate above the first points where diked salt ponds restrict the marsh to belts along the sloping slough banks.
These blocks are all separated from other sizeable habitat blocks by distances or channels wide enough that they probably constitute a deterrent to free dispersal by the sedentary song sparrows, even though the full-grown birds would have the capability of flying the distance involved. These larger blocks can be grouped with intervening smaller ones to obtain a more geographic comparison. When this is done, the “West Bay” optimum of Bair and Greco islands south through Palo Alto has a total of 1,544.3 acres of “good quality” tidal marsh, while the nearly contiguous marshes in the Dumbarton Point-Newton Slough to Mowry Slough and south to outermost Coyote Creek (where still not very brackish) has 1,719.7 acres.

Using Johnston’s (1956b) published population density figures for samuelis (1.11 territories per acre), Marshall and Dedrick (1994) estimated the total pusillula population to be 7,412 pairs – a little over 10% of that preceding the diking and/or filling of the tidal marshes. Johnston’s density figures assumed no song sparrows in those often rather large parts of a tidal marsh more than 10 meters or so distant from a channel. Additional fieldwork is yet to be done to check the actual species population densities at the locations which I quality-rated in my study; until such quantitative checks are completed, no better overall population estimate for pusillula than those arrived at by Marshall and Dedrick (1994) can be derived.

**M. m. samuelis** - Marshall and Dedrick (1994) gave the results of planimeter measurements on the Nichols and Wright (1971) map of historic tidal marshes within the range of this subspecies as 63,690 acres, as well as Dedrick’s (1993) 1992 measurement of the tidal marshes remaining – 14,060 acres. At 22.1% of the original, this is the best record of marsh retention among the three salt marsh song sparrow ranges, despite the fact that many blocks are narrow and/or isolated about the shores of San Pablo Bay. These authors reported examples of locations where birds of this subspecies were found and some captured in a 1986 survey, also noting that some “verdant marsh” [but diked] areas (e.g., on Tubbs Island) lacked them. Nor were any found in the 2,416-acre bayfront marsh between Sears Point and Mare Island, although Marshall’s notes from 1947 in-2,416-acre bayfront marsh between Sears Point and Tubbs Island) lacked them. Nor were any found in the

An area of 80 ha in the central part of the Petaluma Marsh was surveyed intensively for song sparrows (Collins and Resh 1985). In this study, the vegetation was sampled along transects perpendicular to channels, and the marsh variations grouped into four habitat-types for analysis of the song sparrow data (sloughs, natural channels, mosquito-control ditches, and areas 10 m or more from any channel or ditch). The sparrow’s territories were mapped by following the birds and the polygon of each male’s mapped song-posts measured. Seven replicate plots in each habitat-type other than the areas beyond 10 m from a waterway (which were found to have no sparrows) were thus surveyed.

In general, territory placement and sizes among natural channels were found by Collins and Resh (1985) to be very similar to that reported by Johnston (1956b), but the plots along the constructed ditches had fewer and larger territories. Collins and Resh (1985) attributed this difference to a lower level of food and nesting-site (and predator-avoidance) resources than was present along the
sinuous natural channels with their much greater vegetation diversity. The density of the samuelis sparrows was also checked in the non-breeding season during the dispersal phase of the young, thus reflecting the relative attractiveness of each habitat-type. Even then, the natural channel and slough-bank areas were occupied by from 1.5 to three times the number of birds as were found near the ditches. Collins and Rish (1985) concluded “ditches are not preferred habitat for the salt marsh song sparrow, primarily because ditches support shorter and spatially less diverse vegetation than apparently occurs along sloughs and natural channels. However, ditches do provide additional habitat that has increased the carrying capacity of Petaluma Marsh for salt marsh song sparrows:’ [because the ditches are better than having no small channels at all]. The amount of marsh along tidal waterways [of any sort] was increased by 300% by such ditches and thus “ditching has added more than two thousand salt marsh sparrow territories to the Petaluma Marsh” (Collins and Rish 1985).

**M. m. maxillaris** - In their petition to list the Suisun song sparrow as endangered, Marshall and M ewaldt (1988) estimated the 1850s extent of tidal marshes within the range of this subspecies, plus those that developed as a result of sedimentation from placer mining at 66,618 to 73,712 acres. Such marshes, they also estimated, would have supported 69,949 to 77,398 pairs of these birds. These authors also cited a 90.4% reduction in area of such marshes, the total areas they had planimetered being 6,762 acres in 1986, which they estimated then supported about 5,666 pairs. Sizes and estimated populations were given for 10 different blocks of such marsh, and the distinctiveness in morphology and ecology of the taxon is summarized. Sites where previous investigators had conducted special studies or censuses were mapped and their results summarized. The petition included a map showing sectors of the Benicia Christmas Bird Count circle that included any tidal and adjacent marshes, and a graph showing the number of song sparrows (probably most, though not all of this subspecies) recorded in those sectors in the years 1977 through 1986 (compiled by Robin Leong). Year-to-year numbers varied considerably as census effort and methods of access varied. Thus they “do not reflect population changes but they show that the birds were constantly found over the years in optimum habitats” (M arshall and M ewaldt 1988). Michael Rippey’s measurements of seeds available to the salt marsh sparrows of San Pablo Bay-Napa River marshes and the Suisun Marsh area were also cited; the largest common seed of Suisun being that of alkali bulrush (Scirpus robustus) at 5/32 inch, which is notably larger than seeds of Distichlis, Grindelia, and Salicornia – a food resource feature that may help to explain the advantage for birds of this subspecies in having a swollen bill shape.

A California Department of Fish and Game staff report (Larsen 1989), forwarded to the Commission with the M arshall-M ewaldt petition, cited a number of additional pertinent studies and supported most of the petition’s findings, but the petition was rejected by the Commission. Nevertheless, these two reports in combination present an amount of detail for this subspecies not yet attained for the other two salt marsh races, for the details of the ecology of samuelis as learned by Johnston (1956a,b) were included as applicable to maxillaris as well. Under “Essential Habitat,” Larsen (1989) listed many details of the description by M arshall (1948a) for this subspecies. She summarized: “Suisun song sparrows use the tallest S. acutus in the centers of patches for song and calling perches, find concealment in the piles of dead stems, and forage on the bare surface of the mud between the stems and along the slough margins at low tide. They do not forage between stems that are only 2.5 to five centimeters [one to two inches] apart, but only forage in areas with stems that are 10 to 15 centimeters [four to six inches] apart. ... Thus they are limited to the area covered by tides, where flow is unimpeded by dikes, levees or channels.”

Larsen (1989) described marginal use of upland plants along levees by the sparrows, “but their territorial headquarters are always at the slough margins.” Their avoidance of diked marsh areas with Salicornia and Grindelia with no or impeded tidal flow was mentioned, although a few birds were found in such an area, on the eastern side of Cordelia Slough, that drained well through a culvert. [As noted above for pusillula, small populations of that subspecies also use such areas.]

M arshall and D edrick (1994) reported a more recent measuring of the tidal marsh areas of the Suisun to Southampton Bay marshes, and the surveys of locations where there were still birds of this subspecies found in or near those marshes, its historic range. Of the 64,552 acres of historic tidal marsh, 8,586 acres (13.4%) remained as of 1992, with a total estimated population of 9,530 pairs. M arshall and D edrick (1994) emphasized the prime habitat as being the fully intertidal brackish marsh, although they did find in a 1990 survey (in just parts of the North Suisun area) some 79 pairs (out of 3,803 total) occupying “non-tidal territories,” 30 of them near Roaring River. As mentioned above, M arshall (1948a) found a few of these in territories centered on a “hedge” in dry grass by a ditch with water only at the two ends.

Using sophisticated Geographic Information System (GIS) software, Scollon (1993) carried out the most thorough analysis of the remaining habitat for this subspecies. This study evaluated pertinent data for the tidal marshes, such as block size, distance from neighboring blocks, and availability of suitable habitat for dispersal between blocks, and rated the blocks and to some extent, the corridors, as to numbers of Suisun song spar-
rows they could support. The rating was based on the pairs-per-acre ranges for five "strata" of habitat quality as given in an unpublished 1992 document by Joshua Collins – the field data in support of which I have not seen. Scollon himself apparently did no field work with the sparrows, but integrated the spatial aspects of the findings of others who have worked with song sparrows in the range of maxillaris. For example, in addition to using Collins' "strata" evaluations, the dispersal distances found in one local population by Johnston in the 1950s [of samuelis, and cited above], plus a single comment by Larsen (1989) that these birds [maxillaris] "will generally not travel more than 50 meters over areas lacking protective cover, such as open water", were the sole basis for his ratings of the suitability of dispersal corridors. Nevertheless, Scollon’s (1993) maps of the various blocks of habitats, each with its rating indicated by type of hatching, and the subsequent "linking" of habitat patches [or, alternatively, not linking them] based on the dispersal capabilities as applied to the maps, result in a set of "population patches" for the subspecies. This is the broadest scale approach to a population analysis of any of the tidal marsh subspecies yet performed. Scollon (1993) presented histograms of all the population patches by area (<30, 30-399, and 400-1200 acres) and by four ranges of population size. However, these were all apparently based on the data from quick surveys of singing birds, or possibly in part on data from Johnston’s and Collins’ separate reports from samuelis populations. What is really needed, of course, is to carry out a number of field checks with actual censuses of maxillaris population blocks to see whether the basic assumptions underlying all these "What if?" manipulations in the GIS system are borne out. Such field checks should be done with blocks of several different sizes and several different degrees of isolation.

Every ecologically oriented investigation of this subspecies has indicated that its prime habitat is tall brackish marsh with full tidal flows. However, the rating system of habitat strata cited (from Collins) by Scollon indicates considerable difference of populations to be expected even within the intertidal brackish category. In addition, although several authors have stressed that birds of this subspecies "avoid" or "do not use" diked marshlands of various sorts, Larsen (1989), Marshall and Dedrick (1994), and Scollon (1993) all cite instances where smaller populations or limited numbers of maxillaris sparrows were indeed using such habitat. It is possible that most or all of such birds in diked (and therefore marginal?) habitat are those excluded by intraspecific competitors already on territory within more optimum habitat, and that their attempts to breed in the marginal situations are doomed to failure or sharply reduced productivity. Field studies are needed to address this critical ecological question, and to determine whether diked marsh could be managed in such a way as to provide good dispersal corridors, even if not breeding habitat. In the meantime, there is undoubtedly optimum value in fully intertidal brackish marsh with tall Scirpus vegetation along channels or sloughs with mud banks not too steep for low-tide foraging, and not too far from overhanging vegetation for protection from predators.

**Recommendations for Conservation and Management**

This section provides suggested goals for wetlands occurring in the range of each of the three subspecies of salt marsh song sparrow. These goals would be highly beneficial to population success of the target subspecies, while not overly detrimental to the populations of other important wetlands species in these same areas. The proposed goals are listed in order of priority for each subspecies/range.

**Range of M. m. pusillula (South to Central San Francisco Bay)**

1. Keep inviolate all bayward and slough boundaries of the existing large blocks of intertidal marsh. In the East Bay, this should include Dumbarton Marsh and its connecting "Aqueduct" and Newark Slough marshes, the Mowry Slough marsh and bayside marsh west and south of the sloughmouth, the "Ideal Marsh" (bayfront west of Coyote Hills), and "Whale's Tail Marsh" (south and north of the mouth of Alvarado Channel). In the West Bay, this should include Outer Bair Island plus Corkscrew Slough; Greco Island; and north and south of Cooley Landing through Palo Alto Baylands marshes (nearly contiguous now). In the far-South Bay, this should include Triangle Marsh (N NW of Alviso), Albrae Slough and nearby shores of Coyote "Creek," outer parts of Alviso and Guadalupe sloughs, Stevens Creek, etc. (all somewhat brackish but occupied). Other parts closer to the major sewage effluent outfall in Artesian Slough east of Alviso are of too low salinity to support vegetation that is usually occupied by this subspecies, as are the parts of Coyote Creek and Mud Slough near Newby Island at present.

One recently proposed change in land use across the largest block (Dumbarton Marsh) is to upgrade the railroad right-of-way there as a part of a new high-speed rail route from the Central Valley (and Los Angeles) to San Francisco. The fill along the unused tracks has been, for over 20 years, both a high-tide refugium for sparrows, rails, etc., and an avenue for access to the marsh by red foxes and feral house cats – although this avenue has been interrupted to the west and east in recent years by keeping "open" the bridges over the Dumbarton Strait and Newark Slough.
2. Expand any of the areas listed under the first priority above, especially in directions that would either: (a) provide habitat linkage for breeding populations of pusillula in locations between any of these areas, or at least good habitat for dispersal between populations in the event any local population suffers a disastrous decline; or (b) enhances the overall quality of habitat available within or at the edge of the habitat block. Examples of the latter might be to provide additional high-tide refugia where not now adequate, as on Greco Island (birds that use the Pacific Gas and Electric boardwalk are very vulnerable to predators), along various sloughs where the existing marsh is only in the form of a strip between the mudflat and open salt pond levee, or next to diked or industrial development.

Also in this priority should be the enhancement of habitat values for these birds in existing or planned smaller blocks. Such areas would include: Burlingame Cove, Belmont Slough and nearby Bird Island, inner Steinberger Slough, Ravenswood Slough and shoreline through the newly acquired gun club marsh near Dumbarton Bridge, La Riviere Marsh, and manyother Landing Tract of the National Wildlife Refuge, "Pond 3" marsh north of new Alameda Creek, M. t. Eden Creek-Baumberg Tract -Alvarado Channel in H ayward, tidal lagoons in both south and north parts of H ayward Re-gional Shoreline (plus possibly the adjacent H.A.R.D. marsh), newly tidal south part of San Leandro shore area, Arrowhead Marsh and other smaller marsh bits in the vicinity of San Leandro Bay and O akland Airport, Emeryville Crescent Marsh, Albay Cove and Hoffman Marsh (in se. Richmond) - the last being very close to the original northernmost point where this subspecies was found.

3. As the opportunities arise, establish new habitat for this subspecies in areas where it does not now exist. This effort should target areas that would add marsh corridors or patches along likely dispersal routes between major blocks of existing good to optimum habitat. Even small blocks would be worthwhile if spaced relatively closely in such areas. Major gaps in habitat acceptable to this form currently occur from the Bay Bridge through the O akland Estuary/Alameda (except the far eastern part of the south shore), salt pond levees not bordered by marsh though much of H ayward and Fremont, and the shoreline near developed areas through Foster City and San M ateo. It seems unlikely much opportunity will arise to create tidal marshes north of San Francisco Airport, but a 20-acre "marsh" (some to be open tidal flat) is a part of the Golden Gate National Reserve Area Plan for Crissy Field in the Presidio [implementation began in late 1999]. If birds of pusillula parentage are to reach it, they would probably have to be introduced there, at what was likely the northwestern-most limit of the range of this subspecies.

4. Restore, to the extent possible, a "natural" range of salt marsh habitats in the location of the operating salt evaporators or diked former salt ponds (e.g., on Bair Island), should any of these areas become available. This would not only greatly benefit the song sparrows of this subspecies, but also the clapper rail, salt marsh harvest mouse, and several other species. The slow development of marsh vegetation on the "scraped" parts of the former salt ponds in the Hayward Regional Shoreline tidal lagoons, and the siting of a number of the too-straight (engineered) channels excavated there, should be compared with the rapid development of salt marsh vegetation on reopened parts of Bair Island when choosing methods for restoration of the desired habitat.

Range of M. m. samuelis (North San Francisco Bay through San Pablo Bay) –

1. Keep the present large blocks of high-quality habitat in that condition. This includes not ditching the parts that already have adequate small channels with the diverse vegetation the sparrows prefer. The "keeping" also applies to the marsh corridors or small blocks that are spaced to provide dispersal opportunities among the larger blocks.

2. Restore sufficient intervening tidal marsh blocks or strips where there are currently the longest gaps in such. For example, suitable tidal marsh should be restored along the eastern side of the lower Napa River, and wherever possible along the entire Contra Costa County shore from Selby to San Pablo Point (the probable southeastern limit of the original range), as well as from San Rafael through San Quentin Cove. Topography prevents any suitable marsh corridor to Richardson Bay, which has some marsh remaining, but none of these sparrows in it according to M arshall (1948a,b).

3. Expand the tidal marsh area by opening to tide action some of the now disused salt evaporator ponds in the area between Napa River and Sonoma Creek. Since all or most of these ponds are now owned by the State, a truly major addition to the habitat for samuelis song sparrows (as well as clapper rails and other tidal marsh inhabitants) could be realized. Since this area adjoins an existing brackish marsh at Fly Bay and fronts on grassy uplands to the north (as does the existing tidal marsh east of the Napa River near Bull's Island), other species requiring the transitional sort of habitat, such as savannah sparrow and black rail, would also be benefited. This tidal marsh to
Upland zone is now all but absent in San Francisco Bay proper, so the goals for San Pablo Bay should certainly include a major provision to preserve and extend it.

**Range of *M. m. maxillaris* (Suisun Bay marshes and vicinity, west through Southampton Bay)**

1. Preserve the acreage and quality of existing habitat blocks used by all significant numbers of birds. To preserve quality, prevent further salinity intrusion into the Suisun marsh areas caused by greater diversions of freshwater flow in or above the Delta. With increased salinity, the tall brackish marsh to which this race is adapted would likely be replaced by shorter, more salt-tolerant vegetation, more like that in the range of *samuelis*. Birds of that race would, however, be unlikely to disperse eastward into the Suisun area because of lack of habitat along the Carquinez Strait.

Scollon (1993) modeled two levels of salinity intrusion, based in turn on salinity levels for May 1965 and February 1971, as mapped by Rumboltz (1979). Rumboltz’ (1979) 1971 map shows a level of 2,500 micromhos along a line extending across the middle of Honker Bay, and also Joice Island. According to Scollon (1993), this is nearly equivalent to the salinity standard of 2 ppt recommended by the San Francisco Estuary Project [for the continued health of the Suisun Marsh?]. At that salinity level, the vegetation, and hence the sparrows, of the marshes along the southern side of Suisun and Honker bays (totaling approximately 1,900 acres) have already suffered heightened salinity impact. However, there is insufficient recent data on maxillaris sparrow populations in this area with which to test this hypothesis. If that level of salinity intrusion holds, Scollon’s model (case A) predicts that the major populations in the northern part of the Suisun Marsh complex would escape severe impacts. These include the population in the largest block of marsh (1,394 acres) in the Rush Ranch-First M allard Branch area, as well as that in Hill Slough and vicinity (468 acres). In spring 1990, M arshall found 159 pairs of maxillaris in the western 154 acres of the former block and 58 pairs in 130 acres in the latter one (M arshall and D edrick 1994).

If, however, salt intrusion above the threshold level of major vegetation change extends to east of Chippis Island and includes most of M onetza Slough (Scollon’s 1993 case B), there would be a major reduction of maxillaris populations in these last strongholds of the subspecies, and in all other smaller blocks in the northern Suisun Marsh. Without knowing whether M arshall’s 1990 censuses were conducted in average quality habitat within the blocks of marsh he sampled, and without census data from other blocks, no firm prediction can be made as to the future survival of this critically restricted form. It seems quite possible, however, that this one factor alone could eliminate it.

2. Improve the contiguity of tidal marsh blocks throughout the Suisun complex. Although still retaining a higher percentage of its original extent of such marsh than the range of *pusillula*, the separation of major blocks is more widespread. On the southern side of Suisun Bay, this is due primarily to industrial and small harbor developments along the shore, but also in some locations to old filling alone. North of eastern Suisun Bay and Honker Bay, and particularly throughout the area from near Benicia and Cordelia east to Nurse Slough and D enverton, the whole wetland area is nearly all behind dikes and managed. Most impediments to dispersal of the sparrows between larger blocks of tidal marsh are interruptions in the narrow bands of tidal marsh along the numerous sloughs. Scollon (1993) assumed that the birds would tend to spread more toward their optimum lower salinity, with increasingly saline conditions in the western part of the Bay, and his recommendations of key areas to provide the best routes for such shift of range are:

- Along the shoreline of Joice Island (both sides) to “provide a critical link between populations along the shoreline of Grizzly Bay and those in the northern reaches of Suisun Marsh;”
- Along the northern shore of Honker Bay and southern shore of Suisun Bay, to link populations west and east; and
- Along the eastern reach of M onetza Slough, to connect the northern populations with those in the eastern Suisun Bay area. Scollon (1993) indicated that a 1991 proposal would have increased tidal marsh in this area, but it was apparently not accomplished, at least by the time of his writing.

3. Evaluate the management practices in the extensive gun club and wildlife agency lands throughout the western and northern parts of the range of maxillaris, with a focus on alternative types of vegetation control. Management practices should be sought that would provide corridors of brackish marsh across strategic areas that now act as barriers to these birds. At a minimum, provide reasonably continuous marsh-mud interfaces (even if freshwater) that might also provide for dispersal when it occurs during the late summer period. Such manipulations could be done in different locations (rotated) in different years. The area is noted for its provision of habitat for waterfowl. The recommendation made here is not intended to diminish that...
value, and might even be found to enhance it as well as habitat for the sparrows.

References


Anderson, J.R. and V.R. Jennings. 1981. Diked coastal saltmarsh [Breeding Bird Census #211]. Amer. Birds 35: 102. [from the authors’ description, the 14.7 acre plot they studied must have been a part of “Triangle Marsh”, which is diked off from the adjacent salt ponds but not from the Bay or Coyote Creek.]


Response of Birds to Managed Water Levels at Charleston Slough - A Case Study

William G. Bousman

Introduction

Charleston Slough is a former tidal estuary at the boundary of Palo Alto and Mountain View. A dike was placed across the outer slough in the 1920s, but a 60-inch pipe in that dike allowed tidal exchange up through the early 1970s, and a healthy salt marsh community of about 40 to 60 acres existed under the muted tidal regime of that period. In the mid-1970s, the 60-inch pipe was replaced with a 48-inch pipe placed higher in the dike, with the result that the tidal flow became highly muted (or non-existent), and the mean water level in the slough increased sufficiently that the salt marsh was inundated and lost.

Based on the destruction of the salt marsh in Charleston Slough, the San Francisco Bay Conservation and Development Commission (BCDC) prepared a Cease and Desist Order for Leslie Salt Company (now Cargill Salt). The company then transferred ownership to Mountain View, leaving that city to comply with the BCDC requirements. These requirements included the development of a new outlet structure in the outer dike that would provide a tidal fluctuation in the slough of 1.0 feet. At this time, the Santa Clara Valley Audubon Society (SCVAS) started a series of weekly censuses of the slough, and these continued through 1995, although the frequency of the censuses was reduced to every two weeks in the early 1980s.

The present case study is based upon a comparison of census data from the fall periods of 1980 and 1981. This comparison shows species composition and abundance changes that resulted from managed changes of water levels in Charleston Slough.

Methods

In the period from October 1980 through the end of 1981, Charleston Slough was censused on a weekly basis by four to five volunteers. Volunteers were given a one-week window in which to perform their censuses, but otherwise were allowed to select both the day of the census and the time of day. No attempt was made to census the birds in relationship to the tidal cycle. The censuses were made either on foot or by car using the public levee between Charleston Slough and the Palo Alto Flood Control Basin. Birds were censused within or above the slough to the centerlines of surrounding levees.

As essentially all portions of the 109-acre slough were visible from the public levee, the census recorded all birds present. Each census required about two hours. Observers were encouraged to count certain species as groups because of identification difficulties. These groups included greater and lesser scaup and short-billed and long-billed dowitchers. In some censuses, when large numbers of gulls were using the private levees for roosting, not all were identified to species.

Results

Census data show the lowest number of birds and species to occur in the summer, with the greatest number observed during fall, winter, and spring. The initial two years of the census were a time of substantial transition as construction for the new outlet structure was completed in February 1981, substantially changing the water levels in the slough. Prior to the removal of the cofferdam around the outlet structure, the water level was approximately 4.6 feet above Mean Lower Low Water (MLLW) and the entire slough was inundated. With the removal of the cofferdam, the water level dropped by approximately 1.8 feet, and 50 to 60 acres of mudflats were exposed. Very little tidal flow occurred in the slough because of siltation outside of the outer levee.

The water level in the slough started to increase in the winter of 1981-82, and this increase continued in later years so that as of 1996, the slough is largely inundated once again.

A comparison of the census data collected in October and November of 1980 (n=7) and October and November of 1981 (n=6) are shown here as representative of two different water regimes. The 1980 data are for the slough at its maximum water level, with essentially no exposure of mudflats. The 1981 data are for the slough at its minimum water level, with 60-80 acres of mudflats. The census data for the two periods are com-
pared in Table 7.9 for the 36 species counted and, included with the mean number of birds within each period, is the rank of that species relative to the others censused. Table 7.9 shows that 33 species were tallied in 1980 and 34 in 1981, while the mean species total for each census in 1980 was 3,221 birds and in 1981 was 3,559 birds.

The species list from Table 7.9 was sorted by the rank obtained in 1980 and the fifteen most common species are shown in Table 7.10 in rank order. The species totals and ranks for 1981 are also included in this table, but not in rank order. The percent of the total number of birds for 1980 is shown in the table as the final column and the cutoff at fifteen species is based on reaching 98% of the total number of birds recorded in all censuses.

The species list from Table 7.9 was sorted by the rank obtained in 1981 and the eighteen most common species are included in Table 7.11 by the 1981 rank order where, again, the inclusion of common species is based on reaching 98% of the total number of birds.

**Discussion**

The total number of birds counted in October and November of 1981 were approximately 10% greater than...
observed in 1980. Although the variance of the data has not been examined, a 10% change in abundance is unlikely to be significant. Similarly, the number of species comprising 98% of the entire bird population increased from 15 to 18 between 1980 to 1981 and, again, changes of this size are not believed to be significant. Thus abundance and diversity, as measured here, did not change substantially between the two different managed water regimes.

The changes that did occur between 1980 and 1981, however, were in species composition. In 1980 just five species comprised 81% of the population: ruddy duck, American coot, northern pintail, American wigeon, and western gull. The first four of these are species that clearly benefit from ponding and inundation. In 1981 censuses, however, these five species represented only 5% of the total population. Similarly, in 1981, a

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<th>1980 n=7 Rank</th>
<th>1981 n=6 Rank</th>
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different five species comprised 81% of the population: California gull, dowitcher species, herring gull, American avocet, and ring-billed gull. The gull species appear to have responded to the extensive mudflats as areas secure for loafing, while the two shorebirds species used the slough for foraging, as well as resting. These five species that were most common under the high water regime accounted for only 6% of the local population in the prior year’s census data.

The first point from this case study is that the two managed regimes used for this 109-acre former slough resulted in equal numbers of birds and species diversity. The water level between the two years was very different. In 1980, the water level was sufficiently high to inundate the entire slough such that no mudflats were available for foraging or secure roosts. In 1981, the water level had been lowered such that about half the slough’s area was available as mudflat for foraging or roosting. The species composition changed drastically between the two managed water levels – but these sort of changes are not quantified by simple ecological measures, such as total abundance (unchanged) or species diversity (unchanged).

A second point to consider is that the potential of Charleston Slough to be productive in terms of species abundance and diversity under two water management regimes is related to its scale compared to the entire estuarine system. Its size, 109 acres, is small compared to the overall South Bay system, and the variety and number of birds that can opportunistically take advantage of changes in such a small area are quite large. Although this case study probably applies to any similarly sized area within the estuarine system, it is not clear that it applies to areas that are substantially larger.

A third point, and one directly related to the first, is that if simple measures of ecological health such as abundance and diversity cannot be used to distinguish between two managed regimes, than what metrics can be used? As a community of individuals, we all may see and voice the need for some sort of balance in our management of estuarine systems. In particular, when we obtain stewardship responsibilities for a new component of the system, we all see the wisdom of studying this component to allow us to make wise and informed decisions. However, in the end, as in this Charleston Slough example, there may be no sound or rational basis for selecting a “correct” management regime. In the absence of sound ecological principals to be used for management, how do we make our choices? Do we use community values? Do we allow some portion of the electorate a vote? Or do we rely upon leaders of the scientifically-informed community to govern our choices?

Epilogue

This case study is concluded by reporting the “final” solution for the management of Charleston Slough, which was achieved recently, twenty years after the original Cease and Desist Order. It was agreed that the former non-functional outlet structure would be replaced with multiple pipes with sufficient tidal capacity, so that with time, a new tidal prism would be established and the blocked outer channel would be opened through scouring. This construction was accomplished during the 1998 summer season. Although there is an increased tidal range in the slough under this new regime, it does not meet the original requirements. The next step will be to assist tidal scouring by removing some of the Bay mud outside the new outlet structure. There is no prediction, presently, as to when a functioning salt-water marsh is likely to be restored.
The Use of Salt Ponds by Some Selected Birds Other than Shorebirds and Waterfowl

Howard L. Cogswell

The term “salt ponds” is traditionally used to cover any or all of the saltwater impoundments around the San Francisco Bay that are used in the various evaporation stages leading to the final crystallization of the salt in the salt company plant-sites (now only one company, Cargill Inc., Salt Division). A number of salt ponds have been taken out of such use by this company and the last other company to operate (Oliver Brothers of Hayward), but many of these remain mostly barren of vegetation. The accumulated salts in their bottoms make the rainwater ponded in them in the winter ecologically somewhat similar to regular evaporators even though the salinity changes over the year are probably greater than in any one pond in the ponds currently in use. I would recommend that all such inoperative saline areas be included in the salt pond habitat category by the Focus Team, as long as they are mostly barren of vegetation, even though they may be completely dry for half or more of the year. I also recommend that they be placed in the “diked seasonal wetlands” habitat category when there has been sufficient growth of vascular plants that birds and such mammals as voles and the salt marsh harvest mouse typical of such “marshes” can be expected to be present in significant numbers.

South Bay examples of places where such former salt ponds are still mostly barren are on parts of Bair Island, the southernmost and northwestern-most ponds of the sequence west of Alviso Slough (the northwest ponds are the “Knapp Tract,” and part of the National Wildlife Refuge (NWR)), parts of the Hickory Tract just west of the head of Newark Slough, and parts of the Baumberg Tract in Hayward. Other parts of Bair Island, and the Hickory and Baumberg tracts have already succeeded to more seasonal marsh than salt pond characteristics. Presumably, the large area of former salt ponds between the lower Napa River and Sonoma Creek also have portions that would fit in each category as well.

As thus restricted, the majority of the salt pond system is heavily used by birds. A large percentage of that use depends on the shoreline of the ponds, as well as the water, and a special value is easily traced to islands and the remote or undisturbed parts of the dikes between the ponds. Such habitat aspects seem inseparable from the ponds themselves for many species, including some of those mentioned below (but especially, of course, the shorebirds – to be addressed by another focus team). For species that nest or roost on these dry land inclusions in the salt ponds, it is the isolated placement of such land within the surrounding water that makes it valuable. A tenth-acre island in a salt pond is far more valuable as bird habitat than an acre of barren ground in an upland situation! In addition, some “Other Birds,” such as herons, often use the shoreline or shallower water adjacent to dikes and islands for foraging. Hence, this report in no way avoids consideration of the dry land parts of the system.

Salt Pond Operations

It is important in defining wetlands goals for salt ponds to understand the basics of operation of those ponds still used in the salt-extraction process. Details seem inappropriate here, except that: 1) the ponds are functionally connected into salinity gradient sequences as a result of systematic transfers of water among them; 2) each evaporator pond can thus be classified as a low, medium, or high-salinity pond, these categories having marked differences in forage value for birds; 3) some ponds are occasionally pumped nearly dry, and when this happens, their function in “isolation” from predators of roosts or, in the breeding season, of nests is sharply reduced; 4) the most highly saline ponds (crystallizers and the “pickle” ponds just before them in the sequence) have essentially no organisms suitable as food for birds, but in some locations still provide roosting protection; and 5) the “bittern ponds” (currently large ones at Newark and small ones at Redwood City) are filled with the brine remaining after sodium chloride crystallizes, and seem to provide essentially no wildlife value at all, but are a necessary part of the system since water quality restrictions now prohibit the disposal of that brine in the Bay.

Salt Pond Numbering Systems

Leslie Salt Company (which was bought by Cargill in the late 1980s) assigned numbers to each evaporator in the sequence of the water movement at each of their separate plants: Newark (plants 1 and 2), Mowry, Alviso, Redwood City, Baumberg (in west Hayward), and Napa. In several areas, additional ponds were added to the system at either the beginning or somewhere in the middle of the water-movement sequence, these ponds being distinguished by additional letters usually in suffix position. Except for the San Pablo Bay-Napa unit (sold to the State in the early 1990s), all these pond numbers continue in use by Cargill, even though the Alviso plant was closed long ago, and the Baumberg and Redwood City plants in 1972 (Redwood City being reopened a few years later). Water is no longer moved in sequence of the pond numbers, and old systems are combined with converging flows at several salinity levels.

Because duplicate numbers existed for many ponds in different areas, I came up with prefix letters making all the ponds in the NWR distinctively numbered. These are shown on the map in Figure 7.7. I have used “K”...
instead of "N" as prefix for the Newark system because N could be confused with the abbreviation for "north" when parts of the ponds are to be referenced. For the system of ponds in Hayward (the old Baumberg plant ponds) currently outside the NWR limits, I use the prefix "H" to avoid confusion with ponds B1 and B2 (Leslie's designations) in Mountain View – a part of the old Alviso system.

Current Studies of Salt Pond Ecology

A few studies have analyzed ecological or ornithological aspects of groups of salt evaporators with consideration of the differences among the ponds. The chief ones are:

1. Carpelan (1957), who studied a sequence of ponds in the Alviso area and reported primarily on the water chemistry, algae (including microscopic forms), and invertebrates, but mentioned some birds.

2. Anderson (1970), who studied five ponds south of Mowry Slough east to the Southern Pacific Railroad with respect to their use by birds in relation to different salinities and seasons of the year. However, his study omitted three ponds that were involved in the water-movement sequence from intake to three or four ponds short of the crystallizers, as the system then operated.

3. Gill (1977), essentially a summary of his 1971 survey of all breeding species found in bay-related habitats south of San Mateo Bridge (his M.A. thesis at San Jose State), with updates through 1975 from scattered later observations by him and others. Does mention salt ponds and their dikes as habitat features for many species.

4. Swarth (1981), who reported waterbird numbers pond by pond on the 11 ponds lying west of Coyote Hills between the new Alameda Creek Channel (which obliterated the former Coyote Hills Slough) and the east approach to Dumbarton Bridge.

5. Swarth, Akagi and Metropulos (1982), who incorporated the results from Swarth (1981) and extended counts of birds on the same set of ponds. The bird numbers were also analyzed for correlation with the biomass of major invertebrate populations (chiefly brine shrimp and water boatmen) as determined by hundreds of plankton hauls in the upper ¼ meter of water within three meters of a canoe, as well as to variations in water temperature, salinity, pH and depth and to wind direction and location about the periphery of each pond.

The last is by far the most thorough study of bird use of salt ponds and should be reviewed for information about each species to be considered in developing wetlands goals for this habitat type. Yet both that study and the earlier one by Swarth (1981) counted birds only during the four to six hours spanning the high tide point on the adjacent Bay, ostensibly to be able to report the "maximum use" of the ponds by birds. They reported in general terms only, e.g., that most shorebird species were essentially absent from the salt ponds when the nearby tidal flats were exposed. Cogswell (1981) found that exposed tidal flats at the lower tide-levels had a drawing power for shorebirds that even exceeded that of higher level tidal mudflats (that had been created by opening dikes of long-abandoned salt ponds at Hayward). This study included data from all tide levels, but of course none from existing salt ponds.

Another limitation of the studies west of Coyote Hills which was not mentioned by Swarth et al. (1982), is that some or all of the ponds they studied had been receiving water since 1972 from the approximately 20 evaporators that lie to the north of new Alameda Creek. That area is the former Baumberg Unit of the Leslie Salt operation, water from which since that year has been sent by siphon under the new creek channel to merge with that in ponds south of it. So, the Swarth et al. classification of ponds into "low, medium, and high" salinity apparently did not include any that were strictly the lowest or "intake only" salinity. In addition, under the Leslie operation since 1972, ponds K3, K2, and K1 south of the Dumbarton Bridge approach have been the final evaporators in this sequence before the water is pumped to the pickle pond and crystallizers. So Swarth et al. also had no bird data from these highest salinity evaporators, although they did sample invertebrates in the western part of Pond K3 (their Pond 14) and two small "pump donut" ponds close to the bridge.

In 1992-93, I did semi-monthly counts of shorebirds and ardeids, and noted general numbers of other birds on Pond K1 and adjacent Newark Slough as a part of the San Francisco Bay Bird Observatory (SFBBO) Shorebird High-tide Habitat Use Study. Other observers did the same in many units around the South Bay in the same periods. The SFBBO has also implemented (for some 17 years) a Colonial Waterbird Breeding Monitor-
Figure 7.7  Numbering System for the Salt Ponds of South San Francisco Bay. Developed by H.L. Cogswell
based on a map from the Cargill Salt 1998-1999 Completion Report.
Use of Salt Ponds by Some Key Bird Species

The following sections summarize information on the use of salt ponds by 16 of the 24 bird species that were selected by the "Other Birds" Focus Team as representative of our group of species. The summaries are based on information from each of the above-mentioned reports, plus from my own memory based on over 30 years of experience with birds on salt ponds (mostly in the Hayward to Fremont area). The literature makes frequent reference to "low," "medium," and "high-salinity" evaporators or salt ponds. Herein, I use the terms "salt ponds" and "evaporators" interchangeably, unless specific reference is made to crystallizers or pickle ponds, and I omit bittern ponds entirely because of their exceedingly low habitat value for any birds.

My "low salinity" pond category includes ponds with apparently higher fish populations than any west of Coyote Hills, and thus also encompasses high-fish ponds (intake ponds or the next few in long strings of evaporators), such as those sampled for fish by Lonzarich (1989). The "high-salinity" category also is extended at least by several ponds from what was considered to be that category by Swarth et al. (1982); but my "medium-salinity" ponds encompass the same range as theirs and a bit more on each end. In the future, a thorough analysis of data over the whole range of salinities might justify recognizing as many as five salinity categories rather than three.

Eared Grebe (Podiceps nigricollis) - Abundant on medium or medium-high saline evaporators from October through April, and present in lesser numbers in late August through September and in May. Anderson (1970) reported a maximum count of 6,330 in November on his "ponds with high-salinity". Swarth et al. (1982) reported their maximum of 5,565 in April 1980, but numbers the following spring were below 3,200. They had average counts of 500 and about 1,950 in November of the two successive years, and a very few were found through the summer. Correlation of eared grebe numbers per 10 hectares in each of the 11 ponds in the same study was strongly correlated (at p<0.02 level) with the grams dry weight of invertebrate biomass sampled in the same ponds. The brine shrimp and water boatmen that constituted nearly all of these biomass samples are apparently the prime food for this species during its stay here. Brine fly larvae and pupae, spending most of the time on or near the bottom of the salt ponds, may also be important foods, but were mostly missed in the surface plankton hauls.

There have been an estimated one to several thousand eared grebes on each of several different evaporators in the medium-salinity range in southwest Hayward in the fall of different years, and the same is probably true for each set of salinity-sequence ponds around the South Bay. In some areas, the numbers are higher in the April migration period. The total number may thus be 50,000 to 100,000 or more birds - a significant portion of the species population, although minor by comparison with its concentration in late summer at Mono Lake and the winter numbers on Salton Sea.

In 1983, a nesting colony of 70+ adult eared grebes on abandoned sewage ponds in Pleasanton, eastern Alameda County, produced at least nearly the same number of young before abandoning them still in flightless condition as the pond was drained in July. A smaller colony the same year also produced young in the Crittenden Marsh (brackish?) near Mountain View (W. Bousman, pers. comm.), and some persisted for several years in a nearby area. Swimming invertebrate or fish food resources, plus at least some emergent or submerged vegetation to which floating nests can be anchored, are required for grebe nesting. The salt ponds that supply the former very rarely have any of the latter, so their value as a breeding habitat for this species is very limited.

American White Pelican (Pelecanus erythrorhynchos) - Present as non-breeding visitors (most probably post-breeding) to the several lowest salinity salt evaporators in most of the intake sequences around the Bay from July through October in considerable numbers. Some 900 or so were regularly in the South Bay from 1965 through about 1980, then somewhat fewer. Numbers diminish through the winter at rates that vary among years (to zero in some years), followed by resurgence in some years in April and early May. There are a few records also through June.

Even in their peak period, local surveys of only one set of low-saline ponds may often reveal no white pelicans, while a few days later (or even later the same day) scores or hundreds may be present. The 900 total estimated above was supported by numerous flights by small airplane which I made in the 1970s and early 1980s, covering most of the salt ponds from Hayward, south to Alviso, and north through Mountain View, and occasionally on through Redwood City. Salt ponds in which white pelicans were seen feeding and/or resting (both activities in many cases) were H 10 and H 11 [H 10 has been an intake pond since 1973], and H 1 and H 2 (and sometimes the adjacent H 4 and H 7) in Hayward, K1A and K2A in Northwestern Fremont, M 1 and M 2 south of Moffy Slough in southwestern Fremont, A9 in Alviso, and A1 and A2W in Mountain View. Flocks
have been observed flying on nearly straight-line routes between some of these locations.

Pond K1 south of upper Newark Slough, but visible from the Dumbarton Bridge approach, was for years a major gathering point – and most of the white pelicans in the old “Dumbarton Bridge” records noted in Audubon Field Notes and other publications were probably there. After the closing of the Baumberg Salt Plant, however, and the re-ordering of the water flow in late 1972, this pond became a highly saline pond and has held no white pelicans that I know of, even though it still has the numerous small islands on which they formerly rested. In the fall of 1964 and 1965, the full 900 were estimated on ponds H4 through H7 near Turk Island hill, which should have been medium-salinity ponds in those years. Several color-dyed birds were observed there that had come from marking programs at Great Salt Lake, Utah, and Malheur Lake, Oregon, and as I remember, one from Yellowstone Lake. Small flocks have continued to use the H3C, H4C, and H5C ponds and rest on adjacent dikes in at least some winters since 1985, perhaps only when extended rainy weather dilutes those ponds or when the salt company moves water into them in particularly accelerated fashion so that many fish are included.

The presence of large numbers of small to medium-sized fish in water less than a meter deep, where they are susceptible to ready capture (by the swim-and-scoop method often used in concert by flocks of this species), is undoubtedly the chief factor that controls just which ponds are used by them. Barren islands or remote dikes seldom traveled by any human or large predator are augmenting favorable factors – but such features are sometimes reached by flights from the feeding areas if need be. For example, up to 100 or so white pelicans often come to sit on the dikes between effluent ponds in a fenced-off area of the Hayward Regional Shoreline, where they preen, sleep, and fly away again without (at least much of the time) swimming on those ponds or attempting to forage. For an account of the variety of fishes [and their relative numbers?] in salt ponds, the research work of David Lonzarich (1989) in the mid-1980s should be reviewed. A short commentary about his findings and enthusiasm for the study appeared in the San Francisco Bay Bird Observatory Newsletter 5(no.8, Sep.1986).

The question of whether white pelicans would ever breed on any islands in the salt pond system if they were located near a favorable food supply should be considered. Perhaps such a situation could be developed at the Napa-Solano County ponds now owned by the State. There were modest size flocks that used the low-salinity ponds there at least at times; but the fish-bearing ponds would have to be maintained as such, which presents an expense and management problem for public agencies not engaged in salt production. There might even be some possibility for a nesting island in a portion of the Baumberg Tract, purchase of which by the Wildlife Conservation Board has just been authorized. This would require maintenance of the site’s island characteristic (water all around it wide and deep enough to deter most predators) and continued presence of good fishing nearby – probably depending on Cargill Salt continuing Pond H10 as an intake.

Brown Pelican (*Pelecanus occidentalis*) – This species has been said (in the December 12, 1995 meeting notice for this Focus Team) to be “restricted to Central Bay.” This is not completely true. Small numbers frequently reach areas on the open Bay south to Dumbarton Bridge, and occasionally even farther. I recall seeing in late summer or fall up to 30 or 40 on Pond H1, an intake pond in Hayward, where they sat with the white pelicans or fished in their midst. Sometimes these birds would use the same surface-feeding technique as the white pelican, but sometimes it would plunge-dive at a shallow angle obviously designed to avoid striking

White pelicans and ducks on a salt pond west of old Alvarado, September 17, 1964.
the bottom in such shallow water. There is probably little that could be done in operating salt ponds to enhance their value to this species, beyond the provision of some islands in or near the fish-bearing ponds, which would be done for the white pelican or for various terns (see accounts for those species). The northernmost nesting ground for this species, at Point Lobos, Monterey County, has long been abandoned, and there seems little likelihood of any area north of that being chosen. Alcatraz or Red Rock Islands in the Central Bay would be more like their traditional sites, and have been “available” for many years.

**Double-Crested Cormorant (Phalacrocorax auritus)** – Birds of this species use the fish-bearing low-salinity salt ponds all year, but in considerable numbers primarily in the fall. Swarth et al. (1982) found a maximum of 82 (on Pond K2A) in their study area in December 1979 and 79 in late September 1981, although mean numbers were less than half of these figures. Numbers sometimes reach well over 100 in the fish-rich ponds H1 through H4 in southwest Hayward, where they rest on the dikes with pelicans, or on the numerous wooden posts in some of the ponds. In the Central Bay, they have roosted at night on power lines or power line towers for years, e.g., over 5,000 on the line to Brooks Island in the early 1940s (Bartholomew 1942, 1943a,b) and late 1940s (Cogswell, pers. obs.). That power-line has since been removed.

Gill (1977) did not find double-crested cormorants breeding in the South Bay during his 1971 survey. However, in more recent years, they have increasingly taken to nesting on the platforms or sometimes at junctions of legs and braces of powerline towers, e.g., many such south of the western part of San Mateo Bridge. In the salt ponds west of the Napa River, a few nests for a number of years in eucalyptus trees that had died when one salt pond was formed around them; and in the same general pond system, small numbers nested at least in the early 1980s on powerline towers over some of the salt ponds. I know of no such nesting yet in the South Bay, but there are many places where it might take place. However, the numbers of double-crested cormorants using salt ponds either for foraging and daytime resting or for nesting on structures within the ponds is probably rather small compared to the total number in or near the deeper parts of the Bay.

**Snowy Egret (Egretta thula)** – Although Swarth et al. (1982) reported a maximum count of 16 in their 11-pond study area, there are times in late summer or fall when just ponds H1 and H2, and H4 through H7 in southwest Hayward (at least H1, H2, and H4 having high fish populations) are utilized by over 100 snowy egrets at a time [and at least half to 2/3 that number of great egrets and many great blue herons as well]. There are probably similar large numbers on the low- to medium-salinity salt ponds near the largest nesting colony. This is in tule marsh within Artesian (M allard) Slough, west of Alviso, where the tremendous flow of San Jose northern Santa Clara County sewage effluent has converted the tidal marsh to that type. Snowy egrets formerly nested on outer Bair Island in the upper part of Salicornia marsh and on next to the outer levee and later in coyote bushes (Baccharis). Gill (1977) counted 340 and 362 active nests in these areas in 1971 and 1973, respectively. The salt ponds next to this site were taken out of use in the early 1970s, and the colony was decimated by predators (probably red foxes) in subsequent years.

Non-breeding snowy egrets will probably continue to use the low-salinity salt ponds for foraging and resting as long as they continue to have fish within their reach. The numbers foraging during the breeding season does drop in those ponds that are far from the colonies, which could perhaps be improved by providing other tall marsh areas in sloughs or freshwater ponds near those salt ponds. The only example of that known to me is the presence in 1995 of over 100 pairs of snowy egrets nesting (many young being produced successfully) in tall tules planted in effluent ponds at the Hayward Shoreline just three years earlier. A fair number of these birds flew south a mile or so to active salt ponds H10 and H11 and perhaps farther to forage. This colony has continued active at least to 1999.

**Black-Crowned Night Heron (Nycticorax nycticorax)** – Birds of this species use the low-salinity, fish-bearing salt ponds for foraging, but it seems in more localized fashion than the snowy egret. They seem to prefer places where water moves past their still-watch perch, such as gates or siphon-flows between ponds. Partly because they do much of their feeding at night, less is known about all the situations they use. Daytime roosting is usually in trees or within marsh growth, in small to fairly large flocks in the non-breeding season, e.g., in the primarily pickleweed marsh south of the outermost part of Alvarado Channel (old Alameda Creek). This marsh is being expanded eastward by 34 acres by Cargill Salt as mitigation for the impact on tidal marshes around the Bay by their levee-maintenance dredge (terms of the permit issued by the U.S. Corps of Engineers and San Francisco Bay Conservation and Development Commission). Several islands have or will be constructed in that vicinity also, and the attraction of all the diverse habitat may increase for this species.

Nesting colonies of night herons are established in very diverse types of vegetation: tall marsh (as in Artesian Slough, east of Alviso); on the ground or in coyote bushes as on outer Bair Island [609 nests in 1973 (Gill 1977) but colony later abandoned]; dense-foliaged shrubs and low trees (as on Alcatraz Island); and dense-canopied cypress trees even in urban areas (as for years in the City of Alameda). For this species, and potentially other herons as well, a grove of such trees, or even densely-growing eucalyptus, might be planted on Turk
Island hill, which is surrounded by salt ponds in southwest Hayward. Some type of predator-proof (or strongly deterrent) structure would have to be placed across each of the several levees that tie into the island. Perhaps the species most likely to respond to such trees, however, would be the great blue heron which already nests in salt ponds in that general vicinity on the scattered old duck blinds, Archimedes’ screw pumps, and one old gun club building.

Northern Harrier (Circus cyaneus) – This species nests in some salt marshes (upper portions, that are not flooded by tides in April or May), as well as in or near freshwater marshes or grassy flats inland. In the non-breeding season around the entire South Bay, and in the breeding period within a probable several-mile radius from their nest sites, they forage frequently over various marshes, fields, roadsides, dikes, and also those salt ponds that have numerous birds. The passage of a harrier in its typical low-level flight is sufficient to cause massed fly-ups of sandpipers and even ducks, and to send sparrows and other songbirds diving into cover. A persistent harrier on the Hayward shore successfully captured a red phalarope that only flew short distances from the surface of a pond on the predator’s first three or four swoops at it. The actual impact on the population of birds using the salt ponds from predation by harriers is unknown, but is probably small for healthy and alert fully grown individuals which see the harrier coming. There might be some serious impact on downy to partly grown young on the open dikes or islands, but I know of no studies addressing this question.

On the other hand, the population levels of the northern harrier itself (and of several other avian predators) are of concern, and should be watched. With reduction of their natural habitats, especially those required for successful nesting, the harrier may be in some trouble where its chief hunting opportunities are over salt ponds. In 1971, Gill (1977) found five nests near the Bay south of San Mateo Bridge, three of them in pickleweed. In 1979 and 1980, a pair nested in “Ideal Marsh” west of the salt pond series studied by Swarth et al. (1982), and in 1985, a pair nested (but doubtfully successfully) in pickleweed marsh of upper Newark Slough close to Coyote Hills. A pair (or two?) nests fairly regularly in or near Coyote Hills Regional Park. North of there, in summer 1999, one female was seen hunting low over a large colony of Forster’s terns, from which many young were found (partly to nearly fully eaten) on nearby levees. All of these birds probably foraged for considerable periods over nearby salt ponds. A study of the nesting success of the species in such situations compared to the more extensive habitat combinations of marshy ponds and fields of the Suisun area would be valuable.

Peregrine Falcon (Falco peregrinus) [and by implication also the wintering Merlin (Falco columbarius)] – Birds of these species forage over any of the salt ponds that harbor many birds of their normally preferred types. Individual peregrines sometimes become specialists and pursue and capture primarily or only ducks, others work on shorebirds, others take readily to rock or mourning doves, etc. Their hunting covers large areas, yet individuals or pairs take up quarters for the winter where there are elevated perches (e.g., powerline towers) to which they return regularly to rest, pluck, and consume their prey, etc. The merlin is from this standpoint a smaller edition of the peregrine, feeding on smaller birds and normally selecting lower perches. Both species forage over salt ponds frequently, but only the peregrine seems susceptible to having its habitat enhanced in the area. This is because more and more of them are now adapting to human structures for nesting (Bay Bridge, high-rise buildings, etc.). Where this is encouraged and protected within the several-mile foraging distance from salt ponds having many birds of suitable size, the salt pond resource would contribute to the breeding success of the peregrines.

California Clapper Rail (Rallus longirostris obsoletus) – This highly endangered subspecies depends almost entirely on tidal salt marsh for its foraging, retreat from danger, and for nesting. The impact on that habitat by salt-pond maintenance operations, now detailed and avoided or mitigated by the terms of the permit recently issued to Cargill Salt, will require ongoing watch to make sure that such damage does not resume. Clapper rails do occasionally step out into the open on a salt pond dike – but normally only when a super-high tide floods the adjacent marsh. This behavior is more prevalent around the dredge locks that the company builds to move the dredge between the outer ponds and the Bay. The low dikes of these locks soon become covered with pickleweed and other high-marsh plants; are they then a part of the salt marsh habitat, rather than the salt pond?

California Gull (Larus californicus) – Traditionally this species was only a non-breeding migrant and winter resident in coastal California, with large numbers beginning to arrive from inland and/or northern breeding areas in late July. Total populations reached 40,000 or more by September or October, the majority gathered for much of each day at or near the numerous solid-waste disposal sites near the Bay (Cogswell 1970, 1974). Numbers of California gulls dwindle somewhat near those sites (except in the Suisun Bay area), as winter numbers of larger gulls (western, and particularly the herring and glaucous-winged) increase; but spring migration of California gulls makes them again the most numerous species in March and early April. Especially in late summer and fall, but to some extent at other seasons also, many California gulls are seen foraging on the surface of medium to medium-high saline salt ponds – pecking here and there at the surface and apparently
obtaining brine shrimp as food in the same fashion as they do, for example, on Mono Lake where thousands of these gulls nest. This species, like other medium- to large-size gulls, also forages on open tidal flats, or aerially over many different habitats, including the salt ponds. Gulls of all species (except the smallest, Bonaparte’s) also use the salt pond dikes and islands for nighttime roosting or just as a protected place for daytime resting. Some flocks in the 1968-71 period even roosted at night standing in the saturated brine of crystallizers at Newark.

In 1982, about 30 pairs of California gulls were discovered nesting on small islands (old, partly submerged, dikes) in the salt ponds of the Knapp Tract, four miles northwest of Alviso. This was the first known breeding by this species in a coastal situation and the first documented record of nesting west of the Sierra Nevada-Cascade mountain ranges. By 1994, that colony had expanded to nearly 4,000 pairs (Layne 1995). Several smaller “satellite” colonies have also arisen—on a dike in the crystallizer area in Newark, two others south and north of Mowry Slough (the latter with the Caspian Terns) with a total 471 nests in early June 1994, and a fourth on a salt pond dike near Triangle Marsh north of Alviso, with 490 birds (but only 43 nests in 1994) (Layne 1995). The last-mentioned colony was for several prior years much larger. Some of these satellite colonies have been decimated in some years by red foxes, but the three largest colonies have persisted at least through 1995, and the one south of Mowry Slough through 1999.

For several years (1983-1988), hundreds of the young California gulls were banded by SFBBO teams, mostly at the original Knapp Tract colony, as part of a detailed study of the increase in numbers and production of young within a small part of that colony (Jones 1986). For the past few years, the Knapp area has been cut out of the saltwater movement in the Cargill system, so it largely dries up by mid-summer. Large numbers of gulls have continued to nest there, but nests were disrupted apparently by red foxes in 1994 [and 1995?]. It is obvious that the breeding range expansion by this species now includes San Francisco Bay—indeed, a few pairs have even bred at the Alameda Naval Air Station (NAS).

Elsewhere, nesting gulls have been noted as having negative impacts on the nesting of other waterbirds in their vicinity. Their possible impact through predation on eggs or young of herons, egrets, and terns should be evaluated here—particularly for the Forster’s and least terns. If it is decided that management of habitat should be directed toward maintaining the gull populations, as well as the other waterbirds, some decisions on just which areas can be provided or enhanced for the gulls will probably be necessary.

A major paper on the present range and status of California and ring-billed gulls throughout California is in preparation by W. David Shuford of Point Reyes Bird Observatory and Valerie Layne of San Francisco Bay Bird Observatory. A poster summarizing the growth of the California gull colonies around southern San Francisco Bay was presented by Hanson and Ryan, SFBBO staff, at the 1997 meeting of the American Ornithologists’ Union.

**Western Gull (Larus occidentalis)** – This is the most “marine” in habits of all the large gulls in the Bay Area. It successfully maintains a more-or-less equal-numbers status through the mid-winter period in the Central Bay, even when large wintering numbers of glaucous-winged and herring gulls are present. Particularly in late summer and fall, considerable numbers of western gulls also spread to the inner reaches of the Bay—south to Alviso and east to at least Antioch. They are always in the minority among gulls in these areas, using the disposal sites, bay and shores, salt ponds, and marsh sloughs as forage areas.

The traditional major nesting grounds of this species, the Farallon Islands, continues to hold by far the largest number of breeding birds, over 10,000 pairs. Some of these nesting birds commute to Bay Area disposal sites for food, as evidenced by market-prepared chop and steak bones regurgitated on the nesting grounds. Nesting colonies existed on several islands in the Central Bay by at least the late 1960s, and on top of Pier 45 in San Francisco by 1971. A few pairs of western gulls also were found nesting amid the California gulls on the Knapp Tract, northwest of Alviso, in the 1980s by SFBBO teams, who banded a few of their young. One nesting pair was there in 1994 (Layne 1995). Thus, the salt pond habitat is at least marginally a breeding habitat for this species, and the remarks pertaining to the California gull (above) also apply to the western gulls, although in lesser degree.

**Caspian Tern (Sterna caspia)** – This largest tern has nested on dikes between or on barren islands within salt evaporators in the South Bay since at least 1922 in a colony that had 287 active nests in 1931 (DeGroot 1931). This colony apparently shifted exact location over the years, but was always near the eastern approach to Dumbarton Bridge. The colony was observed to have 378 nests on 21 May 1943 (Miller 1943), 188 nests plus 202 young out of nests on 21 June 1952 (C. Sather, oral report in my field notes), and 499 active nests (most with eggs, few with young) on 14 May 1954 (personal field notes). At least on these later dates, the colony was on the dike separating salt ponds K5 and K7, north of the bridge approach, and persisted there until at least the mid-1960s.

Anderson (1970) discovered a thriving colony of Caspian terns on the southern part of the curving dike between ponds M4 and M5, east of Albrae Slough. This site continued in use, at least intermittently, to 1996, when it was decimated by red fox predation a second time.
Large numbers of this species have also nested in the 1970s and 1980s (at each site for a few years only) on the dike between ponds H10 and H11 (and low islands in H10), on the western end of a long peninsular dike between ponds H4 and H7, and on a still barren area of a former salt evaporator on outer Bair Island, Redwood City (estimated 500-600 pairs in July 1975 per Gill 1977). The H10/H11 colony (1969-74) was plagued by salt foam blowing across the nesting birds and was abandoned abruptly. The H4/H7 colony (1976-86?) was perhaps the new location of these same birds and continued expansion to over 100 pairs, including some on the still drivable levee between H4 and H2 - until all the levees in this area were retopped with dredge spoils. The SFBBO observers checking the colony in 1986 (or 1987?) recorded the mostly failed efforts of the terns to nest on the deeply cracked drying mud. Swarth et al. (1982) recorded a maximum of 9 Caspian terns in the salt ponds west of Coyote Hills in 1980 and 1981 - at which time the colony was only a mile or two to the north. The Bair Island colony suffered from probable red fox predation, but some birds continued to gather at the site for several years after any successful nesting. Caspian terns were also regularly seen in the San Pablo Bay-NapaRiver Unit of salt ponds (at least along Highway 37), and probably nested somewhere in that system.

Foraging by birds of this species is wide-ranging, but a majority apparently seeks fish in the open Bay. Certainly, compared to the Forster’s terns, relatively few are seen diving over salt ponds, even the fish-rich low-salinity ones. Small numbers of Caspian terns also appear during the nesting season over reservoirs in the hills quite far from the Bay, and are known to carry fish from such locations back to the colonies (e.g., tags from fish stocked at Del Valle Reservoir have been found on the major South Bay colonies). Hence, the nesting and roosting safety of islands and remote dikes in the salt pond system are the prime ways in which this species is benefited by this habitat.

**Forster’s Tern (Sterna forsteri)** - This species is found mostly from May through September in or near salt pond habitats, when it is nesting or the fledged young are still under intensive care by the adults. A few are present through the winter in favored locations around the Bay, but are seldom seen on salt ponds then. Nesting takes place at numerous locations, mostly on small islands within the low- to medium-low salinity ponds (where fish are abundant, and where the newly fledged young may first try their own plunge-dives). Some colonies, however, are on islands within medium-high to high-salinity ponds, e.g., K1, K2 and K3 just south of the eastern approach to Dumbarton Bridge and Newark Slough. There are no fish in those ponds, and foraging is entirely in the slough or the open Bay. The total number of active nests tallied south of San Mateo Bridge in 1971 by Gill (1977) was 935, while in 1994 somewhat over 1,000 were in 27 colonies there (Layne 1995), nearly all of them on islands or remote dikes in salt ponds. Hence, there is no obvious increase or decrease in the total population in recent years.

Colonies of Forster’s terns sometimes persist for many years at the same sites, but in other cases shift to new locations in the same general region. The species seems more able to succeed with smaller colonies than the Caspian, and is thus able to use even quite small islands. However, where these are in salt ponds subject to spring or early summer draw-down by the pond operators, their success is jeopardized by the relatively much easier access to the sites by predators. This took place, for example, in at least two years between the middle and end of May in Pond H8A and a colony of over 100 pairs disappeared. A goal for optimal habitats for this species...
would certainly include some careful planning and cooperation to keep higher water levels in ponds with nesting islands until the young terns were flying.

**California Least Tern (Sterna antillarum browni)** - This endangered subspecies has its northernmost ongoing breeding colony at Alameda NAS, where they are normally present from May to August. Some pairs with the earlier fledglings apparently move from this colony by mid-August to other sites with abundant small fish and nearby resting sites barren of vegetation and free of most disruption by predators or humans. These post-breeding assembly areas are considered important for the successful maturing of the young birds, including their development of adequate foraging skills. In the salt pond system, the areas most regularly used for this sort of activity are ponds H 10 and H 11 (resting on the dike between them or on islands), and H 1 and H 2 in southwest H ayward.

A few late nests of this species, probably by pairs that had nests interrupted at Alameda, have been found (June - August) in both of these areas; but I believe none persisted through hatching of the eggs. For several years small numbers of least terns gathered in summer on barren islands in the experimental ponds (treated sewage effluent) on the H ayward shore; and in 1990 one pair nested there successfully (young fledged) on the one island to which crushed oyster-shells had been added to attract them. There has not been any subsequent nesting there through 1999, however. In 1972-75 (Gill 1977), a small colony with at least 14 active scrapes in 1975 did nest successfully on the barren flat (former salt pond) near the Caspian tern colony on outer Bair Island— the least terns perhaps depending on the vigorous defense by the larger species against predator intrusion into the area. A few pairs also have nested near Pittsburg, on or near industrial wastewater ponds in the marsh zone.

The question of whether some parts of the salt pond system could be managed to provide enhanced habitat for this species is made more difficult to answer by the varying success of the “outliers” from the main Bay Area colony itemized above. The only procedure likely to succeed would be the provision of low, barren islands in or very close to low-salinity (intake) salt ponds and/or large channel-mouths at the bayshore. Elsewhere in its range, the least tern is known to take advantage of new dredge spoil islands (but of sandy spoils), and has done so to some extent in the Alameda- O akland Airport area. The pervasively muddy nature of the substrate in the salt pond areas, and the vigorous growth of marsh or other halophytic vegetation on islands that are just above the waterline would have to be overcome, since these are ecological features which this species tends to avoid.

**Coast Savannah Sparrow (Passerculus sandwich- ensis alaudinus)** - Formerly known (at least in part) as the Bryant’s marsh sparrow (P. s. bryanti), this is the form of the continent-wide savannah sparrow that breeds in the coastal strip (especially the summer fog belt) of California from Humboldt County south to the vicinity of Morro Bay in San Luis Obispo County. It is somewhat more “marsh-adapted” than most subspecies of savannah sparrows, but notably less so than beldingi of southern California. Savannah sparrows of several other subspecies occur in migration or winter in grasslands and weed fields and to some extent in the marshes around the Bay. Many of these are essentially impossible to distinguish from alaudinus in the field, so habitat-use differences are very poorly known for these seasons.

The nesting habitat of bryanti [now alaudinus] was originally described as “tidal marshes” around San Francisco Bay, but gradually the form was found to occupy also the more moist grasslands of nearby valleys and outer Coast Range hills. Johnston (1968) provides the best summary - “It maintains populations in two main types of habitat in coastal California: the Salicornia association of tidal marshes and the grassland associations of the coastal fog belt.” He gives some details from his own research on San Pablo M arsh and from earlier work elsewhere by Marshall (1948), and further compares the habitat niche of this form with the overlapping salt marsh form of song sparrow: “… on salt marshes [it] is nearly limited to the broad expanses of low-lying salicornia (Salicornia ambigua) on the older and higher parts of marshes... [that] lie back of that salt marsh vegetation (cordgrass, Spartina foliosa) best suited to frequent submergence by tidal flooding.” The song sparrow’s favorite forage area is the mud banks of the small channels within the latter type of marsh, but Johnston (1968) does mention that savannah sparrows are occasionally seen in that habitat in the higher marsh.

Eight nests were found in 1971 by Gill (1977) in his intensive survey of breeding birds south of San M ateo Bridge, where he cited the preferred nesting habitat as “levee tops grown to annual grasses and high pickleweed growing on the levee banks.” Gill (1977) further estimated the overall nesting population of the area at 800 to 1,000 pairs. How many of these would be within the salt pond zone is unclear.

With respect to the salt pond habitats, savannah sparrows are often seen in winter and migration periods along those dikes that have at least frequent patches of herbaceous vegetation or salicornia, sometimes well away from marshes or herblands of greater width. The numbers that use such linear habitat zones are much greater, however, where there is upper-zone tidal marsh or seasonal wetland adjacent across the dike from the salt pond. Presumably, some of these birds are of the breeding form alaudinus, but this should be verified by in-hand identification. Where there is as much as an acre or two of mixed herbs and salicornia, such as along the Dum barton Bridge highway fill through the salt pond zones, scattered individual savannah sparrows sing on territory (and
are thus presumably alaudinus) through May and June. Elsewhere, in my experience, singing savannah sparrows are found in the diked-off “seasonal wetland” habitat as well as in the upper parts of the tidal marshes, and the species appears to be a marginal one with respect to use of even the “upland” bits included in the salt-pond complex.

Alameda Song Sparrow (Melospiza melodia pusillula) and Samuel’s Song Sparrow (M. m. samuelis) - These are two of the three subspecies of song sparrows endemic to the tidal marshes and immediate vicinity in the San Francisco Bay system. The third one is M. m. maxillaris of the Suisun Bay area, where there are no salt ponds. All three forms were studied in detail from the standpoint of validity as subspecies, distribution, and habitats by Marshall (1948), and have been further updated with respect to ties to the remaining tidal marshes by Marshall and Dedrick (1994). Only samuelis, found in the marshes about San Pablo Bay and the Marin County side of San Francisco Bay, has been studied with respect to its breeding cycle, territory, and foraging habits (Johnston 1956). This involved three years of intensive work with banded birds on the San Pablo Marsh, Contra Costa County, and illuminated several ways, other than morphology, in which birds of that subspecies differ from their upland neighbors: 1) their territories are smaller and usually linearly arranged along the small curving channels in the marsh; 2) birds use the Grindelia or other higher plants of the marsh for nesting and/or high-tide refuge, or even leave their territories for nearby upland edges during the higher high-tides; 3) the peak period of first brood egg-laying is March-April and renesting laying peaks in May, thus most nests avoid being flooded during the extra-high tides of June, whereas the upland song sparrows usually have eggs or nestlings from April to early July; and 4) dispersal of the young birds is much more limited than in the upland song sparrows, averaging only 185 meters from their hatching place. Presumably similar adaptations to the intertidal habitat exist in pusillula and maxillaris.

In the South Bay, Gill (1977) found 17 nests of pusillula in 1971, all between 12 M and 7 M May, and estimated the total population south of San Mateo Bridge at 1,800 pairs. Birds of that race are commonly seen using the levees and dikes adjacent to its normal marsh habitats, and presumably samuelis in the San Pablo Bay–Napa River salt pond system does the same. No special study of the use of such habitats has been done, so the comments that follow are rather random recollections from my own experience with pusillula.

During the higher high tides of spring tide periods, when all the small channels in the tidal salt marsh are flooded, and even much of the salicornia zone is underwater, song sparrows that live or forage in such areas at other times of tide move to any nearby “above-water” refugium available. Where that is a salt pond dike, the birds gather and forage in and near the uppermost bits of vegetation – a few sometimes crossing the dike to forage on brine flies, e.g., at the edge of the salt pond itself. Occupancy that is more permanent at all tide levels, even singing on territory, occurs sporadically along the bayfront levees of the outermost salt evaporators, even up to a half-mile or so from any real salt marsh. The few birds I have seen in such places forage amid rip-rap and the wisps of salicornia and ruderal herbs that grow in such places. This sort of habitat is also occupied where the habitat landward of the levee is seasonal wetland, with at least some “marsh” vegetation. I have also noted several song sparrows (presumably pusillula) on the dike with no rip-rap between Pond K1 (or PP1 of Leslie’s system) and the former salt ponds of the Hickory Tract in Newark. This dike is now partly covered by salicornia and cuts off bits of the original head of N ewark Slough, where there is always some residual water. Elsewhere, pusillula sparrows (or presumably such) are sometimes seen taking refuge or foraging on small walkways, pump structures, or associated fences within the edges of salt ponds themselves. All of these instances are, however, marginal to the main habitat occupied by these forms—the tidal salt marshes – and these salt marsh song sparrows actually are more restricted to the vicinity of such marshes than are the savannah sparrows. Hence, conservation goals should emphasize that type of habitat for all three endemic salt marsh adapted subspecies of the song sparrow.

References

________. 1981. Populations of birds using new tidal lagoons compared to an outer tide flat on San Francisco Bay at Hayward, California. Tiburon


Jones, P.A. 1986. Aspects of the reproductive biology of the California gull in Alviso, California. M.A. Thesis, San Francisco State Univ. 96pp. [has data on colony growth, clutch and egg sizes, growth of young in relation to various factors, and nest success compared to that in Mono Lake and Great Salt Lake colonies].

Layne, V. 1995. Colonial waterbird monitoring on South San Francisco Bay, a 1994 summary. San Francisco Bay Bird Observatory report to San Francisco Bay N atl. Wildl. Refuge. 9pp. [Giving for each species the range of observation dates, maximum count of adults and nests and date of such for each of numerous sectors around the South Bay. Comparable reports for prior years also on file at the NWR and at SFBBO.]


Miller, M.T. 1943. Census of a colony of Caspian terns. Condor 45: 220-225 [the same colony as described by DeGroot 1931].


Swarth, C.W. 1981. A study of waterbird abundance and distribution on salt ponds in South San Francisco Bay. M.S. report for independent study class, Calif. State Univ., Dept. of Biol. Sci. 34pp. [pp. 24-34 present the raw data by each pond and census date].

Appendix A

Author Contact Information
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David G. Ainley
H.T. Harvey and Associates
3150 Almaden Expressway, Suite 145
San Jose, California 95118.
408-448-9450
dainley@harveyecology.com

Joy D. Albertson
Don Edwards SF Bay National Wildlife Refuge
P.O. Box 524
Newark, California 94560-0524
510-792-0222
joy_albertson@fws.gov

Janice M. Alexander
University of California, Berkeley
151 Hilgard Hall
Berkeley, California 94720-3110
510-643-5430
jalexand@nature.berkeley.edu

Peter R. Baye
U.S. Fish and Wildlife Service
Endangered Species Division
P.O. Box 2012
Mare Island, California 94592
707-562-3003
peter_baye@fws.gov

Dennis R. Becker
California Department of Fish and Game
Grizzly Island Refuge
2548 Grizzly Island Road
Suisun, California 94585-9539
707-425-3828
grizzlyisland@dfg.ca.gov

William G. Bousman
321 Arlington Way
Menlo Park, California 94025
650-322-5282

Andrée M. Breaux
SF Bay Regional Water Quality Control Board
1515 Clay Street, Suite 1400
Oakland, California 94612
510-622-2324
ab@rb2.swrcb.ca.gov

Michael L. Casazza
U.S. Geological Survey
Western Ecological Research Center
6924 Tremont Road
Dixon, California 95620
707-678-0682 ext.629
mike_casazza@usgs.gov

Steven C. Chappel
Suisun Resource Conservation District
2544 Grizzly Island Road
Suisun, California 94585
707-425-9302
srcd@castles.com

Howard L. Cogswell
1548 East Avenue
Hayward, California 94541-5313
510-581-2201
CABIRDS@aol.com

Ron Duke
H.T. Harvey and Associates
3150 Almaden Expressway, Suite 145
San Jose, California 95118
408-448-9450
rduke@harveyecology.com

Jules G. Evens
Avocet Research Associates
P.O. Box 839
Point Reyes California 94956
415-663-1148
jevens@svn.net

Phyllis M. Faber
212 Del Casa
Mill Valley, California 94941
415-388-6002
pmfaber@aol.com

Leora Feeney
1330 Eighth Street
Alameda, California 95401
510-522-8525

Steve Foreman
LSA Associates, Incorporated
157 Park Place
Point Richmond, California 94801
510-236-6810
steve.foreman@lsa-assoc.com

Stephen L. Granholm
LSA Associates, Incorporated
157 Park Place
Point Richmond, California 94801
510-236-6810
stephen.granholm@lsa-assoc.com

Brenda J. Grewell
University of California, Davis
Environmental Study & Policy Development
Davis, California 95616
530-752-4326
bjgrewell@ucdavis.edu
Michael F. McGowan
San Francisco State University
Tiburon Center for Environmental Studies
P.O. Box 855
Tiburon, California 94920
415-338-3514
mcgowan@sfsu.edu

A. Keith Miles
U.S. Geological Survey-Davis Field Station
Western Group in Ecology
1 Shields Avenue, Kerr Hall, Rm 278
Davis, California 95616
530-752-5365
keith_miles@usgs.gov

Michael R. Miller
U.S. Geological Survey-Dixon Field Station,
6924 Tremont Road
Dixon, California 95620
michael_r_miller@usgs.gov

Gary W. Page
Point Reyes Bird Observatory
4990 Shoreline Highway
Stinson Beach, California 94970
415-868-1221 ext.23
gpage@prbo.org

Thomas P. Ryan
San Francisco Bay Bird Observatory
P.O. Box 247
Alviso, California 95002
408-946-6548
wtswift@aol.com

Michael K. Saiki
U.S. Geological Survey-Dixon Field Station
Western Fisheries Research Center
6924 Tremont Road
Dixon, California 95620
707-678-0682 ext.617
michael_saiki@usgs.gov

Howard S. Shellhammer
San Jose State University
Department of Biological Sciences
San Jose, California 95192-0100
408-924-4897
shellham@email.sjsu.edu

Ted R. Sommer
California Department of Water Resources
Environmental Services Office
3251 S Street
Sacramento, California 95816-7017
916-227-7537
tsommer@water.ca.gov

Lynne E. Stenzel
Point Reyes Bird Observatory
4990 Shoreline Highway
Stinson Beach, California 94970
415-858-1221
lstenzel@prbo.org

John Y. Takekawa
U.S. Geological Survey-SF Estuary Field Station
P.O. Box 2012
Vallejo, California 94592
707-562-2000
john_takekawa@usgs.gov

Robert N. Tasto
California Department of Fish and Game
20 Lower Ragsdale Drive, Suite 100
Monterey, California 93940
831-649-7142
btasto@dfg.ca.gov

Scott Terrill
H.T. Harvey and Associates
3150 Almaden Expressway, Suite 145
San Jose, California 95118
408-448-9450
sterrill@harveyeology.com

Lynne A. Trulio
San Jose State University
1 Washington Square
San Jose, California 95192-0116
650-964-5944
ltrulio@email.sjsu.edu

Nils Warnock
Point Reyes Bird Observatory
4990 Shoreline Highway
Stinson Beach, California 94970
415-868-1221 ext.37
nilsw@prbo.org

Sarah E. Warnock
Point Reyes Bird Observatory
4990 Shoreline Highway
Stinson Beach, California 94970
415-868-1221 ext.37

Frank G. Wernette
California Department of Fish and Game
4001 North Wilson Way
Stockton, California 95205
209-948-7800
fwernett@delta.dfg.ca.gov
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Teams of scientists have developed habitat goals for the baylands ecosystem of San Francisco Bay. In creating this long-term vision, they assembled and synthesized information on more than 100 species of fish and wildlife; they also described the habitats that support these species. This report summarizes this information and is suited for anyone interested in learning more about the Bay’s plants and animals.